



Whole-Genome Sequence of Toxic Freshwater Cyanobacterium *Chrysosporum ovalisporum* Strain UAM-MAO

Soledad Sanz-Alférez,ª Carolina E. Rodríguez-Sanz,ª Ángel Barón-Sola,ª Francisca F. del Campoª

^aDepartamento de Biología, Universidad Autónoma de Madrid, Madrid, Spain

ABSTRACT Here, we report the complete nucleotide sequence of *Chrysosporum ovalisporum* UAM-MAO, a filamentous, cylindrospermopsin-producing cyanobacterium involved in bloom forming in freshwater systems worldwide. It was isolated from an artificial pond in Madrid, Spain. The genome sequence contains 336 contigs, consisting of 7,478,035 bp and 2,851 putative protein-coding genes.

The development of toxic harmful algal blooms is frequently correlated with climate change and eutrophication, in which cyanobacteria are significant, since a high concentration of nitrogen and phosphorous contribute to the massive proliferation of toxic cyanobacteria in water reservoirs (1). Cyanobacteria are organisms producing a great number of secondary metabolites with biological activity, including toxic products denominated cyanotoxins, which are common pollutants in freshwater systems. Among them, cylindrospermopsin (CYN), a potent alkaloid and protein synthesis inhibitor, is of increasing concern due to the growing number of detections reported worldwide in the last few years (2). Various cyanobacterium species have been identified as CYN producers, and *Chrysosporum ovalisporum* is one of them. This bacterium has an invasive behavior and is becoming an important health hazard because most strains are toxic (3). The gene clusters (*cyr* and *aoa*) involved in CYN synthesis have been completely described in several cyanobacteria (4, 5), showing several rearrangements in gene order and different flanking regions.

Chrysosporum ovalisporum (formerly Aphanizomenon ovalisporum) strain UAM-MAO was isolated from an artificial pond in Juan Carlos Park, Madrid, Spain, during a bloom formation. The production of CYN has been detected, and the aoaA, aoaB, and aoaC gene sequences and expression have been characterized (6, 7). DNA was extracted following mechanical disruption in cetyltrimethylammonium bromide (CTAB) buffer and treatment with proteinase K and lysozyme. A MiSeq paired-end genomic library was prepared and sequenced on an Illumina MiSeq platform (Parque Científico de Madrid, Spain). The reads were processed by Prinseq, and a de novo assembly was performed using SPAdes (8). Complementary metrics were examined by applying QUAST (9) to complete the annotation of the full genome using the BG7 system (10). Bioinformatic analysis revealed that the genome of UAM-MAO is approximately 7.47 Mbp in size, distributed in 336 contigs (\geq 1,000 bp), with a GC content of 50.39%. The annotation identified 2,851 coding sequences. Furthermore, studies of the UAM-MAO cyr gene cluster have been done, bearing a nucleotide sequence (up to 96% identity) similar to those of to Aphanizomenon sp. strain 10E6 (4) and Rhaphidiopsis curvata (5), but their genes are arranged in a different manner. In addition, secondary metabolites and other toxin biosynthesis genes were predicted by antiSMASH (11) using nonribosomal peptide synthetase (NRPS) and/or polyketide synthase (PKS) gene identification.

The availability of this genome may allow for a greater understanding of gene diversity and evolution within cyanobacterium organisms; also, it will improve our

Received 5 June 2018 Accepted 13 September 2018 Published 11 October 2018

Citation Sanz-Alférez S, Rodríguez-Sanz CE, Barón-Sola Á, del Campo FF. 2018. Wholegenome sequence of toxic freshwater cyanobacterium *Chrysosporum ovalisporum* strain UAM-MAO. Microbiol Resour Announc 7:e00819-18. https://doi.org/10.1128/MRA .00819-18.

Editor Irene L. G. Newton, Indiana University Bloomington

Copyright © 2018 Sanz-Alférez et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Soledad Sanz-Alférez, soledad.sanz@uam.es. knowledge of the *cyr* cluster gene organization, as well as help to predict the regulation of cyanotoxins and secondary metabolite biosynthesis.

Data availability. This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession no. CDHJ00000000. The version described in this paper is the first version, CDHJ01000000.

ACKNOWLEDGMENT

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

REFERENCES

- 1. Paerl HW, Otten TG. 2013. Harmful cyanobacterial blooms: causes, consequences, and control. Microb Ecol 65:995–1010. https://doi.org/10 .1007/s00248-012-0159-y.
- Kinnear S. 2010. Cylindrospermopsin: a decade of progress on bioaccumulation research. Mar Drugs 8:542–564. https://doi.org/10.3390/ md8030542.
- Mehnert G, Leunert F, Cirés S, Jöhnk KD, Rücker J, Nixdorf B, Wiedner C. 2010. Competitiveness of invasive and native cyanobacteria from temperate freshwaters under various light and temperature conditions. J Plankton Res 32:1009. https://doi.org/10.1093/plankt/fbq033.
- Stüken A, Jakobsen KS. 2010. The cylindrospermopsin gene cluster of *Aphanizomenon* sp. strain 10E6: organization and recombination. Micro-biology 156:2438–2451. https://doi.org/10.1099/mic.0.036988-0.
- Jiang Y, Xiao P, Yu G, Sano T, Pan Q, Li R. 2012. Molecular basis and phylogenetic implications of deoxycylindrospermopsin biosynthesis in the cyanobacterium *Raphidiopsis curvata*. Appl Environ Microbiol 78: 2256–2263. https://doi.org/10.1128/AEM.07321-11.
- Barón-Sola A, Gutiérrez-Villanueva MA, del Campo FF, Sanz-Alférez S. 2013. Characterization of *Aphanizomenon ovalisporum* amidinotransferase involved in cylindrospermopsin synthesis. MicrobiologyOpen 2:447–458. https://doi.org/10.1002/mbo3.78.

- Barón-Sola Á, del Campo FF, Sanz-Alférez S. 2017. Influence of glycine and arginine on cylindrospermopsin production and *aoa* gene expression in *Aphanizomenon ovalisporum*. Toxins 9:355. https://doi.org/10 .3390/toxins9110355.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. https:// doi.org/10.1093/bioinformatics/btt086.
- Pareja-Tobes P, Manrique M, Pareja-Tobes E, Pareja E, Tobes R. 2012. BG7: a new approach for bacterial genome annotation designed for next generation sequencing data. PLoS One 7:e49239. https://doi.org/10 .1371/journal.pone.0049239.
- Medema MH, Blin K, Cimermancic P, de Jager V, Zakrzewski P, Fischbach MA, Weber T, Takano E, Breitling R. 2011. antiSMASH: rapid identification, annotation and analysis of secondary metabolite biosynthesis gene clusters in bacterial and fungal genome sequences. Nucleic Acids Res 39:W339–W346. https://doi.org/10.1093/nar/gkr466.