GENOME SEQUENCES





Whole-Genome Sequencing of *Pantoea* sp. Strain RIT388, a Potential Oral Opportunistic Pathogen Isolated from a Chewing Stick (*Distemonanthus benthamianus*)

Han Ming Gan,^{a,b} Anutthaman Parthasarathy,^c Kurtis R. Henry,^d Michael A. Savka,^c Bolaji N. Thomas,^d André O. Hudson^c

^aCentre for Integrative Ecology–School of Life and Environmental Sciences, Deakin University, Victoria, Australia ^bGenomics Facility, Monash University, Selangor, Malaysia

^cThe Gosnell School of Life Sciences, Rochester Institute of Technology, Rochester, New York, USA

^dCollege of Health Science and Technology, Rochester Institute of Technology, Rochester, New York, USA

ABSTRACT In this study, we report the isolation, identification, characterization, and whole-genome sequence of the endophyte *Pantoea* sp. strain RIT388, isolated from *Distemonanthus benthamianus*, a plant known for its antifungal and antibacterial properties that is commonly used for chewing sticks.

The genus *Pantoea* is made up of Gram-negative bacteria within the *Erwiniaceae* family of enterobacteria and contains both free-living and host-associating species (1, 2). These bacteria form yellow mucoid colonies and associate with a variety of hosts, which include plants, insects, larger animals, and humans (1, 3). Some *Pantoea* species are well-known plant pathogens (4–6). *Pantoea agglomerans* has been isolated from patients with septic conditions, catheters, and trauma wounds, as well as from those with nosocomial infections (7, 8). Fatal outbreaks in neonates are known (9, 10). Apart from *Pantoea agglomerans*, *Pantoea septica*, *Pantoea dispersa*, and *Pantoea latae*, strains previously considered plant-associated or environmental isolates, *Pantoea allii* and *Pantoea eucalypti* are considered clinical specimens (11, 12).

Pantoea sp. strain RIT388 was isolated on tryptic soy agar during a study to identify endophytic bacteria from *Distemonanthus benthamianus* (13). *Pantoea* sp. RIT388 cells are rods and are approximately 1.5 μ m long (Fig. 1). *D. benthamianus*, used for oral hygiene, is a semideciduous perennial tree found in second-growth forests in Nigeria, Cameroon, and Ghana (14). A recent study showed that extracts from the bark possess bactericidal activity against *Staphylococcus aureus* and *Streptococcus mutans*, two bacterial species that are often associated with skin and dental infections, respectively (15).

Genomic DNA (gDNA) was isolated from a 5-ml culture of *Pantoea* sp. RIT388 grown in tryptic soy broth using the GenElute bacterial gDNA isolation kit (Sigma-Aldrich, USA) according to the manufacturer's protocol. The gDNA was normalized to a concentration of 0.1 ng/µl based on a Qubit reading and processed with the Nextera XT library preparation kit (Illumina, San Diego, CA). The library was subsequently sequenced on a MiSeq sequencer located at the Monash University Malaysia Genomics Facility using a run configuration of 2 × 250 bp. Default parameters were used for all software unless otherwise noted. The raw sequencing reads were adapter trimmed and assembled *de novo* using Trimmomatic v0.39 (16) and Unicycler v0.4.7 (17), respectively. A total of 1,543,820 paired-end reads (~387 Mb and 77× genome coverage) were generated and assembled into 68 contigs with a total length of 5,010,327 bp (GC content, 56.96%; N_{50} length, 221,699 bp). The assembled genome was then submitted to NCBI for annotation using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v4.7 (18).

Some *Pantoea* species use quorum sensing via acyl homoserine lactone (AHL) signals to control gene expression based on cell density (19). However, despite har-

Citation Gan HM, Parthasarathy A, Henry KR, Savka MA, Thomas BN, Hudson AO. 2020. Whole-genome sequencing of *Pantoea* sp. strain RIT388, a potential oral opportunistic pathogen isolated from a chewing stick (*Distemonanthus benthamianus*). Microbiol Resour Announc 9:e01468-19. https://doi.org/ 10.1128/MRA.01468-19.

Editor Vincent Bruno, University of Maryland School of Medicine

Copyright © 2020 Gan et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to André O. Hudson, aohsbi@rit.edu.

Received 26 November 2019 Accepted 4 February 2020 Published 27 February 2020

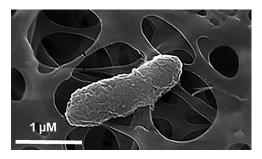


FIG 1 Scanning electron micrograph of *Pantoea* sp. RIT388, showing an elongated rod-shaped cell. Scanning voltage, 20 kV; magnification, ×74,100.

boring a *luxl* homolog (gene locus tag BBB56_18675), RIT388 does not accumulate AHL signals, as determined with a biosensor strain using the TraR receptor (20). This could mean that either a novel signal is produced or that the RIT388 *luxl* homolog is mutated. The latter has been shown in *Vibrio fischeri*, where mutations in *luxl* result in a nonfunctional protein (20).

Data availability. This whole-genome assembly of *Pantoea* sp. RIT388 has been deposited in GenBank under the accession number RMVG00000000 (assembly number GCF_003813865). Raw sequencing reads have been deposited in the SRA database under accession number SRR10522315 (BioProject number PRJNA327264).

ACKNOWLEDGMENTS

A.P., M.A.S., and A.O.H. acknowledge ongoing support from the Gosnell School of Life Sciences (GSoLS) and the College of Science (COS) at the Rochester Institute of Technology (RIT).

REFERENCES

- Walterson AM, Stavrinides J. 2015. Pantoea: insights into a highly versatile and diverse genus within the Enterobacteriaceae. FEMS Microbiol Rev 39:968–984. https://doi.org/10.1093/femsre/fuv027.
- Delétoile A, Decré D, Courant S, Passet V, Audo J, Grimont P, Arlet G, Brisse S. 2009. Phylogeny and identification of *Pantoea* species and typing of *Pantoea agglomerans* strains by multilocus gene sequencing. J Clin Microbiol 47:300–310. https://doi.org/10.1128/JCM.01916-08.
- Nadarasah G, Stavrinides J. 2014. Quantitative evaluation of the hostcolonizing capabilities of the enteric bacterium *Pantoea* using plant and insect hosts. Microbiology 160:602–615. https://doi.org/10.1099/mic.0 .073452-0.
- Roper MC. 2011. Pantoea stewartii subsp. stewartii: lessons learned from a xylem-dwelling pathogen of sweet corn. Mol Plant Pathol 12:628–637. https://doi.org/10.1111/j.1364-3703.2010.00698.x.
- Brady CL, Venter SN, Cleenwerck I, Engelbeen K, Vancanneyt M, Swings J, Coutinho TA. 2009. Pantoea vagans sp. nov., Pantoea eucalypti sp. nov., Pantoea deleyi sp. nov. and Pantoea anthophila sp. nov. Int J Syst Evol Microbiol 59:2339–2345. https://doi.org/10.1099/ijs.0.009241-0.
- Coutinho TA, Venter SN. 2009. Pantoea ananatis: an unconventional plant pathogen. Mol Plant Pathol 10:325–335. https://doi.org/10.1111/j .1364-3703.2009.00542.x.
- Dutkiewicz J, Mackiewicz B, Kinga Lemieszek M, Golec M, Milanowski J. 2016. Pantoea agglomerans: a mysterious bacterium of evil and good. Part III. Deleterious effects: infections of humans, animals and plants. Ann Agric Environ Med 23:197–205. https://doi.org/10.5604/12321966 .1203878.
- Cruz AT, Cazacu AC, Allen CH. 2007. Pantoea agglomerans, a plant pathogen causing human disease. J Clin Microbiol 45:1989–1992. https://doi.org/10 .1128/JCM.00632-07.
- Van Rostenberghe H, Noraida R, Wan Pauzi WI, Habsah H, Zeehaida M, Rosliza AR, Fatimah I, Nik Sharimah NY, Maimunah H. 2006. The clinical picture of neonatal infection with *Pantoea* species. Jpn J Infect Dis 59:120–121.
- 10. Bergman KA, Arends JP, Schölvinck EH. 2007. Pantoea agglomerans

septicemia in three newborn infants. Pediatr Infect Dis J 26:453-454. https://doi.org/10.1097/01.inf.0000261200.83869.92.

- 11. Rezzonico F, Stockwell VO, Tonolla M, Duffy B, Smits TH. 2012. *Pantoea* clinical isolates cannot be accurately assigned to species based on metabolic profiling. Transpl Infect Dis 14:220–221. https://doi.org/10.1111/j.1399-3062.2011.00684.x.
- Rezzonico F, Smits TH, Montesinos E, Frey JE, Duffy B. 2009. Genotypic comparison of *Pantoea agglomerans* plant and clinical strains. BMC Microbiol 9:204. https://doi.org/10.1186/1471-2180-9-204.
- Harvey D, Bardelang P, Goodacre SL, Cockayne A, Thomas NR. 2017. Antibiotic spider silk: site-specific functionalization of recombinant spider silk using "click" chemistry. Adv Mater 29. https://doi.org/10.1002/ adma.201604245.
- Adeniyi BO, John OO, Lawal TO. 2011. *In-vitro* antifungal activity of Distemonanthus benthamianus stem. Int J Pharm Pharm Sci 3:51–55.
- Adeniyi BA, Odumosu BT. 2012. Antibacterial and antifungal properties of *Distemonanthus* benthamianus (Fabaceae) crude extract. Glob J Pharm Res 1:567–574.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi .org/10.1093/nar/gkw569.
- Wen-Si T, Yunos M, Yusrina N, Pui-Wan T, Nur Izzati M, Tan-Guan-Sheng A, Yin W, Chan K. 2014. *Pantoea* sp. isolated from tropical fresh water exhibiting *N*-acyl homoserine lactone production. ScientificWorldJournal 2014:828971. https://doi.org/10.1155/2014/828971.
- Hanzelka BL, Stevens AM, Parsek MR, Crone TJ, Greenberg EP. 1997. Mutational analysis of the Vibrio fischeri Luxl polypeptide: critical regions of an autoinducer synthase. J Bacteriol 179:4882–4887. https://doi.org/ 10.1128/jb.179.15.4882-4887.1997.