

Research Article

Immunomodulatory Role of Adipose-Derived Stem Cells on Equine Endometriosis

Maria Elena Falomo,¹ Letizia Ferroni,² Ilaria Tocco,³ Chiara Gardin,² and Barbara Zavan²

¹Department of Veterinary Clinical Sciences, Viale dell'Università 16, 35100 Padua, Italy

²Department of Biomedical Sciences, University of Padua, Via G. Colombo 3, 35100 Padua, Italy

³Department of Neurosciences, University of Padua, Via Giustiniani 2, 35100 Padua, Italy

Correspondence should be addressed to Barbara Zavan; barbara.zavan@unipd.it

Received 12 January 2015; Accepted 9 March 2015

Academic Editor: Aaron W. James

Copyright © 2015 Maria Elena Falomo et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Endometriosis is a degenerative process due to a chronic inflammatory damage leading to extracellular matrix components deposition and glandular fibrosis. It is known that mesenchymal stem cells secrete a wide range of bioactive molecules, some of them modulating the immune inflammatory response, and others providing regeneration and remodeling of injured tissue. We have performed *in vitro* experiments in order to analyze the capability of allogenic equine adipose-derived stem cells (ADSCs) to infiltrate mares' endometrial tissues and to stimulate the expression of cytokines and metalloproteinases. Differences in the biologic response to the exposure to ADSCs between pathological and healthy endometrial tissue have been identified. These results could challenge researchers to progress forward with future studies for the development of a biological therapy with a possible application in translational medicine.

1. Introduction

Mesenchymal stem cells (MSCs) are nowadays isolated from many different tissues, including bone marrow, umbilical cord blood and tissue, and adipose tissue, which constitute the most promising alternative for research and clinical purposes due to the easy accessibility. MSCs secrete a wide range of bioactive molecules, some of them modulating the immune inflammatory response, others providing regeneration and remodeling of injured tissue, and finally mitogens acting on lymphatic progenitors differentiation [1–10]. Thus, MSCs gained progressively higher attention in consideration of their bioactive features and started to be vastly used in research to test the effects of their multifactor dispensaries on clinical settings.

Endometriosis is a multifactorial degenerative process of uterine glands and surrounding stroma, and it is considered a major cause of equine infertility [11, 12]. Endometrial tissue degeneration is due to a chronic inflammatory damage leading to extracellular matrix (ECM) components deposition and glandular fibrosis. In mares, the disease seems to be age-related, although a correlation with hormonal

changes has not been demonstrated [13–15]. Etiology is still only partially clear, as studies show differences in mares' immune inflammatory response and thus susceptibility of the development of endometriosis [16–18]. Hypothesis has been made that differences in the expression rate of pro- and anti-inflammatory gene products in chronic inflammation are implied [17–19], involving especially ECM degrading-endopeptidases known as metalloproteinases (MMPs) and their inhibitors (TIMPs) [20, 21].

We previously and successfully isolated and expanded *in vitro* human adipose-derived stem cells (ADSCs) and observed their regenerative potential [22–24]. Based on current knowledge about the expression profile of inflammatory proteins, we aimed at analyzing the capacity of allogenic equine MSCs to integrate and influence the expression of these proteins in mares' endometrial tissue, possibly obtaining interesting data for translational purposes.

2. Materials and Methods

2.1. Samples Collection. Endometrial biopsies (40 × 40 × 3 mm) included in this study were collected postmortem

TABLE 1: Equine primer sequences.

Gene symbol	Forward primer (5' → 3')	Reverse primer (5' → 3')	Product length (bp)
ACTB	CTCCCAGCACGATGAAGA	GTA CTCTCTGCTTGCTGATCC	125
IL10	GGCACCCAGTCTGAGAACA	GCTGGTCCTTCATTTGAAAAGAAAGTC	115
IL1RN	TGTGTCAAGTCTGGTGATGAGA	TGTTTGAGCGGATGAAGGTGA	108
IL1B	CAACGGAGAGAATACAAACCAACAAG	GGCTTCCCATCTTTCATCCCA	141
IL6	AGAACACAACAACCTCACCTCATCC	AGAGGAAGGAATGCCCATGAAC	125
IL8	TCCAAGAATTCCTCAGTAAAGATGCC	CCTAGATACTGCGTGGGACAATG	173
MMP14	CAAGATGCCTCCTCAACCCA	GCCACGGTGTCAAAGTTCC	107
MMP2	TGGTCCGTGTGAAGTATGGC	TCGAAGTTGTAGGTGGTGA	139
MMP9	ACAGTGCCTTTGGGTCCAG	GTACCTCCCGTCCTTGAAGA	102
TIMP2	GCCAAAGCGGTCAAGTGAAGA	GTAGATGAACCTCGATGTCCTTGTCA	132
TNFA	ACGGTGCTTGTGCCTCA	CGGTAACCTGCTCTCCCTCTG	112

ACTB: actin beta; IL10: interleukin 10; IL1RN: interleukin 1 receptor antagonist; IL1B: interleukin 1 beta; IL6: interleukin 6; IL8: interleukin 8; MMP14: matrix metalloproteinase 14; MMP2: matrix metalloproteinase 2; MMP9: matrix metalloproteinase 9; TIMP2: TIMP metalloproteinase inhibitor 2; TNFA: tumor necrosis factor alpha.

from 12 mares: 6 younger than 6 years old and 6 older than 15 years old. Animals were classified according to Kenney's histological classes between healthy (classes I and IIA; $n = 6$) and pathological (class IIB and greater; $n = 6$) mares [25].

2.2. Equine ADSCs Isolation. Equine ADSCs were isolated from the fat of mares aging less than 5 years. The region above the dorsal gluteal muscle, at the base of the tail, was chosen as the adipose tissue collection site because of the availability of material, the absence of large veins, and ease of access. Adipose tissue samples (approximately 100 gr each) were preserved in a sterile phosphate buffered saline (PBS) (Lonza, Walkersville, MD, USA) solution enriched with 1% penicillin/streptomycin (P/S) (EuroClone, Milan, Italy) and 1% amphotericin B (EuroClone) until processing. The samples were digested using a solution of 0.075% collagenase from *Clostridium histolyticum* type II (Sigma-Aldrich, St. Louis, MO, USA) in Hank's Balanced Salt Solution (HBSS) (Lonza), for 3 h at room temperature and in slow agitation. At the end of the digestion, the collagenase activity was blocked with an equal volume of complete Dulbecco's modified Eagle's medium (DMEM) (completed: cDMEM). cDMEM consisted of Dulbecco's modified Eagle's medium (DMEM, Lonza) supplemented with 10% fetal bovine serum (FBS, Bidachem S.p.A., Milano, Italy) and 1% P/S. After centrifugation for 7 min at 500 g, the pellet was washed in PBS and filtered with a 70 μ M cell strainer (BD Biosciences, Mississauga, Ontario, Canada). The cell suspension was resuspended in cDMEM, transferred to a 25 cm² tissue culture flask, and then incubated at 37°C and 5% CO₂ for 15 days. Culture medium was changed every 2 days.

2.3. Equine ADSCs Labeling with Quantum Dots. Equine ADSCs at p3 were labeled with Qtracker 705 Cell Labeling Kit (Life Technologies, Carlsbad, CA, USA) following the manufacturer's protocol. Quantum dots are nanocrystals made of semiconducting materials that, once inside the cells, provide stable fluorescence that can be traced through several

generations and are not transferred to adjacent cells in a population [26].

2.4. Equine ADSCs Seeding on Endometrial Tissues. Endometrial biopsies from healthy and pathological mares were cut into 10 × 10 × 3 mm pieces. 5 × 10⁵ equine ADSCs were then seeded onto each piece in cDMEM and incubated at 37°C and 5% CO₂ for 3 days.

2.5. Histological Analyses. Endometrial biopsies were frozen in isopentane and liquid nitrogen. Samples were cut with cryostat (Leica CM1950, Leica Biosystems, Nußloch, Germany) in slivers of 7–10 μ m.

Healthy and pathological endometrial tissues before equine ADSCs seeding were stained with hematoxylin and eosin. On the contrary, endometrial samples treated with labeled equine ADSCs were stained with Hoechst 33342. All reagents were purchased from Sigma-Aldrich (Saint Louis, MO, USA).

2.6. Real-Time PCR. Total RNA was extracted using the TRIzol Reagent (Life Technologies) from healthy and pathological endometrial tissues before and after ADSCs seeding. The samples were quantified using the NanoDrop spectrophotometer (NanoDrop 1000, Thermo Scientific). For the first-strand cDNA synthesis, 1000 ng of total RNA was reverse transcribed using M-MLV RT (Moloney Murine Leukemia Virus Reverse Transcriptase, Life Technologies) according to the manufacturer's protocol. Equine primers were selected for each target gene with Primer 3 software (Table 1). Real-time PCRs were carried out using the designed primers at a concentration of 300 nM and FastStart SYBR Green Master (Roche Diagnostics, Mannheim, Germany) on a Rotor-Gene 3000 (Corbett Research, Sydney, Australia). Thermal cycling conditions were as follows: 15 min denaturation at 95°C, followed by 40 cycles of 15 s denaturation at 95°C, annealing for 30 s at 60°C, and 20 s elongation at 72°C. Values were normalized to the expression of the β -actin internal reference, whose abundance did not change under our experimental

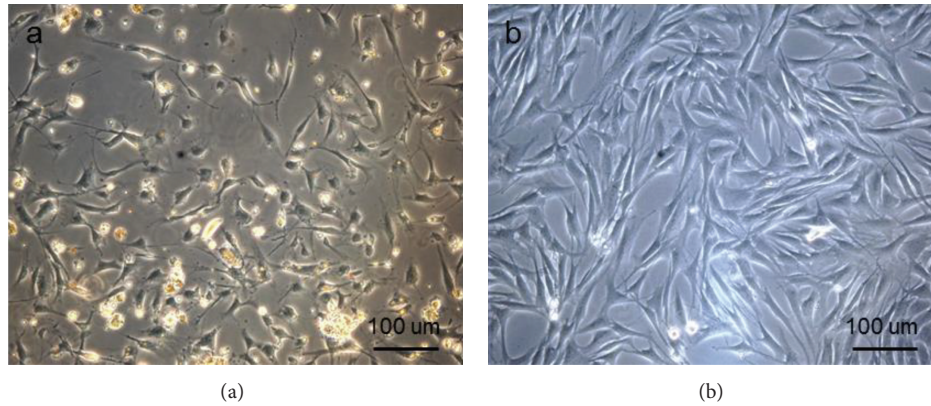


FIGURE 1: Equine ADSCs at phase-contrast microscope. (a) Equine ADSCs at p0. (b) Equine ADSCs at p3.

conditions. Experiments were performed with 3 different cell preparations and repeated at least 3 times.

2.7. Statistical Analysis. Data are presented as mean \pm SD and MedCalc 9.0 software was used. The effect of ADSCs therapy on the total population was evaluated using paired matched Wilcoxon's rank test. A P value < 0.05 for the analysis was considered significant.

3. Results and Discussion

3.1. Equine ADSCs Isolation and Expansion. Adipose-derived stem cells (ADSCs) were isolated after enzymatic digestion from equine abdominal fat and plated on a 25 cm² tissue culture flask. After 3 days, the nonadherent cells were removed by replacing the medium with fresh complete medium. The adherent cells are defined as passage 0 (p0) cells. At this stage, the cell morphology varies between cuboidal shape and spindle shape (Figure 1(a)). After reaching 80% confluence, ADSCs were trypsinized and expanded until p3. Figure 1(b) shows that the cells acquire a spindle-shaped fibroblast-like morphology.

3.2. Equine ADSCs Labeling and Seeding on Endometrial Tissues. In order to verify if equine ADSCs could be able to infiltrate the endometrial tissue, we treated *in vitro* endometrial biopsies with ADSCs previously labeled with quantum dots. Equine ADSCs labeled at p3 show a well distribution of quantum dots nanocrystals in vesicles in the cytoplasm (Figure 2), thus suggesting the efficacy of the method for tracking the ADSCs seeded on the endometrial tissues.

The presence of fluorescent labeled ADSCs cultivated on endometrial tissue from both healthy and pathological mares has been observed after 3 days from seeding. As reported in Figures 3(a) and 3(b), the equine ADSCs were detected in both of the tissues, although in major quantity in the pathological samples. Furthermore, in the endometriosis-affected tissues, the labeled ADSCs were infiltrated in the periglandular space as well as in single glands. Hematoxylin and eosin staining of the endometrial tissues before ADSCs seeding confirms the different morphological structure between the

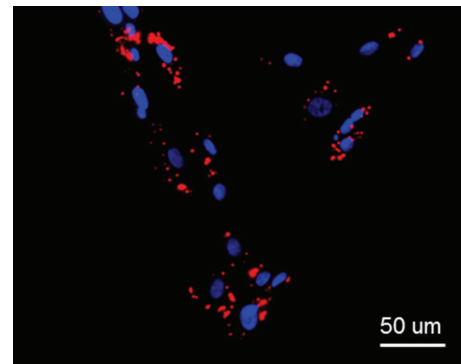


FIGURE 2: Equine ADSCs at fluorescence microscope. Equine ADSCs at p3 stained with quantum dots (red) and Hoechst 33342 (blue).

healthy and the pathological tissues (Figures 3(c) and 3(d)), along with an evident inflammatory condition in the latter.

3.3. Gene Expression of Inflammatory Factors before and after ADSCs Seeding. Endometriosis in mares is a degenerative process characterized by epithelial and stromal alterations and fibrosis. Although the disease is well-known and causes severe consequences such as early embryo death, contradictory hypotheses have been made around its etiology, the more supported being a malfunctioning of the immune system and a dysregulation of the inflammatory processes [16].

In order to test this hypothesis, the expression of different cytokines and MMPs has been evaluated in pathological in comparison to healthy endometrial tissues (Figure 4). Basal expression of interleukin 1 beta (IL1B), interleukin 6 (IL6), interleukin 8 (IL8), interleukin 10 (IL10), tumor necrosis factor alpha (TNFA), and MMP9 was strong in pathologic samples compared to healthy endometrial tissue and particularly significant for IL10 ($P = 0.02$) and TNFA ($P = 0.003$). These results confirmed the observations made by Fumuso et al. [27, 28] and Feghali and Wright [29] regarding a higher expression of IL1B, IL8, and TNFA in endometriosis. Conversely, the higher expression of IL6 and

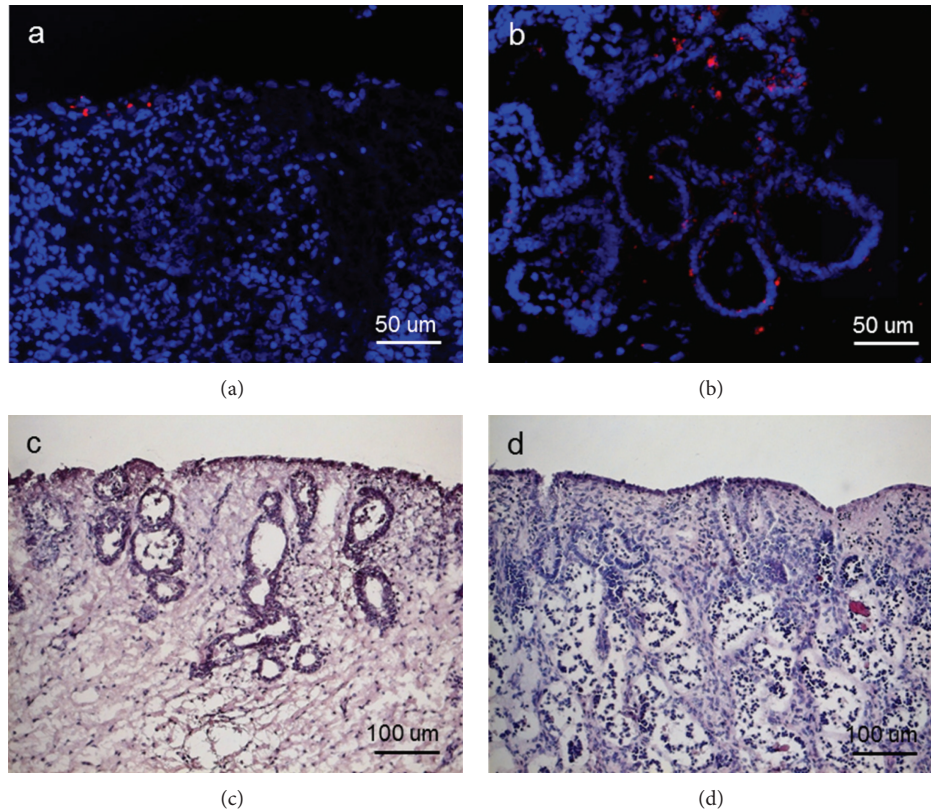


FIGURE 3: Equine endometrial biopsy from (a, c) healthy and (b, d) pathological mares incubated 3 days with equine ADSCs. (a, b) Fluorescent imaging of equine ADSCs labeled with quantum dots (red); nuclei stained with Hoechst 33342 (blue). (c, d) Hematoxylin and eosin staining of ADSCs-seeded healthy and pathological endometrial tissues.

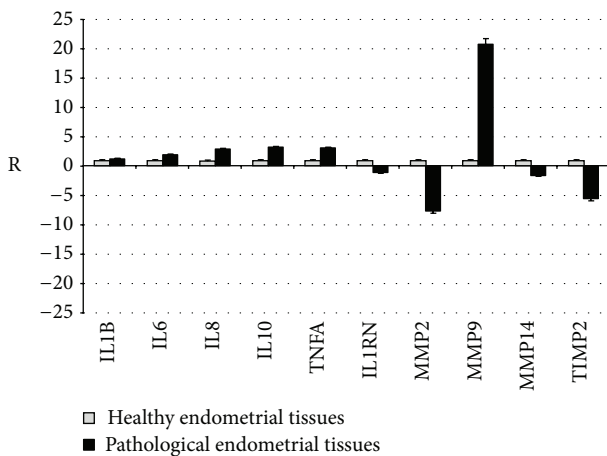


FIGURE 4: Gene expression levels (R) of cytokines and MMPs in pathological endometrial tissues (black) compared to healthy endometrial tissues (grey) before ADSCs seeding.

IL10 in pathologic samples contrasted with previous observations, where a low level of IL6 was thought to be associated with an increased risk of the development of endometriosis and high levels of IL10 were auspicated in consideration of its anti-inflammatory properties [27, 28]. Interleukin 1 receptor

antagonist (IL1RN) expression levels were lower in pathologic endometrium compared to healthy tissue, confirming its role as a protective factor [17, 30]. MMPs expression levels gave contradictory results, as MMP2 and MMP14 were detected at lower levels in pathologic endometrium, whereas MMP9 was much higher when compared to healthy tissue values. TIMP2 expression followed the MMP2 and MMP14 trend. These discrepancies suggest that a profound dysregulation is active in endometriosis, leading to ECM accumulation and glandular fibrosis [12].

In order to evaluate the effects of ADSCs seeding on endometrial tissue of healthy and pathological mares, the expression pattern of the same panel of inflammatory secretory factors has been analyzed (Figures 5(a) and 5(b)). The profile obtained from equine endometrial tissues after the treatment with allogenic ADSCs showed a general reduction for expression levels of IL1B, IL10, TNFA, and IL1RN and an increase of expression levels of IL6 and IL8 in both groups. These results suggest both positive and negative effects of equine ADSCs on the inflammatory processes regulation in the endometrium, as a reduction in IL1B and TNFA is auspicious to downregulate chronic inflammatory processes [29], whereas high levels of IL10 and IL1RN are desirable in consideration of their anti-inflammatory properties [17, 28]. A recent study showed contrasting effects for IL6 in the inflammatory process, being related to the recruitment of

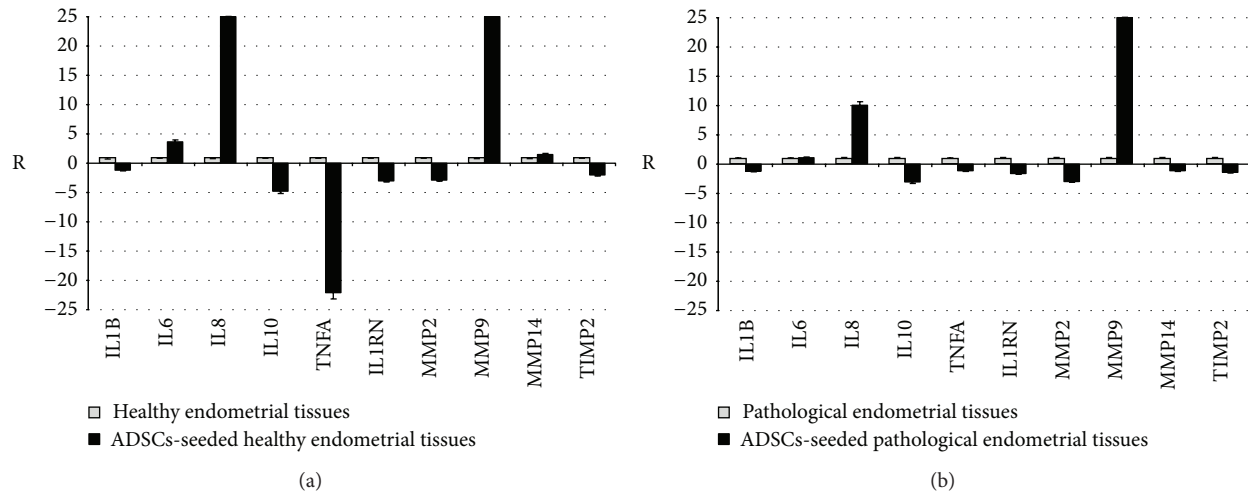


FIGURE 5: Gene expression profile of cytokines and MMPs after seeding of equine ADSCs in (a) healthy endometrial tissues and (b) pathological endometrial tissues. The results are expressed as ratio (R) with respect to mRNA expression of endometrial tissue without ADSCs.

neutrophils in the early inflammatory phase while inducing neutrophils apoptosis and monocyte phagocytic properties in the late stages [31]. Another cytokine which showed an increased expression rate in both groups after ADSCs seeding was IL8. In consideration of its proinflammatory activity, the effect could be potentially negative in the treatment of endometriosis.

Regarding MMPs and their inhibitors, the expression of MMP2 and TIMP2 decreased, whereas expression levels of MMP9 increased in both pathological and healthy endometrial tissues after ADSCs seeding. As far as MMPs are concerned, the reduced expression of MMP2 after ADSCs culture should have a positive effect in treating endometriosis [12, 19, 21]. The positive effect comprises also the increased expression of MMP9 in both groups. In fact, previous observations made by Aresu et al. [12] detected a standard expression of MMP9 in endometriosis, hypothesizing a positive role coming from an increased activity of tissue remodeling [21]. However we must notice that our pathologic samples already showed a higher expression of MMP9 compared to healthy ones. MMP14 expression levels increased only in healthy tissues. MMP14 can degrade different matrix components and activate other MMPs, in particular MMP2 and MMP9 [12]. Furthermore, TIMPs can influence all MMPs-related processes. Little is published regarding TIMP2 expression in mares' endometriosis [12], but studies on human hepatic fibrosis proved that the recovery phase is characterized by a decrease of TIMP2 expression levels [21].

4. Conclusions

All things considered, in the present work we could not detect a clear potential clinical effect of ADSCs on equine endometriosis. The balance between pro- and anti-inflammatory, lytic, and fibrotic environment is very subtle, and many actors are involved in the scene, providing a complex pattern of interactions to consider when trying to

influence biologic processes. Nevertheless, we highlighted important differences in the biologic response of endometrial tissue to the exposure to ADSCs, challenging researchers to progress forward with future studies hopefully for the development of a biological therapy in clinical trials.

Conflict of Interests

The authors declare no conflict of interests.

Authors' Contribution

Letizia Ferroni and Chiara Gardin carried out the biological and molecular studies; Maria Elena Falomo partook in samples collection and data analyses; Barbara Zavan and Maria Elena Falomo conceived the study and participated in its design and coordination. Iliaria Tocco helped to draft the paper. All authors read and approved the final paper. Maria Elena Falomo and Letizia Ferroni contributed equally to this work.

Acknowledgment

This research was supported by funds from the University of Padova, Progetto di Ateneo, to Barbara Zavan.

References

- [1] D. D. Carrade, M. W. Lame, M. S. Kent, K. S. Clark, N. J. Walker, and D. L. Borjesson, "Comparative analysis of the immunomodulatory properties of equine adult-derived mesenchymal stem cells," *Cell Medicine*, vol. 4, no. 1, pp. 1-11, 2012.
- [2] M. D. Nicola, C. Carlo-Stella, M. Magni et al., "Human bone marrow stromal cells suppress T-lymphocyte proliferation induced by cellular or nonspecific mitogenic stimuli," *Blood*, vol. 99, no. 10, pp. 3838-3843, 2002.
- [3] M. Krampera, S. Glennie, J. Dyson et al., "Bone marrow mesenchymal stem cells inhibit the response of naive and memory

- antigen-specific T cells to their cognate peptide," *Blood*, vol. 101, no. 9, pp. 3722–3729, 2003.
- [4] P. Comoli, F. Ginevri, R. MacCario et al., "Human mesenchymal stem cells inhibit antibody production induced in vitro by allostimulation," *Nephrology Dialysis Transplantation*, vol. 23, no. 4, pp. 1196–1202, 2008.
 - [5] A. J. Nauta, A. B. Kruisselbrink, E. Lurvink, R. Willemze, and W. E. Fibbe, "Mesenchymal stem cells inhibit generation and function of both CD34⁺-derived and monocyte-derived dendritic cells," *Journal of Immunology*, vol. 177, no. 4, pp. 2080–2087, 2006.
 - [6] E. Gonzalez-Rey, P. Anderson, M. A. González, L. Rico, D. Büscher, and M. Delgado, "Human adult stem cells derived from adipose tissue protect against experimental colitis and sepsis," *Gut*, vol. 58, no. 7, pp. 929–939, 2009.
 - [7] B. Parekkadan, A. W. Tilles, and M. L. Yarmush, "Bone marrow-derived mesenchymal stem cells ameliorate autoimmune enteropathy independently of regulatory T cells," *Stem Cells*, vol. 26, no. 7, pp. 1913–1919, 2008.
 - [8] F. Benvenuto, S. Ferrari, E. Gerdoni et al., "Human mesenchymal stem cells promote survival of T cells in a quiescent state," *Stem Cells*, vol. 25, no. 7, pp. 1753–1760, 2007.
 - [9] M. J. Crop, C. C. Baan, S. S. Korevaar, J. N. M. Ijzermans, W. Weimar, and M. J. Hoogduijn, "Human adipose tissue-derived mesenchymal stem cells induce explosive T-Cell proliferation," *Stem Cells and Development*, vol. 19, no. 12, pp. 1843–1853, 2010.
 - [10] L. I. Mambelli, G. H. Z. Winter, A. Kerkis, E. Malschitzky, R. C. Mattos, and I. Kerkis, "A novel strategy of mesenchymal stem cells delivery in the uterus of mares with endometriosis," *Theriogenology*, vol. 79, no. 5, pp. 744–750, 2013.
 - [11] C. Hoffmann, C. Ellenberger, R. C. Mattos et al., "The equine endometriosis: new insights into the pathogenesis," *Animal Reproduction Science*, vol. 111, no. 2–4, pp. 261–278, 2009.
 - [12] L. Aresu, S. Benali, D. Giannuzzi, R. Mantovani, M. Castagnaro, and M. E. Falomo, "The role of inflammation and matrix metalloproteinases in equine endometriosis," *Journal of Veterinary Science*, vol. 13, no. 2, pp. 171–177, 2012.
 - [13] S. M. Fiala, C. A. Pimentel, R. M. Gregory, and R. C. Mattos, "Does equine sperm concentration influence the sperm migration to the oviducts?" *Animal Reproduction Science*, vol. 89, no. 1–4, pp. 261–264, 2005.
 - [14] S. M. Fiala, C. A. Pimentel, A. L. G. Mattos, R. M. Gregory, and R. C. Mattos, "Effect of sperm numbers and concentration on sperm transport and uterine inflammatory response in the mare," *Theriogenology*, vol. 67, no. 3, pp. 556–562, 2007.
 - [15] M. E. Cadario, W. W. Thatcher, E. Klapstein et al., "Dynamics of prostaglandin secretion, intrauterine fluid and uterine clearance in reproductively normal mares and mares with delayed uterine clearance," *Theriogenology*, vol. 52, no. 7, pp. 1181–1192, 1999.
 - [16] M. H. T. Troedsson, "Uterine clearance and resistance to persistent endometritis in the mare," *Theriogenology*, vol. 52, no. 3, pp. 461–471, 1999.
 - [17] E. M. Woodward, M. Christoffersen, J. Campos et al., "Endometrial inflammatory markers of the early immune response in mares susceptible or resistant to persistent breeding-induced endometritis," *Reproduction*, vol. 145, no. 3, pp. 289–296, 2013.
 - [18] H. Zerbe, F. Engelke, E. Klug, H.-A. Schoon, and W. Leibold, "Degenerative endometrial changes do not change the functional capacity of immigrating uterine neutrophils in mares," *Reproduction in Domestic Animals*, vol. 39, no. 2, pp. 94–98, 2004.
 - [19] I. Walter, J. Handler, I. Miller, and C. Aurich, "Matrix metalloproteinase 2 (MMP-2) and tissue transglutaminase (TG 2) are expressed in periglandular fibrosis in horse mares with endometriosis," *Histology and Histopathology*, vol. 20, no. 4, pp. 1105–1113, 2005.
 - [20] W. H. Rodgers, L. M. Matrisian, L. C. Giudice et al., "Patterns of matrix metalloproteinase expression in cycling endometrium imply differential functions and regulation by steroid hormones," *The Journal of Clinical Investigation*, vol. 94, no. 3, pp. 946–953, 1994.
 - [21] A. L. Clutterbuck, P. Harris, D. Allaway, and A. Mobasheri, "Matrix metalloproteinases in inflammatory pathologies of the horse," *Veterinary Journal*, vol. 183, no. 1, pp. 27–38, 2010.
 - [22] B. Zavan, V. Vindigni, C. Gardin et al., "Neural potential of adipose stem cells," *Discovery Medicine*, vol. 10, no. 50, pp. 37–43, 2010.
 - [23] B. Zavan, L. Michelotto, L. Lancerotto et al., "Neural potential of a stem cell population in the adipose and cutaneous tissues," *Neurological Research*, vol. 32, no. 1, pp. 47–54, 2010.
 - [24] V. Vindigni, L. Michelotto, L. Lancerotto et al., "Isolation method for a stem cell population with neural potential from skin and adipose tissue," *Neurological Research*, 2009.
 - [25] R. M. Kenney and P. A. Doig, "Equine endometrial biopsy," *Current Therapy in Theriogenology*, vol. 2, pp. 726–729, 1986.
 - [26] A. P. Alivisatos, W. Gu, and C. Larabell, "Quantum dots as cellular probes," *Annual Review of Biomedical Engineering*, vol. 7, pp. 55–76, 2005.
 - [27] E. Fumuso, S. Giguère, J. Wade, D. Rogan, I. Videla-Dorna, and R. A. Bowden, "Endometrial IL-1 β , IL-6 and TNF- α , mRNA expression in mares resistant or susceptible to post-breeding endometritis: effects of estrous cycle, artificial insemination and immunomodulation," *Veterinary Immunology and Immunopathology*, vol. 96, no. 1-2, pp. 31–41, 2003.
 - [28] E. A. Fumuso, J. Aguilar, S. Giguère, M. Rivulgo, J. Wade, and D. Rogan, "Immune parameters in mares resistant and susceptible to persistent post-breeding endometritis: effects of immunomodulation," *Veterinary Immunology and Immunopathology*, vol. 118, no. 1-2, pp. 30–39, 2007.
 - [29] C. A. Feghali and T. M. Wright, "Cytokines in acute and chronic inflammation," *Frontiers in Bioscience*, vol. 2, pp. 12–26, 1997.
 - [30] N. Gérard, M. Caillaud, A. Martoriati, G. Goudet, and A.-C. Lalmanach, "The interleukin-1 system and female reproduction," *The Journal of Endocrinology*, vol. 180, no. 2, pp. 203–212, 2004.
 - [31] G. Kaplanski, V. Marin, F. Montero-Julian, A. Mantovani, and C. Farnarier, "IL-6: a regulator of the transition from neutrophil to monocyte recruitment during inflammation," *Trends in Immunology*, vol. 24, no. 1, pp. 25–29, 2003.