

Effects of Nitric Oxide (NO) Synthesis Inhibition on Antitumor Responses during Interleukin-2 (IL-2) Treatment of Mice

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Objectives: To evaluate if L-arginine: NO pathway is activated in tumor tissues during IL-2 therapy and to evaluate whether IL-2 induced NO synthesis represents an antitumor effector mechanism or an inhibitory factor against therapeutic effects of IL-2.

Methods: Four groups [untreated control, N^G-monomethyl-L-arginine (MLA) therapy only, IL-2 therapy only, IL-2/MLA therapy groups] of BALB/c mice were injected intraperitoneally with 2×10^6 Meth A tumor cells on day 0. MLA was administered subcutaneously with Alzet continuous infusion pumps on day 2. IL-2 therapy (180,000 IU s.c. every 12h for 5 days) was started on day 3. NO production within ascites tumors was assessed by measuring nitrite concentrations in cultures of ascites cells harvested on day 8. Survival and the rate of body weight increment of the mice were measured to evaluate therapeutic responses. Daily urinary nitrate excretion was monitored to demonstrate the effectiveness of MLA in inhibiting NO synthesis.

Results: Nitrite production in supernatants of Meth A ascites cell cultures was $63 \pm 14 \mu\text{M}$ in IL-2 treated mice and $3.2 \pm 1.5 \mu\text{M}$ in untreated controls ($p < 0.001$). MLA prevented the IL-2 therapy induced increase in nitrite production. IL-2 therapy did not decrease the rate of body weight increment and marginally prolonged mean survival to 18.2 days, compared to 16.6 days in control mice ($p = 0.255$). MLA administration decreased the rate of body weight increment and prolonged mean survival of IL-2 treated mice (21.8 days, $p = 0.001$ versus IL-2 alone). Interestingly, the MLA treatment increased the rate of body weight increment and diminished the survival of control mice to 11.6 days ($p = 0.003$). MLA administration via Alzet continuous infusion pumps achieved approximately 60% suppression of urinary nitrate excretion by control mice. Subcutaneous IL-2 treatment strongly induced nitric oxide synthesis (up to $3.5 \mu\text{moles}$ of urinary nitrate/mouse/day). MLA also effectively suppressed IL-2 induced NO production.

Conclusion: L-arginine: NO pathway can be activated in malignant ascites by IL-2 therapy and NO synthesis functions as an inhibitory mechanism against IL-2 induced anti-tumor effects.

Key Words: Nitric Oxide, Interleukin-2, Arginine, MLA, Tumor

INTRODUCTION

One of the major paradoxes in understanding the clinical effects of IL-2 remains the dichotomy between almost universal susceptibility of tumor cells to IL-2 activated lymphocytes (termed lymphokine activated killer or LAK cells) in vitro and the

low response rates observed in clinical trials¹⁻⁹. In the two most susceptible cancers, renal cell carcinoma and malignant melanoma, only 10~25% response rates and 5~10% complete response rates have been reported⁷⁻⁹. These findings have raised the possibility that there may be inhibitory factors against LAK cell activation in vivo, which therefore diminish anti-tumor effects of IL-2 therapy.

NO is a well known cytotoxic effector molecule which may contribute to the development of cell mediated immune responses in a number of ways including tumor cell killing^{10,11}. On the other hand,

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This study was supported by the Chong-Ram Grant in 1993.

NO is potentially immunosuppressive, resulting in decreased lymphocyte proliferation and cytotoxic activity¹²⁻¹⁵. Thus, both anti-tumor and tumor promoting activities of NO appear possible *in vivo*.

IL-2 therapy is known to induce synthesis of proinflammatory cytokines such as IFN γ , TNF and IL-1 by LAK cells^{16,17}. The same mediators are known to induce NO synthase expression in macrophages¹⁸. Hibbs and co-workers demonstrated that IL-2 treated patients developed marked (6-10 fold) increases in NO synthesis, peaking on days 5-7 following a 5 day course of high-dose intravenous bolus IL-2 treatment¹⁹. These observations have raised a question whether high output NO produced during IL-2 therapy acts as an anti-tumor effector mechanism or an inhibitory mechanism against the anti-tumor effect of IL-2 therapy by suppressing LAK cell activities.

We, therefore, developed a murine ascites tumor model to evaluate the role of NO synthesis during IL-2 treatment, and demonstrated that L-arginine: NO synthesis pathway is activated in the local tumor tissue itself as well as systemically, and that NO synthesized during IL-2 treatment may be an inhibitory factor against anti-tumor effects of IL-2 therapy.

MATERIALS AND METHODS

1. Animals

Specific pathogen free BALB/c mice (ages 6-8 weeks) were obtained from Harlan-Sprague Dawley (Indianapolis, IN) and housed at the Chonbuk National University Hospital Animal Care Facility. Mice were maintained under guidelines established by the Chonbuk National University Hospital Animal Care Committee, which also approved experimental protocols. Mice were age and sex matched at the onset of each experiment. All experiments were performed at least twice with highly concordant results.

2. Tumor Cell Lines

Meth A tumor (a gift from Dr. Lloyd Old, Memorial Sloan Kettering Cancer Center, NY) is a methylcholanthrene-induced spindle cell skin cancer arising in a BALB/c mouse²⁰. Tumor cells were maintained by serial intraperitoneal passage in syngeneic BALB/c mice or by culture RPMI 1640 supplemented with 5% fetal calf serum (Hyclone

Laboratories, Inc., Logan, UT), 100 units/ml penicillin G (Sigma Chemical Co., St. Louis, MO), 50 μ g/ml streptomycin (Sigma) and 2 mM glutamine (Sigma) (working medium). The ascites variant of this tumor is extremely aggressive and is rapidly lethal to mice in 12-18 days following injection of $2-3 \times 10^6$ cells. Less virulent strains of this tumor exist, that have been attenuated by long-term passage in cell cultures.

Mycoplasmal infection was excluded by periodic surveillance cultures of supernatants derived from the tumor cell line grown in antibiotic free media on pleuropneumonia-like organism agar (PPL0 agar, Baxter Health Care Corporation, McGaw Park, IL) supplemented with 10% horse serum (Hyclone), 25% yeast extract (Gibco BRL, Gaithersburg, MD) and 10 units penicillin G/ml.

3. Treatment of Meth A Ascites-Bearing Mice with Subcutaneous IL-2

At the beginning (day 0) of each experiment, normal BALB/c mice were injected intraperitoneally with 2×10^6 Meth A tumor cells. Beginning on day 3, groups of 5 mice were treated with a 5 day course of IL-2 (180,000 IU subcutaneously every 12h for 5 days). The recombinant human IL-2 was a generous gift from Chiron Corporation, Emeryville, CA. Some mice were implanted with osmotic minipumps (Model 2001, Alza Corporation, Palo Alto, CA) containing 0.2 ml of 3.38 M N^G-monomethyl-L-arginine (MLA; a NOS inhibitor)²¹. In previous studies, we have demonstrated that these infusions block both baseline and cytokine-induced NO synthesis for 8-9 days²¹.

4. Analysis of NO Synthesis by Ascites Cells

Ascites cells were cultured in working medium (7.5×10^5 cells/ml) in 96 well flat bottom plates (at a final volume of 0.2 ml/well). After a 48h incubation at 37°C in humidified 95% air/5% CO₂ atmosphere, nitrite, a stable metabolite of NO, was quantified in culture supernatants using a previously described colorimetric microtiter assay²¹. Briefly, 50 μ l samples of cell culture supernatant from each well were dispensed into flat bottomed microtiter plates, and incubated with 100 μ l of a 1:1 mixture of 1% sulfanilamide in 30% acetic acid with 0.1% N-(1-naphthyl) ethylenediamine dihydrochloride in 60% acetic acid. Once mixed, this reagent was used within 5 min. Following a 5 min incubation with

experimental samples or standards, absorbance at 570 nm was evaluated in a microtiter plate reader. Concentrations were determined from a linear standard curve generated from 6.25-100 μM sodium nitrite in working medium. Results are presented as mean \pm SD. Working medium contained $<0.5 \mu\text{M}$ nitrite.

6. Effect of Nitric Oxide Synthase Inhibition on IL-2 Induced Anti-Tumor Responses

Four groups (untreated control, MLA therapy only, IL-2 therapy only, IL-2/MLA therapy groups) of 5 normal BALB/c mice were acclimated to a nitrate/nitrite free diet (Ziegler Brothers, Gardeners, PA) in metabolic cages (Nalgene, Rochester, NY). Following a 7 day acclimation (on day 0), mice were injected intraperitoneally with 2×10^6 Meth A tumor cells. On day 2, two groups (MLA therapy only and IL-2/MLA therapy groups) of mice were implanted with subcutaneous Alzet continuous infusion pumps (Model 2001) containing 0.2 ml of 3.38 M MLA. The other groups of mice underwent subcutaneous tunnel formation but no pump implantation. On day 3, IL-2 therapy (180,000 IU s.c. every 12h for 5 days) was begun in IL-2 therapy only and IL-2/MLA therapy groups. Daily body weight measurement and survival of the mice were evaluated to assess therapeutic responses. The concentration of nitrate in sequential 24 hour urine samples was measured using a previously described method²¹. Briefly, 50 μl of urine samples were dispensed into triplicate wells of round bottom microtiter plates and incubated for 90 min with 50 μl of a frozen suspension of *Escherichia coli*, induced for the enzyme nitrate reductase²². Following this incubation, plates were centrifuged at 1,000 xg for 10 min and 50 μl of supernatant were transferred to clean flat bottom microtiter plates and analyzed for nitrite as described previously.

7. Statistical Analysis

Statistical analysis was performed using Student's ttest and differences were considered to be significant at $p < 0.05$.

RESULTS

1. Subcutaneous IL-2 Administration Induces NO Synthesis by Cells in Meth A Ascites

Cellular ascites from Meth A bearing mice were

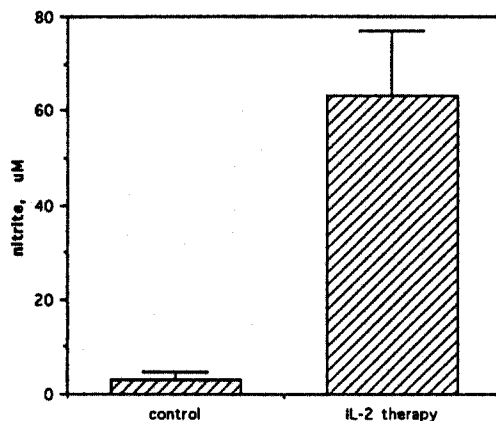


Fig. 1. Induction of high output NO synthesis in Meth A ascites tumor cells by IL-2 therapy. Meth A ascites cells harvested from three IL-2 treated mice were cultured at 1.5×10^5 cells/well in microtiter plates in triplicate. After a 48h culture, nitrite was measured in 50 μl culture supernatants by a colorimetric assay. Results are mean \pm SD. Meth A ascites cells harvested from three untreated mice served as controls.

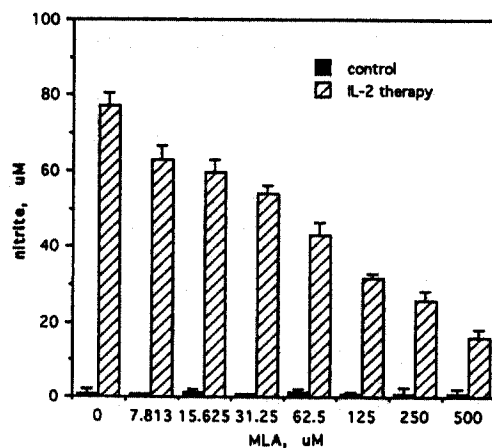


Fig. 2. Effect of MLA on NO synthesis of Meth A ascites cells from IL-2 treated mice. Meth A ascites cells harvested from IL-2 treated mice were cultured at 1.5×10^5 cells/well in microtiter plates in triplicate with varying concentrations (0-500 μM) of MLA. After a 48h culture, nitrite was measured in 50 μl culture supernatants by a colorimetric assay. Results are mean \pm SD. Ascites tumor cells harvested from untreated mice served as controls.

harvested under sterile conditions on day 8 following the last dose of IL-2. Following erythrocyte lysis, cells were cultured at 1.5×10^5 cells/well in 0.2 ml working medium for 48h and analyzed for nitrite synthesis. Ascites cells derived from untreated mice served as controls. Significant nitrite accumulation ($63 \pm 14 \mu\text{M}$) was observed in supernatants of Meth A ascites cells from IL-2 treated mice (Fig. 1). In contrast, ascites cells from control mice synthesized little nitrite ($3.2 \pm 1.5 \mu\text{M}$). This result demonstrated that nitrite synthesis was strongly induced in malignant ascites following subcutaneous IL-2 administration in mice. Further experiments were performed to confirm the origin of the nitrite from nitric oxide synthase.

2. Effect of MLA on NO Synthesis of Meth A Ascites Cells from IL-2 Treated Mice

The nitric oxide synthase inhibitor MLA was used

to establish that the source of nitrite in cell cultures was L-arginine: NO pathway. Ascites cells from IL-2 treated mice were cultured with increasing concentrations (0-500 μM) of MLA. Nitrite production in these cultures was found to be inhibited in a dose dependent manner (Fig. 2). In contrast, MLA had no effect on the trace amounts of nitrite produced by ascites cells from non-IL-2 treated control mice ($< 0.5 \mu\text{M}$). Control experiments performed in the presence of 10 $\mu\text{g/ml}$ polymyxin B gave similar results, excluding inadvertent endotoxin contamination.

Furthermore, bacterial lipopolysaccharide is undetectable in our working medium by Limulus lysate assay (sensitivity, 100 pg/ml). This result demonstrated that the nitrite in ascites cell cultures from IL-2 treated mice was a product of the cytokine induced L-arginine: NO pathway.

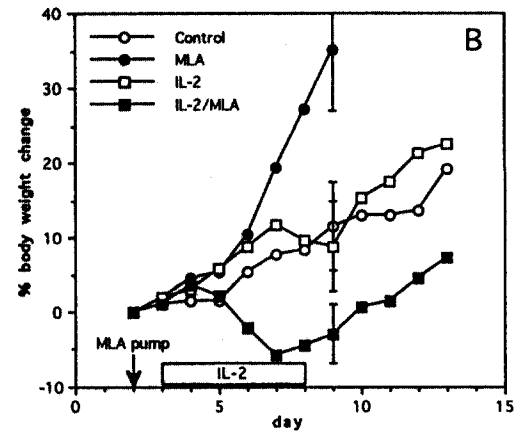
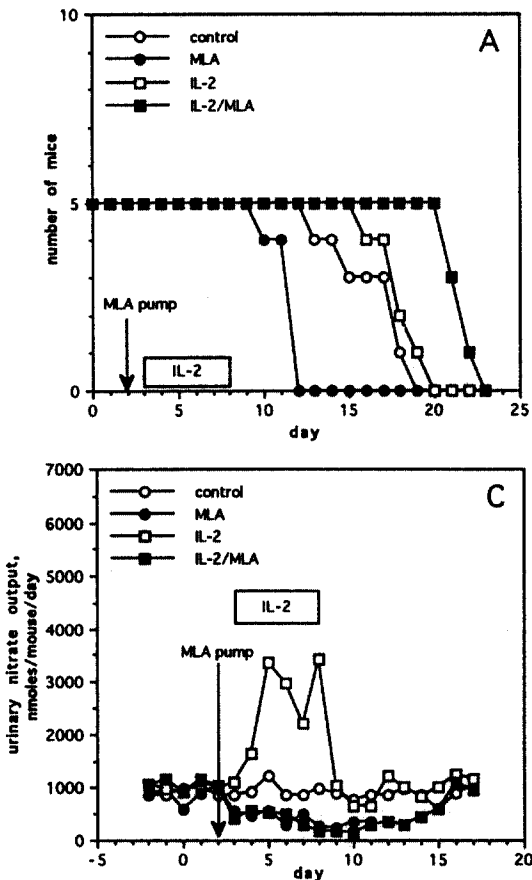


Fig. 3. Evaluation of the role of IL-2 therapy-induced NO synthesis in the treatment of intraperitoneal Meth A tumor. Four groups of BALB/c mice bearing Meth A ascites tumor were treated (untreated control, MLA therapy only, IL-2 therapy only, and IL-2/MLA therapy). Survival (Panel A) and the rate of body weight increment (Panel B) (Bars express mean \pm SD, $p < 0.05$ on day 9, control vs MLA and IL-2 vs IL-2/MLA) of the mice were evaluated and daily urinary nitrate excretion (Panel C) was monitored.

3. Effect of NO Synthesis Inhibition on IL-2 Induced Anti-Tumor Responses *in vivo*

To evaluate whether IL-2 induced NO synthesis represents an anti-tumor effector mechanism or an inhibitory factor against therapeutic effects of IL-2, mice were acclimated to metabolic cages on a nitrate/nitrite-free diet. On day 0, four groups of mice (5 mice in each group) were injected intraperitoneally with 2×10^6 Meth A tumor cells. On day 2, mice receiving MLA treatment were implanted subcutaneously with Alzet continuous infusion pumps (Model 2001) containing 0.2 ml of 3.38 M MLA. The remaining groups of mice underwent anesthesia and subcutaneous tunnel formation but no pump implantation. On day 3, IL-2 therapy was begun (180,000 IU s.c. every 12h for 5 days). This experiment demonstrated that IL-2 therapy of Meth A ascites-bearing mice induced no significant changes in the rate of body weight increment (ascites growing) (on day 9, $8.8 \pm 6.1\%$ in IL-2 therapy group versus $11.5 \pm 6.2\%$ in control mice, $p = 0.365$) and in mean survival (18.2 days in IL-2 therapy group versus 16.6 days in control mice, $p = 0.255$) (Fig. 3A, 3B). In contrast, chronic MLA administration significantly decreased the rate of body weight increment (on day 9, $-2.9 \pm 5.0\%$ in IL-2/MLA therapy group versus $8.8 \pm 6.1\%$ in IL-2 therapy group, $p < 0.001$) and prolonged mean survival of IL-2 treated mice (21.8 days in IL-2/MLA therapy group versus 18.2 days in IL-2 therapy group, $p = 0.001$).

Interestingly, the MLA treatment increased the rate of body weight increment (on day 9, $11.5 \pm 6.2\%$ in control group versus $35.1 \pm 7.2\%$ in MLA therapy group, $p < 0.001$) and diminished the survival of untreated control mice (16.6 days in control group versus 11.6 days in MLA therapy group, $p = 0.003$). In order to confirm the effectiveness of NOS inhibition, pooled daily urines from experimental mice were analyzed for urinary nitrate excretion (Fig. 3C). MLA administration via Alzet continuous infusion pumps achieved approximately 60% suppression of urinary nitrate excretion by control mice. Subcutaneous IL-2 treatment strongly induced nitric oxide synthesis (up to $3.5 \mu\text{moles/mouse/day}$), a 4-fold increase. MLA effectively suppressed NO production due to cytokine stimulation when given as subcutaneous continuous infusion.

DISCUSSION

NO is a well known cytotoxic effector molecule which may contribute to the development of cell mediated immune responses in a number of ways including the primary defense against facultative and obligate intracellular pathogens²³ and tumor cell killing^{10,11}. On the other hand, NO has potentially immunosuppressive effects on anticancer immune responses. A large number of studies have demonstrated that macrophage NO synthesis can decrease lectin and antigen stimulated lymphocyte proliferation¹²⁻¹⁵. In high concentration, NO appears to inhibit antigen-specific cytotoxic T cell activation during infection^{14,15} and during the course of graft-versus host disease^{24,25}. NO may also decrease macrophage Class II MHC antigen expression, which may impede the process by which antigen is presented to T cells²⁶. Our own studies have established that direct exposure of splenocytes to high concentrations of NO strongly inhibits IL-2 induced LAK cell activation and expansion (manuscript in preparation). Thus, both anti-tumor and tumor promoting activities of NO appear possible *in vivo*.

The observation that MLA is a potent competitive inhibitor of the NOS enzyme provided an additional experimental tool to differentiate effector functions mediated by the L-arginine: NO pathway^{27,28}.

LAK cell induction is observed in most IL-2 treated patients^{29,30} and has been thought to be important in mediating anti-tumor responses of IL-2 therapy. Hibbs and co-workers recently demonstrated that IL-2 treatment of patients with malignant melanoma and renal cell carcinoma strongly induces NO synthesis¹⁹. We hypothesized that cytokine induced NO synthesis represented a novel inhibitory factor against LAK cell activation and therefore diminished the anti-tumor effect of IL-2 therapy. In the present study, we employed Meth A ascites tumor, which is relatively resistant to IL-2 therapy, as a model. Subcutaneous injections of IL-2 into a remote site were used to produce a systemic immune response. This mode of treatment avoided recurrent intraperitoneal injections, that could have disseminated ascites tumor.

Experiments reported herein established that significant nitrite synthesis was induced in malignant ascites during IL-2 treatment. Addition of the NO

synthase inhibitor MLA to cell cultures demonstrated a dose dependent inhibition of the nitrite synthesis, confirming the origin of the nitrite to be from the L-arginine: NO pathway. These results strongly indicate that NO synthesis can be induced in ascites tumor cells by IL-2: This experimental result is consistent with earlier observation by Cox et al. who demonstrated that IL-2 could activate murine macrophage NO synthesis, particularly in the presence of IFNg³¹. Further studies will be necessary to establish whether IL-2 is acting directly on macrophages, via IL-2 receptors³². Alternatively, secondarily released inflammatory mediators, such as TNF, IL-1 and IFNg, produced by lymphocytes within the ascites, may play an obligate role in IL-2 induced NO synthesis by macrophages^{31,33}. Cytokine-activated macrophages are known to increase secretion of IL-1, TNF and IL-6, which may also enhance induction of iNOS activity within macrophages themselves and tumor cells^{34,35}. The cellular source of NO produced during IL-2 therapy remains to be studied. Tumor infiltrating macrophages and tumor cells may be the first candidates for the NO synthesis because there are precedents for macrophages and tumor cells to synthesize NO following cytokine induction^{10,11,34,35}. It seems unlikely that T or NK lymphocytes contribute significantly to intraperitoneal NO synthesis. Our unpublished data represented that the cytokine exposure, which are well known stimulus to iNOS expression in macrophages^{10,11,19}, did not induce NO synthesis in lymphocyte cultures.

This model demonstrated that IL-2 treatment marginally increased mean survival of tumor bearing mice from 16.6 to 18.2 days ($p=0.225$). The subcutaneous regimen of IL-2 administration induced marked (up to 4 fold) increases in urinary nitrate excretion, demonstrating potent induction of NO synthesis *in vivo*. This increase was markedly inhibited by concomitant MLA administration. At the same time, the marginally beneficial effects of IL-2 treatment on survival was significantly increased by MLA administration, prolonging mean survival from 18.2 to 21.8 days ($p=0.001$). Likewise, the rate of body weight increment of IL-2 treatment group was significantly delayed by the treatment with MLA. These results indicated that high output NO synthesized during IL-2 therapy is an inhibitory factor against the therapeutic anti-tumor responses. In contrast, survival and the rate of body weight

increment of tumor-bearing control mice were diminished and increased respectively by MLA administration, suggesting the possibility of small but definite anti-tumor effect of baseline NO synthesis in these animals. The latter finding is consistent with earlier observations by Yim et al. who demonstrated that endogenous baseline NO production in mice bearing antigenic subcutaneous tumor decreases the rate of tumor progression^{21,37}.

Mechanisms which can explain the contrasting difference of the role of NO in tumor immunology between the absence and presence of IL-2 therapy remain to be clarified. To unify these conflicting series of observations, we have proposed that, in the absence of IL-2 therapy, there may be little or no activated cytotoxic lymphocytes in tumor tissues and lymphocytes may, therefore, not be a major anti-tumor effector mechanism. Baseline endogenous NO synthesis may then be an important mechanism which delays tumor growth in this setting. In contrast, in the presence of IL-2 therapy, both LAK cell activation, as well as high output NO synthesis, are induced in the tumor microenvironment. This high output NO can be cytotoxic not only to tumor cells but also prominently inhibit induction of LAK cell activation and proliferation. Based on the previous reports indicating that the anti-tumor effects of NO are mainly cytostatic rather than cytolytic¹¹, but those of LAK cells are cytolytic rather than cytostatic¹⁻⁶, it may be possible that LAK cells are more effective than high output NO in inhibiting tumor growth. Inhibition of NO synthesis during IL-2 therapy, may therefore, result in increases in LAK cell activation and be beneficial to the therapeutic anti-tumor responses. The interrelationship of the anti-tumor and immunosuppressive roles of NO is likely to be complex and will require further evaluation.

Systemic high output NO synthesis is very likely to contribute to dose limiting toxicities of IL-2 therapy, such as hypotension, vascular leak and decreased myocardial performance (Yim, et al., manuscript submitted). Thus, the possibility exists that MLA administration during IL-2 therapy is beneficial in preventing the toxicities, as well as in promoting therapeutic anti-tumor responses.

REFERENCES

1. Yim C-Y, Hibbs JB Jr, McGregor JR, Galinsky RE,

EFFECTS OF NITRIC OXIDE (NO) SYNTHESIS INHIBITION ON ANTITUMOR RESPONSES DURING INTERLEUKIN-2 (IL-2) TREATMENT OF MICE

- Samlowski WE. Use of N-actyl cysteine to increase intracellular glutathione during the induction of anti-tumor responses by IL-2. *J Immunol* 1994; 152:5796.
2. Lotze MT, Grimm EA, Mazumder A, Strausser JL, Rosenberg SA. Lysis of fresh and cultured autologous tumor by human lymphocytes cultured in T-cell growth factor. *Cancer Res* 1981; 41:4420.
 3. Rayner AA, Grimm EA, Lotze MT, Wilson DJ, Rosenberg SA. Lymphokine-activated killer (LAK) cell phenomenon. IV. Lysis by LAK cell clones of fresh human tumor cells form autologous and multiple allogenic tumors. *J Natl Cancer Inst* 1985; 75:67.
 4. Grimm EA, Mazumder A, Zhang HZ, Rosenberg SA. Lymphokine-activated killer cell phenomenon: Lysis of natural killer-resistant fresh solid tumor cells by interleukin-2 activated autologous human peripheral blood lymphocytes. *J Exp Med* 1982; 155:1823.
 5. Harker WG, Tom C, McGregor JR, Slade L, Samlowski WE. Human tumor cell line resistance to chemotherapeutic agents does not predict resistance to natural killer or lymphokine-activated killer cell mediated cytotoxicity. *Cancer Res* 1990; 50:5931.
 6. Gambacorti-Passerini C, Rivoltini L, Supino R, Rodolfo M, Radizzani M, Fossati G, Parmiani G. Susceptibility of chemoresistant murine and human tumor cells to lysis by interleukin-2 activated lymphocytes. *Cancer Res* 1988; 48:2372.
 7. Rosenberg SA. Adoptive cellular therapy in patients with advanced cancer: An update. *Biol Ther Cancer* 1991; 1:1.
 8. Dutcher JP, Creekmore S, Weiss GR, Margolin K, Markowitz AB, Roper M, Parkinson D, Ciobanu N, Fisher RI, Boldt DH, Doroshow JH, Rayner AA, Hawkins M, Atkins M. A phase II study of interleukin-2 and lymphokine-activated killer cells in patients with metastatic malignant melanoma. *J Clin Oncol* 1989; 7:477.
 9. McCabe MS, Stablein D, Hawkins MJ and the NCI Modified Group C Investigators. The modified Group C experience-Phase III randomized trials of IL-2 vs IL-2/LAK in advance renal cell carcinoma and advanced melanoma. [Abstract] *Am Soc Clin Oncol Proc* 1991; 10:213.
 10. Hibbs JB Jr, Taintor RR, Vavrin Z, Rachlin EM. Nitric oxide: a cytotoxic activated macrophage effector molecule. *Biochem. Biophys. Res Commun* 1988; 157: 87.
 11. Stuehr DJ, Nathan CF. Nitric oxide: A macrophage product responsible for cytostasis and respiratory inhibition in tumor target cells. *J Exp Med* 1989; 169: 1543.
 12. Mills CD. Molecular basis of "suppressor" macrophages. Arginine metabolism via the nitric oxide synthase pathway. *J Immunol* 1991; 146:2719.
 13. Albina JE, Abate JA, Henry WL Jr. Nitric oxide production is required for murine resident peritoneal macrophages to suppress mitogen stimulated T cell proliferation. Role of IFN-gamma in the induction of the nitric oxide-synthesizing pathway. *J Immunol* 1991; 147:144.
 14. Gregory SH, Wing EJ, Hoffman RA, Simmons RL. Reactive nitrogen intermediates suppress the primary immunologic response to *Listeria*. *J Immunol* 1993; 150:2901.
 15. Stefani MM, Muller I, Louis JA. *Leishmania major*-specific CD8+ T cells are inducers and targets of nitric oxide produced by parasitized macrophages. *Eur J Immunol* 1994; 24:746.
 16. Gemlo BT, Palladino MA, Jaffe HS, Espevik TP, Rayner AA. Circulating cytokines in patients with metastatic cancer treated with recombinant interleukin-2 and lymphokine-activated killer cells. *Cancer Res* 1988; 48:5864.
 17. Mier JW, Vachino G, Van der Meer JWM, Numerof RP, Adams S, Cannon JG, Bernheim HA, Atkins MB, Parkinson DR, Dinarello CA. Induction of circulating tumor necrosis factor (TNF α) and the mechanism for the febrile response to interleukin-2 (IL-2) in cancer patients. *J Clin Immunol* 1988; 8:426.
 18. Deng W, Thiel B, Tannenbaum CS, Hamilton TA, Stuehr DJ. Synergistic cooperation between T cell lymphokines for induction of the nitric oxide synthase gene in murine peritoneal macrophages. *J Immunol* 1993; 151:32.
 19. Hibbs JB Jr, Westenfelder C, Taintor R, Vavrin Z, Kablitz C, Baranowski RL, Ward JH, Menlove RL, McMurry MP, Kushner JP, Samlowski WE. Evidence for cytokine inducible nitric oxide synthesis from L-arginine in patients receiving interleukin-2 therapy. *J Clin. Invest* 1992; 89:867.
 20. Borberg H, Oettgen HF, Choudry K, Beattie EJ. Inhibition of established transplants of chemically induced sarcomas in syngeneic mice by lymphocytes from immunized donors. *Int J Cancer* 1972; 10:539.
 21. Yim C-Y, Bastian NR, Smith JC, Hibbs JB Jr, Samlowski WE. Macrophage nitric oxide synthesis delays progression of ultraviolet light induced murine skin cancers. *Cancer Res* 1993; 65:5507.
 22. Granger DL, Hibbs JB Jr, Perfect JR, Durack DT. Metabolic fate of L-arginine in relation to microbistatic capability of murine macrophages. *J Clin Invest* 1990; 85:264.
 23. Nathan CF, Hibbs JB Jr. Role of nitric oxide synthesis in macrophage antimicrobial activity. In *Current Opinion in Immunology*. Silverstein S and Unkeless J, eds. Current Science, London p 65.

24. Radi R, Beckman JS, Bush KM, Freeman BA. Peroxynitrite oxidation of sulfhydryls: The cytotoxic potential of superoxide and nitric oxide. *J Biol Chem* 1991; 266:4244.
25. Hoffman RA, Langrehr JM, Wren SM, Dull KE, Ildstad ST, McCarthy SA, Simmons RL. Characterization of the immunosuppressive effects of nitric oxide in graft vs host disease. *J Immunol* 1993; 151: 1508.
26. Sicher SC, Vazquez MA, Lu CY. Inhibition of macrophage Ia expression by nitric oxide. *J Immunol* 1994; 153:1293.
27. Hibbs JB Jr, Vavrin Z, Taintor RR. L-arginine is required for expression of the activated macrophage effector mechanism causing selective metabolic inhibition in target cells. *J Immunol* 1987; 138:550.
28. Hibbs JB Jr, Taintor RR, Vavrin Z. Macrophage cytotoxicity: role for L-arginine deiminase and imino nitrogen oxidation to nitrite. *Science* 1987; 235:473.
29. Boldt DH, Mills BJ, Gemlo BT, Holden H, Mier J, Paietta E, McMannis JD, Escobedo LV, Sniecinski I, Rayner AA, Hawkins MJ, Atkins MB, Ciobanu N, Ellis TM. Laboratory correlates of adoptive immunotherapy with recombinant interleukin-2 and lymphokine-activated killer cells in humans. *Cancer Res* 1988; 48: 4409.
30. Ghosh AK, Dazzi H, Thatcher N, Moor M. Lack of correlation between peripheral blood lymphokine-activated killer (LAK) cell function and clinical response in patients with advanced malignant melanoma receiving recombinant interleukin-2. *Int J Cancer* 1989; 43:410.
31. Cox GW, Melillo G, Chattopadhyay U, Mullet D, Fertel RH, Varesio L. Tumor necrosis factor- α -dependent production of reactive nitrogen intermediates mediates IFN- γ plus IL-2-induced murine macrophage tumoricidal activity. *J Immunol* 1992; 149:3290.
32. Espinoza-Delgado I, Ortaldo JR, Winkler-Pickett R, Sugamura K, Varesio L, Longo DL. Expression and role of p75 interleukin-2 receptor on human monocytes. *J Exp Med* 1990; 171:1821.
33. Velasquez WS, Cabanillas F, Salvador P, McLaughlin P, Fridrik M, Tucker S, Jagannath S, Hagemester FB, Redman JR, Swan F, Barlogie B. Effective salvage therapy for lymphoma with cisplatin in combination with high-dose Ara-C and dexamethasone (DHAP). *Blood* 1988; 71:117.
34. Amber IJ, Hibbs JB Jr, Taintor RR, Vavrin Z. Cytokines induce an L-arginine-dependent effector system in nonmacrophage cells. *J Leukoc Biol* 1988; 44:58.
35. Amber IJ, Hibbs JB Jr, Taintor RR, Vavrin Z. The L-arginine dependent effector mechanism is induced in murine adenocarcinoma cells by culture supernatant from cytotoxic activated macrophages. *J Leukoc Biol* 1988; 43:187.
36. Drapier J-C, Wietzerbin J, Hibbs JB Jr. Interferon- γ and tumor necrosis factor induce the L-arginine-dependent cytotoxic effector mechanism in murine macrophages. *Eur J Immunol* 1988; 18:1587.
37. Yim CY, Lee CH, Choi SM, Yoo WH, Lee SJ, Lim SH, Sohn MH. Nitric oxide synthesis in murine skin cancers. *J Kor Cancer Assoc* 1995; 27:101.