



REVIEW ARTICLE OPEN

Signaling pathways in the regulation of cytokine release syndrome in human diseases and intervention therapy

Xia Li^{1,2,3,4}, Mi Shao^{1,2,3,4}, Xiangjun Zeng^{1,2,3,4}, Pengxu Qian^{1,2,3,4,5}✉ and He Huang^{1,2,3,4}✉

Cytokine release syndrome (CRS) embodies a mixture of clinical manifestations, including elevated circulating cytokine levels, acute systemic inflammatory symptoms and secondary organ dysfunction, which was first described in the context of acute graft-versus-host disease after allogeneic hematopoietic stem-cell transplantation and was later observed in pandemics of influenza, SARS-CoV and COVID-19, immunotherapy of tumor, after chimeric antigen receptor T (CAR-T) therapy, and in monogenic disorders and autoimmune diseases. Particularly, severe CRS is a very significant and life-threatening complication, which is clinically characterized by persistent high fever, hyperinflammation, and severe organ dysfunction. However, CRS is a double-edged sword, which may be both helpful in controlling tumors/viruses/infections and harmful to the host. Although a high incidence and high levels of cytokines are features of CRS, the detailed kinetics and specific mechanisms of CRS in human diseases and intervention therapy remain unclear. In the present review, we have summarized the most recent advances related to the clinical features and management of CRS as well as cutting-edge technologies to elucidate the mechanisms of CRS. Considering that CRS is the major adverse event in human diseases and intervention therapy, our review delineates the characteristics, kinetics, signaling pathways, and potential mechanisms of CRS, which shows its clinical relevance for achieving both favorable efficacy and low toxicity.

Signal Transduction and Targeted Therapy (2021)6:367; <https://doi.org/10.1038/s41392-021-00764-4>

INTRODUCTION

Cytokine release syndrome (CRS) is characterized by a mixture of clinical manifestations, including elevated circulating cytokine levels, acute systemic inflammatory symptoms, and secondary organ dysfunction, while severe CRS is a very significant and life-threatening complication. Given the lack of a unified and widely accepted definition of CRS, Fajgenbaum and his colleagues proposed the following three criteria to identify CRS: (1) elevated circulating cytokine levels, (2) acute systemic inflammatory symptoms, and (3) secondary organ dysfunction (often renal, hepatic, or pulmonary) due to inflammation beyond that which could be attributed to a normal response to a pathogen (if a pathogen is present) or any cytokine-driven organ dysfunction (if no pathogen is present).¹ However, CRS is a double-edged sword, which may be both helpful in controlling tumors/viruses/infections and harmful to the host.

It has important clinical relevance to acknowledge that CRS facilitates disease control and may be harmful to the host. To better understand the cause of CRS, the history and general characteristic of CRS were initially introduced. Then, we summarized clinical features and risk factors of CRS caused by iatrogenic factors, pathogenic factors, monogenic disorders, and autoimmune diseases. The cellular and molecular mechanisms of CRS among these human diseases were further

highlighted. Finally, CRS models, cutting-edge technologies, management of CRS, and future direction were also discussed.

THE HISTORY AND GENERAL CHARACTERISTICS OF CRS

In 1993, Ferrara et al. coined the term ‘cytokine storm’ to describe the clinical manifestations of acute graft-versus-host disease after allogeneic hematopoietic stem-cell transplantation.² Later, cytokine storm was also known as cytokine release syndrome (CRS) and reported in the influenza pandemic³ and severe acute respiratory syndrome coronavirus (SARS-CoV).⁴ In addition, chimeric antigen receptor T cell (CAR-T cell) immunotherapy was the fourth pillar in cancer therapeutics,^{5–11} in which CRS was the most common clinical complication and was first reported in 2010.¹² Meanwhile, coronavirus disease 2019 (COVID-19), which is characterized by severe respiratory illness in humans, is also accompanied by CRS.^{13–15} Furthermore, the significant boom in the publication of studies with the key words ‘cytokine release syndrome’, ‘chimeric antigen receptor T cell’, or ‘COVID-19’ in PubMed (<https://pubmed.ncbi.nlm.nih.gov/>) in recent decades suggests that CRS, CAR-T, and COVID-19 are important issues in the fields of cancer, immunotherapy, and viral infection (Fig. 1). Meanwhile, CRS phenomenon can be also observed in other human diseases, such as monogenic disorders¹⁶ and autoimmune diseases.¹⁷

¹Bone Marrow Transplantation Center, The First Affiliated Hospital, Zhejiang University School of Medicine, Hangzhou, People's Republic of China; ²Liangzhu Laboratory, Zhejiang University Medical Center, 1369 West Wenyi Road, Hangzhou 311121, People's Republic of China; ³Institute of Hematology, Zhejiang University, Hangzhou, Zhejiang, People's Republic of China; ⁴Zhejiang Province Engineering Laboratory for Stem Cell and Immunity Therapy, Hangzhou, Zhejiang, People's Republic of China and ⁵Center of Stem Cell and Regenerative Medicine, Zhejiang University School of Medicine, Hangzhou, People's Republic of China

Correspondence: Pengxu Qian (axu@zju.edu.cn) or He Huang (huanghe@zju.edu.cn)

These authors contributed equally: Xia Li, Mi Shao, Xiangjun Zeng

Received: 11 February 2021 Revised: 9 August 2021 Accepted: 5 September 2021

Published online: 20 October 2021

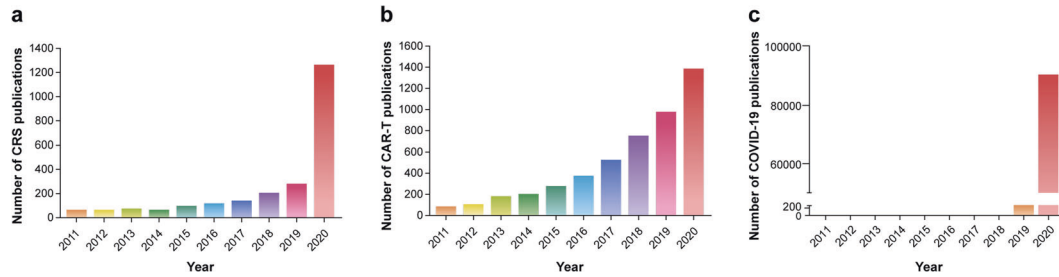


Fig. 1 The increasing number of publications searched by the key words ‘cytokine release syndrome’ (a), ‘chimeric antigen receptor T (CAR-T) cell’ (b) or ‘COVID-19’ (c) in PubMed (<https://pubmed.ncbi.nlm.nih.gov>) in recent decades

Table 1. The sources and functions of CRS-related cytokines in different CRS studies

Cytokines	Sources	Functions	Reference
Antitumor effector cytokines			
GZMB	Cytotoxic T lymphocytes, NK cells	Antitumor, proapoptotic, and antiviral activities	Ma et al. (2020); ¹⁸ Gust et al. (2019) ¹⁹
IFN-γ	Th1 cells, NK cells, CD8 T cells	Activates macrophages, supports Th1 differentiation, antiproliferative activities, weakens antiviral activity	Zhang et al. (2019) ²⁰
TNF-α	Monocytes, macrophages, activated T cells, neutrophils	Strong mediator of inflammatory and immune functions, regulates cell growth and differentiation	Zhang et al. (2018) ²¹
MIP-1α	Eosinophils	Attracts leukocytes	Xue et al. (2017) ²²
Stimulatory and regulatory cytokines			
GM-CSF	Bone marrow stromal cells, macrophages	Essential for the growth and differentiation of neutrophils	Sachdeva et al. (2019); ²³ Sterner et al. (2019) ²⁴
IL-2	Activated T cells	Stimulates the proliferation and differentiation of T and B cells, activates NK cells	Golumba-Nagy et al. (2018) ²⁵
MCP-1	Monocytes, macrophages, tumor cells	Stimulates the motility of T cells, NK cells and basophils; induces the recruitment and activation of monocytes and macrophages during inflammation	Hirayama et al. (2019) ²⁶
NO	T cells, macrophages, fibroblasts, endothelial cells	Triggers apoptosis	Giavridis et al. (2018) ²⁷
IL-15	Primarily dendritic cells, cells of the monocyte lineage	Stimulates the proliferation and development of NK cells and T cells	Atilla et al. (2020); ¹⁹⁴ Giuffrida et al. (2020) ²⁸
Inflammatory cytokines			
IL-1α/IL-1β	Monocytes, macrophages, dendritic cells, endothelial cells, fibroblasts, adipocytes	Induction of local inflammation and systemic effects such as fever, the acute-phase response and stimulation of neutrophil production	Norelli et al. (2018); ²⁹ Giavridis et al. (2018) ²⁷
IL-6	T cells, B cells, macrophages, bone marrow stromal cells, fibroblasts	Regulates B and T cell functions, effects on hematopoiesis in vivo, induces inflammation and the acute-phase response	Norelli et al. (2018) ²⁹
IL-17α	CD4, CD8, and γδ T cells; NK cells	Promotes inflammation by increasing the production of pro-inflammatory cytokines and chemokines that attract monocytes and neutrophils	Rossi et al. (2018) ³⁰

In general, CRS caused by various factors involves a mixture of common elevated cytokines, including tumor necrosis factor (TNF)-α, interleukin (IL)-1, IL-6, and interferon gamma (IFN-γ). To further explore the CRS-related cytokine functions, we summarized 12 cytokines and divided them into three groups: antitumor effectors (granzyme B (GZMB), IFN-γ, macrophage inflammatory protein (MIP)-1α, and TNF-α),^{18–22} stimulatory and regulatory cytokines (GM-CSF, IL-2, MCP-1, NO, and IL-15),^{23–28} and inflammatory cytokines (IL-1, IL-6, and IL-17A).^{27,29,30} Their sources, functions and related studies are summarized in Table 1.

CRS IN IATROGENIC DISEASES

CRS in CAR-T-cell therapy

Clinical features. The clinical manifestations of CRS are characterized by high fever, hypotension, hypoxia, capillary leakage, coagulopathy, respiratory distress and severe organ dysfunction.³¹ Severe complications, including liver transaminitis and renal

insufficiency, are life-threatening and frequently require ICU care with vasopressors and/or ventilation support.^{32,33} To better understand the dynamic changes in CRS, the present review artificially divided CRS into three stages based on the general changing dynamics of CRS-related cytokine levels and CAR-T-cell counts: CRS initiation stage, CRS peak stage, and CRS recovery stage. In the CRS initiation stage, the levels of serum TNF-α, IL-1, IL-2, IL-10, and C-reactive protein (CRP) first increase. Then, temperature and the levels of ferritin, IFN-γ, and IL-6 are upregulated from the CRS initiation stage to the peak stage. During the CRS recovery stage, IL-4 and IL-17α levels are increased in turn, while the levels of the other above-mentioned cytokines gradually decline (Fig. 2).

To explore the distribution of different CRS grades, in 2017, Hay et al. reported data for 133 adult patients who received CD19 CAR-T cells.³⁴ According to the consensus classification criteria of CRS grade proposed by Lee and his colleagues,³⁵ they found that CRS developed in 70% patients, including 62.5% with grade 1~3

CRS and 7.6% with grade 4~5 CRS. In another study, Curran et al. found that 16% (4 of 25) of patients with relapsed/refractory (r/r) B-cell acute lymphoblastic leukemia (ALL) experienced severe CRS.³⁶ The percentages of CRS of different grades in different diseases and their classification criteria are also summarized in Table 2.^{8,34,37-45} It is worth noting that the risk of severe CRS in B-cell malignancies is the highest for ALL, lymphoma, and multiple myeloma.

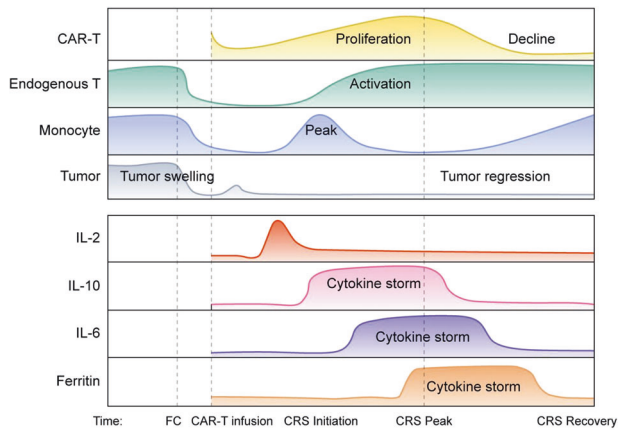


Fig. 2 The in vivo kinetics of cell counts and cytokine levels in the serum during the CRS process after CAR-T-cell therapy. Patients were treated with tocilizumab or corticosteroids when CRS reached at grade 3–4

Risk factors. Current studies suggest that the severity of CRS is associated with tumor burden (the number of malignant bone marrow cells), basic characteristics of patients (age, gut microflora, etc.), and quality/quantity of CAR-T cells (including the construct, infusion dose and amplification ability).

Tumor burden. In general, CRS is more frequent and severe in patients with a high tumor burden. The tumor burden may not only determine the peak number of CAR-T cells but also affect the time course of CAR-T-cell proliferation. However, Curran et al. concluded that the dose intensity of conditioning chemotherapy and the minimal pretreatment disease burden positively impacted the response without increasing toxicity.³⁶ These phenomena may occur because fludarabine/cyclophosphamide (FC)-mediated lymphodepletion provides more 'space' for CAR-T-cell proliferation, reducing competition with other lymphocytes for growth factors and removing inhibitory immune cells.⁴⁶

Basic characteristics in patients. In addition to the tumor burden, patient immune status should also be considered for CAR-T-cell therapy and CRS control. Regarding patient aging-related factors, Zettler et al. reviewed a total of 804 cases, which included 471 young patients (<65 years old) and 333 aged patients (>65 years old).⁴⁷ CRS was the most common adverse event reported in both young and aged groups. Although CRS severity was not significantly different, some individual clinical features of CRS, including pyrexia, tachycardia, and thrombocytopenia, were significantly more common in the younger age group. For gut microflora related to individual patient immune status, Kale et al. demonstrated that 'small' intestinal immunopathology played a 'big' role in lethal CRS in a mouse model.⁴⁸ Furthermore, they found that IFN- γ -JAK/STAT-

Table 2. Studies on the incidence of CRS of different grades and the classification criteria

Disease/Target antigen	Costimulatory domain	CRS classification criteria	CRS percentage (severe rate)	Reference
ALL				
CD19	4-1BB	Consensus criteria and NCI CTCAE v4.03	70% (G3–5: 12%)	Hay et al. (2017) ³⁴
CD19	CD28	NCI consensus CRS grading system and NCI CTCAE v4.03	80% (G3-4: 16%)	Curran et al. (2019) ³⁶
CD19	CD28	Modified criteria of Lee and colleagues ³⁵ and NCI CTCAE v4.02	76% (G3-4: 28%)	Lee et al. (2015) ³⁷
CD19	4-1BB		77% (G3-4: 47%)	Maude et al. (2018) ³⁸
CD19	CD28	MSKCC CRS grading system and NCI CTCAE v4.03	85% (G3-4: 26%)	Park et al. (2018) ⁴⁴
CD19	4-1BB	Modified criteria of Lee and colleagues and NCI CTCAE v4.03	66.7% (G3-4: 40%)	Hu et al. (2017) ⁴⁵
CD22	4-1BB		76%	Fry et al. (2018) ³⁹
Lymphoma				
CD19	CD28	Modified criteria of Lee and colleagues and NCI CTCAE v4.03	92% (G3-4: 11%)	Locke et al. (2019) ⁴⁰
CD19	4-1BB	University of Pennsylvania grading scale and NCI CTCAE v4.03	58% (G3-4: 22%)	Schuster et al. (2019) ⁸
Multiple myeloma				
BCMA	4-1BB	Modified criteria of Lee and colleagues and NCI CTCAE v4.03	90% (G3-4: 7%)	Zhao et al. (2018) ¹⁹⁵
BCMA	4-1BB	Modified criteria of Lee and colleagues and NCI CTCAE v4.03	76% (G3-4: 6%)	Raje et al. (2019) ⁴²
CD19 & BCMA	4-BB & 4-1BB	Modified criteria of Lee and colleagues and NCI CTCAE v4.03	90% (G3-4: 5%)	Yan et al. (2019) ⁴³
CLL				
CD19	4-1BB	Consensus criteria and NCI CTCAE v4.03	70% (G3-5: 12%)	Hay et al. (2017) ³⁴

CRS grading is performed using the classification proposed by Lee and his colleagues, while grading of organ toxicities is performed according to Common Terminology Criteria for Adverse Events (CTCAE) Version 4.03

Table 3. Drugs triggered cytokine release syndrome

Antibody	Antigen	Driven cytokine and approximate time of CRS appearance, and Ref.
Muromonab	CD3	TNF- α , IFN- γ , IL-2, and IL-6; 2~4 h after infusion (Yan et al., 2019) ⁵⁵
Rituximab	CD20	TNF- α , IFN- γ , and IL-6; 2 h after infusion (Byrd et al., 2001) ⁵⁸
TGN1412	CD28	TNF- α , IFN- γ , IL-8, IL-10, and DIC; 1.5 h after infusion (Suntharalingam et al., 2006) ⁵⁹
CP-870,893	CD40	TNF- α and IL-6; minutes to hours after infusion (Vonderheide et al., 2007; ⁶⁴ and Nowak et al., 2015) ⁶³
Alemtuzumab	CD52	TNF- α , IFN- γ , and IL-6; 2~4 h after infusion (Ferrajoli et al., 2001; ⁶⁵ and Moreau et al., 1996) ⁶⁶
Blinatumomab	CD3 & CD19	TNF- α and IL-6; no time information (Topp et al., 2015) ⁶⁰
TDB	CD3 & HER2	TNF- α , IL-1, IL-2, and IL-6; 2 h after infusion (Li et al., 2019) ⁶¹
Nivolumab & ipilimumab	PD1 & CTLA4	CRP, aspartate aminotransferase, alanine aminotransaminase, D-dimer; minutes to day 2 after infusion (Urasaki et al., 2021) ⁶²

driven pathways and IL-17 α deficiency contributed to lethal small intestinal immunopathology in T cell-driven CRS.

Quality and quantity of CAR-T cells. CAR-T cells themselves are also important in CRS. In regard to CAR-T-cell constructs, our group demonstrated that dual-targeted CD19/CD22 CAR-T-cell therapy had a lower incidence of CRS than single-target CD19 CAR-T-cell therapy, especially in severe CRS (≥ 3 grade CRS).⁴⁹ Additionally, Magnani et al. reported that sleeping beauty-engineered CAR-T cells achieved activity without severe toxicities, with two grade 1 CRS cases and one grade 2 CRS case occurring in 13 patients with B-cell ALL.⁵⁰ On the other hand, the CD4⁺/CD8⁺ ratio of CAR-T-cell infusion products (IPs)⁵¹ and the dose of administered CAR-T cells⁵² are always positively correlated with the treatment effect and CRS severity. Furthermore, the CAR-T-cell infusion strategy is also a factor to consider. In 2020, Frey et al. demonstrated that compared with a single infusion, fractionated dosing of CAR-T cells with inpatient dose modification (CAR-T-cell infusion: day 1, 10%; day 2, 30%; and day 3, 60%) optimized safety (low and manageable CRS) without compromising efficacy (higher survival rate) in adults with ALL.⁵³

CRS in other immune therapy. CRS is not restricted to CAR-T-cell therapy and is also associated with other monoclonal antibody (mAb) treatment. These antibodies include: muromonab (OKT3), rituximab, TGN1412, CP-870,893, blinatumomab, and TDB.

OKT3, an anti-CD3 antibody mainly indicated for the treatment of acute organ transplant rejection, may also lead to severe CRS with increased serum levels of cytokines (e.g., TNF- α , IL-2, IL-6, and IFN- γ).^{54,55} Rituximab, a chimeric anti-CD20 mAb, is widely used to treat hematological malignancies, including CD20-positive diffuse large B-cell lymphoma and⁵⁶ and chronic lymphocytic leukemia.⁵⁷ Severe CRS has occurred in patients receiving rituximab, who had severe dyspnea, fever and blood pressure changes. Cytokine of TNF- α , IL-6, and IFN- γ reached the peak at 2 h after rituximab infusion in chronic lymphocytic leukemia (CLL) patients.⁵⁸ Besides, TGN1412, a superagonist anti-CD28 humanized mAb, can directly stimulate T cells and treatment of healthy volunteers with TGN1412 led to severe CRS and increased disseminated intravascular coagulation (DIC).⁵⁹

On the other hand, the Kantarjian's group demonstrated that single-agent blinatumomab (a bispecific antibody that binds to CD19 tumor cells and CD3 T cells) showed antileukemia activity in adult patients with relapsed or refractory B-precursor ALL, in which 2% patients had grade 3 CRS.⁶⁰ In 2019, Li et al. found that CRS was also induced after the treatment of anti-HER2/CD3 T cell-dependent bispecific (TDB) antibody.⁶¹ It is worth noting that systemic serum expression of TNF- α , IL-2, and IL-6 was substantially increased only at 2 h after treatment. Furthermore, TNF- α blockade can prevent macrophage activation without

affecting T cell killing capacity and effectively mitigates CRS in anti-CD3 bispecific antibody therapy. In addition, immune checkpoint inhibitors with nivolumab- (anti-programmed cell death-1 mAb) and ipilimumab- (anti-cytotoxic T lymphocyte antigen-4 mAb) induced CRS were also reported in 2021, in which CRP, aspartate aminotransferase, alanine aminotransaminase, and D-dimer were significantly increased after nivolumab and ipilimumab infusion.⁶² At last, other mAbs⁶³⁻⁶⁶ most commonly associated with CRS are also listed in Table 3.

CRS IN PATHOGENIC DISEASES

CRS in COVID-19

Clinical features. CRS is observed in infections and a significant part of the pathogen-related diseases, including bacterial sepsis, HHV-8-associated MCD, and COVID-19. Coronavirus disease 2019 (COVID-19), which is caused by SARS-CoV-2, has become one of the worst pandemics of 2020 and has caused a large number of deaths.⁶⁷ COVID-19 is characterized by heterogeneous symptoms ranging from mild fatigue to life-threatening pneumonia, CRS syndrome, and multi-organ failure.¹ The laboratory features of COVID-19 include lymphocytopenia, leukopenia, and thrombocytopenia.^{68,69}

The elevated cytokines in the serum of patients with COVID-19 include IL-1 β , IL-6, IP-10, TNF, IFN- γ , MIP 1 α /1 β , and VEGF.^{70,71} Furthermore, patients with severe COVID-19 may have significantly higher levels of inflammatory parameters, including CRP, ferritin, D-dimer, and pro-inflammatory cytokines (such as TNF- α , GM-CSF, IL-6, IP-10, MCP-1, and MCP-1a), than patients with mild COVID-19, which demonstrated the presence of a CRS in severe affected patients. Higher IL-6 levels (hyperinflammation) and tissue damage are strongly associated with shorter survival in COVID-19.^{72,73} Wang et al.⁷⁴ found that the increase in IL-6 occurred 1–2 days prior to the decreases in CD8 and CD4 T cells. Liu et al.⁷⁵ observed that CD4 and CD8 T-cell numbers dropped to their lowest levels after 4–6 days of illness, whereas IL-10, IL-2, TNF, and other cytokines reached peak levels.

Risk factors. Understanding the possible intricate risk factors associated with COVID-19 severity is helpful in identifying those patients who are at a high risk and require prioritized management to prevent disease progression and severe adverse outcomes. In this section, the severity of the COVID-19 would be discussed in terms of viral load and basic characteristics in patients (age, sex, and underlying diseases).

Viral load. Viral load varies greatly among different populations and is closely related to the course of disease.⁷⁶ The cycle threshold values of severe COVID-19 patients were significantly lower than mild COVID-19 patients, and the mean viral load of severe patients was much higher than mild COVID-19 patients.⁷⁷

The kinetics of viral load and exposure to large numbers of infective SARS-CoV-2 were highly predictive markers of severe outcomes in older patients.^{78,79} In addition, Lieberman et al. reported that variability in the antiviral response in males and elderly patients with COVID-19 was dependent on viral load and infection time.⁸⁰

Basic characteristics in patients. Higher severity was found to be associated with older age⁸¹ and male sex.⁸² Data from 79,394 confirmed COVID-19 patients in China showed that those aged below 30 and above 59 years were 0.6 and 5.1 times more likely to die than patients aged 30–59 years, respectively.⁸³ Biological age, which was comprised of chronological age and nine clinical chemistry biomarkers, was an optimal predictor of disease severity.⁸⁴

In addition to age and sex, preexisting comorbidities, such as hypertension, diabetes and obesity, are also external risk factors for the survival of critically ill patients with COVID-19. Hypertension, the most frequent comorbidity, was an independent risk factor for assessing the severity of COVID-19,⁸⁵ while a history of chronic obstructive pulmonary disease, hypercholesterolemia, and diabetes was also independently associated with mortality.⁸⁶

CRS in other pathogenic diseases. Infectious diseases are a leading cause of death in the whole world. The triggering of the inflammatory response, especially through cytokine production, is essential for the elimination of pathogens during infection. However, uncontrolled cytokine production would result in a CRS. Here, we summarized infection-associated CRS into two common categories: bacterial infections and viral infections.

The involvement of CRS and subsequent immunoparalysis has been reported in the development of bacterial infections with leptospirosis,⁸⁷ streptococcus,⁸⁸ and staphylococcus.⁸⁹ Disseminated bacterial infections can lead to fever, cell death, coagulopathies, and multi-organ dysfunction by producing many cytokines and inducing CRS. For the cellular mechanisms, macrophages are a major cell type driving the CRS during infection. In the development of sepsis, activated macrophages release a great number of pro-inflammatory cytokines and chemokines, including IL-1 α , IL-1 β , IL-6, and TNF- α , and initiate an inflammatory response.⁹⁰ Certain bacteria can lead to polyclonal activation of T cells, cytokine production, and toxic shock syndrome by producing super antigens that cross-link the major histocompatibility complex and T-cell receptors. For the molecular mechanisms, microbial pathogen-associated molecular patterns (PAMPs) will be recognized by the pattern recognition receptors (PRRs), which are expressed on the surface of innate immune cells.⁹¹ The PAMPs/PRR triggers an inflammatory cascade by activating multiple signaling modules, including NF- κ B and AP-1 transcription factors,⁹² which in turn regulate the expression of cytokines, PGs, and NO.

CRS was also observed in patients infected with influenza virus and coronavirus, such as H1N1, H1N5, SARS-CoV, and MERS-CoV.⁹³ Serum levels of IL-8, IL-9, IL-17, IL-6, IL-15, TNF- α , and IL-10 were increased in H1N1 influenza virus infection, while IL-8, IP-10, MCP-1, MIP-1, and CXCL-9 were elevated in H5N1 influenza virus infection.⁹⁴ Meanwhile, the release of IL-6 and IL-1 β from other immune cells promotes recruitment of T cells and neutrophils.⁹⁵ In addition, patients infected with EBV or cytomegalovirus may have accompanying perforin functional defects, which leads to defective clearance of antigen-bearing dendritic cells and prolonged engagement between lymphocytes and antigen-presenting cells.⁹⁶ These further resulted in continuous activation and proliferation of T cells and macrophages, and an autocrine loop of pro-inflammatory cytokines.⁹⁷ For the molecular mechanisms during the virus infection, both PAMPs and damage-associated molecular patterns (DAMPs) participate in innate immune responses through the cascade amplification of IFN.

Downstream production of IFN promotes intracellular antiviral defenses in neighboring epithelial cells which may limit viral dissemination.

CRS in monogenic and autoimmune diseases. CRS phenomena can also be observed in other human diseases, such as monogenic disorders and autoimmune diseases.

CRS in monogenic disorders. Recent years have witnessed a broad explosion of new autoinflammatory disorders. The monogenic autoinflammatory diseases are spontaneous or minimally triggered immune activation due to either loss-of-function mutations in genes that suppress inflammation or gain-of-function mutations in genes that propagate inflammation. According to the underlying mechanisms, monogenic autoinflammatory diseases are mainly divided into inflammasome/IL-1 diseases, the interferonopathies, and the NF- κ B/TNF disorders.⁹⁸

Inflammasome/IL-1 diseases include familial Mediterranean fever (with MEFV gene disorder), familial cold autoinflammatory syndrome (with NLRP3 disorder), autoinflammation with infantile enterocolitis (with NLRC4 gene disorder), and deficiency of IL-1 receptor antagonist (with L1RN gene disorder), and so on. As a typical inflammasome/IL-1 monogenic disorder, hemophagocytic lymphohistiocytosis (HLH) is characterized by uncontrolled activation of NK cells, cytotoxic T lymphocytes and macrophages, leading to immune-mediated injury of multiple organ systems.^{16,99} Clinically, patients with HLH are characterized by fever, hepatosplenomegaly, cytopenia, and activated macrophages in hemopoietic organs.¹⁰⁰ The pathophysiology of HLH is that enhanced antigen presentation and repeated IFN- γ -dependent stimulation of Toll-like receptors lead to the uncontrolled activation of antigen-presenting cells and T cells and produce an exaggerated inflammatory response (such as IL-1, IL-4, IL-6, IL-8, IL-10, IL-18, IFN- γ , and TNF- α).¹⁰⁰ Therefore, medications to block the cytokine network achieved considerable efficacy for controlling HLH. Glucocorticoids, cyclosporine, etoposide, anti-thymocyte globulin, and cytotoxic chemotherapies are effective in controlling both primary and secondary HLH by suppressing or eliminating T cells and other immune cells.¹⁰¹ In addition, anti-IL-1 antibodies, anti-IL-6 antibodies, JAK1 and JAK2 inhibitors, and anti-IFN- γ antibodies are also effective in blocking specific cytokines and controlling HLH.¹

Interferonopathies include Singleton-Merten syndrome (with IFIH1 or DDX58a gene disorder) and STING-associated vasculopathy of infancy (with TMEM137 gene disorder). NF- κ B/TNF disorders include haploinsufficiency of A20 (with TNFAIP3 gene disorder), OTULIN-related autoinflammatory syndrome (with OTULIN gene disorder), Blau syndrome (with NOD2 gene disorder), TNF receptor-associated periodic syndrome (with TNFRSF1A gene disorder), and deficiency of adenosine deaminase 2 (with ADA2 gene disorder). Moreover, Janus kinase inhibitor and TNF inhibitor are the main targeted therapy for the above two kinds of monogenic autoinflammatory diseases, respectively.

CRS in autoimmune diseases. Besides autoinflammatory diseases, autoimmune diseases are another axis of immune dysfunction which mistakenly target autologous tissues as if they were foreign. Autoimmune diseases would lead to seemingly unprovoked inflammation and CRS but do not show any signs of infection or autoimmunity. Systemic lupus erythematosus (SLE), a classical kind of autoimmune disease, is characterized by exaggerated B-cell and T-cell responses and loss of immune tolerance against self-antigens, leading to variable clinical symptoms, including low-grade fever, fatigue, weight loss and other target organ manifestations, which are accompanied by elevated levels of autoimmune antibodies and a series of cytokines.¹⁰² Of note, the profiles of cytokine levels, including IL-6, IL-8, and IL-18, could be used to monitor the disease activity of SLE.^{103,104} Intervention

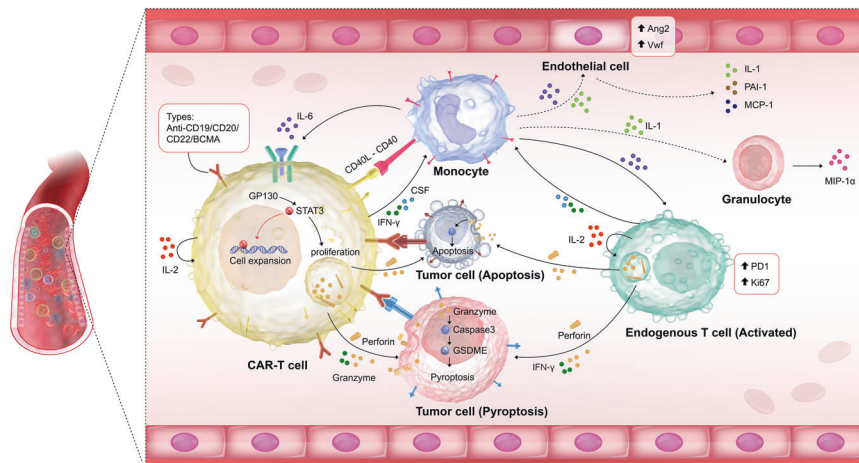


Fig. 3 Cellular mechanisms of CAR-T-cell therapy triggering CRS. IL-6 *trans*-signaling promoted the expansion and antitumor activity of CAR-T cells via the GP130/STAT3 pathway, while apoptosis and pyroptosis were found in tumor cells after CAR-T-cell therapy; monocytes, endogenous T cells, endothelial cells, and granulocyte were all activated

strategies include glucocorticoids, antimalarial agents, nonsteroidal anti-inflammatory drugs, immunosuppressive agents, and B-cell-targeting biologics.¹⁰² Furthermore, CD19-targeted CAR-T cells also have the potential to eliminate B cells and control SLE.¹⁰⁵ Other autoimmune diseases, such as autoimmune pancreatitis (AIP), psoriasis, rheumatoid arthritis, systemic vasculitis and ulcerative colitis, also show different patterns of cytokine elevation and different degrees of fever during disease progression. AIP is associated with increased production of type I IFN and is characterized by elevated IFN-I-dependent IL-33 production, which leads to fibrotic and inflammation responses.¹⁰⁶ Psoriasis, an immune-mediated chronic disease, is pathogenically driven by pro-inflammatory cytokines, in which the IL-23/IL-17A axis plays the critical important role in pathogenesis.¹⁰⁷

In summary, germline mutations in genes regulating the granule-mediated cytotoxicity, innate immune system, and inflammasome activation were the leading causes of autoinflammatory diseases and autoimmune diseases triggering CRS, while CD8 T cells and innate cells are most likely the primary driver cells, which produce and release TNF, IFN- γ , IL-1, and a combination of other cytokines to drive pathogenesis of these diseases.¹

CELLULAR MECHANISMS OF CRS

Cellular mechanisms of CRS in immunotherapy

CRS usually occurs at 1–10 days after CAR-T-cell infusion, accompanied by CAR-T-cell expansion and exhaustion. CAR-T cells and other cell subtypes are important in CAR-T-cell immunotherapy. In this section, we summarized new developments related to different cell types, levels (in vivo/in vitro levels; transcriptomic, epigenomic, and proteomic levels; and different species), dynamic changes, and potential interactions in CRS. Based on the interactions of CAR-T cells, endogenous T cells, monocytes, and endothelial cells, we proposed potential mechanisms of CRS in Fig. 3. The detailed information was provided as follows.

Characterization and dynamics of CAR-T

Xue et al. performed single-cell multiplexed proteomic profiling of CD19 CAR-T-cell products from four patients in vitro with a 16-plex cytokine microfluidics device in 2017. They revealed that CAR-T cells showed high polyfunctionality in antitumor effector and stimulatory functions.²² In 2019, Xhangolli et al. demonstrated that CAR-T-cell activation involved a mixed T helper 1/2 cell response independent of differentiation in vitro.¹⁰⁸ In their studies, both

CD4 and CD8 CAR-T cells were equally effective in directly killing target tumor cells in vitro, and cytotoxic activity was associated with elevated coproduction of a wide range of cytokines (IFN- γ , TNF- α , GM-CSF, IL-5, IL-8, and IL-13). Additionally, cytotoxic T-lymphocyte-associated protein 4 (CTLA4) was the most upregulated immune checkpoint gene upon CAR-T-cell activation, which further correlated with the upregulation of IL-10 expression in the CD4 subset and that of TGFB1 in the CD8 subset.

Furthermore, Sheih et al. showed that the clonal diversity of CAR-T cells was highest in CAR-T-cell IPs and declined following infusion in vivo.¹⁰⁹ Moreover, the clones that expanded after CAR-T infusion mainly originated from infused clusters with relatively high expression of cytotoxicity- and proliferation-related genes (CCL4, CD27, IFN- γ , GZMH, and GZMK). Furthermore, compared with the IPs, CD8 CAR-T cells at later times after infusion expressed higher proportions of inhibitory receptors (PD-1, LAG-3, TIM-3, KLRG1, TIGIT, 2B4, and CD160). At late and very late time points after CAR-T-cell infusion, the proliferative ability of CAR-T cells was decreased (decreasing gene expression of MKi67), which was accompanied by depletion of the target antigen.

Characterization and dynamics of endogenous T cells

Our group longitudinally profiled the transcriptome of T cells in a plasma cell leukemia patient treated with anti-BCMA CAR-T cells and found that CAR-T cells in the CRS peak phase transition from a proliferative to a cytotoxic intermediate state, while the expression of GZMB and PRF1 was observed in endogenous T cells at the CRS peak phase.¹¹⁰ Meanwhile, in 2020, Chen et al. found that CAR-T cells composed only 1–5% of the total T-cell population within the intact tumor microenvironment (TME) in diffuse large B-cell lymphoma patients following CAR-T-cell therapy.¹¹¹ Unexpectedly, a large number of non-CAR-T cells (endogenous T cells) within the TME were activated, and these cells were positive for Ki-67, IFN- γ , GZMB, and/or PD-1 and associated with beneficial and pathological effects.

Characterization and dynamics of monocytes and macrophages

In 2018, Norelli et al. found that monocytes accumulated concomitantly with leukemia clearance in a humanized mouse CRS model.²⁹ In detail, the counts of CD14 monocytes increased from day 1 to day 4 after CAR-T-cell infusion. Then, they started to decline and reached a minimum value at 7 days after CAR-T-cell infusion. Next, the counts of CD14 monocytes significantly increased and were maintained at a relatively high level during CRS recovery. Furthermore, monocyte ablation

protected mice from CRS. These results suggested that CRS severity correlated with monocyte counts. In addition, the critical cytokines IL-6 and IL-1 were found to be mainly produced by monocytes during CRS, while the concentration of IL-1 β in the serum increased approximately 24 h earlier than that of IL-6, and the level of IL-1 β in the serum was lower than that of IL-6 after CAR-T-cell infusion. Another study also showed a similar phenomenon for monocytes. Giavridis et al. demonstrated that CRS severity was mediated not by CAR-T-cell-derived cytokines but by IL-6, IL-1, and nitric oxide (NO) produced by recipient macrophages (especially in the mouse peritoneum).²⁷ Furthermore, they concluded that myeloid activation required proximity between CAR-T cells (CD40L) and macrophages (CD40), and indirect cytokine contact.

Characterization and dynamics of endothelial cells

In 2017, Hay et al. demonstrated that severe CRS was accompanied by endothelial activation, in which the biomarkers of endothelial activation (angiopoietin-2 and von Willebrand factor) increased significantly.³⁴ Additionally, the presence of vascular leakage and disseminated intravascular coagulation in patients with severe CRS was consistent with widespread endothelial activation.

CRS mechanisms based on the interactions of different cell types The pathways involved in severe CRS are mainly enriched in myeloid cell differentiation, cytokine production, and acute inflammatory response processes.¹¹² We speculated that the mechanisms of CRS involved the interactions of CAR-T cells, monocytes, macrophages, endothelial and tumor cells, which included direct contact and indirect interactions. In 2018, Norelli et al.²⁹ and Giavridis et al.²⁷ reported that activation of monocytes and macrophages was a major contributor to the ‘amplification’ of the inflammatory response in CAR-T-cell therapy. Direct contact between CAR-T cells and monocytes/macrophages plays an important role in the activation of monocytes/macrophages. For example, CD40-CD40LG, CD69, lymphocyte activation gene 3 (LAG3), and membrane-expressed TNF- α have been demonstrated to activate monocytes/macrophages through contact-dependent mechanisms.²⁷

On the other hand, indirect interactions (cytokine interactions) also play significant roles in CAR-T-cell-driven CRS. Among these cytokine interactions, CAR-T-cell-derived IL-6 is one of the most important initiators that amplifies the release of IL-6 from monocytes and further drives the development of severe CRS. In 2019, Jiang et al. revealed that IL-6 *trans*-signaling promoted the expansion and antitumor activity of CAR-T cells via the GP130/

STAT3 pathway.¹¹³ Furthermore, in 2020, Kang et al. found that IL-6 knockdown in CAR-T cells via shRNA technology significantly reduced IL-6 release from monocytes *in vitro*.¹¹⁴ In addition to IL-6, the IL-1 and CSF pathways also play significant roles in CAR-T-cell-driven CRS via indirect interactions. For example, GM-CSF inactivation in CAR-T cells by TALEN-mediated genetic engineering prevents monocyte-dependent release of key CRS mediators *in vitro*.²³ Then, our group showed significant interactions between highly cytotoxic CAR-T cells and endogenous T cells through the ligand–receptor pairs of CCR5-CCL5, CCL3-CCR5, CD70-CD27, and ICAM1-SPN/ITGAL.¹¹⁰ In addition to the interactions among CAR-T cells, monocytes and endogenous T cells, the crosstalk between CAR-T cells and tumors has also been studied. In 2020, Liu et al. demonstrated that CAR-T cells released GZMB, which activated caspase 3 to cleave GSDME in target tumor cells and caused pyroptosis. GSDME further leads to activation of caspase 1 and GSDMD in macrophages, which is critical for triggering CRS.¹¹⁵

Cellular mechanisms of CRS in COVID-19

Understanding immune responses at the cellular and molecular level in COVID-19 helps to elucidate the mechanisms of host responses and interpret disease pathogenesis.¹¹⁶ Circulating T and B lymphocytes decreased significantly in COVID-19 patients, particularly in severe and critical stages of COVID-19 infection.¹¹⁷ Of note, such lymphopenia in patients with severe COVID-19 frequently occurs accompanying abnormal activation of monocytes/macrophages and an increase in neutrophil numbers.¹¹⁸ Other cells including NK and endothelial cells have also been reported to play an indispensable role in COVID-19 disease. The characteristics and comparisons of cytokine and cell types between mild and severe patients with COVID-19 were also shown in Fig. 4.

Characterization and dynamics of T cells

Sekine et al. demonstrated that SARS-CoV-2-specific T cells were important for long-term immune protection against COVID-19,¹¹⁹ in which acute-phase SARS-CoV-2-specific T cells were highly activated and correlated with various clinical markers of disease severity, whereas convalescent-phase T cells were polyfunctional and displayed a stem-like memory phenotype. On the other hand, Zhu L et al. observed XIAP-associated factor 1 (XAF1)-, TNF-, and FAS-induced T cell apoptosis in COVID-19 patients.¹²⁰

CD8 T resident memory and CD4 T helper 17 cells in mild COVID-19 undergo active expansion towards the end of the trajectory and are characterized by good effector functions, while in critical COVID-19 these remain more naive.¹²¹ Also, Meckiff et al. analyzed CD4 T cells from 40 COVID-19 patients,

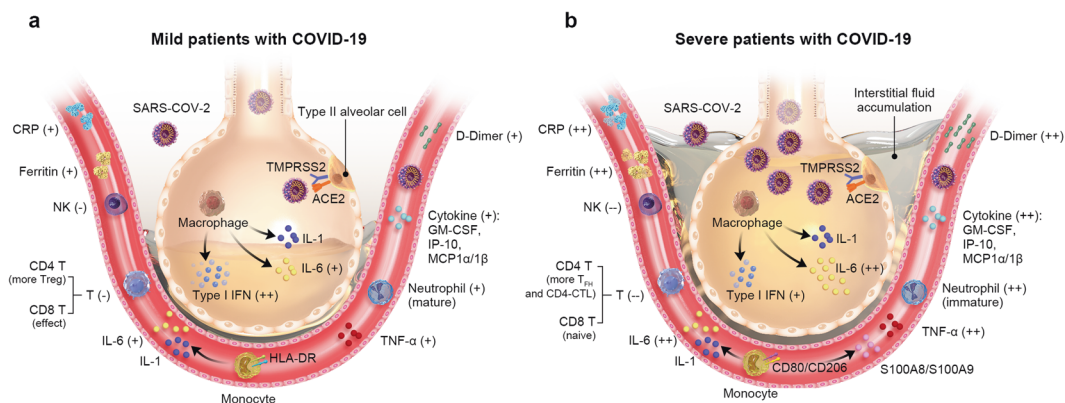


Fig. 4 Cellular mechanisms of CRS in mild and severe patients with COVID-19. Mild patients have more NK and T cells in the peripheral blood (a), while severe patients are exposed to more COVID-19 and have more interstitial fluid accumulation (b). Neutrophil, C-reactive protein (CRP), ferritin, IL-6, TNF- α , D-dimer, and other cytokine in severe patients with COVID-19 were higher than that in mild patients

and showed that hospitalization (severe patients with COVID-19) is associated with increased cytotoxic follicular helper cells and cytotoxic T helper cells and a reduction in regulatory T cells.¹²² Moreover, in severe patients, the immune landscape featured a deranged IFN response, profound immune exhaustion with skewed T-cell receptor repertoire, and broad T cell expansion. Intensive expansion of highly cytotoxic effector T-cell subsets, such as granulysin (GNLY)⁺ CD4⁺ effector T cells, GNLY⁺CD8⁺ effector T cells and CD160 NKT cells, was associated with convalescence in moderate patients.¹²³

Characterization and dynamics of NK cells

It was reported that NK cells in the peripheral blood (PB) decreased in patients infected with SARS-CoV-2, especially in severe cases.¹²⁴ However, another detached study showed no difference in the number of CD16⁺CD56⁺ NK cells in mild versus severe cases.¹²⁵ On the other hand, Maucourant et al. illustrated a detailed NK cell activation landscape in bronchoalveolar lavage from COVID-19 patients¹²⁶ and identified the hallmarks of these immunotypes, including perforin, NKG2C, and KSP37. They also observed arming of CD56 bright NK cells was driven by a defined protein-protein interaction network of inflammatory soluble factors in COVID-19.¹²⁶

Characterization and dynamics of monocytes and macrophages Compared with normal healthy individuals, monocytes in PB from COVID-19 patients show an activated phenotype and secrete more IL-6, IL-10, and TNF- α ,¹²⁷ while the number of monocytes shows no significant difference. These activated monocytes in COVID-19 patients show features of mixed M1/M2 macrophage polarization, which is particularly associated with disease severity and poor prognosis. Besides, a large number of CD14⁺IL-1 β ⁺-activated monocytes were increased in COVID-19 patients.¹¹⁷ Furthermore, it was predicted that TNFSF13, IL-18, IL-2, and IL-4 may help to recover COVID-19 patients. Also, Guo Chuang et al. revealed a monocyte-associated CRS by single-cell sequencing.¹²⁸ Lee et al. found that type I IFN response co-existed with the TNF/IL-1 β -driven inflammation in classical monocytes from patients with severe COVID-19, while this was not seen in patients with milder COVID-19.¹²⁹

Monocyte-to-macrophage trajectories showed that chronic hyperinflammatory monocytes accumulated, while anti-inflammatory and antigen-presenting macrophages in alveoli were depleted in critical COVID-19. Besides, monocytes may lead to fibrosis and worsen disease severity by ATP purinergic signaling.

Characterization and dynamics of neutrophils

Neutrophils in circulation gradually increase as COVID-19 progresses and play a vital role in pathophysiology, particularly in patients with severe disease courses.¹³⁰ In severe COVID-19 patients, dysfunctional neutrophils were associated with emergency myelopoiesis.¹³¹ Guo Qirui et al. found that coronavirus infection induced pathological damage by producing a certain group of neutrophils.¹³² Zhang et al.¹³³ reported that the neutrophil-to-lymphocyte ratio (NLR) combined with IgG also helped to predict the severity of COVID-19.

Moreover, some studies have explored the function of neutrophil extracellular traps (NETs). Middleton et al. revealed that COVID-19 plasma would trigger NET formation by neutrophils and may lead to prothrombotic clinical presentations in COVID-19 patients.¹³⁴ In addition, Barnes et al. also showed lung infiltration of neutrophils in an autopsy specimen from a COVID-19 patient and proposed that the ability to form NETs may contribute to organ damage and mortality in COVID-19.¹³⁵

In conclusion, compared with mild patients, severe patients with COVID-19 are affected by higher load of SARS-CoV-2 at first, through the viral receptor ACE2 for entry into the cell and

the serine protease TMPRSS2 for viral spike protein priming. Secondly, the pulmonary interstitial fluid accumulation and low type I IFN were observed in severe patients when compared with mild patients. Then, the cytokines (including CRP, Ferritin, IL-6, TNF- α , GM-CSF, IP-10, and MCP1 α /1 β) in the serum of severe patients with COVID-19 were higher than that in the mild patients with COVID-19, while the number of T and NK cells decreased and the number of neutrophils increased. In addition, the CD8 T cells and neutrophils in severe patients were of naive state, while these in mild patients were effective and mature. Finally, CD80/CD206 monocyte and their release factors S100A8 and S100A9 are positively associated with the severity of COVID-19, while HLA-DR monocyte is negatively associated with the severity of COVID-19.¹³⁶

Molecular mechanisms of CRS

According to the important cytokines of CRS mentioned above, the present review illustrated the possible molecular and cellular mechanisms of CRS from several key cytokine-triggered signaling pathways, including TNF/NF- κ B, IL-1/NF- κ B, IL-6/JAK-STAT, and IFN- γ /JAK/STAT signaling pathways.

TNF/NF- κ B signaling pathways

TNF- α is a well-known pro-inflammatory cytokine, mainly produced by monocytes and T cells and is involved in early inflammatory events. As an initial driver of nuclear factor κ B (NF- κ B) signaling pathway, TNF- α can induce the expression of a variety of anti-apoptotic and pro-inflammatory genes by activating its receptor TNFR1 and intermediate adapters. NF- κ B plays critical roles in the immune system, particularly in regulating the expression of some inflammation-related cytokines. Hence, NF- κ B in turn can induce TNF- α expression.¹³⁷

TNF- α is closely associated with iatrogenic diseases, pathogenic diseases, and monogenic and autoimmune diseases. TNF- α blockade (adalimumab) exerted separated and synergistic effects on preventing endothelial activation induced by CAR-T, myeloid cells, and tumor cells.¹³⁸ Besides, adalimumab prevents macrophage activation and effectively mitigates CRS in anti-CD3 bispecific antibody therapy.⁶¹ Furthermore, in SARS-CoV and MERS cases, excessive TNF- α represented a poor prognosis and inhibition of NF- κ B can increase survival in SARS-CoV-infected mice.¹³⁹ Moreover, deregulation of TNF- α characterizes many autoimmune diseases.¹⁴⁰ Therefore, TNF α /NF- κ B signaling may play pathological roles in the initiation of CRS and the over-activation of immune cells during CRS.

IL-1/NF- κ B signaling pathways

IL-1 family cytokines play a vital role in immune systems and mediate inflammation in response to various stimuli. From the cellular level, IL-1 is mainly secreted by monocytes, macrophages and dendritic cells, which were encoded by IL1A and IL1B.^{29,141} Then, IL-1 β from activated myeloid cells further induced endothelial activation and neutrophil production. On the other hand, the basic molecular mechanism of IL-1/NF- κ B signal initiation is a stepwise process: (1) IL-1 β was formed from inactivate IL-1 β precursors by cleavage of NLRP3 inflammasomes. (2) IL-1 α / β bind to the same IL-1 receptor, activating a cascade of intracellular NF- κ B.

In CAR-T therapy, CRS severity correlated with monocyte/macrophage counts and critical cytokines IL-1 and IL-6 were mainly produced by monocytes during CRS. IL-1 β may also contribute to CRS in coronavirus infections.¹⁴² In many autoinflammatory disorders, IL-1 is the critical player of the innate immune response and inflammation.¹⁴³ Anti-IL1 β antibody exerted separated and synergistic effects in preventing endothelial activation induced by tumor cells, CAR-T, and myeloid cells.¹³⁸ In severe COVID-19, it was proposed that NLRP3 activation and IL-1 β precursor cleavage potentially aggravate CRS.¹⁴⁴

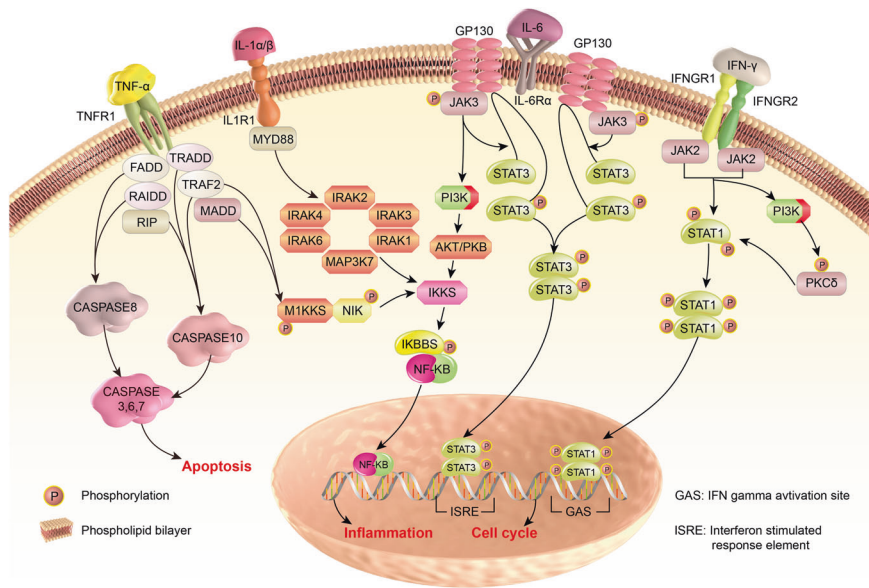


Fig. 5 Molecular mechanisms of CRS from TNF/NF-κB, IL-1/NF-κB, IL-6/JAK-STAT, and INF-γ/JAK-STAT pathway

IL-6/JAK-STAT signaling pathways

IL-6 is an inflammation-related cytokines and activates the JAK/STAT3 pathway by deploying *cis*-signaling and *trans*-signaling. In *cis*-signaling, IL-6 binds to membrane-bound IL-6R that was restrictedly expressed on immune cells, and then combined with gp130 and activated downstream JAK/STAT3 and other signaling pathways. In *trans*-signaling, circulating IL-6 firstly binds to soluble IL-6Rs and then binds with the gp130 dimer which was expressed on almost all cell subpopulation. This *trans*-signaling of the JAK-STAT3 pathway expanded inflammation from immune cells to cells without mIL-6R expression, such as endothelial cells and vascular smooth muscle cells. The crushing activation of the IL-6-IL-6R/JAK-STAT3 signaling results in systemic hyperinflammation and provokes the secretion of various mediators.¹⁴⁵ These results may further contribute to vascular hyperpermeability, leakiness, hypotension, and pulmonary dysfunction during CRS.

IL-6 has been shown to be markedly elevated in patients with multiple CRS. Besides CAR-T-induced CRS in adults and children,^{29,146} many studies also showed that the serum of IL-6 increased remarkably in patients with COVID-19.¹⁴⁷⁻¹⁴⁹ Besides, targeting IL-6 pathway has led to innovative therapeutic approaches for various rheumatic diseases and related therapy is on the way.

INF-γ/JAK/STAT signaling pathways

INF-γ is a potent activator of macrophages, mainly secreted by NK cells and activated T cells.¹⁵⁰ INF-γ is involved in inflammation and other immunological processes, which plays a major role in protective immunity against bacterial and viral infections by activating JAK1/STAT cascades. More specifically, INF-γ activates JAK/STAT-associated signaling pathway by binding to its two receptor-subtypes (INF-γ receptor (IFNGR)-1 and -2), and achieve transcriptional activation of INF-γ-inducible genes. The major STAT protein activated by INF-γ is STAT1 and the INF-γ-inducible genes are IFN regulatory factor (IRF)-1, CXCL10, CXCL11, and IFIT2.^{151,152}

Different studies have demonstrated that INF-γ is highly involved in a variety of CRS-related diseases. In CAR-T therapy, the production of INF-γ was mediated by myeloid-derived macrophages and plays a critical role in CRS pathogenesis.¹⁵³ Elevated levels of the above core cytokines have been confirmed in patients in clinical¹⁴⁶ and in murine models.³² In primary HLH,

large quantities of INF-γ were produced and excessive T cells were activated.¹⁵⁴ In COVID-19, numerous studies have reported elevated levels of INF-γ in patients, which may mainly from macrophages, not T cells.

To further strengthen the understanding of CRS, the molecular mechanisms of CRS were summarized from TNF/NF-κB, IL-1/NF-κB, IL-6/JAK-STAT, and INF-γ/JAK-STAT pathways and shown in Fig. 5.

CRS MODELS

CRS models in CAR-T-cell immunotherapy

CRS models are essential for the analysis of CRS mechanisms. In this section, we summarize the detailed methods of CRS mouse model studies, including mouse models (mouse strain, hematopoietic stem cells (HSCs), leukemia cells, CAR-T cells, and injection methods), cytokine detection panels, time points, and weight and fever information.

In the severe combined immunodeficiency (SCID)-beige mouse model, Sterner et al. found that GM-CSF inhibition not only reduced CRS severity but also enhanced CAR-T-cell functions in xenografts.²⁴ Furthermore, compared with CD19 CAR-T cells, GM-CSF-deficient CD19 CAR-T cells maintained normal functions and enhanced antitumor activity and improved overall survival. In another mouse CRS model study, Mestermann et al. demonstrated that a short treatment course of dasatinib protected a subset of mice from lethal CRS, indicating that this treatment may be a broadly applicable pharmacologic on/off switch for CAR-T cells.¹⁵⁵

The mouse myeloid system (monocytes and macrophages) in the SCID-beige mouse model is not well matched with human CAR-T cells killing leukemia cells. Therefore, the humanized NSG mouse model is more widely used in different studies.^{29,156,157} In 2017, Diaconu et al. demonstrated that in a humanized mouse model, the inducible caspase-9 (iC9) safety switch could eliminate CD19 CAR-T cells in a dose-dependent manner, allowing either selective containment of CD19 CAR-T-cell expansion in the case of CRS or complete deletion on demand to allow normal B-cell reconstitution.¹⁵⁶ In 2018, by using a humanized NSG mouse model, Norelli et al. demonstrated that CRS was characterized by high fever and elevated IL-6 levels and that the major source of IL-1 and IL-6 during CRS is monocytes.²⁹ In addition, in 2020, Sun et al. revealed that the kinase LCK and phosphatase SHP1 can be engineered in both CD28 and 4-1BB CAR-T cells to tune CAR-T-cell

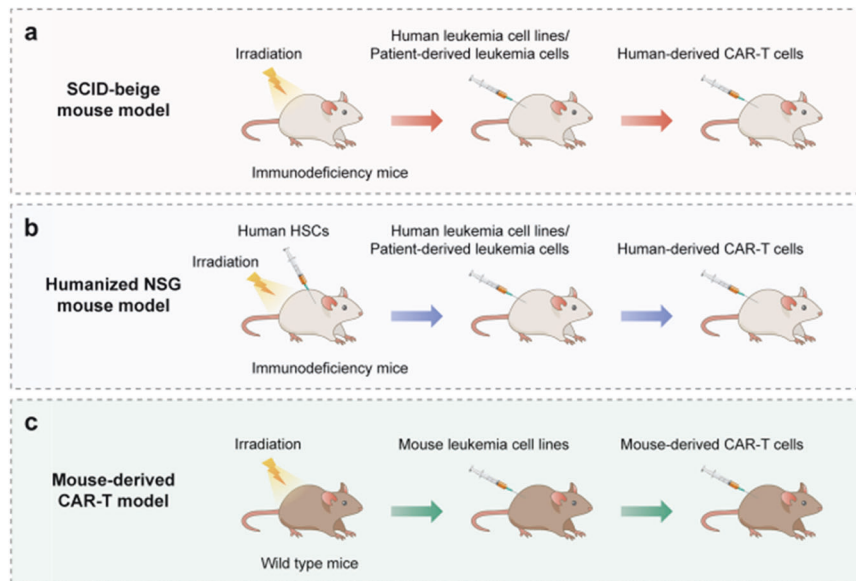


Fig. 6 CRS mouse models. **a** Severe combined immunodeficiency (SCID)-beige mouse model. **b** Humanized NSG mouse model. **c** Mouse-derived CAR-T model

activity by affecting the magnitude of CAR-T-cell activation. In their humanized mouse model, SHP1 recruitment induced by the transient administration of AP21967 alleviated weight loss and significantly reduced the release of human IFN- γ , IL-6, GM-CSF, and TNF- α into the plasma.¹⁵⁷

In addition to the above models, there is another mouse-derived CAR-T model in which mouse CAR-T cells recognize mouse leukemia cells in the mouse microenvironment, which would be beneficial for exploration of CRS mechanisms (Fig. 6).

CRS model in COVID-19

Animal models of SARS-CoV-2 infection are important and urgently needed to improve research and drug/vaccine development. In 2020, the Qin Chua's group established animal models of SARS-CoV-2 in transgenic mice that express human ACE2 (hACE2 mice). Weight loss, virus replication, viral antigens, and interstitial pneumonia were observed in hACE2 mice after SARS-CoV-2 infection, while these phenomena were not found in wild-type mice infected with SARS-CoV-2.¹⁵⁸ In addition, the Shi Zheng-Li's group developed a mouse model with symptoms similar to those of patients with COVID-19 by transgenic technology (HFH4-hACE2 in C3B6 mice).¹⁵⁹ In the HFH4-hACE2 COVID-19 model, HFH4 was used to control human ACE2 expression. In addition to ACE2, other genes (such as TMRPSS2 and FcyRT) are also critical for COVID-19 infection. Therefore, multiple humanized COVID-19 models will be a direction for future study.

CUTTING-EDGE TECHNOLOGIES IN CRS-RELATED STUDIES

Single-cell technologies

Single-cell RNA sequencing. Single-cell RNA sequencing, especially 10 \times genomic single-cell transcriptome sequencing, has become a powerful tool for revealing the dynamic changes in various subpopulations and the source of key cytokine both in the CAR-T-cell therapy and COVID-19.^{129,136,160–162} The sample types and time points of the above single-cell studies on CAR-T- and COVID-19-induced CRS were summarized in Table 4. Single-cell studies of CAR-T mainly focus on the changes of CAR-T cells, while the single-cell studies of COVID-19 mainly focus on the comparison of severe patients, mild patients, and healthy donors among different tissues, especially the lung and PB.

In addition to the above single-cell studies, Boroughs et al.¹⁶³ profiled the transcriptome of CAR-T cells in vitro and found that

the differentially expressed genes of 4-1BBz CAR CD8 T cells were enriched in central memory cell phenotype and fatty acid metabolism genes compared to that of 28z CAR-T cells. Meanwhile, Deng et al. performed single-cell RNA sequencing of anti-CD19 CAR-T-cell IPs in 24 patients with LBCL. Apart from finding that a rare cell population with monocyte-like transcriptional features in CAR-T products was associated with high-grade ICANS, they also observed that high-grade CRS was negatively correlated with exhausted CD8 T cells and positively correlated with exhausted CD4 T cells.¹⁶⁴

Single-cell TCR and ATAC sequencing. In addition to scRNA-seq, single-cell T-cell receptor (TCR) sequencing and single-cell assays for transposase-accessible chromatin using sequencing (scATAC-seq) have also been employed in studies of CAR-T-cell expansion. Sheih et al.¹⁰⁹ described the alterations of clonal diversity of CAR-T cells during the course of the disease by single-cell TCR sequencing. To be specific, the clonal diversity of CAR-T cells was highest in the IPs and declined following infusion. For COVID-19, Nguyen et al. analyzed ex vivo CD8⁺ T cells specific for COVID-19 epitopes and found that B7/N105+CD8⁺ T cells were seen in high numbers during COVID19 and persisted for a long time, by the integrative analysis of high naive frequency and TCR plasticity.¹⁶⁵

Using scATAC-seq, Wang et al. showed that the precise location of the CAR vector in the patient genome may play an essential role in treatment outcomes.¹⁶⁶ Combined with bulk and single-cell ATAC sequencing, Chen et al. found that IRF7 could affect CAR-T-cell persistence across T-cell subsets by regulating chronic IFN signaling, and that the TCF7 regulon is maintained in effector T cells among patients with long-term CAR-T-cell persistence.¹⁶⁷ The dynamic epigenetic changes in various cell types during the CRS process are also worthy of further study.

Single-cell multiplexed cytokine assay. A multiplexed single-cell cytokine assay method has been employed to identify and predict CRS biomarkers after CAR-T-cell therapy. In 2016, Teachey et al. found that the expression levels of 24 cytokines were highly associated with different CRS grades in CAR-T-cell therapy by using the above technology. Furthermore, prediction models based on IFN- γ /sgp130/sIL-1RA expression levels could accurately predict which patients would develop severe CRS.¹⁶⁸ For COVID-19, Mathew et al. analyzed 125 COVID-19 patients at the protein

Table 4. Single-cell studies on CAR-T-cell therapy and COVID-19

Cell type	Sample/Time	Conclusions
CAR-T-cell therapy		
CAR-T cells	CRS initiation stage	CD8 41-BBz CAR-T enriched in central memory cell phenotype and fatty acid metabolism genes (Boroughs et al., 2020) ¹⁶³
CAR-T cells	CRS initiation stage	CD4 helper and CD8 cytotoxic CAR-T cells are equally effective in directly killing tumor cells (Xhangolli et al., 2019) ¹⁰⁸
Leukocyte	CRS peak stage	High fever and elevated IL-6 levels are hallmarks of CRS; Human monocytes are the major source of IL-1 and IL-6 during CRS (Norelli et al., 2018) ²⁹
CAR-T and endogenous T	CRS peak stage	CAR-T cells in the CRS peak phase transition from a proliferative to a cytotoxic intermediate state; Endogenous T presents an active state in the CRS peak stage (Li et al., 2021) ¹¹⁰
CAR-T cells	CRS recovery stage	Clonal diversity of CAR-T cells is highest in products and declines following infusion; Amplified oligoclonal at last mainly originate from CAR-T products with high expression of cytotoxicity and proliferation genes (Sheih et al., 2020) ¹⁰⁹
CAR-T cells	CRS recovery stage	Heterogeneity of CAR-T cells contributes to variations in efficacy and toxicity in LBCL (Deng et al., 2020) ¹⁶⁴
COVID-19		
PBMC	76 COVID-19 patients and 69 healthy donors	Enhanced plasma levels of inflammatory mediators, including EN-RAGE, TNFSF14, and oncostatin M, were correlated with COVID-19 severity (Arunachalam et al., 2020) ¹⁶⁰
PBMC	3 COVID-19 patients and 3 healthy donors	Immature neutrophil and nonclassical monocyte pools, with levels of the protein calprotectin linked to disease severity (Silvin et al., 2020) ¹⁶¹
PBMC	9 COVID-19 patients, 5 influenza patients and 4 healthy donors	TNF/IL-1 β -driven inflammation was defining characteristics of COVID-19. The type I IFN response plays a pivotal role in exacerbating inflammation in severe COVID-19 (Lee et al., 2020) ¹²⁹
PBMC	40 COVID-19 patients and 13 healthy donors	Hospitalization is associated with increased cytotoxic follicular helper cells and cytotoxic T helper cells and a reduction in regulatory T cells (Meckiff et al., 2020) ¹²²
BALF	6 Severe- and 3 mild- COVID-19 patients	Dramatic differences between the mild and severe COVID-19 patients, including an inflammatory signature. SARS-CoV-2 infects epithelial cells and alters immune landscape in severe patients (Bost et al., 2020) ¹²³
PBMC	5 COVID-19 patients, 2 influenza virus patients and 2 healthy donors	COVID-19 patient features XAF1-, TNF-, and FAS-induced T cell apoptosis. COVID-19 activates distinct pathway (STAT1/IRF3) versus influenza, and substantial differences were revealed like expression of interleukin (IL)6R and IL6ST (Zhu L et al., 2020) ¹²⁰
PBMC	2 COVID-19 patients and 2 healthy donors	A monocyte subpopulation contributes to the inflammatory CRS and tocilizumab treatment can attenuate the CRS in COVID-19 patients (Guo et al., 2020) ¹²⁸
PBMC and whole blood	53 COVID-19 patients and 56 healthy donors	HLA-DR ^{hi} CD11c ^{hi} inflammatory monocytes with an IFN-stimulated gene signature were elevated in mild COVID-19. Severe COVID-19 was marked by the occurrence of neutrophil precursors (Schulte-Schrepping et al., 2020) ¹³¹
PBMC	13 COVID-19 patients and 5 healthy donors	COVID-19 showed a strong IFN- α response and an overall acute inflammatory response. In severe patients, the immune landscape featured a deranged interferon response (Zhang et al., 2020) ¹³³
PBMC	7 Severe COVID-19 patients and 6 healthy donors	Severe disease has been associated with changes in peripheral immune activity, including increased levels of pro-inflammatory cytokines, while peripheral monocytes and lymphocytes do not express substantial amounts of pro-inflammatory cytokines (Wilk et al., 2020) ¹⁶²
PBMC, BALF/ PFMC, sputum	196 COVID-19 patients and 25 healthy donors	Systemic upregulation of S100A8/A9, mainly from megakaryocytes and monocytes in the peripheral blood, may contribute to the CRS frequently in severe patients (Ren et al., 2021) ¹³⁶

level by high-dimensional flow cytometry and identified three prominent and distinct immunotypes that are related to disease severity and clinical parameters.¹⁶⁹ Among them, immunotypes with robust activated CD4 T cells, activated CD8 T cells, and plasmablasts were associated with disease severity of COVID-19.

Other cutting-edge technologies. Gene editing is another new advanced technology and the introduction of gene editing into CAR-T cells would be a new direction for preventing severe CRS.

Kang et al. found that IL-6 knockdown of CAR-T cells significantly reduced IL-6 release from monocytes in vitro.¹¹⁴ Drug-screening is also an effective technology to temporarily switch off CAR-T cells. In a mouse model of CRS, Mestermann et al. found that dasatinib halted cytokine production and proliferation of CAR-T cells and protected mice from otherwise fatal CRS, while the inhibitory effect was rapidly and completely reversed upon discontinuation of dasatinib.¹⁵⁵ In addition, Sun et al. found that SHP1 recruitment induced by the transient administration of AP21967 significantly

reduced the release of human IFN- γ , IL-6, GM-CSF, and TNF- α in the plasma of a humanized mouse model.¹⁵⁷ At last, novel types of CAR are also a direction to fundamentally prevent CRS. For example, CAR NK cells can induce responses in patients with r/CD19-positive cancers without severe CRS.¹⁷⁰

Management of CRS by intervention targeting CRS-related pathways. CRS management is essential for CAR-T-cell therapy and patients with COVID-19 and monogenic and autoimmune diseases. Here, we summarized drugs targeting specific cytokines and overlapping signaling pathways for intervening.

Drugs targeting TNF/NF- κ B signaling pathways. TNF, a key upstream factor mediating monocyte activation, can result in inflammation, fever, and cellular proliferation. The TNF-TNF receptor superfamily can lead to the expression of multiple pro-inflammatory genes by inducing NF- κ B. In anti-CD3 bispecific antibody therapy, adalimumab (anti-TNF- α drug) prevents macrophage activation and effectively mitigates CRS.⁶¹ Furthermore, the effect of adalimumab on COVID-19 patients has also been investigated.¹⁷¹

Drugs targeting IL-1/NF- κ B signaling pathways. IL-1 activates a cascade involving intracellular nuclear factor κ B (NF- κ B) and other significant pathways. The IL-1R antagonist anakinra is effective for the treatment of CRS in CAR-T-cell therapy,²⁹ secondary HLH,¹⁷² and COVID-19.^{173,174}

Drugs targeting IL-6/JAK-STAT signaling pathways. IL-6 plays a vital role in acute inflammatory response and CRS and is highly elevated in CAR-T-cell therapy patients, COVID-19 patients, and monogenic and autoimmune diseases.¹⁷⁵ Targeted IL-6 receptor drug (tocilizumab) and neutralizing IL-6 drug (siltuximab) have already been in clinical use to treat CRS disorders, including HLH- and CAR-T-cell-induced CRS.¹⁴⁶ In addition, in severe and critical COVID-19 patients, the Wei Haiming's group suggested that tocilizumab improved clinical outcomes immediately and was an effective treatment to reduce mortality.¹⁷⁶ However, in moderately ill patients with COVID-19, tocilizumab was not effective for preventing mechanical ventilation or death.¹⁷⁷

Drugs targeting IFN- γ /JAK/STAT signaling pathways. IFN- γ , a potent activator of macrophages is primarily secreted by activated T and NK cells. The US FDA approved anti-IFN- γ antibody emapalumab have been used to treat patients with relapsed/refractory HLH, based on favorable results in a single-arm, open-label Phase II/III trial (NCT01818492; NCT02069899).¹⁷⁸ In addition, neutralizing antibodies against IFN- γ improved animal survival in a mouse model of LPS-induced sepsis.¹⁷⁹ Both interventions in animal models and clinical trials of therapies targeting the type I IFN signaling pathway have shown efficacy in the treatment of autoimmune diseases, such as SLE, rheumatoid arthritis, juvenile idiopathic arthritis, juvenile dermatomyositis, and systemic sclerosis.¹⁸⁰ Furthermore, anti-IFN- γ antibody treatment improved survival and reduced pro-inflammatory cytokines in experimental model of macrophage activation syndrome associated with juvenile idiopathic arthritis.¹⁸¹

In summary, the most common clinical management approach for CRS after CAR-T-cell therapy is supportive care. When severe CRS occurs, drugs such as tocilizumab, siltuximab, or corticosteroids are always used. In CAR-T-cell therapy, anti-IL-6 drug therapy is recommended to treat hypotension and hypoxia for patients with a higher grade of CRS.³² When patients are refractory to anti-IL-6 therapy, the usage of corticosteroids^{182,183} can be considered to inhibit T cell activation by downregulating signal transduction through the IL-2 receptor. For the management of CRS in COVID-19, except for the above methods, some specific interventions or targets were provided, including early type I IFN response.⁶⁷ Preliminary reports showed that IFN alpha-2b

combined with LPV/r accelerated viral clearance in COVID-19 patients and shortened the length of hospitalization.¹⁸⁴ In addition, paquinimod, a specific inhibitor of S100A8/A9, could significantly reduce viral load and the activation of aberrant neutrophils in SARS-CoV-2-infected mice, and rescue pneumonia.¹³² At last, as patients treated with CAR-T cell are intrinsically more susceptible to SARS-CoV-2 infection, our group recommend that high-dose steroids should be avoided because they could worsen CAR-T-cell therapy outcomes and disrupt CAR-T-cell persistence. Furthermore, tocilizumab or artificial liver treatment is recommended for severe COVID-19 patients with elevated cytokine levels.¹⁸⁵

Perspectives. A subset of patients receiving CAR-T/antibodies or with COVID-19, monogenic and autoimmune diseases display severe CRS. In the present review, we summarized the clinical features, risk factors, cellular and molecular mechanisms, and management of CRS in the above human diseases, as well as CRS models and cutting-edge technologies in CRS studies.

The elevated serum cytokines in the patients with COVID-19, monogenic and autoimmune diseases or CAR-T/antibody treatment were similar (including IL-6, TNF, IFN- γ , CRP, and D-dimer), while signaling pathways involved in the regulation of CRS also overlapped (such as IL-1/NF- κ B, TNF/NF- κ B, IL-6/JAK-STAT, and INF- γ /JAK-STAT signaling pathway). In addition, the differences in CRS among phenotype and etiologies should also be noted. For example, lymphopenia (low blood lymphocyte count) is one of the key characteristics of severe COVID-19 patients,¹⁸⁶ while lymphocytes (including CAR-T and endogenous T cells) are activated and increase in number in CAR-T-cell therapy.¹¹⁰ Our group also compared the clinical characteristics and risk factors associated with severe CRS induced by COVID-19 and CAR-T-cell therapy.¹⁸⁷ We found a significantly higher incidence of grade 4 CRS in the COVID-19 group than in the CAR-T-cell therapy group. In addition, COVID-19 group showed increased TNF- α level and decreased IL-2, IL-6, IL-10, and IFN- γ levels.

Intervention therapy for CRS is also an important issue. Several strategies to diminish undesirable CRS were summarized: (a) drug intervention, including multitarget drugs, targeted mAb drugs, targeted monocyte-clearing drugs (liposomal clodronate,²⁹) and small molecules; (b) the structure of the CAR-T vector, including new structure of CAR-T vector,¹⁸⁸ novel CAR-NK cells, CAR-macrophage cells¹⁸⁹ and dual-target CAR-T cells, and their infusion strategy and dose; (c) gene editing of CAR-T cells, including IL-6^{114,190} and GM-CSF²⁴ knockout and naive memory-related gene (LPR6 targeting¹⁹¹) overexpression; and (d) low-molecular-weight adapters.¹⁹²

In addition to CRS management, some unclear issues related to CRS and future directions are also summarized in the present review. For example, the crosstalk of endothelial cells, granulocytes, and CAR-T cells is worthy of further verification and exploration. Meanwhile, transcriptome-level data alone cannot solve all the problems of CAR-T-cell expansion and exhaustion issues.¹⁹³ Therefore, in the future, single-cell multiomics and other new cutting-edge technologies, including CAR-T-cell integration site analysis, scRNA-seq, scATAC-seq, CyTOF and imaging technology, should be combined for the CRS studies.

CRS is a double-edged sword, which facilitates disease control and may be harmful to the host. Our goal is to illustrate the framework, characteristics and mechanism of CRS in human diseases and achieve both favorable efficacy and low toxicity.

FUNDING

We are grateful for grants from multiple sources: National Natural Science Foundation of China (81900176, 81730008, 81870080, and 91949115), National Key R&D Program

of China, Stem Cell and Translation Research (2018YFA0109300), Zhejiang Provincial Key Research and Development Program (2018C03016-2, 2019C03016), Zhejiang Province Science Foundation for Distinguished Young Scholars (LR19H080001).

ADDITIONAL INFORMATION

Competing interests: The authors declare no competing interests.

REFERENCES

- Fajgenbaum, D. C. & June, C. H. Cytokine Storm. *N. Engl. J. Med.* **383**, 2255–2273 (2020).
- Ferrara, J. L., Abhyankar, S. & Gilliland, D. G. Cytokine storm of graft-versus-host disease: a critical effector role for interleukin-1. *Transplant. Proc.* **25**, 1216–1217 (1993).
- Tejaro, J. R. et al. Endothelial cells are central orchestrators of cytokine amplification during influenza virus infection. *Cell* **146**, 980–991 (2011).
- Pedersen, S. F. & Ho, Y. C. SARS-CoV-2: a storm is raging. *J. Clin. Invest.* **130**, 2202–2205 (2020).
- Grupp, S. A. et al. Chimeric antigen receptor-modified T cells for acute lymphoid leukemia. *N. Engl. J. Med.* **368**, 1509–1518 (2013).
- Maude, S. L. et al. Chimeric antigen receptor T cells for sustained remissions in leukemia. *N. Engl. J. Med.* **371**, 1507–1517 (2014).
- Schuster, S. J. et al. Chimeric antigen receptor T cells in refractory B-cell lymphomas. *N. Engl. J. Med.* **377**, 2545–2554 (2017).
- Schuster, S. J. et al. Tisagenlecleucel in adult relapsed or refractory diffuse large B-cell lymphoma. *N. Engl. J. Med.* **380**, 45–56 (2019).
- Neelapu, S. S. et al. Axicabtagene ciloleucel CAR T-cell therapy in refractory large B-cell lymphoma. *N. Engl. J. Med.* **377**, 2531–2544 (2017).
- Johnson, L. A. & June, C. H. Driving gene-engineered T cell immunotherapy of cancer. *Cell Res.* **27**, 38–58 (2017).
- Titov, A. et al. Adoptive Immunotherapy beyond CAR T-Cells. *Cancers* **13**, 743 (2021).
- Morgan, R. A. et al. Case report of a serious adverse event following the administration of T cells transduced with a chimeric antigen receptor recognizing ERBB2. *Mol. Ther.* **18**, 843–851 (2010).
- Moore, J. B. & June, C.H. Cytokine release syndrome in severe COVID-19. *Science* **368**, 473–474 (2020).
- Mehta, P. et al. COVID-19: consider cytokine storm syndromes and immunosuppression. *Lancet.* **395**, 1033–1034 (2020).
- Wang, M. et al. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Res.* **30**, 269–271 (2020).
- Zhang, M. et al. A heterozygous RAB27A mutation associated with delayed cytolytic granule polarization and hemophagocytic lymphohistiocytosis. *J. Immunol.* **196**, 2492–2503 (2016).
- Kaufman, K. M. et al. Whole-exome sequencing reveals overlap between macrophage activation syndrome in systemic juvenile idiopathic arthritis and familial hemophagocytic lymphohistiocytosis. *Arthritis Rheumatol.* **66**, 3486–3495 (2014).
- Ma, X. et al. Interleukin-23 engineering improves CAR T cell function in solid tumors. *Nat. Biotechnol.* **38**, 448–459 (2020).
- Gust, J. et al. Glial injury in neurotoxicity after pediatric CD19-directed chimeric antigen receptor T cell therapy. *Ann. Neurol.* **86**, 42–54 (2019).
- Zhang, Z. et al. Modified CAR T cells targeting membrane-proximal epitope of mesothelin enhances the antitumor function against large solid tumor. *Cell Death Dis.* **10**, 476 (2019).
- Zhang, E. et al. Accurate control of dual-receptor-engineered T cell activity through a bifunctional anti-angiogenic peptide. *J. Hematol. Oncol.* **11**, 44 (2018).
- Xue, Q. et al. Single-cell multiplexed cytokine profiling of CD19 CAR-T cells reveals a diverse landscape of polyfunctional antigen-specific response. *J. Immunother. Cancer* **5**, 85 (2017).
- Sachdeva, M. et al. Granulocyte-macrophage colony-stimulating factor inactivation in CAR T-cells prevents monocyte-dependent release of key cytokine release syndrome mediators. *J. Biol. Chem.* **294**, 5430–5437 (2019).
- Sterner, R. M. et al. GM-CSF inhibition reduces cytokine release syndrome and neuroinflammation but enhances CAR-T cell function in xenografts. *Blood* **133**, 697–709 (2019).
- Golumba-Nagy, V., Kuehle, J., Hombach, A. A. & Abken, H. CD28-ζ CAR T cells resist TGF-β repression through IL-2 signaling, which can be mimicked by an engineered IL-7 autocrine loop. *Mol. Ther.* **26**, 2218–2230 (2018).
- Hirayama, A. V. et al. The response to lymphodepletion impacts PFS in patients with aggressive non-Hodgkin lymphoma treated with CD19 CAR T cells. *Blood* **133**, 1876–1887 (2019).
- Giavridis, T. et al. CAR T cell-induced cytokine release syndrome is mediated by macrophages and abated by IL-1 blockade. *Nat. Med.* **24**, 731–738 (2018).
- Giuffrida, L. et al. IL-15 preconditioning augments CAR T cell responses to checkpoint blockade for improved treatment of solid tumors. *Mol. Ther.* **28**, 2379–2393 (2020).
- Norelli, M. et al. Monocyte-derived IL-1 and IL-6 are differentially required for cytokine-release syndrome and neurotoxicity due to CAR T cells. *Nat. Med.* **24**, 739–748 (2018).
- Rossi, J. et al. Preinfusion polyfunctional anti-CD19 chimeric antigen receptor T cells are associated with clinical outcomes in NHL. *Blood* **132**, 804–814 (2018).
- Porter, D. et al. Grading of cytokine release syndrome associated with the CAR T cell therapy tisagenlecleucel. *J. Hematol. Oncol.* **11**, 35 (2018).
- Neelapu, S. S. et al. Chimeric antigen receptor T-cell therapy - assessment and management of toxicities. *Nat. Rev. Clin. Oncol.* **15**, 47–62 (2018).
- Neelapu, S. S. et al. Toxicity management after chimeric antigen receptor T cell therapy: one size does not fit 'ALL'. *Nat. Rev. Clin. Oncol.* **15**, 218 (2018).
- Hay, K. A. et al. Kinetics and biomarkers of severe cytokine release syndrome after CD19 chimeric antigen receptor-modified T-cell therapy. *Blood* **130**, 2295–2306 (2017).
- Lee, D. W. et al. Current concepts in the diagnosis and management of cytokine release syndrome. *Blood* **124**, 188–195 (2014).
- Curran, K. J. et al. Toxicity and response after CD19-specific CAR T-cell therapy in pediatric/young adult relapsed/refractory B-ALL. *Blood* **134**, 2361–2368 (2019).
- Lee, D. W. et al. T cells expressing CD19 chimeric antigen receptors for acute lymphoblastic leukaemia in children and young adults: a phase 1 dose-escalation trial. *Lancet* **385**, 517–528 (2015).
- Maude, S. L. et al. Tisagenlecleucel in children and young adults with B-cell lymphoblastic leukemia. *N. Engl. J. Med.* **378**, 439–448 (2018).
- Fry, T. J. et al. CD22-CAR T cells induce remissions in CD19-CAR Naïve and resistant B-ALL. *Nat. Med.* **24**, 20–28 (2018).
- Locke, F. L. et al. Long-term safety and activity of axicabtagene ciloleucel in refractory large B-cell lymphoma (ZUMA-1): a single-arm, multicentre, phase 1–2 trial. *Lancet Oncol.* **20**, 31–42 (2019).
- Dong, J. et al. miR-10a rejuvenates aged human mesenchymal stem cells and improves heart function after myocardial infarction through KLF4. *Stem Cell Res Ther.* **9**, 151–151 (2018).
- Raje, N. et al. Anti-BCMA CAR T-cell therapy bb2121 in relapsed or refractory multiple myeloma. *N. Engl. J. Med.* **380**, 1726–1737 (2019).
- Yan, Z. et al. A combination of humanized anti-CD19 and anti-BCMA CAR T cells in patients with relapsed or refractory multiple myeloma: a single-arm, phase 2 trial. *Lancet Haematol.* **6**, e521–e529 (2019).
- Park, J. H. et al. Long-term follow-up of CD19 CAR therapy in acute lymphoblastic leukemia. *N. Engl. J. Med.* **378**, 449–459 (2018).
- Hu, Y. et al. Potent anti-leukemia activities of chimeric antigen receptor-modified T cells against CD19 in Chinese patients with relapsed/refractory acute lymphocytic leukemia. *Clin. Cancer Res.* **23**, 3297–3306 (2017).
- Neelapu, S. S. CAR-T efficacy: is conditioning the key? *Blood* **133**, 1799–1800 (2019).
- Zettler, M. E. et al. Real-world adverse events associated with CAR T-cell therapy among adults age ≥ 65 years. *J. Geriatr. Oncol.* **12**, 239–242 (2021).
- Kale, S. D. et al. “Small” intestinal immunopathology plays a “big” role in lethal cytokine release syndrome, and its modulation by interferon-γ, IL-17A, and a Janus kinase inhibitor. *Front. Immunol.* **11**, 1311 (2020).
- Wang, Y. et al. A retrospective comparison of CD19 single and CD19/CD22 bispecific targeted chimeric antigen receptor T cell therapy in patients with relapsed/refractory acute lymphoblastic leukemia. *Blood Cancer J.* **10**, 105 (2020).
- Magnani, C. F. et al. Sleeping beauty-engineered CAR T cells achieve antileukemic activity without severe toxicities. *J. Clin. Invest.* **130**, 6021–6033 (2020).
- Turtle, C. J. et al. Immunotherapy of non-Hodgkin's lymphoma with a defined ratio of CD8⁺ and CD4⁺ CD19-specific chimeric antigen receptor-modified T cells. *Sci. Transl. Med.* **8**, 355ra116 (2016).
- Pan, J. et al. High efficacy and safety of low-dose CD19-directed CAR-T cell therapy in 51 refractory or relapsed B acute lymphoblastic leukemia patients. *Leukemia* **31**, 2587–2593 (2017).
- Frey, N. V. et al. Optimizing chimeric antigen receptor T-cell therapy for adults with acute lymphoblastic leukemia. *J. Clin. Oncol.* **38**, 415–422 (2020).
- Bugelski, P. J. et al. Monoclonal antibody-induced cytokine-release syndrome. *Expert Rev. Clin. Immunol.* **5**, 499–521 (2009).
- Yan, H., Semple, K. M., González, C. M. & Howard, K. E. Bone marrow-liver-thymus (BLT) immune humanized mice as a model to predict cytokine release syndrome. *Transl. Res.* **210**, 43–56 (2019).
- Bromberg, J. E. C. et al. Rituximab in patients with primary CNS lymphoma (HOVON 105/ALLG NHL 24): a randomised, open-label, phase 3 intergroup study. *Lancet Oncol.* **20**, 216–228 (2019).
- Eltahir, M. et al. Profiling of donor-specific immune effector signatures in response to rituximab in a human whole blood loop assay using blood from CLL patients. *Int. Immunopharmacol.* **90**, 107226 (2021).

58. Byrd, J. C. et al. Rituximab using a thrice weekly dosing schedule in B-cell chronic lymphocytic leukemia and small lymphocytic lymphoma demonstrates clinical activity and acceptable toxicity. *J. Clin. Oncol.* **19**, 2153–2164 (2001).
59. Suntharalingam, G. et al. Cytokine storm in a phase 1 trial of the anti-CD28 monoclonal antibody TGN1412. *N. Engl. J. Med.* **355**, 1018–1028 (2006).
60. Topp, M. S. et al. Safety and activity of blinatumomab for adult patients with relapsed or refractory B-precursor acute lymphoblastic leukaemia: a multicentre, single-arm, phase 2 study. *Lancet Oncol.* **16**, 57–66 (2015).
61. Li, J. et al. CD3 bispecific antibody-induced cytokine release is dispensable for cytotoxic T cell activity. *Sci. Transl. Med.* **11**, eaax8861 (2019).
62. Urasaki, T. et al. Case report: A case of trimethoprim/sulfamethoxazole-triggered hypotensive shock: cytokine release syndrome related to immune checkpoint inhibitors and drug-induced hypersensitivity syndrome. *Front. Oncol.* **11**, 681997 (2021).
63. Nowak, A. K. et al. A phase 1b clinical trial of the CD40-activating antibody CP-870,893 in combination with cisplatin and pemetrexed in malignant pleural mesothelioma. *Ann. Oncol.* **26**, 2483–2490 (2015).
64. Vonderheide, R. H. et al. Clinical activity and immune modulation in cancer patients treated with CP-870,893, a novel CD40 agonist monoclonal antibody. *J. Clin. Oncol.* **25**, 876–883 (2007).
65. Ferrajoli, A., O'Brien, S. & Keating, M. J. Alemtuzumab: a novel monoclonal antibody. *Expert Opin. Biol. Ther.* **1**, 1059–1065 (2001).
66. Moreau, T. et al. Transient increase in symptoms associated with cytokine release in patients with multiple sclerosis. *Brain* **119**, 225–237 (1996).
67. Andreaskos, E. & Tsiodras, S. COVID-19: lambda interferon against viral load and hyperinflammation. *EMBO Mol. Med.* **12**, e12465 (2020).
68. Guan, W. J. et al. Clinical characteristics of coronavirus disease 2019 in China. *N. Engl. J. Med.* **382**, 1708–1720 (2020).
69. Klok, F. A. et al. Confirmation of the high cumulative incidence of thrombotic complications in critically ill ICU patients with COVID-19: an updated analysis. *Thrombosis Res.* **191**, 148–150 (2020).
70. Huang, C. et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* **395**, 497–506 (2020).
71. Caricchio, R. et al. Preliminary predictive criteria for COVID-19 cytokine storm. *Ann. Rheum. Dis.* **80**, 88–95 (2021).
72. Del Valle, D. M. et al. An inflammatory cytokine signature predicts COVID-19 severity and survival. *Nat. Med.* **26**, 1636–1643 (2020).
73. Lucas, C. et al. Longitudinal analyses reveal immunological misfiring in severe COVID-19. *Nature* **584**, 463–469 (2020).
74. Wang, W. et al. Definition and risks of cytokine release syndrome in 11 critically ill COVID-19 patients with pneumonia: analysis of disease characteristics. *J. Infect. Dis.* **222**, 1444–1451 (2020).
75. Liu, J. et al. Longitudinal characteristics of lymphocyte responses and cytokine profiles in the peripheral blood of SARS-CoV-2 infected patients. *EBioMedicine* **55**, 102763 (2020).
76. Kociolek, L. K. et al. Comparison of upper respiratory viral load distributions in asymptomatic and symptomatic children diagnosed with SARS-CoV-2 infection in pediatric hospital testing programs. *J. Clin. Microbiol.* **59**, e02593-20 (2020).
77. Liu, Y. et al. Viral dynamics in mild and severe cases of COVID-19. *The Lancet. Infect. Dis.* **20**, 656–657 (2020).
78. Hagman, K. et al. SARS-CoV-2 RNA in serum as predictor of severe outcome in COVID-19: a retrospective cohort study. *Clin. Infect. Dis.* ciae1285 (2020).
79. Azkur, A. K. et al. Immune response to SARS-CoV-2 and mechanisms of immunopathological changes in COVID-19. *Allergy* **75**, 1564–1581 (2020).
80. Lieberman, N. A. P. et al. In vivo antiviral host transcriptional response to SARS-CoV-2 by viral load, sex, and age. *PLoS Biol.* **18**, e3000849 (2020).
81. Zhang, J. J. et al. Clinical, radiological, and laboratory characteristics and risk factors for severity and mortality of 289 hospitalized COVID-19 patients. *Allergy* **76**, 533–550 (2021).
82. Mughal, M. S. et al. COVID-19 patients in a tertiary US hospital: assessment of clinical course and predictors of the disease severity. *Respir. Med.* **172**, 106130 (2020).
83. Wu, J. T. et al. Estimating clinical severity of COVID-19 from the transmission dynamics in Wuhan, China. *Nat. Med.* **26**, 506–510 (2020).
84. Kuo, C. L. et al. COVID-19 severity is predicted by earlier evidence of accelerated aging. Preprint at <https://www.medrxiv.org/content/10.1101/2020.07.10.2014777v1> (2020).
85. Zhu, Z. et al. Clinical value of immune-inflammatory parameters to assess the severity of coronavirus disease 2019. *Int. J. Infect. Dis.* **95**, 332–339 (2020).
86. Grasselli, G. et al. Risk factors associated with mortality among patients with COVID-19 in intensive care units in Lombardy, Italy. *JAMA Intern. Med.* **180**, 1345–1355 (2020).
87. Cagliero, J., Villanueva, S. & Matsui, M. Leptospirosis pathophysiology: into the storm of cytokines. *Front. Cell. Infect. Microbiol.* **8**, 204 (2018).
88. Walker, M. J. et al. Disease manifestations and pathogenic mechanisms of Group A Streptococcus. *Clin. Microbiol. Rev.* **27**, 264–301 (2014).
89. Spaulding, A. R. et al. Staphylococcal and streptococcal superantigen exotoxins. *Clin. Microbiol. Rev.* **26**, 422–447 (2013).
90. Hörber, S. et al. The atypical inhibitor of NF- κ B, I κ B ζ controls macrophage Interleukin-10 expression. *J. Biol. Chem.* **291**, 12851–12861 (2016).
91. Akira, S., Uematsu, S. & Takeuchi, O. Pathogen recognition and innate immunity. *Cell* **124**, 783–801 (2006).
92. Schroder, K. & Tschopp, J. The inflammasomes. *Cell* **140**, 821–832 (2010).
93. Channappanavar, R. & Perlman, S. Pathogenic human coronavirus infections: causes and consequences of cytokine storm and immunopathology. *Semin. Immunopathol.* **39**, 529–539 (2017).
94. Liu, Q., Zhou, Y. H. & Yang, Z. Q. The cytokine storm of severe influenza and development of immunomodulatory therapy. *Cell. Mol. Immunol.* **13**, 3–10 (2016).
95. Vardhana, S. A. & Wolchok, J. D. The many faces of the anti-COVID immune response. *J. Exp. Med.* **217**, e20200678 (2020).
96. Lykens, J. E. et al. Perforin is a critical physiologic regulator of T-cell activation. *Blood* **118**, 618–626 (2011).
97. Terrell, C. E. & Jordan, M. B. Perforin deficiency impairs a critical immunoregulatory loop involving murine CD8⁺ T cells and dendritic cells. *Blood* **121**, 5184–5191 (2013).
98. Nigrovic, P. A., Lee, P. Y. & Hoffman, H. M. Monogenic autoinflammatory disorders: conceptual overview, phenotype, and clinical approach. *J. Allergy Clin. Immunol.* **146**, 925–937 (2020).
99. Al-Samkari, H. & Berliner, N. Hemophagocytic lymphohistiocytosis. *Annu. Rev. Pathol.* **13**, 27–49 (2018).
100. Ramos-Casals, M. et al. Adult haemophagocytic syndrome. *Lancet* **383**, 1503–1516 (2014).
101. Johnson, T. S. et al. Etoposide selectively ablates activated T cells to control the immunoregulatory disorder hemophagocytic lymphohistiocytosis. *J. Immunol.* **192**, 84–91 (2014).
102. Kiriakidou, M. & Ching, C. L. Systemic lupus erythematosus. *Ann. Intern. Med.* **172**, Itc81–itc96 (2020).
103. Ruchakorn, N. et al. Performance of cytokine models in predicting SLE activity. *Arthritis Res. Ther.* **21**, 287 (2019).
104. Reynolds, J. A. et al. Cytokine profiling in active and quiescent SLE reveals distinct patient subpopulations. *Arthritis Res. Ther.* **20**, 173 (2018).
105. Jin, X. et al. Therapeutic efficacy of anti-CD19 CAR-T cells in a mouse model of systemic lupus erythematosus. *Cell. Mol. Immunol.* **18**, 1896–1903 (2021).
106. Watanabe, T. et al. Mechanistic insights into autoimmune pancreatitis and IgG4-related disease. *Trends Immunol.* **39**, 874–889 (2018).
107. Blauvelt, A. & Chiricozzi, A. The immunologic role of IL-17 in psoriasis and psoriatic arthritis pathogenesis. *Clin. Rev. Allergy Immunol.* **55**, 379–390 (2018).
108. Xhangolli, I. et al. Single-cell analysis of CAR-T cell activation reveals a mixed T_H1/T_H2 response independent of differentiation. *Genomics Proteomics Bioinformatics* **17**, 129–139 (2019).
109. Sheih, A. et al. Clonal kinetics and single-cell transcriptional profiling of CAR-T cells in patients undergoing CD19 CAR-T immunotherapy. *Nat. Commun.* **11**, 219 (2020).
110. Li, X. et al. Single-cell transcriptomic analysis reveals BCMA CAR-T cell dynamics in a patient with refractory primary plasma cell leukemia. *Mol. Ther.* **29**, 645–657 (2021).
111. Chen, P. H. et al. Activation of CAR and non-CAR T cells within the tumor microenvironment following CAR T cell therapy. *JCI Insight.* **5**, e134612 (2020).
112. Zhang, Q. et al. Transcriptome and regulatory network analyses of CD19-CAR-T immunotherapy for B-ALL. *Genomics Proteomics Bioinformatics* **17**, 190–200 (2019).
113. Jiang, Z. et al. IL-6 trans-signaling promotes the expansion and anti-tumor activity of CAR T cells. *Leukemia* **35**, 1380–1391 (2021).
114. Kang, L. et al. Interleukin-6-knockdown of chimeric antigen receptor-modified T cells significantly reduces IL-6 release from monocytes. *Exp. Hematol. Oncol.* **9**, 11 (2020).
115. Liu, Y. et al. Gasdermin E-mediated target cell pyroptosis by CAR T cells triggers cytokine release syndrome. *Sci. Immunol.* **5**, eaax7969 (2020).
116. Zmievskaia, E. et al. Application of CAR-T cell therapy beyond oncology: autoimmune diseases and viral infections. *Biomedicine* **9**, 59 (2021).
117. Wen, W. et al. Immune cell profiling of COVID-19 patients in the recovery stage by single-cell sequencing. *Cell Discov.* **6**, 31 (2020).
118. Chen, G. et al. Clinical and immunologic features in severe and moderate forms of coronavirus disease 2019. *J. Clin. Invest.* **130**, 2620–2629 (2020).
119. Sekine, T. et al. Robust T cell immunity in convalescent individuals with asymptomatic or mild COVID-19. *Cell* **183**, 158–168.e114 (2020).
120. Zhu, L. et al. Single-cell sequencing of peripheral mononuclear cells reveals distinct immune response landscapes of COVID-19 and influenza patients. *Immunity* **53**, 685–696.e683 (2020).

121. Wauters, E. et al. Discriminating mild from critical COVID-19 by innate and adaptive immune single-cell profiling of bronchoalveolar lavages. *Cell Res.* **31**, 272–290 (2021).
122. Meckiff, B. J. et al. Imbalance of regulatory and cytotoxic SARS-CoV-2-reactive CD4⁺ T cells in COVID-19. *Cell* **183**, 1340–1353.e1316 (2020).
123. Bost, P. et al. Host-viral infection maps reveal signatures of severe COVID-19 patients. *Cell* **181**, 1475–1488.e1412 (2020).
124. Qin, C. et al. Dysregulation of immune response in patients with coronavirus 2019 (COVID-19) in Wuhan, China. *Clin. Infect. Dis.* **71**, 762–768 (2020).
125. Zheng, Y. et al. Study of the lymphocyte change between COVID-19 and non-COVID-19 pneumonia cases suggesting other factors besides uncontrolled inflammation contributed to multi-organ injury. Preprint at <https://www.medrxiv.org/content/10.1101/2020.02.19.20024885v4> (2020).
126. Maucourant, C. et al. Natural killer cell immunotypes related to COVID-19 disease severity. *Sci. Immunol.* **5**, eabd6832 (2020).
127. Zhang, D. et al. Frontline science: COVID-19 infection induces readily detectable morphologic and inflammation-related phenotypic changes in peripheral blood monocytes. *J. Leukoc. Biol.* **109**, 13–22 (2021).
128. Guo, C. et al. Single-cell analysis of two severe COVID-19 patients reveals a monocyte-associated and tocilizumab-responding cytokine storm. *Nat. Commun.* **11**, 3924 (2020).
129. Lee, J. S. et al. Immunophenotyping of COVID-19 and influenza highlights the role of type I interferons in development of severe COVID-19. *Sci. Immunol.* **5**, eabd1554 (2020).
130. Reusch, N. et al. Neutrophils in COVID-19. *Front. Immunol.* **12**, 652470 (2021).
131. Schulte-Schrepping, J. et al. Severe COVID-19 is marked by a dysregulated myeloid cell compartment. *Cell* **182**, 1419–1440.e1423 (2020).
132. Guo, Q. et al. Induction of alarmin S100A8/A9 mediates activation of aberrant neutrophils in the pathogenesis of COVID-19. *Cell Host Microbe* **29**, 222–235.e224 (2021).
133. Zhang, B. et al. Immune phenotyping based on the neutrophil-to-lymphocyte ratio and IgG level predicts disease severity and outcome for patients with COVID-19. *Front. Mol. Biosci.* **7**, 157 (2020).
134. Middleton, E. A. et al. Neutrophil extracellular traps contribute to immunothrombosis in COVID-19 acute respiratory distress syndrome. *Blood* **136**, 1169–1179 (2020).
135. Barnes, B. J. et al. Targeting potential drivers of COVID-19: Neutrophil extracellular traps. *J. Exp. Med.* **217**, e20200652 (2020).
136. Ren, X. et al. COVID-19 immune features revealed by a large-scale single-cell transcriptome atlas. *Cell* **184**, 1895–1913.e1819 (2021).
137. Shakhov, A. N. et al. Kappa B-type enhancers are involved in lipopolysaccharide-mediated transcriptional activation of the tumor necrosis factor alpha gene in primary macrophages. *J. Exp. Med.* **171**, 35–47 (1990).
138. Chen, Y. et al. Therapeutic potential of TNF α and IL1 β blockade for CRS/ICANS in CAR-T therapy via ameliorating endothelial activation. *Front. Immunol.* **12**, 623610 (2021).
139. DeDiego, M. L. et al. Inhibition of NF- κ B-mediated inflammation in severe acute respiratory syndrome coronavirus-infected mice increases survival. *J. Virol.* **88**, 913–924 (2014).
140. Chatzantoni, K. & Mouzaki, A. Anti-TNF-alpha antibody therapies in autoimmune diseases. *Curr. Top. Med. Chem.* **6**, 1707–1714 (2006).
141. Mridha, A. R. et al. NLRP3 inflammasome blockade reduces liver inflammation and fibrosis in experimental NASH in mice. *J. Hepatol.* **66**, 1037–1046 (2017).
142. Alosaimi, B. et al. MERS-CoV infection is associated with downregulation of genes encoding Th1 and Th2 cytokines/chemokines and elevated inflammatory innate immune response in the lower respiratory tract. *Cytokine* **126**, 154895 (2020).
143. Scambler, T. et al. Autoinflammatory disease in the lung. *Immunology* **154**, 563–573 (2018).
144. Freeman, T. L. & Swartz, T. H. Targeting the NLRP3 Inflammasome in Severe COVID-19. *Front. Immunol.* **11**, 1518 (2020).
145. Tanaka, T., Narazaki, M. & Kishimoto, T. Immunotherapeutic implications of IL-6 blockade for cytokine storm. *Immunotherapy* **8**, 959–970 (2016).
146. Hu, Y. et al. Predominant cerebral cytokine release syndrome in CD19-directed chimeric antigen receptor-modified T cell therapy. *J. Hematol. Oncol.* **9**, 70 (2016).
147. Blanco-Melo, D. et al. Imbalanced host response to SARS-CoV-2 drives development of COVID-19. *Cell* **181**, 1036–1045.e1039 (2020).
148. Ruan, Q. et al. Clinical predictors of mortality due to COVID-19 based on an analysis of data of 150 patients from Wuhan, China. *Intensive Care Med.* **46**, 846–848 (2020).
149. Herold, T. et al. Elevated levels of IL-6 and CRP predict the need for mechanical ventilation in COVID-19. *J. Allergy Clin. Immunol.* **146**, 128–136.e124 (2020).
150. Luo, W. et al. Targeting JAK-STAT signaling to control cytokine release syndrome in COVID-19. *Trends Pharmacol. Sci.* **41**, 531–543 (2020).
151. Schroder, K., Hertzog, P. J., Ravasi, T. & Hume, D. A. Interferon-gamma: an overview of signals, mechanisms and functions. *J. Leukoc. Biol.* **75**, 163–189 (2004).
152. Holtzman, M. J., Patel, D. A., Zhang, Y. & Patel, A. C. Host epithelial-viral interactions as cause and cure for asthma. *Curr. Opin. Immunol.* **23**, 487–494 (2011).
153. Shimabukuro-Vornhagen, A. et al. Cytokine release syndrome. *J. Immunother. Cancer* **6**, 56 (2018).
154. Passarelli, C. et al. IFNAR2 deficiency causing dysregulation of NK cell functions and presenting with hemophagocytic lymphohistiocytosis. *Front. Genet.* **11**, 937 (2020).
155. Mestermann, K. et al. The tyrosine kinase inhibitor dasatinib acts as a pharmacologic on/off switch for CAR T cells. *Science Transl. Med.* **11**, eaa5907 (2019).
156. Diaconu, I. et al. Inducible caspase-9 selectively modulates the toxicities of CD19-specific chimeric antigen receptor-modified T cells. *Mol. Ther.* **25**, 580–592 (2017).
157. Sun, C. et al. THEMIS-SHP1 recruitment by 4-1BB tunes LCK-mediated priming of chimeric antigen receptor-redirection T cells. *Cancer Cell* **37**, 216–225.e216 (2020).
158. Bao, L. et al. The pathogenicity of SARS-CoV-2 in hACE2 transgenic mice. *Nature* **583**, 830–833 (2020).
159. Jiang, R. D. et al. Pathogenesis of SARS-CoV-2 in transgenic mice expressing human angiotensin-converting enzyme 2. *Cell* **182**, 50–58.e58 (2020).
160. Arunachalam, P. S. et al. Systems biological assessment of immunity to mild versus severe COVID-19 infection in humans. *Science* **369**, 1210–1220 (2020).
161. Silvini, A. et al. Elevated calprotectin and abnormal myeloid cell subsets discriminate severe from mild COVID-19. *Cell* **182**, 1401–1418.e1418 (2020).
162. Wilk, A. J. et al. A single-cell atlas of the peripheral immune response in patients with severe COVID-19. *Nat. Med.* **26**, 1070–1076 (2020).
163. Boroughs, A. C. et al. A distinct transcriptional program in human CAR T cells bearing the 4-1BB signaling domain revealed by scRNA-seq. *Mol. Ther.* **28**, 2577–2592 (2020).
164. Deng, Q. et al. Characteristics of anti-CD19 CAR T cell infusion products associated with efficacy and toxicity in patients with large B cell lymphomas. *Nat. Med.* **26**, 1878–1887 (2020).
165. Nguyen, T. H. O. et al. CD8⁺ T cells specific for an immunodominant SARS-CoV-2 nucleocapsid epitope display high naive precursor frequency and TCR promiscuity. *Immunity* **54**, 1066–1082.e1065 (2021).
166. Wang, W. et al. Joint profiling of chromatin accessibility and CAR-T integration site analysis at population and single-cell levels. *Proc. Natl Acad. Sci. USA* **117**, 5442–5452 (2020).
167. Chen, G. M. et al. Integrative bulk and single-cell profiling of pre-manufacture T-cell populations reveals factors mediating long-term persistence of CAR T-cell therapy. *Cancer Discov.* **11**, 2186–2199 (2021).
168. Teachey, D. T. et al. Identification of predictive biomarkers for cytokine release syndrome after chimeric antigen receptor T-cell therapy for acute lymphoblastic leukemia. *Cancer Discov.* **6**, 664–679 (2016).
169. Mathew, D. et al. Deep immune profiling of COVID-19 patients reveals distinct immunotypes with therapeutic implications. *Science* **369**, eabc8511 (2020).
170. Liu, E. et al. Use of CAR-transduced natural killer cells in CD19-positive lymphoid tumors. *N. Engl. J. Med.* **382**, 545–553 (2020).
171. Mahase, E. Covid-19: anti-TNF drug adalimumab to be trialled for patients in the community. *BMJ* **371**, m3847 (2020).
172. Eloseily, E. M. et al. Benefit of anakinra in treating pediatric secondary hemophagocytic lymphohistiocytosis. *Arthritis Rheumatol.* **72**, 326–334 (2020).
173. Cavalli, G. et al. Interleukin-1 blockade with high-dose anakinra in patients with COVID-19, acute respiratory distress syndrome, and hyperinflammation: a retrospective cohort study. *Lancet Rheumatol.* **2**, e325–e331 (2020).
174. Huet, T. et al. Anakinra for severe forms of COVID-19: a cohort study. *Lancet Rheumatol.* **2**, e393–e400 (2020).
175. Kang, S., Tanaka, T., Narazaki, M. & Kishimoto, T. Targeting Interleukin-6 signaling in clinic. *Immunity* **50**, 1007–1023 (2019).
176. Xu, X. et al. Effective treatment of severe COVID-19 patients with tocilizumab. *Proc. Natl Acad. Sci. USA* **117**, 10970–10975 (2020).
177. Stone, J. H. et al. Efficacy of tocilizumab in patients hospitalized with Covid-19. *N. Engl. J. Med.* **383**, 2333–2344 (2020).
178. Locatelli, F. et al. Emapalumab in children with primary hemophagocytic lymphohistiocytosis. *N. Engl. J. Med.* **382**, 1811–1822 (2020).
179. Karki, R. et al. Synergism of TNF- α and IFN- γ triggers inflammatory cell death, tissue damage, and mortality in SARS-CoV-2 infection and cytokine shock syndromes. *Cell* **184**, 149–168.e117 (2021).
180. Jiang, J. et al. Type I interferons in the pathogenesis and treatment of autoimmune diseases. *Clin. Rev. Allergy Immunol.* **59**, 248–272 (2020).
181. Prencipe, G. et al. Neutralization of IFN- γ reverts clinical and laboratory features in a mouse model of macrophage activation syndrome. *J. Allergy Clin. Immunol.* **141**, 1439–1449 (2018).

182. Locke, F. L., Go, W. Y. & Neelapu, S. S. Development and use of the anti-CD19 chimeric antigen receptor T-cell therapy axicabtagene ciloleucel in large B-cell lymphoma: a review. *JAMA Oncol.* **6**, 281–290 (2020).
183. Paliogianni, F. et al. Novel mechanism for inhibition of human T cells by glucocorticoids. Glucocorticoids inhibit signal transduction through IL-2 receptor. *J. Immunol.* **151**, 4081–4089 (1993).
184. Wang, B. et al. Subcutaneous injection of IFN alpha-2b for COVID-19: an observational study. *BMC Infect. Dis.* **20**, 723 (2020).
185. Hu, Y. et al. CAR T-cell treatment during the COVID-19 pandemic: management strategies and challenges. *Curr. Res. Transl. Med.* **68**, 111–118 (2020).
186. Yang, X. et al. Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: a single-centered, retrospective, observational study. *The Lancet. Respiratory Med.* **8**, 475–481 (2020).
187. Hong, R. et al. Clinical characterization and risk factors associated with cytokine release syndrome induced by COVID-19 and chimeric antigen receptor T-cell therapy. *Bone Marrow Transplant.* **56**, 570–580 (2021).
188. Ying, Z. et al. A safe and potent anti-CD19 CAR T cell therapy. *Nat. Med.* **25**, 947–953 (2019).
189. Zhang, L. et al. Pluripotent stem cell-derived CAR-macrophage cells with antigen-dependent anti-cancer cell functions. *J. Hematol. Oncol.* **13**, 153 (2020).
190. Chen, L. Y. et al. Successful application of anti-CD19 CAR-T therapy with IL-6 knocking down to patients with central nervous system B-cell acute lymphocytic leukemia. *Transl. Oncol.* **13**, 100838 (2020).
191. He, P. et al. Co-expressing LRP6 with anti-CD19 CAR-T cells for improved therapeutic effect against B-ALL. *Front. Oncol.* **10**, 1346 (2020).
192. Lee, Y. G. et al. Regulation of CAR T cell-mediated cytokine release syndrome-like toxicity using low molecular weight adapters. *Nat. Commun.* **10**, 2681 (2019).
193. Shah, N. N. & Fry, T. J. Mechanisms of resistance to CAR T cell therapy. *Nat. Rev. Clin. Oncol.* **16**, 372–385 (2019).
194. Ataca Atilla, P. et al. Modulating TNF α activity allows transgenic IL15-Expressing CLL-1 CAR T cells to safely eliminate acute myeloid leukemia. *J. Immunother. Cancer.* **8**, (2020).
195. Zhao, W. H. et al. A phase 1, open-label study of LCAR-B38M, a chimeric antigen receptor T cell therapy directed against B cell maturation antigen, in patients with relapsed or refractory multiple myeloma. *J. Hematol. Oncol.* **11**, 141 (2018).



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2021, corrected publication 2021