Hormesis and Endothelial Progenitor Cells

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Abstract

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Hormetic-biphasic dose response relationships are reported herein for human endothelial progenitor cells involving estradiol, nicotine, the anti-diabetic agent pioglitazone, resveratrol, and progesterone. In general, these studies demonstrate the capacity of these agents to enhance EPC proliferation and angiogenesis functional applications, having a focus on repairing endothelial tissue damage due to acute injury (e.g., stroke), as well as damage from chronic conditions (e.g., atherosclerosis) and normal aging processes.

Keywords

hormesis, stem cells, endothelial stem cells, cell proliferation, cell differentiation

Introduction

Endothelial repair was originally seen as a local type of process involving the migration and proliferation of endothelial cells from a nearby uninjured cellular zone. However, it is now known that vascular healing requires not only cells located within vessel wall environments but also circulating¹ bone marrow-derived stem cells (BMSCs).²⁻⁵ A subset of these BMSCs, called endothelial progenitor cells, are present within the peripheral blood, with the potential to proliferate and differentiate into endothelial cells,⁶ leading to the restoration of endothelial function, enhancement of angiogenesis and overall vascular repair, preventing atherosclerosis. The present paper provides an assessment of the occurrence of chemically induced hormetic dose responses in human endothelial progenitor cell functions, including cell proliferation, cell migration, cell adhesion, and the process of angiogenesis, their underlying mechanistic foundations and therapeutic applications.

Hormesis Overview

While hormesis has been substantially evaluated in the biological, toxicological, and biomedical literature, the use of the terms hormesis or hormetic within the stem cell literature is extremely limited, affecting the capacity for a standard key word search strategy, further suggesting the need to provide a brief overview of the hormesis concept for those researchers in the area of stem cells with particular interest in cell proliferation and cell differentiation and their dose response features.

Hormesis is a biphasic dose/concentration response, displaying a low dose/concentration stimulation and a high-dose/ concentration inhibition.⁷⁻⁹ Its quantitative characteristics include a maximum stimulatory response typically between 30% and 60% greater than the control group (Figure 1) along with a stimulatory width that is usually in the 10-20-fold range but may show considerable variability, not uncommonly being greater than 50-fold.¹⁰⁻¹² The hormetic response results from a direct subtoxic (hormetic) dose or a subtoxic (hormetic) preconditioning dose and a subsequent toxic dose.^{13,14} The hormesis concept shows considerable generality, being independent of biological model (e.g., microbes, plants, animal models, and humans), endpoint, level of biological organization (i.e., cell, organ, and organism), in vitro and in vivo evaluations, inducing agent, and mechanism.¹⁵⁻¹⁷ Comprehensively integrated evaluations of hormetic dose responses for both chemicals and ionizing radiation provide historical

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foundations of hormesis from first reporting in the 1880s concerning the effects of multiple chemical disinfectants on the growth/metabolism of yeast to the present.¹⁸⁻²⁴

As a result of the general lack of linkage of hormesis and stem cells in the literature, including EPCs, it was necessary to develop a broader and more general search strategy. Using principally PubMed and Web of Science databases, key words such as stem cells, cell proliferation, cell differentiation, EPCs, biphasic dose responses, low dose stimulation, as well as hormesis and hormetic and their combinations were used. In addition, all papers obtained were cross-referenced. Further, all relevant papers were assessed for each article that cited these papers (using Web of Science) and checked for relevance. Finally, all active research groups in the area were followed for all their relevant publications.

Estrogen

Estrogens provide protective roles in the vascular system, including functioning of the endothelial cell layer. Within this biological context, estrogens play an important role in the endothelial cell restoration process following damage.²⁶⁻³⁰ The capacity of estrogen to affect EPC proliferation was assessed by Foresta et al.¹ using EPCs from



Figure 1. General representation of the hormetic dose response (modified from: Calabrese and Baldwin, 1998²⁵).



Figure 2. Effects of β -estradiol on human endothelial progenitor cell proliferation (modified from: Foresta et al., 2007) *= P \leq .05.

healthy adult donors over a 10,000-fold concentration range (Figure 2). These preliminary findings indicate that the estrogen treatment enhanced cell proliferation in a biphasic manner, with the response being optimized between 10–1000 pM, a 100-fold concentration range. The stimulatory response was mediated by the estrogen receptor- α (ER- α), with no involvement of the ER- β receptor. The estrogen concentration range that was associated with the enhanced EPC proliferation effect was similar to that reported for women under hyperstimulation with in vitro fertilization techniques.

Progesterone

Progesterone, an endogenous sex hormone, has significant protective effects in the vascular system. Such protection is mediated, in part, due to its capacity to enhance EPC proliferation.³¹ In an effort to explore further the effects of progesterone on EPC, Yu et al.³² assessed its capacity to affect multiple functions including tube formation and length, cell adhesion, cell migration, and protection against induced damage due to cortical impact injury. The progesterone treatment biphasically affected multiple endpoints including cell tube formation, cell adhesion, and cell migration, with all showing the optimal concentration at 10^{-9} M (Figure 3). Follow-up experiments with UPA, a progesterone receptor antagonist, blocked the stimulatory response for multiple endpoints at the optimal 10^{-9} M concentration. This was also the case for the effect of progesterone on the secretion of vascular epithelial growth factor (VEGF). These findings

suggested that low concentrations of progesterone enhanced EPC angiogenesis activities via the progesterone receptor B (PR B) while the inhibitory effect at the higher concentrations was hypothesized to be mediated by PR A.

Nicotine

While high concentrations of nicotine can cause endothelial injury, lower concentrations can increase endothelial proliferation, reduce apoptosis, and enhance capillary network formation.³³ These findings lead Wang et al.³³ to explore the effects of nicotine on circulating human EPCs. This study provided the first report that nicotine induced biphasic concentration responses on cell proliferation, migration, adhesion, and vasculogenesis at physiologically relevant concentrations, being maximal at 10^{-8} mol/L (Figure 4). This concentration of nicotine is comparable to those found in the blood of smokers (i.e., ~60 to 100 nmol/L).³⁴ Follow-up studies by Heeschen et al.³⁵ suggested that the mechanism for the low concentration effects was related to the capacity of nicotine to stimulate the endothelial nicotinic acetylcholine receptor (hACHR) which induces angiogenesis.

Pioglitazone: Anti-Diabetic Agent

Circulating EPCs have a critical role in the process of endothelial regeneration following arterial damage.^{36,37} The capacity to repair such damage is affected by multiple factors such as age, gender, and physical training status. Coronary artery disease is the leading cause of mortality of those with



Figure 3. Effects of progesterone on endothelial progenitor cell adhesion and migration (modified from: Yu et al., 2017) * = P < .05.



Figure 4. Effects of nicotine on endothelial progenitor cells (modified from: Wang et al., 2004) * = $P \le .05$.



Figure 5. Effects of pioglitazone on human endothelial progenitor cell adhesion on fibronectin (modified from: Redondo et al., 2007) * = $P \le .05$.

type 2 diabetes, a condition associated with significant deficits in EPC functions (e.g., altered proliferation and adhesion). Within the context of treating diabetic patients, considerable attention has been directed toward the thiazolidinediones (e.g., rosiglitazone and pioglitazone), peroxisome proliferatoractivated receptor (PPAR)- γ agonists, and insulinsensitizing agents. Further interest in these agents emerged from studies showing that they offered benefits beyond affecting only glycemia. Such additional benefits include the capacity to enhance the differentiation of EPCs, an effect mediated by PPAR- γ . Further, these agents affect potential benefits via the activation of transforming growth factor (TGF)- β 1, enhancing the proliferation of EPCs in an hormetic-like biphasic manner.³⁸ These observations led Redondo et al.³⁹ to assess the effects of pioglitazone on human EPC function while elaborating the roles and interactions of PPAR- γ and TGF- β 1. While evaluating the effects of pioglitazone over a 100-fold concentration range (.1 to 10 μ M) these authors reported that pioglitazone induced a biphasic concentration effect, with the optimal response at 1.0 μ M (Figure 5). Since the pioglitazone did not significantly affect proliferation and apoptosis, the increase of EPC number may be due to enhanced adhesion and differentiation, conclusions that



Figure 6. A Effects of resveratrol on cell proliferation of endothelial progenitor cells (modified from: Xia et al., 2008) * = $P \le .05$. B Effects of resveratrol on migration of endothelial progenitor cells (modified from: Xia et al., 2008) * = $P \le .05$.

were supported by experiments blocking PPAR- γ and TGF- β 1 by antagonists. An integrated mechanistic assessment suggests that the pioglitazone low concentration stimulatory effect was mediated by PPAR- γ whereas TGF- β 1-mediated the inhibition.

Resveratrol

Since endothelial cell injury/death can play an important role in the pathogenesis of intimal hyperplasia in response to vascular injury, a therapeutic strategy that stimulates early reendothelialization of the damage area would prevent intimal lesion development while compromising long-term patency. Since there was a strong relationship of moderate consumption of red wine with a decreased occurrence of coronary artery disease in experimental models and in epidemiological studies, Gu et al.⁴⁰ assessed the effects of resveratrol on the angiogenesis activation and eNOS expression of isolated human endothelial cells in vitro and the effects of resveratrol on the mobilization of endothelial cells from bone marrow. Employing 4 concentrations (1, 5, 15, and 60 μ M), Gu et al.⁴⁰ reported the occurrence of an hormetic-like biphasic concentration response for multiple endpoints: endothelial cell proliferation, cell migration, cell adherence, and eNOS expression/concentrations. These findings were striking in their qualitative and quantitative consistency along with the optimal stimulatory response for each endpoint being $1.0 \,\mu$ M. Likewise, the maximum stimulatory response for each endpoint was in the 40 to 60% range. The follow-up findings of Gu et al.⁴⁰ were supported in follow-up *in vivo* experimental investigation using a rat model. These experiments indicated that low doses of resveratrol increased the mobilization of endothelial cells, facilitated re-endothelialization, and diminished the occurrence of neointimal formation and up-regulation of eNOS following an induced balloon injury. The Gu et al.⁴⁰ findings were strongly supported by Xia et al.⁴¹ who also reported hormetic-like biphasic dose response of resveratrol on endothelial progenitor cell proliferation and cell migration (Figures 6(a) and (b)). They also reported that resveratrol induced hormetic effects by altering teleromerase activity.

Discussion

This paper documents the occurrence of hormetic dose responses of endothelial stem cells. The nature of research in this area has focused on the capacity of EPCs to enhance the repair of damaged endothelial cells due to traumatic injury such as from a stroke or in the course of chronic disease such as type 2 diabetes, which can damage endothelial cells, and contributes to the development of atherosclerosis. The quantitative features of the hormetic dose response (N=18) reported herein are consistent with those reported in the general hormesis literature with the median maximum stimulation 162.5% and with median stimulatory range of 100-fold.

The present analysis demonstrates that the hormetic potential of EPCs may affect either a post-trauma beneficial or undesirable response. For example, in the case of estrogens, there is the capacity to enhance tissue repair of damaged vasculature. However, it is well known that estrogen may have a tumor promotional effect.^{42,43} One of the mechanisms by which this occurs could be via the enhancement of angiogenesis in developing tumors. While the negative features of nicotine have been emphasized in the literature for tumor promotion via enhanced angiogenesis⁴⁴ it may also be the case for other agents with EPC proliferation potential. Thus, the role of hormetic effects on endothelial cells has the potential to promote health by repairing damaged endothelial cells due to normal aging and enhance some disease processes or other conditions or to play a role in tumor promotion.

The present findings complement and extend a recent extensive effort to discover, document, and assess the occurrence and biological/biomedical significance of hormetic dose responses in stem cell biology. Hormetic dose responses have been now reported to be commonly reported for adipose derived stem cells (ADSCs),⁴⁵ apical papilla stem cells (APSCs),⁴⁶ bone marrow stem cells (BMSCs),⁴⁷ dental pulp stem cells (DPSCs),⁴⁸ embryonic stem cells (ESCs),⁴⁹ neuronal stem cells (NSCs),⁵⁰ and periodontal ligament stem cells (PDLSCs).⁵¹ Based on the substantial occurrence of hormetic dose responses affecting cell viability, cell survival, cell proliferation, cell differentiation, and a capacity to enhance resilience to a broad spectrum of inflammatory conditions, a broad spectrum of stem cell types use adaptive strategies that conform to the quantitative features of the hormetic dose response, displaying a substantial degree of generality. However, despite these widespread occurrences of hormesis within the stem cell literature these findings are generally unrecognized as general dose response concept clinical implications. However, the recent spate of papers on the role of hormesis in stem cell functioning is likely to enhance the recognition of the significance of hormetic-biphasic dose responses and their role in stem cell biology.

Declaration of Conflicting Interests

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References

- Foresta C, Zuccarello D, Biagioli A, De Toni L, Prana E, Nicoletti V, et al. Oestrogen stimulates endothelial progenitor cells via oestrogen receptor-alpha. *Clin Endocrinol.* 2007;67: 520-525.
- Iwakura A, Shastry S, Luedemann C, Hamada H, Kawamoto A, Kishore R, et al. Estradiol enhances recovery after myocardial infarction by augmenting incorporation of bone marrow-derived endothelial progenitor cells into sites of ischemia-induced neovascularization via endothelial nitric oxide synthase-mediated activation of matrix metalloproteinase-9. *Circulation*. 2006;113: 1605-1614.
- Fontaine V, Filipe C, Werner N, Gourdy P, Billon A, Garmy-Susini B, et al. Essential role of bone marrow fibroblast growth factor-2 in the effect of estradiol on reendothelialization and endothelial progenitor cell mobilization. *Am J Pathol*. 2006;169: 1855-1862.
- Hamada H, Kim MK, Iwakura A, Ii M, thorne T, Qin G, et al. Estrogen Receptors α and β Mediate Contribution of Bone Marrow-Derived Endothelial Progenitor Cells to Functional Recovery After Myocardial Infarction. *Circulation*. 2006;114:2261-2270.
- Strehlow K, Werner N, Berweiler J, Link A, Dirnagl U, Priller J, et al. Estrogen increases bone marrow-derived endothelial progenitor cell production and diminishes neointima formation. *Circulation*. 2003;107:3059-3065.
- Asahara T, Murohara T, Sullivan A, Silver M, van der Zee R, Li T, et al. Isolation of putative progenitor endothelial cells for angiogenesis. *Science*. 1997;275:964-966.
- Calabrese EJ, Baldwin LA. Defining hormesis. *Hum Exp Toxicol.* 2002;21:91-97.
- Calabrese EJ, Mattson MP. Hormesis provides a generalized quantitative estimate of biological plasticity. *Journal of Cell Communication and Signaling*. 2011;5:25-38.
- 9. Mattson MP. Hormesis defined. Ageing Res Rev. 2008;7:1-7.
- Calabrese E, Blain R. The occurrence of hormetic dose responses in the toxicological literature, the hormesis database: An overview. *Toxicol Appl Pharmacol*. 2005;202:289-301.
- Calabrese EJ, Blain RB. The hormesis database: The occurrence of hormetic dose responses in the toxicological literature. *Regul Toxicol Pharmacol.* 2011;61(1):73-81.
- Calabrese EJ, Agathokleous E, Kozumbo WJ, Stanek EJ, Leonard D. Estimating the range of the maximum hormetic stimulatory response. *Environ Res.* 2019;170:337-343. doi:10. 1016/j.envres.2018.12.020.
- 13. Calabrese EJ. Preconditioning is hormesis part I: Documentation, dose-response features and mechanistic foundations. *Pharmacol Res.* 2016a;110:242-264.
- Calabrese EJ. Preconditioning is hormesis part II: How the conditioning dose mediates protection: Dose optimization within temporal and mechanistic frameworks. *Pharmacol Res.* 2016b;110:265-275.

- Calabrese EJ. Hormesis: Why it is important to toxicology and toxicologists. *Environ Toxicol Chem.* 2008;27:1451-1474.
- Calabrese EJ. Hormetic mechanisms. *Crit Rev Toxicol*. 2013;43: 580-606.
- Calabrese EJ, Kozumbo WJ. The hormetic dose-response mechanism: Nrf2 activation. *Pharmacol Res.* 2021;167:105526.
- Calabrese EJ, Baldwin LA. Chemical hormesis: Its historical foundations as a biological hypothesis. *Hum Exp Toxicol*. 2000a; 19:2-31.
- Calabrese EJ, Baldwin LA. The marginalization of hormesis. *Hum Exp Toxicol*. 2000b;19:32-40.
- Calabrese EJ, Baldwin LA. Radiation hormesis: its historical foundations as a biological hypothesis. *Hum Exp Toxicol*. 2000c; 19:41-75.
- Calabrese EJ, Baldwin LA. Radiation hormesis: The demise of a legitimate hypothesis. *Hum Exp Toxicol*. 2000d;19:76-84.
- Calabrese EJ, Baldwin LA. Tales of two similar hypotheses: The rise and fall of chemical and radiation hormesis. *Hum Exp Toxicol.* 2000e;19:85-97.
- 23. Calabrese EJ. Toxicology rewrites its history and rethinks its future: giving equal focus to both harmful and beneficial effects. *Environ Toxicol Chem.* 2011;30:2658-2673.
- Calabrese E. Dose-Response: A Fundamental Concept in Toxicology. In: Hayes AW, Kruger CL, eds *Principles and Methods of Toxicology*. 6th Edition. CRC Press; 2014:89-140.
- Calabrese EJ, Baldwin LA. A general classification of U-shaped dose-response relationships in toxicology and their mechanistic foundations. *Hum Exp Toxicol.* 1998;17:353-364.
- Orshal JM, Khalil RA. Gender, sex hormones, and vascular tone. *Am J Physiol Regul Integr Comp Physiol*. 2004;286:R233-R249.
- Mendelsohn ME, Karas RH. The protective effects of estrogen on the cardiovascular system. N Engl J Med. 1999;340: 1801-1811.
- Iwakura A, Luedemann C, Shastry S, Hanley A, Kearney M, Aikawa R, et al. Estrogen-mediated, endothelial nitric oxide synthase-dependent mobilization of bone marrow-derived endothelial progenitor cells contributes to reendothelialization after arterial injury. *Circulation*. 2003;108: 3115-3121.
- Karas RH, Schulten H, Pare G, Aronovitz MJ, Ohlsson C, Gustafsson J-A, et al. Effects of Estrogen on the Vascular Injury Response in Estrogen Receptor α,β (Double) Knockout Mice. *Circ Res.* 2001;89:534-539.
- Brouchet L, Krust A, Dupont S, Chambon P, Bayard F, Arnal JF. Estradiol Accelerates Reendothelialization in Mouse Carotid Artery Through Estrogen Receptor-α but Not Estrogen Receptor-β. *Circulation*. 2001;103:423-428.
- Matsubara Y, Matsubara K. Estrogen and progesterone play pivotal roles in endothelial progenitor cell proliferation. *Reprod Biol Endocrinol.* 2012;10:2.
- 32. Yu P, Li S, Zhang Z, Wen X, Quan W, Tian Q, et al. Progesterone-mediated angiogenic activity of endothelial progenitor cell and angiogenesis in traumatic brain injury rats were antagonized by progesterone receptor antagonist. *Cell Proliferation*. 2017;50:e23362.

- Wang X, Zhu J, Chen J, Shang Y. Effects of nicotine on the number and activity of circulating endothelial progenitor cells. J Clin Pharmacol. 2004;44:881-889.
- Benowitz NL Biomarkers of cigarette smoking Smoking and Tobacco Control monograph. National Cancer Institute; 1996:93-111.
- Heeschen C, Chang E, Aicher A, Cooke JP. Endothelial progenitor cells participate in nicotine-mediated angiogenesis. *J Am Coll Cardiol.* 2006;48:2553-2560.
- Hristov M, Weber C. Endothelial progenitor cells: Characterization, pathophysiology, and possible clinical relevance. *J Cell Mol Med.* 2004;8:498-508.
- Hristov M, Zernecke A, Bidzhekov K, Liehn EA, Shagdarsuren E, Ludwig A, et al. Importance of CXC chemokine receptor 2 in the homing of human peripheral blood endothelial progenitor cells to sites of arterial injury. *Circ Res.* 2007;100: 590-597.
- Henrich D, Hahn P, Wahl M, Wilhelm K, Dernbach E, Dimmeler S, et al. Serum Derived from Multiple Trauma Patients Promotes the Differentiation of Endothelial Progenitor Cells In Vitro: Possible Role of Transforming Growth Factor-??1 and Vascular Endothelial Growth Factor165. *Shock*. 2004;21:13-16. doi:10. 1097/01.shk.0000101669.49265.50.
- Redondo S, Hristov M, Gumbel D, Tejerina T, Weber C. Biphasic effect of pioglitazone on isolated human endothelial progenitor cells: Involvement of peroxisome proliferatoractivated receptor-γ and transforming growth factor-β1. *Thromb Haemostasis*. 2007;97:979-987.
- J G, Cq W, Hh F, Hy D, XI X, Ym X, et al. Effects of resveratrol on endothelial progenitor cells and their contributions to reendothelialization in intima-injured rats. *J Cardiovasc Pharmacol.* 2006;47(5):711-721.
- Xia L, Wang XX, Hu XS, Guo XG, Shang YP, Chen HJ, et al. Resveratrol reduces endothelial progenitor cells senescence

through augmentation of telomerase activity by Akt-dependent mechanisms. *Br J Pharmacol*. 2008;155:387-394.

- Leung BS. Roles of estrogen in mammary development tumorigenesis and promotion of tumor growth. In: Leung BS, ed. *Hormonal Regulation of Mammary Tumors*. Eden Press; 1982: 30-72.
- Yager JD, Zurlo J, Ni N. Sex hormones and tumor promotion in liver. Proceedings of the Society for Experimental Biology and Medicine. Society for Experimental Biology and Medicine. 1991;198:667-674.
- Heeschen C, Jang JJ, Weis M, Pathak A, Kaji S, Hu RS, et al. Nicotine stimulates angiogenesis and promotes tumor growth and atherosclerosis. *Nat Med.* 2001;7:833-839.
- Calabrese EJ. Hormesis and adult adipose-derived stem cells. *Pharm Res (N Y)*. 2021a;172:105802. doi:10.1016/j.phrs.221/ 105803.
- Calabrese EJ. Hormesis and apical papilla stem cells. *Chem-Biol Inter* 2021b;18. (submitted).
- Calabrese EJ. Hormesis and Bone Marrow Stem Cells: Enhancing cell proliferation, differentiation and resilience to inflammatory stress. *Chem Biol Interact*. 2022c;351:109730. doi: 10.1016/j.cbi/2021.109730.
- Calabrese EJ, Agathokleous E, Dhawan G, Kapoor R, Calabrese V. Human dental pulp stem cells and hormesis. *Ageing Res Rev.* 2021a;73:101540.
- Calabrese EJ. Hormesis and embryonic stem cells. *Chem-Biol Inter*. 2021d;352:109783.
- Calabrese EJ, Calabrese V, Dhawan G, Kapoor R, Giordano J. Hormesis and neural stem cells. *Free Rad Biol Med.* 2021b;178: 314-329.
- Calabrese EJ. Human periodontal ligament stem cells and hormesis: Enhancing cell renewal and cell differentiation. *Pharmacol Res.* 2021e;173:105914. doi:10.1016/j.phrs.2021.105914.