

Exploring the promoter regions of cancer predisposition genes in patients with triple-negative breast cancer reveals the presence of rare germline variants

Michela Palleschi¹, Alessandra Virga²,*,⑩, Emanuela Scarpi³, Eugenio Fonzi³, Antonino Musolino¹,⁴, Filippo Merloni¹, Samanta Sarti¹, Rita Danesi⁵, Mila Ravegnani⁵, Chiara Casadei¹, Marianna Sirico¹, Caterina Gianni¹, Roberta Maltoni¹, Sara Bravaccini²,⑩, Daniele Calistri², Valentina Arcangeli⁵, Valentina Zampiga², Ilaria Cangini², Erika Bandini², Francesca Mannozzi³, Fabio Falcini⁵,⁶, Giovanni Martinelli², Ugo De Giorgi³, Paola Ulivi²,‡, Gianluca Tedaldi²,‡

¹Medical Oncology, Breast & GYN Unit, IRCCS Istituto Romagnolo per lo Studio dei Tumori (IRST) "Dino Amadori", Meldola, 47014, Italy,

²Biosciences Laboratory, IRCCS Istituto Romagnolo per lo Studio dei Tumori (IRST) "Dino Amadori", Meldola, 47014, Italy, ³Unit of Biostatistics and Clinical Trials, IRCCS Istituto Romagnolo per lo Studio dei Tumori (IRST) "Dino Amadori", 47014, Meldola, Italy,

"Unit of Biostatistics and Clinical Irials, INCCS Istituto Romagnolo per lo Studio del Tumori (INST) - Dino Amadori , 47014, Meldola, Italy, 4Department of Medical and Surgical Science, University of Releana, Releana, 40129, Italy

⁴Department of Medical and Surgical Science, University of Bologna, Bologna, 40138, Italy

⁵Emilia-Romagna Cancer Registry, Romagna Unit, IRCCS Istituto Romagnolo per lo Studio dei Tumori (IRST) "Dino Amadori", Meldola, 47014, Italy,

⁶Local Health Authority, Cancer Prevention Unit, Forlì, 47121, Italy,

Department of Hematology and Sciences Oncology, Institute of Haematology "L. and A. Seràgnoli", S. Orsola University Hospital, Bologna, 40138, Italy,

Department of Medical Oncology, IRCCS Istituto Romagnolo per lo Studio dei Tumori (IRST) "Dino Amadori", Meldola, 47014, Italy,

*Corresponding author: Alessandra Virga, PhD Biosciences Laboratory, IRCCS Istituto Romagnolo per lo Studio dei Tumori (IRST) "Dino Amadori", Via Piero Maroncelli, 40, 47014, Meldola (FC), Italy (alessandra.virga@irst.emr.it).

[‡]Contributed equally.

Abstract

Background: Current genetic screening for predisposition to breast cancer (BC) is limited to *BRCA1/2* exons and intron/exon boundaries, and limited information exists about the impact of variants in *BRCA1/2* non-coding regions. The majority of alterations identified in these regions remain unclassified, but evidence of the impact of variants in the regulatory regions on cancer risk and response to treatment is emerging.

Patients and methods: This project aimed to investigate the prevalence of germline variants in the non-coding regulatory regions of *BRCA1/2* and other BC predisposition genes in patients with triple-negative BC (TNBC) selected for age at cancer diagnosis and/or family history of cancer. The study also aims to investigate the relationship between these variants and clinical outcomes such as overall survival, disease-free survival (DFS), and response to treatment. We analyzed a Next-Generation Sequencing (NGS) custom panel of promoter regions of 28 genes involved in BC predisposition on 144 patients with TNBC previously tested wild type for coding regions of *BRCA1/2*.

Results: The NGS analysis identified 635 rare variants in promoter regions of the 28 genes. Among the 144 patients, for 75 with available clinical data, rare germline variants in BRCA2 promoter were statistically significantly related to worse overall survival (OS) (P-value = .017). No differences in DFS and OS were found for the other genes. Rare variants in the CDH1 promoter were related to the highest percentage of non-pathological complete response after neoadjuvant chemotherapy (P = .0273); MLH1 and PALB2 rare non-coding variants were found to be both related to bilateral BC (P = .0146 and P = .0005, respectively) and ATM promoter variants were associated with a positive family history (P = .041).

Conclusion: Our results underscore the importance of searching for rare germline variants in regulatory regions of cancer predisposition genes in patients with TNBC, since these variants can be associated with an increased cancer risk.

Key words: hereditary breast cancer; triple-negative breast cancer; promoter variants; regulatory regions; cancer predisposition.

Implications for practice

Current genetic screening for inherited diseases is mainly focused on exonic regions, in particular the available commercial tests cover principally the coding sequence or sometimes only hotspot position in several genes. The clinical importance of studying germline variants in the promoter regions of *BRCA1/2* and other predisposition genes in patients with TNBC is critical for risk stratification and optimizing treatment response in hereditary cancer. Current genetic screening of noncoding regions faces limitations in understanding the effective role of molecular alterations for patients with inherited syndromes. Our study examined the regulatory and promoter regions of 28 cancer

predisposition genes to highlight the potential correlation between gene variants and clinical outcomes. We identified several variants in the promoter regions of cancer predisposition genes that could increase breast cancer risk. The results suggested a significant association between *BRCA2* variants and worse OS, while *CDH1*, *MLH1*, *PALB2* and *ATM* variants showed relationships with treatment response, disease characteristics and family history, respectively. However, more research is needed into the role of genetic alterations in the regulatory regions of cancer susceptibility genes.

Introduction

Triple-negative breast cancer (TNBC) is a type of breast cancer (BC) in which cells do not have estrogen or progesterone receptors (ER or PR) and do not contain high levels of HER2 receptor. TNBC accounts for about 10%-15% of all BCs.¹ Women under 40 years, who are Black, or who have a *BRCA1*/2 pathogenic variant are more likely to develop TNBC.² More than 75% of BCs that develop in carriers of a *BRCA1*/2 mutation (*BRCAmut*) are TNBC. TNBC is usually more aggressive, harder to treat, and more likely to recur than cancers that are hormone receptor-positive.

The presence of *BRCA1/2* variants can affect the treatment of TNBC in several ways:

- PARP inhibitors (PARPi): women with a *BRCAmut* TNBC may benefit from targeted therapies such as PARPi. PARPi are drugs that block an enzyme called PARP, which helps repair damaged DNA. In women with a *BRCAmut* TNBC, PARPi can be effective because they target cells that are already deficient in DNA repair mechanisms. Several studies have shown that PARPi can improve survival in women with *BRCAmut* TNBC.³
- Chemotherapy: standard neoadjuvant chemotherapy remains the standard of care for early-stage TNBC, regardless of *BRCA1/2* status. However, some studies have suggested that women with *BRCAmut* TNBC may respond differently to chemotherapy than non-*BRCAmut* TNBC. For example, a systematic review and meta-analysis found that *BRCAmut* TNBC had a higher response rate to platinum-based chemotherapy than non-*BRCAmut* TNBC.⁴
- Clinical trials: women with *BRCAmut* TNBC may be eligible for clinical trials of new treatments. For example, a recent study found that the combination of a PARPi and immunotherapy (pembrolizumab) improved progression-free survival in women with *BRCAmut* TNBC.⁵

Current genetic screening for BRCA1/2 variants is limited to the coding exons and intron/exon boundaries of BRCA1 and BRCA2 genes. However, thanks to the wide use of Next-Generation Sequencing (NGS), the number of genes and genomic alterations suspected to be involved in cancer predisposition has dramatically increased.^{6,7} However, evidence of non-coding variants' impact on cancer risk and response to treatment begins to emerge.8 Despite this, limited information currently exists about the impact of variants in promoter regions, and the majority of variants that were identified in these regions remain unclassified. Recent studies have demonstrated that alterations in these regions, may alter the transcriptional activities of BRCA1 and BRCA2, potentially leading to an increased susceptibility to breast and ovarian cancers (BOCs), as well as other malignancies such as pancreatic and prostate cancers. 9,10 Promoter and regulatory regions are essential components of the genome that control gene expression by facilitating the binding of transcription factors and RNA polymerase, thereby influencing cellular functions and

developmental processes (Figure 1). These regions can modulate the timing, location, and level of gene expression, which is crucial for maintaining normal physiological functions and responding to environmental changes.¹¹ Understanding the dynamics of these regulatory elements is vital for elucidating the complex mechanisms underlying gene regulation and its implications for health and disease.¹²

Our study aims to investigate the prevalence of rare germline variants in the regulatory regions of BC predisposition genes in patients with risk factors: TNBC, early onset (< 35 years) and/or family history of BOCs, and/or bilateral BC. The study also aims to investigate the relationship between these variants and clinical outcomes such as overall survival (OS), disease-free survival (DFS), and response to treatment.

Patients and methods

Ethics statement

The study was performed in accordance with the Good Clinical Practice and the Declaration of Helsinki and approved by the AVR Ethics Committee (protocol L3P2210, GifT). All the patients signed informed consent for the genetic analyses and the use of the results for research purposes.

Patients and samples

We enrolled 144 consecutive withpatients TNBC (years 2019-2021) previously subjected to standard germline *BRCA1/2* testing and resulted wild type (WT) for the presence of alterations in the coding regions, after selection by the Genetic Counseling Service of IRCCS IRST.

The following data were collected from all consenting patients after registration:

- Demographic data: birthday, weight and height at the time of treatment initiation, ECOG performance status;
- Tumor information: date of diagnosis, cancer histology, grade and stage, date of second malignancy onset, type of tumor and its main characteristics;
- Treatment information: use of neoadjuvant treatment, date of start and end of chemotherapy, chemotherapeutic regimen with doses, number of cycles administered, type of surgery, date of surgery, date of progression/relapse (if any), number and types of further therapeutic regimens;
- Date of death or last follow-up (if still alive).

Next-generation sequencing

All the experiments were performed in the IRST Biosciences Laboratory

Blood was stored at – 80°C until genomic DNA was extracted. DNA was purified by QIAamp DNA Mini Kit (Qiagen) and quantified using Qubit fluorometer (Thermo Fisher Scientific) with Qubit dsDNA BR Assay Kit.

Sequencing libraries were created starting from 200 ng of genomic DNA, following the protocol Illumina DNA

Prep with Enrichment (Illumina) with an NGS custom panel (Integrated DNA Technologies).

The custom panel was designed including the promoter regions of 28 genes associated with a predisposition towards BOCs, retrieving the genomic coordinates of the promoters from UCSC and Ensembl genome browsers (**Table 1**). Sequencing was performed by using the NextSeq550 platform (Illumina) with NextSeq 500/550 Mid Output Kit v2.5 (300 Cycles) configured 2 × 151 cycles.

Bioinformatics analysis

Raw reads were analyzed with Illumina DRAGEN Bio-IT Platform v4.0 (1), on the hg19 reference genome, using the following command line parameters:

-- read-trimmers quality,adapter --trim-min-quality 20 --trim-adapter-read1 < adapter_file> --trim-adapter-read2 < adapter_file>

- -- enable-duplicate-marking true
- -- enable-variant-caller true
- -- vc-target-bed < bed_file>
- -- vc-target-bed-padding 50

Resulting variant calls were annotated with ANNOVAR v2020-06-07 (2) on the databases refGene, gnomad211_exome, clinvar_20221231. Then, the variants in the promoters were manually filtered with the following exclusion criteria:

• Variants present in >5 patients;

Intron

Exon

- Variants with a population frequency >5% (based on the gnomAD database);
- Variants with a variant allele frequency (VAF) <0.3 (considered unreliable calls on germline DNA);
- Variants with a coverage <10 (deemed unreliable calls);

DNA

• Variants classified as benign/likely benign on ClinVar;

Exon

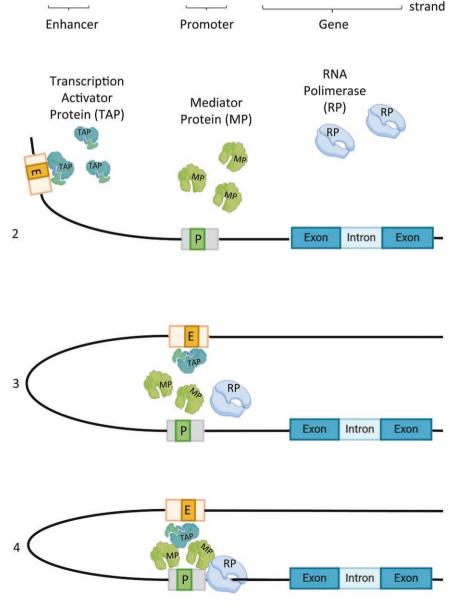


Figure 1. The explanatory figure illustrating the role and structure of enhancers, promoters, and regulatory regions in human genes.

Table 1. NGS custom panel.

Gene	Chromosome	Transcript	Promoter coordinates (hg19)		
ABRAXAS1	4q21.23	NM_139076.3	84405004	_	84407579
ATM	11q22.3	NM_000051.4	108090352	_	108102824
BAP1	3p21.1	NM_004656.4	52443528	_	52445254
BARD1	2q35	NM_000465.4	215670586	_	215676228
BLM	15q26.1	NM_000057.4	91259223	_	91261915
BRCA1	17q21.31	NM_007294.4	41275950	_	41279128
BRCA2	13q13.1	NM_000059.4	32888279	_	32891522
BRIP1	17q23.2	NM_032043.3	59938392	_	59942231
CDH1	16q22.1	NM_004360.5	68770451	_	68781341
CHEK2	22q12.1	NM_007194.4	29136832	_	29139889
MLH1	3p22.2	NM_000249.4	37029343	_	37036702
MRE11	11q21	NM_005591.4	94225645	_	94228482
MSH2	2p21-p16.3	NM_000251.3	47628819	_	47632070
MSH6	2p16.3	NM_000179.3	48008831	_	48012262
NBN	8q21.3	NM_002485.5	90992947	_	90997773
NF1	17q11.2	NM_001042492.3	29420791	_	29423873
NTHL1	16p13.3	NM_002528.7	2096199	_	2099088
PALB2	16p12.2	NM_024675.4	23650914	_	23654441
PMS2	7p22.1	NM_000535.7	6046956	_	6053558
PTEN	10q23.31	NM_000314.8	89619851	_	89629136
RAD50	5q31.1	NM_005732.4	131891483	_	131895115
RAD51C	17q22	NM_058216.3	56768997	_	56771624
RAD51D	17q12	NM_002878.4	33445311	_	33447640
RECQL4	8q24.3	NM_004260.4	145741765	_	145744684
STK11	19p13.3	NM_000455.5	1202623	_	1210888
TP53	17p13.1	NM_000546.6	7587180	_	7593026
WRN	8p12	NM_000553.6	30890166	_	30893433
XRCC2	7q36.1	NM_005431.2	152371781	-	152374441

List of the 28 cancer predisposition genes included in the NGS custom panel with the chromosome position, the name of the transcript and the genomic coordinates of the promoter regions analyzed.

Variants located in highly repetitive regions (microsatellites).

The remaining variants were used to produce Oncoprint plots (R package ComplexHeatmap v2.16.0) and Lolliplots (R package trackViewer v1.38.1).

Statistical analysis

Median values (with interquartile range-IQR) were reported for continuous variables, while absolute values and percentages were reported for non-continuous variables. DFS was calculated in months as the difference between the date of diagnosis and the date of disease progression for patients experiencing progression. For patients who did not experience progression or had no evidence of disease at the time of death, DFS was calculated as the difference between the date of diagnosis and the date of the last disease re-evaluation or the date of death. Events were defined as instances of disease progression or deaths without evidence of disease. OS was calculated in months as the difference between the date of diagnosis and the date of death for deceased patients. For living patients, OS was calculated as the difference between the date of diagnosis and the date of the last follow-up. Events were represented by

deaths. The association between clinical characteristics and genetic variants was assessed using the Chi-square test. The probability percentages for DFS and OS were calculated using the Kaplan-Meier product limit method.¹³ The proportional hazards Cox regression model was used to calculate Hazard Ratios (HR) and their corresponding 95% confidence intervals (95% CI) for covariates, both in univariate and multivariate analyses. Due to the exploratory nature of the study, no multiple testing corrections were made. All *P*-values were determined using two-tailed tests, and the statistical analyses were conducted using the SAS statistical software, version 9.4 (SAS Institute).

Results

Between January 2019 and December 2021, 144 patients with TNBC were recruited in this study.

Complete clinical characteristics were available for 75 patients and are summarized in **Table 2**. Among the 75 patients, with a median age of 53 years (range, IQR 47-60), 29 (38.7%) had a family history of cancer, 3 (4%) presented bilateral tumors, 22 (29.3%) had residual disease after neoadjuvant chemotherapy, only 3 patients (4%) developed a

second tumor. Most of the patients received anthracycline therapy in a (neo) adjuvant setting (85.3%); relapse occurred in 22 patients (29%).

The NGS analysis on the promoter regions of 28 genes of the 144 patients with TNBC, revealed the presence of 635 rare variants: 54 small deletions, 28 small insertions and 553 nucleotide changes, as shown in **Supplementary Table S1**. Out of these 635 rare variants, the most frequently alterations were in the promoters of the CDH1 (40%), ATM (35%), STK11 (31%), PTEN (18%), PMS2 (21%), MSH2 (15%), and BARD1 (16%) genes (**Supplementary Figure S1**). A more in-depth analysis in regulatory regions of 4 genes of interest was conducted to highlight the position of alterations (**Figure 2**).

Rare germline variants in *BRCA2* promoter were statistically significantly related to worse DFS (**Figure 3A**) and OS (HR = 4.76, 95% CI, 1.32-17.15, P = .017) (**Figure 3B**). No differences in DFS and OS were found for other genes. Rare variants in *CDH1* promoter were related to the highest percentage of non-pathological complete response (pCR) (P = .027) (**Figure 3C**). Rare variants in *MLH1* and *PALB2* promoters were found to be related to bilateral BC (P = .015 and P < 0.001, respectively—data not shown). Rare variants of the regulatory regions in *ATM* gene were associated with a positive family history (P = .041), as shown in **Figure 3D**.

As regards the patient clinical characteristics, the stage at diagnosis and the residual disease were unfavorable prognostic factor for both univariate DFS and OS (Table 3).

Table 2. Patient clinical characteristics.

	N. (%)	
Age: median value (range, IQR)	53 (34-71, 47-60	
Family history of cancer		
No	46 (61.3)	
Yes	29 (38.7)	
Bilateral tumors		
No	72 (96.0)	
Yes	3 (4.0)	
Stage		
I	25 (33.3)	
II	31 (41.4)	
III	19 (25.3)	
Anthracycline therapy:		
No	11 (14.7)	
Yes	64 (85.3)	
Neoadjuvant chemotherapy		
No	42 (56.0)	
Yes	33 (44.0)	
Pathological complete response (pCR)		
Yes	11 (33.0)	
No	22 (67.0)	
Adjuvant chemotherapy		
No	23 (30.7)	
Yes	52 (69.3)	

List of the patient's clinical characteristics. The table shows family history of cancer, presence of tumor bilaterality, tumor stage, treatment data, and response to therapy. N: number, IQR: interquartile range.

In multivariate analysis, including BRCA2 promoter, residual disease, and age, after backward stepwise selection, rare germline variants of BRCA2 promoter are confirmed to be an independent prognostic factor correlated with shorter DFS (HR = 4.39, 95% CI, 1.17-16.52, P = .028) and OS (HR = 6.23, 95% CI, 1.49-26.09, P = 0.012) (Table 4).

Discussion

Inherited pathogenic variants in the BRCA1 and BRCA2 genes are well-established risk factors for BC, particularly in women with a family history of the disease.¹⁴ TNBC is an aggressive subtype of BC that is associated with a poorer prognosis compared to other subtypes. 13 Recent studies have shown that rare germline variants in non-coding regions of BRCA1/2 and other genes can contribute to the development of the disease.^{8,9,14} Traditional genetic screening methods, such as those used in commercial tests (eg, FoundationOne), focus primarily on exonic regions, which may miss significant non-coding variants that can affect gene expression and contribute to cancer risk. Emerging evidence suggests that variants in these regulatory regions may influence not only the likelihood of developing BC, but also treatment response and overall clinical outcomes. 15,16 This highlights the need to expand genetic testing to include regulatory regions, as understanding these elements could lead to more comprehensive risk assessments and personalized treatment strategies, ultimately improving patient management and outcomes. In particular, a recent study has shown that variants in the 5' region of BRCA1 and BRCA2 genes can alter promoter activity and protein binding, which can affect gene expression, ultimately impacting BC risk.8 Another study found that non-coding variants in BRCA1 and BRCA2 genes can affect the splicing of RNA, which can lead to the production of abnormal proteins and contribute to cancer development. These variants are spread throughout the regulatory and promoter regions of the genes and it can be difficult to detect them using traditional genetic testing methods.¹⁸ Moreover, there is limited information on ongoing clinical trials investigating the impact of non-coding variants in BRCA1 and BRCA2 regulatory regions on BC prognosis. While these studies suggest that variants in BRCA1 and BRCA2 genes can impact BC risk, there is limited research on their impact on BC prognosis. 19,20 To date, the risks associated with rare variants in BC predisposition genes have been largely unclear.

We aimed to investigate the impact of rare germline variants in promoter regions of BRCA1/2 and other 26 cancer predisposition genes on TNBC. In our series of 144 patients with early TNBC previously tested WT for the presence of germline variants in the coding regions of BRCA1/2 and other cancer predisposition genes, all patients were found to have at least one rare variant in the promoter regions of 28 BC predisposition genes. In line with the literature data, among the most altered regulatory regions we identified mainly genes involved in tumor suppression and DNA damage repair systems.^{21,22} For 75 patients, complete clinical data were available and we correlated the presence of the rare variants in the regulatory regions of 28 genes with the clinical variables of interest and disease aggressiveness. Rare non-coding germline variants in BRCA2 promoter were found to significantly worsen OS (P = .017; HR = 4.76, 95% CI, 1.32-17.15). In the POSH study,²³ patients with a BRCAmut had a similar prognosis as patients without these alterations, but several studies

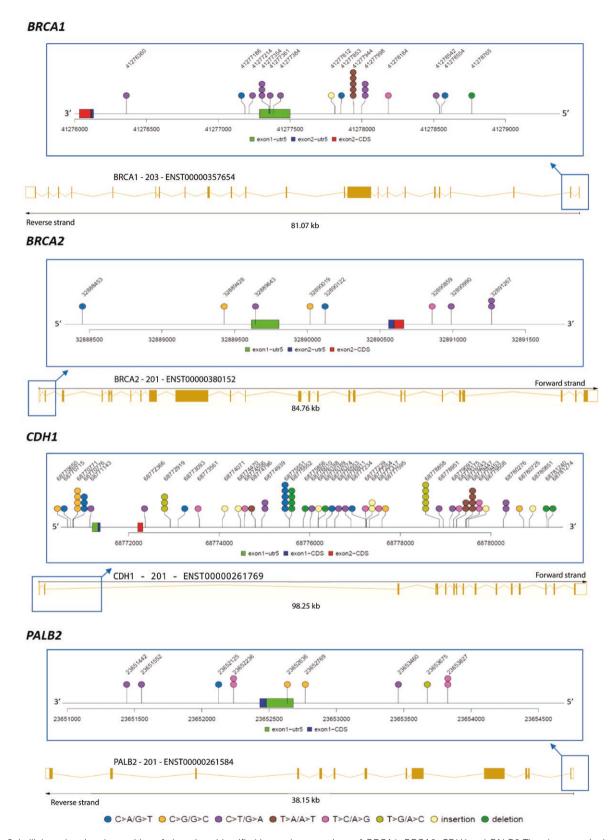


Figure 2. Lolliplots showing the position of alterations identified in regulatory regions of *BRCA1*, *BRCA2*, *CDH1* and *PALB2*. The plot range is the promoter region of each gene (see Table 1), with a downstream padding of 300 bp.

have indicated that variants in *BRCA1* and *BRCA2* may impact the effectiveness of chemotherapy. The GeparSixto Study²⁴ randomly assigned patients to either standard chemotherapy containing anthracycline and taxane alone or

with the addition of carboplatin. The trial demonstrated that in patients with TNBC, the presence of a *BRCA1/2* variant was linked to a higher percentage of patients achieving pCR and improved survival when compared to those without

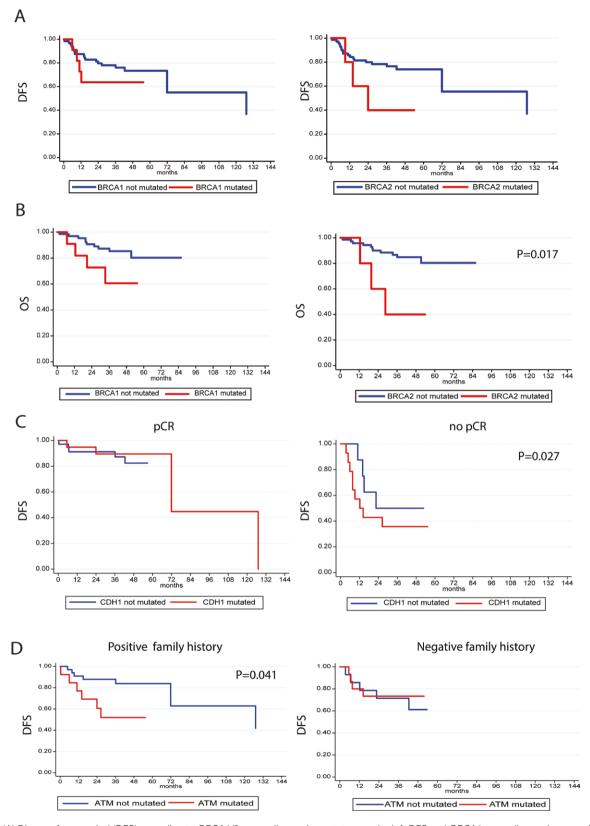


Figure 3. (A) Disease-free survival (DFS) according to *BRCA1/2* non-coding regions status: on the left DFS and *BRCA1*non-coding regions: on the right DFS and regulatory regions of *BRCA2*. (B) Overall survival (OS) according to *BRCA1/2* non-coding regions status on the left relationship between alterated non-coding regions of *BRCA1* and OS, on the right relationship between regulatory regions of *BRCA2* and OS (C) DFS according to non-coding regions of *CDH1* status and pathological complete response (pCR): on the left pCR, on the right no pCR. (D) DFS according to *ATM* status in regulatory regions and family history: on the left Positive family history; on the right Negative family history.

Table 3. Univariate analysis of DFS and OS according to clinical characteristics

	DFS		OS	
	HR (95% CI)	P	HR (95% CI)	P
Family history of cancer				
No	1.00		1.00	
Yes	1.34 (0.56-3.24)	.511	0.88 (0.29-2.63)	.822
Bilateral tumors				
No	1.00		1.00	
Yes	NE	-	NE	_
Residual disease				
No	1.00		1.00	
Yes	6.09 (2.41-15.38)	.0001	3.80 (1.31-10.98)	.014
Stage				
I	1.00		1.00	
II	3.57 (0.99-12.90)		NE	
III	7.42 (1.94-28.42)	.013	NE	_

Univariate analysis of DFS and OS according to clinical characteristics. Abbreviations: DFS, disease -free survival; NE, not estimable, OS, overall survival.

Table 4. Multivariate analysis of DFS and OS according to age, residual disease, and germline BRCA2 rare variants.

	DFS		OS	
	HR (95% CI)	P	HR (95% CI)	P
Residual disease				
No	1.00		1.00	
Yes	4.41 (1.64-11.89)	.003	3.46 (1.03-11.65)	.045
BRCA2				
WT	1.00		1.00	
mutated	4.39 (1.17-16.52)	.028	6.23 (1.49-26.09)	.012

Abbreviations: DFS, disease-free survival; OS, overall survival; WT, wild type.

alterations, irrespective of other factors (eg, chemotherapy). Patients without BRCA1/2 alterations who received standard chemotherapy without carboplatin had lower DFS rates than those who received chemotherapy plus carboplatin.²⁵ Therefore, patients with a BRCAmut TNBC might have a survival advantage because of the higher efficacy of systemic chemotherapy. The GeparOLA study²⁶ aimed to assess the efficacy and safety of neoadjuvant olaparib in combination with paclitaxel compared to carboplatin in patients with HER2-negative BC and homologous recombination deficiency. Although the study did not meet its primary endpoint, it demonstrated that olaparib in a neoadjuvant setting is comparable to carboplatin for patients with a BRCAmut while exhibiting reduced toxicity. This finding supports the notion that by continuing to select patients who are sensitive to PARPi, there is potential for a de-escalation strategy regarding treatment toxicity for a broader patient population. None of our patients received platinum salts as chemotherapy. In addition, an Italian study observed a lower BC-specific OS rate in BRCA2 variant carriers after the first two years after diagnosis.²⁷ Most of the deaths in our case series were observed in the first 2 years from diagnosis. Consistent with

literature data, rare variants in regulatory region of *PALB2* were present in 10% of patients, and 29% of the carriers had bilateral tumors with a positive statistical association (*P* < .001).²⁸ Prophylactic bilateral mastectomy is indicated in several clinical scenarios for women with *PALB2* mutations, particularly when BC is identified in one breast. This surgical procedure has the potential to lower the risk of developing neoplasms in the contralateral breast by as much as 95%. The National Comprehensive Cancer Network guidelines advocate that individuals with *PALB2* mutations should undergo genetic counseling to assess the advantages of bilateral mastectomy along with other risk-reducing strategies. These recommendations emphasize the importance of a personalized approach to cancer risk management, considering both individual and familial cancer histories.²⁸

Similarly, MLH1 regulatory variants were found in 16% of our cases and were related to a higher risk of bilateral tumors (P = .015). MLH1 gene, involved in Lynch syndrome plays a significant role in cancer predisposition, since MLH1 mutation carriers have a high risk for multiple primary cancers, including colorectal, endometrial, ovarian and breast cancers. 29,30 Moreover, variants in ATM regulatory regions were

strongly associated with a positive family history (P = 0.041) whilst alterations in the regulatory region of CDH1 were strongly associated with residual after neoadjuvant chemotherapy (P = .027) being present in 44% of all cases. In the EpiTax-trial, CDH1 mutations predicted an inferior response in the paclitaxel arm (P = .01) as well as the epirubicin arm (P = .04). Our findings, together with literature data, support the predictive value of CDH1 mutations in relation to treatment outcomes after neoadjuvant chemotherapy, which aligns the significance of alterations also in the regulatory non-coding region of CDH1 and residual disease.

Overall, our results underline the importance of extended genetic testing using panels including the promoter regions of genes involved in cancer predisposition, especially in BC patients lacking variants in the coding regions of BRCA1/2. The last American College of Medical Genetics guidelines³² do not provide specific recommendations for the reporting and classification of variants identified in BRCA1/2 promoter, intronic, and untranslated regions. Therefore, carriers should be managed exclusively based on their personal and family history, which allows for the estimation of cancer risk. The identification of genetic variants in the regulatory regions of cancer predisposition genes could modify the clinical, personal, and familial history in terms of surveillance, prevention strategies, and also personalized treatments, such as the use of PARP inhibitors in patients with alterations in homologous recombination genes. Variants of uncertain significance represent a challenge, since the disease risk associated with them may be over-interpreted or misinterpreted. As a result, it should not be used for clinical decisionmaking. To date, there is limited available data concerning sequence alterations in non-coding regions of BRCA1/2. Even less information is available about the outcome of carriers who should be managed based on their lifetime cancer risk once their genetic screening remains inconclusive. In the future, the use of genetic tests including the regulatory and non-coding regions of cancer predisposition genes will improve the management and treatment of patients with cancer predisposition.

Conclusion

To summarize, our study underscores the growing significance of rare germline variants in the regulatory regions of genes such as BRCA1/2 in contributing to TNBC predisposition. Identifying individuals at increased risk due to these variants can guide clinical management, potentially improving patient outcomes. Our study highlighted the importance of analyzing the non-coding and regulatory regions of cancer predisposition genes in patients with a suspected hereditary tumor. The associations identified between genetic alterations and clinical characteristics resulted to be statistically weak, due to the limited number of patients. However, the trends observed were in line with the literature data on carriers of variants in the coding regions of cancer predisposition genes, suggesting that also variants in non-coding regions can affect the gene function. Further research is necessary to fully understand their role in BC risk and to develop enhanced screening and prevention approaches for at-risk individuals. Given the limitations, our analyses should be regarded as preliminary, and larger studies are needed to validate these findings, including functional assays on the biological role of the identified alterations.

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Author contributions

Gianluca Tedaldi, Michela Palleschi, Alessandra Virga, Paola Ulivi, Ugo De Giorgi, Daniele Calistri, Fabio Falcini, Giovanni Martinelli (Conceptualization). Michela Pallesch, Alessandra Virga, Gianluca Tedaldi, Eugenio Fonzi, Emanuela Scarpi, Filippo Merloni, Samanta Sarti, Chiara Casadei, Sara Bravaccini (Data curation), Michela Palleschi, Alessandra Virga, Gianluca Tedaldi, Eugenio Fonzi, Emanuela Scarpi (Formal Analysis). NA (Funding acquisition). Michela Pallesch, Alessandra Virga, Gianluca Tedaldi, Eugenio Fonzi, Emanuela Scarpi. Methodology Gianluca Tedaldi, Alessandra Virga, Valentina Zampiga, Ilaria Cangini, Erika Bandini (Investigation). Michela Pallesch, Alessandra Virga, Eugenio Fonzi, Gianluca Tedaldi. Resources Valentina Zampiga, Ilaria Cangini, Erika Bandini, Daniele Calistri, Rita Danesi, Mila Ravegnani, Valentina Arcangeli, Marianna Sirico, Caterina Gianni, Roberta Maltoni, Francesca Mannozzi Software Eugenio Fonzi, Emanuela Scarpi Supervision Paola Ulivi, Ugo De Giorgi, Gianluca Tedaldi, Daniele Calistri, Fabio Falcini, Giovanni Martinelli (Project administration). Michela Pallesch, Alessandra Virga, Gianluca Tedaldi, Eugenio Fonzi, Emanuela Scarpi (Validation). Michela Pallesch, Alessandra Virga, Gianluca Tedaldi, Eugenio Fonzi, Emanuela Scarpi (Visualization). Michela Pallesch, Alessandra Virga, Eugenio Fonzi, Gianluca Tedaldi, Emanuela Scarpi, Ugo De Giorgi, Paola Ulivi (Writing-original draft). Michela Pallesch, Alessandra Virga, Emanuela Scarpi, Eugenio Fonzi, Filippo Merloni, Samanta Sarti, Rita Danesi, Mila Ravegnani, Chiara Casadei, Marianna Sirico, Caterina Gianni., Roberta Maltoni, Filippo Merloni, Sara Bravaccini, Daniele Calistri, Valentina Arcangeli, Valentina Zampiga, Ilaria Cangini, Erika Bandini, Eugenio Fonzi, Giovanni Martinelli, Ugo De Giorgi, Paola Ulivi, and Gianluca Tedaldi (Writing—review & editing).

Conflicts of Interest

U.D.G.: Consultant: Janssen, AstellasPharma, Sanofi, Bayer, Pfizer, Bristol-Myers Squibb, Novartis, Ipsen, Merck; Institutional funding: Roche, Sanofi, AstraZeneca. M.P.: reports honoraria for educational events/materials from Novartis, Daiichy-Sankyo, Gilead and travel, accommodations, and/or expenses from Grant from Novartis, Astrazeneca and Lilly. A.M.: reports research funding: Lilly, Roche, Pfizer Scientific Advisory Board: Lilly, Roche, MSD, Daiichi.

Data availability

The data that support the findings of this study are available in the supplementary material of this article, and additional data are available upon request. No restrictions apply.

Ethics approval

All methods used in our studies involving human participants adhered strictly to the ethical guidelines of our institutional

research committee, aligning with the principles outlined in the 1964 Helsinki declaration and its subsequent modifications, or equivalent ethical norms. This prospective study, with protocol code IRST B112 (L3P2210), was approved on 30/07/2020 by the Institutional Review Board of IRCCS Istituto Romagnolo per lo Studio dei Tumori (IRST) "Dino Amadori," Meldola, Italy.

Supplementary material

Supplementary material is available at *The Oncologist* online.

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