

sampling and measurement will allow progress in development of methods to minimise personal exposure to aeroallergen.

References

- Wahn U, Lau S, Bergmann R *et al*. Indoor allergen exposure is a risk factor for sensitisation during the first three years of life. *J Allergy Clin Immunol* 1997; **99**: 763-769.
- Platts-Mills TAE, *et al*. Indoor allergens and asthma: Report of the Third International Workshop. *J Allergy Clin Immunol* 1997; **100**: S1-S24.
- van der Heide S, *et al*. Seasonal differences in airway hyperresponsiveness in asthmatic patients: relationship with allergen exposure and sensitization to house dust mites. *Clin Exp Allergy* 1997; **27**: 627-633.
- Custovic A, *et al*. Effect of environmental manipulation in pregnancy and early life on respiratory symptoms and atopy during first year of life: a randomised trial. *Lancet* 2001; **358**: 188-193.
- Harving H, *et al*. House dust mite and atopic dermatitis: a case-control study on the significance of house dust mites as etiologic allergens in atopic dermatitis. *Ann Allergy* 1990; **65**: 25-31.
- Platts-Mills TAE, de Weck AL, *et al*. Dust mite allergens and asthma - a worldwide problem. Report of an International Workshop, Bad Kreuznach, Federal Republic of Germany, September 1987. *J Allergy Clin Immunol* 1989; **83**: 416-427.
- Arlian L. Water balance and humidity requirements of house dust mites. *Exp Appl Acarol* 1992; **16**: 15-35.
- Tovey ER, Chapman MD, Platts-Mills TAE. The distribution of house dust mite allergen in the houses of patients with asthma. *Am Rev Respir Dis* 1981; **124**: 630-635.
- Ingram JM, *et al*. Quantitative assessment of exposure to dog (Can F) and cat (Fel d) allergens: relationship to sensitization and asthma among children living in Los Alamos, New Mexico. *J Allergy Clin Immunol* 1995; **96**: 1-26, 449-456.
- Munir A, *et al*. Cat (Fel d 1), dog (Can f 1) and cockroach allergens in homes of asthmatic children from three climatic zones in Sweden. *Allergy* 1994; **49**: 508-516.
- Rosenstreich DL, *et al*. The role of cockroach allergy and exposure to cockroach allergen in causing morbidity among inner-city children with asthma. *N Engl J Med* 1997; **336**(19): 1356-1363.
- Quirce S, *et al*. Major cat allergen (Fel D1) levels in the homes of patients with asthma and their relationship to sensitization to cat dander. *Ann Allergy Asthma Immunol* 1995; **75**(4): 325-330.
- Custovic A, *et al*. Distribution, aerodynamic characteristics, and removal of the major cat allergen Fel d 1 in British homes. *Thorax* 1998; **53**(1): 33-38.
- Mitakakis T, *et al*. Personal exposure to allergenic pollen and mould spores in inland New South Wales, Australia. *Clin Exp Allergy* 2000; **30**: 1733-1739.
- Wood RA, *et al*. Antigenic analysis of household dust samples. *Am Rev Respir Dis* 1988; **137**(2): 358-363.
- von Mutius E, *et al*. Exposure to endotoxin or other bacterial components might protect against the development of atopy. *Clin Exp Allergy* 2000; **30**: 1230-1235.
- Eggleston PA, Bush RK. Environmental allergen avoidance: an overview. *J Allergy Clin Immunol* 2001; **107**(3): S403-S405.
- Gotzsche P, Hammarquist C, Bur M. House dust mite control measures in the management of asthma: meta-analysis. *Br Med J* 1998; **317**: 1105-1110.
- Platts-Mills T, *et al*. Sensitisation, asthma and modified Th2 response in children exposed to cat allergen a population based cross sectional study. *Lancet* 2001; **357**(9258): 752-756.
- De Lucca S, Tovey ER. Quantitative modelling of domestic allergen Der p 1 using amplified ELISA and nasal sampling. *J Allergy Clin Immunol* 2001; **107**(2): S219.
- Arlian L, Platts-Mills T. The biology of house dust mite allergens in allergic disease. *J Allergy Clin Immunol* 2001; **107**(3): S406-S413.
- Price JA, *et al*. Measurement of airborne mite antigen in homes of asthmatic children. *Lancet* 1990; **336**: 895-897.
- Oliver J, *et al*. Allergen levels in airborne and surface dust. *Int Arch Allergy Immunol* 1995; **107**(1-3): 452-453.
- Sakaguchi M, *et al*. Measurement of airborne mite allergen exposure in individual subjects. *J Allergy Clin Immunol* 1996; **97**(5): 1040-1044.
- Bollinger M, *et al*. Measurement of cat allergen levels in the home by use of an amplified ELISA. *J Allergy Clin Immunol* 1998; **101**(1): 124-125.
- O'Meara T, Tovey ER. Monitoring personal allergen exposure. In: Gershwin M, ed. *Clinical Reviews in Allergy and Immunology*. Humana Press, 2000: 341-395.
- Siebers R, *et al*. Reduction in airborne house dust mite allergen (Der p 1) after occlusive covering of bedding. *Allergy Clin Immunol Int* 2000; **2**: 54.
- Arlian L, *et al*. Lowering humidity in homes reduces dust mites and their allergens. *J Allergy Clin Immunol* 2001; **107**: 99-104.
- Owen S, *et al*. Control of house dust mite antigen in bedding. *Lancet* 1990; **335**: 396-397.
- Arlian LG, Veselica MM. Re-evaluation of the humidity requirements of the house-dust mite *Dermatophagoides farinae* (Acari: Pyroglyphidae). *J Med Entomol* 1981; **18**: 351-352.
- Colloff MJ, Taylor C, Merrett TG. The use of domestic steam cleaning for the control of house dust mites. *Clin Exp Allergy* 1995; **25**(11): 1061-1066.
- Tovey ER, Woolcock AJ. Direct exposure of carpets to sunlight can kill all mites. *J Allergy Clin Immunol* 1993; **93**: 1072-1074.
- Colloff MJ. Use of liquid nitrogen in the control of house dust mite populations. *Clin Allergy* 1986; **16**: 41-47.
- Woodfolk JA, *et al*. Chemical treatments of carpets to reduce allergen: comparison of effects of tannic acid and other treatments on proteins derived from dust mites and cats. *J Allergy Clin Immunol* 1995; **96**: 325-333.
- Tovey ER, *et al*. Changes in mite allergen Der p I in house dust following spraying with a tannic acid/acaricide solution. *Clin Exp Allergy* 1992; **22**: 67-74.
- Vanlaar CH, *et al*. Domestic control of house dust mite allergen in children's beds. *J Allergy Clin Immunol* 2000; **105**: 1130-1133.
- Rains N, *et al*. House dust mite allergen (Der p 1) accumulation on new synthetic and feather pillows. *Clin Exp Allergy* 1999; **29**: 182-185.
- Hallam C, *et al*. House dust mite allergens in feather and synthetic pillows. *Allergy* 1998; **54**(1): 407-408.
- Kemp TJ, *et al*. House dust mite allergen in pillows. *Br Med J* 1996; **313**: 916.
- De Lucca S, O'Meara T, Tovey E. Exposure to mite and cat allergens on a range of clothing items at home and the transfer of cat allergen in the workplace. *J Allergy Clin Immunol* 2000; **106**: 874-879.
- Vojta P, *et al*. Effects of physical interventions on house dust mite allergen levels in carpet, bed, and upholstery dust in low income, urban homes. *Environ Health Perspect* 2001; **109**(8): 815-819.
- Lewis R, *et al*. Factors affecting the retention of dust mite allergen on carpet. *Am Ind Hyg Assoc J* 1998; **59**: 606-613.
- Vyszynski-Moher D, *et al*. Management of house dust mites and their allergens by benzyl benzoate products. *Ann Allergy Asthma Immunol* 2000; **84**: 136.
- Woodfolk JA, *et al*. Chemical treatment of carpets to reduce allergen: a detailed study of the effects of tannic acid on indoor allergens. *J Allergy Clin Immunol* 1994; **94**: 19-26.
- Warner JA. Double blind trial of ionisers in children with asthma sensitive to the house dust mite. *Thorax* 1993; **48**: 330-333.
- Harving H, Korsgaard J, Dahl R. Mechanical ventilation in dwellings as preventive measures in mite asthma. *Allergy Proc* 1988; **9**: 283.
- Tovey ER, *et al*. Distribution of mite allergen Der p I in Australian communities. *J Allergy Clin Immunol* 1995; **95**: 325.
- Girgis S, *et al*. Thunderstorm associated asthma in an inland town in south eastern Australia. *Eur Respir J* 2000; **16**(1): 3-8.
- Platts-Mills TAE, *et al*. Reduction of bronchial hyperreactivity during prolonged allergen avoidance. *Lancet* 1982; **25**: 675-678.

Role of house-dust endotoxin exposure in aetiology of allergy and asthma

Olivier Michel

Clinic of Allergology and Respiratory Diseases, Saint-Pierre University Hospital (ULB, Free University of Brussels), Rue Haute 322, B-1000 Brussels, Belgium

Tel: +32 2 535 42 03

Fax: +32 2 535 41 74

E-mail: omichel@ulb.ac.be

Endotoxin and its purified derivative lipopolysaccharide (LPS) are Gram-negative bacterial potent pro-inflammatory constituents continuously shed into the environment.¹ A number of different Gram-negative bacteria inhabits the normal body surfaces including the skin, oral cavity, respiratory tract, gastrointestinal tract, vagina and urinary tract. Humans can be exposed to endotoxin via several ways. In addition to the septic shock frequently

caused by translocation of Gram-negative microorganisms normally present in the gut of the host to the circulation, there is continuous exposure to airborne endotoxin. The release of endotoxin from Gram-negative bacteria that colonise the respiratory tract in the majority of patients with chronic bronchitis can contribute to the lung function decrease by initiating release of inflammatory mediators from bronchial epithelial cells.² High levels of airborne (up to $1 \mu\text{g}/\text{m}^3$) endotoxin have been reported from a variety of occupational environments (e.g. swine confinement, poultry farm, cotton mill, brewery, waste processing).¹ A number of cross-sectional studies reports an association between exposure to endotoxin measured in the dust from those occupational settings and the risk to develop non-atopic chronic obstructive pulmonary diseases,³⁻⁶ toxic pneumonitis and systemic effects.^{7,8} In the domestic environment, there is also endotoxin contaminating house dust⁹⁻¹² that, by itself or in association with allergen exposure, could be an important determinant of asthma severity.^{9,10} Recently, Hasday *et al.* reported that high levels of endotoxin are produced by cigarette smoke.¹³

Inhalation of pure endotoxin may elicit, in some individuals, dyspnea, chest tightness, myalgia, shivers, fatigue and malaise associated or not with fever.^{14,15} A similar clinical response is observed after exposure to dust containing endotoxin such as grain handlers,¹⁶ cotton workers,⁵ fibre-glass manufacturing employees,⁸ or animal farmers.¹⁷ A large inter-individual variability in the sensitivity to endotoxin has been reported.¹⁸⁻²⁰ In humans, inhalation of pure endotoxin is associated with bronchoconstriction,^{14,15,18-21} change in the level of non-specific airways responsiveness,^{14,20,21} and reduction in alveolar-capillary diffusion.²² Compared with asthmatics, in normal subjects a higher dose of pure endotoxin is required to produce bronchoconstriction.^{14,15,18,19,23} Although the endotoxin response is reproducible in a given subject,¹⁹ there is a large between-subjects variability^{14,18,19,20} at least partially related to the airways inflammatory status²⁴ and to the level of non-specific airways responsiveness.²⁵

Local and systemic inflammatory responses have been measured after endotoxin inhalation in normal and asthmatic subjects. Significant blood leukocytosis and neutrophilia were observed 4-8 h after inhalation of endotoxin both in normal^{14-16,18,22,26} and asthmatic subjects.^{21,27} This neutrophilia was not related with the change in lung function.^{21,27} *In vitro*, small amounts of endotoxin ($< 1 \text{ ng}/\text{ml}$) activate human airways macrophages, releasing several pro-inflammatory cytokines (tumour necrosis factor- α (TNF- α), interleukin (IL)-1, IL-6) and metabolites of arachidonic acid. The presence of LPS-binding protein and the soluble fraction of CD14 receptor (sCD14) in the airways²⁴ may increase the macrophage activation by

endotoxin.^{24,28} Six hours after an inhalation of endotoxin-contaminated dust, high concentrations of IL-1, IL-1 RA, IL-6, IL-8 and TNF- α and their specific mRNAs were measured in the bronchoalveolar lavage.²⁹ These cytokines are potential activators of the hepatic acute-phase protein response, consistent with the rise in the blood concentration of the C-reactive protein (CRP) 24 h after endotoxin inhalation.^{13,17,21,26} We speculate that cytokines produced into the airways are released in the blood and stimulate the hepatocytes.

Airway inflammation characterized by neutrophil recruitment in bronchoalveolar lavage (BAL) was observed after bronchial challenge with endotoxin-contaminated dusts like allergen extracts,³⁰ grain dust²⁹ and swine dust,¹⁷ while in normal subjects 100 μg of inhaled pure endotoxin induced a 100-fold increase in neutrophils from BAL.³¹ A significant increase in neutrophils measured in the induced sputum occurred after 5^{15,32} to 60 μg ³³ endotoxin. The sputum concentrations in myeloperoxidase (MPO) (from neutrophils), eosinophil cationic protein (ECP) (from eosinophils) and TNF- α rose significantly 6 h after endotoxin.¹⁵

There are some published data suggesting that environmental endotoxin could be a synergic factor on the amplitude of immunoglobulin E-mediated response. On one hand, in allergic mild asthmatics, an exposure to air containing low levels of endotoxin ($250 \text{ ng}/\text{m}^3$) for 4 h before bronchial challenge with allergen increases significantly both bronchial reactivity and antigen-induced airway eosinophilia.³⁴ The airways cellular inflammation to inhaled allergen is modified by endotoxin contamination of the allergen extract. Indeed, while detoxified pure allergen extract results in bronchial eosinophil recruitment, endotoxin contamination ($1 \text{ ng}/\text{ml}$) causes neutrophilia.³⁰ On the other hand, inhalation of allergen in sensitized subjects leads to airways plasma exsudation including extravasation of sCD14 and lipopolysaccharide-binding protein (LBP).²⁴ These proteins may enhance the capacity of inhaled endotoxin to activate an inflammatory cascade that may amplify the inflammatory response to inhaled antigen in some asthmatics, as was suggested by several field studies.^{9,10,35} In the home environment, the amount of endotoxin in house dust has been related to the severity of asthma both in atopic^{9,10} and non-atopic subjects.³⁵ In dust-mite-sensitized subjects, the level of exposure to mite allergen was higher in subjects with asthma than in those with rhinitis, while the severity of the asthmatic disease was significantly associated with a low forced expiratory volume in 1 sec (FEV1) and FEV1/forced vital capacity, and the daily need for oral and topical corticosteroid, as well as with the asthma score.⁹ More recently, Douwes *et al.*³⁶ did not find an association between endotoxin exposure and peak

expiratory flow (PEF) variability in a group of children defined by asthma symptoms. However, the daily PEF variability was very low (6.4%), suggesting asthma was intermittent or doubtful. Therefore, endotoxin should be considered as an enhancing rather than inducing factor in asthma.

We recently challenged 15 normal subjects with inhaled endotoxin (0.5, 5, 50 µg).¹⁸ Subjects who developed significant increase in body temperature had a larger increase in the systemic inflammatory response (blood neutrophilia and blood concentrations of CRP and LBP), while subjects who developed a significant increase in airways responsiveness had an increase in the sputum concentration of ECP. The amplitude of the systemic response and decrease in FEV1 were inversely associated with the atopic status, suggesting a link between atopy and LPS responsiveness. This observation reinforces the hypothesis for a mechanism linking the macrophage susceptibility to LPS stimulation with the increase in macrophage production of cytokines that inhibits the T helper cell (Th)-2 response and, consequently, the risk to become atopic. Environmental exposure to LPS and other bacterial wall products, present in house dust^{9-12,35,36} and/or from the intestinal tract,³⁷ could be a necessary step for maturation the immune system and the development of a Th1-like response through the presentation of antigen in conjunction with IL-12.³⁸⁻⁴⁰ In mice, endotoxin sensitivity is genetically determined, involving mutation in the Toll-like receptor-4 (TLR4) gene,⁴¹ a co-receptor essential for the LPS signalling. In human, recent data suggest that polymorphisms in the genes encoding the TLR4⁴² or the LPS receptor CD14⁴³ may be related to symptoms and diseases. exposure to endotoxin in early life could be protective for the risk of atopy, while in symptomatic asthma it could be a risk factor of a severe disease.

Finally, available data on the protective effect of anti-asthmatic drugs on the endotoxin-induced response are quite limited. An acute pre-treatment with sodium cromoglycate⁴⁴ or with short or long acting B₂-agonists completely prevents the bronchoconstriction induced by an acute exposure to LPS⁴⁵ while, given in a single dose, an inhaled corticosteroid does not prevent the endotoxin-induced blood inflammation.⁴⁵ Studies are in progress to evaluate the possible protective effect of chronic treatment with oral corticosteroids on the response to endotoxin.

References

- Rylander R, ed. Endotoxins in the environment: a criteria document. *Int J Occup Environ Health* 1997; **3** (Suppl 1): S1-S48.
- Kheir OA, Davis RJ, Devalia JL. Bacterial-induced release of inflammatory mediators by bronchial epithelial cells. *Eur Respir J* 1996; **9**: 1913-1922.
- Castellan RM, Olenchock SA, Kinsley KB, Hankinson JL. Inhaled endotoxin and decreased spirometric values. *N Engl J Med* 1987; **317**: 605-610.
- Theelin A, Tegler O, Rylander R. Lung reaction during poultry handling related to dust and bacterial endotoxin levels. *Eur J Respir Dis* 1984; **65**: 266-271.
- Rylander R, Haglund P, Lundholm M. Endotoxin in cotton dust and respiratory function decrement among cotton workers in an experimental cardroom. *Am Rev Respir Dis* 1985; **131**: 209-213.
- Vogelzang PFJ, van der Gulden JWJ, Folgering H, et al. Endotoxin exposure as a major determinant of lung function decline in pig farmers. *Am J Respir Crit Care Med* 1998; **157**: 15-18.
- Von Essen S, Robbins RA, Thompson AB, Rennan SI. Organic dust toxic syndrome: an acute febrile reaction to organic dust exposure distinct from hypersensitivity pneumonitis. *J Toxicol Clin Toxicol* 1990; **28**: 389-420.
- Milton DK, Amsel J, Reed CE, Enright PL, Brown LR, Aughenbaugh GL, Morey PR. Cross-sectional follow-up of a flu-like respiratory illness among fiberglass manufacturing employees: endotoxin exposure associated with two distinct sequelae. *Am J Ind Med* 1995; **28**: 469-488.
- Michel O, Kips J, Duchateau J, et al. Severity of asthma is related to endotoxin in house dust. *Am J Respir Crit Care Med* 1996; **154**: 1641-1646.
- Rizzo MC, Naspitz CK, Fernandez-Caldas E, Lockey RF, Mimi a I, Solé D. Endotoxin exposure and symptoms in asthmatic children. *Pediatr Allergy Immunol* 1997; **8**: 121-126.
- Gereda JE, Leung DYM, Thatayatikom A, Streib JE, Price MR, Klinnert MD, Liu AH. Relation between house-dust endotoxin exposure, type 1 T-cell development, and allergen sensitisation in infants at high risk of asthma. *Lancet* 2000; **355**: 1680-1683.
- von Mutius E, Braun-Fahrlander, Schierl R, et al. Exposure to endotoxin or other bacterial components might protect against the development of atopy. *Clin Exp Allergy* 2000; **30**: 1205-1212.
- Hasday JD, Bascom R, Costa JJ, Fitzgerald T, Dubin W. Bacterial endotoxin is an active component of cigarette smoke. *Chest* 1999; **115**: 829-835.
- Rylander R, Bake B, Fisher JJ, Helander JM. Pulmonary function and symptoms after inhalation of endotoxin. *Am Rev Respir Dis* 1989; **140**: 981-986.
- Michel O, Nagy A-M, Schroeven M, Duchateau J, Nève J, Fondu P, Sergysels R. Dose-response relationship to inhaled endotoxin in normal subjects. *Am J Respir Crit Care Med* 1997; **156**: 1157-1164.
- Jagiello PJ, Thorne PS, Watt JL, Frees KL, Quinn TJ, Schwartz DA. Grain dust and endotoxin inhalation challenges produce similar inflammatory response in normal subjects. *Chest* 1996; **110**: 263-270.
- Larsson KA, Eklund AG, Hansson LO, Isaksson BM, Malmberg PO. Swine dust causes intense airways inflammation in healthy subjects. *Am J Respir Crit Care Med* 1994; **150**: 973-977.
- Michel O, Dentener M, Corazza F, Buurman W, Rylander R. Healthy subjects express differences in clinical responses to inhaled lipopolysaccharide which are related with inflammation and with atopy. *J Allergy Clin Immunol* 2001; **107**: 797-804.
- Kline JN, Cowden J, Hunninghake GW, et al. Variable airway responsiveness to inhaled lipopolysaccharide. *Am J Respir Crit Care Med* 1999; **160**: 297-303.
- Michel O, Duchateau J, Sergysels R. Effect of inhaled endotoxin on bronchial reactivity in asthmatic and normal subjects. *J Appl Physiol* 1989; **66**: 1059-1064.
- Michel O, Ginanni R, Le Bon B, Content J, Duchateau J, Sergysels R. Inflammatory response to acute inhalation of endotoxin in asthmatic patients. *Am Rev Respir Dis* 1992; **146**: 352-357.
- Herbert A, Carvalho M, Rubenowitz E, Bake B, Rylander R. Reduction of alveolar-capillary diffusion after inhalation of endotoxin in normal subjects. *Chest* 1992; **102**: 1095-1098.
- Cavagna G, Foa V, Vigliani EC. Effects in man and rabbits of inhalation of cotton dusts or extracts and purified endotoxin. *Br J Ind Med* 1969; **26**: 314-321.
- Dubin W, Martin TR, Swoveland P, et al. Asthma and endotoxin: lipopolysaccharide-binding protein and soluble CD14 in bronchoalveolar compartment. *Am J Physiol* 1996; **270**: L736-L744.
- Michel O, Ginanni R, Sergysels R. Relation between the bronchial obstructive response to inhaled lipopolysaccharide and bronchial responsiveness to histamine. *Thorax* 1992; **47**: 288-291.
- Michel O, Duchateau J, Plat G, Cantinieux B, Hotimsky A, Gerain J, Sergysels R. Blood inflammatory response to inhaled endotoxin in normal subjects. *Clin Exp Allergy* 1995; **25**: 73-79.
- van der Zwan JC, Orié NG, Kauffman HF, Wiers PWJ, De Vries K. Bronchial obstructive reactions after inhalation with endotoxin and precipitinogens of *Haemophilus influenzae* in patients with chronic non specific lung disease. *Clin Allergy* 1982; **12**: 547-559.
- Martin TR, Mathison JC, Tobias PS, Leturcq DJ, Moriarty AM, Maunder RJ, Ulevitch RJ. Lipopolysaccharide binding protein enhances the responsiveness of alveolar macrophages to bacterial lipopolysaccharide. *J Clin Invest* 1992; **90**: 2209-2219.
- Clapp WD, Becker S, Quay J, et al. Grain dust-induced airflow obstruction and inflammation of the lower respiratory tract. *Am J Respir Crit Care Med* 1994; **150**: 611-617.

30. Hunt LW, Gleich GJ, Ohnishi T, Weiler DA, Mansfield ES, Kita H, Sur S. Endotoxin contamination causes neutrophilia following pulmonary allergen challenge. *Am J Respir Crit Care Med* 1994; **149**: 1471-1475.
31. Sandstrom T, Bjermer L, Rylander R. Lipopolysaccharide (LPS) inhalation in healthy subjects increases neutrophils, lymphocytes and fibronectin levels in bronchoalveolar lavage fluid. *Eur Respir J* 1992; **5**: 992-996.
32. Thorn J, Rylander R. Inflammatory response after inhalation of bacterial endotoxin assessed by the induced sputum technique. *Thorax* 1998; **53**: 1047-1052.
33. Nightingale JA, Rogers DF, Hart LA, Kharitonov SA, Chung KF, Barnes PJ. Effect of inhaled endotoxin on induced sputum in normal, atopic, and atopic asthmatic subjects. *Thorax* 1998; **53**: 563-571.
34. Boehlecke BA, Peden D, Hazucha M, Alexis N, Tucker K, Bromberg P. Exposure to low level of endotoxin for four hours increased response to inhaled mite allergen in mild asthmatics [abstract]. *Am J Respir Crit Care Med* 1999; **159**: A699.
35. Michel O, Ginnani R, Duchateau J, Vertongen F, Le Bon B, Sergysels R. Domestic endotoxin exposure and clinical severity of asthma. *Clin Exp Allergy* 1991; **21**: 441-448.
36. Douwes J, Zuidhof A, Doekes G, van der Zee S, Wouters I, Boezen HM, Brunekreef B. (1-3)- β -D-Glucan and endotoxin in house dust and peak flow variability in children. *Am J Respir Crit Care Med* 2000; **162**: 1348-1354.
37. Sudo N, Sawamura S, Tanaka K, Aiba Y, Kubo C, Koga Y. The requirement of intestinal bacterial flora for the development of an IgE production system fully susceptible to oral tolerance induction. *J Immunol* 1997; **159**: 1739-1745.
38. Martinez FD. Maturation of immune responses at the beginning of asthma. *J Allergy Clin Immunol* 1999; **103**: 355-361.
39. Holt PG, Sly PD, Björkstén B. Atopic versus infectious diseases in childhood: a question of balance? *Pediatr Allergy Immunol* 1997; **8**: 53-58.
40. Liu AH, Leung DY. Modulating the early allergic response with endotoxin. *Clin Exp Allergy* 2000; **30**: 1535-1539.
41. Poltorak A, He X, Smirnova I, et al. Defective LPS signaling in C3H/HeJ and C57BL/10ScCr mice: mutations in *Tlr4* gene. *Science* 1998; **282**: 2085-2088.
42. Arbour NC, Lorenz E, Schutte BC, et al. TLR4 mutations are associated with endotoxin hyporesponsiveness in humans. *Nat Genet* 2000; **25**: 187-191.
43. M Baldini, IC Lohman, M Halonen, RP Erickson, PG Holt, FD Martinez. A polymorphism in the 5' (flanking region of the CD14 gene is associated with circulating soluble CD14 levels and with total serum Immunoglobulin E. *Am J Respir Cell Mol Biol* 1999; **20**: 976-983.
44. Michel O, Ginanni R, Sergysels R. Protective effect of sodium cromoglycate on lipopolysaccharide-induced bronchial obstruction in asthmatics. *Int Arch Allergy Immunol* 1995; **108**: 298-302.
45. Michel O, Olbrecht J, Moulard D, Sergysels R. Effect of anti-asthmatic drugs on the response to inhaled endotoxin. *Ann Allergy Asthma Immunol* 2000; **85**: 305-310.

Asthma, atopy, antibiotics and the bowel

Julian Crane

Department of Medicine, School of Medicine,
Mein Street, Newtown, Wellington, New
Zealand

Tel: +64 4 385 5999 5258

Fax: +64 4 389 5427

E-mail: crane@wnmeds.ac.nz

The past three decades have seen an increase in reported asthma and allergic diseases from many studies around the world, recently described as an epidemic.¹ While many hypotheses have been developed to explain these changes, the hygiene hypothesis has for the past decade encompassed an expanding link between the epidemiology and immunology of both atopic sensitisation and atopic

diseases. These associations and the utility of the hypothesis have recently been reviewed by David Strachan,² who first reported the associations between birth order and hayfever, and articulated the hygiene hypothesis as an explanation.³ Inverse relationships between atopic and infectious disease was first raised a decade earlier by Gerrard *et al.* in a comparison of atopic disease amongst the Metis (native Indian) and white communities of northern Saskatchewan.⁴ The immunological basis for the hypothesis rests on the concept of immune deviation in early life towards T helper cell (Th)1 immune responses induced by microbial exposure, with Th1 responses suppressing Th2 responses and immunoglobulin E (IgE) production. The hypothesis therefore refers to IgE-mediated diseases such as hayfever but is less applicable to asthma, where atopy plays an important but not exclusive role.

The relationships between asthma, bronchial hyperresponsiveness (BHR) and atopy have recently been examined among 20- to 44-year-old adults, in five Spanish centres involved in the European Community Respiratory Health Survey. The adjusted proportion of BHR attributable to atopy was 21% and the proportion of asthma symptoms and BHR attributable to atopy was 42%.⁵ Factors associated with the hygiene hypothesis such as birth order or specific infections will vary in their strength of association with asthma depending on the proportion of asthma attributable to atopy. An important feature of any useful hypothesis is that it should unify disparate observations. The hygiene hypothesis does this, suggesting explanations for socio-economic variations in atopic disease both within and between countries, and a plausible explanation of some of the long-term upward trends in prevalence. Studies of the relationship between infection or microbial exposures and atopic disease also tend to support the hygiene hypothesis.

The influence of antibiotics on these associations has recently been studied. Farooqi *et al.* found a twofold risk of doctor-diagnosed atopic diseases with antibiotic treatment in the first 2 years of life, among a general practice birth cohort.⁶ The increased risk was apparent for all classes of antibiotics, although greater for cephalosporins and macrolides; it was independent of the underlying condition being treated, and was similar for those with and without a history of maternal atopy. Antibiotic exposure was the strongest predictor of atopic disease in this study.

The other two studies have examined antibiotic use among children in Sweden and New Zealand, whose families have some association with an anthroposophic lifestyle. Families embracing this lifestyle, whose tenets were set out by Rudolph Steiner in the nineteenth century, tend to minimise their involvement with conventional medical