Alternative splicing factors and cardiac disease: more than just missplicing?

ZACHERY R. GREGORICH and WEI GUO

Department of Animal and Dairy Sciences, University of Wisconsin-Madison, Madison, Wisconsin 53706, USA

ABSTRACT

Alternative splicing (AS) is the process wherein the exons from a single gene are joined in different combinations to produce nonidentical, albeit related, RNA transcripts. This process is important for the development and physiological function of many organs and is particularly important in the heart. Notably, AS has been implicated in cardiac disease and failure, and a growing number of genetic variants in AS factors have been identified in association with cardiac malformation and/or disease. With the field poised to interrogate how these variants affect cardiac development and disease, an understandable point of emphasis will undoubtedly be on downstream target gene missplicing. In this Perspective article, we would like to encourage consideration not only of the potential for novel disease mechanisms, but also for contributions from disruption of the ever-expanding list of nonsplicing functions ascribed to many AS factors. We discuss the emergence of a novel cardiac disease mechanism based on pathogenic RNA granules and speculate on the generality of such a mechanism among localization-disrupting AS factor genetic variants. We also highlight emerging nonsplicing functions attributed to several AS factors with cardiac disease-associated genetic variants in the hopes of pointing to avenues for exploration of mechanisms that may contribute to disease alongside target gene missplicing.

Keywords: alternative splicing; cardiomyopathy; RBM20; RNA granules

INTRODUCTION

Alternative splicing (AS) is an important posttranscriptional mechanism that plays key roles in generating protein diversity (Nilsen and Graveley 2010). It is estimated that over 95% of multiexon genes are regulated via AS (Pan et al. 2008; Wang et al. 2008). Additionally, AS can regulate gene expression through nonsense-mediated decay (Lewis et al. 2003). It has been found that AS is important not only for coding genes but also for long noncoding RNAs as well (Deveson et al. 2018). For an overview of regulatory mechanisms in AS, the reader is referred to the many excellent reviews published on this topic (Licatalosi and Darnell 2010; Irimia and Blencowe 2012; Kornblihtt et al. 2013; Fu and Ares 2014; Baralle and Giudice 2017; Kastner et al. 2019; Shenasa and Hertel 2019; Ule and Blencowe 2019; Gordon et al. 2021). AS is crucial for the development of multiple organs but contributes the greatest to the development of the brain and striated muscles, including cardiac muscle (Wang et al. 2008). In fact, cardiac genes, such as TNNT2, which encodes the cardiac-specific isoform of troponin T, were among the first found to be

regulated by this process (Medford et al. 1984). Subsequent transcriptome-wide studies have revealed that broad and dramatic changes in protein isoform expression, brought about through AS, are crucial for the development of the heart (Kalsotra et al. 2008; Giudice et al. 2014). This importance is further underscored by the fact that the genetic ablation of various AS factors in the hearts of rodents either manifests or predisposes the heart to dysfunction and/or failure, as reviewed elsewhere (Montañés-Agudo et al. 2023; Cao et al. 2024).

Given the important role that AS plays in the development and function of the heart, it is perhaps not surprising that this process has also been implicated in cardiac disease and failure (Kong et al. 2010; Lee et al. 2011; Song et al. 2012). Studies have shown that certain AS factors are differentially expressed in various cardiomyopathies, as well as in heart failure, with a recent example being SLM2, which is upregulated in dilated cardiomyopathy (DCM) (Boeckel et al. 2022). Furthermore, a growing number of genetic variants in AS factors have been identified in human patients in association with cardiac malformation and/or disease (Brauch et al. 2009; Johnston et al. 2010; Homsy et al. 2015; Wang et al. 2016; Au et al. 2018; Yu

Corresponding author: wguo2@wisc.edu

Article is online at http://www.rnajournal.org/cgi/doi/10.1261/rna .080332.124. Freely available online through the RNA Open Access option.

© 2025 Gregorich and Guo This article, published in RNA, is available under a Creative Commons License (Attribution 4.0 International), as described at http://creativecommons.org/licenses/by/4.0/.

et al. 2018; Chen et al. 2024); and this number will undoubtedly continue to rise as high-throughput sequencing becomes increasingly affordable and achieves greater use in clinical diagnostics (Pasche and Absher 2011).

When seeking to establish the molecular mechanisms associated with disease-causing genetic variants in AS factors, a logical first step is to focus on the role that downstream target gene missplicing plays in pathobiology as exemplified by our own studies (Guo et al. 2012). While this is an understandable and likely fruitful place to begin, it is important to keep in mind that this may be only one part of the equation. To exemplify this, we provide a brief overview of our evolving understanding of the molecular mechanisms in cardiac disease caused by pathogenic genetic variants in the AS factor RNA-binding motif protein 20 (RBM20), often referred to as RBM20 cardiomyopathy (Gregorich et al. 2024). We highlight the identification of pathogenic RBM20 granules as a causative agent in cardiac disease and discuss whether such a mechanism could be shared among localization-disrupting variants in AS factors. We conclude by pointing out several additional nonsplicing functions of AS factors with genetic variants implicated in cardiac malformation and disease, beginning with RBM20. Importantly, the purpose here is not to arque that target gene missplicing downstream from AS factor variants does not contribute to cardiac pathobiology and disease. Instead, we wish to encourage investigation of other mechanisms and processes that may also be involved.

Pathogenic cytoplasmic granules and missplicing: dual contributors to RBM20 cardiomyopathy

Loss of the Rbm20 gene, which encodes the musclespecific splicing factor RBM20, was identified by our laboratory over a decade ago as the cause of a DCM-like phenotype in a rat strain deficient in splicing of the giant sarcomeric protein titin (Guo et al. 2012). We immediately recognized that the phenotype in Rbm20 knockout (KO) rats resembled, at least in some respects, that previously reported in individuals with DCM caused by pathogenic genetic variants in RBM20 with cardiac remodeling, increased susceptibility to arrhythmia, and premature mortality (Brauch et al. 2009). Beyond titin, this study and others identified approximately 30 other genes subject to AS regulated by RBM20 (Guo et al. 2012; Maatz et al. 2014). Critically, these included several genes important for the regulation of Ca²⁺-handling, such as Camk2d, Ryr2, and Trdn (Guo et al. 2012; Maatz et al. 2014). Hence, a putative mechanistic basis for RBM20 cardiomyopathy was established based on target gene missplicing. Specifically, this disease was presumed to arise because of (1) reduced ventricular wall tension resulting from altered titin splicing and (2) impaired contractility secondary to changes in the splicing of genes important for Ca²⁺-handling (Guo et al. 2012, 2018, 2021; Linke and Bücker 2012; Methawasin et al. 2014). This splicing-centric mechanism received further support with the demonstration that *Rbm20* KO mice phenocopy KO rats with DCM and proarrhythmic changes expected to underlie the propensity for arrhythmias in individuals carrying pathogenic *RBM20* genetic variants (van den Hoogenhof et al. 2018).

Yet, it quickly became apparent that not all data supported target gene missplicing as the sole mechanism of disease pathogenesis in RBM20 cardiomyopathy. Firstly, there was a pronounced phenotypic discrepancy between Rbm20 KO rodents and patients carrying pathogenic genetic variants in RBM20, with the latter exhibiting a much more severe phenotype. To illustrate, even homozygous Rbm20 KO rats do not exhibit signs of systolic dysfunction and approximately 83% of rats live over 18 months without developing heart failure (Guo et al. 2012). Conversely, all reported human carriers of pathogenic genetic variants in RBM20 are heterozygous and most develop severe disease, as evidenced by the requirement for heart transplantation at earlier age in RBM20 variant carriers compared to that in carriers of pathogenic genetic variants in other DCM-associated genes (Kayvanpour et al. 2017). Secondly, mice expressing RBM20 lacking the RRM, which renders the protein splicing deficient, did not develop DCM despite similar missplicing of major RBM20 target genes like Ttn and Camk2d (Methawasin et al. 2014). This lack of phenotype received further support recently with the generation of mice expressing a variant of uncertain significance (I536T) in the RRM of RBM20, previously identified in a case of sudden cardiac death (Yamamoto et al. 2019). Like mice lacking the RRM, mice homozygous for the I536T variant display marked alterations in the splicing of well-established RBM20 target genes but develop neither cardiac dysfunction nor heart failure (Yamamoto et al. 2022). Collectively, these findings suggested that the missplicing of RBM20 target genes alone is not sufficient to explain the severe phenotype in pathogenic RBM20 variant carriers.

It was not until the first Rbm20 variant knock-in (KI) animal models were generated that answers were finally provided (Ihara et al. 2020; Schneider et al. 2020). Two variant KI animals were reported simultaneously by different laboratories, each carrying a different genetic variant in a DCMassociated hotspot in exon 9 (c.1881–1920 in humans) (Parikh et al. 2019), which encodes a portion of the arginine/serine-rich (RS) domain in RBM20. Unlike rodents with Rbm20 ablation, these genetic variant KI animals developed severe DCM with cardiac dysfunction and arrhythmia, even in heterozygous carriers (Ihara et al. 2020; Schneider et al. 2020), mirroring disease in human patients. Critically, these KI animals not only unambiguously demonstrated that pathogenic variants in this hotspot are causative in severe DCM but also brought to light a novel disease paradigm in DCM based on the formation of pathological cytoplasmic RNA granules (Ihara et al. 2020; Schneider et al. 2020). It was subsequently found that these variants disrupt a critical nuclear localization signal located in the RS domain (Zhang et al. 2023a). The presence of cytoplasmic granules has now been confirmed in human patient tissue (Gaertner et al. 2020), as well as in additional animal and human-induced pluripotent stem cell-derived cardiomyocyte (hiPSC-CM) models by our laboratory and others (Wyles et al. 2016; Streckfuss-Bömeke et al. 2017; Briganti et al. 2020; Fenix et al. 2021; Lennermann et al. 2022; Nishiyama et al. 2022; Wang et al. 2022; Zhang et al. 2022; Grosch et al. 2023; Kornienko et al. 2023).

An important question remaining after this discovery is to what extent each of these mechanisms—target gene missplicing and cytoplasmic RBM20 granules—contribute to the pathogenesis of RBM20 cardiomyopathy. For instance, individuals carrying genetic variants in a second DCM-associated hotspot in RBM20 located in exon 11 (c. 2721–2760 in humans) develop DCM (Beggali et al. 2016; Gaertner et al. 2020; Robyns et al. 2020), yet these variants do not appear to block nuclear import or lead to the formation of cytoplasmic granules like those in the exon 9 hotspot (Gaertner et al. 2020; Zhang et al. 2023a). Instead, limited evidence indicates that exon 11 variants may cause disease through haploinsufficiency that gives rise to missplicing (Beqqali et al. 2016). It remains unclear whether granule formation (unique to exon 9 variants) and haploinsufficiency or missplicing (associated with both exon 9 and 11 variants) lead to phenotypic differences in human patients. However, this distinction has been observed in rodent models, and emerging evidence from studies in hiPSC-CMs suggests that similar differences may also exist in humans. Specifically, 3D-engineered heart tissue models generated with hiPSC-CMs harboring pathogenic RBM20 KI genetic variants exhibit greater contractile dysfunction than those generated with RBM20 KO hiPSC-CMs (Fenix et al. 2021).

Pathogenic RNA granules: a potential common mechanism in cardiac disease driven by localizationdisrupting genetic variants in splicing factors

Over a decade ago, pathogenic genetic variants in the FUS and TARDBP genes, which encode RNA-binding proteins, were linked to the formation of cytoplasmic granules that underlie amyotrophic lateral sclerosis (Kabashi et al. 2008; Kwiatkowski et al. 2009; Vance et al. 2009). The discovery that certain genetic variants in RBM20 also promote the formation of pathogenic granules in the cytoplasm establishes a new paradigm for cardiac disease. An important question is whether this is unique to RBM20 or whether localization-disrupting genetic variants in other RNA-binding proteins, such as AS factors, could also cause cardiac disease through such a mechanism. From a theoretical perspective, it seems likely that this is

the case, as many AS factors carry domains/regions capable of supporting the multivalent interactions necessary for granule formation (Lin et al. 2015). Intriguingly, there may already be evidence for another AS factor for which this is the case. A previous study identified a genetic variant in the AS factor Rbfox2 associated with hypoplastic left heart syndrome (HLHS) (Verma et al. 2016), a condition wherein the left ventricle does not form correctly and cannot adequately support systemic circulation. This variant results in premature truncation of the Rbfox2 protein and loss of an NLS at the extreme C terminus of the protein (Verma et al. 2016). Interestingly, such mislocalization may also occur through AS of the Rbfox2 gene as has been demonstrated in calcific tendons (Cho et al. 2019). Whether these granules are detrimental as they are in RBM20 cardiomyopathy remains to be determined, but this finding provides at least some support that variants in other AS factors may also disrupt heart formation or promote failure through pathogenic cytoplasmic granules.

Disease-associated variants in splicing factors and their roles beyond splicing

In the following section, we will highlight several AS factors with cardiac disease-associated genetic variants, beginning with RBM20, and their emerging functions beyond splicing. This is not intended to be exhaustive, but instead to point to interesting avenues for exploration of mechanisms that could contribute to cardiac disease alongside missplicing in the context of AS factor genetic variants.

RBM20

A growing list of nonsplicing functions has been uncovered for RBM20 in recent years. These include the generation of important circular RNAs from the titin gene (Khan et al. 2016), regulation of alternative polyadenylation (Fenix et al. 2021), and the formation of splicing factories (Bertero et al. 2019). It must be mentioned that these nonsplicing functions appear to depend on the splicing function of RBM20, at least to some extent. Namely, ablation of Rbm20 completely abolishes the production of circular RNAs from the titin gene in mice (Khan et al. 2016). Similarly, there appears to be some degree of coordination between AS and alternative polyadenylation (Zhang et al. 2023b). In the case of splicing factory formation, this also depends on the titin mRNA (Bertero et al. 2019), yet whether transcript splicing is required has not been investigated. Regardless, disruption of these functions has the potential to have far-reaching consequences beyond those attributable to direct missplicing of RBM20's target genes. For instance, it has been shown that while pathogenic variant KI and RBM20 KO give rise to similar missplicing events in hiPSC-CMs, there are events that are unique in the genetic variant KI cells (Fenix et al. 2021). It is tempting to speculate that this may be due to disruption of events that depend on the RBM20 splicing factories, although other mechanisms cannot be ruled out without additional studies. Therefore, how these functions are impacted by pathogenic genetic variants in the protein warrants further investigation.

Rbfox family proteins

Members of the Rbfox family of RNA-binding proteins have been shown to play a crucial role in AS events in neuronal and striated muscle, including in the heart (Jin et al. 2003; Underwood et al. 2005; Gallagher et al. 2011). The Rbfox family comprises three members with Rbfox1 and Rbfox2 being expressed in the heart (Jin et al. 2003; Underwood et al. 2005). Exome sequencing of 1213 congenital heart disease probands and their unaffected parents identified three predicted loss-of-function variants in Rbfox2 (Homsy et al. 2015). Interestingly, all three probands, as well as a previously identified proband with a de novo copy number loss encompassing RBFOX2 identified by the same laboratory (Glessner et al. 2014), manifested HLHS. Dysregulation of Rbfox2 has also been identified in patients with Type 2 diabetes (Nutter et al. 2016). While the pathogenicity of *Rbfox2* variants remains to be validated, knockdown of Rbfox2 in zebrafish gives rise to an HLHS-like phenotype (Huang et al. 2022). Similarly, conditional ablation of Rbfox2 in the embryonic mouse heart resulted in the failure to develop normal cardiac chambers (Verma et al. 2022). Current studies have focused largely on the missplicing of Rbfox2 target genes. For instance, Rbfox2 KO in animal models resulted in aberrant splicing of transcripts coding for mitochondrial, cytoskeletal, and sarcomere components (Huang et al. 2022; Verma et al. 2022). Aside from potential contributions from cytoplasmic granules in the case of variants that disrupt the C-terminal nuclear localization signal (Verma et al. 2016) as discussed above, Rbfox proteins have also been shown to play a role in microRNA biogenesis (Chen et al. 2016). MicroRNAs have emerged as important regulators of cardiac function and disease (van Rooij et al. 2006; Boon and Dimmeler 2015; Zhou et al. 2018). Thus, after establishing the pathogenicity of disease-associated variants, it will be imperative to determine how/whether pathogenic variants affect the processing and maturation of Rbfox-regulated microRNAs, as well as the downstream consequences.

CELF family proteins

Six different CELF isoforms have been identified with tissue-specific expression patterns that are developmentally regulated (Good et al. 2000; Ladd et al. 2001, 2004). Among CELF isoforms, CELF4 is expressed in multiple tissues, including in the heart (Ladd et al. 2001). Interestingly,

a genome-wide association study conducted in childhood cancer survivors identified a single nucleotide polymorphism in CELF4 that was associated with anthracycline-related cardiomyopathy (Wang et al. 2016). This SNP may affect a splice donor site in the CELF4 gene potentially leading to the production of a truncated protein (Wang et al. 2016). CELF proteins are well-known splicing regulators of a developmental transition in the AS of the TNNT2 gene (Philips et al. 1998; Ladd et al. 2004), which encodes the cardiac isoform of troponin T. Investigation of TNNT2 splicing in myocardial tissue from patients in this study revealed a significant association between the presence of the SNP and the coexistence of both the embryonic and adult splicing variants of TNNT2 (Wang et al. 2016). Considering prior findings, it was hypothesized that the continued coexpression of multiple troponin T isoforms enhances cardiotoxicity in response to treatment with high-dose anthracyclines (Wang et al. 2016). Nevertheless, it was recently found that CELF4 plays an important role in translational regulation (Salamon et al. 2023). This is perhaps not surprising given that other members of the CELF protein family are broadly implicated in many facets of RNA biology, including AS, deadenylation, mRNA decay, and translation, among others (Vlasova and Bohjanen 2008; Dasgupta and Ladd 2012). Yet, to what extent disruption of CELF4-mediated translational regulation contributes to predisposition to disease remains an open and intriguing question.

CONCLUSIONS

The role of AS in cardiac development and disease has garnered increasing attention over the last several decades. The growing identification of genetic variants in AS factors linked to cardiac developmental defects and disease has set the stage for deeper investigations into the underlying molecular mechanisms (Brauch et al. 2009; Johnston et al. 2010; Homsy et al. 2015; Wang et al. 2016; Au et al. 2018; Yu et al. 2018; Chen et al. 2024). Target gene missplicing downstream from pathogenic variants in AS factors is likely to contribute to disease pathogenesis to some extent. Nevertheless, taking a lesson from our own studies on the molecular mechanisms associated with pathogenic variants in RBM20, we advocate for a broader investigation into the contributions of AS factor functions, particularly their nonsplicing roles, to disease pathogenesis. Missplicing may not be the sole mechanism at play. This approach is especially relevant given the expanding roles that many AS factors are now recognized to have in various stages of the RNA life cycle. However, such investigations are not without challenges. Innovative strategies will be necessary to dissect the various nonsplicing functions mediated by AS factors and their contributions to cardiac development and disease.

ACKNOWLEDGMENTS

This work was supported by the National Institutes of Health (NIH) National Heart, Lung, and Blood Institute (HL148733 to W.G.); the American Heart Association Foundation (19TPA3480072 and 23TPA1069731 to W.G.); the Wisconsin Alumni Research Foundation (AAH4884 to W.G.); the University of Wisconsin Foundation (AAH5964 to W.G.); and the Salm Gift fund (AAP1679 to W.G.).

REFERENCES

- Au PYB, Goedhart C, Ferguson M, Breckpot J, Devriendt K, Wierenga K, Fanning E, Grange DK, Graham GE, Galarreta C, et al. 2018. Phenotypic spectrum of Au-Kline syndrome: a report of six new cases and review of the literature. Eur J Hum Genet 26: 1272–1281. doi:10.1038/s41431-018-0187-2
- Baralle FE, Giudice J. 2017. Alternative splicing as a regulator of development and tissue identity. *Nat Rev Mol Cell Biol* **18:** 437–451. doi:10.1038/nrm.2017.27
- Beqqali A, Bollen IA, Rasmussen TB, van den Hoogenhof MM, van Deutekom HW, Schafer S, Haas J, Meder B, Sørensen KE, van Oort RJ, et al. 2016. A mutation in the glutamate-rich region of RNA-binding motif protein 20 causes dilated cardiomyopathy through missplicing of titin and impaired Frank-Starling mechanism. *Cardiovasc Res* **112**: 452–463. doi:10.1093/cvr/cvw192
- Bertero A, Fields PA, Ramani V, Bonora G, Yardimci GG, Reinecke H, Pabon L, Noble WS, Shendure J, Murry CE. 2019. Dynamics of genome reorganization during human cardiogenesis reveal an RBM20-dependent splicing factory. *Nat Commun* 10: 1538. doi:10.1038/s41467-019-09483-5
- Boeckel JN, Möbius-Winkler M, Müller M, Rebs S, Eger N, Schoppe L, Tappu R, Kokot KE, Kneuer JM, Gaul S, et al. 2022. SLM2 is a novel cardiac splicing factor involved in heart failure due to dilated cardiomyopathy. *Genomics Proteomics Bioinformatics* 20: 129–146. doi:10.1016/j.gpb.2021.01.006
- Boon RA, Dimmeler S. 2015. MicroRNAs in myocardial infarction. *Nat Rev Cardiol* **12:** 135–142. doi:10.1038/nrcardio.2014.207
- Brauch KM, Karst ML, Herron KJ, de Andrade M, Pellikka PA, Rodeheffer RJ, Michels VV, Olson TM. 2009. Mutations in ribonucleic acid binding protein gene cause familial dilated cardiomyopathy. J Am Coll Cardiol 54: 930–941. doi:10.1016/j.jacc.2009.05 .038
- Briganti F, Sun H, Wei W, Wu J, Zhu C, Liss M, Karakikes I, Rego S, Cipriano A, Snyder M, et al. 2020. iPSC modeling of RBM20-deficient DCM identifies upregulation of RBM20 as a therapeutic strategy. *Cell Rep* **32:** 108117. doi:10.1016/j.celrep.2020.108117
- Cao J, Wei Z, Nie Y, Chen HZ. 2024. Therapeutic potential of alternative splicing in cardiovascular diseases. *EBioMedicine* **101**: 104995. doi:10.1016/j.ebiom.2024.104995
- Chen Y, Zubovic L, Yang F, Godin K, Pavelitz T, Castellanos J, Macchi P, Varani G. 2016. Rbfox proteins regulate microRNA biogenesis by sequence-specific binding to their precursors and target downstream Dicer. Nucleic Acids Res 44: 4381–4395. doi:10 .1093/nar/gkw177
- Chen Y, Yang B, Zhang XM, Chen S, Wang M, Hu L, Pan N, Li S, Shi W, Yang Z, et al. 2024. Biallelic variants in *RBM42* cause a multisystem disorder with neurological, facial, cardiac, and musculoskeletal involvement. *Protein Cell* **15:** 52–68. doi:10.1093/procel/pwad034
- Cho N, Kim JO, Lee S, Choi S, Kim J, Ko MS, Park SJ, Ji JH, Kim KK. 2019. Alternative splicing induces cytoplasmic localization of RBFOX2 protein in calcific tendinopathy. *Exp Mol Pathol* **109:** 36–41. doi:10.1016/j.yexmp.2019.104264

- Dasgupta T, Ladd AN. 2012. The importance of CELF control: molecular and biological roles of the CUG-BP, Elav-like family of RNA-binding proteins. *Wiley Interdiscip Rev RNA* **3:** 104–121. doi:10.1002/wma.107
- Deveson IW, Brunck ME, Blackburn J, Tseng E, Hon T, Clark TA, Clark MB, Crawford J, Dinger ME, Nielsen LK, et al. 2018. Universal alternative splicing of noncoding exons. *Cell Syst* **6:** 245–255.e45. doi:10.1016/j.cels.2017.12.005
- Fenix AM, Miyaoka Y, Bertero A, Blue SM, Spindler MJ, Tan KKB, Perez-Bermejo JA, Chan AH, Mayerl SJ, Nguyen TD, et al. 2021. Gain-of-function cardiomyopathic mutations in RBM20 rewire splicing regulation and re-distribute ribonucleoprotein granules within processing bodies. *Nat Commun* 12: 6324. doi:10.1038/s41467-021-26623-y
- Fu XD, Ares M. 2014. Context-dependent control of alternative splicing by RNA-binding proteins. *Nat Rev Genet* 15: 689–701. doi:10.1038/nrg3778
- Gaertner A, Klauke B, Felski E, Kassner A, Brodehl A, Gerdes D, Stanasiuk C, Ebbinghaus H, Schulz U, Dubowy KO, et al. 2020. Cardiomyopathy-associated mutations in the RS domain affect nuclear localization of RBM20. Hum Mutat 41: 1931–1943. doi:10.1002/humu.24096
- Gallagher TL, Arribere JA, Geurts PA, Exner CR, McDonald KL, Dill KK, Marr HL, Adkar SS, Garnett AT, Amacher SL, et al. 2011. Rbfox-regulated alternative splicing is critical for zebrafish cardiac and skeletal muscle functions. *Dev Biol* 359: 251–261. doi:10 .1016/j.ydbio.2011.08.025
- Giudice J, Xia Z, Wang ET, Scavuzzo MA, Ward AJ, Kalsotra A, Wang W, Wehrens XH, Burge CB, Li W, et al. 2014. Alternative splicing regulates vesicular trafficking genes in cardiomyocytes during postnatal heart development. *Nat Commun* 5: 3603. doi:10.1038/ncomms4603
- Glessner JT, Bick AG, Ito K, Homsy J, Rodriguez-Murillo L, Fromer M, Mazaika E, Vardarajan B, Italia M, Leipzig J, et al. 2014. Increased frequency of de novo copy number variants in congenital heart disease by integrative analysis of single nucleotide polymorphism array and exome sequence data. *Circ Res* **115**: 884–896. doi:10.1161/CIRCRESAHA.115.304458
- Good PJ, Chen Q, Warner SJ, Herring DC. 2000. A family of human RNA-binding proteins related to the *Drosophila* Bruno translational regulator. *J Biol Chem* **275:** 28583–28592. doi:10.1074/jbc..M003083200
- Gordon JM, Phizicky DV, Neugebauer KM. 2021. Nuclear mechanisms of gene expression control: pre-mRNA splicing as a life or death decision. Curr Opin Genet Dev 67: 67–76. doi:10.1016/j.gde.2020.11.002
- Gregorich ZR, Zhang Y, Kamp TJ, Granzier HL, Guo W. 2024. Mechanisms of RBM20 cardiomyopathy: insights from model systems. Circ Genom Precis Med 17: e004355. doi:10.1161/CIRCGEN.123.004355
- Grosch M, Schraft L, Chan A, Küchenhoff L, Rapti K, Ferreira AM, Kornienko J, Li S, Radke MH, Krämer C, et al. 2023. Striated muscle-specific base editing enables correction of mutations causing dilated cardiomyopathy. *Nat Commun* **14:** 3714. doi:10.1038/s41467-023-39352-1
- Guo W, Schafer S, Greaser ML, Radke MH, Liss M, Govindarajan T, Maatz H, Schulz H, Li S, Parrish AM, et al. 2012. RBM20, a gene for hereditary cardiomyopathy, regulates titin splicing. Nat Med 18: 766–773. doi:10.1038/nm.2693
- Guo W, Zhu C, Yin Z, Wang Q, Sun M, Cao H, Greaser ML. 2018. Splicing factor RBM20 regulates transcriptional network of titin associated and calcium handling genes in the heart. *Int J Biol Sci* **14:** 369–380. doi:10.7150/ijbs.24117
- Guo W, Zhu C, Yin Z, Zhang Y, Wang C, Walk AS, Lin YH, McKinsey TA, Woulfe KC, Ren J, et al. 2021. The ryanodine receptor stabilizer

- S107 ameliorates contractility of adult Rbm20 knockout rat cardiomyocytes. *Physiol Rep* **9:** e15011. doi:10.14814/phy2.15011
- Homsy J, Zaidi S, Shen Y, Ware JS, Samocha KE, Karczewski KJ, DePalma SR, McKean D, Wakimoto H, Gorham J, et al. 2015. De novo mutations in congenital heart disease with neurodevelopmental and other congenital anomalies. *Science* 350: 1262– 1266. doi:10.1126/science.aac9396
- Huang M, Akerberg AA, Zhang X, Yoon H, Joshi S, Hallinan C, Nguyen C, Pu WT, Haigis MC, Burns CG, et al. 2022. Intrinsic myocardial defects underlie an Rbfox-deficient zebrafish model of hypoplastic left heart syndrome. *Nat Commun* 13: 5877. doi:10 .1038/s41467-022-32982-x
- Ihara K, Sasano T, Hiraoka Y, Togo-Ohno M, Soejima Y, Sawabe M, Tsuchiya M, Ogawa H, Furukawa T, Kuroyanagi H. 2020. A missense mutation in the RSRSP stretch of Rbm20 causes dilated cardiomyopathy and atrial fibrillation in mice. Sci Rep 10: 17894. doi:10.1038/s41598-020-74800-8
- Irimia M, Blencowe BJ. 2012. Alternative splicing: decoding an expansive regulatory layer. *Curr Opin Cell Biol* **24:** 323–332. doi:10.1016/j.ceb.2012.03.005
- Jin Y, Suzuki H, Maegawa S, Endo H, Sugano S, Hashimoto K, Yasuda K, Inoue K. 2003. A vertebrate RNA-binding protein Fox-1 regulates tissue-specific splicing via the pentanucleotide GCAUG. EMBO J 22: 905–912. doi:10.1093/emboj/cdg089
- Johnston JJ, Teer JK, Cherukuri PF, Hansen NF, Loftus SK, Chong K, Mullikin JC, Biesecker LG; NIH Intramural Sequencing Center (NISC). 2010. Massively parallel sequencing of exons on the X chromosome identifies RBM10 as the gene that causes a syndromic form of cleft palate. Am J Hum Genet 86: 743–748. doi:10.1016/j.ajhg.2010.04.007
- Kabashi E, Valdmanis PN, Dion P, Spiegelman D, McConkey BJ, Vande Velde C, Bouchard JP, Lacomblez L, Pochigaeva K, Salachas F, et al. 2008. TARDBP mutations in individuals with sporadic and familial amyotrophic lateral sclerosis. Nat Genet 40: 572–574. doi:10.1038/ng.132
- Kalsotra A, Xiao X, Ward AJ, Castle JC, Johnson JM, Burge CB, Cooper TA. 2008. A postnatal switch of CELF and MBNL proteins reprograms alternative splicing in the developing heart. Proc Natl Acad Sci 105: 20333–20338. doi:10.1073/pnas.0809045105
- Kastner B, Will CL, Stark H, Lührmann R. 2019. Structural insights into nuclear pre-mRNA splicing in higher eukaryotes. *Cold Spring Harb Perspect Biol* **11:** a032417. doi:10.1101/cshperspect.a032417
- Kayvanpour E, Sedaghat-Hamedani F, Amr A, Lai A, Haas J, Holzer DB, Frese KS, Keller A, Jensen K, Katus HA, et al. 2017. Genotype-phenotype associations in dilated cardiomyopathy: meta-analysis on more than 8000 individuals. Clin Res Cardiol 106: 127–139. doi:10.1007/s00392-016-1033-6
- Khan MA, Reckman YJ, Aufiero S, van den Hoogenhof MM, van der Made I, Beqqali A, Koolbergen DR, Rasmussen TB, van der Velden J, Creemers EE, et al. 2016. RBM20 regulates circular RNA production from the titin gene. *Circ Res* **119**: 996–1003. doi:10.1161/CIRCRESAHA.116.309568
- Kong SW, Hu YW, Ho JW, Ikeda S, Polster S, John R, Hall JL, Bisping E, Pieske B, dos Remedios CG, et al. 2010. Heart failure-associated changes in RNA splicing of sarcomere genes. *Circ Cardiovasc Genet* **3:** 138–146. doi:10.1161/CIRCGENETICS.109.904698
- Kornblihtt AR, Schor IE, Alló M, Dujardin G, Petrillo E, Muñoz MJ. 2013. Alternative splicing: a pivotal step between eukaryotic transcription and translation. Nat Rev Mol Cell Biol 14: 153–165. doi:10.1038/nrm3525
- Kornienko J, Rodríguez-Martínez M, Fenzl K, Hinze F, Schraivogel D, Grosch M, Tunaj B, Lindenhofer D, Schraft L, Kueblbeck M, et al. 2023. Mislocalization of pathogenic RBM20 variants in dilated cardiomyopathy is caused by loss-of-interaction with Transportin-3. *Nat Commun* **14:** 4312. doi:10.1038/s41467-023-39965-6

- Kwiatkowski TJ, Bosco DA, Leclerc AL, Tamrazian E, Vanderburg CR, Russ C, Davis A, Gilchrist J, Kasarskis EJ, Munsat T, et al. 2009. Mutations in the FUS/TLS gene on chromosome 16 cause familial amyotrophic lateral sclerosis. Science 323: 1205–1208. doi:10 .1126/science.1166066
- Ladd AN, Charlet N, Cooper TA. 2001. The CELF family of RNA binding proteins is implicated in cell-specific and developmentally regulated alternative splicing. *Mol Cell Biol* **21**: 1285–1296. doi:10.1128/MCB.21.4.1285-1296.2001
- Ladd AN, Nguyen NH, Malhotra K, Cooper TA. 2004. CELF6, a member of the CELF family of RNA-binding proteins, regulates muscle-specific splicing enhancer-dependent alternative splicing. J Biol Chem 279: 17756–17764. doi:10.1074/jbc.M310687200
- Lee JH, Gao C, Peng G, Greer C, Ren S, Wang Y, Xiao X. 2011. Analysis of transcriptome complexity through RNA sequencing in normal and failing murine hearts. *Circ Res* **109:** 1332–1341. doi:10.1161/CIRCRESAHA.111.249433
- Lennermann DC, Pepin ME, Grosch M, Konrad L, Kemmling E, Hartmann J, Nolte JL, Clauder-Münster S, Kayvanpour E, Sedaghat-Hamedani F, et al. 2022. Deep phenotyping of two preclinical mouse models and a cohort of RBM20 mutation carriers reveals no sex-dependent disease severity in *RBM20* cardiomyopathy. *Am J Physiol Heart Circ Physiol* 323: H1296–H1310. doi:10.1152/ajpheart.00328.2022
- Lewis BP, Green RE, Brenner SE. 2003. Evidence for the widespread coupling of alternative splicing and nonsense-mediated mRNA decay in humans. *Proc Natl Acad Sci* **100**: 189–192. doi:10 .1073/pnas.0136770100
- Licatalosi DD, Darnell RB. 2010. RNA processing and its regulation: global insights into biological networks. *Nat Rev Genet* **11:** 75–87. doi:10.1038/nrg2673
- Lin Y, Protter DS, Rosen MK, Parker R. 2015. Formation and maturation of phase-separated liquid droplets by RNA-binding proteins. *Mol Cell* **60**: 208–219. doi:10.1016/j.molcel.2015.08.018
- Linke WA, Bücker S. 2012. King of hearts: a splicing factor rules cardiac proteins. *Nat Med* **18:** 660–661. doi:10.1038/nm.2762
- Maatz H, Jens M, Liss M, Schafer S, Heinig M, Kirchner M, Adami E, Rintisch C, Dauksaite V, Radke MH, et al. 2014. RNA-binding protein RBM20 represses splicing to orchestrate cardiac pre-mRNA processing. J Clin Invest 124: 3419–3430. doi:10.1172/JCI74523
- Medford RM, Nguyen HT, Destree AT, Summers E, Nadal-Ginard B. 1984. A novel mechanism of alternative RNA splicing for the developmentally regulated generation of troponin T isoforms from a single gene. *Cell* 38: 409–421. doi:10.1016/0092-8674(84)90496-3
- Methawasin M, Hutchinson KR, Lee EJ, Smith JE, Saripalli C, Hidalgo CG, Ottenheijm CA, Granzier H. 2014. Experimentally increasing titin compliance in a novel mouse model attenuates the Frank-Starling mechanism but has a beneficial effect on diastole. Circulation 129: 1924–1936. doi:10.1161/CIRCULATIONAHA.113.005610
- Montañés-Agudo P, Pinto YM, Creemers EE. 2023. Splicing factors in the heart: uncovering shared and unique targets. *J Mol Cell Cardiol* **179:** 72–79. doi:10.1016/j.yjmcc.2023.04.003
- Nilsen TW, Graveley BR. 2010. Expansion of the eukaryotic proteome by alternative splicing. *Nature* 463: 457–463. doi:10.1038/ nature08909
- Nishiyama T, Zhang Y, Cui M, Li H, Sanchez-Ortiz E, McAnally JR, Tan W, Kim J, Chen K, Xu L, et al. 2022. Precise genomic editing of pathogenic mutations in *RBM20* rescues dilated cardiomyopathy. *Sci Transl Med* **14:** eade1633. doi:10.1126/scitranslmed.ade1633
- Nutter CA, Jaworski EA, Verma SK, Deshmukh V, Wang Q, Botvinnik OB, Lozano MJ, Abass IJ, Ijaz T, Brasier AR, et al. 2016. Dysregulation of RBFOX2 is an early event in cardiac pathogenesis of diabetes. *Cell Rep* 15: 2200–2213. doi:10.1016/j.celrep.2016.05.002

- Pan Q, Shai O, Lee LJ, Frey BJ, Blencowe BJ. 2008. Deep surveying of alternative splicing complexity in the human transcriptome by high-throughput sequencing. *Nat Genet* **40:** 1413–1415. doi:10 1038/ng 259
- Parikh VN, Caleshu C, Reuter C, Lazzeroni LC, Ingles J, Garcia J, McCaleb K, Adesiyun T, Sedaghat-Hamedani F, Kumar S, et al. 2019. Regional variation in *RBM20* causes a highly penetrant arrhythmogenic cardiomyopathy. *Circ Heart Fail* 12: e005371. doi:10.1161/CIRCHEARTFAILURE.118.005371
- Pasche B, Absher D. 2011. Whole-genome sequencing: a step closer to personalized medicine. JAMA 305: 1596–1597. doi:10.1001/ iama.2011.484
- Philips AV, Timchenko LT, Cooper TA. 1998. Disruption of splicing regulated by a CUG-binding protein in myotonic dystrophy. *Science* **280**: 737–741. doi:10.1126/science.280.5364.737
- Robyns T, Willems R, Van Cleemput J, Jhangiani S, Muzny D, Gibbs R, Lupski JR, Breckpot J, Devriendt K, Corveleyn A. 2020. Whole exome sequencing in a large pedigree with DCM identifies a novel mutation in RBM20. Acta Cardiol 75: 748–753. doi:10.1080/ 00015385.2019.1674490
- Salamon I, Park Y, Miškić T, Kopić J, Matteson P, Page NF, Roque A, McAuliffe GW, Favate J, Garcia-Forn M, et al. 2023. Celf4 controls mRNA translation underlying synaptic development in the prenatal mammalian neocortex. Nat Commun 14: 6025. doi:10.1038/ s41467-023-41730-8
- Schneider JW, Oommen S, Qureshi MY, Goetsch SC, Pease DR, Sundsbak RS, Guo W, Sun M, Sun H, Kuroyanagi H, et al. 2020. Dysregulated ribonucleoprotein granules promote cardiomyopathy in RBM20 gene-edited pigs. Nat Med 26: 1788–1800. doi:10 .1038/s41591-020-1087-x
- Shenasa H, Hertel KJ. 2019. Combinatorial regulation of alternative splicing. *Biochim Biophys Acta Gene Regul Mech* **1862:** 194392. doi:10.1016/j.bbagrm.2019.06.003
- Song HK, Hong SE, Kim T, Kim DH. 2012. Deep RNA sequencing reveals novel cardiac transcriptomic signatures for physiological and pathological hypertrophy. PLoS ONE 7: e35552. doi:10.1371/jour nal.pone.0035552
- Streckfuss-Bömeke K, Tiburcy M, Fomin A, Luo X, Li W, Fischer C, Özcelik C, Perrot A, Sossalla S, Haas J, et al. 2017. Severe DCM phenotype of patient harboring RBM20 mutation S635A can be modeled by patient-specific induced pluripotent stem cell-derived cardiomyocytes. J Mol Cell Cardiol 113: 9–21. doi:10.1016/j.yjmcc.2017.09.008
- Ule J, Blencowe BJ. 2019. Alternative splicing regulatory networks: functions, mechanisms, and evolution. *Mol Cell* **76:** 329–345. doi:10.1016/j.molcel.2019.09.017
- Underwood JG, Boutz PL, Dougherty JD, Stoilov P, Black DL. 2005. Homologues of the Caenorhabditis elegans Fox-1 protein are neuronal splicing regulators in mammals. Mol Cell Biol 25: 10005–10016. doi:10.1128/MCB.25.22.10005-10016.2005
- Vance C, Rogelj B, Hortobágyi T, De Vos KJ, Nishimura AL, Sreedharan J, Hu X, Smith B, Ruddy D, Wright P, et al. 2009. Mutations in FUS, an RNA processing protein, cause familial amyotrophic lateral sclerosis type 6. Science 323: 1208–1211. doi:10.1126/science.1165942
- van den Hoogenhof MMG, Beqqali A, Amin AS, van der Made I, Aufiero S, Khan MAF, Schumacher CA, Jansweijer JA, van Spaendonck-Zwarts KY, Remme CA, et al. 2018. RBM20 mutations induce an arrhythmogenic dilated cardiomyopathy related to disturbed calcium handling. *Circulation* **138**: 1330–1342. doi:10.1161/CIRCULATIONAHA.117.031947
- van Rooij E, Sutherland LB, Liu N, Williams AH, McAnally J, Gerard RD, Richardson JA, Olson EN. 2006. A signature pattern of stress-

- responsive microRNAs that can evoke cardiac hypertrophy and heart failure. *Proc Natl Acad Sci* **103:** 18255–18260. doi:10 .1073/pnas.0608791103
- Verma SK, Deshmukh V, Nutter CA, Jaworski E, Jin W, Wadhwa L, Abata J, Ricci M, Lincoln J, Martin JF, et al. 2016. Rbfox2 function in RNA metabolism is impaired in hypoplastic left heart syndrome patient hearts. Sci Rep 6: 30896. doi:10.1038/srep30896
- Verma SK, Deshmukh V, Thatcher K, Belanger KK, Rhyner AM, Meng S, Holcomb RJ, Bressan M, Martin JF, Cooke JP, et al. 2022. RBFOX2 is required for establishing RNA regulatory networks essential for heart development. *Nucleic Acids Res* 50: 2270–2286. doi:10.1093/nar/gkac055
- Vlasova IA, Bohjanen PR. 2008. Posttranscriptional regulation of gene networks by GU-rich elements and CELF proteins. RNA Biol 5: 201–207. doi:10.4161/rna.7056
- Wang ET, Sandberg R, Luo S, Khrebtukova I, Zhang L, Mayr C, Kingsmore SF, Schroth GP, Burge CB. 2008. Alternative isoform regulation in human tissue transcriptomes. *Nature* 456: 470– 476. doi:10.1038/nature07509
- Wang X, Sun CL, Quiñones-Lombraña A, Singh P, Landier W, Hageman L, Mather M, Rotter JI, Taylor KD, Chen YD, et al. 2016. CELF4 variant and anthracycline-related cardiomyopathy: a children's oncology group genome-wide association study. J Clin Oncol 34: 863–870. doi:10.1200/JCO.2015.63.4550
- Wang C, Zhang Y, Methawasin M, Braz CU, Gao-Hu J, Yang B, Strom J, Gohlke J, Hacker T, Khatib H, et al. 2022. RBM20^{S639G} mutation is a high genetic risk factor for premature death through RNA-protein condensates. J Mol Cell Cardiol 165: 115–129. doi:10.1016/j.yjmcc.2022.01.004
- Wyles SP, Li X, Hrstka SC, Reyes S, Oommen S, Beraldi R, Edwards J, Terzic A, Olson TM, Nelson TJ. 2016. Modeling structural and functional deficiencies of *RBM20* familial dilated cardiomyopathy using human induced pluripotent stem cells. *Hum Mol Genet* **25:** 254–265. doi:10.1093/hmg/ddv468
- Yamamoto T, Miura A, Itoh K, Takeshima Y, Nishio H. 2019. RNA sequencing reveals abnormal *LDB3* splicing in sudden cardiac death. *Forensic Sci Int* **302:** 109906. doi:10.1016/j.forsciint.2019 .109906
- Yamamoto T, Sano R, Miura A, Imasaka M, Naito Y, Nishiguchi M, Ihara K, Otani N, Kominato Y, Ohmuraya M, et al. 2022. I536T variant of RBM20 affects splicing of cardiac structural proteins that are causative for developing dilated cardiomyopathy. J Mol Med (Berl) 100: 1741–1754. doi:10.1007/s00109-022-02262-8
- Yu Z, Tang PL, Wang J, Bao S, Shieh JT, Leung AW, Zhang Z, Gao F, Wong SY, Hui AL, et al. 2018. Mutations in *Hnmpa1* cause congenital heart defects. *JCI Insight* 3: e98555. doi:10.5353/th_991044178483203414
- Zhang Y, Wang C, Sun M, Jin Y, Braz CU, Khatib H, Hacker TA, Liss M, Gotthardt M, Granzier H, et al. 2022. RBM20 phosphorylation and its role in nucleocytoplasmic transport and cardiac pathogenesis. *FASEB J* **36:** e22302. doi:10.1096/fj.202101811RR
- Zhang Y, Gregorich ZR, Wang Y, Braz CU, Zhang J, Liu Y, Liu P, Shen J, Aori N, Hacker TA, et al. 2023a. Disruption of the nuclear localization signal in RBM20 is causative in dilated cardiomyopathy. *JCI Insight* 8: e170001. doi:10.1172/jci.insight.170001
- Zhang Z, Bae B, Cuddleston WH, Miura P. 2023b. Coordination of alternative splicing and alternative polyadenylation revealed by targeted long read sequencing. *Nat Commun* **14:** 5506. doi:10.1038/s41467-023-41207-8
- Zhou SS, Jin JP, Wang JQ, Zhang ZG, Freedman JH, Zheng Y, Cai L. 2018. miRNAS in cardiovascular diseases: potential biomarkers, therapeutic targets and challenges. *Acta Pharmacol Sin* **39**: 1073–1084.