Research Article

Pyrolytic and Kinetic Analysis of Two Coastal Plant Species: *Artemisia annua* and *Chenopodium glaucum*

Lili Li,¹ Xiaoning Wang,^{1,2} Jinsheng Sun,^{2,3} Yichen Zhang,² and Song Qin¹

¹ Yantai Institute of Coastal Zone Research, Chinese Academy of Sciences, 17 Chunhui Road, Laishan District, Yantai 264003, China

² Tianjin Key Laboratory of Animal and Plant Resistance/College of Life Science, Tianjin Normal University, Tianjin 300387, China

³ Tianjin Aquatic Animal Infectious Disease Control and Prevention Center, Tianjin 300221, China

Correspondence should be addressed to Song Qin; sqin@yic.ac.cn

Received 13 July 2013; Revised 29 September 2013; Accepted 22 October 2013

Academic Editor: Wei Zhang

Copyright © 2013 Lili Li et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The large amount of coastal plant species available makes them ideal candidates for energy production. In this study, thermogravimetric analysis was used to evaluate the fuel properties of two coastal plant species, and the distributed activation energy model (DAEM) was employed in kinetic analysis. The major mass loss due to devolatilization started at 154 and 162°C at the heating rate of 10°C min⁻¹ for *Artemisia annua* and *Chenopodium glaucum*, respectively. The results showed that the average activation energies of *Artemisia annua* and *Chenopodium glaucum* were 169.69 and 170.48 kJ mol⁻¹, respectively. Furthermore, the activation energy changed while the conversion rate increased, and the frequency factor k_0 decreased greatly while the activation energy decreased. The results also indicated that the devolatilization of the two coastal plant species underwent a set of first-order reactions and could be expressed by the DAEM. Additionally, a simplified mathematical model was proposed to facilitate the prediction of devolatilization curves.

1. Introduction

Biomass is one of the most promising feedstocks for biofuel production because of its merits of renewability and wide distribution. *Artemisia annua* and *Chenopodium glaucum* are coastal plant species that can adapt to several stressful environmental conditions, including high saline content, drought, and heavy metal pollution. They are annual halophytes and distributed widely in North America, Europe, Africa, and Asia. *Artemisia annua* and *Chenopodium glaucum* obtained our interests because of their high yields, appropriate biomass characteristics, low input demands, and positive environmental impacts, such as helping to improve the soil texture and reduce soil salinity. The large amount of these plant species available makes them ideal candidates for energy production.

Among various energy conversion technologies, pyrolysis is one of the most commonly used techniques, which is characterized by thermal conversion of biomass into useful fuel at high temperature in the absence of oxidizing atmosphere [1]. Furthermore, thermogravimetric analysis is a highly precise method for the study of pyrolysis, and it is shown that each kind of biomass has unique pyrolysis characteristics because of its specific proportions of the components [2]. Pyrolytic and kinetic data from thermogravimetry analysis not only are useful for understanding the processes and mechanisms of the thermal degradation but also can be used as input parameters for a thermal degradation reaction system [3].

Numerous models have been used for the pyrolytic analysis. However, it has been found that distributed activation energy model (DAEM) is more accurate than some pseudomechanistic models, especially when the decomposition is carried out under inert atmosphere [4–6]. The present study has evaluated the fuel properties of *Artemisia annua* and *Chenopodium glaucum*. The kinetic parameters were determined by the DAEM method, and a simplified mathematical model was proposed to facilitate the prediction of devolatilization curves.

2. Materials and Methods

2.1. Materials. Artemisia annua and Chenopodium glaucum were collected from a coastal zone of Yantai, Shandong

province, China. The plants were oven-dried at 60° C to constant weight and then grounded with a Mini-Mill to pass through a 125 μ m sieve.

2.2. Proximate and Ultimate Analysis. The moisture analysis was conducted according to ASTM E871-82 (2006). The ash content was determined according to ASTM E1755-01 (2007). The volatile matter content was analyzed according to ASTM E872-82 (2006). The fixed carbon was expressed as the 100%-ash content-volatile matter-moisture content. The C, H, O, N, and S contents in the samples were measured according to our previous study [7]. All measurements were replicated three times.

2.3. Thermogravimetric Analysis. The powders of Artemisia annua and Chenopodium glaucum were analyzed by a Mettler Toledo TGA/DSC1 STARe thermoanalyzer. The pyrolysis experiments were performed at heating rates of 5, 10, 20, and 30° C min⁻¹ in a dynamic high purity nitrogen flow of 50 mL min⁻¹. The temperature of the furnace was programmed to rise from room temperature to 900°C.

2.4. Kinetic Analysis Using DAEM. DAEM has been widely used in analyzing the thermal reaction system of biomasses [6, 8]. It assumes that many irreversible first-order parallel reactions that have different rate parameters occur simultaneously. The model is expressed as

$$1 - \frac{m_t}{m_0} = \int_0^\infty \exp\left(-k_0 \int_0^t \exp\left(-\frac{E}{RT}\right) dt\right) f(E) dE, \quad (1)$$

where m_t is the mass loss by time t, m_0 is the total mass loss at the end of pyrolysis, f(E) is the distribution function of activation energy E that represents the difference in the activation energies of many first-order parallel reactions, and k_0 is the frequency factor corresponding to E value. After a series of reduction, including transformation and approximation [9], DAEM can be expressed as

$$\ln\left(\frac{\beta}{T^2}\right) = \ln\left(\frac{k_0 R}{E}\right) + 0.6075 - \frac{E}{RT}.$$
 (2)

Equation (2) establishes a linear relationship between $\ln(\beta/T^2)$ and (1/T) with the slope of (-E/R), where β is the heating rate and *R* is the universal gas constant. Activation energy *E* and frequency factor k_0 can be determined by the slope and intercept of the Arrhenius plots.

3. Results and Discussion

3.1. Proximate and Ultimate Analysis. Proximate and ultimate analysis facilitates the correlation of the individual composition to the thermal behavior of the biomass materials. Proximate analysis showed that the ash contents of *Artemisia annua* and *Chenopodium glaucum* were 4.4 and 5.1 wt%, respectively. *Artemisia annua* was a little lower in ash content than *Chenopodium glaucum* (Table 1). Ultimate analysis showed that these coastal plant species consisted of moderately high contents of carbon and oxygen and low amounts of nitrogen, hydrogen, and sulfur (Table 1).

3.2. Characterization of Pyrolysis. The shape of the thermogravimetry (TG) curves did not change with different heating rates. However, TG curves shifted toward the right while the heating rate increased (Figure 1). This was due to the effect that the time to reach a given temperature became shorter by an increased heating rate and caused the entire curve to shift to higher temperatures. The temperatures corresponding to the maximum mass loss rate were also increased with the increasing of heating rate (Table 2). Additionally, the maximum rate of decomposition tended to increase at higher heating rate because there was more thermal energy to facilitate better heat transfer between the surroundings and the insides of the samples (Figure 2).

Table 2 showed characteristics of the thermal degradation for the two coastal plant species at different heating rates. The average rate of mass loss at the same heating rate for *Artemisia annua* was higher than that for *Chenopodium* glaucum, which indicated that *Artemisia annua* had a little higher pyrolysis reactivity. Furthermore, the major mass loss due to devolatilization started at lower temperature for *Artemisia annua* than *Chenopodium* glaucum. At the heating rate of 10° C min⁻¹, for example, the onset of devolatilization occurred at 154 and 162°C for *Artemisia annua* and *Chenopodium* glaucum, respectively.

3.3. Kinetic Analysis. The linear and parallel development for different conversion rates from 0.1 to 0.9 at various heating rates was shown in Figure 3. All plots had fairly high linear correlation coefficients ($R^2 \ge 0.98$), indicating that the devolatilization of two plant species underwent a set of first-order reactions. The results showed that *Artemisia annua* and *Chenopodium glaucum* were fairly similar in the average activation energies, which were 169.69 and 170.48 kJ mol⁻¹, respectively (Table 3).

While the conversion rate increased from 0.1 to 0.9, the *E* value of *Artemisia annua* decreased from 180.03 to 161.45 kJ mol⁻¹, whereas that of *Chenopodium glaucum* decreased from 185.84 to 147.17 kJ mol⁻¹ (Table 3). It can be seen that the frequency factor k_0 decreased greatly while *E* values decreased. For *Artemisia annua*, for example, the k_0 value decreased from 2.82×10^{12} to $1.12 \times 10^8 \text{ s}^{-1}$ when the *E* value decreased from 180.03 to 161.45 kJ mol⁻¹.

3.4. Prediction of Devolatilization Curves. When Figure 3 was available, a simplified mathematical model could be established to predict the devolatilization curves. The full line in Figure 3(a) showed that the data points obtained for each particular devolatilization rate could be linearized. And the dotted line in Figure 3(a) showed that the data points obtained for each heating rate could also be linearized, which could be described in the form of $y = a(1/T) + b + \ln \beta$ (where *a* was the slope of the line and $b + \ln \beta$ was the intercept of the line) [10]. The simplified process was exhibited in Table 4. Then the temperature at which devolatilization occurred can be determined with the intersection between the linearization

Analysis	Properties/%	Artemisia annua	Chenopodium glaucum	
	Water content	2.97 ± 0.036	3.10 ± 0.090	
Proximate analysis	Volatile	81.32 ± 3.520	79.95 ± 1.728	
	Ash	4.41 ± 0.038	5.10 ± 0.115	
	Fixed carbon	11.30	11.85	
	С	44.45 ± 0.005	42.77 ± 0.021	
Ultimate analysis	Н	6.26 ± 0.001	6.03 ± 0.035	
	N	0.76 ± 0.004	0.86 ± 0.036	
	S	Not Detected	Not Detected	
	О	31.31 ± 0.007	35.45 ± 0.014	

TABLE 1: Proximate and ultimate analysis of Artemisia annua and Chenopodium glaucum.

TABLE 2: Characteristics of pyrolysis for Artemisia annua and Chenopodium glaucum.

Species	Heating rate/°C min ⁻¹	$T_i/^{\circ}C^{a}$	$T_f/^{\circ}C^{b}$	$T_1/^{\circ}C^{c}$	$(-d\alpha/dt)_{average}$ /% s ⁻¹	$(-d\alpha/dt)_{max}$ /% s ⁻¹	Mass loss/% ^d
Artemisia annua	5	140	396	318	0.0070	0.0672	73.35
	10	154	410	333	0.0135	0.1247	65.41
	20	167	433	344	0.0252	0.2566	66.18
	30	173	448	351	0.0363	0.3794	63.65
Chenopodium glaucum	5	148	391	330	0.0065	0.0568	68.47
	10	162	401	337	0.0128	0.1049	67.67
	20	172	420	353	0.0238	0.2054	63.10
	30	179	465	360	0.0358	0.2742	63.91

 ${}^{a}T_{i}$ was the initial temperature of the major mass loss stage.

 ${}^{b}T_{f}$ was the final temperature of the major mass loss stage. ${}^{c}T_{1}$ was the temperature corresponding to the larger peak of the DTG curve.

^dMass loss = (Initial mass – Residue mass)/Initial mass × 100%.



FIGURE 1: TG curves for Artemisia annua (a) and Chenopodium glaucum (b).



FIGURE 2: DTG curves for Artemisia annua (a) and Chenopodium glaucum (b).



FIGURE 3: Determination of the temperature at which devolatilization occurred for *Artemisia annua* (a) and *Chenopodium glaucum* (b). The dotted lines of 1, 2, 3, and 4 were at the heating rates of 5, 10, 20, and 30° C min⁻¹, respectively.



FIGURE 4: Prediction of the TG curves for Artemisia annua (a) and Chenopodium glaucum (b).



FIGURE 5: Difference between experimental temperature and predicted temperature for Artemisia annua (a) and Chenopodium glaucum (b).

for a heating rate and for each devolatilization rate. *T* can be described in

$$T = \frac{E/R + 1166.55}{\ln(k_0 R/E) + 15.35 - \ln(\beta)}.$$
 (3)

The experimental devolatilization curves at the heating rate of 10° C min⁻¹ were plotted together with the curves determined by the simplified mathematical model (Figure 4), which matched the experimental data very well for the two coastal plant species. The results were similar for the

Convertion rate	Artemisia annua			Chenopodium glaucum		
	$E/kJ \text{ mol}^{-1}$	k_0 / s^{-1}	R^2	$E/kJ mol^{-1}$	$k_0 / { m s}^{-1}$	R^2
0.1	180.03	2.82×10^{12}	1.00	185.84	1.03×10^{13}	0.99
0.2	173.17	$1.44 imes 10^{11}$	1.00	178.55	4.20×10^{11}	0.99
0.3	173.72	5.81×10^{10}	1.00	176.15	$8.19 imes 10^{10}$	0.99
0.4	171.54	$1.54 imes 10^{10}$	1.00	175.49	2.91×10^{10}	1.00
0.5	169.66	5.30×10^{9}	1.00	171.24	5.73×10^{9}	1.00
0.6	166.57	1.67×10^{9}	1.00	169.97	2.44×10^{9}	1.00
0.7	164.73	7.48×10^{8}	1.00	167.81	9.65×10^{8}	1.00
0.8	166.29	6.53×10^{8}	1.00	162.11	1.94×10^{8}	1.00
0.9	161.45	1.12×10^{8}	1.00	147.17	5.71×10^{6}	0.98
Average	169.69			170.48		

TABLE 3: Kinetic parameters analyzed by DAEM for Artemisia annua and Chenopodium glaucum.

TABLE 4: Derivation of prediction equation.

Line	Equation: $y = a(1/T) + b + \ln\beta$	а	b
1	$y = 1193.22(1/T) - 14.78 + \ln(30/60)$	1193.22	-14.78
2	$y = 1178.49(1/T) - 14.76 + \ln(20/60)$	1178.49	-14.76
3	$y = 1158.18(1/T) - 14.73 + \ln(10/60)$	1158.18	-14.73
4	$y = 1136.31(1/T) - 14.69 + \ln(5/60)$	1136.31	-14.69
Average		1166.55	-14.74

heating rate of 5, 20 and 30°C min⁻¹. Moreover, the maximum relative errors between the experimental temperature and those obtained by the simplified mathematical model were 0.85% and 0.88% for *Artemisia annua* and *Chenopodium glaucum*, respectively (Figure 5). From this validation, it was evident that the developed simplified mathematical model closely predicted the devolatilization curves of *Artemisia annua* and *Chenopodium glaucum*. Therefore, the simplified mathematical model is a useful tool for the prediction of devolatilization curves.

4. Conclusions

The fuel properties of *Artemisia annua* and *Chenopodium glaucum* were evaluated. The major mass loss due to devolatilization started at 154 and 162°C at the heating rate of 10°C min⁻¹ for *Artemisia annua* and *Chenopodium glaucum*, respectively. The results showed that average activation energies of *Artemisia annua* and *Chenopodium glaucum* were 169.69 and 170.48 kJ mol⁻¹, respectively. The activation energy changed while the conversion rate increased, and the frequency factor k_0 decreased greatly while activation energy decreased. The results also indicated that the devolatilization of the two coastal plant species underwent a set of first-order reactions and could be expressed by the DAEM. Additionally, a simplified mathematical model was proved to be a credible tool for the prediction of devolatilization curves.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgment

This work was supported by the National Key Technology R&D Program of China (Grant no. 2013BAB01B00).

References

- [1] D. Mourant, Z. Wang, M. He et al., "Mallee wood fast pyrolysis: effects of alkali and alkaline earth metallic species on the yield and composition of bio-oil," *Fuel*, vol. 90, no. 9, pp. 2915–2922, 2011.
- [2] T. S. Lira, K. G. Santos, V. V. Murata, M. Gianesella, and M. A. S. Barrozo, "The use of nonlinearity measures in the estimation of kinetic parameters of sugarcane bagasse pyrolysis," *Chemical Engineering & Technology*, vol. 33, no. 10, pp. 1699–1705, 2010.
- [3] S. Munir, S. S. Daood, W. Nimmo, A. M. Cunliffe, and B. M. Gibbs, "Thermal analysis and devolatilization kinetics of cotton stalk, sugar cane bagasse and shea meal under nitrogen and air atmospheres," *Bioresource Technology*, vol. 100, no. 3, pp. 1413–1418, 2009.
- [4] D. K. Shen, S. Gu, B. Jin, and M. X. Fang, "Thermal degradation mechanisms of wood under inert and oxidative environments using DAEM methods," *Bioresource Technology*, vol. 102, no. 2, pp. 2047–2052, 2011.
- [5] J. Cai and R. Liu, "New distributed activation energy model: numerical solution and application to pyrolysis kinetics of some types of biomass," *Bioresource Technology*, vol. 99, no. 8, pp. 2795–2799, 2008.
- [6] G. Várhegyi, B. Bobály, E. Jakab, and H. Chen, "Thermogravimetric study of biomass pyrolysis kinetics: a distributed activation energy model with prediction tests," *Energy & Fuels*, vol. 25, no. 1, pp. 24–32, 2011.
- [7] L. Li, N. Zhao, X. Fu, M. Shao, and S. Qin, "Thermogravimetric and kinetic analysis of *Spirulina* wastes under nitrogen and air atmospheres," *Bioresource Technology*, vol. 140, pp. 152–157, 2013.
- [8] J. Giuntoli, S. Arvelakis, H. Spliethoff, W. De Jong, and A. H. M. Verkooijen, "Quantitative and kinetic thermogravimetric fourier transform infrared (TG-FTIR) study of pyrolysis of agricultural residues: influence of different pretreatments," *Energy & Fuels*, vol. 23, no. 11, pp. 5695–5706, 2009.
- [9] K. Miura and T. Maki, "A simple method for estimating f(E) and $k_0(E)$ in the distributed activation energy model," *Energy & Fuels*, vol. 12, no. 5, pp. 864–869, 1998.

BioMed Research International

[10] A. Soria-Verdugo, N. Garcia-Hernando, L. Garcia-Gutierrez, and U. Ruiz-Rivas, "Analysis of biomass and sewage sludge devolatilization using the distributed activation energy model," *Energy Conversion and Management*, vol. 65, pp. 239–244, 2013.