

Whole-genome analysis of influenza A(H1N1)pdm09 viruses isolated in Uganda from 2009 to 2011

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We report a whole-genome analysis of 19 influenza A(H1N1)pdm09 isolates from four Ugandan hospitals between 2009 and 2011. The isolates differed from the vaccine strain A/California/07/2009 by three amino acid substitutions P100S, S220T, and I338V in the hemagglutinin and by two amino acid substitutions V106I and N248D in the neuraminidase proteins with consistent mutations in all gene segments distinguishing isolates from the 2009/2010 to 2010/2011 seasons. Phylogenetic analysis showed low genetic evolution, with genetic distances of 0%–1.3% and 0.1%–1.6% for HA and NA genes, respectively. The amino acid substitutions did not lead to antigenic differences from the reference strains.

KEYWORDS

influenza A(H1N1)pdm09, Uganda, whole-genome sequencing

1 | INTRODUCTION

Despite increased capabilities for the detection and surveillance of influenza viruses in many parts of the world,¹ whole-genomic data on the viruses from sub-Saharan Africa remain limited. Yet whole-genome sequencing provides the needed evidence for scientifically sound public health interventions including vaccine development. Due to constant viral antigenic evolution, it is important to continually examine the evolutionary changes in the strains for appropriate vaccine selection each year.² The influenza A(H1N1)pdm09 virus was first reported in Uganda in July 2009 and subsequently spread and

established rapidly within the population where it was thereafter routinely detected. In this study, we report the whole-genome analysis of and genetic variations among the pandemic influenza A(H1N1)pdm09 viruses isolated between July 2009 and May 2011 from four hospital sites in Uganda.

2 | MATERIALS AND METHODS

The study was conducted in two influenza seasons: the 2009/2010 season from July 2009 to February 2010 and the 2010/2011 season

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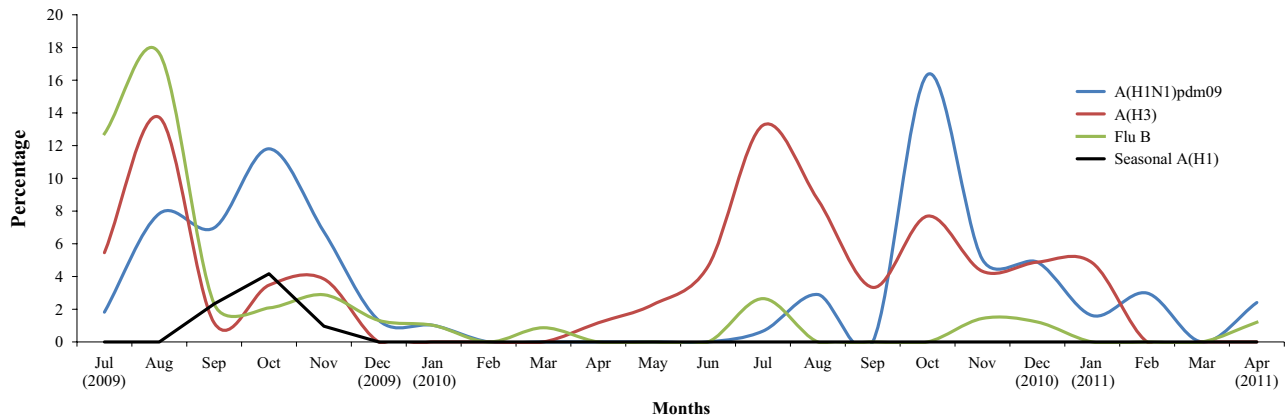


FIGURE 1 Occurrence and seasonality of the influenza A(H1N1)pdm09 isolates during the study period (July 2009 to April 2011)

from July 2010 to April 2011. Samples from patients were collected from four Ugandan hospital-based sentinel surveillance sites: Mulago National Referral Hospital in Kampala (central Uganda), Jinja Regional Referral Hospital in Jinja District (eastern Uganda), Bugiri District Hospital in Bugiri District (eastern Uganda), and Gulu Regional Referral Hospital in Gulu District (northern Uganda). Only individuals who had a fever (temperature $\geq 38^{\circ}\text{C}$) plus either a cough or a sore throat in the last 72 hour of reporting to the outpatient clinic were enrolled in the study. Samples were collected and screened for pandemic influenza using the Centers for Disease Control and Prevention (CDC) protocol for real-time reverse transcription (RT-PCR) for influenza A (H1N1).³ The viruses were isolated in MDCK cells. Three representative isolates were compared against the reference strains in a hemagglutination inhibition (HI) test⁴ using 0.5% turkey erythrocytes for antigenic differences. For whole-genome sequencing, RNA was extracted from isolates with the RNeasy[®] mini kit (Qiagen) as per the manufacturer's protocol. All the eight segments were amplified in a single reaction by RT-PCR using Uni-12, Uni-13, and polymerase gene primers as previously described⁵ and SuperScript III One-Step RT-PCR System with Platinum[®] Taq High Fidelity DNA Polymerase (Invitrogen). PCR products were gel-extracted and purified using the Illustra GFX[™] PCR DNA and Gel Band Purification Kit (GE Healthcare, Pittsburgh, Pennsylvania, USA) per the manufacturer's instructions. PCR products were sequenced by the Sanger sequencing method at the National Institute of Allergy and Infectious Diseases Center of Excellence in Influenza Research and Surveillance/WHO Collaborating Center for the Ecology of Influenza in Animals at St. Jude Children's Research Hospital (St. Jude), Memphis, TN. Nucleotide sequences obtained were aligned by ClustalW and edited with Bioedit Software version 5.0.9.⁶ Phylogenetic analysis of the Ugandan, the pandemic H1N1 vaccine strain (A/California/07/2009), and other related sequences from the GenBank was performed. Sequence feature (SF) variant type analysis was performed from the Influenza Research Database as described previously⁷ to predict whether the isolates carried specific SFs of importance. The sequences were deposited into GenBank with accession numbers KJ690389–KJ690546.

3 | ETHICAL APPROVAL

The study was approved by Makerere University School of Public Health Institutional Review Board (MUSPH #020), the US Army Research and Material Command (MRMC HSRRB/HRPO #RV231/A-1427.1), and the Uganda National Council for Science and Technology (UNCST #HS377).

4 | RESULTS

Influenza A(H1N1)pdm09 was the second most predominant strain during the period (36%; 73/199 influenza isolates) which by December 2009 had completely replaced the seasonal influenza H1N1 (Fig 1). Virus isolates did not show significant antigenic differences from the vaccine strain A/California/07/2009 (Table 1) in antigenic tests. Whole-genome analysis of 19 influenza A(H1N1)pdm09 isolates

TABLE 1 Antigenic comparison of representative Ugandan influenza A(H1N1)pdm09 virus isolates with reference strains

| Virus strain | HI titers | |
|------------------------------------|---|---|
| | Reference antisera | |
| Reference strain | F.2009-4 α A/ California/4/2009 | F.2009-8 α A/ Tennessee 1-560/2009 |
| A/Tennessee/1-560/ 2009 (H1N1) | 160 | 160 |
| A/California/04/2009 (H1N1) | 80 | 160 |
| Test strain | | |
| A/Uganda/MUWRP- 098/2009 (H1N1) | 320 | 160 |
| A/Uganda/MUWRP- 220/2010 (H1N1) | 640 | 320 |
| A/Uganda/MUWRP- 233/2010 (H1N1) | 320 | 320 |

HI, hemagglutination inhibition.

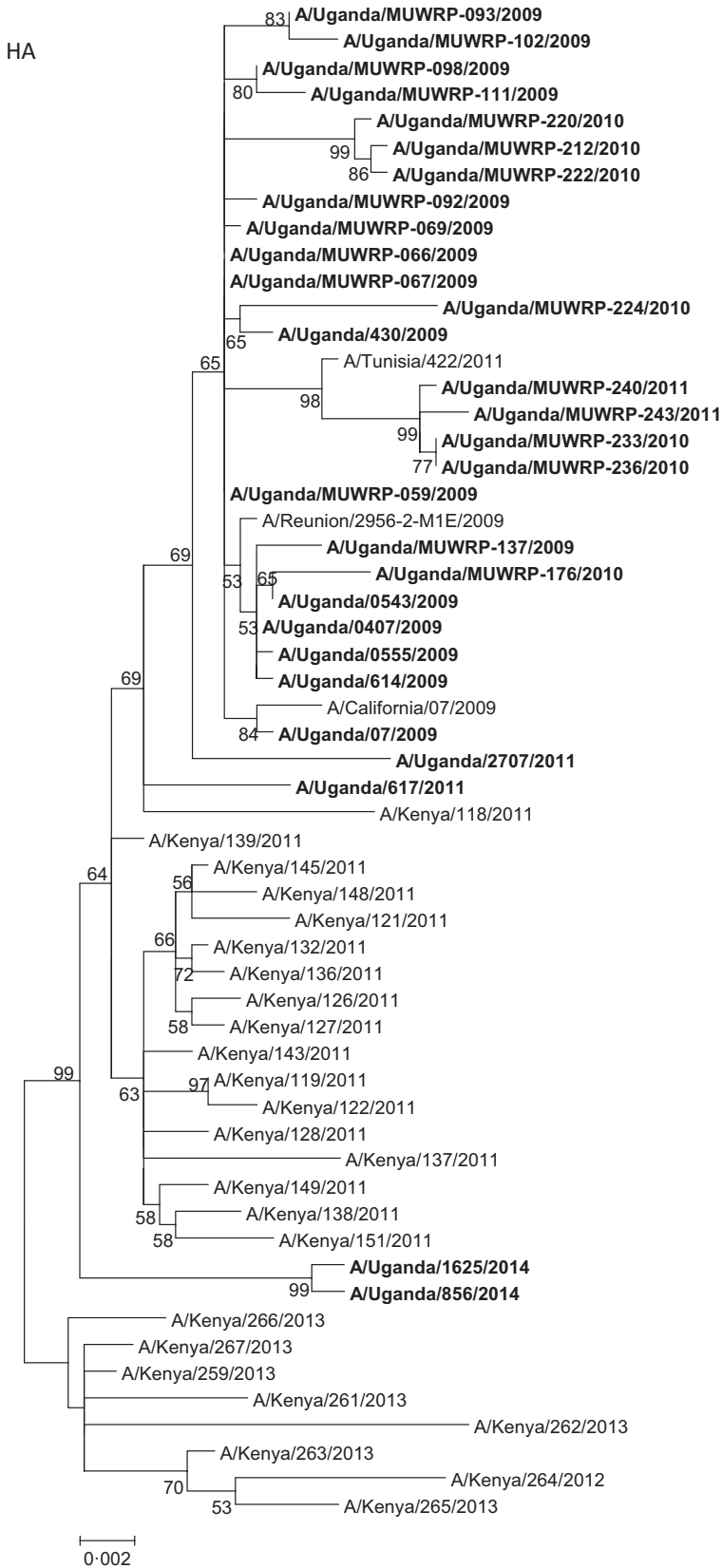


FIGURE 2 Phylogenetic tree of the hemagglutinin (HA) gene segment of the Ugandan influenza A(H1N1)pdm09 viruses (in bold font) at the nucleotide level. The HA sequences of Ugandan H1N1 isolates were compared with relevant virus sequences available in GenBank: sequences from representative influenza A(H1N1)pdm09 isolates from Africa and closest blast hits from different regions of the world. Ugandan sequences available in the GISAID Epiflu database were also used (GISAID isolates ids: EPI_ISL_94767, EPI_ISL_105881, EPI_ISL_176803, EPI_ISL_176806). Sequences of vaccine strains [A/California/07/2009 (H1N1pdm09) and A/Brisbane/59/2007 (seasonal H1N1)] were also included. Bootstrap values (500 replicates) >50 are indicated on the nodes.*indicates partial sequence data

showed that they differed from A/California/07/2009 by three amino acid substitutions P100S, S220T, and I338V in the HA segment and two amino acid substitutions V106I and N248D in the neuraminidase (NA) segment. The genetic variation among the isolates recovered

was low, with genetic distances of 0%–1.3% for HA (Fig. 2). The influenza A(H1N1)pdm09 strains isolated in July and August 2009 closely matched the A/California/07/2009 vaccine strain (except for the few amino acid differences detected in all Uganda isolates),

but those isolated subsequently had additional differences from the A/California/07/2009 strain (Table 2). While there were no age- or gender-related differences in the isolates, it was noted that the strains isolated from Jinja Hospital could be differentiated from the strains from the other hospital sites by an amino acid Y366F substitution in the HA gene. Most of the 2010 influenza A(H1N1)pdm09 strains had the amino acid substitutions V36I, S145P, I303V, and I341V. The late

2010 and the 2011 influenza A(H1N1)pdm09 strains were further defined by amino acid substitutions A151, S200P, and E516K, and there were additional amino acid substitutions in the 2011 isolates as shown in Table 2.

Phylogenetic analysis of NA also revealed a low genetic variation (genetic distance 0.1%–1.6%) at the nucleotide level (Fig. 3), but the strains isolated in different years could also be distinguished by their

TABLE 2 Ugandan influenza A(H1N1)pdm09 isolates (n = 19) included in the study and their genetic substitutions in the HA and NA proteins compared with the A/California/07/2009 vaccine strain

| Virus strain | Date of sample collection | Patient age (months) | Patient gender | Hospital | Amino acid substitutions in the HA compared to A/California/07/2009 | Amino acid substitutions in the NA compared to A/California/07/2009 |
|-------------------------------|---------------------------|----------------------|----------------|----------|---|---|
| A/Uganda/MUWRP-059/2009(H1N1) | July 22, 2009 | 39 | F | Mulago | P100S, S220T, I338V | V106I, N248D |
| A/Uganda/MUWRP-092/2009(H1N1) | Sep 21, 2009 | 84 | M | Mulago | P100S, S220T, I338V | |
| A/Uganda/MUWRP-066/2009(H1N1) | Aug 5, 2009 | 36 | M | Mulago | P100S, S220T, I338V | V106I, N248D, N309D |
| A/Uganda/MUWRP-067/2009(H1N1) | Aug 5, 2009 | 67 | F | Mulago | P100S, S220T, I338V | V106I, N248D, N309D |
| A/Uganda/MUWRP-069/2009(H1N1) | Aug 7, 2009 | 56 | F | Mulago | P100S, S220T, I338V, H45Q | V106I, N248D, N309D |
| A/Uganda/MUWRP-111/2009(H1N1) | Oct 19, 2009 | 12 | F | Jinja | P100S, S220T, I338V, Y366F | V106I, N248D |
| A/Uganda/MUWRP-098/2009(H1N1) | Sep 28, 2009 | 168 | M | Jinja | P100S, S220T, I338V, Y366F | V106I, N248D, I393V, G454S |
| A/Uganda/MUWRP-102/2009(H1N1) | Oct 5, 2009 | 240 | F | Jinja | P100S, S220T, I338V, V64I, A65S, N146R, G187, Y366F, H370L | V106I, N248D |
| A/Uganda/MUWRP-093/2009(H1N1) | Sep 23, 2009 | 15 | F | Mulago | P100S, S220T, I338V, V64I, A65S, N146R, G187 | V106I, N248D |
| A/Uganda/MUWRP-137/2009(H1N1) | Nov 3, 2009 | 42 | M | Jinja | P100S, S220T, I338V, D239E | V106I, N248D, M15V, I17V, I188T |
| A/Uganda/MUWRP-176/2010(H1N1) | July 6, 2010 | 29 | F | Mulago | P100S, S220T, I338V, I133M, H155, D239E | V106I, N248D, M15V, I17V, I188T |
| A/Uganda/MUWRP-212/2010(H1N1) | Oct 22, 2010 | 84 | M | Gulu | P100S, S220T, I338V, V36I, S145P, I303V, I341V | V106I, N248D |
| A/Uganda/MUWRP-220/2010(H1N1) | Oct 26, 2010 | 6 | M | Gulu | P100S, S220T, I338V, V36I, S145P, I303V, I341V | V106I, N248D |
| A/Uganda/MUWRP-222/2010(H1N1) | Oct 28, 2010 | 24 | M | Gulu | P100S, S220T, I338V, V36I, S145P, I303V, I341V | V106I, N248D |
| A/Uganda/MUWRP-224/2010(H1N1) | Nov 11, 2010 | 7 | F | Jinja | P100S, S220T, I338V, V36I, S145P, I303V, I341V | Q39R, S70N; N190S, Q309S, N386S |
| A/Uganda/MUWRP-233/2010(H1N1) | Dec 9, 2010 | 18 | M | Mulago | P100S, S220T, I338V, A151, S200P, E516K | R220K, Q313R, I389K, V394I |
| A/Uganda/MUWRP-236/2010(H1N1) | Dec 17, 2010 | 22 | M | Mulago | P100S, S220T, I338V, A151, S200P, E516K | Q78R, R220K, Q313R, I389K, V394I |
| A/Uganda/MUWRP-240/2011(H1N1) | Feb 7, 2011 | 18 | F | Mulago | A151, S200P, E516K, S207G | N44I, R220K, Q313R, I389K, V394I |
| A/Uganda/MUWRP-243/2011(H1N1) | Apr 11, 2011 | 228 | F | Jinja | A151, S200P, E516K, E252G, K475E | R220K, Q313R, I389K, V394I |

HA, hemagglutinin; NA, neuraminidase.

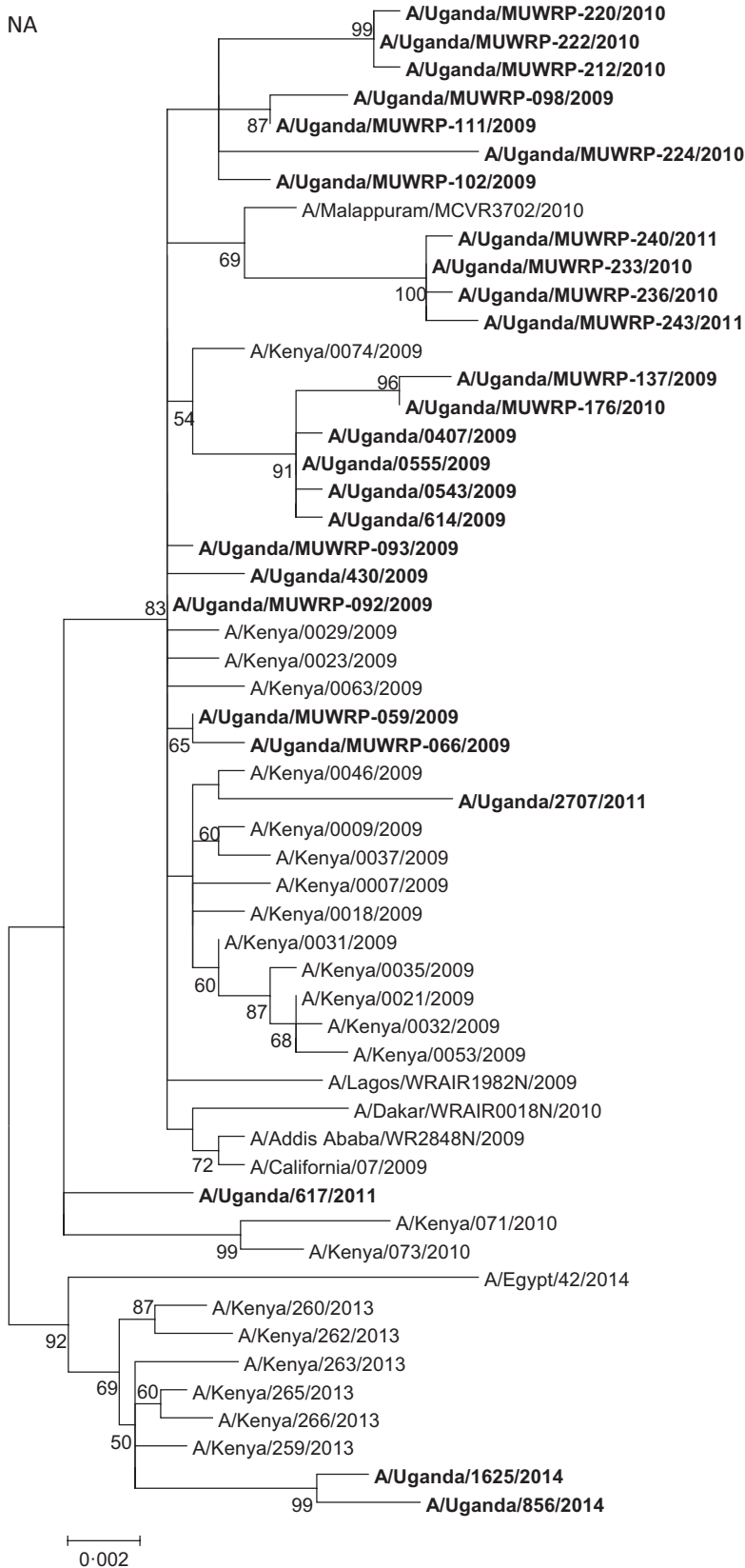
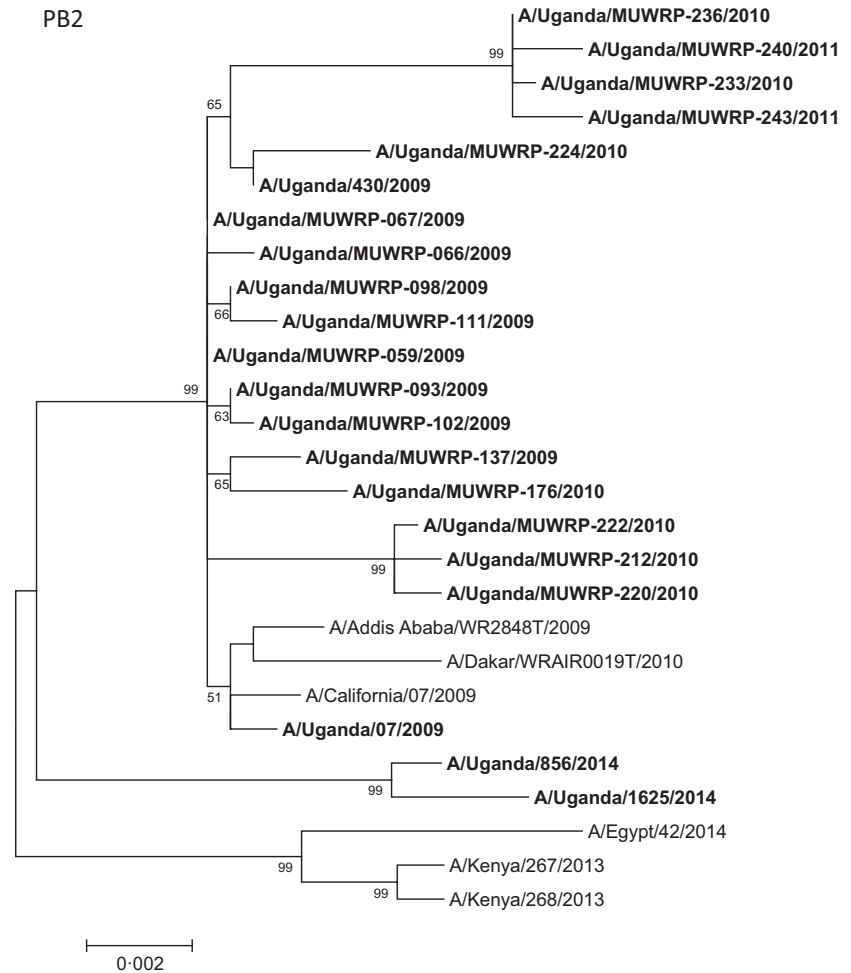


FIGURE 3 Phylogenetic tree of the neuraminidase (NA) gene segment of the Ugandan influenza A(H1N1)pdm09 viruses (in bold font) at the nucleotide level. The NA sequences of Ugandan H1N1 isolates were compared with relevant virus sequences available in GenBank: sequences from representative influenza A(H1N1)pdm09 isolates from Africa and closest blast hits from different regions of the world. Ugandan sequences available in the GISAID Epiflu database were also used (GISAID isolates ids: EPI_ISL_94767, EPI_ISL_105881, EPI_ISL_176803, EPI_ISL_176806, EPI_ISL_62338, EPI_ISL_65033, EPI_ISL_65034, EPI_ISL_68760, and EPI_ISL_71429). Sequences from vaccine strains [A/California/07/2009 (H1N1pdm09) and A/Brisbane/59/2007 (seasonal H1N1)] were also included. Bootstrap values (500 replicates) >50 are indicated on the nodes. * indicates partial sequence data

amino acid substitutions. Besides the common amino acid differences mentioned above, strains isolated from early 2009 were defined by the amino acid substitution N309D, and two isolates also had the amino acid substitutions M15V, I17V, and I188T, whereas strains isolated

from late 2010 and 2011 were defined by amino acid substitutions R220K, Q313R, I389K, and V394I. One strain isolated from 2010 had a unique NA genotype, with amino acid substitutions Q39R, S70N, N190S, Q309S, and N386S.

FIGURE 4 Phylogenetic tree of the polymerase 2 (PB2) gene segment of Ugandan influenza A(H1N1)pdm09 viruses (in bold font) at the nucleotide level. The PB2 sequences of Ugandan H1N1 isolates were compared with relevant virus sequences available in GenBank: sequences from representative influenza A(H1N1)pdm09 isolates from Africa and closest blast hits from different regions of the world. Ugandan sequences available in the GISAID Epiflu database were also used (GISAID isolates ids: EPI_ISL_176803, EPI_ISL_176806, EPI_ISL_62338, EPI_ISL_35548). Sequences from vaccine strains [A/California/07/2009 (H1N1)pdm09] and A/Brisbane/59/2007 (seasonal H1N1)] were also included. Bootstrap values (500 replicates) >50 are indicated on the nodes. * indicates partial sequence data



The phylogenetic tree of PB2 (Fig. 4) and other gene segments (data not shown) of the influenza A(H1N1)pdm09 strains exhibited a similar pattern of clustering of isolates. SF variant type analysis did not predict significant variations associated with increased virus fitness or transmission efficiency or significant antigenic differences. No molecular determinant of oseltamivir resistance was identified, but the determinant of amantadine resistance was suspected because of the Ser31Asn substitution in the M2 gene.

5 | DISCUSSION

Genome analysis helps understand the molecular evolution of viruses and other genetic factors unique to isolates that might help inform the formulation of vaccines for specific regions. While the influenza A(H1N1)pdm09 strains isolated in this study had specific amino acids that defined them, they were generally similar to the reference strains, with no significant antigenic variation from the vaccine strain A/California/07/2009.⁸ Consistent with findings from previous reports,^{9–12} the isolates from our study showed low genetic diversity. Despite this, the viruses isolated in different seasons clustered closely

in the phylogenetic trees and could fairly be separated from each other. The influenza A(H1N1)pdm09 strains completely replaced the seasonal H1N1 strains shortly after the 2009 pandemic, and these strains have not been isolated since that time, a phenomenon reported in other regions of the world.¹³

The limited information on whole-genome sequences from influenza virus strains from sub-Saharan Africa has constrained the region's contribution toward generating the information needed to develop new vaccines, therapies, and diagnostics for influenza. There are reported mutations in the influenza A(H1N1)pdm09 strains that can lead to increased fitness.¹⁴ However, on the basis of SF analysis, the isolates in our study did not carry amino acid substitutions that are associated with efficient transmission or fitness, although biological experiments would be needed to further confirm this observation. Genetic changes in influenza viruses often arise as a result of several years of re-assortment among different subtypes of viruses.¹⁵ Most of the isolates in our study shared high similarity with contemporary reference strains from other regions, which indicates that the Uganda viruses could be because of the introduction of new viruses into Uganda from the rest of the world rather than independent evolution of variants within Uganda.

6 | CONCLUSION

The sequence feature (SF) variant type analysis of the genomes of our isolates did not predict key mutations associated with functional, epitope, and structural features that are related to increased fitness and antigenic variations. Nevertheless, accumulated amino acid substitutions were identified and with the ability of these viruses to undergo rapid mutations, it is critical to maintain continuous surveillance to identify any important changes that might be associated with increase viral fitness and transmissibility.

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DISCLAIMER

The views expressed herein are those of the authors and do not represent those of the US Army Medical Research Directorate – Kenya, the Walter Reed Army Institute of Research, the US Army Medical Command, the US Department of the Army, or the US Department of Defense.

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