

# Immune-Associated Gene Signatures Serve as a Promising Biomarker of Immunotherapeutic Prognosis for Renal Clear Cell Carcinoma

Qi Wang<sup>1</sup>, Hanmin Tang<sup>1</sup>, Xuehui Luo<sup>1</sup>, Jie Chen<sup>1</sup>, Xinyue Zhang<sup>1</sup>, Xinyue Li<sup>1</sup>, Yuesen Li<sup>1</sup>, Yuetong Chen<sup>1</sup>, Yungang Xu<sup>2\*</sup> and Suxia Han<sup>1\*</sup>

<sup>1</sup> Department of Radiation Oncology, First Affiliated Hospital of Xi'an Jiaotong University, Xi'an, China, <sup>2</sup> Department of Cell Biology and Genetics, School of Basic Medical Sciences, Xi'an Jiaotong University Health Science Center, Xi'an, China

#### **OPEN ACCESS**

Edited by:

Guoqing Wang, Jilin University, China

#### Reviewed by: Zhen Tian,

Zhen Han, Zhengzhou University, China Liang Yu, Xidian University, China

> \*Correspondence: Suxia Han

shan87@mail.xjtu.edu.cn Yungang Xu yungang.xu@xjtu.edu.cn

#### Specialty section:

This article was submitted to Systems Immunology, a section of the journal Frontiers in Immunology

Received: 05 March 2022 Accepted: 22 April 2022 Published: 24 May 2022

#### Citation:

Wang Q, Tang H, Luo X, Chen J, Zhang X, Li X, Li Y, Chen Y, Xu Y and Han S (2022) Immune-Associated Gene Signatures Serve as a Promising Biomarker of Immunotherapeutic Prognosis for Renal Clear Cell Carcinoma. Front. Immunol. 13:890150. doi: 10.3389/fimmu.2022.890150 As the most common type of renal cell carcinoma (RCC), the renal clear cell carcinoma (ccRCC) is highly malignant and insensitive to chemotherapy or radiotherapy. Although systemic immunotherapies have been successfully applied to ccRCC in recent years, screening for patients who can benefit most from these therapies is still essential and challenging due to immunological heterogeneity of ccRCC patients. To this end, we implemented a series of deep investigation on the expression and clinic data of ccRCC from The Cancer Genome Atlas (TCGA) International Consortium for Cancer Genomics (ICGC). We identified a total of 946 immune-related genes that were differentially expressed. Among them, five independent genes, including SHC1, WNT5A, NRP1, TGFA, and IL4R, were significantly associated with survival and used to construct the immune-related prognostic differential gene signature (IRPDGs). Then the ccRCC patients were categorized into high-risk and low-risk subgroups based on the median risk score of the IRPDGs. IRPDGs subgroups displays distinct genomic and immunological characteristics. Known immunotherapy-related genes show different mutation burden, wherein the mutation rate of VHL was higher than 40% in the two IRPDGs subgroups, and SETD2 and BAP1 mutations differed most between two groups with higher frequency in the high-risk subgroup. Moreover, IRPDGs subgroups had different abundance in tumorinfiltrating immune cells (TIICs) with distinct immunotherapy efficacy. Plasma cells, regulatory cells (Tregs), follicular helper T cells (Tfh), and M0 macrophages were enriched in the high-risk group with a higher tumor immune dysfunction and rejection (TIDE) score. In contrast, the low-risk group had abundant M1 macrophages, mast cell resting and dendritic cell resting infiltrates with lower TIDE score and benefited more from immune checkpoint inhibitors (ICI) treatment. Compared with other biomarkers, such as TIDE and tumor inflammatory signatures (TIS), IRPDGs demonstrated to be a better biomarker for assessing the prognosis of ccRCC and the efficacy of ICI treatment with the promise in screening precise patients for specific immunotherapies.

Keywords: renal clear cell carcinoma (ccRCC), immune checkpoint inhibition, IRPDGs, tumor microenvironment, prognostic biomarker

## INTRODUCTION

Renal cell carcinoma (RCC) is one of the most common malignancies in humans. It is divided into renal papillary cell carcinoma, renal clear cell carcinoma (ccRCC), and chromophobe renal cell carcinoma. ccRCC is the most common type, accounting for over 80% of renal cancers (1). ccRCC is highly malignant and insensitive to chemotherapy or radiotherapy. At present, surgical resection is an effective treatment for early limited ccRCC. However, approximately 25% to 50% of primary patients experience recurrence five years after nephrectomy, and onethird of patients develop metastases; the overall 5-year survival rate is only 14% (1–3). ccRCC has distinct immunological features, such as a high degree of immune infiltration, relatively low mutational load, and relative sensitivity to anti-angiogenic therapy and immunotherapy (4).

Systemic immunotherapy has been successfully applied to ccRCC in recent years and has shown great benefits. Cytokinebased immunotherapy, including interleukin 2 (IL-2) and recombinant human interferon-alpha 2a (IFN-α2a), is the first approved immunotherapy for renal cell carcinoma. The combined results of several studies have shown an overall objective response rate (ORR) of 14% in IL-2-treated ccRCC patients, with 5% having complete response (CR) and 9% partial response (PR) (5). IFN- $\alpha$ 2a treatment has an ORR of 7.5%–10% in patients with advanced ccRCC (6, 7). The ORR of both cytokine treatments is generally low and is accompanied by significant toxicity to multiple major organs (2). Moore et al. discovered that the Von Hippel-Lindau (VHL) gene is mutated in more than 80% of renal cell carcinomas owing to promoter methylation, resulting in VHL protein dysfunction, which in turn induces hypoxia-inducible factor (HIF) activation and leads to abnormal expression of downstream vascular endothelial growth factor (VEGF) (8-10). After this discovery, tyrosine kinase inhibitors (TKIs), such as sunitinib and cabozantinib, which target the VEGF pathway and neovascularization, have been progressively used in ccRCC. The ORR of sunitinib monotherapy was 27%; however, previous studies have reported grade 3 or higher adverse reactions in 65%-70.6% of patients (11-13). Although the efficacy of TKIs over previous cytokine therapy has been confirmed, most patients with ccRCC develop drug resistance within one year (14).

The advent of immune checkpoint inhibitors (ICI) has solved the problems of low response rate and high toxicity in both cytokine and TKI treatments, and they are now the primary treatment option for ccRCC in clinical settings. Previous studies have demonstrated that immune checkpoints are involved in regulating cytotoxic T lymphocyte (CTL) activation and effector function to maintain autoimmune tolerance, and tumor cells evade the body's immunosurveillance in this way (15). ICI therapy achieves the antitumor effect by reactivating the immune response effect of CTL. ICIs currently used in the treatment of ccRCC mainly include pembrolizumab and nivolumab (PD-1 suppressants); avelumab and atezolizumab (PD-L1 suppressants); and ipilimumab (CTLA-4 suppressant). The ORR improves to 20%–42% with ICI monotherapy (13, 16–18). With the approval of TKIs and ICI in combination for ccRCC, multiple phase-III trials have provided many combination options of ICI and TKIs for clinical first- and second-line immunotherapy, with an ORR of 39.1%–71%. However, previous studies have reported that more than 90% of patients receiving the combination treatment of TKI and ICI had adverse events, of which 46%–82.4% had grade 3 or higher adverse events (13, 19–25). Although the ORR of ICI monotherapy is not satisfactory, the combination of ICI and TKIs improves the ORR to some extent, but also increases toxicity.

Screening for patients who can benefit most from these therapies is essential. Several studies have found that many factors, including tumor microenvironment (TME), influence the efficacy of ICI treatment (26, 27). Tumor-infiltrating immune cells (TIICs) in the TME of ccRCC play a key role in both pro- and anti-tumor processes, and are closely associated with clinical regression and response to immunotherapy (8, 10, 28, 29). According to Gulati et al., the TME of ccRCC is extremely heterogeneous. Patients with the same degree of progression may show different treatment responses and prognoses when receiving immunotherapy (30). Thus, there is a need to elucidate the interactions of various TIICs in the TME and to screen patients who are more suitable for immunotherapy. Current clinical treatment options usually rely on the International Metastatic Kidney Cancer Database (IMDC) or Memorial Sloan-Kettering Cancer Center (MSKCC) criteria (31, 32). The prognosis of ccRCC is primarily based on pathological staging (33); however, the current pathological staging system is inadequate to identify patients who are prone to adverse effects and low response rate when receiving ICI therapy. To advance the individualization of immunotherapy, it is urgent to discover new potential biomarkers to predict clinical response to ICI and prognosis.

In this study, we collected KIRC-mRNA data and related clinical information for ccRCC from TCGA. We initially screened 24 immune-related hub genes by applying WGCNA and univariate Cox regression analysis. Then, we constructed an independent immune-related prognostic differential gene signature (IRPDGs) containing five genes using multivariate Cox regression analysis. Data from ICGC were used for external validation. We defined high- and low-risk subgroups of the IRPDGs based on the median risk score and used gene set enrichment analysis (GSEA) to explore important signaling pathways and potential molecular mechanisms enriched in the IRPDGs subgroups. We also downloaded the corresponding mutation data in TCGA for a comprehensive analysis of somatic mutations. We used the deconvolution algorithm CIBERSORT (34) and single sample genes set enrichment analysis (ssGSEA) to analyze the relative infiltration abundance of 22 different TIICs and 29 immune-associated functional indicators in different subgroups. Receiver operating characteristic (ROC) analysis was performed to further assess the prognostic value of the IRPDGs. Compared with other biomarkers such as tumor immune dysfunction and rejection (TIDE) and 18-gene tumor inflammatory signatures (TIS), the IRPDGs demonstrated significant advantages in predicting the

prognosis of patients treated with immunotherapy. The workflow for this study is shown in **Figure 1**.

## MATERIALS AND METHODS

### **Patients and Data**

We downloaded RNA-sequencing data and detailed clinical information for ccRCC from TCGA (https://portal.gdc.cancer. gov/), which contains 539 cancer samples and 72 paracancerous samples. A total of 91 specimens were obtained from the ICGC database (https://dcc.icgc.org/releases) for external validation. Perl language (version 5.30.2) was used to extract clinical information. The list of immune-related genes was obtained from the Immport (https://www.Immport.org/home/) and the InnateDB (https://www.innateDBdb.com/) databases.

### Search for Immune-Related Hub Genes

The DEGs were screened from 539 ccRCC samples and 72 paracancerous samples with the LIMMA package (p-value < 0.05, |log2FC| > 1), and were intersected with the downloaded list of the immune-related genes to obtain immune-related DEGs. The Gene Ontology (GO) and the Kyoto Encyclopedia of Genes and Genomes (KEGG) were used to explore the major pathways enriched in immune-related hub genes. We used WGCNA (35) to identify hub genes. We first calculated Pearson correlation coefficients, constructed a similarity matrix using the expression data, and converted the similarity matrix into a neighborhood matrix with a soft threshold of 3; we then built TOM matrix based on the neighborhood matrix, clustered genes with a distance of 1 -TOM, and identified and constructed different gene modules for visualization. Seven modules were obtained, and the top two modules (brown and green modules) with the most significant correlations were chosen. We filtered the top 40 core genes in the two modules by using CytoHuba (36). Finally, the best cutoff values of the core genes were calculated by the R package, and a total of 24 immune-related hub genes closely related to ccRCC survival were screened for further analysis.

## **Construction and Validation of IRPDGs**

Five genes independently and significantly associated with ccRCC prognosis were screened from 24 hub genes by multivariate Cox regression analysis. The IRPDGs was defined as the result of multiplying the expression value of each gene separately by the sum of the weights in the Cox model. Different subgroups of IRPDGs were defined according to the median risk score. The IGCG data (n = 91) were used for validation. Kaplan-Meier survival analysis with log-rank tests was performed to assess the prognostic power of the IRPDGs (P < 0.05).

# Molecular Profiling in the IRPDGs Subgroups

The GSEA was used to reveal important signaling pathways and potential molecular mechanisms enriched in different subgroups. A reference genome (c2.Cp.kegg.v7.4.Symbols.Gmt) was taken to compare immunophenotypes between the IRPDGs subgroups (with p < 0.05 and FDR < 0.25 as cutoff values). We downloaded mutation information associated with ccRCC from TCGA, and the Maftools package in the R software was used to comprehensively analyze somatic mutations between the IRPDGs subgroups.



# Assessment of Immune Characteristics and Immunotherapy

Based on ccRCC gene expression profiles, the abundance of 22 TIICs in different subgroups was quantified by using CIBERSORT (https://CIBERSORT.stanford). We used the GSVA package for ssGSEA of the enrichment levels of 29 immune-related functional indicators. Using the "survival ROC" R-package for survival analyses, we calculated areas under the curve (AUC) at different cutoff time points. To further explore the value of the IRPDGs in prognosis, we analyzed differences in TIDE score, microsatellite instability (MSI), T-cell dysfunction, and T-cell rejection effects in the IRPDGs subgroups. We compared the prognostic ability of the IRPDGs, TIDE, and TIS. The TIDE score was calculated online (http://tide.dfci.atherard.edu/), and the TIS score was defined as the mean of the log2-scaled normalized expression of 18 marker genes (37).

### **Statistical Analysis**

R program was used in most of our research. Pearson correlation tests were used to analyze the characteristics of immune-related hub genes. Differences between variables were analyzed using independent t-tests and chi-square tests. The prognostic value of the IRPDGs compared with clinicopathological features of ccRCC was assessed by the K–M survival and Cox regression analysis. Two-tailed P < 0.05 was considered significant.

## RESULTS

### Screening of Immune-Related Hub Genes

We screened a total of 9459 DEGs from the TCGA cohort (539 cancer samples, 72 paracancerous samples). The list of the immunerelated genes was obtained from InnateDB and ImmPort databases. We obtained 946 immune-related DEGs by taking intersections (Figures 2A, B). Then, we performed functional enrichment analysis and identified 2137 GOs and 87 KEGG-related pathways (Figures 2C, D). Circle diagrams and specific tabular data are shown in Figure S1 and Table S1. Candidate genes (n = 946) were then subjected to WGCNA. Logarithm log(K) had nodes with connectivity K and negatively correlated with logarithm log(P(K)) of node probabilities, with a correlation coefficient greater than 0.9. Based on the relationship between soft-threshold power and average connectivity, the optimal power for scale-free network-based networks was determined as 3. Seven modules were finally settled. Based on Pearson correlation coefficients between the modules and sample features, brown and green modules closely related to ccRCC were selected, and the top 40 core genes were identified for a followup study by CytoHuba (Figures 2E-G). The Kaplan-Meier survival analysis and univariate Cox analyses identified 24 immune-related hub genes that were closely associated with RCC survival (Figure 3A, Figure S2).

# Construction of the IRPDGs and Clinical Variability Analysis

Multivariate Cox regression analysis of the 24 immune-related hub genes finally identified 5 independent prognostic genes (SHC1,

WNT5A, NRP1, TGFA, IL4R) that significantly affected ccRCC survival (**Figure 3B**). We then constructed prognostic models for the IRPDGs with the following formula: IRPDGs = expression value of SHC1 \* (0.12) + expression value of WNT5A \* (0.37) + expression value of NRP1 \* (-0.12) + expression value of TGFA \* (-0.06) + expression value of IL4R \* (0.12). The high-risk versus low-risk IRPDGs subgroups were defined based on the median risk score as risk threshold. The results showed that patients in the low-risk subgroup had a better prognosis than those in the high-risk subgroup (P < 0.001, log-rank test) (**Figure 3C**). Using ICGC data as external validation yielded results consistent with those of the TCGA dataset, both with P-values < 0.05 (**Figure 3D**).

The univariate Cox regression analysis including clinical and pathological information showed that age, IRPDGs, grade, and stage were high-risk factors associated with ccRCC prognosis. Further multivariate Cox analysis revealed that the IRPDGs remained an independent prognostic marker after adjusting for other factors (**Figures 4A, B**). The distribution of clinicopathological characteristics of the TCGA patient cohort across the IRPDGs subgroups is shown in **Figure 4C**, with detailed results in **Table S3**. **Figures 4D, E** specifically demonstrate the significant differences in the distribution of tumor stage and grade between the two risk subgroups.

# Molecular Characterization in the IRPDGs Subgroups

Molecular pathways and potential mechanisms enriched in the IRPDGs subgroups were identified by the GSEA. Pathways related to arachidonic acid metabolism and cytokine-cytokine interactions were enriched in the IRPDGs-high subgroup, while citratetricarboxylic acid (TCA) cycle and neuroactive ligand-receptor interaction pathways were enriched in the IRPDGs-low subgroup (Figures 5A, B). The detailed results of the GSEA are presented in Table S2. To further explore the immunological properties, we analyzed the somatic mutations between the subgroups. The number of mutations was higher overall in the high-risk subgroup than in the low-risk subgroup of the IRPDGs. Of these, the frequency of missense mutations was the highest, followed by shift deletions and nonsense mutations. Figures 5C, D shows the top 20 genes with the highest mutation rate in both subgroups. The mutation rate in VHL was higher than 40% in both subgroups. BAP1 and SETD2 mutation rates differed most between the two subgroups and were mostly seen in the high-risk subgroup.

# Immunological Characteristics Among the IRPDGs Subgroups

The CIBERSORT is an analysis tool developed in Java and R for characterizing cell composition from tissue gene expression profiles. It uses a deconvolution algorithm to estimate the abundance of member cell types in mixed cell populations based on gene expression data (34). Here, we use CIBERSORT to explore the proportion of various TIICs in the different IRPDGs subgroups. The high-risk subgroup was mainly rich in Tfh, Tregs, plasma cells, and M0 macrophages; the low-risk subgroup had more M1 macrophages, dendritic cells resting, and mast cells resting (**Figure 6A**). The ssGSEA was applied to



explore indicators of immune function involved in the two risk subgroups. The results showed that the IRPDGs-high subgroup was more enriched in CD8+ T cells, cytolytic activity, promotion of inflammogenesis, T cell co-inhibition, and costimulation, and type I IFN responses; in contrast, the IRPDGs-low subgroup was enriched in mast cells and functions related to type II IFN responses (**Figure 6B**). We then explored the value of the abovementioned differential TIICs for prognosis and found that patients with higher expression scores for mast cells resting and dendritic cells resting had a better prognosis, while those with high expression scores for plasma cells, follicular helper T cells, Tregs, and M0 macrophages had a worse prognosis. This was consistent with the prognostic model results that the low-risk subgroup of IRPDGs had a better prognosis (**Figures 6C-H**).

# Survival Analysis Between the IRPDGs Subgroups

Immunotherapy efficacy between the subgroups of IRPDGs was assessed using TIDE. A higher TIDE score is associated with



poorer ICI therapy outcomes and poorer patient survival (38). Calculation of TIDE score for the IRPDGs subgroups showed higher TIDE scores in the high-risk subgroup than in the lowrisk subgroup. This means that patients with low IRPDGs scores have a greater benefit from immunotherapy. In addition, the IRPDGs-low subgroup had a higher MSI and lower T-cell rejection and T-cell dysfunction, with p-values less than 0.05 for all of the outcomes (**Figures 7A–D**). Using the ROC analysis to evaluate the model for predicting disease progression, we showed that our model was able to predict the survival rates of ccRCC patients at 1, 2, and 3 years with good accuracy and that the predictive value of the IRPDGs for overall survival (OS) was significantly better than that of TIDE or TIS. The AUC for IRPDGs, TIDE, and TIS was 0.741, 0.543, and 0.516, respectively (**Figures 7E, F**).

### DISCUSSION

Considering that ccRCC is an immunogenic and vascularized tumor (27), ICI therapy has shown great potential and has been approved by the US Food and Drug Administration (FDA) as the standard treatment for patients with early and advanced metastatic ccRCC (4, 8). However, the ORR to ICI therapy remains low, with only a small proportion of patients achieving long-term, sustained outcomes after the treatment. The combination of ICIs and TKIs is now increasingly studied and is superior to ICI monotherapy in terms of OS and ORR, but it is also associated with greater toxicity. Screening patients with potentially greater benefit from immunotherapy is necessary. The TME consists of tumor cells and other nonmalignant cells, including the surrounding TIICs, fibroblasts, and stromal cells (39, 40). Among them, TIICs regulate tumor development at different stages (41). We have knowledge from previous studies that ccRCC patients treated with ICI show different treatment responses probably due to the heterogeneity of TME (30). Therefore, an in-depth investigation of the interaction mechanisms and biological functions of TIICs in the TME can help to optimize current immunotherapy. Given the limited assessment tools available, new biomarkers that effectively predict the clinical response and prognosis of ICI therapy are urgently needed to achieve precision immunotherapy.

A total of 946 immune-related DEGs were identified in our study. The final IRPDGs was constructed based on the five independent prognostically significantly associated genes and was validated in the ICGC dataset. After adjusting for clinical and pathological variables, the IRPDGs remained an independent prognostic factor. The five genes used to construct the IRPDGs were WNT5A, NRP1, TGFA, SHC1, and IL4R. The Wnt family member 5A (WNT5A) is a classical noncanonical WNT ligand



grades in the IRPDGs subgroup of the TCGA cohort.

that belongs to a large family of WNT cysteine-rich secretory glycoproteins and is involved in multiple signaling pathways that regulate a variety of cellular processes (42, 43). WNT5A mediates the activation of Tregs and tumor-associated macrophages (TAM), which thereby induces immunosuppression during the progression of castration-resistant prostate cancer (44). WNT5A may be a potential target for the prevention of gastrointestinal radiotoxicity because its absence impairs the ability of individual intestinal stem cells (ISCs) to form crypt spheroids after irradiation, whereas the addition of WNT5A saves ISCs from radiation-induced cell death (45). Tumor angiogenesis is considered an important phenomenon in the progression of tumors and is associated with overexpression of Neuropilin1 (NRP1) receptors. NRP1 is a nonenzymatic transmembrane glycoprotein, and recent studies have shown that high NRP1 surface expression leads to more suppressive Tregs and reduced progression-free survival (PFS). NRP1 is an independent novel marker in RCC (46), which forms a complex with transexpressed VEGFR2 and inhibits tumor angiogenesis, thereby improving

patient survival. NRP1 is also highly expressed in glioblastoma multiforme (GBM) and is closely associated with the prognosis of GBM patients (47). As a member of the epidermal growth factor family, TGFA binds to EGFR to activate a series of signaling pathways that regulate cell biological processes such as cell proliferation, migration, differentiation, and energy metabolism (48, 49). TGFA also promotes cell proliferation and epithelialmesenchymal transition (EMT) in prostate, liver, and breast cancers (50-53). SHC1 is a kind of skeletal protein that is associated with cell membrane metabolism and exerts its regulatory role mainly through the EGFR pathway (54, 55). Overexpression of SHC1 is associated with poor survival in stage IIA colon cancer (56). SHC1 encodes a splice protein that acts on various tyrosine kinase signaling pathways and is thought to be a key mediator in promoting immunosuppressed breast cancer (55). Interleukin 4 receptor (IL4R) is a type I cytokine receptor produced by activated Th2 cells and mast cells. The IL-4/ IL-4R axis plays a role in immunity and inflammation, and IL4R has been shown to interact with SHC1 (57, 58).



Our survival analysis revealed that patients in the low-risk subgroup of the IRPDGs had a better prognosis. We further explored the prognostic characteristics of ccRCC patients in the IRPDGs subgroups using TIDE. Peng Jiang established the online TIDE score combining T cell dysfunction and T cell exclusion characteristics, which predicts immunotherapy responsiveness by calculating genome-wide expression profiles of pre-treatment patients, simulating tumor immune escape with different levels of CTLs (38). TIDE has shown outstanding advantages over other biomarkers for the prediction of ICI therapy responsiveness in prospective clinical trials. Our results showed higher TIDE scores and lower T-cell dysfunction and Tcell rejection in the high-risk subgroup of the IRPDGs. This suggests that patients in IRPDGs-high groups had more immune dysfunction and were less responsive to immunotherapy. In contrast, the IRPDGs-low group with a lower TIDE score and higher MSI score had a greater benefit from immunotherapy and a better prognosis. Interferon-gamma-7-related 18-gene T-cell inflammatory signaling (TIS) is a good predictor of ICI therapy reaction and prognosis in a variety of tumors (37, 59, 60). Through the analysis of the ROC curve, we found that the prediction value of total survival time was significantly better for the IRPDGs than for TIDE and TIS, with the AUC of IRPDGs, TIDE, and TIS being 0.741, 0.543, and 0.516, respectively. This means that the IRPDGs can effectively identify high-risk patients and select those who are more suitable for immunotherapy. Moreover, the IRPDGs, which consist of only five genes, are also easier to detect.

The TME plays a crucial role in the tumors' growth and migration. The GO enrichment and KEGG pathway analyses of the immune-related DEGs revealed a variety of immune-associated pathways and functions, including leukocyte-mediated adaptive immunity, cytokine-cytokine receptor interactions, chemokine, Tcell receptor, and B-cell receptor signaling pathways. Based on the results above, we further explored the mechanisms of TIICs in ccRCC immunomodulation through the GSEA. Our study showed large differences in the composition of TIICs between the IRPDGs subgroups. Six TIICs that differed significantly between the subgroups were significantly associated with the survival of patients with ccRCC. Tfh, Tregs, plasma cells, and M0 macrophages were more abundant in the IRPDGs high-risk subgroup, while M1 macrophages, mast cell resting, and dendritic cell resting was more frequently seen in the low-risk subgroup. As one of the important immunosuppressive cell types, Tregs are highly immunosuppressive to effector cells. It has been confirmed that with the development of ccRCC, the infiltration of Treg cells increases, which indicates a poor prognosis (61, 62); and the degree of Treg infiltration is lower in some patients treated with ICI to CR (63). Previous studies have suggested that the infiltration of ccRCC by memory B cells and Tfh indicates a poor prognosis; in contrast, dendritic cell resting, mast cell resting, and eosinophilia are significantly associated with improved prognosis (64). Stefanie Regine Dannenmann et al. showed that pro-inflammatory macrophage phenotype (M1) is induced by IFN $\gamma$  and lipopolysaccharide (LPS), and some factors associated with M1 correlate with prolonged survival in ccRCC (65). Our findings



support these conclusions. Further exploration of the molecular immune functions between the IRPDGs subgroups revealed that the high-risk subgroup was more likely to involve CD8+ T cells and tumor-infiltrating lymphocytes (TILs), whereas the low-risk subgroup involved more mast cells and type II IFN responses. CD8+ T cell infiltration is associated with longer PFS and OS in most solid tumors (66); in contrast, in ccRCC, infiltration by CD8+ T cells predicts a poor prognosis (67–69). Type II interferon IFN-γ is released by T cells, NK cells, and macrophages. A recent study has explored the relationship between PD-L1 expression and IFN- $\gamma$  by RT-PCR and western blot analysis; it showed that PD-L1 was induced by typical IFN-y signaling in ccRCC cell lines, and IFNγ-positive tumor tissues had high levels of PD-L1-mRNA expression and showed better OS (70). These findings further supported that the low-risk subgroup of the IRPDGs had better prognosis and immune efficacy than the high-risk subgroup.

A prerequisite for the efficacy of ICI is the generation of immunogenic neoantigens by tumor-specific mutations and the promotion of immune infiltration. Tumor mutational load (TMB) is an important predictor of ICI efficacy in many malignant tumors (71). We explored differences in somatic mutations between the subgroups and showed that missense mutations occurred most frequently between the two subgroups, followed by shift deletions and nonsense mutations. Missense mutations are associated with increased tumor antigen presentation and CD8+ T cell infiltration in most of the solid tumors (61, 72, 73). The VHL mutation rate was higher than 40% in both IRPDGs subgroups, and genes with the largest mutational differences were SETD2 and BRCA1-associated protein-1 (BAP1), which accounted for 18%:4% and 14%:5%, respectively. SETD2 is a histone methyltransferase whose normal function is of great importance in genome stability and DNA damage repair (74, 75). SETD2 mutations are associated with





shorter PFS and OS in metastatic RCC and breast cancer (76). BAP1 is a kind of deubiquitinase that regulates various cellular functions including proliferation, differentiation, and metabolism (77, 78). BAP1 is mutated in lung cancer, thyroid cancer, kidney cancer, melanoma, and mesothelioma (79–81). Previous studies have found that the rate of BAP1 mutation increases with the increase in the ccRCC stage. BAP1 and SETD2 mutations are associated with disease progression and indicate a worse prognosis (82, 83). These data are consistent with our survival results; namely, patients with high SETD2 and BAP1 mutations in the IRPDGs-high subgroup had a poorer prognosis than those in the low IRPDGs subgroup.

In conclusion, with the advent of the era of accurate immunotherapy, IRPDGs shows obvious advantages in evaluating the response to ICI treatment and in predicting the prognosis. TIICs, which are significantly related to the prognosis of ccRCC in the IRPDGs subgroups, are helpful to guide future immunotherapy. The IRPDGs is a promising tool to guide clinical practice. More indepth studies are needed to further confirm this point.

#### DATA AVAILABILITY STATEMENT

The raw datasets used in this study can be found in online databases, respectively, which are described in details in Methods sections. The source data and codes for reproducing the results and figures are freely accessible from https://www.jianguoyun.com/p/Df1PX6AQq5usChjNyrAEIAA.

### REFERENCES

 Hsieh JJ, Purdue MP, Signoretti S, Swanton C, Albiges L, Schmidinger M, et al. Renal Cell Carcinoma. Nat Rev Dis Primers (2017) 3:17009. doi: 10.1038/nrdp.2017.9

## **AUTHOR CONTRIBUTIONS**

QW: Research design and drafting of manuscripts. HT, XLu, and JC: Data analysis. XZ, XLi, YC, and YL: Literature review. YX, SH: Reviewing, revising, and guiding the writing of the manuscript. All authors contributed to the article and approved the submitted version.

### FUNDING

This study was funded by the Shaanxi Provincial Innovation Capacity Support Program (Approval No. 2018TD-002).

### ACKNOWLEDGMENTS

The authors would like to thank Xi'an Jiaotong University for supporting bioinformatics and related training. We thank LetPub (www.letpub.com) for its linguistic assistance during the preparation of this manuscript.

### SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2022.890150/ full#supplementary-material

 Janzen NK, Kim HL, Figlin RA, Belldegrun AS. Surveillance after Radical or Partial Nephrectomy for Localized Renal Cell Carcinoma and Management of Recurrent Disease. Urol Clin North Am (2003) 30(4):843–52. doi: 10.1016/ s0094-0143(03)00056-9

- Gong J, Maia MC, Dizman N, Govindarajan A, Pal SK. Metastasis in Renal Cell Carcinoma: Biology and Implications for Therapy. *Asian J Urol* (2016) 3 (4):286–92. doi: 10.1016/j.ajur.2016.08.006
- Kim MC, Jin Z, Kolb R, Borcherding N, Chatzkel JA, Falzarano SM, et al. Updates on Immunotherapy and Immune Landscape in Renal Clear Cell Carcinoma. *Cancers* (2021) 13(22):5856. doi: 10.3390/cancers13225856
- 5. Malek TR. The Biology of Interleukin-2. Annu Rev Immunol (2008) 26:453–79. doi: 10.1146/annurev.immunol.26.021607.090357
- Minasian LM, Motzer RJ, Gluck L, Mazumdar M, Vlamis V, Krown SE. Interferon Alfa-2a in Advanced Renal Cell Carcinoma: Treatment Results and Survival in 159 Patients with Long-Term Follow-Up. J Clin Oncol Off J Am Soc Clin Oncol (1993) 11(7):1368–75. doi: 10.1200/jco.1993.11.7.1368
- Negrier S, Escudier B, Lasset C, Douillard JY, Savary J, Chevreau C, et al. Recombinant Human Interleukin-2, Recombinant Human Interferon Alfa-2a, or Both in Metastatic Renal-Cell Carcinoma. Groupe Fr D'immunoth'er N Engl J Med (1998) 338(18):1272–8. doi: 10.1056/nejm199804303381805
- Vuong L, Kotecha RR, Voss MH, Hakimi AA. Tumor Microenvironment Dynamics in Clear-Cell Renal Cell Carcinoma. *Cancer Discov* (2019) 9 (10):1349–57. doi: 10.1158/2159-8290.Cd-19-0499
- Moore LE, Nickerson ML, Brennan P, Toro JR, Jaeger E, Rinsky J, et al. Von Hippel-Lindau (Vhl) Inactivation in Sporadic Clear Cell Renal Cancer: Associations with Germline Vhl Polymorphisms and Etiologic Risk Factors. *PloS Genet* (2011) 7(10):e1002312. doi: 10.1371/journal.pgen.1002312
- van den Heuvel C, van Ewijk A, Zeelen C, de Bitter T, Huynen M, Mulders P, et al. Molecular Profiling of Druggable Targets in Clear Cell Renal Cell Carcinoma through Targeted Rna Sequencing. *Front Oncol* (2019) 9:117. doi: 10.3389/fonc.2019.00117
- Choueiri TK, Hessel C, Halabi S, Sanford B, Michaelson MD, Hahn O, et al. Cabozantinib Versus Sunitinib as Initial Therapy for Metastatic Renal Cell Carcinoma of Intermediate or Poor Risk (Alliance A031203 Cabosun Randomised Trial): Progression-Free Survival by Independent Review and Overall Survival Update. *Eur J Cancer (Oxf Engl)* (2018) 1990):94. doi: 10.1016/j.ejca.2018.02.012
- Hutson TE, Lesovoy V, Al-Shukri S, Stus VP, Lipatov ON, Bair AH, et al. Axitinib Versus Sorafenib as First-Line Therapy in Patients with Metastatic Renal-Cell Carcinoma: A Randomised Open-Label Phase 3 Trial. *Lancet* Oncol (2013) 14(13):1287–94. doi: 10.1016/s1470-2045(13)70465-0
- Motzer RJ, Tannir NM, McDermott DF, Arén Frontera O, Melichar B, Choueiri TK, et al. Nivolumab Plus Ipilimumab Versus Sunitinib in Advanced Renal-Cell Carcinoma. N Engl J Med (2018) 378(14):1277–90. doi: 10.1056/NEJMoa1712126
- Mollica V, Di Nunno V, Gatto L, Santoni M, Scarpelli M, Cimadamore A, et al. Resistance to Systemic Agents in Renal Cell Carcinoma Predict and Overcome Genomic Strategies Adopted by Tumor. *Cancers* (2019) 11(6):830. doi: 10.3390/cancers11060830
- Li Y, Li F, Jiang F, Lv X, Zhang R, Lu A, et al. A Mini-Review for Cancer Immunotherapy: Molecular Understanding of Pd-1/Pd-L1 Pathway & Translational Blockade of Immune Checkpoints. *Int J Mol Sci* (2016) 17 (7):1151. doi: 10.3390/ijms17071151
- Motzer RJ, Rini BI, McDermott DF, Redman BG, Kuzel TM, Harrison MR, et al. Nivolumab for Metastatic Renal Cell Carcinoma: Results of a Randomized Phase Ii Trial. J Clin Oncol Off J Am Soc Clin Oncol (2015) 33 (13):1430–7. doi: 10.1200/jco.2014.59.0703
- Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, et al. Safety, Activity, and Immune Correlates of Anti-Pd-1 Antibody in Cancer. *N Engl J Med* (2012) 366(26):2443–54. doi: 10.1056/NEJMoa1200690
- Motzer RJ, Escudier B, McDermott DF, George S, Hammers HJ, Srinivas S, et al. Nivolumab Versus Everolimus in Advanced Renal-Cell Carcinoma. *N Engl J Med* (2015) 373(19):1803–13. doi: 10.1056/NEJMoa1510665
- Rini BI, Plimack ER, Stus V, Gafanov R, Hawkins R, Nosov D, et al. Pembrolizumab Plus Axitinib Versus Sunitinib for Advanced Renal-Cell Carcinoma. N Engl J Med (2019) 380(12):1116-27. doi: 10.1056/ NEJMoa1816714
- Choueiri TK, Powles T, Burotto M, Escudier B, Bourlon MT, Zurawski B, et al. Nivolumab Plus Cabozantinib Versus Sunitinib for Advanced Renal-Cell Carcinoma. N Engl J Med (2021) 384(9):829–41. doi: 10.1056/ NEJMoa2026982

- 21. Choueiri TK, Motzer RJ, Rini BI, Haanen J, Campbell MT, Venugopal B, et al. Updated Efficacy Results from the Javelin Renal 101 Trial: First-Line Avelumab Plus Axitinib Versus Sunitinib in Patients with Advanced Renal Cell Carcinoma. Ann Oncol Off J Eur Soc Med Oncol (2020) 31(8):1030–9. doi: 10.1016/j.annonc.2020.04.010
- 22. Powles T, Plimack ER, Soulières D, Waddell T, Stus V, Gafanov R, et al. Pembrolizumab Plus Axitinib Versus Sunitinib Monotherapy as First-Line Treatment of Advanced Renal Cell Carcinoma (Keynote-426): Extended Follow-Up from a Randomised, Open-Label, Phase 3 Trial. *Lancet Oncol* (2020) 21(12):1563–73. doi: 10.1016/s1470-2045(20)30436-8
- Motzer RJ, Penkov K, Haanen J, Rini B, Albiges L, Campbell MT, et al. Avelumab Plus Axitinib Versus Sunitinib for Advanced Renal-Cell Carcinoma. N Engl J Med (2019) 380(12):1103–15. doi: 10.1056/ NEJMoa1816047
- Motzer R, Alekseev B, Rha SY, Porta C, Eto M, Powles T, et al. Lenvatinib Plus Pembrolizumab or Everolimus for Advanced Renal Cell Carcinoma. N Engl J Med (2021) 384(14):1289–300. doi: 10.1056/NEJMoa2035716
- 25. Tomita Y, Motzer RJ, Choueiri TK, Rini BI, Miyake H, Uemura H, et al. Efficacy and Safety of Avelumab Plus Axitinib (a + Ax) Versus Sunitinib (S) in Elderly Patients with Advanced Renal Cell Carcinoma (Arcc): Extended Follow-Up Results from Javelin Renal 101. *J Clin Oncol* (2021) 39 (6\_suppl):301. doi: 10.1200/JCO.2021.39.6\_suppl.301
- Nishino M, Ramaiya NH, Hatabu H, Hodi FS. Monitoring Immune-Checkpoint Blockade: Response Evaluation and Biomarker Development. *Nat Rev Clin Oncol* (2017) 14(11):655–68. doi: 10.1038/nrclinonc.2017.88
- Díaz-Montero CM, Rini BI, Finke JH. The Immunology of Renal Cell Carcinoma. Nat Rev Nephrol (2020) 16(12):721–35. doi: 10.1038/s41581-020-0316-3
- Borcherding N, Vishwakarma A, Voigt AP, Bellizzi A, Kaplan J, Nepple K, et al. Mapping the Immune Environment in Clear Cell Renal Carcinoma by Single-Cell Genomics. *Commun Biol* (2021) 4(1):122. doi: 10.1038/s42003-020-01625-6
- 29. Chen DS, Mellman I. Elements of Cancer Immunity and the Cancer-Immune Set Point. *Nature* (2017) 541(7637):321–30. doi: 10.1038/nature21349
- Gulati S, Martinez P, Joshi T, Birkbak NJ, Santos CR, Rowan AJ, et al. Systematic Evaluation of the Prognostic Impact and Intratumour Heterogeneity of Clear Cell Renal Cell Carcinoma Biomarkers. *Eur Urol* (2014) 66(5):936-48. doi: 10.1016/j.eururo.2014.06.053
- Motzer RJ, Mazumdar M, Bacik J, Berg W, Amsterdam A, Ferrara J. Survival and Prognostic Stratification of 670 Patients with Advanced Renal Cell Carcinoma. J Clin Oncol Off J Am Soc Clin Oncol (1999) 17(8):2530–40. doi: 10.1200/jco.1999.17.8.2530
- 32. Heng DY, Xie W, Regan MM, Warren MA, Golshayan AR, Sahi C, et al. Prognostic Factors for Overall Survival in Patients with Metastatic Renal Cell Carcinoma Treated with Vascular Endothelial Growth Factor-Targeted Agents: Results from a Large, Multicenter Study. J Clin Oncol Off J Am Soc Clin Oncol (2009) 27(34):5794–9. doi: 10.1200/jco.2008.21.4809
- 33. Frank I, Blute ML, Cheville JC, Lohse CM, Weaver AL, Zincke H. An Outcome Prediction Model for Patients with Clear Cell Renal Cell Carcinoma Treated with Radical Nephrectomy Based on Tumor Stage, Size, Grade and Necrosis: The Ssign Score. J Urol (2002) 168(6):2395–400. doi: 10.1097/01.ju.0000035885.91935.d5
- Newman AM, Liu CL, Green MR, Gentles AJ, Feng W, Xu Y, et al. Robust Enumeration of Cell Subsets from Tissue Expression Profiles. *Nat Methods* (2015) 12(5):453–7. doi: 10.1038/nmeth.3337
- Langfelder P, Horvath S. WGCNA: An R Package for Weighted Correlation Network Analysis. BMC Bioinf (2008) 9:559. doi: 10.1186/1471-2105-9-559
- 36. Chin CH, Chen SH, Wu HH, Ho CW, Ko MT, Lin CY. Cytohubba: Identifying Hub Objects and Sub-Networks from Complex Interactome. BMC Syst Biol (2014) 8 Suppl 4(Suppl 4):S11. doi: 10.1186/1752-0509-8-s4s11
- Ayers M, Lunceford J, Nebozhyn M, Murphy E, Loboda A, Kaufman DR, et al. Ifn-Γ-Related Mrna Profile Predicts Clinical Response to Pd-1 Blockade. *J Clin Invest* (2017) 127(8):2930–40. doi: 10.1172/jci91190
- Jiang P, Gu S, Pan D, Fu J, Sahu A, Hu X, et al. Signatures of T Cell Dysfunction and Exclusion Predict Cancer Immunotherapy Response. *Nat Med* (2018) 24(10):1550–8. doi: 10.1038/s41591-018-0136-1

- Zheng B, Xie F, Cheng F, Wang J, Yao Z, He W, et al. Integrative Analysis of Immune-Related Genes in the Tumor Microenvironment of Renal Clear Cell Carcinoma and Renal Papillary Cell Carcinoma. *Front Mol Biosci* (2021) 8:760031. doi: 10.3389/fmolb.2021.760031
- Wu T, Dai Y. Tumor Microenvironment and Therapeutic Response. Cancer Lett (2017) 387:61–8. doi: 10.1016/j.canlet.2016.01.043
- Lei X, Lei Y, Li JK, Du WX, Li RG, Yang J, et al. Immune Cells within the Tumor Microenvironment: Biological Functions and Roles in Cancer Immunotherapy. *Cancer Lett* (2020) 470:126–33. doi: 10.1016/j.canlet. 2019.11.009
- 42. Shen C, Liu J, Wang J, Zhong X, Dong D, Yang X, et al. Development and Validation of a Prognostic Immune-Associated Gene Signature in Clear Cell Renal Cell Carcinoma. *Int Immunopharmacol* (2020) 81:106274. doi: 10.1016/ j.intimp.2020.106274
- van Amerongen R, Mikels A, Nusse R. Alternative Wnt Signaling Is Initiated by Distinct Receptors. Sci Signal (2008) 1(35):re9. doi: 10.1126/ scisignal.135re9
- 44. Ji Z, Zhao W, Lin HK, Zhou X. Systematically Understanding the Immunity Leading to Crpc Progression. *PloS Comput Biol* (2019) 15(9):e1007344. doi: 10.1371/journal.pcbi.1007344
- García García CJ, Acevedo Diaz AC, Kumari N, Govindaraju S, de la Cruz Bonilla M, San Lucas FA, et al. Hif2 Regulates Intestinal Wnt5a Expression. *Front Oncol* (2021) 11:769385. doi: 10.3389/fonc.2021.769385
- Morin E, Lindskog C, Johansson M, Egevad L, Sandström P, Harmenberg U, et al. Perivascular Neuropilin-1 Expression Is an Independent Marker of Improved Survival in Renal Cell Carcinoma. J Pathol (2020) 250(4):387–96. doi: 10.1002/path.5380
- Liu Y, Liu Y, Gao Y, Wang L, Shi H, Xuan C. H19- and Hsa-Mir-338-3p-Mediated Nrp1 Expression Is an Independent Predictor of Poor Prognosis in Glioblastoma. *PloS One* (2021) 16(11):e0260103. doi: 10.1371/journal. pone.0260103
- Zhang X, Li X, Fu X, Yu M, Qin G, Chen G, et al. Circular Rna Taf4b Promotes Bladder Cancer Progression by Sponging Mir-1298-5p and Regulating Tgfa Expression. Front Oncol (2021) 11:643362. doi: 10.3389/fonc.2021.643362
- Badawy AA, El-Hindawi A, Hammam O, Moussa M, Gabal S, Said N. Impact of Epidermal Growth Factor Receptor and Transforming Growth Factor-A on Hepatitis C Virus-Induced Hepatocarcinogenesis. *APMIS Acta Pathol Microbiol Immunol Scand* (2015) 123(10):823–31. doi: 10.1111/apm.12431
- Giricz O, Calvo V, Peterson EA, Abouzeid CM, Kenny PA. Tace-Dependent Tgfα Shedding Drives Triple-Negative Breast Cancer Cell Invasion. Int J Cancer (2013) 133(11):2587–95. doi: 10.1002/ijc.28295
- DeHaan AM, Wolters NM, Keller ET, Ignatoski KM. Egfr Ligand Switch in Late Stage Prostate Cancer Contributes to Changes in Cell Signaling and Bone Remodeling. *Prostate* (2009) 69(5):528–37. doi: 10.1002/pros.20903
- Qin Z, He W, Tang J, Ye Q, Dang W, Lu Y, et al. Micrornas Provide Feedback Regulation of Epithelial-Mesenchymal Transition Induced by Growth Factors. *J Cell Physiol* (2016) 231(1):120–9. doi: 10.1002/jcp.25060
- 53. Jin Y, Peng D, Shen Y, Xu M, Liang Y, Xiao B, et al. Microrna-376c Inhibits Cell Proliferation and Invasion in Osteosarcoma by Targeting to Transforming Growth Factor-Alpha. DNA Cell Biol (2013) 32(6):302–9. doi: 10.1089/dna.2013.1977
- 54. Zhao Y, Wang Y, Zhao E, Tan Y, Geng B, Kang C, et al. Ptrf/Cavin1, Regulated by Shc1 through the Egfr Pathway, Is Found in Urine Exosomes as a Potential Biomarker of Ccrcc. *Carcinogenesis* (2020) 41(3):274–83. doi: 10.1093/carcin/ bgz147
- 55. Ahn R, Sabourin V, Bolt AM, Hébert S, Totten S, De Jay N, et al. The Shc1 Adaptor Simultaneously Balances Stat1 and Stat3 Activity to Promote Breast Cancer Immune Suppression. *Nat Commun* (2017) 8:14638. doi: 10.1038/ ncomms14638
- Grossman SR, Lyle S, Resnick MB, Sabo E, Lis RT, Rosinha E, et al. P66 Shc Tumor Levels Show a Strong Prognostic Correlation with Disease Outcome in Stage IIa Colon Cancer. *Clin Cancer Res Off J Am Assoc Cancer Res* (2007) 13 (19):5798–804. doi: 10.1158/1078-0432.Ccr-07-0073
- Junttila IS. Tuning the Cytokine Responses: An Update on Interleukin (II)-4 and II-13 Receptor Complexes. Front Immunol (2018) 9:888. doi: 10.3389/ fimmu.2018.00888
- 58. Horsophonphong S, Sritanaudomchai H, Nakornchai S, Kitkumthorn N, Surarit R. Odontogenic Gene Expression Profile of Human Dental Pulp-

Derived Cells under High Glucose Influence: A Microarray Analysis. J Appl Oral Sci Rev FOB (2021) 29:e20201074. doi: 10.1590/1678-7757-2020-1074

- Jamieson NB, Maker AV. Gene-Expression Profiling to Predict Responsiveness to Immunotherapy. *Cancer Gene Ther* (2017) 24(3):134–40. doi: 10.1038/cgt.2016.63
- Cristescu R, Nebozhyn M, Zhang C, Albright A, Kobie J, Huang L, et al. Transcriptomic Determinants of Response to Pembrolizumab Monotherapy across Solid Tumor Types. *Clin Cancer Res* (2022) 28(8):1680–9. doi: 10.1158/ 1078-0432.Ccr-21-3329
- 61. Şenbabaoğlu Y, Gejman RS, Winer AG, Liu M, Van Allen EM, de Velasco G, et al. Tumor Immune Microenvironment Characterization in Clear Cell Renal Cell Carcinoma Identifies Prognostic and Immunotherapeutically Relevant Messenger Rna Signatures. *Genome Biol* (2016) 17(1):231. doi: 10.1186/ s13059-016-1092-z
- Braun DA, Street K, Burke KP, Cookmeyer DL, Denize T, Pedersen CB, et al. Progressive Immune Dysfunction with Advancing Disease Stage in Renal Cell Carcinoma. *Cancer Cell* (2021) 39(5):632–48.e8. doi: 10.1016/j.ccell. 2021.02.013
- Krishna C, DiNatale RG, Kuo F, Srivastava RM, Vuong L, Chowell D, et al. Single-Cell Sequencing Links Multiregional Immune Landscapes and Tissue-Resident T cells in Ccrcc to Tumor Topology and Therapy Efficacy. *Cancer Cell* (2021) 39(5):662–77.e6. doi: 10.1016/j.ccell.2021.03.007
- Pan Q, Wang L, Chai S, Zhang H, Li B. The Immune Infiltration in Clear Cell Renal Cell Carcinoma and Their Clinical Implications: A Study Based on Tcga and Geo Databases. J Cancer (2020) 11(11):3207–15. doi: 10.7150/jca.37285
- Dannenmann SR, Thielicke J, Stöckli M, Matter C, von Boehmer L, Cecconi V, et al. Tumor-Associated Macrophages Subvert T-Cell Function and Correlate with Reduced Survival in Clear Cell Renal Cell Carcinoma. *Oncoimmunology* (2013) 2(3):e23562. doi: 10.4161/onci.23562
- 66. Jansen CS, Prokhnevska N, Master VA, Sanda MG, Carlisle JW, Bilen MA, et al. An Intra-Tumoral Niche Maintains and Differentiates Stem-Like Cd8 T Cells. *Nature* (2019) 576(7787):465–70. doi: 10.1038/s41586-019-1836-5
- Murakami T, Tanaka N, Takamatsu K, Hakozaki K, Fukumoto K, Masuda T, et al. Multiplexed Single-Cell Pathology Reveals the Association of Cd8 T-Cell Heterogeneity with Prognostic Outcomes in Renal Cell Carcinoma. *Cancer Immunol Immunother CII* (2021) 70(10):3001–13. doi: 10.1007/s00262-021-03006-2
- 68. Giraldo NA, Becht E, Pagès F, Skliris G, Verkarre V, Vano Y, et al. Orchestration and Prognostic Significance of Immune Checkpoints in the Microenvironment of Primary and Metastatic Renal Cell Cancer. *Clin Cancer Res Off J Am Assoc Cancer Res* (2015) 21(13):3031–40. doi: 10.1158/1078-0432.Ccr-14-2926
- Remark R, Alifano M, Cremer I, Lupo A, Dieu-Nosjean MC, Riquet M, et al. Characteristics and Clinical Impacts of the Immune Environments in Colorectal and Renal Cell Carcinoma Lung Metastases: Influence of Tumor Origin. *Clin Cancer Res Off J Am Assoc Cancer Res* (2013) 19(15):4079–91. doi: 10.1158/1078-0432.Ccr-12-3847
- Hänze J, Wegner M, Noessner E, Hofmann R, Hegele A. Co-Regulation of Immune Checkpoint Pd-L1 with Interferon-Gamma Signaling Is Associated with a Survival Benefit in Renal Cell Cancer. *Target Oncol* (2020) 15(3):377– 90. doi: 10.1007/s11523-020-00728-8
- Havel JJ, Chowell D, Chan TA. The Evolving Landscape of Biomarkers for Checkpoint Inhibitor Immunotherapy. *Nat Rev Cancer* (2019) 19(3):133–50. doi: 10.1038/s41568-019-0116-x
- Rooney MS, Shukla SA, Wu CJ, Getz G, Hacohen N. Molecular and Genetic Properties of Tumors Associated with Local Immune Cytolytic Activity. *Cell* (2015) 160(1-2):48–61. doi: 10.1016/j.cell.2014.12.033
- Matsushita H, Vesely MD, Koboldt DC, Rickert CG, Uppaluri R, Magrini VJ, et al. Cancer Exome Analysis Reveals a T-Cell-Dependent Mechanism of Cancer Immunoediting. *Nature* (2012) 482(7385):400–4. doi: 10.1038/ nature10755
- Fahey CC, Davis IJ. Setting the Stage for Cancer Development: Setd2 and the Consequences of Lost Methylation. *Cold Spring Harb Perspect Med* (2017) 7 (5):a026468. doi: 10.1101/cshperspect.a026468
- Hong CS, Elsamadicy AA, Fisayo A, Inzucchi SE, Gopal PP, Vining EM, et al. Comprehensive Genomic Characterization of a Case of Granular Cell Tumor of the Posterior Pituitary Gland: A Case Report. *Front Endocrinol* (2021) 12:762095. doi: 10.3389/fendo.2021.762095

- Newbold RF, Mokbel K. Evidence for a Tumour Suppressor Function of Setd2 in Human Breast Cancer: A New Hypothesis. *Anticancer Res* (2010) 30 (9):3309–11.
- Masclef L, Ahmed O, Estavoyer B, Larrivée B, Labrecque N, Nijnik A, et al. Roles and Mechanisms of Bap1 Deubiquitinase in Tumor Suppression. *Cell Death Differ* (2021) 28(2):606–25. doi: 10.1038/s41418-020-00709-4
- Ventii KH, Devi NS, Friedrich KL, Chernova TA, Tighiouart M, Van Meir EG, et al. Brca1-Associated Protein-1 Is a Tumor Suppressor That Requires Deubiquitinating Activity and Nuclear Localization. *Cancer Res* (2008) 68 (17):6953–62. doi: 10.1158/0008-5472.Can-08-0365
- 79. Landry AP, Wang JZ, Nassiri F, Patil V, Gao A, Zadeh G. Bap1-Deficient Meningioma Presenting with Trabecular Architecture and Cytokeratin Expression: A Report of Two Cases and Review of the Literature. J Clin Pathol (2021), jclinpath–2021-207952. doi: 10.1136/jclinpath-2021-207952
- Murali R, Wiesner T, Scolyer RA. Tumours Associated with Bap1 Mutations. Pathology (2013) 45(2):116–26. doi: 10.1097/PAT.0b013e32835d0efb
- Abdel-Rahman MH, Pilarski R, Cebulla CM, Massengill JB, Christopher BN, Boru G, et al. Germline Bap1 Mutation Predisposes to Uveal Melanoma, Lung Adenocarcinoma, Meningioma, and Other Cancers. J Med Genet (2011) 48 (12):856–9. doi: 10.1136/jmedgenet-2011-100156
- 82. Hakimi AA, Ostrovnaya I, Reva B, Schultz N, Chen YB, Gonen M, et al. Adverse Outcomes in Clear Cell Renal Cell Carcinoma with Mutations of 3p21 Epigenetic Regulators Bap1 and Setd2: A Report by Mskcc and the Kirc

Tcga Research Network. Clin Cancer Res Off J Am Assoc Cancer Res (2013) 19 (12):3259–67. doi: 10.1158/1078-0432.Ccr-12-3886

 Brugarolas J. Pbrm1 and Bap1 as Novel Targets for Renal Cell Carcinoma. Cancer J (Sudbury Mass) (2013) 19(4):324-32. doi: 10.1097/PPO. 0b013e3182a102d1

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Wang, Tang, Luo, Chen, Zhang, Li, Li, Chen, Xu and Han. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.