

RESEARCH

Open Access



# The sensitivity of different methods for detecting abnormalities in auditory nerve function

Tianhao Lu<sup>1,2</sup>, Qiang Li<sup>1,2</sup>, Chen Zhang<sup>1,2</sup>, Min Chen<sup>1,2</sup>, Zhengming Wang<sup>1,2</sup> and Shufeng Li<sup>1,2\*</sup> 

\*Correspondence:  
lisf@fudan.edu.cn

<sup>1</sup> Department  
of Otolaryngology, Eye  
& ENT Hospital of Fudan  
University, 83 Fenyang Road,  
Shanghai 200031, China  
Full list of author information  
is available at the end of the  
article

## Abstract

**Background:** Cochlear implants (CIs) have become important for the treatment of severe-to-profound sensorineural hearing loss (SNHL). Meanwhile, electrically evoked compound action potentials (ECAPs) and electrically evoked auditory brainstem responses (EABRs), which can be examined and evaluated with minimal patient cooperation, have become more reliable for tone measurement and speech recognition postoperatively. However, few studies have compared the electrophysiological characteristics of the auditory nerve using ECAPs and EABRs under different functional states of the auditory nerve (FSANs). We used guinea pig models in which six electrodes were implanted unilaterally with continuous electrical stimulation (ES) for 4 h. The amplitude growth functions (AGFs) of the alternating polarity ECAP (AP-ECAP) and forward-masking subtraction ECAP (FM-ECAP), as well as the EABR waves under “normal” and “abnormal” FSANs, were obtained.

**Results:** Both the AP-ECAP and FM-ECAP thresholds were significantly higher than those measured by EABR under both “normal” FSAN and “abnormal” FSANs ( $p < 0.05$ ). There was a significant difference in the slope values between electrodes 1 and 2 and electrodes 3 and 4 in terms of the AP-ECAP under the “abnormal” FSAN ( $p < 0.05$ ). The threshold gaps between the AP-ECAP and FM-ECAP were significantly larger under the “abnormal” FSAN than under the “normal” FSAN ( $p < 0.05$ ).

**Conclusions:** Both of the ECAP thresholds were higher than the EABR thresholds. The AP-ECAP was more sensitive than the FM-ECAP under the “abnormal” FSAN.

**Keywords:** Electrically evoked compound action potential, Electrically evoked auditory brainstem response, Electrical stimulation, Different functional states of auditory nerve

## Background

Cochlear implants (CIs) are the most effective aids that can help people with sensorineural hearing loss (SNHL) achieve auditory reconstruction [1–3]. Postoperative examinations and evaluations are also important components of the treatment process. To clinically evaluate the level of hearing recovery after CI, subjective audiometry is often performed by collecting a large amount of data in a short time and then assessing the



© The Author(s) 2020. This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

hearing status of the individuals with the implants daily [4, 5]. However, individuals need CIs at increasingly younger ages; these patients cannot accurately respond to subjective evaluation problems, and it is difficult for doctors to determine their comfort level during the CI adjustment process postoperatively [6]. Therefore, objective evaluations, which can be performed with minimal patient cooperation, are more reliable than subjective evaluations for tone measurement and speech recognition [7].

Electrically evoked compound action potentials (ECAPs) and electrically evoked auditory brainstem responses (EABRs) are both valuable in evaluating the functional states of the auditory nerve (FSAN), as they correspond to the responses of the auditory pathway evoked by intracochlear electrical stimulation [8–10]. ECAPs correspond to compound action potentials of the auditory nerve that are recorded through the electrodes of the cochlear implant [11], while EABRs correspond to the responses of the integrity of auditory pathways that are recorded via electrodes placed on the scalp [12]. Measuring ECAPs is currently one of the most commonly used objective intraoperative evaluation methods. At the end of an operation, the compound action potentials that are produced by auditory nerves and detected by the electrodes of the CIs are recorded to determine whether the individual has a spectrum disorder of auditory neuropathy, and the action potentials can act as a stimulation reference for the initial cochlear activation [13]. There are two common stimulus artefact reduction paradigms that are used to record ECAPs, i.e., alternating polarity ECAPs (AP-ECAPs) and forward-masking subtraction ECAPs (FM-ECAPs) [14, 15]. EABRs can be used to monitor intraoperative auditory function, which can allow us to determine whether a CI has been successfully implanted. Specifically, EABRs are stimulated by the processor and electrodes of the CI to mimic sound processing in the brainstem under ES, and the CI can be determined to be effective or not according to the waveform differentiation results [16].

Few studies have investigated the differences in electrophysiological characteristics between ECAPs and EABRs, and no studies have demonstrated a consistent relationship between these two measures [17, 18]. Several studies have also compared the electrophysiological characteristics of AP-ECAPs and FM-CPAPs [19–22]. AP-ECAPs were found to have smaller amplitudes, higher thresholds and steeper slopes than FM-ECAPs for cochlear devices but not for advanced bionic devices [19, 22]. However, these previous studies were conducted in human cochlear recipients with hearing loss, who exhibit a variety of different pathophysiological mechanisms. Therefore, their FSANs might be different, which may have an impact on the consistency of the results.

Previous research from our laboratory has shown that the continuous stimulation of charge-balanced biphasic pulses to the cochlea can significantly elevate the ECAP threshold in guinea pigs [23]. These findings indicate that acute electrical stimulation (ES) has an inhibitive effect on the excitability of the auditory nerve. We refer to this kind of inhibited excitability as an “abnormal” FSAN relative to the original level of excitability, namely, the “normal” FSAN.

To evaluate the sensitivity of different methods in detecting abnormalities in auditory nerve function, we established guinea pig models with “normal” and “abnormal” FSANs by simply implanting intracochlear electrodes with a specific density and administering continuous ES with a specific duration. Then, we compared the electrophysiological characteristics of the AP-ECAPs, FM-ECAPs and EABRs of the two FSANs.

## Results

The average values of the AP-ECAP, FM-ECAP and EABR thresholds under the “normal” FSAN were 159.2, 155.0 and 116.3 CL, while those under the “abnormal” FSAN were 194.1, 183.1 and 144.7 CL, respectively (Table 1). There was no significant difference between the AP-ECAP and FM-ECAP thresholds at all electrodes under the “normal” and “abnormal” FSANs ( $p > 0.05$ ) (Fig. 1a). However, there was a significant difference between the EABR thresholds and the two kinds of ECAP thresholds ( $p < 0.0001$ ) (Fig. 1b). The mean gap between the AP-ECAPs and EABRs and between the FM-ECAPs and EABRs under the “normal” FSAN was 42.9 and 38.7 CL, respectively, while for those under the “abnormal” FSAN was 49.4 and 38.4 CL, respectively. These results suggested that the EABR thresholds were lower than the ECAP thresholds with the methods used in the present study.

The mean AGF slope values of the AP-ECAP and FM-ECAP were 14.5 and 20.3 under the “normal” FSAN, respectively, and 20.4 and 26.4 under the “abnormal” FSAN, respectively (Table 2). The slope values of the AGFs were not significantly different between the AP-ECAPs and FM-ECAPs under the “normal” FSAN or among the electrodes within groups ( $p > 0.05$ , Fig. 2a). However, there was an exception in the situation mentioned above under the “abnormal” FSAN. There was a significant difference between the AP-ECAP slope values of electrodes 1 and 2 and electrodes 3 and 4 ( $p < 0.01$ , Fig. 2b). These results indicated that the slope of the AP-ECAP may be more sensitive in reflecting the “abnormal” FSAN.

The gaps among the thresholds under the “normal” FSAN and under “abnormal” FSAN were not significantly different for the AP-ECAPs, FM-ECAPs and EABRs ( $p > 0.05$ ), the mean values of which were 45.2, 44.4 and 44.8 CL, respectively (Table 3, Fig. 3a). Similarly, the gaps between the AGF slopes of the AP-ECAP and FM-ECAP were not significantly different ( $p > 0.05$ ), as the mean values were 7.5 and 8.2, respectively (Table 4, Fig. 3b). These results suggested that the threshold gaps in the ECAP and EABR for the “normal” FSAN and “abnormal” FSAN were equally effective in reflecting the severity of the “abnormal” FSAN. The AGF slope gaps of the AP-ECAP and FM-ECAP were equally effective as well.

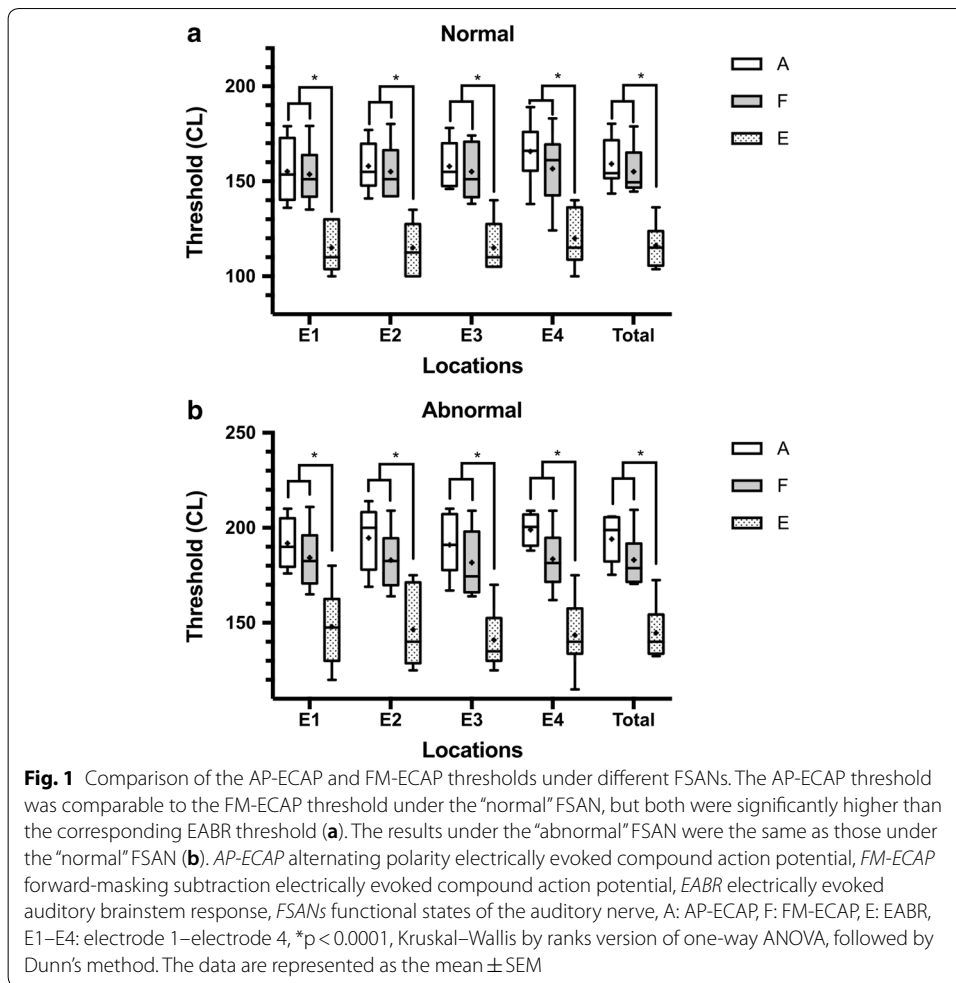
The threshold gaps between the AP-ECAP and FM-ECAP under the “abnormal” FSAN were significantly larger than those under the “normal” FSAN ( $p < 0.05$ ), and the mean values were 11.0 and 4.2 CL for the “abnormal” and “normal” FSANs, respectively (Fig. 4a). The corresponding slope gaps of the AGF were comparable ( $p > 0.05$ ), as the mean value was 5.8 under the “normal” FSAN and 2.2 under the “abnormal” FSAN (Fig. 4b). These results suggested that the “abnormal” FSAN augments the difference between the thresholds of the AP-ECAP and FM-ECAP. In other words, AP-ECAP is likely more sensitive in detecting changes in ECAP thresholds under the “abnormal” FSAN.

## Discussion

To explore the electrophysiological characteristics of ECAPs and EABRs under different FSANs, we compared and analysed the AP-ECAP, FM-ECAP and EABR thresholds as well as the slopes of the AGF for the AP-ECAPs and FM-ECAPs.

**Table 1 ECAP and EABR thresholds under different FSANs**

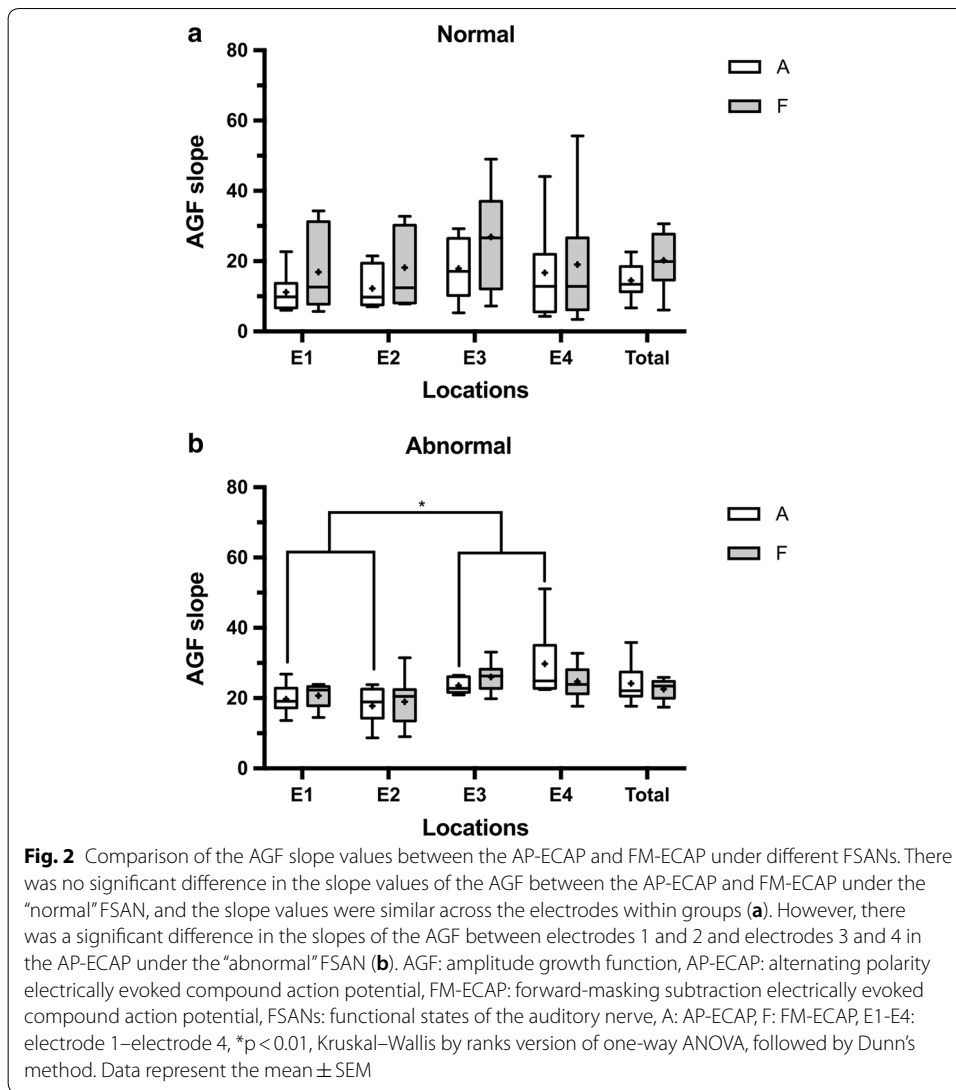
Electrode locations	AP-ECAP			FM-ECAP			EABR						
	Mean (CL)	SD (CL)	N	Mean (CL)	SD (CL)	N	Mean (CL)	SD (CL)	N	SE			
Normal FSAN	E1	155.2	15.8	10	5.0	153.5	14.1	10	4.5	115.0	13.3	10	4.2
	E2	158.0	12.1	10	3.8	154.9	13.9	10	4.4	115.0	14.1	10	4.5
	E3	157.9	12.1	10	3.8	155.0	14.0	10	4.4	115.0	13.3	10	4.2
	E4	165.7	14.9	10	4.7	156.5	18.4	10	5.8	120.0	15.1	10	4.8
	Total	159.2	13.9	40	2.2	155.0	14.7	40	2.3	116.3	13.6	40	2.2
Abnormal FSAN	E1	191.8	13.1	10	4.2	184.3	15.0	10	4.8	148.0	19.5	10	6.2
	E2	194.6	16.0	10	5.1	183.0	15.2	10	4.8	146.5	20.1	10	6.4
	E3	191.0	15.7	10	5.0	181.7	16.6	10	5.3	141.0	14.9	10	4.7
	E4	199.0	7.7	10	2.4	183.5	15.5	10	4.9	143.5	17.5	10	5.5
	Total	194.1	13.4	40	2.1	183.1	15.0	40	2.4	144.7	17.6	40	2.8



**Table 2** AGF slopes

	Electrode locations	AP-ECAP				FM-ECAP			
		Mean	SD	N	SE	Mean	SD	N	SE
Normal FSAN	E1	11.2	5.6	10	1.8	16.9	11.1	10	3.5
	E2	12.2	5.7	10	1.8	18.2	10.7	10	3.4
	E3	17.9	8.8	10	2.8	26.9	14.6	10	4.6
	E4	16.7	14.3	10	4.5	19.1	19.3	10	6.1
	Total	14.5	9.4	40	1.5	20.3	14.3	40	2.3
Abnormal FSAN	E1	19.7	3.8	10	1.2	25.4	15.4	10	4.9
	E2	17.8	5.2	10	1.7	23.2	15.0	10	4.7
	E3	29.5	12.6	10	4.0	29.4	11.2	10	3.5
	E4	29.8	11.2	10	3.5	27.8	10.7	10	3.4
	Total	24.2	10.3	40	1.6	26.4	12.9	40	2.0

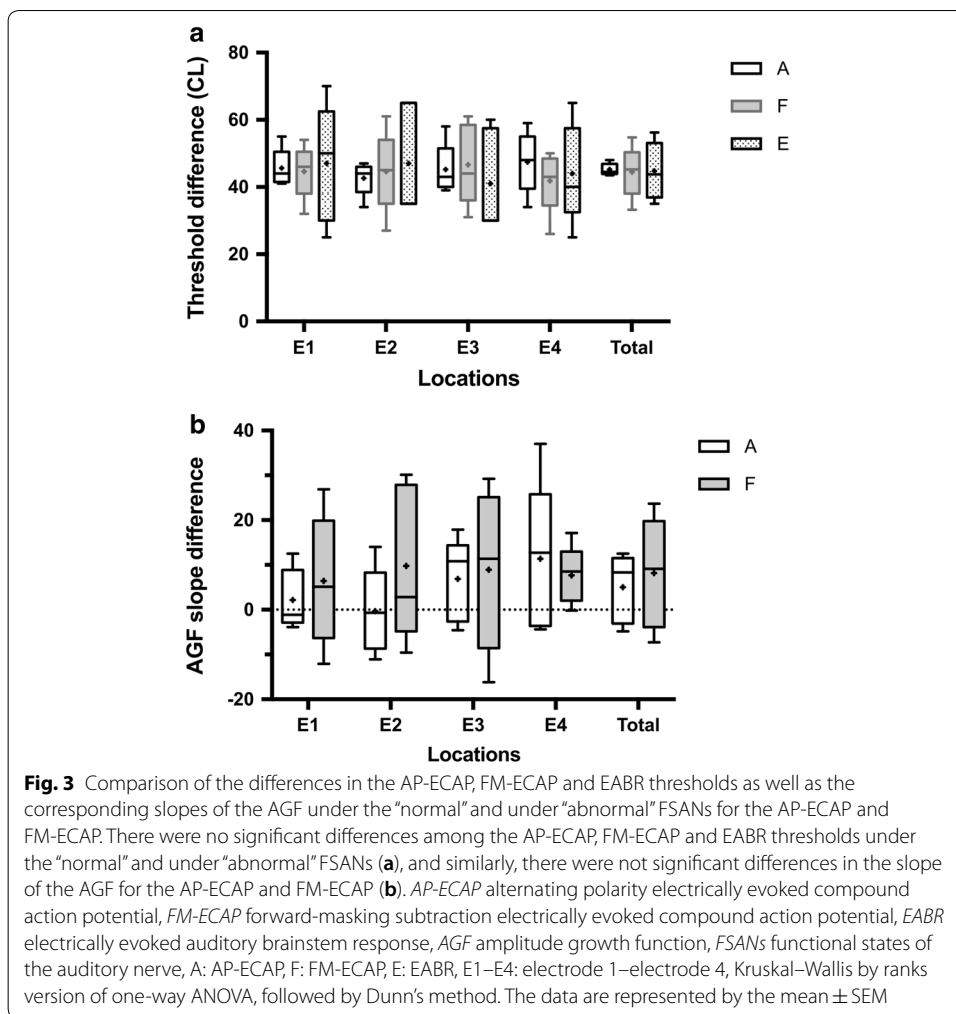
There was no significant difference between the AP-ECAP and FM-ECAP thresholds at any of the electrodes under the two different auditory nerve functions, but both thresholds were significantly higher than the EABR threshold by approximately



**Table 3** Threshold gaps between normal and abnormal FSAN

Electrode locations	AP-ECAP				FM-ECAP				EABR			
	Mean (CL)	SD (CL)	N	SE	Mean (CL)	SD (CL)	N	SE	Mean (CL)	SD (CL)	N	SE
E1	45.6	5.6	5	2.5	44.6	8.0	5	3.6	47.0	17.5	5	7.8
E2	42.6	5.0	5	2.2	44.6	12.1	5	5.4	47.0	16.4	5	7.3
E3	45.2	7.5	5	3.4	46.6	12.0	5	5.4	41.0	15.2	5	6.8
E4	47.4	9.1	5	4.1	41.8	9.3	5	4.2	44.0	14.7	5	6.6
Total	45.2	6.7	20	1.5	44.4	9.8	20	2.2	44.8	14.9	20	3.3

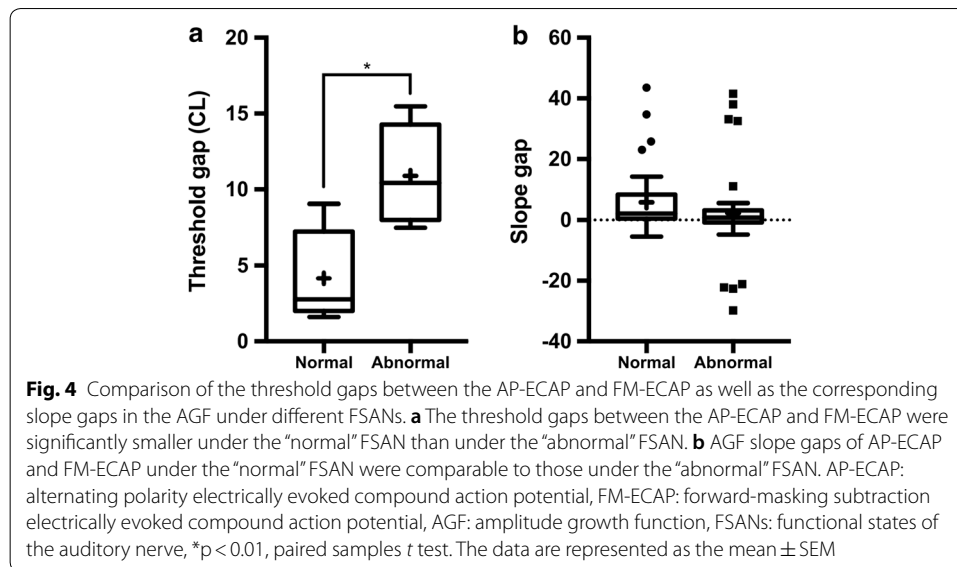
40 CL. Differences in the stimulation parameters, for example, the stimulation frequency, might account for this result. Next, the methods used to determine the threshold were also different: a gap larger than 50  $\mu$ V between the P wave and N wave was considered a positive indicator for the ECAP threshold, while the minimum



**Table 4** AGF slope gap between normal and abnormal FSAN

	Electrode locations	Mean	SD	N	SE
AP-ECAP	E1	3.3	6.3	5	2.8
	E2	2.0	7.5	5	3.3
	E3	11.1	17.1	5	7.6
	E4	13.6	21.2	5	9.5
	Total	7.5	14.2	20	3.2
FM-ECAP	E1	6.4	14.6	5	6.5
	E2	9.8	17.2	5	7.7
	E3	8.9	18.0	5	8.0
	E4	7.7	6.4	5	2.9
	Total	8.2	13.6	20	3.0

intensity of the electrical stimulation that elicited a V wave was considered the EABR threshold. Moreover, P waves, N waves and V waves occurred at different locations. In addition, the difference in the recording method also affected the results. ECAPs



have near-field recording potentials in the vicinity of the auditory nerve with few superpositions, typically 50 superpositions. However, EABRs are far-field potentials recorded on the skin, requiring the superposition of 500–1000 signals [17].

When we compared the slope values of the AGF between electrodes 1 and 2 and electrodes 3 and 4 under the “abnormal” FSAN, we found a significant difference between them. Miller et al. found that the threshold of each nerve fibre to electrical stimulation was normally distributed and that the ECAP slope reflected the number of SGNs reaching the discharge threshold level with an increasing intensity of electrical stimulation [24]. These results indicated that AP-ECAPs and FM-ECAPs have an equal ability to reflect the number of SGNs at the level of the discharge threshold under the “normal” FSAN, but the slope values of the AGF for the AP-ECAP are more sensitive under the “abnormal” FSAN. Moreover, the distance between electrodes and the modiolus cochleae or the number of SGNs and auditory nerve fibres might be critical factors affecting these slopes [25].

There was no significant difference in the AP-ECAP, FM-ECAP and EABR thresholds or in the ECAP slope between the “normal” and “abnormal” FSANs. This result suggested that the structure and function of the auditory nerve can be equally represented by the AP-ECAP, FM-ECP and EABR and that the abilities of these measures to reflect the number of SGNs at the level of the discharge threshold were not affected by the FSAN.

The difference between AP-ECAP and FM-CAP thresholds under the “normal” FSAN was significantly smaller than that under the “abnormal” FSAN. The difference between the AP-ECAP and FM-CAP slopes under the “normal” FSAN was comparable to that under the “abnormal” FSAN. The former result suggested that the AP-ECAP threshold was more sensitive than the FM-ECAP threshold under the “abnormal” FSAN and that it was more helpful to assess the damage to the FSAN. The difference in the algorithms used for processing ECAPs might account for this result: for the FM-ECAP approach, the refractory period in the auditory nerve was utilized to record the auditory nerve



responses sequentially under a combination of four masking pulses and detection pulses. After four kinds of response components were subtracted, artefacts of electrical stimulation could be removed, and then the ECAP waveform elicited from detection pulses could be retained [26]. However, for the AP-ECAP approach, the auditory nerve was stimulated alternately by electrical impulses with opposite polarity. The polarity of the artefact was opposite, and the polarity of the nerve response did not change with the polarity of the stimulation signal. After superposition, most stimulation artefacts were cancelled out, and the ECAP components were enhanced [21]. Moreover, the latencies of the responses to anodic- and cathodic-leading pulses differ; averaging the responses together results in amplitudes that are smaller than that for either polarity alone due to temporal smearing [22, 27, 28]. This process led to a higher threshold for the AP-ECAP. The latter result suggested that the sensitivity of the AP-ECAP slope was equal to that of the FM-ECAP slope under the two kinds of FSANs. However, the results might not be accurate because of the small sample size and the large dispersion shown in the box plot.

## Conclusion

In this study, we established guinea pig models with cochlear implants and then analysed the electrophysiological characteristics of auditory nerves using the ECAPs and EABRs under different FSANs. Our results suggested that the AP-ECAP responded equivalently to the FM-ECAP in determining the threshold under the “normal” FSAN, but they were both significantly higher than those measured by the EABRs; the AP-ECAP was equal to the FM-ECAP in sensitivity under the “normal” FSAN, while the former was more sensitive than the latter in reflecting the FSAN under the “abnormal” FSAN or in different electrode locations.

## Methods

### Establishment of the guinea pig model with a unilateral cochlear implant

The following procedures, which involve Dunkin Hartley guinea pigs, were approved by the Ethics Review Board of the Eye and ENT Hospital at Fudan University. Guinea pigs aged 2–3 postnatal months and weighing 250–350 g were injected intraperitoneally with 0.2 ml of a 0.1% atropine solution, followed by 0.1 ml of a mixed tiletamine/zolazepam and xylazine hydrochloride solution per 100 g of their body weight (the 1.25 ml xylazine hydrochloride solution was added to the 5 ml tiletamine/zolazepam solution). Hairs on the post aurem were shaved after the guinea pigs were under deep anaesthesia, an incision was made, and the muscle and connective tissue on the auditory bulla were separated. The round window in the cochlea was then exposed, and the round window membrane was punctured with a sterile syringe needle. Electrodes (Listent Medical Tech. Co., Ltd.) were completely implanted into the cochlea through the round window unilaterally. The round window and auditory bulla were sealed with the muscles to fix the electrodes. The receiving stimulator and external electrode were implanted subcutaneously before the incision was sutured.

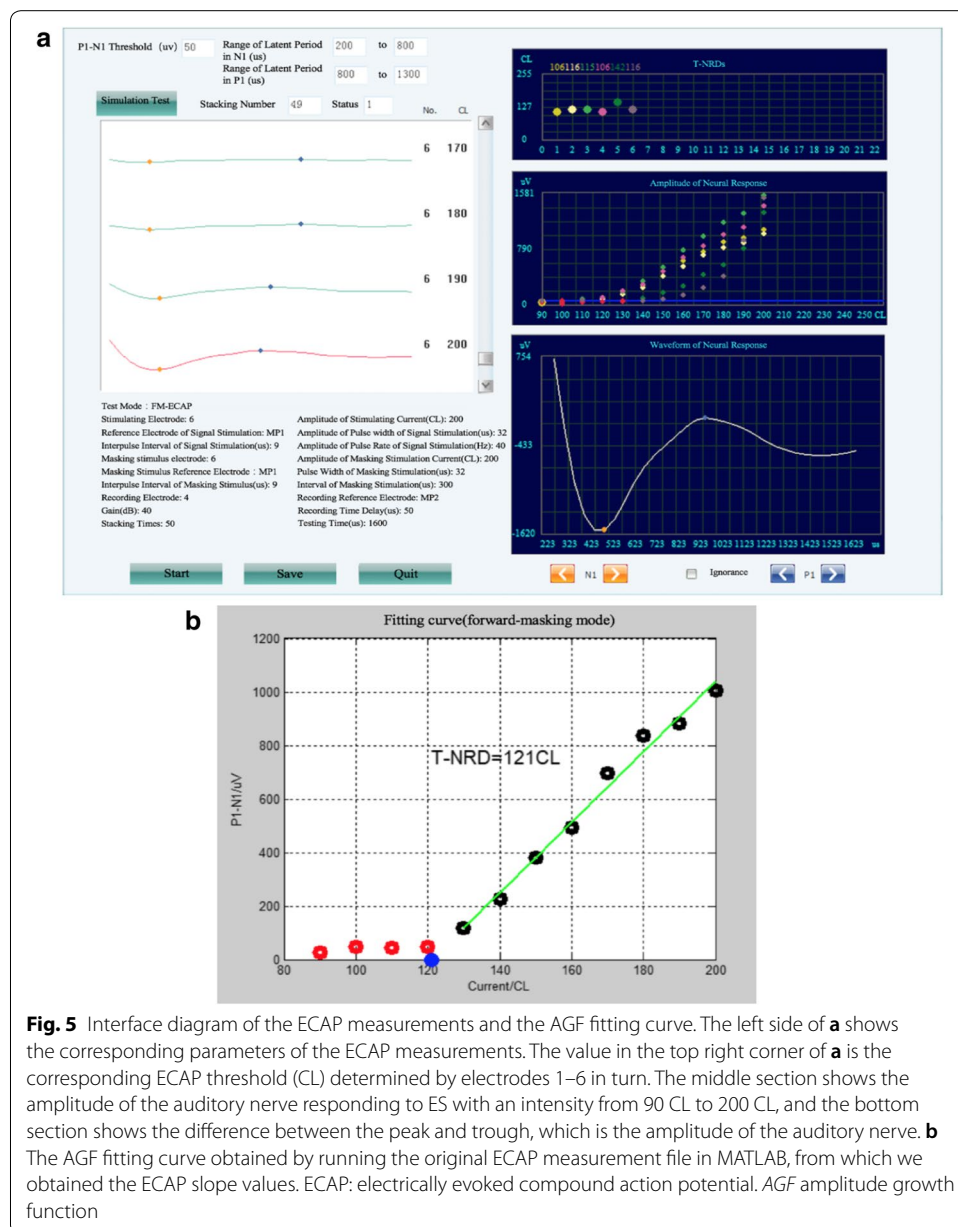
### Measurement of the FM-ECAPs, AP-ECAPs and EABRs

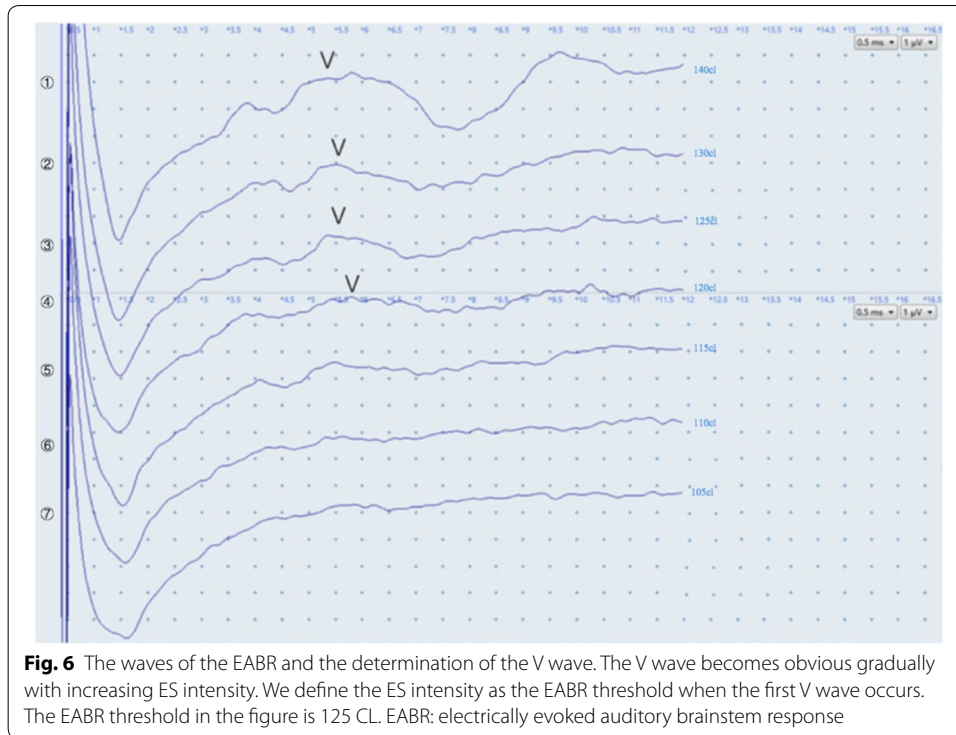
The electrodes were connected to an ECAP measurement instrument, which used MAP V3.00 software (Listent Medical Tech. Co., Ltd.). The impedances of the electrodes

were measured first to ensure proper functioning. The FM-ECAP mode was selected, and the ECAP threshold of each electrode was measured successively (Fig. 5). Then, the AP-ECAP mode was selected, and the ECAP thresholds of electrodes 1–4 were measured successively. Subsequently, the EABR thresholds of electrodes 1–4 were measured sequentially using MAP V3.00 software and Neuro-MEP-Micro equipment (Neurosoft. Co., Ltd., RUS.). We obtained the EABR thresholds by determining V wave (Fig. 6).

### Application of ES to the guinea pig model with the electrodes

The electrodes were connected to the electrical pulse generator, which used V1.00 software (Listent Medical Tech. Co., Ltd.). The intensity of the ES (for electrodes 1–6) was





set to be 6 dB higher than the FM-ECAP threshold. The electrical pulse generator was turned off after 4 h and then the AGFs of the AP-ECAPs and FM-ECAPs as well as the EABR waves were obtained again. We only recorded the data from electrodes 1–4 under the “abnormal” FSAN to reduce deviations in the measurements.

### Statistical analysis

All the original AP-ECAP and FM-ECAP data were processed in MATLAB to obtain the ECAP thresholds and slope values of the AGFs. The statistical analysis was performed by GraphPad Prism 7 (GraphPad Software, Inc.; CA, USA). The variables are reported as the mean  $\pm$  SEM. The significance of the differences was determined by the paired samples *t* test or the Kruskal–Wallis by ranks version of one-way ANOVA, followed by Dunn’s method.

### Abbreviations

CI: Cochlear implant; SNHL: Severe-to-profound sensorineural hearing loss; ECAP: Electrically evoked compound action potential; EABR: Electrically evoked auditory brainstem response; FSAN: Functional states of auditory nerve; ES: Electrical stimulation; AGF: Amplitude growth function; AP-ECAP: Alternating polarity electrically evoked compound action potential; FM-ECAP: Forward-masking subtraction electrically evoked compound action potential; SGNs: Spiral ganglion neurons.

### Acknowledgements

We are grateful to Z. Sun, C. Xu and W. Fan (Shanghai Listent Medical Technology Co., LTD.) for technical support of ECAP measurement and analysis. We further thank Shanghai Listent Medical Technology Co., LTD. for providing customized electrode arrays and pulse generators.

### Authors’ contributions

TL designed and performed experiments, analyzed data, and drafted the manuscript; QL generated the mouse model. CZ and MC analyzed the data. ZW acquired funding and assumed supervision. SL conceived the project, acquired funding, designed experiments, analyzed data, edited the manuscript, and assumed supervision. All authors read and approved the final manuscript.

**Funding**

This work was supported by National Natural Science Foundation of China Grant No. 81670927 and the Science and Technology Commission of Shanghai Municipality No. 19411961400.

**Availability of data and materials**

The data sets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

**Ethics approval and consent to participate**

The procedures using Dunkin Hartley guinea pigs were approved by the Ethics Review Board of Eye and ENT Hospital of Fudan University.

**Consent for publication**

All subjects provided consent for publication.

**Competing interests**

The authors declare that they have no competing interests.

**Author details**

<sup>1</sup> Department of Otolaryngology, Eye & ENT Hospital of Fudan University, 83 Fenyang Road, Shanghai 200031, China.

<sup>2</sup> NHC Key Laboratory of Hearing Medicine, Shanghai, China.

Received: 22 October 2019 Accepted: 22 January 2020

Published online: 03 February 2020

**References**

- Konerding WS, Janssen H, Hubka P, Tornoe J, Mistrik P, Wahlberg L, Lenarz T, Kral A, Scheper V. Encapsulated cell device approach for combined electrical stimulation and neurotrophic treatment of the deaf cochlea. *Hear Res.* 2017;350:110–21.
- Burghard A, Lenarz T, Kral A, Paasche G. Insertion site and sealing technique affect residual hearing and tissue formation after cochlear implantation. *Hear Res.* 2014;312:21–7.
- Roche JP, Hansen MR. On the horizon cochlear implant technology. *Otolaryng Clin N Am.* 2015;48(6):1097.
- Friesen LM, Shannon RV, Baskent D, Wang X. Speech recognition in noise as a function of the number of spectral channels: comparison of acoustic hearing and cochlear implants. *J Acoust Soc Am.* 2001;110(2):1150–63.
- Green T, Faulkner A, Rosen S. Spectral and temporal cues to pitch in noise-excited vocoder simulations of continuous-interleaved-sampling cochlear implants. *J Acoust Soc Am.* 2002;112(5):2155–64.
- de Andrade KCL, Muniz LF, Menezes PD, Neto SDC, Carnauba ATL, Leal MD. The value of electrically evoked stapedius reflex in determining the maximum comfort level of a cochlear implant. *J Am Acad Audiol.* 2018;29(4):292–9.
- Valencia DM, Rimell FL, Friedman BJ, Oblander MR, Helmbrecht J. Cochlear implantation in infants less than 12 months of age. *Int J Pediatr Otorhi.* 2008;72(6):767–73.
- Basta D, Dahme A, Todt I, Ernst A. Relationship between intraoperative eCAP thresholds and postoperative psychoacoustic levels as a prognostic tool in evaluating the rehabilitation of cochlear implantees. *Audiol Neuro-Otol.* 2007;12(2):113–8.
- Cinar BC, Yarali M, Atay G, Bajin MD, Sennaroglu G, Sennaroglu L. The role of eABR with intracochlear test electrode in decision making between cochlear and brainstem implants: preliminary results. *Eur Arch Oto-Rhino-L.* 2017;274(9):3315–26.
- Westen AA, Dekker DMT, Briaire JJ, Frijns JHM. Stimulus level effects on neural excitation and eCAP amplitude. *Hear Res.* 2011;280(1–2):166–76.
- Hughes ML, Stille LJ. Effect of stimulus and recording parameters on spatial spread of excitation and masking patterns obtained with the electrically evoked compound action potential in cochlear implants. *Ear Hearing.* 2010;31(5):679–92.
- Lassaletta L, Polak M, Huesers J, Diaz-Gomez M, Calvino M, Varela-Nieto I, Gavilan J. Usefulness of electrical auditory brainstem responses to assess the functionality of the cochlear nerve using an intracochlear test electrode. *Otol Neurotol.* 2017;38(10):E413–20.
- de Vos JJ, Biesheuvel JD, Briaire JJ, Boot PS, van Gendt MJ, Dekkers OM, Fiocco M, Frijns JHM. Use of electrically evoked compound action potentials for cochlear implant fitting: a systematic review. *Ear Hearing.* 2018;39(3):401–11.
- Good GM, Isaacson G. Otoendoscopy for improved pediatric cholesteatoma removal. *Ann Otol Rhinol Laryngol.* 1999;108(9):893–6.
- Hunter JB, Zuniga MG, Sweeney AD, Bertrand NM, Wanna GB, Haynes DS, Wootten CT, Rivas A. Pediatric endoscopic cholesteatoma surgery. *Otolaryngol Head Neck Surg.* 2016;154(6):1121–7.
- Anwar A, Singleton A, Fang YX, Wang BH, Shapiro W, Roland JT, Waltzman SB. The value of intraoperative EABRs in auditory brainstem implantation. *Int J Pediatr Otorhi.* 2017;101:158–63.
- Brown CJ, Hughes ML, Luk B, Abbas PJ, Wolaver A, Gervais J. The relationship between EAP and EABR thresholds and levels used to program the nucleus 24 speech processor: data from adults. *Ear Hear.* 2000;21(2):151–63.
- Hay-McCutcheon MJ, Brown CJ, Clay KS, Seyle K. Comparison of electrically evoked whole-nerve action potential and electrically evoked auditory brainstem response thresholds in nucleus CI24R cochlear implant recipients. *J Am Acad Audiol.* 2002;13(8):416–27.

19. Baudhuin JL, Hughes ML, Goehring JL. A comparison of alternating polarity and forward masking artifact-reduction methods to resolve the electrically evoked compound action potential. *Ear Hearing*. 2016;37(4):E247–55.
20. Eisen MD, Franck KH. Electrically evoked compound action potential amplitude growth functions and HiResolution programming levels in pediatric CII implant subjects. *Ear Hear*. 2004;25(6):528–38.
21. Frijns JHM, Briare JJ, de Laat JAPM, Grote JJ. Initial evaluation of the Clarion CII cochlear implant: speech perception and neural response imaging. *Ear Hearing*. 2002;23(3):184–97.
22. Hughes ML, Goehring JL, Baudhuin JL. Effects of stimulus polarity and artifact reduction method on the electrically evoked compound action potential. *Ear Hear*. 2017;38(3):332–43.
23. Li Q, Lu T, Zhang C, Hansen MR, Li S. Electrical stimulation induces synaptic changes in the peripheral auditory system. *J Comp Neurol*. 2019. <https://doi.org/10.1002/cne.24802>.
24. Miller CA, Robinson BK, Rubinstein JT, Abbas PJ, Runge-Samuels CL. Auditory nerve responses to monophasic and biphasic electric stimuli. *Hear Res*. 2001;151(1–2):79–94.
25. Prado-Guitierrez P, Fewster LM, Heasman JM, McKay CM, Shepherd RK. Effect of interphase gap and pulse duration on electrically evoked potentials is correlated with auditory nerve survival. *Hear Res*. 2006;215(1–2):47–55.
26. Abbas PJ, Brown CJ, Shallop JK, Firszt JB, Hughes ML, Hong SH, Staller SJ. Summary of results using the nucleus CI24M implant to record the electrically evoked compound action potential. *Ear Hear*. 1999;20(1):45–59.
27. Macherey O, Carlyon RP, van Wieringen A, Deeks JM, Wouters J. Higher sensitivity of human auditory nerve fibers to positive electrical currents. *JARO-J Assoc Res Oto*. 2008;9(2):241–51.
28. Undurraga JA, van Wieringen A, Carlyon RP, Macherey O, Wouters J. Polarity effects on neural responses of the electrically stimulated auditory nerve at different cochlear sites. *Hear Res*. 2010;269(1–2):146–61.

### Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Ready to submit your research? Choose BMC and benefit from:**

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

**At BMC, research is always in progress.**

Learn more [biomedcentral.com/submissions](https://biomedcentral.com/submissions)

