## **Tandem Inhibition of PKC in Diαβetic Nephropathy** It Takes Two to Tango?

Vicki Thallas-Bonke<sup>1,2</sup> and Mark E. Cooper<sup>1,2,3</sup>

t is now more than 15 years since the initial reports of upregulation of protein kinase C (PKC) isoforms in the diabetic kidney and positive preclinical data with the PKC isoform inhibitor, ruboxistaurin. However, subsequent clinical studies with ruboxistaurin that failed to reach key primary end points and the failure of the PKC- $\alpha$  or PKC- $\beta$  isoform knockout mice in the presence of diabetes to totally prevent the diabetes-associated renal lesions have reduced the enthusiasm for targeting PKC isoforms in diabetic nephropathy. However, as reported in this issue of Diabetes, Menne et al. (1) have provided positive provocative data suggesting that dual inhibition of PKC- $\alpha$  and - $\beta$ isoforms is a novel approach that warrants consideration in diabetic nephropathy. The work is strengthened by using not only a genetic approach, i.e., a double PKC isoform knockout mouse, but by also using a PKC- $\alpha/\beta$  inhibitor, albeit its specificity remains questionable.

PKC has been implicated in the pathogenesis of diabetic nephropathy. PKC is a family of closely related enzymes that phosphorylate serine or threenine residues of various intracellular proteins and thus is involved in a wide range of cellular functions that may be relevant to the pathophysiology of diabetes complications including basement membrane production and signal transduction for hormones and growth factors (2). At least 11 isoforms have been identified and classified into three groups. The first group is the conventional PKCs, including the isoforms studied by Menne et al. (1) such as  $\alpha$ ,  $\beta I$ ,  $\beta II$ , as well as  $\gamma$ . The second group is composed of  $\delta$ ,  $\varepsilon$ ,  $\eta$ ,  $\theta$ , and  $\mu$ , and these are the novel PKCs. There is a third group or atypical PKCs, which include  $\zeta$ ,  $\iota$ , and  $\lambda$  (3). Although hyperglycemia is one of the major players in PKC activation, a number of other stimuli have been demonstrated to play a role, including advanced glycation end products (AGEs) (4), AGE receptors (5), angiotensin II (6), and reactive oxygen species (7,8) (Fig. 1).

Debate has been raging over many years to determine which PKC isoform(s) are primarily upregulated and directly implicated in diabetes complications. One of the earliest in vivo studies showed an increase in PKC activity in renal glomeruli from streptozotocin (STZ) diabetic rats within 1 week after the induction of diabetes (9). The first

Corresponding author: Mark E. Cooper, mark.cooper@bakeridi.edu.au. DOI: 10.2337/db12-1666

See accompanying original article, p. 1167.

inhibitor developed for potential clinical use was against the PKC-B isoform, LY333531, now known as ruboxistaurin. In 1996 it was demonstrated that this competitive reversible inhibitor to PKC- $\beta$ I and - $\beta$ II was able to improve albuminuria, glomerular filtration rate, and retinal circulation in diabetic rats when administered orally for 2 weeks (10). In a longer study in the db/db mouse model, treatment with LY333531 ameliorated albuminuria and mesangial expansion possibly by attenuating the expression of transforming growth factor (TGF)-β leading to reduced fibronectin and collagen IV (11). Subsequently, a study in diabetic transgenic Ren-2 rats' inhibition of PKC-β with ruboxistaurin resulted in amelioration of albuminuria, structural injury, and TGF-β expression despite continued hyperglycemia and hypertension (12). Clinical studies with ruboxistaurin in patients with diabetic nephropathy have been disappointing with at best a modest effect on albuminuria (13,14).

Though there have been a number of supporters of the prime importance of the PKC- $\beta$  isoform, other researchers have implicated PKC- $\alpha$  as a major culprit for diabetes complications. PKC- $\alpha$  was shown to be increased in glomerular mesangial cells cultured under high glucose conditions (15). This was followed by a study by Kang et al. (2) who demonstrated an increase in PKC- $\alpha$  and - $\varepsilon$  in the kidney after 4 weeks of STZ diabetes but no increase in the PKC- $\beta$  isoforms. Furthermore, a link between PKC- $\alpha$  and activation of reactive oxygen species was identified (16).

Generation of PKC isoform-specific knockout mice (17) initiated the next phase of the investigation into PKC isoform specificity in diabetes. It was demonstrated in PKC- $\alpha$ knockout mice that glucose-induced albuminuria is mediated at least in part by PKC- $\alpha$  signaling via downregulation of proteoglycans and vascular endothelial growth factor (VEGF) (18). Our own group demonstrated renal activation of PKC- $\alpha$  in a STZ diabetic rat model, which was attenuated by an AGE inhibitor and correlated with renal VEGF expression (4). Further studies by Menne et al. (19) demonstrated no loss of nephrin with the deletion of PKC- $\alpha$  in diabetic mice, further linking PKC- $\alpha$  to previously identified mechanisms of diabetes-associated albuminuria. To further elucidate the differences in deleting various PKC isoforms, a study by Meier et al. (20) in 8-week-old diabetic and control PKC-β knockout mice showed that this isoform is linked to the regulation of renal hypertrophy with no effect on albuminuria or nephrin loss. These findings are consistent with an article by King and colleagues (21) in which a longer duration study in diabetic PKC- $\beta$  knockout mice for up to 24 weeks revealed reduced glomerular and renal hypertrophy, although there was a modest effect on albuminuria.

These studies on PKC- $\alpha$  and PKC- $\beta$  knockout mouse models led the investigators to hypothesize that PKC- $\alpha$ – dependent signaling is involved in perlecan and nephrin expression leading to albuminuria in diabetes, whereas the

From the <sup>1</sup>Diabetes Complications Division, Baker IDI Heart & Diabetes Institute, Melbourne, Victoria, Australia; the <sup>2</sup>Department of Medicine, Australia; and Northern Clinical Schools, University of Melbourne, Victoria, Australia; and the <sup>3</sup>Department of Medicine and Immunology, Alfred Medical Research & Education Precinct, Monash University, Melbourne, Australia.

<sup>© 2013</sup> by the American Diabetes Association. Readers may use this article as long as the work is properly cited, the use is educational and not for profit, and the work is not altered. See http://creativecommons.org/licenses/by -nc-nd/3.0/ for details.

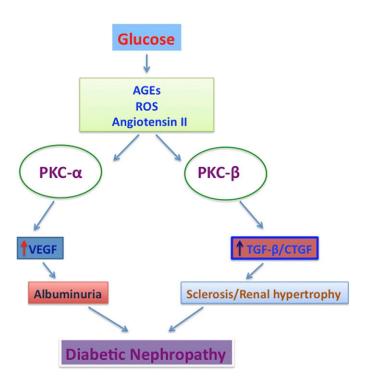


FIG. 1. In diabetic nephropathy, hyperglycemia initiates various intracellular signaling pathways resulting in further downstream activation of different PKC isoforms. ROS, reactive oxygen species; CTGF, connective tissue growth factor.

PKC-β isoform is involved in TGF-β-mediated renal hypertrophy (22). This postulate led to experiments involving the generation of double knockout mice (PKC- $\alpha/\beta^{-/-}$ ) in which glomerular hypertrophy, TGF- $\beta$ , and extracellular matrix were diminished when compared with wild-type mice after 8 weeks of diabetes (1). Interestingly-and as yet not fully explained-albuminuria was not completely prevented with the double  $\alpha/\beta$  PKC isoform knockout mice when compared with the single diabetic PKC- $\alpha$  knockout mice. To strengthen these findings in the knockout mice, treatment with the PKC- $\alpha/\beta$  inhibitor CGP41252 in the STZ 129/SV and the *db/db* mice was associated with improvement in albuminuria but only a mild reduction in renal hypertrophy. These findings were interpreted as occurring as a result of CGP41252 being a pan-PKC inhibitor rather than specific for PKC- $\alpha/\beta$  (1). Such an agent may inhibit novel PKC isoforms such as PKC-*\varepsilon*, whose upregulation has been shown to be protective in diabetic nephropathy (23).

As diabetic nephropathy continues to be a major complication of type 1 and type 2 diabetes and represents the major cause of end-stage renal disease in the Western world, new therapies are urgently needed. Improved control of hyperglycemia and blood pressure are critical for preserving renal function in diabetes, and interruption of the renin-angiotensin system remains the gold standard for the treatment of diabetic nephropathy. However, this new study highlights the need for further development of isoform-specific PKC inhibitors with dual PKC- $\alpha/\beta$  action, which could potentially be used in diabetic nephropathy.

## ACKNOWLEDGMENTS

V.T.-B. is a recipient of a JDRF Advanced Postdoctoral Fellowship. M.E.C. is a National Health and Medical

Research Council Australian Fellow and the recipient of a JDRF Scholar Award.

No potential conflicts of interest relevant to this article were reported.

## REFERENCES

- 1. Menne J, Shushakova N, Bartels J, et al. Dual inhibition of classical protein kinase C- $\alpha$  and protein kinase C- $\beta$  isoforms protects against experimental murine diabetic nephropathy. Diabetes 2013;62:1167–1174
- Kang N, Alexander G, Park JK, et al. Differential expression of protein kinase C isoforms in streptozotocin-induced diabetic rats. Kidney Int 1999; 56:1737–1750
- Koya D, King GL. Protein kinase C activation and the development of diabetic complications. Diabetes 1998;47:859–866
- 4. Thallas-Bonke V, Lindschau C, Rizkalla B, et al. Attenuation of extracellular matrix accumulation in diabetic nephropathy by the advanced glycation end product cross-link breaker ALT-711 via a protein kinase C-alpha-dependent pathway. Diabetes 2004;53:2921–2930
- Li YM, Mitsuhashi T, Wojciechowicz D, et al. Molecular identity and cellular distribution of advanced glycation endproduct receptors: relationship of p60 to OST-48 and p90 to 80K-H membrane proteins. Proc Natl Acad Sci USA 1996;93:11047–11052
- Osicka TM, Yu Y, Panagiotopoulos S, et al. Prevention of albuminuria by aminoguanidine or ramipril in streptozotocin-induced diabetic rats is associated with the normalization of glomerular protein kinase C. Diabetes 2000;49:87–93
- Noh H, King GL. The role of protein kinase C activation in diabetic nephropathy. Kidney Int Suppl. 2007;(Suppl.):S49–S53
- Thallas-Bonke V, Thorpe SR, Coughlan MT, et al. Inhibition of NADPH oxidase prevents advanced glycation end product-mediated damage in diabetic nephropathy through a protein kinase C-alpha-dependent pathway. Diabetes 2008;57:460–469
- Craven PA, DeRubertis FR. Protein kinase C is activated in glomeruli from streptozotocin diabetic rats. Possible mediation by glucose. J Clin Invest 1989;83:1667–1675
- Ishii H, Jirousek MR, Koya D, et al. Amelioration of vascular dysfunctions in diabetic rats by an oral PKC beta inhibitor. Science 1996;272:728–731
- Koya D, Haneda M, Nakagawa H, et al. Amelioration of accelerated diabetic mesangial expansion by treatment with a PKC beta inhibitor in diabetic db/ db mice, a rodent model for type 2 diabetes. FASEB J 2000;14:439–447
- 12. Kelly DJ, Zhang Y, Hepper C, et al. Protein kinase C beta inhibition attenuates the progression of experimental diabetic nephropathy in the presence of continued hypertension. Diabetes 2003;52:512–518
- 13. Bakris GL. Protein kinase C-beta inhibition: a promise not yet fulfilled. Clin J Am Soc Nephrol 2007;2:619–620
- 14. Tuttle KR, McGill JB, Haney DJ, Lin TE, Anderson PW, PKC-DRS, PKC-DMES, and PKC-DRS 2 Study Groups. Kidney outcomes in long-term studies of ruboxistaurin for diabetic eye disease. Clin J Am Soc Nephrol 2007;2:631–636
- 15. Kikkawa R, Haneda M, Uzu T, Koya D, Sugimoto T, Shigeta Y. Translocation of protein kinase C alpha and zeta in rat glomerular mesangial cells cultured under high glucose conditions. Diabetologia 1994;37:838–841
- 16. Li PF, Maasch C, Haller H, Dietz R, von Harsdorf R. Requirement for protein kinase C in reactive oxygen species-induced apoptosis of vascular smooth muscle cells. Circulation 1999;100:967–973
- Leitges M, Schmedt C, Guinamard R, et al. Immunodeficiency in protein kinase cbeta-deficient mice. Science 1996;273:788–791
- Menne J, Park JK, Boehne M, et al. Diminished loss of proteoglycans and lack of albuminuria in protein kinase C-alpha-deficient diabetic mice. Diabetes 2004;53:2101–2109
- Menne J, Meier M, Park JK, et al. Nephrin loss in experimental diabetic nephropathy is prevented by deletion of protein kinase C alpha signaling in-vivo. Kidney Int 2006;70:1456–1462
- Meier M, Park JK, Overheu D, et al. Deletion of protein kinase C-beta isoform in vivo reduces renal hypertrophy but not albuminuria in the streptozotocin-induced diabetic mouse model. Diabetes 2007;56:346–345
- Ohshiro Y, Ma RC, Yasuda Y, et al. Reduction of diabetes-induced oxidative stress, fibrotic cytokine expression, and renal dysfunction in protein kinase Cbeta-null mice. Diabetes 2006;55:3112–3120
- Meier M, Menne J, Park JK, Haller H. Nailing down PKC isoform specificity in diabetic nephropathy two's company, three's a crowd. Nephrol Dial Transplant 2007;22:2421–2425
- Meier M, Menne J, Park JK, et al. Deletion of protein kinase C-epsilon signaling pathway induces glomerulosclerosis and tubulointerstitial fibrosis in vivo. J Am Soc Nephrol 2007;18:1190–1198