

Good Clinical Response to Erlotinib in a Non-Small Cell Lung Cancer Patient Harboring Multiple Brain Metastases and a Double Active Somatic Epidermal Growth Factor Gene Mutation

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Key Words

Epidermal growth factor receptor inhibitor · Erlotinib · Squamous cell lung cancer · Pharmacokinetics · Pleural effusion

Abstract

Recently, 2 small molecule kinase inhibitors (TKIs), targeting epidermal growth factor receptor (EGFR), have proven effective in the treatment of non-small cell lung cancer. However, it is unknown whether the EGFR double activating mutation of L858R in exon 21 and the in-frame deletion in exon 19 is a predictor of the effectiveness of EGFR-TKIs. We report for the first time a case of non-small cell lung cancer with central nervous system metastases harboring a rare EGFR double activating mutation who showed a good clinical response to erlotinib, regardless of his poor performance status, as swallowing is not possible. Therefore, we suggest that erlotinib may become a therapeutic choice in cases of central nervous system metastases even with poor performance status.

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Introduction

Lung cancer is a major cause of cancer-related mortality worldwide and is expected to remain a major health problem for the foreseeable future. Chemotherapy is the cornerstone of the management of the disease; however, its therapeutic impact on patient survival has been modest. Recent discoveries have provided greater understanding of the molecular basis of the disease, and the success of 2 small molecule kinase inhibitors (TKIs), gefitinib (Iressa®, AstraZeneca, London, UK) and erlotinib (Tarceva®, OSI Pharmaceuticals, Melville City, N.Y., USA), against epidermal growth factor receptor (EGFR) in the treatment of non-small cell lung cancer (NSCLC) has provided evidence for the effectiveness of the strategy. Erlotinib and gefitinib inhibit the tyrosine kinase activity of EGFR and have been studied extensively. Since Lynch et al. have identified specific activating mutations within the tyrosine kinase domain of EGFR [1], the missense mutation L858R in exon 21 and the in-frame deletion in exon 19, nested around the amino acid residues 747 to 750 of the EGFR polypeptide, account for >85% of all clinically important mutations related to TKI sensitivity [2]. The detection of EGFR mutations in tumor tissues has been applied for predicting the response of TKI treatment and hence guiding the treatment for advanced NSCLC. A panel of 30 EGFR kinase domain mutations that were recently reported in NSCLC patients was cloned and expressed for analysis of kinase activity, transforming potential, and drug sensitivity. These mutations affect the N-lobe (exons 18–20) and the C-lobe (exon 21) of the EGFR kinase domain [3], however it is uncertain whether the EGFR double activating mutation is a predictor of the effectiveness of EGFR-TKIs.

Patients with central nervous system (CNS) metastases in general suffer from deterioration of performance status (PS) and do not have a long survival time. The primary treatment for CNS metastases in patients with NSCLC has traditionally consisted of whole-brain radiotherapy, surgery, or radiosurgery, while systemic chemotherapy has been thought to play a limited role because of the belief that the brain is a pharmacologic sanctuary site [4, 5]. Several articles in which CNS metastases were improved by erlotinib have been reported [6–11]; however, it remains unknown whether CNS metastases are improved by erlotinib in a NSCLC case harboring EGFR double activating mutation with a poor PS.

We report, for the first time, a case of NSCLC with CNS metastases harboring an EGFR double activating mutation that showed a good clinical response to erlotinib even with a poor PS.

Case Report

The patient was a 60-year-old Japanese man with no smoking history and PS 1 ([table 1](#)). He suffered from prolonged left chest pain on our 1st visit. Evaluation at another facility revealed a nodule in the left lung measuring 3.5 cm in diameter, multiple mediastinal contralateral lymph node metastases, multiple brain metastases, and left pleural effusion and disseminations. Pathologic examination of specimens obtained by transbronchial biopsy from the left lung mass revealed adenocarcinoma ([fig. 1](#)) with a disease stage of cT4N3M1. To choose a therapeutic strategy, we investigated the EGFR gene mutation status of the transbronchial biopsy specimen with the PNA-LNA clamp method [12–14] and detected an EGFR gene double activating mutation; L747-S752 deletion in exon 19 and L858R in exon 21.

Cancer cells were obtained from paraffin-embedded biopsy specimens by manual microdissection. The definition of a small specimen is that the quantity of the specimen is sufficient to make a pathologic diagnosis; at most several hundred tumor cells are necessary. Formalin-fixed paraffin-embedded tissue was cut in 6–8 µm sections, and mounted on pre-treated glass slides. Noncancer cells and necrotic parts were manually removed from the slide under the microscope. The slides were deparaffinized, then DNA

was extracted with phenol-chloroform and ethanol precipitation. A PNA-LNA PCR clamp, which was designed to detect 11 different EGFR mutations, was used for determination of the EGFR gene mutation status [12–14]. In this study, we adopted the PNA-LNA PCR clamp for analysis of the EGFR somatic mutations. In the PNA-LNA PCR clamp, other types of EGFR mutations are used to control for amplification by the clamp primer. In this case, a direct sequencing method was employed to seek other types of EGFR mutations.

On the basis of the results of molecular analysis, the patient received carboplatinate (area under the concentration-time curve 5) and paclitaxel (180 mg/m^2) intravenously on day 1 of a 3-week cycle. Both the pulmonary lesion and the left pleural effusion and disseminations showed stable disease, as assessed by response evaluation criteria in solid tumors [15]. After he was on medication for 3 courses with stable disease, he developed seizures and multiple brain lesions (fig. 2a, b, c). He then received the oral EGFR-TKI erlotinib as the 2nd-line treatment. Upon administration of erlotinib, he could not swallow because of disturbance of consciousness so we administered erlotinib dissolved in water from a nasal tube.

He was also administered hydantoin as an anticonvulsant which induced cytochrome CYP3A4 and might promote metabolism of erlotinib. Therefore, we performed pharmacokinetic analyses of erlotinib and OSI-420 on days 1 and 8 after starting its administration. Blood samples were obtained at 5 points a day: just before administration (C0), 2 h (C2), 4 h (C4), 8 h (C8), and 24 h (C24) after administration. The concentrations were determined by high-performance liquid chromatography with ultraviolet detection, as previously reported [16]. Before analysis, we got written informed consent from the patient. Surprisingly, although the serum concentration of erlotinib was quite low (C_{\max} 925 ng/ml on day 8, fig. 3 and table 2), the signs of brain metastases improved dramatically (fig. 2) and his PS recovered from 4 to 1. He has remained on erlotinib and has been without evidence of disease progression for about 2 months.

Discussion

In this case, the C_{\max} of erlotinib on day 8 was 956 ng/ml, which was remarkably lower than the plasma concentration previously reported [17]. The low plasma concentration may be due to CYP3A4 induced by hydantoin. In spite of the low plasma concentration, the patient showed a good clinical response of his multiple brain metastases and his PS recovered dramatically from 4 to 1.

Erlotinib inhibits the activity of the EGFR tyrosine kinase in intact tumor cells, with 50% inhibitory concentration values of 20 nmol/l (7.9 ng/ml) [18]. In a study of athymic mice with human carcinomas, the plasma concentration of erlotinib associated with a 50% inhibition of EGFR tyrosine kinase was estimated at 8–12 $\mu\text{mol/l}$ (3.1–4.7 $\mu\text{g/ml}$) [19]. According to these reports, although the plasma concentration was relatively low compared to previously reported results [17], 956 ng/ml on day 8 is above the IC50. Therefore, this may be the 1st reason for the patient's good clinical response.

Erlotinib is metabolized in the liver, mainly by the cytochrome CYP3A4 (approximately 80%). Erlotinib and its metabolites are excreted predominantly via the feces [20]. Although the administration of erlotinib dissolved in water is expected to yield a good absorption, the plasma concentration of erlotinib was quite low. In our opinion, the reason may be cytochrome CYP3A4. Thus in administration of erlotinib, coadministration with medicine that might induce cytochrome CYP3A4 may affect the plasma concentration of erlotinib more than we expected. Such a coadministration may contribute to a poor clinical response to erlotinib.

In terms of EGFR-TKI sensitivity, Kancha et al. have identified 4 sets of mutations based on their drug sensitivity profiles: (a) mutations that are very sensitive to all 3 drugs tested with IC50 values in the low nanomolar range (L858R and Del 747–753insS), (b) mutations that are less sensitive to gefitinib (IC50 >100 nmol/l) but sensitive to both erlotinib and AEE788 (G719S, V742A, and R776C; IC50 < 100 nmol/l), (c) mutations that

are less sensitive to both gefitinib and erlotinib but sensitive to AEE788 (D761N, S768I, S784F, L838V, and L861Q), and (d) mutations that are resistant to all three drugs tested (N826S and T790M) [3]. To our knowledge, reports concerning cases with an EGFR double activating mutation are few. Sriuranpong et al. reported one case with an EGFR double activating mutation in Thailand [21]. Zhang et al. described 5 cases with EGFR double activating mutation in Chinese patients [22]. In the Zhang report, exon 19 deletion + L858R double mutations were located on the same allele and the double mutant had a higher sensitivity to TKIs than the other two single activating mutation in the *in vitro* study. Although it remains unknown whether the double mutant shows good clinical response to erlotinib, this may be the 2nd reason of the patient's good clinical response. To our knowledge, this is the first case with a good clinical response to erlotinib harboring this EGFR double activating mutation.

In addition, several articles in which CNS metastases were improved by erlotinib have been reported [6–11] and Katayama et al. reported that erlotinib was a reasonable option for the treatment of CNS diseases that appeared after a good initial response of extra CNS lesions to gefitinib [9]. However, very little is known about the CNS penetration of and exposure to this drug [23–25]. In our case, CNS metastases were improved by erlotinib regardless of low plasma concentration, poor PS, and disturbance of consciousness. Therefore, in cases with CNS metastases, even if they suffer from deterioration of PS, erlotinib may become a therapeutic choice when the cytotoxic agents are not appropriate.

Table 1. Patient's background

EGFR mutation		Age	Gender	Histology	Smoking status	PS	Stage	Metastatic lesion
exon 19	exon 21							
L747-S752del	2573T>G:L858R	60	male	adenocarcinoma	never smoker	1	cTN3M1/IV	brain

Table 2. The pharmacokinetics parameters and serum concentrations of erlotinib and OSI-420

	Erlotinib				OSI-420			
	AUC ₀₋₂₄ ng × h/ml	C _{max} ng/ml	t _{max} h	t _{1/2} h	AUC ₀₋₂₄ ng × h/ml	C _{max} ng/ml	t _{max} h	t _{1/2} h
Day 1	14,178	824	8	15.7	1,029	59	8	20.1
Day 8	16,936	925	2	64.6	1,895	101	4	69.4

C_{max} = Maximal plasma concentration; t_{max} = time to C_{max}; t_{1/2} = half period.

Fig. 1. High-power magnification of a tumor specimen of the left lung nodule shows adenocarcinoma.

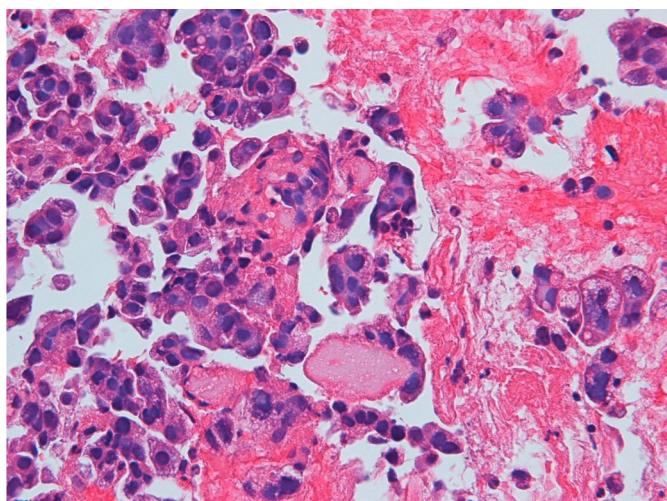


Fig. 2. Brain MRI of the current case. **a** On our 1st visit. **b** The signs of CNS metastases were revealed before administration of erlotinib. **c** 4 weeks after initiation of erlotinib, the signs had improved.

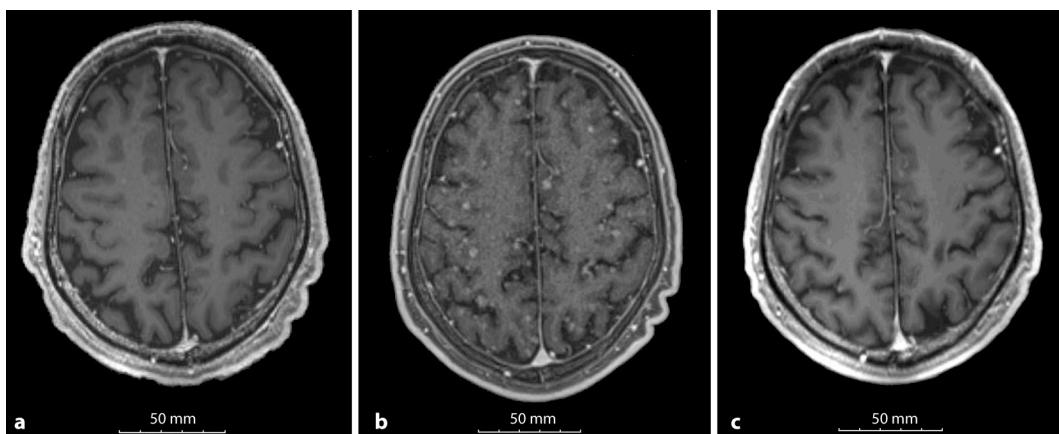
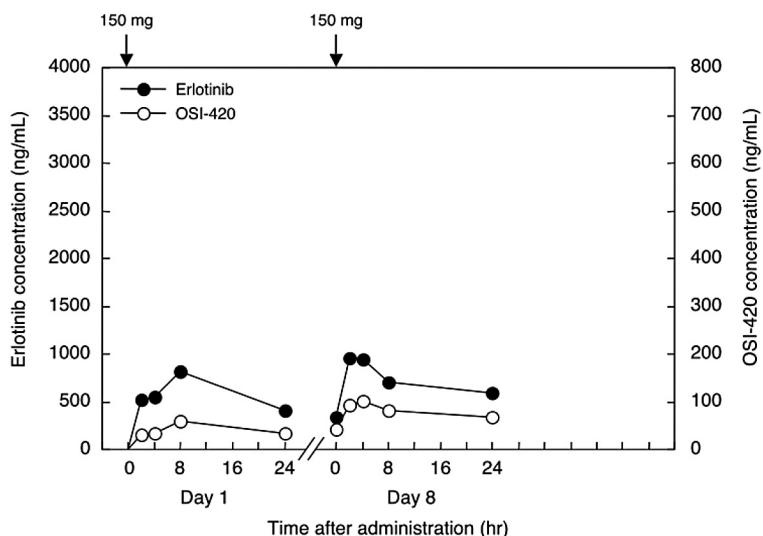


Fig. 3. Plasma concentration-time plots of erlotinib and OSI-420 in case 1 on days 1 and 8. Arrows indicate administration of 150 mg erlotinib.



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