

# Pharyngotonsillitis

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Acute pharyngotonsillitis is a common illness that often leads patients to consult general practitioners, pediatricians, internists, ear, nose and throat physicians, and other types of primary-care doctors. The disease results in a high degree of absence from daycare, school and work.

The highest incidence of pharyngotonsillitis occurs during the winter season and among individuals in the 5–15 years age-group (28). Pharyngotonsillitis can be caused by a wide variety of pathogens (Table 1). Viruses account for over 50% of all cases of pharyngotonsillitis (2). Of bacterial pathogens, beta-hemolytic group A streptococci (e.g. *Streptococcus pyogenes*) are responsible for 15–30% of all cases of pharyngotonsillitis, with higher rates of occurrence during the winter months in temperate climates (3, 13, 86, 92). Patients with pharyngotonsillitis caused by beta-hemolytic group A streptococci are candidates for antibiotic treatment. However, it is of utmost importance to prevent the over-prescription of antibiotics for pharyngotonsillitis that is not caused by beta-hemolytic group A streptococci, and prescription of antibiotics should especially be avoided for patients in whom the disease is caused by a virus (50). As described below, patients in whom sore throat is caused by bacteria other than beta-hemolytic group A streptococci may also benefit from antibiotic therapy.

Patients with severe symptoms, such as difficulties in swallowing, or dysphonia or stridor, must be evaluated for 'deeper' throat infections, such as epiglottitis, peritonsillitis, parapharyngeal abscess and Lemierre's syndrome. These illnesses, however, are not discussed in this article. This review deals exclusively with pharyngotonsillitis.

## Beta-hemolytic group A streptococci

### Grouping of streptococci

Identification of bacterial strains constitutes the basic tool in studies of the epidemiology and treatment

outcome of pharyngotonsillitis. The streptococcal group of organisms are classified into Lancefield's serological groups A–U according to carbohydrate antigens in the cell wall. Based on the presence of T antigen, beta-hemolytic group A streptococci are divided into about 30 T types (46).

Further subdivision of group A streptococci is made on the basis of the M-protein (45). DNA technologies of the past decade has resulted in the classification of the M-type of beta-hemolytic group A streptococci based upon the sequence of the M-type gene *emm*, and more than 120 *emm*-sequence types have been identified (20).

### Virulence factors

Beta-hemolytic group A streptococci possess a multitude of extracellular and cell-bound virulence factors, which aid in various stages of tissue invasion and are responsible for different disease manifestations.

The cell wall M-protein, an extended alpha-helical protein with anticomplementary and antiphagocytic properties, is considered as the main virulence determinant of beta-hemolytic group A streptococci. M-proteins, by interacting with plasma proteins such as immunoglobulin G, fibrinogen and C4-binding protein, are able to interfere with innate and adaptive host immune responses (9). As recognized for decades, only type-specific antibodies directed towards the N-terminal part of the M-protein will be opsonic and protect against beta-hemolytic group A streptococcal disease (24).

The hyaluronic capsule, although poorly expressed *in vitro*, is a second antiphagocytic factor of beta-hemolytic group A streptococci (90). The T-protein, previously not implicated as biologically important, was recently shown to be present in fimbriae-like structures in beta-hemolytic group A streptococci and probably plays a role in tissue adhesion (56). Pyrogenic exotoxins (erythrogenic toxins) are superantigens capable of triggering T lymphocytes to

**Table 1.** Infectious agents of pharyngotonsillitis

Beta-hemolytic streptococcus group A
Beta-hemolytic streptococcus group C and group G
<i>Arcanobacterium hemolyticum</i>
<i>Corynebacterium diphtheriae</i>
Spirochetes and fusiform bacteria (Vincent's angina)
<i>Neisseria gonorrhoeae</i>
<i>Mycoplasma pneumoniae</i>
<i>Chlamydia pneumoniae</i>
Adenovirus
Epstein-Barr virus
Cytomegalovirus
Rhinovirus
Parainfluenza virus
Respiratory syncytial virus
Coronavirus
Herpes simplex virus
Coxsackie A virus
Influenza A and B viruses
Human immunodeficiency virus (primary infection)

release massive amounts of cytokines and interleukins, thereby generating severe symptoms such as fever, the scarlatiniform rash, tissue necrosis, hypotension and organ failure (4). The cysteine protease (identical to exotoxin SpeB) according to experimental work may be essential for circulatory shock and lung damage (31, 32). This enzyme, and a second cysteine protease of beta-hemolytic group A streptococci, may cleave immunoglobulin G, thereby interfering with immune opsonization of beta-hemolytic group A streptococci. Streptolysins S and O are capable of lysing erythrocytes as well as leukocytes and platelets (26, 77). Streptokinase, which converts plasminogen to plasmin, may significantly contribute to a rapid spread of beta-hemolytic group A streptococci within infected tissue by lysing blood clots (51).

### Internalization

The ability of beta-hemolytic group A streptococci, especially in the stationary phase, to invade respiratory epithelial cells was recently demonstrated (47, 94). Beta-hemolytic group A streptococci are mainly extracellular bacteria, but by specifically binding

fibronectin, a protein that exists in human blood plasma and in the extracellular matrix, beta-hemolytic group A streptococci may be efficiently internalized into human mucosal cells. The fibronectin bound to the bacterial surface thereby acts like a bridging molecule with host cell integrins, which in turn initiate the uptake process that leads to internalization (45).

Sela & Barzilai (76) found that beta-hemolytic group A streptococcal strains were able to survive for 4–7 days inside cultured epithelial cells. They also found that beta-hemolytic group A streptococcal strains from patients with treatment failure harboured an internalization-associated gene in higher prevalence than strains recovered from patients in whom the streptococcal strains had been successfully eradicated. It is not known if internalization into host cells can influence the severity of beta-hemolytic group A streptococcal infections. Strains of streptococci show varying capacity to internalize (47, 94), and internalized beta-hemolytic group A streptococci have been found in asymptomatic carriers as well as in patients with pharyngotonsillitis (95). Strains from patients with treatment failure exhibit a significantly increased ability to survive intracellularly in cultured epithelial cells compared with strains from successfully treated patients (53). On the other hand, a beta-hemolytic group A streptococcal strain that was able to internalize in an animal model was less able to cause serious disease than beta-hemolytic group A streptococci without the internalizing capacity (59).

### Disease manifestations

Beta-hemolytic group A streptococci are strict human pathogens giving rise to a wide range of infectious diseases. Impetigo, pharyngotonsillitis and erysipelas may be comparatively mild and are effectively treated with antibiotics. However, since the late 1980s, a rising number of life-threatening invasive beta-hemolytic group A streptococcal infections have been encountered, such as necrotizing fasciitis and streptococcal toxic shock syndrome (12, 36). Streptococci in these diseases are often restricted to certain M-types, in particular M1, and produce powerful superantigens, such as the pyrogenic exotoxin A, SpeA. Surgical intervention is often needed in the case of necrotizing fasciitis. However, in spite of antibiotics and intensive care, the mortality is high for patients with necrotizing fasciitis or streptococcal toxic shock syndrome (14, 19). Acute rheumatic fever, the most serious nonsuppurative complication of beta-hemolytic group A streptococcal

pharyngotonsillitis, is the leading cause of acquired heart disease among children in developing countries (5). Although no longer a significant health problem in most socioeconomically advanced countries, limited outbreaks of acute rheumatic fever occurred in the USA in the 1980s (88).

Acute poststreptococcal glomerulonephritis, a major cause of child renal failure, occurs after throat and skin infections with beta-hemolytic group A streptococci (87). Large epidemics are still occurring in developing countries, compared with sporadic cases in the developed world.

## Clinical diagnosis

Signs and symptoms of acute pharyngotonsillitis include fever, throat angina, redness of the tonsils and pharynx, tonsillar exudate, enlarged and tender cervical lymph nodes, dysphagia and headache. Concurrent symptoms from the respiratory tract (e.g. cough or rhinorrhea) indicate a viral origin of the disease. Established beta-hemolytic group A streptococcal throat and skin infections in close surroundings of the patient such as at home, or in school or daycare centres, increases the probability of a beta-hemolytic group A streptococcal origin of the infection. Certain symptoms are more pronounced in pharyngotonsillitis of a beta-hemolytic group A streptococcal origin than in pharyngotonsillitis of another etiology. Thus, a high degree of redness in the throat (71), fever (30) and a shorter duration of symptoms before seeking medical care (79) correlate significantly with a beta-hemolytic group A streptococci etiology. The so-called Centor criteria have been widely used to implicate beta-hemolytic group A streptococci in adult patients and these are as follows: tonsillar exudates, tender anterior cervical adenopathy, fever by history and absence of cough. If three of these criteria are met, the positive predictive value for a beta-hemolytic group A streptococcal infection is 40–60% (11). The findings are, however, inconsistent and although the clinical picture may be of some guidance, it is seldom sufficient for establishing a reliable etiological diagnosis.

## Microbiological diagnosis of beta-hemolytic group A streptococcal pharyngotonsillitis

As there are no pathognomonic signs or symptoms that can provide a definite diagnosis of beta-hemolytic group A streptococcal pharyngotonsillitis, the diagnosis depends on the identification of the bac-

terium. Bacterial identification can be performed by obtaining a throat culture or by a rapid antigen detection test. A good view of the pharynx and obtaining proper sampling technique are essential to achieve a representative sample. The specimen should be obtained from the tonsillar surface because beta-hemolytic group A streptococci in pharyngotonsillitis are predominantly localized on the tonsils and on the posterior oropharyngeal wall (48). The rapid antigen test requires more bacteria than required for culture. The quantity of sample material thus influences the sensitivity and specificity of the rapid tests, which are currently reported as being 74–97% and 89–95%, respectively (49, 58).

An asymptomatic carriage stage of beta-hemolytic group A streptococci is reported to occur in about 10% of adults and 25% of children during the winter season, although the estimates vary considerably (71). During large outbreaks of beta-hemolytic group A streptococcal pharyngotonsillitis, the carrier rate can be as high as 60% (21). A 4-year longitudinal study of 5–15-year-old schoolchildren found that the period during which a child carried beta-hemolytic group A streptococci of the same *emm* type averaged 10.8 weeks (range: 3–34 weeks). Many children, however, experienced several periods of carriage during the study and frequently exhibited switches in *emm* type (54). The risk of becoming a carrier of beta-hemolytic group A streptococci, as well as of contracting pharyngotonsillitis, is related to the period of time spent in close contact with a patient during the week preceding the onset of illness (18, 89). The reason why some individuals become carriers is unknown, but the carrier state appears to be a relatively harmless condition because it probably does not result in a clinical infection (39) and the streptococci are present in low numbers (71), which probably does not support a person-to-person transmission of the organism (21). However, problems arise if a carrier of beta-hemolytic group A streptococci acquires viral pharyngitis, because a positive test for beta-hemolytic group A streptococci raises the issue of antibiotic treatment. This example highlights the importance of a careful evaluation of symptoms in order to avoid unnecessary antibiotic treatment.

## Laboratory findings

A correlation with leukocytosis (33, 71) or an increased level of C-reactive protein and beta-hemolytic group A streptococcal pharyngotonsillitis has been reported (40), whereas other investigators have

failed to verify such relationships (61, 70). Serological tests of anti-streptolysin O and anti-DNase B are of no diagnostic value in acute pharyngotonsillitis, but may be useful in the investigation of complications of the disease, such as rheumatic fever (69).

### Reasons to treat beta-hemolytic group A streptococcal pharyngotonsillitis

Beta-hemolytic group A streptococcal pharyngotonsillitis is a self-limiting disease and the routine use of penicillin V has therefore been questioned (25). However, beta-hemolytic group A streptococci are amongst the most virulent human pathogens, and patients with pharyngotonsillitis caused by infection with this bacterium can be seriously affected with high fever, dysphagia and severe pain. Although a majority of patients become free of symptoms within a week, irrespective of therapy, antibiotic treatment of pharyngotonsillitis caused by beta-hemolytic group A streptococci can significantly shorten the duration of symptoms (13, 16). Antibiotic treatment may also, to some extent, reduce the risk of purulent complications, such as peritonsillitis, otitis and sinusitis (13, 15, 93). In acute rheumatic fever, it is claimed that a majority of the patients have a history of pharyngotonsillitis. The decline of acute rheumatic fever in the developed world may indeed be the result of routine antibiotic use for beta-hemolytic group A streptococcal pharyngotonsillitis, supporting the present principles of treatment. Rheumatic fever is still a major health problem in many developing countries, and beta-hemolytic group A streptococci have been estimated to be the eighth most common source of global mortality caused by a single pathogen (10). In necrotizing fasciitis and streptococcal toxic shock syndrome, however, the port of entry of beta-hemolytic group A streptococci is seldom reported to be pharyngotonsillitis (14, 19).

In sum, the reasons for antibiotic treatment of beta-hemolytic group A streptococcal pharyngotonsillitis are: (i) more rapid alleviation of symptoms; (ii) reducing the spread of beta-hemolytic group A streptococci; and (iii) reducing the risk of suppurative and nonsuppurative complications. It is generally agreed that the benefits of antibiotic treatment outweigh the disadvantages (15, 34, 72).

### Treatment of beta-hemolytic group A streptococcal pharyngotonsillitis

Penicillin V orally for 10 days (12.5 mg/kg, two to four times daily) is currently the antibiotic therapy of

choice for beta-hemolytic group A streptococcal pharyngotonsillitis. Despite over 50 years of use of penicillin, no penicillin-resistant beta-hemolytic group A streptococcal strains have so far been encountered. A possible explanation for this is that penicillin resistance in this species is not compatible with a virulent phenotype (29). In penicillin V treatment of beta-hemolytic group A streptococcal pharyngotonsillitis, a treatment period of at least 10 days should be imposed in order to achieve an acceptably low recurrence rate (27, 75, 82, 93). Despite the absence of penicillin resistance, treatment failure of beta-hemolytic group A streptococcal pharyngotonsillitis is as high as 5–25% (82). A second course of penicillin V treatment is followed by still higher failure rates (41), in some cases necessitating tonsillectomy.

Cephalosporins have been shown to be more effective than penicillin V in treating primary beta-hemolytic group A streptococcal pharyngotonsillitis (37, 55, 67). Cephalosporins may enable shorter treatment regimens than penicillin V, and some patients may only need to be dosed once daily (68). Cephalosporins are less susceptible to the  $\beta$ -lactamases produced by oral bacteria and probably exert a lesser impact on bacteriocin-producing alpha-hemolytic streptococci in the throat (37, 73).

Treatment of recurrent beta-hemolytic group A streptococcal pharyngotonsillitis with clindamycin instead of penicillin V seems to produce a significantly better clinical outcome (6, 62). This may be a result of the intracellular accumulation and a high concentration of clindamycin in tonsillar surface fluid (61).

Penicillin V, as a result of its narrow antimicrobial spectrum and the absence of penicillin resistance among beta-hemolytic group A streptococci, is still regarded as the drug of choice for primary beta-hemolytic group A streptococcal pharyngotonsillitis. Cephalosporins or clindamycin appear to be appropriate alternatives for patients experiencing recurrence of pharyngotonsillitis after penicillin V treatment.

The problem of frequent recurrence of beta-hemolytic group A streptococcal pharyngotonsillitis in patients is sometimes resolved by performing tonsillectomy, although opinions differ as to the benefit and risks of this treatment (8, 65, 66). In a Cochrane Review from 2000, no studies fulfilled the inclusion criteria for evaluating the effectiveness of tonsillectomy in adults with chronic/recurrent acute pharyngotonsillitis. Only two studies fulfilled the

criteria assessing the benefit of tonsillectomy in children with chronic/recurrent acute pharyngotonsillitis, and only limited conclusions could be drawn from these studies (8). Further studies are needed to evaluate the effectiveness of tonsillectomy in relation to pharyngotonsillitis.

### Reasons for recurrence of beta-hemolytic group A streptococcal pharyngotonsillitis after penicillin V treatment

Possible factors that may lead to recurrence after penicillin V treatment of pharyngotonsillitis include low patient compliance, re-infection from the environment, eradication of alfa-hemolytic streptococci with inhibitory effects on beta-hemolytic group A streptococci, an increase in beta-lactamase-producing bacteria inactivating the drug, penicillin-tolerant streptococci, a low antibiotic concentration at the site of infection and intracellular beta-hemolytic group A streptococci surviving therapy (Table 2).

#### Low compliance

Treatment of beta-hemolytic group A streptococcal pharyngotonsillitis with penicillin V results in rapid recovery (16, 93). As the patient is often free of symptoms after 2–3 days of treatment, they may consider further medication as unnecessary and prematurely discontinue the antibiotic treatment, raising the odds of treatment failure.

#### Re-infection from the environment

As family members and other close contacts of patients with beta-hemolytic group A streptococcus pharyngotonsillitis are often infected with the same strain, recurrence may be caused by re-infection from other individuals (22).

**Table 2.** Possible causes of recurrence of beta-hemolytic group A streptococcal pharyngotonsillitis after treatment with penicillin V

Low compliance
Re-infection from the environment
Eradication of alfa-hemolytic streptococci by penicillin V
Increase in beta-lactamase-producing bacteria capable of inactivating penicillin V
Penicillin-tolerant beta-hemolytic group A streptococci
Low antibiotic concentration at the site of infection
Intracellular beta-hemolytic group A streptococci capable of surviving therapy with penicillin V

### Alpha-hemolytic streptococci with inhibitory effect on beta-hemolytic group A streptococci

Some alpha-hemolytic streptococci produce bacteriocins with inhibitory activity against beta-hemolytic group A streptococci. Eradication of alpha-hemolytic streptococci by penicillin V will theoretically reduce bacterial interference and thus increase the risk of treatment failure (74). However, some investigations have failed to show that lack of bacterial interference was related to bacterial treatment failure in beta-hemolytic group A streptococcal pharyngotonsillitis (27). On the other hand, administration of alpha-hemolytic streptococci into the throat following  $\beta$ -lactam treatment of beta-hemolytic group A streptococcal pharyngotonsillitis has been able to reduce the recurrence rate (23, 73). However, replacement treatment with alpha-hemolytic streptococci is not yet commercially available.

### Increase in $\beta$ -lactamase-producing bacteria inactivating the drug

Treatment with penicillin V will promote selection of bacterial species producing  $\beta$ -lactamase, conceivably accounting for the inactivation of penicillin V (7, 84). However, studies on penicillin V vs. amoxicillin/clavulanic acid in the treatment of pharyngotonsillitis have been inconclusive (41, 83). Gerber et al. (27) studied penicillin V and cefadroxil in the treatment of primary beta-hemolytic group A streptococcus pharyngotonsillitis and found no evidence that  $\beta$ -lactamases produced by the normal pharyngeal flora were related to bacterial treatment failure. The role of  $\beta$ -lactamases in treatment failure thus remains unclear.

### Penicillin tolerance

Tolerance to  $\beta$ -lactam antibiotics is a known phenomenon in some medically important species, such as *Enterococcus faecalis*, *Streptococcus pneumoniae* and various alpha-hemolytic streptococci (85), and appears to account for the failure of penicillin therapy of *Arcanobacterium haemolyticum* infections (60). Penicillin tolerance of beta-hemolytic group A streptococci has been suggested to account for the failure in penicillin V treatment of beta-hemolytic group A streptococcal pharyngotonsillitis, but reports have been contradictory, conceivably because of the variability in the definition of ‘tolerance’ as well as the technical pitfalls of the methods used (42, 44, 63, 78, 80). The existence of penicillin tolerance in

beta-hemolytic group A streptococci has also been questioned (91).

#### Low antibiotic concentration at the site of infection

The beta-hemolytic group A streptococci causing acute pharyngotonsillitis are mainly present in the secretion on the tonsillar surface and in the crypts rather than in the parenchyma (17, 48). Penicillin V was detected in tonsillar surface fluid in a majority of patients on the first day of treatment of acute beta-hemolytic group A streptococcus pharyngotonsillitis, but despite a high concentration in serum, was rarely present on the 10th day or in healthy, treated subjects (81). Insufficient concentrations of antibiotics in tonsillar surface fluid may contribute to treatment failure of beta-hemolytic group A streptococcal pharyngotonsillitis (64).

#### Intracellular beta-hemolytic group A streptococcal surviving therapy

Beta-hemolytic group A streptococci may reside in epithelial cells of pharyngotonsillitis lesions, and may not be reached by penicillin V. Österlund et al. (95) found that beta-hemolytic group A streptococci, which were internalized in human respiratory epithelial cells and grown in an antibiotic supplemented medium, were externalized as soon as the extracellular antibiotic was removed and an extracellular infection developed rapidly thereafter. Thus, respiratory epithelial cells may act as a reservoir for internalized beta-hemolytic group A streptococci and have the potential to cause infection after termination of a penicillin V treatment. The extent to which epithelial cell-embedded beta-hemolytic group A streptococci cause recurrence of pharyngotonsillitis after penicillin V treatment remains to be determined.

## Bacteria other than beta-hemolytic Group A streptococci

### Nongroup A beta-hemolytic streptococci

Group C and group G streptococci may cause pharyngotonsillitis, especially in older children and adults (35). Rapid tests for detecting these streptococcal groups are not available and thus bacterial culture is needed for diagnosis. Evidence suggests that group C and group G streptococci can cause acute poststreptococcal glomerulonephritis, but not rheumatic fever.

## *Arcanobacterium hemolyticum*

*A. hemolyticum* is an uncommon cause of pharyngotonsillitis and is often associated with a rash that may mimic scarlatina (1). The disease is found most commonly in individuals of the 15–18 years age-group and the incidence is about 2.5% among patients with pharyngotonsillitis in this age group (52). The organism must be cultured on specific media and is not sensitive to penicillin but is sensitive to macrolides. *A. hemolyticum*-related pharyngotonsillitis is self-limiting with no known complications.

## *Corynebacterium diphtheriae*

Pharyngotonsillitis caused by *C. diphtheriae* was relatively common during World War II, but has rapidly declined in Europe since that time. In a vaccinated population the disease is very rare. However, *C. diphtheriae*-related pharyngotonsillitis is not uncommon in the former Soviet Union countries where its incidence is probably under-reported.

The pharyngotonsillitis caused by *C. diphtheriae* is characterized by grey membranes in the throat, larynx and around the nostrils. Bleeding occurs if attempts are made to remove the membranes. The bacteria produce an exotoxin that gives rise to myocarditis and neurological complications with paralysis and polyneuropathia. Treatment with parenteral antibiotics and specific immunoglobulin should be started immediately and even before the microbiological diagnosis in cases of a high suspicion of *C. diphtheriae*-related pharyngotonsillitis.

## Vincent's angina

Vincent's angina is an uncommon infection presenting with unilateral necrotizing tonsillitis, caused by a mixed anaerobic and aerobic flora of spirochetes, bacteroides, streptococci and fusiform bacteria. Penicillin is the drug of choice. The patient must be followed-up after a couple of weeks to rule out malignancy, because carcinoma of the tonsils can show a similar clinical picture.

## *Neisseria gonorrhoeae*

*N. gonorrhoeae* may be a cause of pharyngotonsillitis in individuals practising oral sex. A study of patients with sore throats in a general medical practice found the prevalence of *N. gonorrhoeae* to be about 2% (43). About 10% of patients with genital gonorrhoea have

throat cultures positive for *N. gonorrhoeae* and are often asymptomatic. *N. gonorrhoeae* must be grown on specific media. Antibiotic treatment is needed to prevent transmission as well as further dissemination of the organism and the disease.

### ***Mycoplasma pneumoniae*, *Chlamydia pneumoniae***

Pharyngotonsillitis involving *M. pneumoniae* or *C. pneumoniae* is frequently accompanied by bronchitis. Infections with these two bacteria often affect children and young adults.

## **Nonbacterial pharyngotonsillitis**

### **Epstein–Barr virus / mononucleosis**

Most individuals become infected with Epstein–Barr virus during childhood and will only experience a subclinical infection or mild pharyngitis. However, about one-third of individuals who become infected during adolescence develop the mononucleosis syndrome. Infectious mononucleosis is a systemic disease characterized by pharyngotonsillitis, nodular fever and septicemia. Patients show prominent exudate and swelling of the tonsils, cervical adenopathy, hepatomegaly and often splenomegaly. Mononucleosis patients with beta-hemolytic group A streptococcal pharyngotonsillitis show a prolonged course of pharyngotonsillitis, fever, fatigue and tender cervical adenopathy. Heterophile antibodies are not detectable until several weeks after the onset of mononucleosis, but immunoglobulin M specific antibodies against capsid and early Epstein–Barr virus antigens can be helpful for early diagnosis. Mononucleosis in most patients will resolve without special treatment. Some patients having difficulty with swallowing and breathing will need hospital care. Cytomegalovirus causes about 10% of mononucleosis cases, and hepatitis is more evident than with Epstein–Barr virus-related disease. Testing for specific cytomegalovirus antibodies are of diagnostic importance.

### **Primary human immunodeficiency virus infection**

The clinical presentation of a primary human immunodeficiency virus (HIV) infection may mimic that of Epstein–Barr virus mononucleosis. The incubation period is about 2–3 weeks. The acute HIV

syndrome presents with pharyngotonsillitis, weight loss, fever, adenopathy, rash and splenomegaly. Laboratory tests often show lymphopenia and increased transaminase levels (38). Tests for anti-HIV immunoglobulins are often negative in the early disease stages but detection of HIV RNA by polymerase chain reaction is helpful in providing a diagnosis. It is important to establish the HIV diagnosis as early as possible to initiate an appropriate antiviral therapy.

### **Influenza virus**

Pharyngotonsillitis caused by influenza virus is often exudative. The disease usually occurs in major outbreaks and is often associated with myalgias, fever, tracheitis and headache. Rapid tests for the detection of influenza A and B viruses may be useful, and antiviral therapy may reduce the illness duration by about 24 h but is seldom advocated.

### **Adenovirus**

Adenovirus is a common cause of pharyngotonsillitis. Pharyngotonsillitis caused by adenovirus often presents as pharyngoconjunctival fever (57). The disease is otherwise difficult to distinguish from pharyngotonsillitis caused by other pathogens.

### **Herpes simplex virus**

Pharyngotonsillitis caused by herpes simplex virus often appears with vesicles and ulcerative lesions of the mouth and gingiva, and the disease occurs most frequently in children. The throat pain can be very severe, and hospitalization and parenteral nutrition may be indicated. Herpes simplex virus type 1 is by far the most common cause of the disease, but herpes simplex virus type 2 can present similar clinical manifestations.

### **Coxsackie A virus / herpangina**

This coxsackie A enterovirus can give rise to pharyngotonsillitis that is characterized by small vesicles surrounded by a red halo in the posterior palate. The disease is virtually always seen in children.

### **Noninfectious pharyngotonsillitis**

Agranulocytosis and leukaemia might occasionally start as a severe pharyngotonsillitis that does not

respond to antibiotics. Laboratory blood tests are needed to confirm the diagnosis.

Periodic fever syndrome characterized by repeated episodes of pharyngotonsillitis with cervical adenopathy, fever, headache, nausea, reduced general condition and elevated C-reactive protein can occur in children with no proven etiological pathogen. There is no evidence-based treatment, but tonsillectomy is considered to have an effect (96).

## Summary

Acute pharyngotonsillitis is a common illness with the highest incidence occurring during the winter season. Viruses account for more than half of the cases of pharyngotonsillitis. Beta-hemolytic group A streptococci are the etiological agents in 15–30% of cases, with higher figures observed during the winter months. Beta-hemolytic group A streptococci are virulent human pathogens, which may cause suppurative and nonsuppurative complications, and sometimes life-threatening diseases such as necrotizing fasciitis and streptococcal toxic shock syndrome. Signs and symptoms of a beta-hemolytic group A streptococcal infection include tonsillar exudate, tender anterior cervical adenopathy, a history of fever and absence of a cough, but the findings are rather inconsistent. Identification of the bacterium by culture or by a rapid antigen test is needed to establish a definite diagnosis. Penicillin V for 10 days is the drug of choice in primary beta-hemolytic group A streptococcal pharyngotonsillitis. Although penicillin resistance is not recorded in beta-hemolytic group A streptococci, penicillin V treatment fails in as many as 5–25% of the patients, and higher failure rates can occur after a second course of penicillin V treatment, sometimes necessitating tonsillectomy. Factors potentially contributing to antibiotic treatment failure include low compliance rate, reinfection from the environment, eradication of alpha-hemolytic streptococci with an inhibitory effect on beta-hemolytic group A streptococci, an increase in beta-lactamase-producing bacteria inactivating the drug, penicillin-tolerant streptococci, low antibiotic concentration at the site of infection, and intracellular beta-hemolytic group A streptococci surviving therapy. Treatment with cephalosporins or clindamycin has been reported to result in a less frequent recurrence of beta-hemolytic group A streptococcal pharyngotonsillitis and is advocated in patients with beta-hemolytic group A streptococcus pharyngotonsillitis not responding to penicillin V treatment.

## References

- Banck G, Nyman M. Tonsillitis and rash associated with *Corynebacterium haemolyticum*. *J Infect Dis* 1986; **154**: 1037–1040.
- Bisno AL. Acute pharyngitis. *N Engl J Med* 2001; **344**: 205–211.
- Bisno AL. Acute pharyngitis. Etiology and diagnosis. *Pediatrics* 1996; **97**: 949–954.
- Bisno AL, Brito MO, Collins CM. Molecular basis of group A streptococcal virulence. *Lancet Infect Dis* 2003; **3**: 191–200.
- Bisno AL. Group A streptococcal infections and acute rheumatic fever. *N Engl J Med* 1991; **325**: 783–793.
- Brook I, Hirokawa R. Treatment of patients with a history of recurrent tonsillitis due to group A  $\beta$ -hemolytic streptococci. A prospective randomized study comparing penicillin, erythromycin, and clindamycin. *Clin Pediatr (Phila)* 1985; **24**: 331–336.
- Brook I. Role of  $\beta$ -lactamase-producing bacteria in the failure of penicillin to eradicate group A streptococci. *Pediatr Infect Dis* 1985; **4**: 491–495.
- Burton MJ, Towler B, Glasziou P. Tonsillectomy versus non-surgical treatment for chronic/recurrent acute tonsillitis. *Cochrane Database Syst Rev* 2000; **2**: CD001802.
- Carlsson F, Sandin C, Lindahl G. Human fibrinogen bound to *Streptococcus pyogenes* M protein inhibits complement deposition via the classical pathway. *Mol Microbiol* 2005; **56**: 28–39.
- Carpetis JR, Steer AC, Mulholland EK, Weber M. The global burden of group A streptococcal diseases. *Lancet Infect Dis* 2005; **5**: 685–694.
- Centor RM, Whitterspoon JM, Dalton HP, Brody CE, Link K. The diagnosis of strep throat in adults in the emergency room. *Med Decis Making* 1981; **1**: 239–246.
- Cone LA, Woodard DR, Schlievert PM, Tomory GS. Clinical and bacteriologic observations of a toxic shock-like syndrome due to *Streptococcus pyogenes*. *N Engl J Med* 1987; **317**: 146–149.
- Dagnelie CF, van der Graaf Y, De Melker RA. Do patients with sore throat benefit from penicillin? A randomized double-blind placebo-controlled clinical trial with penicillin V in general practice *Br J Gen Pract* 1996; **46**: 589–593.
- Davis HD, McGeer A, Schwartz B, Green K, Cann D, Simor AE, Low DE. Invasive group A streptococcal infections in Ontario, Canada. Ontario Group A streptococcal Study Group. *N Engl J Med* 1996; **335**: 547–554.
- Del Mar CB, Glasziou PP, Spinks AB. Antibiotics for sore throat. *Cochrane Database Syst Rev* 2006; **4**: CD000023.
- De Meyere M, Mervielde Y, Verschraegen G, Bogaert M. Effect of penicillin on the clinical course of streptococcal pharyngitis in general practice. *Eur J Clin Pharmacol* 1992; **43**: 581–585.
- Ebenfelt A, Ericson LE, Lundberg C. Acute pharyngotonsillitis is an infection restricted to the crypt and surface secretion. *Acta Otolaryngol (Stockh)* 1998; **118**: 264–271.
- Engelgau MM, Woernle CH, Schwartz B, Vance NJ, Horan JM. Invasive group A streptococcus carriage in a child care centre after a fatal case. *Arch Dis Child* 1994; **71**: 318–322.
- Eriksson BK, Andersson J, Holm SE, Norgren M. Epidemiological and clinical aspects of invasive group A



- streptococcal infections and the streptococcal toxic shock syndrome. *Clin Infect Dis* 1998; **27**: 1428–1436.
20. Facklam RF, Martin DR, Lovgren M, Johnson DR, Efstratiou A, Thompson TA, Gowan S, Kriz P, Tyrrell GJ, Kaplan E, Beall B. Extension of the Lancefield classification for group A streptococci by addition of 22 new M protein gene sequence types from clinical isolates: emm103 to emm124. *Clin Infect Dis* 2002; **34**: 28–38.
  21. Falck G, Kjellander J. Outbreak of group A streptococcal infection in a day-care center. *Pediatr Infect Dis J* 1992; **11**: 914–919.
  22. Falck G, Holm SE, Kjellander J, Norgren M, Schwan Å. The role of household contacts in the transmission of group A streptococci. *Scand J Infect Dis* 1997; **29**: 239–244.
  23. Falck G, Grahn-Håkansson E, Holm SE, Roos K, Lagergren L. Tolerance and efficacy of interfering alpha-streptococci in recurrence of streptococcal pharyngotonsillitis: a placebo-controlled study. *Acta Otolaryngol (Stockh)* 1999; **119**: 944–948.
  24. Fischetti VA. Streptococcal M protein: molecular design and biological behaviour. *Clin Microbiol Rev* 1989; **2**: 285–314.
  25. Flottorp S, Oxman AD, Cooper JG, Hjortdahl P, Sandberg S, Vorland LH. Guidelines for diagnosis and treatment of sore throat. *Tidsskr Nor Laegeforen* 2000; **10**: 1754–1760.
  26. Fontaine MC, Lee JJ, Kehoe MA. Combined contributions of streptolysin O and streptolysin S to the virulence of serotype M5 *Streptococcus pyogenes* strain Manfredo. *Infect Immun* 2003; **71**: 3857–3865.
  27. Gerber MA, Tanz RR, Kabat W, Bell GL, Siddiqui B, Lerer TJ, Lepow ML, Kaplan EL, Shulman ST. Potential mechanisms for failure to eradicate group A streptococci from the pharynx. *Pediatrics* 1999; **104**: 911–917.
  28. Glezen WP, Clyde WA Jr, Senior RJ, Sheaffer CI, Denny FW. Group A streptococci, mycoplasmas, and viruses associated with acute pharyngitis. *JAMA* 1967; **202**: 119–124.
  29. Gutman L, Tomasz A. Penicillin-resistant and penicillin-tolerant mutants of group A streptococci. *Antimicrob Agents Chemother* 1982; **22**: 128–136.
  30. Hansen J, Schmidt H, Bitsch N. Sore throat. Principles of diagnosis and treatment. *Practitioner* 1983; **227**: 937–948.
  31. Herwald H, Mörgelin M, Olsen A, Rhen M, Dahlbäck B, Muller-Esterl W, Björck L. Activation of the contact-phase system on bacterial surfaces – a clue to serious complications in infectious diseases. *Nat Med* 1998; **4**: 298–302.
  32. Herwald H, Cramer H, Mörgelin M, Russel W, Sollenberg U, Norrby-Teglund A, Flodgaard H, Lindbom L, Björck L. M protein, a classical bacterial virulence determinant, forms complexes with fibrinogen that induce vascular leakage. *Cell* 2004; **116**: 367–379.
  33. Hjortdahl P, Melbye H. Does near-to-patient testing contribute to the diagnosis of streptococcal pharyngitis in adults? *Scand J Prim Health Care* 1994; **12**: 70–76.
  34. Hofkosh D, Wald ER, Chiponis DM. Prevalence of non-group A beta-hemolytic streptococci in childhood pharyngitis. *South Med J* 1988; **81**: 329–331.
  35. Hoffman S, Kolmos HJ. Effect of antibiotics on symptoms and complications of sore throat. Comments on a meta-analysis from the Cochrane Collaboration. *Läkartidn* 2000; **97**: 2730–2732.
  36. Hoge CW, Schwartz B, Talkington DF, Breiman RF, McNeill EM, Engleender SJ. The changing epidemiology of invasive group A streptococcal infections and the emergence of streptococcal toxic shock-like syndrome. A retrospective population-based study. *JAMA* 1993; **269**: 384–389.
  37. Holm SE, Roos K, Strömberg A. A randomized study of treatment of streptococcal pharyngotonsillitis with cefadroxil or phenoxymethylpenicillin (penicillin V). *Pediatr Infect Dis J* 1991; **10**: S68–S71.
  38. Kahn JO, Walker BD. Current concepts. Acute human immunodeficiency virus type 1 infection. *N Engl J Med* 1998; **339**: 33–39.
  39. Kaplan EL, Gastanaduy AS, Huwe BB. The role of carrier in treatment failures after antibiotic for group A streptococci in the upper respiratory tract. *J Lab Clin Med* 1981; **98**: 326–335.
  40. Kaplan EL, Wannamaker LW. C-reactive protein in streptococcal pharyngitis. *Pediatrics* 1977; **60**: 28–32.
  41. Kaplan EL, Johnson DR. Eradication of group A streptococci from the upper respiratory tract by amoxicillin with clavulanate after oral penicillin V treatment failure. *J Pediatr* 1988; **113**: 400–403.
  42. Kim KS, Kaplan EL. Association of penicillin tolerance with failure to eradicate group A streptococci from patients with pharyngitis. *J Pediatr* 1985; **107**: 681–684.
  43. Komaroff A, Aronson M, Pass T, Ervin C. Prevalence of pharyngeal gonorrhoea in general medical patients with sore throats. *Sex Transm Dis* 1980; **7**: 116–119.
  44. Krasinski K, Hanna B, LaRussa P, Desiderio D. Penicillin tolerant group A streptococci. *Diagn Microbiol Infect Dis* 1986; **4**: 291–297.
  45. Kreikemeyer B, Klenk M, Podbielski A. The intracellular status of *Streptococcus pyogenes*: role of extracellular matrix-binding proteins and their regulation. *Int J Med Microbiol* 2004; **294**: 177–188.
  46. Lancefield RC. The antigenic complex of *Streptococcus haemolyticus*. I. Demonstration of a type-specific substance in extracts of *Streptococcus haemolyticus*. *J Exp Med* 1928; **47**: 91–103.
  47. La Penta D, Rubens XC, Chi E, Cleary P. Group A streptococci efficiently invade human respiratory epithelial cells. *Proc Natl Acad Sci USA* 1994; **91**: 12115–12119.
  48. Lilja M, Myklebust R, Raisanen S, Stenfors LE. Selective attachment of  $\beta$ -haemolytic streptococci group A to oropharyngeal epithelium in health and disease. *Acta Otolaryngol (Stockh)* 1997; **117**: 744–749.
  49. Lindbaek M, Hoiby EA, Lermark G, Steinsholt IM, Hjortdahl P. Which is the best method to trace group A streptococci in sore throat patients: culture or GAS antigen test? *Scand J Prim Health Care* 2004; **22**: 233–238.
  50. Linder JA, Stafford RS. Antibiotic treatment of adults with sore throat by community primary care physicians: a national survey, 1989–1999. *JAMA* 2001; **286**: 1181–1186.
  51. Lottenberg R, Minning-Wenz D, Boyle MD. Capturing host plasmin(ogen): a common mechanism for invasive pathogens. *Trends Microbiol* 1994; **2**: 20–24.
  52. Mackenzie A, Fuite LA, Chan FT, King J, Allen U, MacDonald N, Diaz-Mitoma F. Incidence and pathogenicity of *Arcanobacterium haemolyticum* during a 2-year study in Ottawa. *Clin Infect Dis* 1995; **21**: 177–181.
  53. Marouni MJ, Barzilai A, Keller N, Rubinstein E, Sela S. Intracellular survival of persistent group A streptococci in cultured epithelial cells. *Int J Med Microbiol* 2004; **294**: 27–33.

54. Martin JM, Green M, Barbadora KA, Wald ER. Group A streptococci among school-aged children: clinical characteristics and the carrier state. *Pediatrics* 2004; **114**: 1212–1219.
55. Milatovic D, Knauer J. Cefadroxil versus penicillin in the treatment of streptococcal tonsillopharyngitis. *Eur J Clin Microbiol Infect Dis* 1989; **8**: 282–288.
56. Mora M, Bensi G, Capo S, Falugi F, Zingaretti C, Manetti AG, Maggi T, Taddei AR, Grandi G, Telford JL. Group A *Streptococcus* produce pilus-like structures containing protective antigens and Lancefield T antigens. *Proc Natl Acad Sci USA* 2005; **102**: 15641–15646.
57. Nakayama M, Miyazaki C, Ueda K, Kusuhara K, Yoshikawa H, Nishima S, Shibata R, Tokugawa K. Pharyngoconjunctival fever caused by adenovirus type 11. *Pediatr Infect Dis J* 1992; **11**: 6–9.
58. Nerbrand C, Jasir A, Schalen C. Are current rapid detection tests for Group A Streptococci sensitive enough? Evaluation of 2 commercial kits *Scand J Infect Dis* 2002; **34**: 797–799.
59. Nyberg P, Sakai T, Cho HK, Caparon MG, Fässler R, Björck L. Interactions with fibronectin attenuate the virulence of *Streptococcus pyogenes*. *EMBO J* 2004; **23**: 2.
60. Nyman M, Banck G, Thore M. Penicillin tolerance in *Arcanobacterium haemolyticum*. *J Infect Dis* 1990; **161**: 261–265.
61. Orrling A, Kamme C, Stjernquist-Desatnik A. Penicillin V, loracarbef and clindamycin in tonsillar surface fluid during acute group A streptococcal pharyngotonsillitis. *Scand J Inf Dis* 2005; **37**: 429–435.
62. Orrling A, Stjernquist-Desatnik A, Schalen C. Clindamycin in recurrent Group A streptococcal pharyngotonsillitis – an alternative to tonsillectomy? *Acta Otolaryngol* 1997; **117**: 618–622.
63. Orrling A, Stjernquist-Desatnik A, Schalen C, Kamme C. Treatment failure in streptococcal pharyngotonsillitis. An attempt to identify penicillin tolerant *Streptococcus pyogenes*. *Scand J Infect Dis* 1996; **28**: 143–147.
64. Orrling A, Kamme C, Stjernquist-Desatnik A. Penicillin V, loracarbef and clindamycin in tonsillar surface fluid during acute group A streptococcal pharyngotonsillitis. *Scand J Infect Dis* 2005; **37**: 429–435.
65. Paradise JL, Bluestone CD, Ruth Z, Bachman RZ, Colborn DK, Bernard BS, Taylor FH, Rogers KD, Schwarzbach RH, Stool SE, Friday GA, Smith IH, Saez CA. Efficacy of tonsillectomy for recurrent throat infection in severely affected children. Results of parallel randomized and nonrandomized clinical trials. *N Eng J Med* 1984; **310**: 674–683.
66. Paradise JL, Bluestone CD, Colborn DK, Bernard BS, Rockette HE, Kurs-Lasky M. Tonsillectomy and adenotonsillectomy for recurrent throat infection in moderately affected children. *Pediatrics* 2002; **110**: 7–15.
67. Pichichero ME, Disney FA, Aronovitz GH, Talpey WB, Green JL, Francis AB. Randomized, single-blind evaluation of cefadroxil and phenoxymethyl penicillin in the treatment of streptococcal pharyngitis. *Antimicrob Agents Chemother* 1987; **31**: 903–906.
68. Pichichero ME, Gooch WM, Rodriguez W, Blumer JL, Aronof SC, Jacobs RF, Musser JM. Effective short-course treatment of acute group A  $\beta$ -haemolytic streptococcal tonsillopharyngitis. Ten days of penicillin V vs 5 days or 10 days of cefpodoxime therapy in children. *Arch Pediatr Adolesc Med* 1994; **148**: 1053–1060.
69. Powel C, Parks D. Anti-streptolysin and anti-DNAse B test. In: Isenberg HD editor. *Clinical microbiology procedures handbook*. Washington DC: ASM, 1994: 9.3 1–4–9.4 1–4.
70. Putto A, Meurman O, Ruuskanen O. C-reactive protein in the differentiation of adenoviral, Epstein–Barr viral and streptococcal tonsillitis in children. *Eur J Pediatr* 1986; **145**: 204–206.
71. Roos K. The diagnostic value of symptoms and signs in acute tonsillitis in children over the age of 10 and in adults. *Scand J Infect Dis* 1985; **17**: 259–267.
72. Roos K, Prellner K, Holm S, Larsson P, Stjernquist-Desatnik A, Strömberg A. Norwegian guidelines for “sore throats” nothing for Swedish throats! *Läkartidn* 2000; **97**: 5144–5145.
73. Roos K, Grahn E, Holm SE, Johansson H, Lind L. Interfering alpha-streptococci as a protection against recurrent streptococcal tonsillitis in children. *Int J Pediatr Otorhinolaryngol* 1993; **25**: 141–148.
74. Sanders C, Sanders E, Harrow D. Bacterial interference: effects of oral antibiotics on the normal throat flora and its ability to interfere with group A streptococci. *Infect Immun* 1976; **13**: 808–812.
75. Schwartz RH, Wientzen RL Jr, Pedreira F, Feroli EJ, Mella GW, Guandolo VL. Penicillin V for group A streptococcal pharyngotonsillitis. A randomized trial of seven vs ten days’ therapy. *JAMA* 1981; **246**: 1790–1795.
76. Sela S, Barzilai A. Who do we fail with penicillin in the treatment of group A streptococcus infections? *Ann Med* 1999; **31**: 303–307.
77. Sierig G, Cywes C, Wessels MR, Asbaugh C. Cytotoxic effects of streptolysin O and streptolysin S enhance the virulence of poorly encapsulated group A streptococci. *Infect Immun* 2003; **71**: 446–455.
78. Smith TD, Huskins WC, Kim KS, Kaplan EL. Efficacy of  $\beta$ -lactamase resistant penicillin and influence of penicillin tolerance in eradicating streptococci from the pharynx after failure of penicillin therapy for group A streptococcal pharyngitis. *J Pediatr* 1987; **110**: 777–782.
79. Stjernquist-Desatnik A, Prellner K, Christensen P. Clinical and laboratory findings in patients with acute tonsillitis. *Acta Otolaryngol (Stockh)* 1987; **104**: 351–359.
80. Stjernquist-Desatnik A, Orrling A, Schalen C, Kamme C. Penicillin tolerance in group A streptococci and treatment failure in streptococcal tonsillitis. *Acta Otolaryngol (Stockh) Suppl* 1992; **492**: 68–71.
81. Stjernquist-Desatnik A, Samuelsson P, Walder M. Penetration of penicillin V tonsillar surface fluid in healthy individuals and in patients with acute tonsillitis. *Laryngol Otol* 1993; **107**: 309–312.
82. Strömberg A, Schwan Å, Cars O. Five versus ten days treatment of group A streptococcal pharyngotonsillitis: a randomized controlled clinical trial with phenoxymethylpenicillin and cefradroxil. *Scand J Infect Dis* 1988; **20**: 37–46.
83. Tanz RR, Shulman ST, Sroka PA, Marubio S, Brook I, Yogev R. Lack of influence of  $\beta$ -lactamase-producing flora on recovery of group A streptococci after treatment of acute pharyngitis. *J Pediatr* 1990; **117**: 859–863.

84. Tunér K, Nord CE. Emergence of  $\beta$ -lactamase producing anaerobic bacteria in the tonsils during penicillin treatment. *Eur J Clin Microbiol* 1986; **5**: 399–404.
85. Tuomanen E, Durack DT, Tomasz A. Antibiotic tolerance among clinical isolates of bacteria. *Antimicrob Agents Chemother* 1986; **30**: 521–527.
86. Wannamaker LW. Perplexity and precision in the diagnosis of streptococcal pharyngitis. *Am J Dis Child* 1972; **124**: 352–358.
87. Wannamaker LW. Differences between streptococcal infections of the throat and of the skin. *N Eng J Med* 1970; **8**: 78–85.
88. Veasy LG, Wiedmeier SE, Orsmond GS, Ruttenberg HD, Boucek MM, Roth SJ, Tait VF, Thompson JA, Daly JA, Kaplan EL, Hill HR. Resurgence of acute rheumatic fever in the intermountain area of the United States. *N Engl J Med* 1987; **316**: 421–427.
89. Weiss K, Laverdiere M, Lovgren M, Delorme J, Poirier L, Beliveau C. Group A Streptococcus carriage among close contacts of patients with invasive infection. *Am J Epidemiol* 1999; **149**: 863–868.
90. Wessels MR, Moses AE, Goldberg JB, DiCesare TJ. Hyaluronic acid capsule is a virulence factor for mucoid group A streptococci. *Proc Natl Acad Sci USA* 1991; **88**: 8317–8321.
91. Woolfrey BJ. Penicillin tolerance in  $\beta$ -streptococci. *Scand J Infect Dis* 1988; **20**: 235–237.
92. Woodruff C. Microbiology of infectious diseases of Waldeyer's ring. *Ear Nose Throat J* 1980; **59**: 454–456.
93. Zwart S, Sachs APE, Ruijs GJHM, Gubbels JW, Hoes AW, de Melker RA. Penicillin for acute sore throat: randomized double blind trial of seven days versus three days treatment of placebo in adults. *BMJ* 2000; **320**: 150–154.
94. Österlund A, Engstrand L. Intracellular penetration and survival of *Streptococcus pyogenes* in respiratory epithelial cells in vitro. *Acta Otolaryngol* 1995; **115**: 685–688.
95. Österlund A, Popa R, Nikkilä T, Scheynius A, Engstrand L. Intracellular reservoir of *Streptococcus pyogenes* in vivo: a possible explanation for recurrent pharyngo-tonsillitis. *Laryngoscope* 1997; **107**: 640–647.
96. Öymar K, Kristoffersen EK. Periodic fever syndrome in children. *Tidsskr Nor Laegeforen* 2007; **12**: 1651–1653.