Review Article

Genetic Polymorphisms and Pharmacotherapy for Prostate Cancer

Masaki Shiota¹), Shusuke Akamatsu²), Shintaro Narita³, Naoki Terada⁴), Naohiro Fujimoto⁵), and Masatoshi Eto¹)

Abstract:

The therapeutic landscape of pharmacotherapy for prostate cancer has dramatically evolved, and multiple therapeutic options have become available for prostate cancer patients. Therefore, useful biomarkers to identify suitable candidates for treatment are required to maximize the efficacy of pharmacotherapy. Genetic polymorphisms such as single-nucleotide polymorphisms (SNPs) and tandem repeats have been shown to influence the therapeutic effects of pharmacotherapy for prostate cancer patients. For example, genetic polymorphisms in the genes involved in androgen receptor signaling are reported to be associated with the therapeutic outcome of androgen-deprivation therapy as well as androgen receptor-pathway inhibitors. In addition, SNPs in genes involved in drug metabolism and efflux pumps are associated with therapeutic effects of taxane chemotherapy. Thus, genetic polymorphisms such as SNPs are promising biomarkers to realize personalized medicine. Here, we overview the current findings on the influence of genetic polymorphisms on the outcome of pharmacotherapy for prostate cancer and discuss current issues as well as future visions in this field.

Key Words:

androgen metabolism, androgen receptor, genetic polymorphism, pharmacotherapy, prostate cancer

Introduction

Androgen-deprivation therapy (ADT) with or without firstgeneration anti-androgen agents has been the gold standard as primary pharmacotherapy for treatment-naïve prostate cancer⁽¹⁾. Recently, the therapeutic landscape of pharmacotherapy for prostate cancer patients has been greatly evolving. Secondgeneration anti-androgen agents such as enzalutamide, apalutamide, and darolutamide as well as the CYP17 inhibitor abiraterone have been developed for castration-resistant prostate cancer (CRPC) ⁽²⁾. Although these drugs were initially developed for the treatment of CRPC, enzalutamide, apalutamide, and abiraterone have expanded for use in hormone-sensitive prostate cancer (HSPC) ⁽³⁾. In addition, taxane chemotherapy (docetaxel and cabazitaxel) and radioisotopes (radium-223) have been applied for the treatment of CRPC, and docetaxel has been indicated for HSPC ⁽²⁾. Thus, multiple therapeutic options for CRPC and HSPC are available. Therefore, useful biomarkers to identify patients that are suitable candidates for these treatments are required to maximize the efficacy of pharmacotherapy.

Genetic polymorphisms are considered one of the most

promising biomarkers for the realization of personalized medicine ⁽⁴⁾. Genetic polymorphisms are inter-individual differences in germline DNA and defined as differences in genomic sequences between individuals that occur at a frequency of 1% or more in a population. Most genetic polymorphisms are single-nucleotide polymorphisms (SNPs), and polymorphisms are also detected in repeated sequences such as microsatellites. SNPs are observed at a frequency of ~1 in 1000 nucleotides, and more than 2 million SNPs exist in the entire human genome. SNPs are classified into the following types according to their function: regulatory SNPs (rSNPs), which are located in promoter regions; coding SNPs (cSNPs), which are located in exons and cause an amino acid substitution; silent SNPs (sSNPs), which are located in an exon but do not cause an amino acid substitution; intron SNPs (iSNPs), which are located in introns; and genome SNPs (gSNPs), which are located in intergenic regions (Figure 1). Accordingly, rSNPs and cSNPs are likely to change gene expression and protein function, which results in functional and phenotypic differences, respectively. In addition, sSNPs and iSNPs may affect expression levels of genes. Conversely, gSNPs are speculated to not play a direct functional role, but these may serve as genomic

¹⁾Department of Urology, Graduate School of Medical Sciences, Kyushu University, Fukuoka, Japan. ²⁾Department of Urology, Graduate School of Medicine, Kyoto University, Kyoto, Japan. ³⁾Department of Urology, Graduate School of Medicine, Akita University, Akita, Japan. ⁴⁾Department of Urology, Faculty of Medicine, University of Miyazaki, Miyazaki, Japan. ⁵⁾Department of Urology, School of Medicine, University of Occupational and Environmental Health, Kitakyushu, Japan

Corresponding author: Masaki Shiota, shiota@uro.med.kyushu-u.ac.jp

JMA J. 2021;4(2):99-111

Received: January 12, 2021 / Accepted: January 15, 2021 / Advance Publication: March 26, 2021 / Published: April 15, 2021 Copyright © Japan Medical Association



Figure 1. Schematic of single-nucleotide polymorphism (SNP) types according to the location and function. UTR, untranslated region.

markers linked with distinct functional SNPs.

Genetic polymorphisms can cause various phenotypic differences through changes of expression and/or activity in the corresponding gene. Genetic polymorphisms are also associated not only with disease susceptibility but also with treatment outcomes. For example, a genetic polymorphism in UGT1A1 (UGT1A1*28 and UGT1A1*6), which encodes UDP-glucuronosyltransferase, decreases enzyme activity, and delays the metabolism of SN-38, the active metabolite of irinotecan, which results in a higher incidence of adverse events by irinotecan ⁽⁵⁾. A test for genetic polymorphisms in UGT1A1 has been approved in Japan for patients who will be treated with irinotecan chemotherapy. A SNP in *Nudix hydrolase 15 (NUDT15)*, which encodes the enzyme involved in the metabolism of thiopurines, was shown to be useful in predicting adverse events of thiopurines. A test for genetic polymorphisms in *NUDT15* was recently approved in Japan for patients who will be treated with thiopurines ⁽⁶⁾. Thus, the significance of testing genetic polymorphisms including SNPs in medical care has been growing.

Several genome-wide associated studies (GWASs) on prostate cancer susceptibility in large cohorts have been reported, showing the value of hundreds of SNPs with prostate cancer incidence ^{(7), (8)}. In addition, various studies have reported the significance of SNPs in the outcome of pharmacotherapy for prostate cancer ⁽⁹⁾. An association of genetic background such as race and family history with the outcome of prostate cancer has been shown, which suggests that genetic factors play an important role in pharmacotherapy ^{(10), (11)}. In this review, we provide an overview of the current findings on the influence of genetic polymorphisms in pharmacotherapy for prostate cancer and discuss current issues and future directions in this field.

Genetic Polymorphisms and Primary ADT for HSPC

Aberrant activation of androgen receptor (AR) signaling is one of the main causes by which prostate cancer acquires castration resistance. Therefore, polymorphisms in genes related to the AR pathway may affect the therapeutic efficacy of primary ADT through influencing AR signaling activity ⁽⁹⁾. To date, 63 SNPs in 49 genes have been reported to be associated with the outcome of primary ADT for HSPC (**Table 1**).

De novo androgen synthesis in prostate cancer cells is a major source of androgen under castrated condition during ADT and is shown to play an important role in the progression to CRPC ⁽⁵⁹⁾. Multiple studies have indicated the association of SNPs in genes involved in androgen metabolism, including CYP17A1, CYP19A1, HSD3B1, HSD17B2, HSD17B3, HSD17B4, AKR1C3, and SRD5A2, with the outcome of ADT (Figure 2). For example, a cSNP (rs1047303, 1245A>C, N367T) in *HSD3B1*, which encodes 3β -hydroxysteroid dehydrogenase 1 (3 β -HSD1), results in a variant of 3 β -HSD1 with high activity, and the prognosis of carriers of this variant is poor ^{(18), (19), (20), (21), (22), (60)}. The prognostic impact of the cSNP (rs1047303) in HSD3B1 in the United States was validated in an Asian cohort ⁽²¹⁾, although variant carriers were rare in Asian patients (~15%) compared with Caucasian patients (~50%) (Table 2). The prognostic impact of the cSNP (rs1047303) in HSD3B1 was validated in primary ADT plus docetaxel for HSPC (22). In addition, the prognostic difference by another SNP (rs1856888) in HSD3B1 was also indicated. Ross et al. initially reported that the variant G allele in rs1856888 was associated with a low risk of disease progression among men in the United States (15); however, a recent study from the United States showed poor prognosis in patients carrying the variant G allele in rs1856888⁽²³⁾. Because of the strong linkage disequilibrium between the SNPs (rs1047303 and rs1856888) in HSD3B1⁽²³⁾, the variant allele in the SNPs (rs1047303 and rs1856888) in HSD3B1 is likely to be associated with poor prognosis in patients treated with primary ADT. In a study on an iSNP (rs1870050) in CYP19A1, Ross et al. reported that the variant C allele in rs1870050 was associated with a high risk of disease progression among men in the United States ⁽¹⁵⁾. However, two recent studies showed a low risk of progression and better prognosis among Asian men with the variant C allele in rs1870050 $^{\scriptscriptstyle (12),\,(16)}$. In addition, the prognostic significance of an rSNP (rs743572) in the 5' untranslated region of CYP17A1 has been shown ^{(13), (14)}.

In addition to enzymes for androgen metabolism, the pump for androgens such as dehydroepiandrosterone (DHEA) and testosterone also plays a key role in the development of CRPC ⁽²⁹⁾. SNPs in *SLCO1B3* and *SLCO2B1* genes,

Table 1. Genetic Polymorphisms Associated with Treatment Outcomes for Hormone-Sensitive Prostate Canc

Gene name	Function	rs number	Polymorphism types	Treatment	Validation	Reference
CYP17A1	Androgen metabolism	rs6162	sSNP	ADT		(12)
		rs743572	rSNP	ADT	Validated	(13), (14)
CYP19A1	Androgen metabolism	rs1870050	iSNP	ADT	Almost validated	(12), (15), (16)
		rs4775936	iSNP	ADT		(17)
HSD3B1		rs1047303	cSNP	ADT	Validated	(18)-(22)
				ADT+Docetaxel		(22)
		rs1856888	gSNP	ADT	Almost validated	(15), (23)
HSD17B2	Androgen metabolism	rs4243229, rs7201637	iSNP	ADT		(12)
HSD17B3	Androgen metabolism	rs2257157	iSNP	ADT		(12)
HSD17B4	Androgen metabolism	rs7737181	iSNP	ADT		(15)
AKR1C3	Androgen metabolism	rs12529	cSNP	ADT	Controversial	(24), (25)
SRD5A2	Androgen metabolism	rs523349	cSNP	ADT		(26)
SLCO1B3	Androgen transporter	rs4149117	cSNP	ADT	Validated	(27)-(29)
SLCO2B1	Androgen transporter	rs1077858	iSNP	ADT	Validated	(30), (31)
		rs1789693	iSNP	ADT		(30)
		rs12422149	cSNP	ADT	Almost validated	(29)-(32)
GNRH2	Androgen synthesis	rs6051545	cSNP	ADT		(33)
SHBG	Androgen-binding protein	rs6259	cSNP	ADT	Controversial	(34), (35)
AR	Steroid receptor	CAG repeat	Coding region	ADT	Almost validated	(24), (36), (37)
ESR1	Steroid receptor	rs1062577	rSNP	ADT		(12)
NR3C2	Steroid receptor	rs5522	cSNP	ADT		(38)
YB-1	Transcription factor	rs12030724	iSNP	ADT	Validated	(39), (40)
HIF1A	Transcription factor	rs11549465	cSNP	ADT		(41)
ARRDC3	Target gene of AR	rs2939244	rSNP	ADT		(42)
FLT1	Target gene of AR	rs9508016	rSNP	ADT		(42)
SKAP1	Target gene of AR	rs6054145	rSNP	ADT		(42)
FBXO32	Target gene of AR	rs7830622	rSNP	ADT		(42)
BNC2	Target gene of ER	rs16934641	rSNP	ADT		(43)
TACC2	Target gene of ER	rs3763763	rSNP	ADT		(43)
ALPK1	Target gene of ER	rs2051778	rSNP	ADT		(43)
LSAMP	Target gene of NFĸB	rs13088089	rSNP	ADT		(44)
CCL17	Target gene of NFĸB	rs223899	rSNP	ADT		(44)
PSMD7	Target gene of NFĸB	rs2387084	rSNP	ADT		(44)
MON1B	Target gene of NFĸB	rs284924	rSNP	ADT		(44)
GSTM3	Antioxidant	rs7483	cSNP	ADT	Validated	(45)
CAT	Antioxidant	rs564250	gSNP	ADT		(45)
SLC28A3	Nucleoside transporter	rs56350726	cSNP	ADT		(46)
LRP2	Sterol and steroid transporter	rs6433107, rs3944004, rs830994, rs3770613, rs831003	iSNP	ADT		(47)
EGF	Growth factor	rs4444903	rSNP	ADT		(48)
IRS2	Growth factor	rs7986346	gSNP	ADT		(49)

(Table continued on next page)

Gene name	Function	rs number	Polymorphism types	Treatment	Validation	Reference
TGFBR2	TGF-β signaling	rs3087465	iSNP	ADT		(50)
BMP5	TGF- β signaling	rs317027	gSNP	ADT		(49)
IL18	Cytokine	rs187238	rSNP	ADT		(51)
APC	Wnt signaling	rs2707765, rs497844	iSNP	ADT		(52)
BGLAP	Bone metabolism	rs1800247	rSNP	ADT		(53)
EDN1	Vasoconstrictor	rs1800541, rs2070699	iSNP	ADT		(54)
CASP3	Apoptosis	rs4862396	gSNP	ADT		(49)
TRMT11	Methyltransferase	rs1268121, rs6900796	iSNP	ADT		(55)
COMT	Methyltransferase	rs4680	cSNP	Estramustine phosphate		(56)
KIF3C	miRNA target site	rs6728684	rSNP	ADT		(57)
CDON	miRNA target site	rs3737336	rSNP	ADT		(57)
IFI30	miRNA target site	rs1045747	rSNP	ADT		(57)
PALLD	miRNA target site	rs1071738	rSNP	ADT		(57)
GABRA1	miRNA target site	rs998754	rSNP	ADT		(57)
SYT9	miRNA target site	rs4351800	rSNP	ADT		(57)
-	-	rs16901979, rs7931342	gSNP	ADT		(58)

Table 1. Continued.

ADT, androgen deprivation therapy; AR, androgen receptor; ER, estrogen receptor; NFKB, nuclear factor-K B; SNP, single-nucleotide polymorphism; TGF, tumor growth factor

which encode proteins responsible for the import of testosterone and DHEA, respectively, were reported to be associated with the prognosis of patients treated with primary ADT (27), (28), (29), (30), (31), (32). Higher testosterone uptake in patients with the variant allele of the cSNP (rs4149117, 334G>T, A112S) in SLCO1B3 was shown, and a causal variant of SLCO1B3 was reported to be associated with poor prognosis (27), (28), (29). Several studies demonstrated that the cSNP (rs12422149, 935G>A, R312Q) in SLCO2B1 resulted in higher activity of DHEA-sulfate uptake and the wild-type allele in SLCO2B1 (rs12422149) was associated with early recurrence and poor prognosis after primary ADT ^{(29), (30), (31), (32)}. Another SNP (rs1077858) in SLCO2B1 was also associated with prognosis (30), (31). Thus, SNPs in the genes involved in androgen metabolism and uptake in prostate cancer cells play a key role in the progression of prostate cancer through persistent androgen synthesis in prostate cancer under castrated condition (**Figure 3**).

SNPs in other molecules related to the AR pathway were also shown to have prognostic impact after primary ADT. For example, the CAG repeat in *AR* correlated with prognosis, although null results were also reported ^{(24), (36), (37), (61)}. In addition, the iSNP (rs12030724) in *YB-1* that regulates YB-1 expression, which results in AR and AR variant expression, was also associated with the prognosis of Japanese men with advanced prostate cancer treated with primary ADT ^{(39), (40)}. The cSNP (rs7483, I224V) in *GSTM3*, which encodes an antioxidant enzyme, was also reported to be prognostic in Japanese patients with nonmetastatic and advanced prostate cancer treated with primary ADT $^{\scriptscriptstyle (45)}$.

Genetic Polymorphisms and Treatment with Novel AR-pathway Inhibitors (ARPIs) for CRPC

Novel ARPIs such as enzalutamide, apalutamide darolutamide, and abiraterone have been demonstrate to improve survival in patients with CRPC (2). Because abiraterone is taken up into cells by OATP2B1, which is encoded by SLCO2B1, and then metabolized by 3β -HSD and 5α -reductase, the therapeutic effect of abiraterone treatment may depend on the activities of the molecules involved in androgen metabolism and uptake (Figure 2) (73), (78). Recent reports showed that SNPs in genes involved in androgen metabolism and transport such as CYP17A1, HSD3B1, SRD5A2, and SLCO2B1 correlate with the outcome of abiraterone treatment (Table 3). An rSNP (rs2486758, -362T>C) in CYP17A1 was associated with prognosis after abiraterone treatment (68), (69). In addition, variant carriers of the cSNP (rs1047303) in HSD3B1 showed poor prognosis after treatment with ARPI (65), (66). The prognostic impact of the cSNP (rs1047303) in HSD3B1 for both primary ADT for HSPC as well as ARPIs for CRPC may be because of hyperactive androgen synthesis in variant carriers. The variant allele in HSD3B1 is expected to lead to increased conversion from abiraterone to the more potent delta-4-abiraterone (78). Accordingly, the cSNP (rs1047303) in HSD3B1 was



Figure 2. Gene function of single-nucleotide polymorphisms (SNPs) associated with therapeutic effects and adverse events of drug therapy. Underlined organs and treatments in parentheses mean target organ and treatment in which the gene function of SNPs is involved, respectively. ADT, androgen-deprivation therapy; ARPI, androgen receptor-pathway inhibitor.

Outcome	Variant carrier	Number	Frequency carrying a variant allele	Reference
Prostate cancer susceptibility	High	626	48% (AC/CC, US)	(62)
Hereditary prostate cancer susceptibility	High	98	53% (AC/CC, US)	(63)
Prognosis in primary ADT	Poor	118/137/118	51% (AC/CC, US)	(18)
Prognosis in primary ADT	Poor	102	53% (AC/CC, US)	(19)
Prognosis in primary ADT	Poor	218	54% (AC/CC, US)	(20)
Prognosis in Abiraterone	Insignificant	76	45% (AC/CC, US)	(64)
Progression in primary ADT or ADT+Docetaxel	Poor in low volume	475	53% (AC/CC, US)	(22)
Prognosis in Ezalutamide or Abiraterone	Poor	266	8% (CC, US/UK)	(65)
Prognosis in Ezalutamide or Abiraterone	Poor	547	15% (CC, Canada/Europe)	(66)
Prognosis in primary ADT	Insignificant	103	18% (AC/CC, China)	(67)
Prognosis in primary ADT	Poor	104	9% (AC/CC, Japan)	(21)
Prognosis in Abiraterone	Favorable	99	14% (AC/CC, Japan)	(21)

Table 2. Outcome and Frequencies of the rs1047303 Variant Allele of HSD3B1.

ADT, androgen deprivation therapy; UK, United Kingdom; US, United States



Figure 3. Schematic of molecules involved in androgen synthesis and uptake. The metabolisms surrounded by red, light blue, and blue are mainly processed in adrenal glands, prostate cancer, and both, respectively. OATP2B1 uptakes DHEA into prostate cancer cells. DHEA, dehydroepiandrosterone; DHT, dihydrotestosterone.

Table 3. Genetic Polymorphisms Associated with the Prognosis of Patients with Castration-Resistant Prostate Cancer Treatedwith Androgen Receptor-Pathway Inhibitors.

Gene name	Function	rs number	Polymorphism types	Treatment	Validation	Reference
CYP17A1	Androgen metabolism	rs2486758	rSNP	Abiraterone	Validated	(68), (69)
		rs10883783	iSNP	Abiraterone		(70)
HSD3B1	Androgen metabolism	rs1047303	cSNP	Abiraterone		(21)
				Abiraterone or Enzalutamide	Validated	(65), (66)
SRD5A2	Androgen metabolism	rs523349	cSNP	Abiraterone		(71)
SLCO2B1	Androgen transporter	rs1077858, rs1789693, rs34550074	iSNP, iSNP, cSNP	Abiraterone		(72)
		rs12422149	cSNP	Abiraterone		(73)
YB-1	Androgen receptor regulator	rs10493112	iSNP	Abiraterone		(74)
CYB5A	CYP17A1 activity regulator	rs1790834	iSNP	Abiraterone		(75)
TSPYL1	CYP17A1 and CYP3A4 regulator	rs3828743	cSNP	Abiraterone		(76)
011171171	The state	Group 1 (rs3775777, rs4149534, rs10019305)	CNID	Abiratoropo		(77)
JULIILI	Estrogen inclabolisiii	Group 2 (rs3775770, rs4149527, rs3775768)	IJINE	Abilatelone		

SNP, single-nucleotide polymorphism

shown to be associated with comparable or better treatment efficacy of abiraterone ${}^{(21),(64)}$.

Interestingly, several genes are overlapping in association with the prognosis between primary ADT and ARPIs, which both target the AR pathway (**Table 4**). SNPs in *CYP17A1* and *YB-1* are associated with the outcome of primary ADT and ARPIs, although the SNPs in each gene are different. Furthermore, the cSNP (rs1047303) in *HSD3B1*, cSNP (rs523349) in *SRD5A2*, and SNPs (rs1077858, rs1789693,

and rs12422149) in *SLCO2B1* were shown to be common prognosticators in both primary ADT for HSPC and ARPIs for CRPC. The prognostic and antitumor impacts of the cSNP (rs523349) in *SRD5A2* and SNPs (rs1077858, rs1789693, and rs12422149) in *SLCO2B1* were consistent between primary ADT and abiraterone. Intriguingly, a variant allele in *HSD3B1* (rs1047303) was differentially associated with prognosis in patients treated with abiraterone and other therapies. These findings suggest that *HSD3B1* (rs1047303) may be a promising marker to select appropriate combination therapy with ADT. Further studies on the prognostic impact of these SNPs will be important to evaluate candidates for personalized medicine.

Genetic Polymorphisms and Taxane Treatment for CRPC

The taxane docetaxel is widely used not only for prostate cancer but also for various cancers such as lung, uterine, and ovarian cancers. Many reports have demonstrated the relationship between genetic polymorphisms and the efficacy and adverse events of docetaxel therapy. Previous studies, including several in prostate cancer, reported associations between drug transport genes (ABCB1, ABCC2, ABCG1, ABCG2, SLCO1B3) or drug metabolism genes (CYP1B1, CYP2C8, CYP3A4, CYP3A5) with therapeutic efficacy or adverse events (Figure 2)⁽⁸¹⁾. As shown in Table 5, the cSNP (rs1056836, 4326C>G, L432V) in CYP1B1 was associated with poor response and prognosis (82), (83). In addition, SNPs in estrogen re*ceptor 1 (ESR1)* were also associated with treatment efficacy in prostate cancer⁽⁸⁰⁾. SNPs in *ESR1* were reported to be associated with the outcome of primary ADT and taxane chemotherapy although the position of SNPs in ESR1 is different (Table 4). Then, SNPs in ESR1 may serve as a predictive marker for taxane chemotherapy. OATP1B3, which is encoded by SLCO1B3, plays a role in taxane uptake into cells and is involved in taxane resistance in prostate cancer cells. SNPs in SLCO1B3 may be associated with the treatment efficacy of taxane (88). However, a recent study showed comparable prognosis after cabazitaxel for CRPC between genotypes in SLCO1B3 (rs4149117) (79). Because prognostic impact in primary ADT has been shown^{(27), (28), (29)}, the cSNP (rs4149117) in SLCO3B1 may serve as a predictive marker in pharmacotherapy for prostate cancer.

Current Research Issues and Future Prospects for Personalized Medicine

The associations between multiple SNPs and therapeutic effects of pharmacotherapy for prostate cancer have been reported, as described above. However, to date, no genetic marker has been clinically utilized in pharmacotherapy for prostate cancer, which suggests potential issues as described in the following. While some SNPs have been reproducible in validation studies, others have not yielded consistent results across studies (**Table 1, 3, 5**). This may be because of racial differences in the frequency of genetic polymorphisms and linkage disequilibrium (a phenomenon in which there is a correlation between genetic polymorphisms in a population). To resolve this issue, multiple studies with large populations and meta-analysis studies are required. In addition, advances in technology such as artificial intelligence may serve as a breakthrough method to resolve the complex linkage disequilibrium among

individuals.

Another problem is that the data in most study cohorts were retrospectively collected in daily practice. A daily clinical follow-up generally shows deviations from the strict follow-up schedule in a clinical trial. To improve the quality of data, collecting clinical data using a strict protocol is desirable to obtain more robust findings. In addition, most studies to date have focused on target genetic polymorphisms of individual genes. Because this method may miss useful SNPs, comprehensive methods such as GWAS are required. In addition, a single marker may be not enough for accurate predictive ability, and this may be overcome by using multiple SNPs. GWASs indicated that a single SNP generally provides only a modest (odds ratio, 1.1-1.5) increased susceptibility risk of prostate cancer, where polygenic risk score (PGS) using multiple risk SNPs was developed and validated (89), (90). Therefore, the PGS approach would be useful to increase diagnostic ability.

Furthermore, the genes of SNPs associated with therapeutic outcome can be the cause of treatment resistance. Therefore, these genes are promising targets to overcome treatment resistance. Genes involved in androgen metabolism such as *CYP17A1*, *HSD3B1*, *AKR1C3*, and *SRD5A2* have been candidate targets for drug discovery and drug development, and the SNPs may be crucial in therapeutic efficacy (**Table 6**).

Conclusion

Here, we summarized the known associations between genetic polymorphisms and the outcomes of pharmacotherapy in prostate cancer patients. Recently, multiple novel therapeutic options for HSPC have emerged, and the stratification of suitable patients for each option will be required. Genetic biomarkers such as SNPs will be beneficial for stratifying patients and for estimating the treatment response of an individual patient. The combination of genetic biomarkers with traditional clinicopathological parameters could improve the prognostication and the choice of the most appropriate treatment for each patient, which will be helpful in clinical decision making. Thus, personalized medicine using genetic biomarkers is expected to be realized in pharmacotherapy for prostate cancer. However, unresolved issues remain, such as inconsistent results among studies as well as the current lack of GWAS and PGS approaches, and these issues should be addressed in future research.

Article Information

This article is based on the study, which received the Medical Research Encouragement Prize of The Japan Medical Association in 2020.

Conflicts of Interest

Masaki Shiota received honoraria from Janssen Pharmaceutical Company, Astellas Pharma, and Sanofi; Shusuke Akamat-

Gene name	rs number	Treatment regimen	Risk allele	Outcome	Reference
CYP17A1	rs6162	ADT	G	OS	(12)
	rs743572	ADT	А	OS	(13)
		ADT	А	PFS	(14)
	rs2486758	Abiraterone	С	PFS	(68)
		Abiraterone	С	PFS	(69)
	rs10883783	Abiraterone	А	PFS	(70)
HSD3B1	rs1047303	ADT	С	PFS, MFS, OS	(18)
		ADT	С	PFS	(19)
		ADT	С	MFS	(20)
		ADT	С	PFS	(21)
		ADT	С	PFS, OS	(22)
		ADT+Docetaxel	С	PFS, OS	(22)
		Abiraterone	Null	PFS	(64)
		Abiraterone	А	PFS, OS	(21)
		Abiraterone/Enzalutamide	С	OS	(65)
		Abiraterone/Enzalutamide	С	PFS	(66)
	rs1856888	ADT	А	PFS	(15)
		ADT	G	OS	(23)
SRD5A2	rs523349	ADT	G	PFS, OS	(26)
		Abiraterone	G	PFS	(71)
SLCO1B3	rs4149117	ADT	Т	OS	(27)
			Т	PFS	(28)
			Т	CSS	(29)
		Cabazitaxel	Null	OS	(79)
SLCO2B1	rs1077858	ADT	G	PFS	(30)
		ADT	G	OS	(31)
		Abiraterone	G	MRD	(72)
	rs1789693	ADT	Т	PFS	(30)
		Abiraterone	Т	MRD	(72)
	rs34550074	Abiraterone	Т	MRD	(72)
	rs12422149	ADT	А	CSS	(29)
		ADT	G	PFS	(30)
		ADT	G	PFS	(32)
		ADT	G	PFS	(31)
		Abiraterone	G	PFS	(73)
YB-1	rs10493112	Abiraterone	А	PFS	(74)
	rs12030724	ADT	А	PFS	(39)
		ADT	А	PFS, OS	(40)
ESR1	rs1062577	ADT	А	OS	(12)
	rs2234693	Docetaxel	С	PFS	(80)
	rs9340799	Docetaxel+Thalidomide	G	PFS	(80)

Table 4. Genetic Polymorphisms Associated with the Outcomes of Multiple Treatments.

ADT, androgen deprivation therapy; MRD, minimal residual disease; OS, overall survival; PFS, progression-free survival

Gene name	Function	rs number	Polymorphism types	Treatment	Validation	Reference
CYP1B1	Drug metabolizing enzyme	rs1056836	cSNP	Docetaxel	Validated	(82), (83)
ABCB1	Drug excretion pump	rs1128503, rs2032582, rs1045642	cSNP	Docetaxel+Thalidomide		(84)
ABCB11	Drug excretion pump	rs7602171	iSNP	Docetaxel+Thalidomide		(85)
ABCG2	Drug excretion pump	rs2231142	cSNP	Docetaxel+Vinorelbine/Estramustine phosphate		(86)
ESR1	Steroid receptor	rs2234693, rs9340799	iSNP	Docetaxel+Thalidomide		(80)
GSTP1	Antioxidant	rs1138272	cSNP	Docetaxel+Thalidomide		(85)
SLC5A6	Transporter	rs1395	cSNP	Docetaxel+Thalidomide		(85)
VEGFA	Angiogenesis	rs1570360	rSNP	Docetaxel, Celecoxib +Cyclophosphamide		(87)

Table 5. Genetic Polymorphisms Associated with the Outcome of Taxane Treatment for Castration-Resistant Prostate Cancer.

SNP, single-nucleotide polymorphism

Table 6. Druggable	Targets in Androgen	Metabolism and	Their Inhibitors.
()()	() ()		

Target enzyme	Inhibitor	Developmental status
CYP17	Abiraterone	Approved
	Orteronel (TAK-700)	Phase III (terminated)
	Galeterone	Phase II (terminated)
3β-HSD	Trilostane	Phase II (terminated)/on market for Cushing's syndrome
AKR1C3	Indometacin	Phase II/on market as NSAIDs
	N-(indolylcarbonyl)-piperidines	Phase I
5α-reductase (types I and II)	Dutasteride	Phase II (terminated)/on market for benign prostatic hyperplasia
5α-reductase (type II)	Finasteride	On market for androgenetic alopecia

NSAID, non-steroidal anti-inflammatory drug

su received grant/research support from Astellas Pharma; Shintaro Narita received honoraria from Janssen Pharmaceutical Company; Naohiro Fujimoto received honoraria from Janssen Pharmaceutical Company and Astellas Pharma and grant/research support from Astellas Pharma and Sanofi; Masatoshi Eto received honoraria from Janssen Pharmaceutical Company and grant/research support from Astellas Pharma and Sanofi.

Sources of Funding

This work was supported by Takeda Science Foundation and Japanese Urological Association to Masaki Shiota.

Acknowledgement

We thank Gabrielle White Wolf, PhD, from Edanz Group (https://en-author-services.edanz.com/ac) for editing a draft of this manuscript.

References

and future perspectives toward upfront therapy for metastatic hormone-sensitive prostate cancer. Int J Urol. 2016;23(5):360-9.

- 2. Moussa M, Papatsoris A, Abou Chakra M, et al. Pharmacotherapeutic strategies for castrate-resistant prostate cancer. Expert Opin Pharmacother. 2020;21(12):1431-48.
- Sathianathen NJ, Koschel S, Thangasamy IA, et al. Indirect comparisons of efficacy between combination approaches in metastatic hormone-sensitive prostate cancer: a systematic review and network meta-analysis. Eur Urol. 2020;77(3):365-72.
- Choudhury AD, Eeles R, Freedland SJ, et al. The role of genetic markers in the management of prostate cancer. Eur Urol. 2012;62(4):577-87.
- Hulshof EC, Deenen MJ, Guchelaar HJ, et al. Pre-therapeutic UGT1A1 genotyping to reduce the risk of irinotecan-induced severe toxicity: ready for prime time. Eur J Cancer. 2020;141:9-20.
- 6. Kakuta Y, Kawai Y, Okamoto D, et al. NUDT15 codon 139 is the best pharmacogenetic marker for predicting thiopurine-

^{1.} Shiota M, Eto M. Current status of primary pharmacotherapy

induced severe adverse events in Japanese patients with inflammatory bowel disease: a multicenter study. J Gastroenterol. 2018;53(9):1065-78.

- 7. Akamatsu S, Takata R, Haiman CA, et al. Common variants at 11q12, 10q26 and 3p11.2 are associated with prostate cancer susceptibility in Japanese. Nat Genet. 2012;44(4):426-9.
- Benafif S, Kote-Jarai Z, Eeles RA; PRACTICAL Consortium. A review of prostate cancer genome-wide association studies (GWAS). Cancer Epidemiol Biomarkers Prev. 2018;27(8):845-57.
- 9. Fujimoto N, Shiota M, Tomisaki I, et al. Gene polymorphismrelated individual and interracial differences in the outcomes of androgen deprivation therapy for prostate cancer. Clin Genitourin Cancer. 2017;15(3):337-42.
- Fukagai T, Namiki TS, Carlile RG, et al. Comparison of the clinical outcome after hormonal therapy for prostate cancer between Japanese and Caucasian men. BJU Int. 2006;97(6):1190-3.
- Hemminki K, Ji J, Försti A, et al. Concordance of survival in family members with prostate cancer. J Clin Oncol. 2008;26(10):1705-9.
- Lévesque É, Huang SP, Audet-Walsh É, et al. Molecular markers in key steroidogenic pathways, circulating steroid levels, and prostate cancer progression. Clin Cancer Res. 2013;19(3):699-709.
- Hamada A, Danesi R, Price DK, et al. Association of a CYP17 polymorphism with overall survival in Caucasian patients with androgen-independent prostate cancer. Urology. 2007;70(2):217-20.
- 14. Yamada T, Nakayama M, Shimizu T, et al. Genetic polymorphisms of CYP17A1 in steroidogenesis pathway are associated with risk of progression to castration-resistant prostate cancer in Japanese men receiving androgen deprivation therapy. Int J Clin Oncol. 2013;18(4):711-7.
- Ross RW, Oh WK, Xie W, et al. Inherited variation in the androgen pathway is associated with the efficacy of androgendeprivation therapy in men with prostate cancer. J Clin Oncol. 2008;26(6):842-7.
- Shiota M, Fujimoto N, Tsukahara S, et al. The impact of genetic polymorphism on CYP19A1 in androgen-deprivation therapy among Japanese men. Cancer Chemother Pharmacol. 2019;83(5):933-8.
- 17. Kanda S, Tsuchiya N, Narita S, et al. Effects of functional genetic polymorphisms in the CYP19A1 gene on prostate cancer risk and survival. Int J Cancer. 2015;136(1):74-82.
- Hearn JWD, AbuAli G, Reichard CA, et al. HSD3B1 and resistance to androgen-deprivation therapy in prostate cancer: a retrospective, multicohort study. Lancet Oncol. 2016;17(10):1435-44.
- Agarwal N, Hahn AW, Gill DM, et al. Independent validation of effect of HSD3B1 genotype on response to androgendeprivation therapy in prostate cancer. JAMA Oncol. 2017;3(6):856-7.
- 20. Hearn JWD, Xie W, Nakabayashi M, et al. Association of

HSD3B1 genotype with response to androgen-deprivation therapy for biochemical recurrence after radiotherapy for localized prostate cancer. JAMA Oncol. 2018;4(4):558-62.

- 21. Shiota M, Narita S, Akamatsu S, et al. Association of missense polymorphism in HSD3B1 with outcomes among men with prostate cancer treated with androgen-deprivation therapy or abiraterone. JAMA Netw Open. 2019;2(2):e190115.
- 22. Hearn JWD, Sweeney CJ, Almassi N, et al. HSD3B1 genotype and clinical outcomes in metastatic castration-sensitive prostate cancer. JAMA Oncol. 2020;6(4):e196496.
- 23. Chen WS, Feng EL, Aggarwal R, et al. Germline polymorphisms associated with impaired survival outcomes and somatic tumor alterations in advanced prostate cancer. Prostate Cancer Prostatic Dis. 2020;23(2):316-23.
- Yu CC, Huang SP, Lee YC, et al. Molecular markers in sex hormone pathway genes associated with the efficacy of androgen-deprivation therapy for prostate cancer. PLoS One. 2013;8(1):e54627.
- 25. Shiota M, Endo S, Fujimoto N, et al. Polymorphisms in androgen metabolism genes with serum testosterone levels and prognosis in androgen-deprivation therapy. Urol Oncol. 2020;38(11):849.e11-8.
- 26. Shiota M, Fujimoto N, Yokomizo A, et al. SRD5A gene polymorphism in Japanese men predicts prognosis of metastatic prostate cancer with androgen-deprivation therapy. Eur J Cancer. 2015;51(14):1962-9.
- 27. Hamada A, Sissung T, Price DK, et al. Effect of SLCO1B3 haplotype on testosterone transport and clinical outcome in caucasian patients with androgen-independent prostatic cancer. Clin Cancer Res. 2008;14(11):3312-8.
- 28. Sharifi N, Hamada A, Sissung T, et al. A polymorphism in a transporter of testosterone is a determinant of androgen independence in prostate cancer. BJU Int. 2008;102(5):617-21.
- 29. Wright JL, Kwon EM, Ostrander EA, et al. Expression of SLCO transport genes in castration-resistant prostate cancer and impact of genetic variation in SLCO1B3 and SLCO2B1 on prostate cancer outcomes. Cancer Epidemiol Biomarkers Prev. 2011;20(4):619-27.
- Yang M, Xie W, Mostaghel E, et al. SLCO2B1 and SLCO1B3 may determine time to progression for patients receiving androgen deprivation therapy for prostate cancer. J Clin Oncol. 2011;29(18):2565-73.
- 31. Wang X, Harshman LC, Xie W, et al. Association of SLCO2B1 genotypes with time to progression and overall survival in patients receiving androgen-deprivation therapy for prostate cancer. J Clin Oncol. 2016;34(4):352-9.
- 32. Fujimoto N, Kubo T, Inatomi H, et al. Polymorphisms of the androgen transporting gene SLCO2B1 may influence the castration resistance of prostate cancer and the racial differences in response to androgen deprivation. Prostate Cancer Prostatic Dis. 2013;16(4):336-40.
- 33. Shiota M, Fujimoto N, Takeuchi A, et al. The association of polymorphisms in the gene encoding gonadotropin-releasing hormone with serum testosterone level during androgen

deprivation therapy and prognosis of metastatic prostate cancer. J Urol. 2018;199(3):734-40.

- 34. Monteiro C, Sousa MV, Ribeiro R, et al. Genetic variants in AR and SHBG and resistance to hormonal castration in prostate cancer. Med Oncol. 2013;30(1):490.
- 35. Shiota M, Fujimoto N, Tsukahara S, et al. Genetic polymorphism in sex hormone-binding globulin with a prognosis of androgen deprivation therapy in metastatic prostate cancer among Japanese men. Clin Genitourin Cancer. 2019;17(3):e387-93.
- 36. Bratt O, Borg A, Kristoffersson U, et al. CAG repeat length in the androgen receptor gene is related to age at diagnosis of prostate cancer and response to endocrine therapy, but not to prostate cancer risk. Br J Cancer. 1999;81(4):672-6.
- 37. Shimbo M, Suzuki H, Kamiya N, et al. CAG polymorphic repeat length in androgen receptor gene combined with pretreatment serum testosterone level as prognostic factor in patients with metastatic prostate cancer. Eur Urol. 2005;47(4):557-63.
- Shiota M, Fujimoto N, Imada K, et al. Prognostic impact of genetic polymorphism in mineralocorticoid receptor and comorbidity with hypertension in androgen-deprivation therapy. Front Oncol. 2018;8:635.
- Shiota M, Fujimoto N, Imada K, et al. Potential role for YB-1 in castration-resistant prostate cancer and resistance to enzalutamide through the androgen receptor V7. J Natl Cancer Inst. 2016;108(7):djw005.
- 40. Shiota M, Narita S, Habuchi T, et al. Validated prognostic significance of YB-1 genetic variation in metastatic prostate cancer. Pharmacogenomics J. 2021;21(1):102-5.
- 41. Fraga A, Ribeiro R, Príncipe P, et al. The HIF1A functional genetic polymorphism at locus +1772 associates with progression to metastatic prostate cancer and refractoriness to hormonal castration. Eur J Cancer. 2014;50(2):359-65.
- 42. Huang CN, Huang SP, Pao JB, et al. Genetic polymorphisms in androgen receptor-binding sites predict survival in prostate cancer patients receiving androgen-deprivation therapy. Ann Oncol. 2012;23(3):707-13.
- 43. Huang CN, Huang SP, Pao JB, et al. Genetic polymorphisms in oestrogen receptor-binding sites affect clinical outcomes in patients with prostate cancer receiving androgen-deprivation therapy. J Intern Med. 2012;271(5):499-509.
- Huang SP, Lin VC, Lee YC, et al. Genetic variants in nuclear factor-kappa B binding sites are associated with clinical outcomes in prostate cancer patients. Eur J Cancer. 2013;49(17):3729-37.
- 45. Shiota M, Fujimoto N, Itsumi M, et al. Gene polymorphisms in antioxidant enzymes correlate with the efficacy of androgendeprivation therapy for prostate cancer with implications of oxidative stress. Ann Oncol. 2017;28(3):569-75.
- 46. Jo JK, Oh JJ, Kim YT, et al. A genetic variant in SLC28A3, rs56350726, is associated with progression to castrationresistant prostate cancer in a Korean population with metastatic prostate cancer. Oncotarget. 2017;8(57):96893-902.

- 47. Holt SK, Karyadi DM, Kwon EM, et al. Association of megalin genetic polymorphisms with prostate cancer risk and prognosis. Clin Cancer Res. 2008;14(12):3823-31.
- Teixeira AL, Ribeiro R, Cardoso D, et al. Genetic polymorphism in EGF is associated with prostate cancer aggressiveness and progression-free interval in androgen blockade-treated patients. Clin Cancer Res. 2008;14(11):3367-71.
- Huang SP, Bao BY, Hour TC, et al. Genetic variants in CASP3, BMP5, and IRS2 genes may influence survival in prostate cancer patients receiving androgen-deprivation therapy. PLoS One. 2012;7(7):e41219.
- Teixeira AL, Gomes M, Nogueira A, et al. Improvement of a predictive model of castration-resistant prostate cancer: functional genetic variants in TGFβ1 signaling pathway modulation. PLoS One. 2013;8(8):e72419.
- Liu JM, Liu JN, Wei MT, et al. Effect of IL-18 gene promoter polymorphisms on prostate cancer occurrence and prognosis in Han Chinese population. Genet Mol Res. 2013;12(1):820-9.
- Geng JH, Lin VC, Yu CC, et al. Inherited variants in Wnt pathway genes influence outcomes of prostate cancer patients receiving androgen deprivation therapy. Int J Mol Sci. 2016;17(12):1970.
- 53. Wu HC, Lin CC, Chen WC, et al. Osteocalcin gene HindIII C/T polymorphism is a biomarker for prostate cancer and responsiveness to hormone therapy. Eur Urol. 2003;43(2):197-200.
- Xu D, Wang X, Lou Y. Association of endothelin-1 gene singlenucleotide polymorphisms and haplotypes with risk of hormone refractory prostate cancer. Pharmazie. 2017;72(2):103-6.
- 55. Kohli M, Riska SM, Mahoney DW, et al. Germline predictors of androgen deprivation therapy response in advanced prostate cancer. Mayo Clin Proc. 2012;87(3):240-6.
- 56. Suzuki M, Mamun MR, Hara K, et al. The Val158Met polymorphism of the catechol-O-methyltransferase gene is associated with the PSA-progression-free survival in prostate cancer patients treated with estramustine phosphate. Eur Urol. 2005;48(5):752-9.
- 57. Bao BY, Pao JB, Huang CN, et al. Polymorphisms inside microRNAs and microRNA target sites predict clinical outcomes in prostate cancer patients receiving androgendeprivation therapy. Clin Cancer Res. 2011;17(4):928-36.
- Bao BY, Pao JB, Huang CN, et al. Significant associations of prostate cancer susceptibility variants with survival in patients treated with androgen-deprivation therapy. Int J Cancer. 2012;130(4):876-84.
- 59. Feng Q, He B. Androgen receptor signaling in the development of castration-resistant prostate cancer. Front Oncol. 2019;9:858.
- 60. Chang KH, Li R, Kuri B, et al. A gain-of-function mutation in DHT synthesis in castration-resistant prostate cancer. Cell. 2013;154(5):1074-84.
- 61. Misra D, Xie W, Regan MM, et al. Germline CAG repeat length of the androgen receptor and time to progression in

patients with prostate cancer treated with androgen deprivation therapy. BJU Int. 2011;108(7):1086-91.

- 62. Chang BL, Zheng SL, Hawkins GA, et al. Joint effect of HSD3B1 and HSD3B2 genes is associated with hereditary and sporadic prostate cancer susceptibility. Cancer Res. 2002;62(6):1784-9.
- 63. Park JY, Tanner JP, Sellers TA, et al. Association between polymorphisms in HSD3B1 and UGT2B17 and prostate cancer risk. Urology. 2007;70(2):374-9.
- 64. Hahn AW, Gill DM, Nussenzveig RH, et al. Germline variant in HSD3B1 (1245 A > C) and response to abiraterone acetate plus prednisone in men with new-onset metastatic castrationresistant prostate cancer. Clin Genitourin Cancer. 2018;16(4):288-92.
- 65. Lu C, Terbuch A, Dolling D, et al. Treatment with abiraterone and enzalutamide does not overcome poor outcome from metastatic castration-resistant prostate cancer in men with the germline homozygous HSD3B1 c.1245C genotype. Ann Oncol. 2020;31(9):1178-85.
- 66. Khalaf DJ, Aragón IM, Annala M, et al. HSD3B1 (1245A>C) germline variant and clinical outcomes in metastatic castrationresistant prostate cancer patients treated with abiraterone and enzalutamide: results from two prospective studies. Ann Oncol. 2020;31(9):1186-97.
- 67. Wu G, Huang S, Nastiuk KL, et al. Variant allele of HSD3B1 increases progression to castration-resistant prostate cancer. Prostate. 2015;75(7):777-82.
- Binder M, Zhang BY, Hillman DW, et al. Common genetic variation in CYP17A1 and response to abiraterone acetate in patients with metastatic castration-resistant prostate cancer. Int J Mol Sci. 2016;17(7):1097.
- Crucitta S, Del Re M, Paolieri F, et al. CYP17A1 polymorphism c.-362T>C predicts clinical outcome in metastatic castration-resistance prostate cancer patients treated with abiraterone. Cancer Chemother Pharmacol. 2020;86(4):527-33.
- Salvi S, Casadio V, Burgio SL, et al. CYP17A1 polymorphisms and clinical outcome of castration-resistant prostate cancer patients treated with abiraterone. Int J Biol Markers. 2016;31(3):e264-9.
- 71. Shiota M, Akamatsu S, Narita S, et al. The association between missense polymorphisms in SRD5A2 and HSD3B1 and treatment failure with abiraterone for castration-resistant prostate cancer. Pharmacogenomics J. Forthcoming 2021.
- 72. Mostaghel EA, Cho E, Zhang A, et al. Association of tissue abiraterone levels and SLCO genotype with intraprostatic steroids and pathologic response in men with high-risk localized prostate cancer. Clin Cancer Res. 2017;23(16):4592-601.
- 73. Hahn AW, Gill DM, Poole A, et al. Germline variant in SLCO2B1 and response to abiraterone acetate plus prednisone (AA) in new-onset metastatic castration-resistant prostate cancer (mCRPC). Mol Cancer Ther. 2019;18(3):726-9.
- 74. Afonso A, Silva J, Lopes AR, et al. YB-1 variant and androgen receptor axis-targeted agents in metastatic castration-resistant

prostate cancer patients. Pharmacogenomics. 2020;21(13):919-28.

- 75. Wu X, Xu QJ, Chen PZ, et al. Association between CYP17A1, CYB5A polymorphisms and efficacy of abiraterone acetate/ prednisone treatment in castration-resistant prostate cancer patients. Pharmgenomics Pers Med. 2020;13:181-8.
- 76. Qin S, Liu D, Kohli M, et al. TSPYL family regulates CYP17A1 and CYP3A4 expression: potential mechanism contributing to abiraterone response in metastatic castration-resistant prostate cancer. Clin Pharmacol Ther. 2018;104(1):201-10.
- 77. Agarwal N, Alex AB, Farnham JM, et al. Inherited variants in SULT1E1 and response to abiraterone acetate by men with metastatic castration refractory prostate cancer. J Urol. 2016;196(4):1112-6.
- Li Z, Bishop AC, Alyamani M, et al. Conversion of abiraterone to D4A drives anti-tumour activity in prostate cancer. Nature. 2015;523(7560):347-51.
- 79. Belderbos BPS, de With M, Singh RK, et al. The influence of single-nucleotide polymorphisms on overall survival and toxicity in cabazitaxel-treated patients with metastatic castration-resistant prostate cancer. Cancer Chemother Pharmacol. 2020;85(3):547-53.
- 80. Sissung TM, Danesi R, Kirkland CT, et al. Estrogen receptor α and aromatase polymorphisms affect risk, prognosis, and therapeutic outcome in men with castration-resistant prostate cancer treated with docetaxel-based therapy. J Clin Endocrinol Metab. 2011;96(2):E368-72.
- Frederiks CN, Lam SW, Guchelaar HJ, et al. Genetic polymorphisms and paclitaxel- or docetaxel-induced toxicities: a systematic review. Cancer Treat Rev. 2015;41(10):935-50.
- Sissung TM, Danesi R, Price DK, et al. Association of the CYP1B1*3 allele with survival in patients with prostate cancer receiving docetaxel. Mol Cancer Ther. 2008;7(1):19-26.
- Pastina I, Giovannetti E, Chioni A, et al. Cytochrome 450 1B1 (CYP1B1) polymorphisms associated with response to docetaxel in castration-resistant prostate cancer (CRPC) patients. BMC Cancer. 2010;10:511.
- 84. Sissung TM, Baum CE, Deeken J, et al. ABCB1 genetic variation influences the toxicity and clinical outcome of patients with androgen-independent prostate cancer treated with docetaxel. Clin Cancer Res. 2008;14(14):4543-9.
- Sissung TM, Deeken J, Leibrand CR, et al. Identification of novel SNPs associated with risk and prognosis in patients with castration-resistant prostate cancer. Pharmacogenomics. 2016;17(18):1979-86.
- 86. Hahn NM, Marsh S, Fisher W, et al. Hoosier Oncology Group randomized phase II study of docetaxel, vinorelbine, and estramustine in combination in hormone-refractory prostate cancer with pharmacogenetic survival analysis. Clin Cancer Res. 2006;12(20 Pt 1):6094-9.
- 87. Derosa L, Galli L, Orlandi P, et al. Docetaxel plus oral metronomic cyclophosphamide: a phase II study with pharmacodynamic and pharmacogenetic analyses in castrationresistant prostate cancer patients. Cancer.

2014;120(24):3923-31.

- de Morrée ES, Böttcher R, van Soest RJ, et al. Loss of SLCO1B3 drives taxane resistance in prostate cancer. Br J Cancer. 2016;115(6):674-81.
- 89. Takata R, Takahashi A, Fujita M, et al. 12 new susceptibility loci for prostate cancer identified by genome-wide association study in Japanese population. Nat Commun. 2019;10(1):4422.
- 90. Fantus RJ, Helfand BT. Germline genetics of prostate cancer:

time to incorporate genetics into early detection tools. Clin Chem. 2019;65(1):74-9.

JMA Journal is an Open Access journal distributed under the Creative Commons Attribution 4.0 International License. To view the details of this license, please visit (http://creativecommons.org/ licenses/by/4.0/).