

The complete mitochondrial genome of *Histiostoma blomquisti* (Acari: Histiostomatidae)

Chih-Chi Lee  and John Wang

Biodiversity Research Center, Academia Sinica, Taipei, Taiwan

ABSTRACT

The mite *Histiostoma blomquisti* is a microorganism feeder that uses the red imported fire ant (*Solenopsis invicta*) as a phoretic carrier for dispersal. We sequenced the *H. blomquisti* mitogenome using next-generation sequencing methods. The circular mitogenome of *H. blomquisti* is 15,892 bp and is composed of 13 protein-coding genes, two ribosomal RNA genes, 22 transfer RNAs, and 6 non-coding regions >100 bp. Most tRNAs are highly reduced, like those found in other Acariformes. Phylogenetic analysis based on the concatenated nucleotide sequence of the 13 protein-coding genes supports Histiostomatid mites forming the basal-most lineage in Astigmata.

ARTICLE HISTORY

Received 12 July 2016
Revised 18 July 2016
Accepted 29 July 2016

KEYWORDS

Histiostomatidae;
mitochondrial genome;
mites; red imported
fire ants

Histiostomatid mites are filter feeders whose phoretic second stage nymphs, or deutonymphs, disperse by attaching to arthropods. The deutonymphs of *Histiostoma blomquisti* can often be found on the cuticle of red imported fire ant (*Solenopsis invicta*) queens (Sokolov et al. 2003; Wirth & Moser 2010). Here, we present the first complete Histiostomatid mitogenome.

For sample collection, we scraped approximately 330 deutonymphs from the abdominal cuticle of *S. invicta* queens. Queens were from colonies of the polygynous social form collected from Taoyuan City, Taiwan (24°56'06.71"N, 121°12'40.39"E). We mounted three mites onto slides (ASIZ01000010 - ASIZ01000012) and deposited them at the Biodiversity Research Museum, Academia Sinica, Taiwan.

To obtain enough DNA for sequencing, we extracted genomic DNA with the QIAamp[®] DNA Micro Kit (QIAGEN) and then amplified the DNA with the GenomiPhi[™] V3 DNA Amplification Kit (GE Healthcare Life Sciences). We sequenced the amplified DNA on the Illumina MiSeq platform (average library insert size 579 bp; paired-end read length 301 bp).

For assembly and annotation of the mitogenome, we pre-processed raw sequence reads (cutadapt v1.9.1 (Martin 2011); parameters: -a -q 20 -m 70) and conducted *de novo* whole genome assembly (IDBA-UD (Peng et al. 2012); parameters: -mink 40 -min_count 4 -min_support 2). We identified two mitogenome contigs from the genome assembly. After, we conducted PCR, cloning, and sequencing to bridge the two contigs into a circular mitogenome. We annotated protein coding genes (PCGs) using MITOS (Bernt et al. 2013) and

OrfFinder (Sayers et al. 2011). We annotated tRNAs using ARWEN (Laslett & Canbäck 2008), tRNAscan-SE (Lowe & Eddy 1997), and manual identification based on the anticodon and predicted secondary structure.

The complete mitogenome of *H. blomquisti* (GenBank: KX452726) is 15,892 bp, which is the largest Sarcopitiformes mitogenome to date. The nucleotide composition is AT biased (70%), similar to other Acariformes. The *H. blomquisti* mitogenome contains 13 PCGs, 2 rRNAs, and 22 tRNAs, typical for most animals. However, the tRNAs range in size from 45–59 bp, which are shorter than usual animal tRNAs (circa 75–85 bp). The smallest four tRNAs (45–50 bp, *trnR*, *trnA*, *trnV*, and *trnS*^(UCN)) could only be annotated manually. Among the 13 PCGs, 12 use ATN as the start codon (N, any nucleotide) while *nad3* uses TTG. The stop codons of five PCGs are incomplete (T- for *nad2* and *CYTb*; TA- for *cox2*, *nad1*, and *nad6*). The two largest non-coding regions (903 and 413 bp) are separated by a tRNA. As the former is adjacent to *rrnS*, we presume it corresponds to the control region.

We inferred the phylogenetic relationship of 15 mites within Acariformes, including at least one representative from each family, using the concatenated nucleotide sequences of the 13 PCGs that were aligned based on the corresponding amino acid translation (Edgar 2004). Both maximum likelihood (GTRGAMMA model, RAxMLGUI (Silvestro & Michalak 2012)) and Bayesian inference (GTR+I+ Γ model, MrBayes 3.2.5 (Ronquist et al. 2012)) yielded the same phylogeny indicating that Histiostomatid mites form the basal-most sequenced lineage within Astigmata (Figure 1).

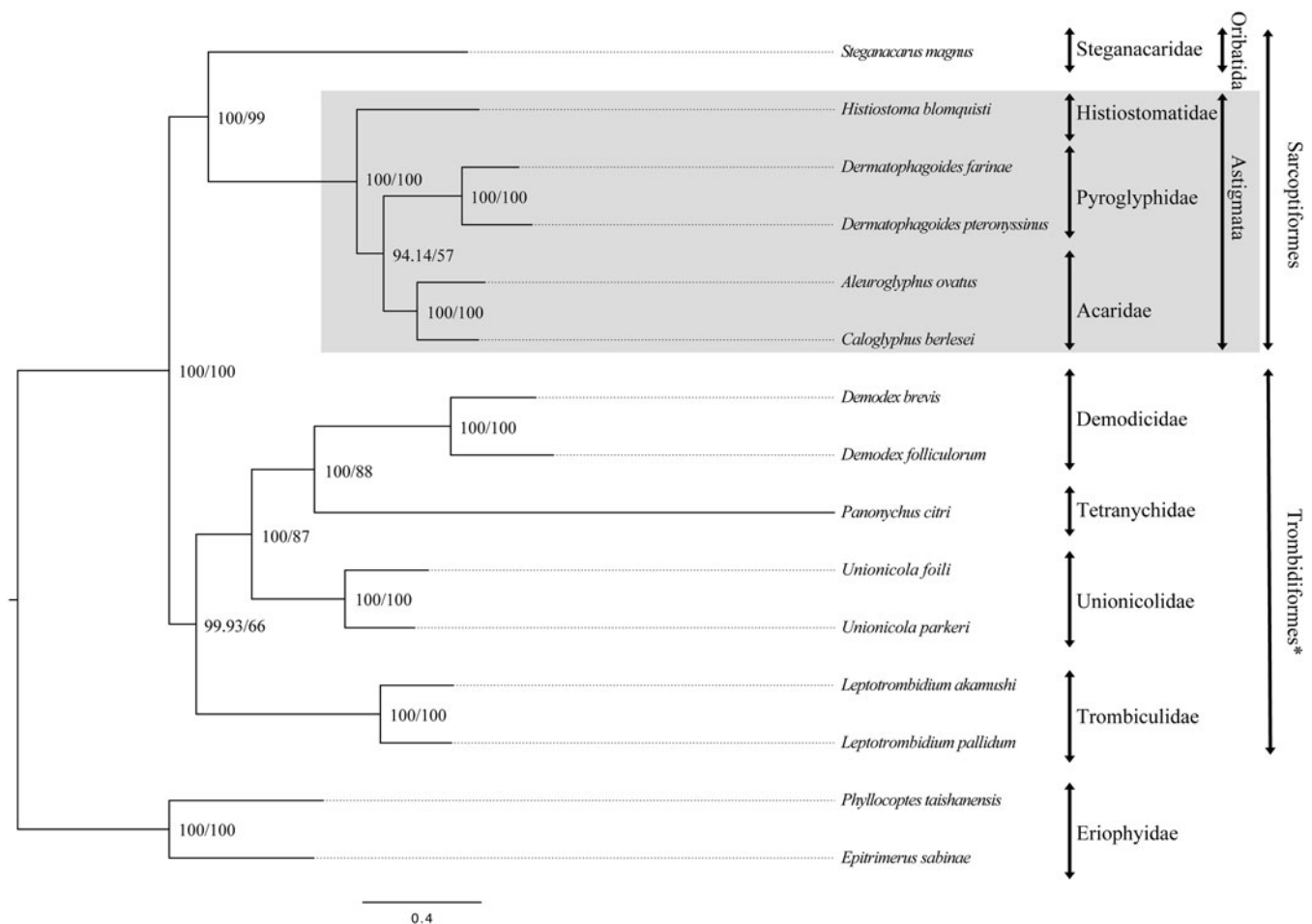


Figure 1. Molecular phylogeny of *Histiostoma blomquisti* and 14 other Acariformes based on the concatenated nucleotide sequences of 13 PCGs. The phylogenetic tree was constructed by the Bayesian inference and maximum-likelihood methods under GTR + I + Γ and GTRGAMMA models, respectively. The numbers at each node indicate the posterior probability (100,000 generations, sampled every 100 generations) and the bootstrap probability (1000 replicates) resulting from the analyses. The mitogenome accession numbers for tree construction are listed as follows: *Aleuroglyphus ovatus* (KC700022), *Caloglyphus berlesei* (KF499016), *Dermatophagoides pteronyssinus* (EU884425), *D. farinae* (NC_013184), *Steganacarus magnus* (EU935607), *Histiostoma blomquisti* (this study: KX452726), *Demodex brevis* (KM114225), *Demodex folliculorum* (KM114226), *Phyllocoptes taishanensis* (KR604967), *Eptrimerus sabinae* (KR604966), *Panonychus citri* (HM189212), *Leptotrombidium pallidum* (AB180098), *L. akamushi* (AB194045), *Unionicola foili* (EU856396), and *U. parkeri* (HQ386015). Astigmata (shaded box). *We rooted the phylogenetic tree using Eriophyiidae based on their exclusion from Trombidiformes in Xue et al. (2016).

Acknowledgements

We thank Dr Mei-Yeh Lu and the High Throughput Genomics Core (BRCAS) for sequencing and advice; and the Biodiversity Research Museum, Academia Sinica for sample preservation.

Disclosure statement

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of this article.

Funding

Academia Sinica, 10.13039/501100001869 [103-CDA-L01]; Ministry of Science and Technology, Taiwan, 10.13039/501100004663 [MOST 101-2621-M-001-006, MOST 103-2311-B-001-018-MY3, MOST 103-2621-M-001-004, MOST 104-2314-B-001-009-MY5]; and Biodiversity Research Center, Academia Sinica

ORCID

Chih-Chi Lee  <http://orcid.org/0000-0002-8778-1449>

References

- Bernt M, Donath A, Jühling F, Externbrink F, Florentz C, Fritsch G, Pütz J, Middendorf M, Stadler PF. 2013. MITOS: improved de novo metazoan mitochondrial genome annotation. *Mol Phylogenet Evol.* 69:313–319.
- Edgar RC. 2004. Muscle: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res.* 32:1792–1797.
- Laslett D, Canbäck B. 2008. ARWEN: a program to detect tRNA genes in metazoan mitochondrial nucleotide sequences. *Bioinformatics.* 24:172–175.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res.* 25:955–964.
- Martin M. 2011. Cutadapt removes adapter sequences from high-throughput sequencing reads. *EMBnet J.* 17:10–12.
- Peng Y, Leung HC, Yiu S-M, Chin FY. 2012. IDBA-UD: a de novo assembler for single-cell and metagenomic sequencing data with highly uneven depth. *Bioinformatics.* 28:1420–1428.
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP. 2012. MrBayes 3.2: efficient bayesian phylogenetic inference and model choice across a large model space. *Syst Biol.* 61:539–542.
- Sayers EW, Barrett T, Benson DA, Bolton E, Bryant SH, Canese K, Chetvernin V, Church DM, DiCuccio M, Federhen S. 2011. Database

- resources of the national center for biotechnology information. Nucleic Acids Res. 39:D38–D51.
- Silvestro D, Michalak I. 2012. raxmlGUI: a graphical front-end for RAxML. Org Divers Evol. 12:335–337.
- Sokolov IM, Sokolova YY, Fuxa JH. 2003. Histiostomatid mites (histiostomatidae: Astigmata: Acarina) from female reproductives of the red imported fire ant. J Entomol Sci. 38:699–702.
- Wirth S, Moser J. 2010. *Histiostoma blomquisti* n. Sp.(acari: Histiostomatidae) a phoretic mite of the red imported ant, *Solenopsis invicta* buren (hymenoptera: Formicidae). Acarologia. 50: 357–371.
- Xue X-F, Guo J-F, Dong Y, Hong X-Y, Shao R. 2016. Mitochondrial genome evolution and tRNA truncation in Acariformes mites: New evidence from eriophyoid mites. Sci Rep. 6: 18920.