

## ORIGINAL RESEARCH

# Effect of foliar application of selenium on morphological and physiological indices of savory (*Satureja hortensis*) under cadmium stress

Iraj Azizi | Behrooz Esmailpour  | Hamideh Fatemi

Department of Horticulture, University of Mohagheh Ardabili, Ardabil, Iran

**Correspondence**

Behrooz Esmailpour, Department of Horticulture, University of Mohagheh Ardabili, Ardabil, Iran.

Email: behsmaiel@yahoo.com

**Abstract**

Cadmium is a heavy metal that pollutes the environment and affects plants physiologically and morphologically. Selenium is considered as a beneficial element, with effective roles in increasing plant tolerance to environmental stresses. A greenhouse factorial pot experiment was conducted to study the impact of selenium on traits of Savory plants under Cd stress. Experimental factors included soil contamination with cadmium (0, 75, 100, and 150  $\mu\text{M}$ ) and foliar spraying of selenium (0, 10, 20, and 40  $\mu\text{M}$  of Sodium Selenate). Biomass, photosynthetic pigments including chlorophyll a, chlorophyll b, total chlorophyll, proline, total soluble solids, cell membrane leakage, relative water content of leaves antioxidant enzymes, and Cd and Zn concentration in shoot and root were recorded. Results revealed that Cd stress decreased vegetative growth criteria, photosynthetic pigments include chlorophyll a, chlorophyll b, total chlorophyll, and carotenoid almost, 55%, 57%, 57%, and 68%, respectively, while proline, cell membrane leakage, peroxidase (POD), and catalase (CAT) antioxidant enzymes were increased with increasing Cd concentrations. Foliar spray of selenium reduced the toxic effects of Cd stress on savory plants via enhancing of proline content and stimulation of CAT and POD enzymes and limitation of cell membrane leakage. Also, selenium foliar spray improved chlorophyll content under Cd stress condition and decreased cadmium accumulation 29% in root, respectively. In general, these results suggest that foliar application of selenium could mitigate Cd toxicity and improve growth and antioxidant capacity of savory under different level of cadmium heavy metal stress.

**KEYWORDS**

cadmium, heavy metals, savory, selenium

## 1 | INTRODUCTION

Cadmium is one of the most toxic heavy metals among major environmental pollutants. Cd is released into water and soil by humans

through urban, industrial, and agricultural activities. Most importantly, contamination by Cd in agriculture happens through long-term use of phosphate fertilizers, contaminated water, and waste water application in irrigation (He et al., 2009; Uruguchi & Fujiwara, 2012). This

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2020 The Authors. *Food Science & Nutrition* published by Wiley Periodicals LLC

unnecessary element which is considered as a highly mobile contaminant is allowed to enter into vegetables easily and can enter into several vital processes of the plant, thereby leading to poor growth, low economic performance of plants, and threats to human health (Di Toppi & Gabbrielli, 1999; Ekmekçi et al., 2008; Shamsi et al., 2008).

Cd-contaminated agricultural land is a major problem because the metal is easily absorbed by the root of the plants and can be translocate to aerial parts. Cd can impair the process of water absorption and cause an imbalance in micronutrient content, photosynthesis, and nitrogen metabolism. Ultimately, it inhibits plant growth and reduces the biomass (Anjum et al., 2008; Ghaghelestany et al., 2020; Gill et al., 2012; Jahanbakhshi & Kheiralipour, 2019). It can lead to plant death in severe cases of contamination (Di Toppi & Gabbrielli, 1999). Cd can inhibit root and branch growth, and reduce chlorophyll biosynthesis (Siedlecka & Krupa, 1999). It usually disrupts photosynthesis, respiration, and relative water content (Gouia et al., 2003). Several studies confirmed that the effects of Cd on decrease in activity of responsible enzymes in the absorption of nitrate and sulfate in plants (Ghnaya et al., 2005; Gouia et al., 2003). Cd can prevent the activity of enzymes which play important roles in the Calvin cycle in Sandalio et al., (2001), the synthesis of carbohydrates, and phosphorus metabolism (Di Toppi & Gabbrielli, 1999).

Cd stimulates the generation of reactive oxygen species (ROS) and leads to oxidative stress in most plants (Gill & Tuteja, 2010) which disrupt photosynthetic pigments and biomolecules such as lipids, proteins, and nucleic acids, along with a significant reduction in growth, production, and even the death of a plant (Foyer & Noctor, 2005). Therefore, plants have evolved antioxidant defense to ameliorate oxidative damage of Cd stress via enzymatic antioxidants (SOD, CAT, APX and GR), nonenzymatic antioxidants (glutathione [GSH] and ascorbate [AsA]; Gill et al., 2012; Mittler et al., 2004).

Selenium is an essential micronutrient with antioxidant, anti-cancer, and antiviral properties for the health of humans and animals, although the need for Se in plants is not proven as yet (Pilon-Smits, 2015). Se at low concentrations plays an important role in antioxidant reactions such as increased glutathione peroxidase activity and hormonal balance in plant cells (Cartes et al., 2010; Filek et al., 2008). The use of low levels of Se (5 and 10  $\mu\text{M}$ ) increased growth and photosynthetic capacity of treated cucumber seedlings under NaCl salinity (Hawrylak-Nowak, 2009).

Selenium plays protective and antioxidant role in decreasing oxidative stress caused by temperature, drought, salinity, mechanical stress, UV radiation, pathogens, and heavy metals. Also, selenium ameliorates stress by increasing the antioxidant capacity of the plant through increasing the activity of enzymatic antioxidants and nonenzymatic antioxidants (Ahangarnezhad et al., 2019; Azarmdel et al., 2020; Cartes et al., 2010; Haghghi et al., 2016; Jahanbakhshi et al., 2018, 2019, 2020; Lin et al., 2012; Momeny et al., 2020; Pandey & Gupta, 2015; Qing et al., 2015; Yao et al., 2009).

Summer savory (*Satureja hortensis* L.) is an annual and herbaceous plant from Lamiaceae family (Mumivand et al., 2013). The leaves and shoots of summer savory contain essential oil (Omidbaigi, 2009), and its essential oil is being used widely in medicinal, food, and health

industrials (Leake et al., 2003). Summer savory is also used in the traditional medicine to treat muscle pains, indigestion, diarrhoea, and infection diseases (Gursoy et al., 2009). The major essential oil compounds of summer savory are carvacrol, thymol,  $\gamma$ -terpinene, and borneol (Kamkar et al., 2013).

Since summer savory is an important medicinal crop, it would be valuable to investigate the responses of this plant to Cd stress. Foliar spraying of Se may alleviate some of the detrimental effects of Cd on *Satureja hortensis* plants. Therefore, this research was conducted to investigate the impacts of foliar spray of Se on some physiological and biochemical characteristics of summer savory under Cd stress.

## 2 | MATERIALS AND METHODS

### 2.1 | Experimental design and soil preparation

A pot experiment was arranged as factorial on the basis of completely randomized design with three replicates in the greenhouse of the Agricultural and Natural Resources Faculty of University of Mohaghegh Ardabili to assess the response of Se foliar spraying on growth and physiology of summer savory (*Satureja hortensis* L.) under Cd stress condition. Cadmium chloride ( $\text{CdCl}_2$ ) and sodium selenate ( $\text{Na}_2\text{SeO}_4$ ) were purchased from Merck. The  $\text{CdCl}_2$  solution was spiked as Cd stress in four concentrations (0, 75, 100, and 150  $\mu\text{M}$ ) to soil, and its moisture content was adjusted to field capacity by adding deionized water. The soils were incubated in stable (darkness, 40°C, in dry and wet) conditions for four months with frequent stirring to allow complete equilibration.

### 2.2 | Seedling cultivation

The seeds of Summer savory native ecotype of Shahr ray were obtained from the Medicinal Plant Research Station of Shahid Beheshti University of Tehran province, Iran. These seeds were surface sterilized with sodium hypochlorite for 5 min and washed five times with distilled water. Savory seeds (*Satureja hortensis*) were planted at a depth of 0.5–1 cm in pots containing 10 kg Cd-polluted soil. Pots then kept in greenhouse at growth conditions consisting the temperature of 22°C, relative humidity of 40%–50%, and average light intensity of 50% (Figure 1). Sodium selenate at four concentrations (0, 10, 20, and 40  $\mu\text{M}$ ) was applied for foliar spray in three growth stage. The first treatment was applied after emergence of two true leaves, and the other two foliar spraying were applied at 2-week intervals.

### 2.3 | Traits measurement

At the end of growth stage (4 months, at flowering stage), the plants were removed and separated into different parts (root, shoot, and leaf), then their morphological traits such as number of leaves, number of lateral branches, plant height by using meter rod, plant



**FIGURE 1** Savory seedlings after foliar application

biomass (weighing balance), and root weight (weighing balance) after oven drying for 72 hr at 70°C.

## 2.4 | Photosynthetic pigments

Photosynthetic pigments and carotenoid were extracted from fresh leaves according to the method of Harmut (1987). About 0.1 g of leaf homogenated with acetone (80%). The absorbance of the pigments was measured by UV visible spectrophotometer (Jenway, Italy) 470, 645, and 663 nm. Photosynthetic pigments and carotenoid content were estimated using the equations proposed by Harmut (1987).

## 2.5 | Membrane stability index

For determination of membrane stability index, leaf disks were prepared from fully expanded leaves and were washed three times with deionized water. The samples were placed in container containing 10 ml deionized water for 24 hr and shaken at 25°C. Then, the EC value of each sample was measured ( $L_t$ ). The samples and solution were autoclaved then EC was measured ( $L_0$ ). The electrolyte leakage from cell membrane was estimated by Equation (1) proposed by Redman et al. (1986).

$$\text{Soluble materials leakage \%} = \frac{L_t}{L_0} \times 100 \quad (1)$$

## 2.6 | Relative water content

Relative water content of plants was determined at the end of experiment according to Ritchie et al. (1990) method. For instance, 0.5 g from the youngest leaf of each plant (FW) was sampled and floated in distilled water for 24 hr then leaf saturation weight was measured (TW). Finally, the leaves were  $n$  placed in an oven at 70°C for 24 hr, and their dry weight was measured according to following equation:

$$\text{RWC\%} = \frac{\text{FW} - \text{DW}}{\text{TW} - \text{DW}} \quad (2)$$

## 2.7 | Proline accumulation

Proline content of leaves was determined by method of Bates et al. (1973). Samples from the youngest leaves (0.5 g) in each treatment was homogenized in 2 ml of 3% sulfosalicylic acid and then was centrifuged at 2000 g for 5 min. The filtered homogenate was mixed with equal volume of ninhydrin and glacial acetic acid in a test tube within a water bath at 100°C for an hour. The reaction was terminated in an ice bath, and the solution was extracted by toluene. The absorbance was recorded at 520 nm. The proline content was calculated according to standard curves of L-proline and reported as  $\mu\text{g}$  per g leaf fresh weight.

## 2.8 | Measurement of carbohydrate content

In order to measure the amount of carbohydrates, at first the extract of the leaves was prepared. For this purpose, 0.1 g of leaf specimen, along with 1 ml ethanol (95%), was pulverized inside a porcelain mortar. Then, 1 ml ethanol (70%) was added to it. The final solution was centrifuged at 2058 g for 15 min. After removing the alcoholic extract containing soluble sugars, 0.1 mg of this extract was added to 3 mg antiroll-sulfuric acid (72%). The mixture was placed in a water bath for 15 min at 100°C. After cooling the mixture, the amounts of soluble sugars were measured at 625 nm (Irigoyen et al., 1992).

## 2.9 | CAT enzyme activity

Protein extraction was carried out based on Sudhakar et al. (2001). Catalase activities (CAT) were estimated using the procedure described by Kar and Mishra (1976). For the catalase enzyme, 60  $\mu\text{l}$  of protein extract was added to 2.5 ml 50 mM buffer with pH = 7 and 0.3 ml 5 mg oxygenated water in an ice bath. The curve of absorption and variation was read at 240 nm. Enzyme activity ( $\mu\text{g}/\text{ml}$ ) was measured of fresh protein.

## 2.10 | POD enzyme activity

The activity of peroxidase was determined by guaiacol and  $\text{H}_2\text{O}_2$  substrates as described by Chance and Maehly (1955). With the extinction coefficient of tetraguaiacol product ( $26.6 \text{ mM}^{-1} \text{ cm}^{-1}$ ), the activity of POX was expressed as mmol produced tetraguaiacol per minute per mg soluble protein (U/mg).

## 2.11 | Measurement of cadmium and zinc elements

Leaf samples from each treatment rinsed with deionized water and then dried at 65°C for 48 hr. The samples were powdered uniformly. Then, (0.1 g) from plant samples were digested with an acid mixture of  $\text{HNO}_3/\text{HClO}_4$  at 100°C and placed in a furnace at 550°C for 5 hr. After cooling, the specimens were removed from the furnace and 10 ml of 2 N normal

chloride acid was added to it. The samples were dissolved in acid with a gentle heat. The solution was filtered through a filter paper (Whatman filter paper No. 42) and was poured into a 50cc laboratory flask. Distilled water was added to a filter paper to wash the remaining material in the funnel in order to transfer the extract back into the laboratory flask. Finally, the extract was diluted with water and reached a volume of 50cc. It was stored for chemical analysis in the refrigerator (Jones, 2001). The amounts of Cd and Zn in the extracts of the aerial parts and roots were read separately by atomic absorption spectrometers.

## 2.12 | Statistical analysis

All data were subjected to two-way analysis of variance (ANOVA) in SAS 9.1. The mean values were separated by LSD (Least Significant Difference) test at  $p < 0.05$  (Jahanbakhshi et al., 2020). The  $p$  values of less than .05 considered statistically significant.

## 3 | RESULTS

### 3.1 | Morphological traits

Cd stress, Se foliar spray, and their interactions significantly influenced biomass (Table 1). Increasing Cd stress resulted in continuous reductions of vegetative traits such as stem height, fresh and dry

weight of root, dry weight of plant, and number of lateral leaves and branches. Se foliar spraying significantly improved the vegetative traits of summer savory plants under different concentrations of Cd stress. According to the results of the, the highest value for all of morphological traits was obtained by foliar spraying of 40  $\mu\text{M}$  Se in plants grown in unpolluted soils and the least amount of these traits was related to 150  $\mu\text{M}$  of Cd and Se control treatment. It seems that Se has increased the amount of carbohydrates, dry and wet weight, and other morphological characteristics of the plant by improving the process of photosynthesis and reducing the damage to chlorophyll under Cd stress.

### 3.2 | Proline

The results show that Proline accumulation significantly increased with increasing the Cd stress in savory plants (Table 2). Likewise, increasing the concentration of Se after foliar application increases the amount of proline, so that the highest proline content  $1.82 \mu\text{g}(\text{FW})^{-1}$  is achieved by 150  $\mu\text{M}$  Cd contamination and 40  $\mu\text{M}$  Se treatment.

### 3.3 | Carbohydrate

The results state that (Table 2), the highest amount of carbohydrates  $1.39 \mu\text{g}(\text{FW})^{-1}$  is seen in the treatment of 40  $\mu\text{M}$  Se and the

**TABLE 1** Comparison of the interactions between selenium soluble and cadmium stress on morphological indices of savory

Treatment ( $\mu\text{M}$ )	Stem height (cm)	Root fresh weight (g)	Number of lateral branches	Number of leaves	Dry plant weight (g)	Root dry weight (g)
<b>Cd 0</b>						
Se 0	45.83 <sup>bc</sup>	2.21 <sup>abc</sup>	22.50 <sup>bc</sup>	146.67 <sup>abcd</sup>	4.90 <sup>abcd</sup>	0.65 <sup>ab</sup>
Se 10	47.83 <sup>abc</sup>	2.52 <sup>ab</sup>	24.16 <sup>ab</sup>	163.33 <sup>ab</sup>	5.46 <sup>ab</sup>	0.82 <sup>ab</sup>
Se 20	49.16 <sup>ab</sup>	2.53 <sup>ab</sup>	25.66 <sup>ab</sup>	172.33 <sup>a</sup>	6.02 <sup>a</sup>	0.90 <sup>a</sup>
Se 40	50.66 <sup>a</sup>	2.76 <sup>a</sup>	27.33 <sup>a</sup>	173.33 <sup>a</sup>	6.02 <sup>a</sup>	0.90 <sup>a</sup>
<b>Cd 75</b>						
Se 0	39.00 <sup>efg</sup>	1.70 <sup>bcd</sup>	21.66 <sup>bcd</sup>	131.67 <sup>abcdef</sup>	4.59 <sup>abcde</sup>	0.65 <sup>ab</sup>
Se 10	39.83 <sup>ef</sup>	1.80 <sup>bcd</sup>	21.83 <sup>bcd</sup>	145.00 <sup>abcde</sup>	4.25 <sup>bcdef</sup>	0.66 <sup>ab</sup>
Se 20	41.16 <sup>de</sup>	2.09 <sup>abc</sup>	22.33 <sup>bcd</sup>	146.67 <sup>abcd</sup>	5.37 <sup>abc</sup>	0.69 <sup>ab</sup>
Se 40	44.50 <sup>dc</sup>	2.21 <sup>abc</sup>	22.33 <sup>bcd</sup>	161.33 <sup>abc</sup>	5.40 <sup>abc</sup>	0.76 <sup>ab</sup>
<b>Cd 100</b>						
Se 0	37.00 <sup>efg</sup>	1.42 <sup>dc</sup>	20.50 <sup>cde</sup>	105.00 <sup>def</sup>	2.85 <sup>f</sup>	0.56 <sup>bc</sup>
Se 10	37.66 <sup>efg</sup>	1.45 <sup>dc</sup>	21.00 <sup>cd</sup>	116.67 <sup>bdef</sup>	3.79 <sup>cdef</sup>	0.56 <sup>bc</sup>
Se 20	38.33 <sup>efg</sup>	1.48 <sup>dc</sup>	22.16 <sup>bcd</sup>	116.67 <sup>bdef</sup>	4.13 <sup>bcdef</sup>	0.57 <sup>bc</sup>
Se 40	39.16 <sup>ef</sup>	1.63 <sup>bcd</sup>	22.00 <sup>bcd</sup>	126.67 <sup>abcdef</sup>	4.28 <sup>bcdef</sup>	0.61 <sup>abc</sup>
<b>Cd 150</b>						
Se 0	34.66 <sup>e</sup>	1.08 <sup>d</sup>	18.00 <sup>e</sup>	95.00 <sup>f</sup>	2.85 <sup>f</sup>	0.33 <sup>c</sup>
Se 10	35.66 <sup>fg</sup>	1.34 <sup>dc</sup>	19.00 <sup>e</sup>	98.33 <sup>ef</sup>	3.14 <sup>ef</sup>	0.33 <sup>c</sup>
Se 20	35.83 <sup>fg</sup>	1.42 <sup>dc</sup>	19.50 <sup>de</sup>	105.00 <sup>d<sup>ef</sup></sup>	3.25 <sup>ef</sup>	0.54 <sup>bc</sup>
Se 40	36.16 <sup>fg</sup>	1.42 <sup>dc</sup>	20.50 <sup>cde</sup>	115.00 <sup>cd<sup>ef</sup></sup>	3.58 <sup>def</sup>	0.55 <sup>bc</sup>

The alphabets in each column do not show statistically significant difference in the 5% probability level based on the LSD test. Nonsimilar letters indicate a significant difference at 5% probability level.

**TABLE 2** Comparison of the interactions between selenium soluble and cadmium stress on Physiological indices of savory

Treatment ( $\mu\text{M}$ )	Proline [ $\mu\text{g}(\text{FW})^{-1}$ ]	Carbohydrate [ $\mu\text{g}(\text{FW})^{-1}$ ]	Chlorophyll A [ $\mu\text{g}(\text{FW})^{-1}$ ]	Chlorophyll b [ $\mu\text{g}(\text{FW})^{-1}$ ]	Total chlorophyll [ $\mu\text{g}(\text{FW})^{-1}$ ]	Carotenoid [ $\mu\text{g}(\text{FW})^{-1}$ ]	Membrane cell leakage (%)
<b>Cd 0</b>							
Se 0	0.41 <sup>h</sup>	0.29 <sup>g</sup>	6.45 <sup>d</sup>	2.42 <sup>c</sup>	9.18 <sup>c</sup>	1.69 <sup>de</sup>	13.30 <sup>fgh</sup>
Se 10	0.46 <sup>gh</sup>	0.33 <sup>fg</sup>	7.19 <sup>bc</sup>	2.46 <sup>c</sup>	9.81 <sup>b</sup>	1.92 <sup>bc</sup>	12.61 <sup>ghi</sup>
Se 20	0.52 <sup>fgh</sup>	0.37 <sup>fg</sup>	7.33 <sup>b</sup>	2.81 <sup>ab</sup>	10.15 <sup>b</sup>	2.19 <sup>ab</sup>	11.74 <sup>ij</sup>
Se 40	0.6 <sup>fg</sup>	0.44 <sup>f<sup>g</sup></sup>	8.18 <sup>a</sup>	3.04 <sup>a</sup>	11.23 <sup>a</sup>	2.38 <sup>a</sup>	10.97 <sup>j</sup>
<b>Cd 75</b>							
Se 0	0.43 <sup>h</sup>	0.35 <sup>fg</sup>	5.6 <sup>ef</sup>	1.76 <sup>efg</sup>	7.33 <sup>f</sup>	1.29 <sup>fg</sup>	15.33 <sup>de</sup>
Se 10	0.55 <sup>fgh</sup>	0.42 <sup>fg</sup>	6.16 <sup>de</sup>	1.83 <sup>def</sup>	8.14 <sup>e</sup>	1.47 <sup>ef</sup>	14.33 <sup>ef</sup>
Se 20	0.66 <sup>f</sup>	0.5 <sup>ef</sup>	6.47 <sup>d</sup>	1.95 <sup>de</sup>	8.48 <sup>de</sup>	1.56 <sup>e</sup>	13.48 <sup>fg</sup>
Se 40	0.87 <sup>e</sup>	0.65 <sup>dce</sup>	6.71 <sup>cd</sup>	2.01 <sup>d</sup>	8.87 <sup>cd</sup>	1.83 <sup>cd</sup>	11.92 <sup>hij</sup>
<b>Cd 100</b>							
Se 0	0.82 <sup>e</sup>	0.47 <sup>ef</sup>	4.69 <sup>gh</sup>	1.47 <sup>hij</sup>	5.83 <sup>hi</sup>	0.84 <sup>ij</sup>	18.33 <sup>b</sup>
Se 10	1.04 <sup>d</sup>	0.62 <sup>ef</sup>	4.89 <sup>gh</sup>	1.83 <sup>def</sup>	6.83 <sup>fg</sup>	0.94 <sup>hij</sup>	16.72 <sup>cd</sup>
Se 20	1.28 <sup>c</sup>	0.8 <sup>cd</sup>	5.05 <sup>fg</sup>	1.59 <sup>ghi</sup>	6.84 <sup>fg</sup>	1.01 <sup>h</sup>	14.43 <sup>ef</sup>
Se 40	1.31 <sup>c</sup>	1.03 <sup>b</sup>	5.49 <sup>f</sup>	1.67 <sup>fgh</sup>	7.27 <sup>f</sup>	1.15 <sup>gh</sup>	12.42 <sup>ghij</sup>
<b>Cd 150</b>							
Se 0	1.02 <sup>d</sup>	0.64 <sup>de</sup>	2.87 <sup>j</sup>	1.04 <sup>k</sup>	3.91 <sup>i</sup>	0.54 <sup>k</sup>	20.65 <sup>a</sup>
Se 10	1.23 <sup>c</sup>	0.83 <sup>de</sup>	3.89 <sup>i</sup>	1.33 <sup>j</sup>	5.29 <sup>i</sup>	0.71 <sup>jk</sup>	18.19 <sup>b</sup>
Se 20	1.52 <sup>b</sup>	1.09 <sup>b</sup>	4 <sup>i</sup>	1.41 <sup>ij</sup>	5.83 <sup>hi</sup>	0.94 <sup>hij</sup>	16.19 <sup>bc</sup>
Se 40	1.82 <sup>a</sup>	1.39 <sup>a</sup>	4.35 <sup>hi</sup>	1.47 <sup>hij</sup>	6.29 <sup>gh</sup>	1.08 <sup>ghi</sup>	15.08 <sup>e</sup>

The alphabets in each column do not show statistically significant difference in the 5% probability level based on the LSD test.

Nonsimilar letters indicate a significant difference at 5% probability level.

contamination level is 150  $\mu\text{M}$  Cd and the lowest amount is in the control level of Se and Cd.

### 3.4 | Chlorophyll

According to the results of comparison of mean (Table 2) of the interaction effect of Cd stress and foliar application of Se on this index, the chlorophyll content (a, b, and carotenoids) also decreased by increasing Cd concentration, but increased Se concentration increases chlorophyll content (a, b, and carotenoids). So that, the highest levels of chlorophyll a, b, and carotenoids are related to the level of Cd without contamination and 40  $\mu\text{M}$  Se treatment.

### 3.5 | Membrane cell leakage

Evidence suggests (Table 2) that increasing the concentration of Cd leads to disruption of membrane protein pumps and with increasing concentration of Cd ion leakage from the cell increases but the application of Se reduces cell leakage. So that, the highest membrane leakage rate 20.65 (%) is related to 150  $\mu\text{M}$  Cd contamination and Se control treatment, and the lowest membrane leakage rate 10.97(%)

is related to the level of non-contamination Cd and 40  $\mu\text{M}$  treatment of Se.

### 3.6 | Antioxidant enzymes

As Cd stress increases compared to the control level, oxidative stress also increases, which leads to the production of reaction oxygen species (ROS), and the plant's antioxidant system is activated to counteract this stress, and Se reduces this stress by improving this activity, as seen the highest amount of POD and CAT enzymes is related to 150  $\mu\text{M}$  Cd contamination and 40  $\mu\text{M}$  Se treatment and the lowest amount of POD enzyme is related to noncontamination level of Cd and control treatment of Se (Table 3).

### 3.7 | Relative water content of leaves

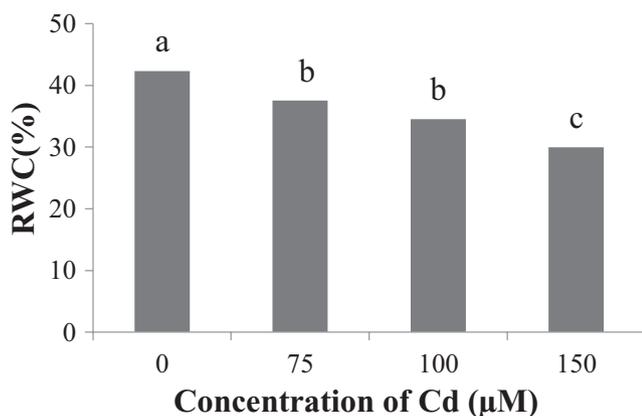
According to the results of (Figure 2) of the effect of Cd stress on relative water content of leaves, the most percentage of relative water content was related to the control level of Cd and the lowest relative water content was related to 150  $\mu\text{M}$  Cd. The relative water content of leaves decreases by increasing the concentration of Cd. But Se

**TABLE 3** Comparison of the interactions between selenium soluble and cadmium stress on antioxidant enzymes of savory

Treatment ( $\mu\text{M}$ )	POD enzyme [ $\mu\text{g protein (min)}^{-1}$ ]	CAT enzyme [ $\mu\text{g protein (min)}^{-1}$ ]
Cd 0		
Se 0	0.00106 <sup>i</sup>	0.000150 <sup>g</sup>
Se 10	0.00126 <sup>hi</sup>	0.000163 <sup>g</sup>
Se 20	0.00143 <sup>gh</sup>	0.000183 <sup>fg</sup>
Se 40	0.00160 <sup>efg</sup>	0.000210 <sup>ef</sup>
Cd 75		
Se 0	0.00126 <sup>hi</sup>	0.000180 <sup>fg</sup>
Se 10	0.00146 <sup>fgh</sup>	0.000203 <sup>f</sup>
Se 20	0.00170 <sup>ef</sup>	0.000243 <sup>e</sup>
Se 40	0.00196 <sup>cd</sup>	0.000286 <sup>d</sup>
Cd 100		
Se 0	0.00140 <sup>gh</sup>	0.000243 <sup>e</sup>
Se 10	0.00173 <sup>de</sup>	0.000296 <sup>d</sup>
Se 20	0.00206 <sup>cd</sup>	0.000343 <sup>c</sup>
Se 40	0.00250 <sup>b</sup>	0.000393 <sup>b</sup>
Cd 150		
Se 0	0.00173 <sup>de</sup>	0.000303 <sup>d</sup>
Se 10	0.00203 <sup>c</sup>	0.000350 <sup>c</sup>
Se 20	0.00243 <sup>b</sup>	0.000416 <sup>b</sup>
Se 40	0.00283 <sup>a</sup>	0.000473 <sup>a</sup>

<sup>a</sup>The alphabets in each column do not show statistically significant difference in the 5% probability level based on the LSD test.

Nonsimilar letters indicate a significant difference at 5% probability level.

**FIGURE 2** Effect of cd stress on leaf relative water content

maintains the relative water content of the leaf by affecting the leaf cell stomata, so that the highest relative water content is related to 40  $\mu\text{M}$  Se (Figure 3).

### 3.8 | Aerial parts Cd

Studies on the effect of Se foliar application on Cd accumulation in aerial parts of the plant showed that the most element of aerial part

Cd was observed in 40  $\mu\text{M}$  Se treatment and the least amount was related to control Se treatment (Figure 4). The results also showed the effect of Cd stress on the Cd aerial part, the highest accumulation of Cd in the aerial part is related to Cd 150  $\mu\text{M}$  and the lowest amount of Cd aerial part is related to the Cd control level (Figure 5).

### 3.9 | The accumulation of root Cd

In this study (Table 4), the highest root Cd accumulation 1.92 mg/L was in 150  $\mu\text{M}$  Cd and Se control and the lowest root Cd concentration 1.07 mg/L was related to the control Cd and 40  $\mu\text{M}$  Se treatment.

### 3.10 | Zinc element accumulation on the aerial parts

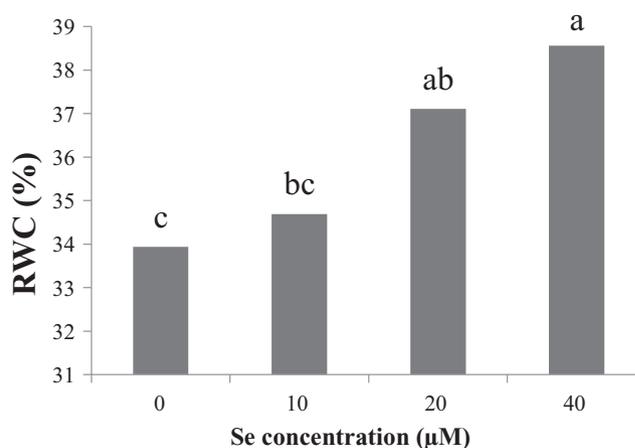
Due to the chemical similarity of Cd to zinc, Cd replaces this element. Therefore, the lowest accumulation of zinc 14 mg/L is observed in the 150  $\mu\text{M}$  Cd treatment and the highest accumulation 1.42 mg/L is observed in the control surface of Cd and 40  $\mu\text{M}$  Se treatment (Table 4).

### 3.11 | Zinc element accumulation on the root

Examination showed the accumulation of zinc on the roots (Figure 6), the most amount of zinc accumulation on the root was related to Se control treatment and the least amount of element on the root is related to 40  $\mu\text{M}$  Se treatment. According to the results (Figure 7) of the effect of Cd stress on root zinc, the most amount of accumulation on the root is related to the control level of Cd and the least root level is related to 150  $\mu\text{M}$  Cd.

## 4 | DISCUSSION

Cadmium is considered as a heavy element in the Earth's crust. It is not only unnecessary for the physiological processes of plants

**FIGURE 3** Effect of Se on leaf relative water content

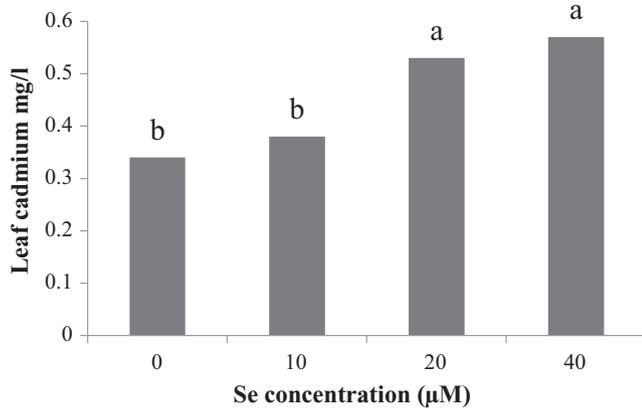


FIGURE 4 Effect of Se on Aerial Cd

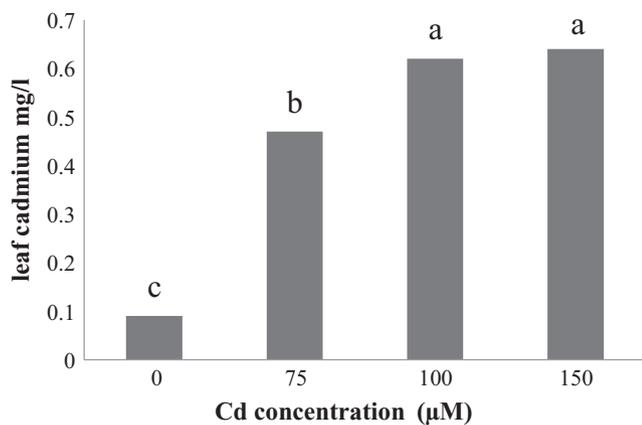


FIGURE 5 Effect of Cd stress on Aerial Cd

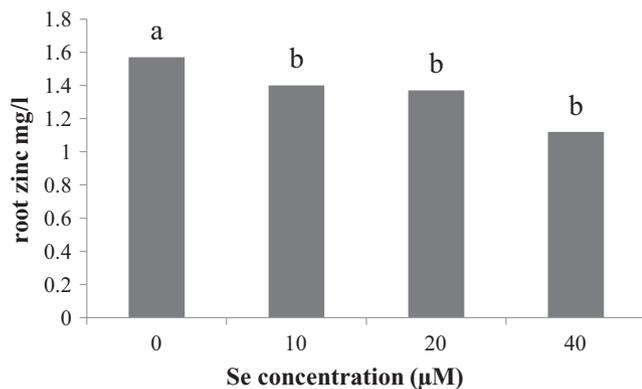


FIGURE 6 Effect of Se on root zinc

but is also toxic to plants and animals even in very low quantities. Reduction in growth and chlorosis is considered as primary effects of Cd on plants. They lead to fewer stems, smaller leaf area, and lower values of fresh and dry weight of the root and other vegetable parts. Some of the most important causes of growth impairment by heavy metals include disorders in the cellular water balance and cell wall elasticity, disorders in the plant's water balance due to reduced cell size, and fewer xylems, along with a reduced absorption of essential nutrients such as  $\text{Fe}^{+2}$ ,  $\text{K}^+$ ,  $\text{Mg}^{+2}$ , and  $\text{Ca}^{+2}$ . Heavy metals

TABLE 4 Comparison of the interactions between selenium soluble and cadmium stress on morphological indices of savory

Treatment (μM)	Root cadmium (mg/L)	Leaf zinc (mg/L)
Cd 0		
Se 0	1.28 <sup>efg</sup>	1.30 <sup>ab</sup>
Se 10	1.13 <sup>gh</sup>	1.30 <sup>ab</sup>
Se 20	1.07 <sup>h</sup>	1.33 <sup>ab</sup>
Se 40	1.07 <sup>h</sup>	1.42 <sup>a</sup>
Cd 75		
Se 0	1.29 <sup>efg</sup>	1.33 <sup>ab</sup>
Se 10	1.21 <sup>efgh</sup>	1.31 <sup>ab</sup>
Se 20	1.51 <sup>bcd</sup>	1.21 <sup>abc</sup>
Se 40	1.33 <sup>def</sup>	1.09 <sup>bcd</sup>
Cd 100		
Se 0	1.54 <sup>bc</sup>	0.86 <sup>defg</sup>
Se 10	1.51 <sup>bcd</sup>	0.96 <sup>cdef</sup>
Se 20	1.36 <sup>cde</sup>	1.09 <sup>bcd</sup>
Se 40	1.33 <sup>def</sup>	1.02 <sup>bcd</sup>
Cd 150		
Se 0	1.92 <sup>a</sup>	0.14 <sup>h</sup>
Se 10	1.68 <sup>b</sup>	0.51 <sup>g</sup>
Se 20	1.39 <sup>cde</sup>	0.61 <sup>fg</sup>
Se 40	1.36 <sup>cde</sup>	0.83 <sup>efg</sup>

The alphabets in each column do not show statistically significant difference in the 5% probability level based on the LSD test.

Nonsimilar letters indicate a significant difference at 5% probability level.

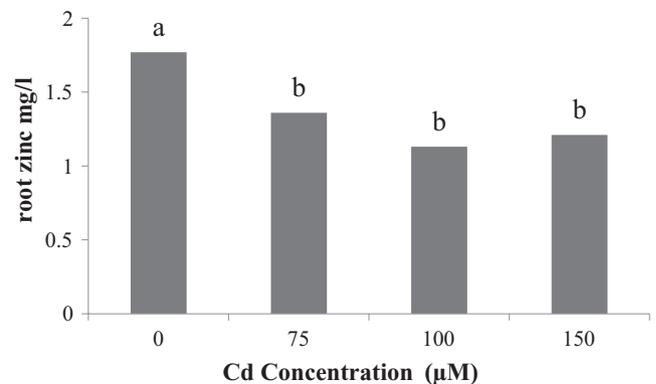


FIGURE 7 Effect of Cd on root zinc

can reduce the growth of plants significantly by severely disrupting photosynthesis, by deterring photosynthetic production and cell division. These ultimately result in shorter internode length and shorter plant height (Chaffei et al., 2003). The interaction of selenite with the plasma membrane, along with changes in membrane permeability regarding ions (potassium, sodium, and calcium), may affect plant growth, respiration, water uptake, and phloem activity

(Dziubinska et al., 2010). Also, Se is a component of several amino acids. It increases the production of ethylene and promotes changes in the composition of membrane lipids, while increasing membrane permeability, potassium leakage, and water content in the intercellular space (Xue et al., 2001).

Proline can act as an antioxidant to reduce the threat of free radicals. It maintains the integrity of the membrane by inhibiting lipid peroxidation (Mehta & Gaur, 1999). In general, there is a special relationship between proline accumulation and water deficiency caused by heavy metals, which ultimately inhibits plant growth (Metwally et al., 2003). There could be a significant relationship between the accumulation of proline under Cd treatment with mechanisms of resistance to osmotic changes. This relationship could also mean a decrease in the activity of the electron transfer system in plants or parts of the plant. Also, a decrease in metabolic activity leads to the accumulation of NADH. Since 2NADH molecules are required for synthesis of a proline molecule of glutamic acid, proline synthesis can serve as a mechanism to reduce acidity and prevent the accumulation of NADH (Saradhi, 1991).

According to previous results (John et al., 2008), the amount of soluble and insoluble carbohydrates in plants can increase or decrease depending on the concentration of heavy metals where the plants grow. Soluble sugars increase by changing the activity of protein channels of water transfer and by closing the leaf stomata, thereby limiting the loss of water from the plant (Zhang & Tyerman, 1999). By decreasing the transfer of water into leaves and after the accumulation of Cd in the cells, the amount of soluble sugars in the plant increases. In addition, the increase in soluble sugars helps the plant keep its carbohydrate content in order to maintain basal metabolism under stress conditions. These results are consistent with a previous report on rice (Verma & Dubey, 2001).

According to the results of this study, the amounts of chlorophyll and carotenoids in savory seedlings decreased when increasing the Cd concentration in the growth medium. Heavy metals disrupt compound formations by inhibiting the biosynthesis of LHCII compound proteins at the transcriptional level (Katznelson et al., 1962). Carotenoids play an important role in protection against oxidative stress. These pigments can assist in detoxification and can reduce the toxic effects of free radicals (Di Toppi & Gabbriellini, 1999).

A low amount of chlorophyll content in the leaf can occur when the synthesis of photosynthetic pigments is inhibited by Cd. This happens because of the lowered absorption of essential nutrients such as iron, manganese, and magnesium (Prasad & Strzałka, 1999). Increasing the amount of Se to appropriate levels can partially reduce chloroplast degradation and increase chlorophyll content (Cai et al., 2011).

Liu et al. (2004) reported that the reaction of Se with ROS can reduce the negative effects of Cd on chlorophyll content in rice leaves. This is caused because free oxygen radicals can break down photosynthetic pigments and structural proteins of the photosynthetic system under Cd stress conditions. This has been proven especially on D1 protein (i.e., one of the first targets for oxidative degradation in the photosystem 2 reaction center; Kim et al., 2006). The

restoration of photosynthesis in plants under stress after application of Se may be associated with reduced levels of ROS (Paciolla et al., 2011).

Heavy metals can damage the membrane by disabling membrane enzymes and inhibiting membrane ATPase activity, thereby affecting membrane integrity (Ruley et al., 2006). Heavy metals cause changes in the cell membrane lipid composition. Thus, damages to the cell membrane can lead to the release of ions out of the cell (Singh et al., 2010). In this study, increasing the cadmium concentration enhanced the amount of ion leakage. The highest ion leakage rate occurred in response to the 150  $\mu\text{M}$  Cd treatment. Meanwhile, Se reduced the ion leakage and increased cell membrane stability by increasing the potassium uptake in plant cells (SaffarYazdy et al., 2012). The lowest amount of ion leakage resulted from using 40  $\mu\text{M}$  Se with no amount of Cd treatment.

It has been proven that peroxidase acts as the main enzyme against stresses caused by heavy metals. This enzyme is known as a stress marker in conditions of heavy metal stress (Choudhary et al., 2007). The formation of ROS can be controlled by the presence of antioxidant enzymes, including peroxidases in plant cells. These stimulate the antioxidant activity in aerial parts of plants and roots where soils contain heavy metals. This is in accordance with previous reports on *Solanum lycopersicum* (Quiroga et al., 2000) and *Raphanus sativus* (Chen et al., 2002).

In this study, observations show that the highest accumulation of cadmium in the roots occurred by 150  $\mu\text{M}$  Cd. In this regard, ions are transferred through the cell membrane by unique proteins called transporters. Of the total ions around the root, only a small part is absorbed by the plant. Most of these ions are physically absorbed by the cell wall where the  $-\text{COO}$  compartment has a negative charge and is responsible for surface absorption in the cell wall. Ions that attach to this compartment cannot enter the cell in the aerial parts of plants. The accumulation of these elements increases the concentration of Cd in cellular vacuoles and prevents their transfer to aerial parts. Therefore, the amount of this element in roots is greater than its amount in aerial parts (Ramos et al., 2002).

Cd is chemically similar to zinc. It imitates the metabolic functions of zinc in the plant (Mengel & Kirkby, 2001) and can be absorbed into the plant instead of zinc (Grant et al., 1998). This similarity in the Cd and zinc properties indicates the importance of their interaction in absorption. It highlights the transfer from roots to the aerial part and the accumulation of Cd in edible tissues, as this element ultimately enters the food chain (An et al., 2005). In this study, it seems that Cd replaced zinc in the aerial parts and that the lowest amount of zinc was observed as a result of the 150  $\mu\text{M}$  Cd treatment.

## 5 | CONCLUSION

Cadmium stress reduced photosynthesis and cell division. It affected the quality of morphological indicators in savory seedlings, whereas spraying selenium on the seedlings reduced the rate of chloroplast destruction. In fact, selenium increased the amount of chlorophyll to

some extent and alleviated the toxic effects of stress caused by cadmium. Nutritional supplementation with selenium enhanced several biochemical features such as proline, carbohydrate, and chlorophyll in savory seedlings under cadmium stress conditions. Selenium reduced cell leakage and increased the activity of catalase and peroxidase enzymes. The treatment worked best when selenium was applied at a concentration of 40  $\mu$ M. Spraying the selenium solution on plants reduced the accumulation of cadmium in the roots and increased the uptake of zinc, followed by the translocation of zinc to the aerial parts of the plant. Therefore, selenium can be an effective agent against the toxic effects of cadmium.

## CONFLICT OF INTEREST

The authors have declared no conflict of interest.

## ETHICAL APPROVAL

This study does not involve any human or animal testing.

## INFORMED CONSENT

Written informed consent was obtained from all study participants.

## DATA AVAILABILITY STATEMENT

Due to the nature of this research, participants of this study did not agree for their data to be shared publicly, so supporting data are not available.

## ORCID

Behrooz Esmailpour  <https://orcid.org/0000-0002-7080-0236>

## REFERENCES

- Ahangarnezhad, N., Najafi, G., & Jahanbakhshi, A. (2019). Determination of the physical and mechanical properties of a potato (the Agria variety) in order to mechanise the harvesting and post-harvesting operations. *Research in Agricultural Engineering*, 65(2), 33–39. <https://doi.org/10.17221/122/2017-RAE>
- An, L., Liu, Y., Zhang, M., Chen, T., & Wang, X. (2005). Effects of nitric oxide on growth of maize seedling leaves in the presence or absence of ultraviolet-B radiation. *Journal of Plant Physiology*, 162(3), 317–326. <https://doi.org/10.1016/j.jplph.2004.07.004>
- Anjum, N. A., Umar, S., Ahmad, A., Iqbal, M., & Khan, N. A. (2008). Ontogenic variation in response of (*Brassica campestris* L.) to cadmium toxicity. *Journal of Plant Interactions*, 3(3), 189–198.
- Azarmdel, H., Jahanbakhshi, A., Mohtasebi, S. S., & Muñoz, A. R. (2020). Evaluation of image processing technique as an expert system in mulberry fruit grading based on ripeness level using artificial neural networks (ANNs) and support vector machine (SVM). *Postharvest Biology and Technology*, 166, 111201. <https://doi.org/10.1016/j.postharvbio.2020.111201>
- Bates, L. S., Waldren, R. P., & Teare, I. D. (1973). Rapid determination of free proline for water-stress studies. *Plant and Soil*, 39(1), 205–207. <https://doi.org/10.1007/BF00018060>
- Cai, Y., Cao, F., Cheng, W., Zhang, G., & Wu, F. (2011). Modulation of exogenous glutathione in phytochelatin and photosynthetic performance against Cd stress in the two rice genotypes differing in Cd tolerance. *Biological Trace Element Research*, 143(2), 1159–1173. <https://doi.org/10.1007/s12011-010-8929-1>
- Cartes, P., Jara, A. A., Pinilla, L., Rosas, A., & Mora, M. L. (2010). Selenium improves the antioxidant ability against aluminium-induced oxidative stress in ryegrass roots. *Annals of Applied Biology*, 156(2), 297–307. <https://doi.org/10.1111/j.1744-7348.2010.00387.x>
- Chaffei, C., Gouia, H., & Ghorbe, M. (2003). Nitrogen metabolism in tomato plants under cadmium stress. *Journal of Plant Nutrition*, 26, 1617–1634. <https://doi.org/10.1081/PLN-120022372>
- Chance, B., & Maehly, A. (1955). Assay of catalases and peroxidases. *Methods in Enzymology*, 11, 764–775.
- Chen, E. L., Chen, Y. A., Chen, L. M., & Liu, Z. H. (2002). Effect of copper on peroxidase activity and lignin content in (*Raphanus sativus*). *Plant Physiology and Biochemistry*, 40(5), 439–444. [https://doi.org/10.1016/S0981-9428\(02\)01392-X](https://doi.org/10.1016/S0981-9428(02)01392-X)
- Choudhary, M., Jetley, U. K., Khan, M. A., Zutshi, S., & Fatma, T. (2007). Effect of heavy metal stress on proline, malondialdehyde, and superoxide dismutase activity in the cyanobacterium *Spirulina platensis*-S5. *Ecotoxicology and Environmental Safety*, 66(2), 204–209. <https://doi.org/10.1016/j.ecoenv.2006.02.002>
- Di Toppi, L. S., & Gabbriellini, R. (1999). Response to cadmium in higher plants. *Environmental and Experimental Botany*, 41(2), 105–130. [https://doi.org/10.1016/S0098-8472\(98\)00058-6](https://doi.org/10.1016/S0098-8472(98)00058-6)
- Dziubinska, H., Filek, M., Krol, E., & Trebacz, K. (2010). Cadmium and selenium modulate slow vacuolar channels in rape (*Brassica napus*) vacuoles. *Journal of Plant Physiology*, 167(18), 1566–1570. <https://doi.org/10.1016/j.jplph.2010.06.016>
- Ekmekçi, Y., Tanyolac, D., & Ayhan, B. (2008). Effects of cadmium on antioxidant enzyme and photosynthetic activities in leaves of two maize cultivars. *Journal of Plant Physiology*, 165(6), 600–611. <https://doi.org/10.1016/j.jplph.2007.01.017>
- Filek, M., Keskinen, R., Hartikainen, H., Szarejko, I., Janiak, A., Miszalski, Z., & Golda, A. (2008). The protective role of selenium in rape seedlings subjected to cadmium stress. *Journal of Plant Physiology*, 165(8), 833–844. <https://doi.org/10.1016/j.jplph.2007.06.006>
- Foyer, C. H., & Noctor, G. (2005). Oxidant and antioxidant signalling in plants: A re-evaluation of the concept of oxidative stress in a physiological context. *Plant, Cell & Environment*, 28(8), 1056–1071. <https://doi.org/10.1111/j.1365-3040.2005.01327.x>
- Ghaghelestany, A. B., Jahanbakhshi, A., & Taghinezhad, E. (2020). Gene transfer to German chamomile (*L. chamomilla* M) using cationic carbon nanotubes. *Scientia Horticulturae*, 263, 109106. <https://doi.org/10.1016/j.scienta.2019.109106>
- Ghnaya, T., Nouairi, I., Slama, I., Messedi, D., Grignon, C., Abdelly, C., & Ghorbel, M. H. (2005). Cadmium effects on growth and mineral nutrition of two halophytes: (*Sesuvium portulacastrum*) and (*Mesembryanthemum crystallinum*). *Journal of Plant Physiology*, 162(10), 1133–1140. <https://doi.org/10.1016/j.jplph.2004.11.011>
- Gill, S. S., Khan, N. A., & Tuteja, N. (2012). Cadmium at high dose perturbs growth, photosynthesis and nitrogen metabolism while at low dose it up regulates sulfur assimilation and antioxidant machinery in garden cress (*Lepidium sativum* L.). *Plant Science*, 182, 112–120. <https://doi.org/10.1016/j.plantsci.2011.04.018>
- Gill, S. S., & Tuteja, N. (2010). Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiology and Biochemistry*, 48(12), 909–930. <https://doi.org/10.1016/j.plaphy.2010.08.016>
- Gouia, H., Suzuki, A., Brulfert, J., & Ghorbal, M. H. (2003). Effects of cadmium on the co-ordination of nitrogen and carbon metabolism in bean seedlings. *Journal of Plant Physiology*, 160(4), 367–376. <https://doi.org/10.1078/0176-1617-00785>
- Grant, C. A., Buckley, W. T., Bailey, L. D., & Selles, F. (1998). Cadmium accumulation in crops. *Canadian Journal of Plant Science*, 78(1), 1–17. <https://doi.org/10.4141/P96-100>
- Gursoy, U. K., Gursoy, M., Gursoy, O. V., Cakmakci, L., Könönen, E., & Uitto, V. J. (2009). Anti-biofilm properties of *Satureja hortensis* L. essential oil against periodontal pathogens. *Anaerobe*, 15(4), 164–167.

- Haghighi, M., Shebanirad, A., & Pessarakli, M. (2016). Effects of selenium as a beneficial element on growth and photosynthetic attributes of greenhouse cucumber. *Journal of Plant Nutrition*, 39(10), 1493–1498. <https://doi.org/10.1080/01904167.2015.1109116>
- Harmut, A. (1987). Chlorophylls and carotenoids: pigments of photosynthetic membranes. *Methods Enzymol.*, 148, 350–383.
- Hawrylak-Nowak, B. (2009). Beneficial effects of exogenous selenium in cucumber seedlings subjected to salt stress. *Biological Trace Element Research*, 132(1–3), 259–269. <https://doi.org/10.1007/s12011-009-8402-1>
- He, J. Y., Ren, Y. F., Wang, F. J., Pan, X. B., Zhu, C., & Jiang, D. A. (2009). Characterization of cadmium uptake and translocation in a cadmium-sensitive mutant of rice (*Oryza sativa* L. ssp. japonica). *Archives of Environmental Contamination and Toxicology*, 57(2), 299–306. <https://doi.org/10.1007/s00244-008-9273-8>
- Irigoyen, J. J., Einerich, D. W., & Sánchez-Díaz, M. (1992). Water stress induced changes in concentrations of proline & total soluble sugars in nodulated alfalfa (*Medicago sativa*) plants. *Physiologia Plantarum*, 84(1), 55–60.
- Jahanbakhshi, A., Abbaspour-Gilandeh, Y., Ghamari, B., & Heidarbeigi, K. (2019). Assessment of physical, mechanical, and hydrodynamic properties in reducing postharvest losses of cantaloupe (*Cucumis melo* var. Cantaloupenis). *Journal of Food Process Engineering*, 42(5), e13091. <https://doi.org/10.1111/jfpe.13091>
- Jahanbakhshi, A., Abbaspour-Gilandeh, Y., & Gundoshmian, T. M. (2018). Determination of physical and mechanical properties of carrot in order to reduce waste during harvesting and post-harvesting. *Food Science & Nutrition*, 6(7), 1898–1903. <https://doi.org/10.1002/fsn3.760>
- Jahanbakhshi, A., & Kheiralipour, K. (2019). Influence of vermicompost and sheep manure on mechanical properties of tomato fruit. *Food Science & Nutrition*, 7(4), 1172–1178. <https://doi.org/10.1002/fsn3.877>
- Jahanbakhshi, A., Momeny, M., Mahmoudi, M., & Zhang, Y. D. (2020). Classification of sour lemons based on apparent defects using stochastic pooling mechanism in deep convolutional neural networks. *Scientia Horticulturae*, 263, 109133. <https://doi.org/10.1016/j.scienta.2019.109133>
- Jahanbakhshi, A., Yeganeh, R., & Momeny, M. (2020). Influence of ultrasound pre-treatment and temperature on the quality and thermodynamic properties in the drying process of nectarine slices in a hot air dryer. *Journal of Food Processing and Preservation*, 44(10), e14818. <https://doi.org/10.1111/jfpp.14818>
- John, R., Ahmad, P., Gadgil, K., & Sharma, S. (2008). Effect of cadmium and lead on growth, biochemical parameters and uptake in (*Lemna polyrrhiza* L.). *Plant Soil and Environment*, 54(6), 262.
- Jones, J. B. (2001). *Laboratory guide for conduction soil tests and plant analysis*. CRC Press LLC.
- Kamkar, A., Tooryan, F., Akhondzadeh Basti, A., Misaghi, A., & Shariatifar, N. (2013). Chemical composition of summer savory (*Satureja hortensis* L.) essential oil and comparison of antioxidant activity with aqueous and alcoholic extracts. *Journal of Veterinary Research*, 68(2), 183–190.
- Kar, M., & Mishra, D. (1976). Catalase, peroxidase, and polyphenoloxidase activities during rice leaf senescence. *Plant Physiology*, 57(2), 315–319. <https://doi.org/10.1104/pp.57.2.315>
- Katznelson, H., Peterson, E. A., & Rouatt, J. W. (1962). Phosphate-dissolving microorganisms on seed and in the root zone of plants. *Canadian Journal of Botany*, 40(9), 1181–1186. <https://doi.org/10.1139/b62-108>
- Kim, S.-H., Sicher, R. C., Bae, H., Gitz, D. C., Baker, J. T., Timlin, D. J., & Reddy, V. R. (2006). Canopy photosynthesis, evapotranspiration, leaf nitrogen, and transcription profiles of maize in response to CO<sub>2</sub> enrichment. *Global Change Biology*, 12(3), 588–600. <https://doi.org/10.1111/j.1365-2486.2006.01110.x>
- Leake, G., Gaspar, F., & Santos, R. (2003). The effect of water on the solubility of essential oils in dense CO<sub>2</sub>. *Journal of Essential Oil Research*, 15(3), 172–177.
- Lin, L., Zhou, W. H., Dai, H. X., Cao, F. B., Zhang, G. P., & Wu, F. B. (2012). Selenium reduce cadmium uptake and mitigates cadmium toxicity in rice. *Journal of Hazardous Materials*, 235, 343–351. <https://doi.org/10.1016/j.jhazmat.2012.08.012>
- Liu, Q., Wang, D. J., Jiang, X. J., & Cao, Z. H. (2004). Effects of the interactions between selenium and phosphorus on the growth and selenium accumulation in rice (*Oryza sativa*). *Environmental Geochemistry and Health*, 26(2), 325–330. <https://doi.org/10.1023/B:EGAH.0000039597.75201.57>
- Mehta, S. K., & Gaur, J. P. (1999). Heavy-metal-induced proline accumulation and its role in ameliorating metal toxicity in (*Chlorella vulgaris*). *The New Phytologist*, 143(2), 253–259.
- Mengel, K., & Kirkby, E. A. (2001). *Principles of plant nutrition*, 5th ed. : Kluwer Academic Publishers.
- Metwally, A., Finkemeier, I., Georgi, M., & Dietz, K. J. (2003). Salicylic acid alleviates the cadmium toxicity in barley seedlings. *Plant Physiology*, 132(1), 272–281. <https://doi.org/10.1104/pp.102.018457>
- Mittler, R., Vanderauwera, S., Gollery, M., & Van Breusegem, F. (2004). Reactive oxygen gene network of plants. *Trends in Plant Science*, 9(10), 490–498. <https://doi.org/10.1016/j.tplants.2004.08.009>
- Momeny, M., Jahanbakhshi, A., Jafarnezhad, K., & Zhang, Y. D. (2020). Accurate classification of cherry fruit using deep CNN based on hybrid pooling approach. *Postharvest Biology and Technology*, 166, 111204. <https://doi.org/10.1016/j.postharvbio.2020.111204>
- Mumivand, H., Nooshkam, A., Moseni, A., & Babalar, M. (2013). Influence of nitrogen and calcium carbonate application rates on nitrate accumulation and yield of summer savory (*Satureja hortensis* L.). *Electronic Journal of Crop Production*, 6(2), 109–124.
- Omidbaigi, R. (2009). *Production and processing of medicinal plants*, Vol. 1, 5th ed. (with complete revision; 347 pp.). : Astan Quds Publication.
- Paciolla, C., De Leonardis, S., & Dipierro, S. (2011). Effects of selenite and selenate on the antioxidant systems in (*Senecio scandens* L.). *Plant Biosystems*, 145(1), 253–259.
- Pandey, C., & Gupta, M. (2015). Selenium and auxin mitigates arsenic stress in rice (*Oryza sativa* L.) by combining the role of stress indicators, modulators and genotoxicity assay. *Journal of Hazardous Materials*, 287, 384–391. <https://doi.org/10.1016/j.jhazmat.2015.01.044>
- Pilon-Smits, E. A. H. (2015). Selenium in plants. In U. Luttge, & W. Beyschlag (Eds.), *Progress in botany* (pp. 93–107). Springer International Publishing.
- Prasad, M. N. V., & Strzałka, K. (1999). Impact of heavy metals on photosynthesis. In *Heavy metal stress in plants* (pp. 117–138). Springer.
- Qing, X., Zhao, X., Hu, C., Wang, P., Zhang, Y., & Zhang, X. (2015). Selenium alleviates chromium toxicity by preventing oxidative stress in cabbage (*Brassica campestris* L.) leaves. *Ecotoxicology and Environmental Safety*, 114, 179–189. <https://doi.org/10.1016/j.ecoenv.2015.01.026>
- Quiroga, M., Guerrero, C., Botella, M. A., Barceló, A., Amaya, I., Medina, M. I., Alonso, F. J., de Forchetti, S. M., Tigier, H., & Valpuesta, V. (2000). A tomato peroxidase involved in the synthesis of lignin and suberin. *Plant Physiology*, 122(4), 1119–1128. <https://doi.org/10.1104/pp.122.4.1119>
- Ramos, I., Esteban, E., Lucena, J. J., & Gárate, A. (2002). Cadmium uptake and subcellular distribution in plants of *Lactuca* sp. Cd–Mn interaction. *Plant Science*, 162(5), 761–767.
- Redman, R. E., Haraldson, J., & Gusta, L. V. (1986). Leakage of UV-absorbing substances as a measure of salt injury in leaf tissue of woody species. *Physiologia Plantarum*, 67(1), 87–91. <https://doi.org/10.1111/j.1399-3054.1986.tb01267.x>
- Ritchie, S. W., Nguyen, H. T., & Holaday, A. S. (1990). Leaf water content and gas-exchange parameters of two wheat genotypes differing in drought resistance. *Crop Science*, 30(1), 105–111. <https://doi.org/10.2135/cropsci1990.0011183X003000010025x>

- Ruley, A. T., Sharma, N. C., Sahi, S. V., Singh, S. R., & Sajwan, K. S. (2006). Effects of lead and chelators on growth, photosynthetic activity and Pb uptake in (*Sesbania drummondii*) grown in soil. *Environmental Pollution*, 144(1), 11–18. <https://doi.org/10.1016/j.envpol.2006.01.016>
- SaffarYazdy, A., Lahouti, M., Ganjeali, A., & Bayat, H. (2012). Impact of selenium supplementation on growth and selenium accumulation on spinach (*Spinacia oleracea* L.) plants. *Notulae Scientia Biologicae*, 4, 95–100. <https://doi.org/10.15835/nsb448029>
- Sandalio, L. M., Dalurzo, H. C., Gomez, M., Romero-Puertas, M. C., & Del Rio, L. A. (2001). Cadmium-induced changes in the growth and oxidative metabolism of pea plants. *Journal of Experimental Botany*, 52(364), 2115–2126. <https://doi.org/10.1093/jexbot/52.364.2115>
- Saradhi, P. P. (1991). Proline accumulation under heavy metal stress. *Journal of Plant Physiology*, 138(5), 554–558.
- Shamsi, I. H., Wei, K., Zhang, G. P., Jilani, G. H., & Hassan, M. J. (2008). Interactive effects of cadmium and aluminum on growth and antioxidative enzymes in soybean. *Biologia Plantarum*, 52(1), 165–169. <https://doi.org/10.1007/s10535-008-0036-1>
- Siedlecka, A., & Krupa, Z. (1999). Cd/Fe interaction in higher plants-its consequences for the photosynthetic apparatus. *Photosynthetica*, 36(3), 321–331. <https://doi.org/10.1023/A:1007097518297>
- Singh, G., Biswas, D. R., & Marwaha, T. S. (2010). Mobilization of potassium from waste mica by plant growth promoting rhizobacteria and its assimilation by maize (*Zea mays*) and wheat (*Triticum aestivum* L.): A hydroponics study under phytotron growth chamber. *Journal of Plant Nutrition*, 33(8), 1236–1251.
- Sudhakar, C., Lakshmi, A., & Giridarakumar, S. (2001). Changes in the antioxidant enzyme efficacy in two high yielding genotypes of mulberry (*Morus alba* L.) under NaCl salinity. *Plant Science*, 161(3), 613–619.
- Uraguchi, S., & Fujiwara, T. (2012). Cadmium transport and tolerance in rice: Perspectives for reducing grain cadmium accumulation. *Rice*, 5(1), 5. <https://doi.org/10.1186/1939-8433-5-5>
- Verma, S., & Dubey, R. S. (2001). Effect of cadmium on soluble sugars and enzymes of their metabolism in rice. *Biologia Plantarum*, 44(1), 117–123. <https://doi.org/10.1023/A:1017938809311>
- Xue, T., Hartikainen, H., & Piironen, V. (2001). Antioxidative and growth-promoting effect of selenium on senescing lettuce. *Plant and Soil*, 237(1), 55–61.
- Yao, X., Chu, J., & Wang, G. (2009). Effects of selenium on wheat seedlings under drought stress. *Biological Trace Element Research*, 130(3), 283–290. <https://doi.org/10.1007/s12011-009-8328-7>
- Zhang, W. H., & Tyerman, S. D. (1999). Inhibition of water channels by HgCl<sub>2</sub> in intact wheat root cells. *Plant Physiology*, 120(3), 849–858.

**How to cite this article:** Azizi I, Esmailpour B, Fatemi H. Effect of foliar application of selenium on morphological and physiological indices of savory (*Satureja hortensis*) under cadmium stress. *Food Sci Nutr*. 2020;8:6539–6549. <https://doi.org/10.1002/fsn3.1943>