



# Cerebral blood flow decrease as an early pathological mechanism in Alzheimer's disease

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## Abstract

Therapies targeting late events in Alzheimer's disease (AD), including aggregation of amyloid beta (A $\beta$ ) and hyperphosphorylated tau, have largely failed, probably because they are given after significant neuronal damage has occurred. Biomarkers suggest that the earliest event in AD is a decrease of cerebral blood flow (CBF). This is caused by constriction of capillaries by contractile pericytes, probably evoked by oligomeric A $\beta$ . CBF is also reduced by neutrophil trapping in capillaries and clot formation, perhaps secondary to the capillary constriction. The fall in CBF potentiates neurodegeneration by upregulating the BACE1 enzyme that makes A $\beta$  and by promoting tau hyperphosphorylation. Surprisingly, therefore, CBF reduction may play a crucial role in driving cognitive decline by initiating the amyloid cascade itself, or being caused by and amplifying A $\beta$  production. Here, we review developments in this area that are neglected in current approaches to AD, with the aim of promoting novel mechanism-based therapeutic approaches.

**Keywords** Alzheimer's · Cerebral blood flow · Capillary · Amyloid  $\beta$  · Pericyte · Neutrophil

## Introduction

Thirty years of research have given us a broad understanding of many mechanisms contributing to Alzheimer's disease [99], but over 400 clinical trials of drugs targeting these pathways have largely failed to reduce cognitive decline [47, 109, 136]. Identification of the amyloid  $\beta$  protein (A $\beta$ ) as the major component of amyloid plaques, together with genetic evidence, initially indicated that dysfunction of the processing of amyloid precursor protein (APP) was the cause of A $\beta$  plaque deposition and downstream tau tangle formation and neuronal dysfunction [59]. Subsequent work led to the conclusion that the level of soluble A $\beta$  oligomers, and of hyperphosphorylation of the cytoskeletal protein tau that is induced by A $\beta$  [62, 91], correlated better with cognitive decline than did plaque level [7, 57, 89, 123].

There are established mechanisms by which A $\beta$  oligomers and hyperphosphorylated tau can contribute to neuronal dysfunction and cognitive decline before synaptic and neuronal damage, and even before A $\beta$  plaque and tau tangle deposition (Fig. 1). A $\beta$  oligomers reduce glutamate uptake [92, 94, 199]. This raises the extracellular glutamate level and increases neuronal excitability [19, 20], which alters synaptic plasticity [92, 94] and in extremis may induce excitotoxicity [60]. Tau phosphorylation leads to soluble tau relocating from axonal microtubules into dendritic spines, where it alters postsynaptic glutamate receptor trafficking or anchoring (of both AMPA and NMDA receptors) and thus suppresses excitatory postsynaptic currents and neuronal activity [21, 67]. These changes may be particularly important when they affect the function of interneurons, which play a key role in generating oscillatory activity that contributes to cognitive function [63, 70, 176].

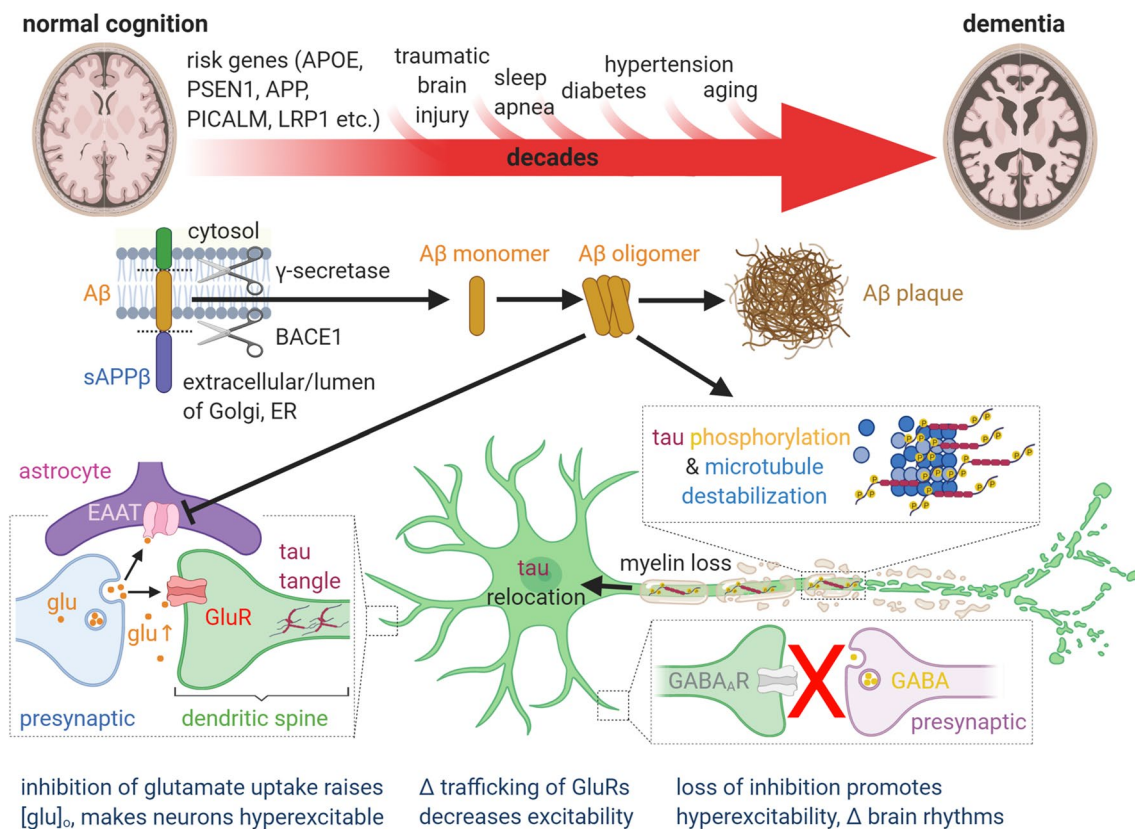
Preclinical AD has therefore been conceptualised as a synaptic disease [157] driven by A $\beta$  and downstream tau phosphorylation, with loss of synapses and cells occurring late in the disorder. However, individuals can be cognitively normal while having plaque levels as high as those in Alzheimer's dementia patients, and the same is true for levels of soluble A $\beta$  oligomers [39]. This could reflect the presence of compensating protective mutations

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**Fig. 1** Current, generally held ideas about the pathology underlying Alzheimer's disease (see main text for details). The transition from normal cognition to dementia, over decades, is promoted by the risk factors shown above the large red arrow. A $\beta$  is produced from amyloid precursor protein (APP) by the action of the  $\gamma$  secretase and  $\beta$  secretase (BACE1) as monomers, but these can then form soluble oligomers, which ultimately form extracellular precipitates as amyloid plaques. A $\beta$  oligomers inhibit astrocyte glutamate uptake (EAAT), thus potentiating the action of synaptically released glutamate (glu). This, together with a loss of GABAergic inhibition, leads to some

neurons becoming hyperexcitable. Meanwhile, A $\beta$  oligomers also induce hyperphosphorylation of axonal microtubule-associated tau, which leads to tau redistributing partly to dendrites where it disrupts trafficking of glutamate receptors and thus depresses excitation and neuronal firing. These synaptic effects, and A $\beta$ - and/or tau-induced loss of axonal myelin, may induce cognitive dysfunction well before synapses are lost and neurons die. The levels of A $\beta$  oligomers and hyperphosphorylated tau correlate better with cognitive decline than does the level of A $\beta$  plaques

or developmental differences in the subjects with high A $\beta$  levels. Alternatively, together with the fact that attempts to prevent cognitive decline—by blocking A $\beta$  production, removing A $\beta$  with antibodies or preventing tau phosphorylation—have all failed clinically (with one possible exception [68]), these data may suggest that there is some other variable that is missing from our understanding of the A $\beta$ -tau cascade. Previously it has been suggested that the vasculature might provide such a factor, in the form of hypertension, impaired blood–brain barrier function, decreased A $\beta$  clearance to the blood, vascular oxidative stress and inflammatory damage, or reduced neurovascular coupling at the arteriolar level [71, 198]. In this review, we show that new evidence reveals that a major missing variable is cerebral blood flow—and specifically its control by capillary pericytes.

## Large decreases of cerebral blood flow occur early in AD

Cerebral blood flow and glucose metabolism are reduced, and the brain's vascular resistance is increased, in human AD [17, 107, 112, 115, 144, 151, 163, 165, 188] and in mice overexpressing amyloid precursor protein (APP) to mimic AD [129]. This also occurs in humans and mice expressing the ApoE4 protein, which predisposes towards AD [111, 148, 162, 163, 172]. The CBF reduction reaches over 50% in some brain areas [5], which is expected to reduce the activity of the Na/K pump (the main consumer of ATP in the CNS: [8]) and all processes dependent on it (including maintenance of the resting potential and glutamate uptake). It will also lead to adenosine generation, which is known to suppress glutamate release [43], and

will produce numerous cell biological changes including changes of the balance of protein synthesis and degradation [173].

Although these changes could simply reflect tissue atrophy in AD [30], with a corresponding loss of blood supply and metabolism, they are associated with hypoxia [114] and it has been reported that the decrease of metabolism is greater than would be expected for the amount of atrophy occurring [165]. Furthermore, the observations of focal constrictions in capillaries from human AD brains [83], constriction of capillaries near plaques in human AD brains [58], and reduced neurovascular coupling and cerebrovascular reactivity in AD mice [48, 174] suggest that blood flow may be reduced by decreases in vessel diameter, and not just by loss of blood vessels.

Chronic blood flow reductions of 50% are expected to cause significant cognitive changes: a sustained reduction in CBF beyond 20% in humans leads to loss of ability to sustain attention, while a reduction beyond 30% in rats impairs spatial memory [105, 177]. A causal influence of blood flow changes on the cognitive changes at the onset of Alzheimer's disease, before synapses or neurons are lost, is suggested by the fact that the reduction of cerebral blood flow starts early in preclinical AD [107, 180], with a faster onset than the deposition of A $\beta$  or tau [76], and the fall of metabolism is also an early event [81, 115]. Furthermore, these changes correlate with cognitive decline [17, 112, 151].

### Cerebral blood flow decreases in AD largely reflect pericytes constricting capillaries

The brain is unusual in that most of the resistance in its vascular bed is in capillaries (Fig. 2a) rather than in arterioles or venules [49], and cerebral blood flow is controlled not only by vascular smooth muscle cells wrapped around arterioles, but also by contractile pericytes which enwrap at least the first 4 branch orders of capillaries from the penetrating arteriole [9, 56, 82, 84, 143, 152, 187]. Contraction of these pericytes produces localised capillary constrictions near the pericyte somata (where most of the circumferential processes of the pericytes are located [133]) and could account for the focal capillary constrictions seen anatomically in capillaries isolated from human AD brains [83].

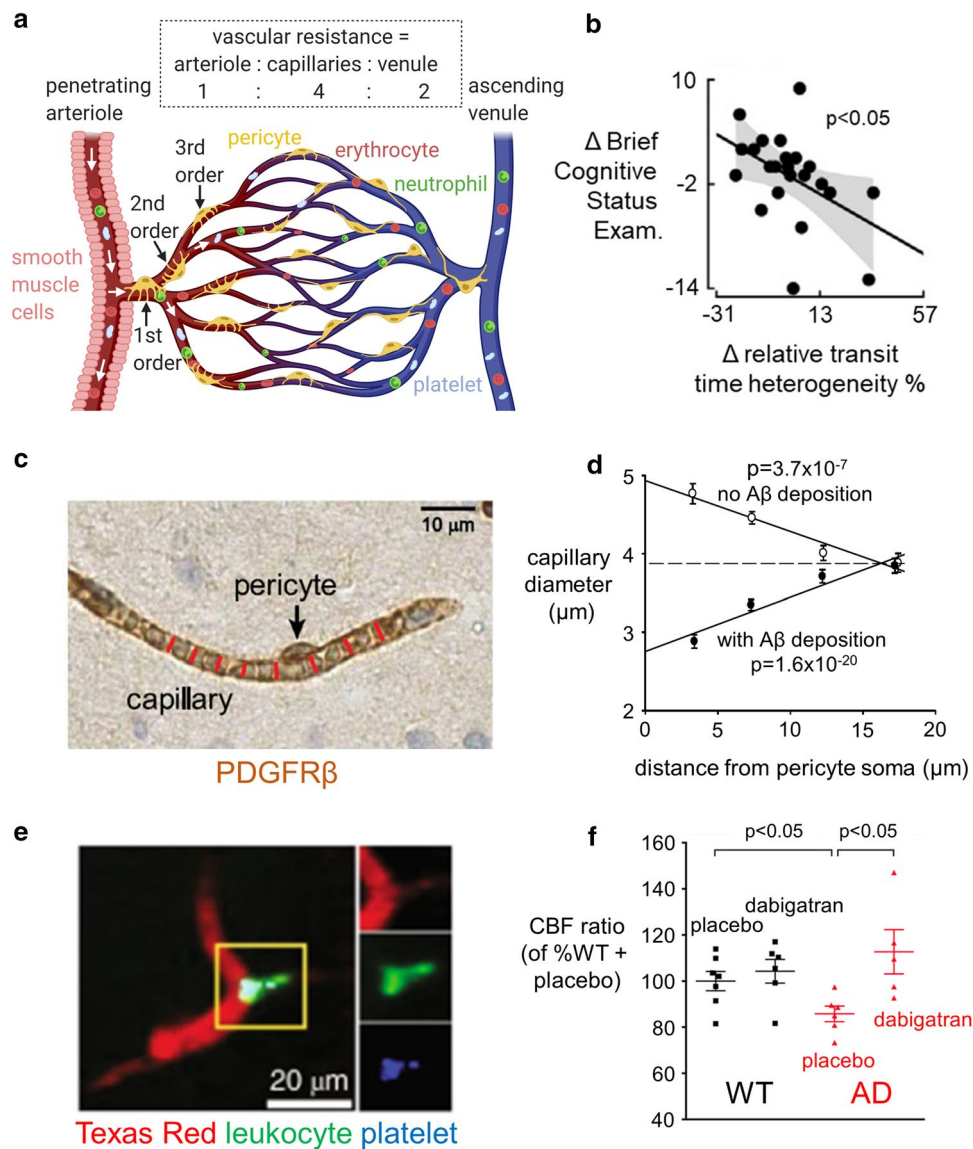
Despite the award of the Nobel Prize to Krogh [87] for his discovery of contractile elements on capillaries which act independently of smooth muscle cells on arterioles, there has been some controversy in the literature about whether pericytes are in fact contractile. However, this debate has now largely been resolved. The Zlokovic group [127] assessed in vitro, ex vivo and in vivo studies on pericyte

contractility and found that 37 out of 39 separate papers reported that pericytes display contractility (and one of the 2 remaining papers [65] actually showed pericytes contracting, but renamed these cells smooth muscle cells: see [9] for discussion). Furthermore, whereas contractility had previously been demonstrated most clearly for pericytes on the 1st–4th branch orders of capillary measured from a penetrating arteriole [56, 65] which express the highest levels of  $\alpha$ -smooth muscle actin, innovations in histochemistry have revealed that even higher branch order pericytes express this contractile protein [3] and optogenetic experiments have shown that these higher branch order pericytes can also regulate capillary diameter and blood flow [[www.biorxiv.org/content/10.1101/2020.03.26.008763v1](http://www.biorxiv.org/content/10.1101/2020.03.26.008763v1)].

Functional indications that capillary pericyte-mediated control of CBF is disrupted in AD have been provided by measurements of the capillary transit time of the blood, and its heterogeneity. Magnetic resonance imaging (MRI) experiments on humans and optical imaging experiments on AD mice have found that AD leads to both a prolongation of the capillary transit time and an increase in its heterogeneity, as if some capillary pericytes became more constricted than others [38, 54]. Furthermore, in humans, these changes correlate with cognitive decline (Fig. 2b), as measured by the Brief Cognitive Status Examination [128].

By analysing images of brain biopsies of patients who consulted neurologists for dementia of unknown cause (Fig. 2c), Nortley et al. [133] demonstrated that patients developing AD have capillary blood flow restricted as a result of capillary constriction. This was shown to be due to pericytes by examining how capillary diameter varied as a function of the distance along the capillary from the pericyte soma (Fig. 2d). Patients depositing A $\beta$  and tau tangles showed a constriction at the pericyte soma relative to positions between pericytes on the capillary. This increased rapidly with the amount of A $\beta$  deposited, suggesting a CBF reduction mechanism that occurs early in the development of the disease (before accumulation of A $\beta$  in and around vascular cells—cerebral amyloid angiopathy—leads to pericyte loss), as is also seen in live imaging of CBF in AD patients [107]. In contrast, in patients lacking A $\beta$  and tau deposition, capillaries showed a larger diameter near the pericyte soma, perhaps because pericytes normally induce growth of the endothelial tube. The difference in the spatial profile of capillary diameter between AD and non-AD patients was estimated to be able to generate a reduction in CBF of ~50%, similar to that found in AD patients in vivo [5].

In AD mouse models, live cortex imaging through a cranial window, or reconstructing the hippocampal vasculature of fixed brains, also showed a reduction of mean capillary diameter compared to normal mice [55, 133, 193], which in



**Fig. 2** The role of pericytes in the physiology and Alzheimer's-related pathology of the brain circulation. **a** Schematic diagram of the vascular bed (colour of blood represents oxygenation), indicating the relative resistance in the capillaries compared to penetrating arterioles and venules, for flow from the pial surface down an arteriole to layer 4, through the capillary bed, and returning to the pial surface through a venule [49]. Capillary diameter can be adjusted by a population of pericytes (yellow) that are contractile, which are located on at least the first four branch orders (see labels) of the capillary bed [56]. Blood flowing through capillaries with pericytes that are contracting to reduce the diameter will flow more slowly and so has a longer capillary transit time than blood flowing through capillaries with relaxed pericytes, thus generating capillary transit time

heterogeneity (CTTH). **b** In patients with AD, CTTH (shown as a % change) increases as cognitive power (assessed with the Brief Cognitive Status Examination) declines (from Fig. 5A of [128], reproduced courtesy of John Wiley and Sons). **c, d** Capillary imaged in right frontal cortex biopsy from a dementing patient lacking A $\beta$  deposition (**c**) and plot of mean capillary diameter versus distance from pericyte somata (**d**) in similar patients lacking or showing A $\beta$  deposition (from Fig. 4A, D of [133]). Patients depositing A $\beta$  show a large constriction near the pericyte somata. **e** Neutrophil (green) occluding a branch (to the right) of a capillary in AD mouse cortex (from Fig. 2A of [26], reproduced courtesy of Springer Nature). **f** Reducing clotting with dabigatran in WT and AD mice (from Fig. 3B of [25], reproduced courtesy of Elsevier Press) increases CBF in AD mice

cortex reflected capillary constriction near pericyte somata [133]. Nortley et al. [133] further demonstrated that, in the AD model mouse they used, neither arterioles nor venules

had an altered diameter, implying that the reduction of CBF is generated by capillaries (although this still remains to be shown for human AD and other AD mouse models).

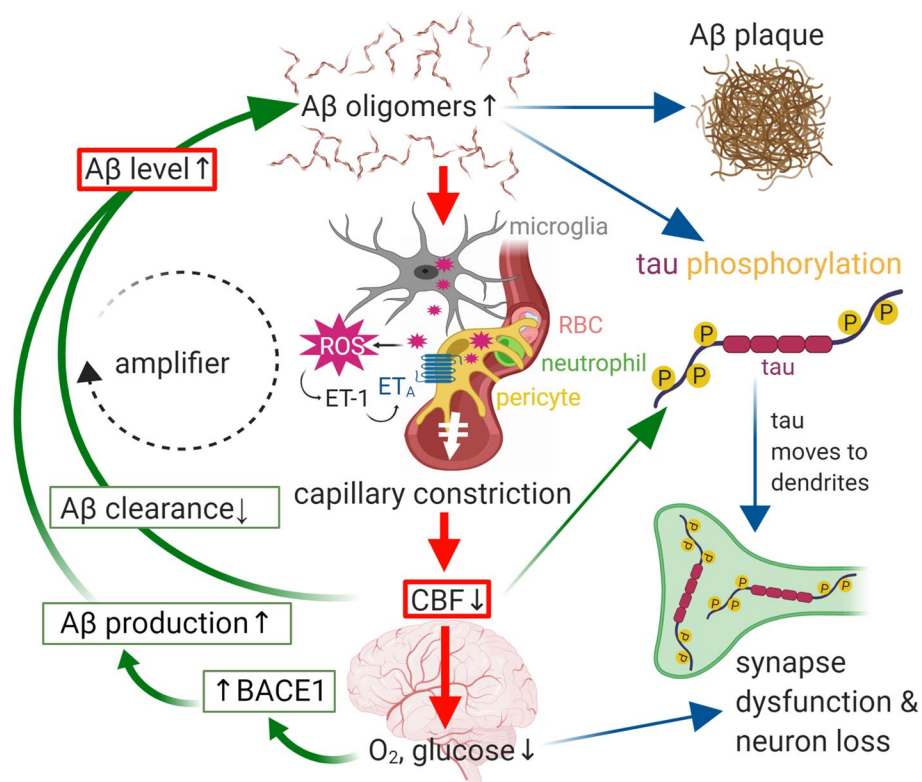


## Mechanism of CBF decrease

Although the mechanism of the long-term pericyte-mediated constriction of capillaries that occurs in human AD brains has not yet been definitively identified, short-term application of A $\beta$  oligomers (both A $\beta_{1-42}$  and A $\beta_{1-40}$ , at nanomolar concentrations similar to those present in AD) to human or rodent brain slices evoked capillary constriction [133] mediated by reactive oxygen species (ROS) generation and activation of endothelin A (ET<sub>A</sub>) receptors (Fig. 3). It is plausible that this signalling pathway is also responsible for capillary constriction in the human AD brain, since the concentrations of both ROS and endothelin-1 are known to be elevated in human AD [10, 114, 135]. The locus of ROS generation is debated, with Park et al. [141] suggesting it to be perivascular macrophages, while Nortley et al. [133] found that ROS are generated by microglia and pericytes. ET<sub>A</sub> receptors are known to

be expressed on all classes of pericyte [190] and their activation in AD is consistent with the elevated level of extracellular endothelin-1 (ET) found in post-mortem AD brains [113, 135].

Release of inflammatory mediators generated during AD may also contribute to the decrease of CBF occurring. Interleukin-1 $\beta$  is generated when microglial and astrocyte inflammasomes are activated by oligomeric A $\beta$ , and (in the context of ischaemia) this cytokine has been shown to decrease CBF by releasing ET [125], although it is unknown whether this decrease is generated by pericytes. Similarly, a mutation in the microglial TREM2 receptor (an AD susceptibility gene) that increases the production of inflammatory mediators also leads to a decrease of CBF [85]. The neuroinflammation occurring in multiple sclerosis can also be associated with hypoperfusion that is correctable by blocking ET<sub>A</sub> receptors or voltage-gated calcium channels [33, 34].



**Fig. 3** Schematic diagram showing how the amyloid beta and tau cascades can be initiated from two entry points (red boxes): (i) a decrease of cerebral blood flow (CBF) which lowers brain O<sub>2</sub> and glucose and thus upregulates the enzyme (BACE1) that makes A $\beta$  or (ii) an increase in A $\beta$  level due to more production or less clearance of A $\beta$ . A $\beta$  oligomers can aggregate into plaques, but also evoke ROS production from microglia and pericytes, which triggers the release of endothelin-1 (ET-1) from a yet-to-be-determined cell type [133].

Activation of ET<sub>A</sub> receptors on pericytes leads to capillary constriction and a decrease of CBF, lowering the levels of O<sub>2</sub> and glucose. Both a rise of A $\beta$  oligomer concentration and a fall of blood flow lead to hyperphosphorylation of tau, which relocates from axonal microtubules to dendrites, causing synapse dysfunction. Together with myelin loss this leads to cognitive decline. The fall of CBF will also contribute to impaired cognition

## The role of upstream arteries and arterioles

Constrictions of rodent cerebral arterioles and middle cerebral artery, resulting in a decrease of cerebral blood flow, have been reported to be evoked by application of exogenous  $A\beta_{1-40}$  [130, 169], but interestingly—at least in the  $APP^{NL-G-F}$  rodent model of AD—the level of  $A\beta$  that occurs in AD is sufficient only to constrict capillaries and not arterioles [133]. Nevertheless, in some AD mice, neurovascular coupling is impaired at the arteriole level [131]. Furthermore, changes in the properties of arteries and arterioles upstream of the brain's capillary beds, and of the downstream venous system, could contribute to the onset of AD. Possible contributing changes include atherosclerosis [69, 182] leading to partial occlusion of large vessels, an increase in arterial stiffness [69] and hypertension [45, 72] (discussed below) resulting in microvascular damage. It is possible that, rather than directly reducing CBF, these changes may promote  $A\beta$  generation or reduce its clearance [45, 69].

## Capillary block by neutrophils and clot formation also reduce CBF in AD

The graded constriction of capillaries by pericytes is predicted to reduce CBF by 50% even in the absence of cells in the blood [133]. In addition, two mechanisms that can produce complete occlusion of vessels have been reported to reduce CBF in AD.

By imaging cell movements in cerebral capillaries, Cruz Hernández et al. [26] observed that in AD ( $APP/PS1$ ), mice capillaries could become blocked by neutrophils (Fig. 2e). In the AD mice 1.8% of capillaries—predominantly of smaller diameter—became blocked, whereas in wild-type mice only 0.4% of capillaries were blocked. It will be important to reproduce these results in human AD patients. In wild-type mice, capillary block increases with ageing and can lead to vessels being pruned [159]. Remarkably, although modelling suggested that the increased block in AD would lead to a decrease of CBF of less than 5%, applying intraperitoneally a high concentration of an antibody to a neutrophil surface marker (Ly6G) led to a relief of capillary block, an increase of blood flow by 26–32% and improved memory. This is surprising because, at least in conditions of inflammation, antibody to Ly6G promotes neutrophil adhesion and aggregation, coagulation and decreased blood flow [132]. The large effect of the antibody on CBF compared with the modelling predictions for relief of capillary block alone may indicate either that the modelling is over-simplified or that

the antibody has effects beyond simply preventing neutrophil blocking of capillaries, perhaps on the effective viscosity of the blood (which leukocytes significantly affect [2, 16]) or on interactions with platelets and endothelial cells [110].

Cortes-Canteli et al. [25] employed long-term anticoagulation with a direct oral anticoagulant, dabigatran, to try to improve outcome in AD mice, based on the observation that excess fibrin is deposited in the AD brain, indicating an excessively prothrombotic environment. Dabigatran preserved CBF and reduced cognitive decline in AD mice (Fig. 2f). While a 15% decrease in CBF was seen at 40 weeks of age in AD mice (a smaller decrease than occurs in affected regions in human AD, possibly because cortical CBF was assessed by measuring it relative to thalamic CBF, which may itself be decreased [11]), after anticoagulation treatment from 2 months of age the CBF was raised above normal by 13%. Interestingly in humans receiving oral anticoagulants, the risk of dementia is reduced by 29% [44].

Given the profound constriction of cerebral capillaries at pericyte somata that is observed in biopsies from human patients developing AD, from a diameter of ~5 to ~2.8  $\mu\text{m}$  [133], it is attractive to hypothesize that both the block of capillaries by neutrophils and the formation of clots that reduce CBF are a consequence of the reduced diameter of capillaries near pericyte somata. Neutrophils are larger and less distensible than red blood cells and pass through capillaries more slowly [16, 37], and so may tend to become lodged at the smallest diameter parts of capillaries. Similarly, although Cortes-Canteli et al. [25] did not image the vasculature to define which vessels exhibited coagulation, the decreased flow expected through pericyte-constricted capillaries would tend to promote clotting, suggesting that thrombi forming in the smallest vessels may contribute to the reduction of CBF occurring.

## Capillary constriction and reduced CBF accelerate AD onset

The capillary constriction seen in AD leads to the neural tissue becoming hypoxic [133], which presumably contributes to the decrease in glucose metabolism observed in AD (see above). Importantly, ischaemia and hypoxia have been shown to upregulate the enzyme (BACE1) responsible for generating  $A\beta$  [168, 197], as schematised in Fig. 3. This leads to more  $A\beta$  production [168, 197], which is expected to promote neurodegeneration and cognitive decline in accordance with the amyloid hypothesis, and indeed this was found [168, 197]. While these mechanistic studies were all in animals or on cell lines expressing human BACE1, the level of BACE1 and its enzymatic activity are increased in humans suffering from AD [78], as expected from the fact

that the capillary constriction in humans developing AD is sufficient to reduce cerebral blood flow by up to 50% [133] and the animal work cited above showing that ischaemia and hypoxia upregulate BACE1. Furthermore, an upregulation of BACE1 has been found to exist in mild cognitive impairment patients, and correlates with A $\beta$  plaque number and cognitive decline [23]. The upregulation of BACE1 by ischaemia and hypoxia occurs as a result of caspase-3 both increasing BACE1 mRNA level and cleaving GGA3, an adaptor protein involved in BACE1 trafficking, to decrease BACE1 degradation [171, 184, 194], and has two conceptual implications.

Firstly, BACE1 upregulation implies that low blood flow or hypoxia—caused by a purely vascular defect, brain injury, sleep apnoea or genetic predisposition—could initiate the production of A $\beta$ . Indeed, bilateral occlusion of the carotid arteries leads to A $\beta$  production and a fall of metabolism in the amygdala, entorhinal cortex and hippocampus [140]. This could explain why subjects with sleep apnoea, or head injury that decreases CBF [155, 178], are more likely to develop AD [95, 189]. Similarly, hypertension leads to a 45% decrease of CBF in selected brain regions [27, 72, 124], and the resulting upregulation of BACE1 may contribute to A $\beta$  accumulation and the increased likelihood of suffering from AD that is associated with hypertension [72]. For the severe ischaemia produced by stroke, however, it is debated [46, 154] whether this evokes A $\beta$  deposition that contributes to the increased incidence of dementia that occurs post-stroke [117]. Some genetic variants may act by reducing CBF. The ApoE4 variant of ApoE is the main susceptibility gene for AD, and has important vascular effects. Expression of ApoE4 leads to a lower CBF even in cognitively normal subjects [111], which will tend to upregulate BACE1 and increase A $\beta$  production (see above). It also promotes accelerated loss of pericytes and consequent breakdown of the blood–brain barrier, which correlate with cognitive decline [119]. Since experimentally reducing CBF also leads to pericyte loss [41, 56, 97] and hence BBB breakdown [4, 13, 97, 118], it is unclear whether the primary effect of ApoE4 on pericytes is to make them constrict capillaries (ApoE4 is known to affect the cytoskeleton and so may affect contractility [22]) with the resulting decrease in CBF causing pericyte loss and subsequent BBB breakdown, or whether the primary effect is the loss of pericytes which somehow causes a decrease of CBF.

Secondly, once A $\beta$  production (or an imbalance between production and removal by various mechanisms described below) has been initiated, the resulting constriction of capillaries by pericytes that it initiates (see above) will reduce CBF, causing an upregulation of BACE1 and production of more A $\beta$  (Fig. 3). This positive feedback loop will amplify the production of A $\beta$ , over an as yet unknown time course, resulting in a further imbalance between A $\beta$  production and removal.

## Capillary constriction as a link between A $\beta$ and tau phosphorylation

Downstream of A $\beta$  production, an important driver of cognitive decline is tau hyperphosphorylation [57, 62, 91], which leads to tau dissociating from microtubules, aggregating and localising more in dendrites (Fig. 3). Importantly, ischaemia (or hypoxia), which is evoked by the pericyte-mediated capillary constriction that A $\beta$  evokes [133], is known to trigger tau phosphorylation [140, 145, 147]. This is unlikely to reflect solely the increase in A $\beta$  level evoked by ischaemia/hypoxia discussed above, because tau phosphorylation occurs in hypertensive rats (which are ischaemic and hypoxic) even without A $\beta$  pathology [147] and is evoked by unilateral carotid artery occlusion in AD mice without a rise in A $\beta$ <sub>1–42</sub> level [145].

Major enzymes phosphorylating tau at AD-related sites include Cdk5 (cyclin-dependent kinase 5) and GSK3 (glycogen synthase kinase 3) [52, 91, 98]. For the following reasons, these may be activated by capillary constriction which evokes ischaemia/hypoxia, and thus inhibits Ca<sup>2+</sup> pumping out of cells and raises [Ca<sup>2+</sup>]<sub>i</sub>. Cdk5 is activated when a raised [Ca<sup>2+</sup>]<sub>i</sub> activates calpain to cleave Cdk5's regulatory subunit p35 [98, 158]. GSK3 is activated by prolonged hypoxia via a decrease in activity of the phosphatidylinositol 3-kinase/Akt pathway [122, 191] and on a shorter time scale by an imipramine-sensitive mechanism [149].

Thus, the A $\beta$ -evoked reduction of CBF, produced by pericyte-mediated capillary constriction in AD, could provide an important link between the rise of extracellular A $\beta$  concentration and the hyperphosphorylation that leads to tau relocating to dendrites and impairing synaptic function (Fig. 3). Consequently, cognitive decline is likely to involve a reduction of CBF, whether the cognitive decline is produced ultimately by A $\beta$  or by tau hyperphosphorylation.

## Effect of reduced blood flow on A $\beta$ clearance and blood–brain barrier (BBB) in AD

The CNS is presumably exposed mainly to A $\beta$  generated within the CNS, rather than A $\beta$  generated peripherally and entering across the BBB (although A $\beta$  transfer in this direction is possible via the receptor for advanced glycation end products (RAGE) [32]), Consequently, the rise of CNS A $\beta$  concentration that occurs in AD depends not only on the rate of A $\beta$  production, but also on the rate at which it is enzymatically degraded within and removed from the CNS [108]. This raises the question of how A $\beta$  clearance will be affected by the up to 50% reductions of CBF that occur in affected areas [5].

Four major clearance routes for A $\beta$  from the CNS have been proposed: via efflux across endothelial cells into the blood; via bulk extracellular flow into the CSF and lymphatic vessels; via movement through the perivascular spaces of either penetrating arterioles or alternatively venules (promoted by cardiac cycle driven pulsation of arterioles and, in the case of exit along venules, also water flow through astrocytes termed the glymphatic system: see below); and via phagocytosis and subsequent degradation by microglia, astrocytes and other cells. Injections of radioactive A $\beta$  into the brain parenchyma have been used to try to quantify the relative importance of these removal mechanisms [160]. Five hours after injecting A $\beta_{1-40}$ , 84.5% of it had been cleared from the CNS and 15.5% was retained. The retained material might include A $\beta$  in the interstitial space and A $\beta$  (or breakdown products) sequestered in microglia, astrocytes and other cells. Of the removed A $\beta$ , 12.7% (i.e. 10.7% of the total injected) was removed by a process that also occurred for the inert tracer inulin, which may include all mechanisms driven by interstitial fluid flow. The remaining 87.3% of removed A $\beta$  was assumed to have exited the BBB across the endothelial cell layer of capillaries. Similar experiments showed that (at 30 min after tracer injection) 30% more A $\beta_{1-42}$  than A $\beta_{1-40}$  was retained in the brain and correspondingly less was cleared across the BBB [196]. Clearance across the BBB involved PICALM (phosphatidylinositol-binding clathrin assembly protein [196]), which is expressed in vascular endothelial cells [190], and LRP1 (low density lipoprotein receptor-related protein 1; but see [75]), which is expressed in perivascular astrocytes and pericytes and to a small extent in capillary endothelial cells (as well as neurons, microglia and oligodendrocyte precursor cells [190]). A major role for endothelial cell LRP1 in mediating A $\beta$  export is shown by knock-out work [167], but astrocyte and neuronal LRP1 may also be involved [79, 96]. There is evidence for association of PICALM and LRP1 gene variants with human AD risk (reviewed by [161] and [196]).

The decrease of CBF that occurs early in preclinical AD could decrease A $\beta$  removal across endothelial cells, thus potentiating A $\beta$  accumulation, by decreasing the level of proteins that mediate the removal. For example, ischaemia will raise [Ca<sup>2+</sup>]<sub>i</sub> which can result in calpain cleaving PICALM [150], and indeed PICALM levels are lower in human AD, correlating both with an increased A $\beta$  level and with cognitive decline as assessed with the Mini Mental State Exam [196]. Similarly, ischaemia leads to the endopeptidase furin cleaving LRP1 [185]. Additionally, a slowing of capillary blood flow could in principle allow A $\beta$  that has exited into the blood to re-enter the brain parenchyma by RAGE-mediated entry across endothelial cells [32], thus again slowing net removal of A $\beta$ .

The CBF decrease in AD is also expected to alter A $\beta$  removal by the other, apparently quantitatively less important [66, 160], mechanisms mentioned above. Pulsation of penetrating arterioles during the cardiac cycle or spontaneous vasomotion has been postulated to power the removal of A $\beta$  (in a retrograde direction with respect to CBF) in the perivascular spaces of penetrating arterioles [36, 156]. Arteriole pulsation is also presumed to promote water flow along the paravascular spaces of arterioles and through both aquaporin-4-expressing glial cells and the extracellular space of the brain [73, 74]. This flow may reach: (i) venules, where it helps to remove A $\beta$  in the perivascular spaces of venules (in the same direction as CBF [73]), and (ii) the CSF and lymphatic vessels [6, 100, 103, 137]. A detailed analysis of these proposals has been provided [66, 164]. In AD, when CBF decreases, decreased pulsatility of the middle cerebral artery has been reported [134] and so, if this extends to penetrating arterioles, less A $\beta$  removal by pulsation-driven mechanisms would be expected. Indeed, removal of A $\beta$  by the CSF, lymphatic and glymphatic systems decreases in AD [88, 142], possibly with contributing factors including increased stiffening of the arterioles with age [179] and ischaemia-induced changes of other key components such as decreased lymphatic function and aquaporin 4 localisation away from astrocyte endfeet abutting blood vessels [28, 88, 186].

The CBF decrease induced by capillary constriction in AD may also alter microglial and astrocyte removal and degradation of A $\beta$ . Ischaemia followed by reperfusion (which may mimic the prolonged decrease of CBF occurring in AD) decreases microglial ramification [106, 121], which could decrease A $\beta$  removal by these cells as surveillance of the brain parenchyma will be reduced [104]. On the other hand, ischaemia upregulates expression of triggering receptor expressed on myeloid cells-2 (TREM2), which is a key molecule by which microglia recognise A $\beta$  and remove it [138, 195], as well as other phagocytosis-related genes [192], suggesting an enhanced ability to remove A $\beta$  by microglia. Similarly ischaemia upregulates ABCA1, MEGF10 and GULP1, which are components of an astrocytic phagocytosis pathway [120], suggesting that the CBF reduction occurring in AD may also enhance A $\beta$  removal by astrocytes [[www.biorxiv.org/content/10.1101/2020.03.29.002857v1](https://www.biorxiv.org/content/10.1101/2020.03.29.002857v1)].

Although this review focuses on the effects of the reduction of CBF that is induced by pericyte-mediated capillary constriction in AD, pericytes themselves are very sensitive to ischaemia [41, 56]. In AD the reduction of CBF, together with intracellular accumulation of A $\beta$  in pericytes [181], may eventually lead to pericyte death [41, 56], which will lead to a loss of BBB function [4, 13, 118, 126] that promotes neurodegeneration [153].



## The role of white matter CBF changes in the onset of AD

Although most attention in the AD field focuses on changes in the grey matter, the A $\beta$  level also increases in the white matter in AD [24], and the CBF decrease early in AD occurs in the white matter as well as the grey matter [80]. Consequently, the CBF decrease might exert some of its effects by generating white matter dysfunction, such as slower action potential propagation. White matter tissue is lost before grey matter tissue in AD [30], and early in AD white matter abnormalities defined by MRI correlate both with cognitive decline and with reduced CBF in the deep and circumventricular white matter [18, 77, 93]. Surprisingly, however, white matter capillary diameter has been reported to increase in AD [61]. These results suggest that it will be important to determine whether, in preclinical human AD, capillary constriction by pericytes occurs in the white matter, as in the grey matter [133], or whether CBF decreases as a result of upstream vessel constriction in the grey matter [101] (possibly with dilation of white matter capillaries as an adaptive response) or for some other reason, and to establish precisely which downstream mechanisms (such as myelin loss [116]) lead to white matter dysfunction early in AD.

## Implications for therapeutic approaches to Alzheimer's disease

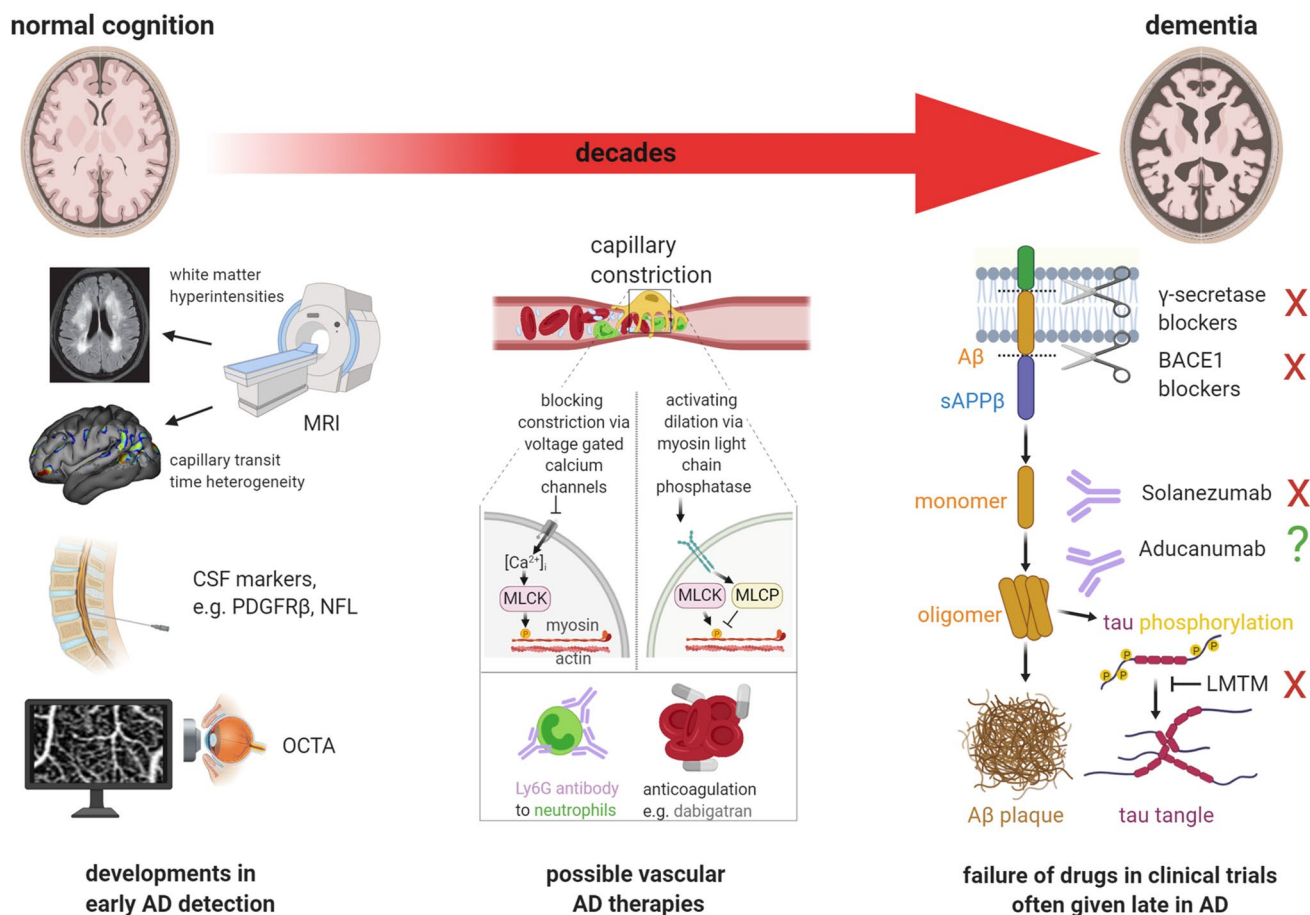
The discoveries that the decrease of CBF in AD occurs early in the disease [76], and is caused by impaired capillary regulation of CBF [26, 54, 128, 133], are consistent with the proposal that impaired capillary blood flow contributes to the onset of AD [31] made soon after the amyloid hypothesis of AD was proposed [59]. These data, including the demonstration that A $\beta$  itself can trigger pericyte-mediated capillary constriction [133], reconcile genetic evidence for the involvement of A $\beta$  in AD with the fact that the first change seen in AD is a decrease of cerebral blood flow [76], and open up new potential therapeutic approaches for this disease. Conceivably, maintaining CBF may prevent cognitive decline if interventions are made early enough to avoid neuronal and glial damage. Just as the risk of stroke is now reduced by giving blood pressure lowering drugs prophylactically, we expect the long-term future of AD therapy to involve—at least partly—prophylactic agents that prevent pericyte-mediated capillary constriction, and thus prevent both direct effects of CBF decreases and the amplification of A $\beta$  production and tau phosphorylation that a fall in CBF generates.

Below, we consider approaches to achieving this and possible biomarkers to use to decide when prophylaxis should be initiated (Fig. 4).

## Preventing pericyte-mediated capillary constriction

The constriction of capillaries by pericytes may be mediated by A $\beta$  evoking the generation of ROS that trigger the release of endothelin-1 (ET), which activates [Ca<sup>2+</sup>]<sub>i</sub>-elevating contractile ET<sub>A</sub> receptors on pericytes [133]. Indeed, in short-term experiments, blocking ROS production and ET<sub>A</sub> receptors prevented development of further A $\beta$ -evoked constriction [133]. However, long-term block of ROS generation is undesirable because ROS are used for signalling in many contexts, as well as for immune defence mechanisms. Furthermore, ET<sub>A</sub> receptor activation is difficult to reverse with blockers [64], and although there is a BBB-permeable ET<sub>A</sub> receptor blocker licenced for clinical use (clazosentan for sub-arachnoid haemorrhage), side effects make this drug unsuitable for long-term administration [175].

A better approach to preventing capillary constriction may therefore be to inhibit the contractile pathways downstream of ET<sub>A</sub> receptors by blocking the release of Ca<sup>2+</sup> from internal stores and increasing the activity of myosin light chain phosphatase to activate relaxation of the contractile filaments. These twin aims can be achieved by using an agonist of guanylate cyclase receptors, such as C-type natriuretic peptide (CNP, [166]). Indeed, CNP rapidly reverses A $\beta$ -evoked constriction of capillaries in brain tissue [133]. An alternative approach is to relax pericytes by inhibiting their voltage-gated Ca<sup>2+</sup> channels (VGCCs). Interestingly, comparing different classes of drugs used to reduce hypertension, it has been claimed that only VGCC blockers slow the progression to dementia in AD ([102], see also [183]), although not all VGCC blockers used for hypertension cross the BBB well and there are numerous mechanisms by which they may slow cognitive decline [90]. One BBB-permeable VGCC inhibitor, nilvadipine, has been shown to restore the CBF of AD mice to normal levels [139]. In human AD, although nilvadipine lowers peripheral blood pressure, it increases CBF in the hippocampus [29], presumably by relaxing pericytes, and shows some slowing of cognitive decline in very mild AD patients [1]. Devising ways of targeting VGCC blockers specifically to CNS pericytes might enhance the efficacy of this approach. Firstly, it would be desirable to avoid inhibiting VGCCs in neurons, which might be achievable by using bivalent drugs that also bind to proteins expressed relatively specifically by pericytes, such as PDGFR $\beta$ . Secondly, if it were possible to avoid inhibiting VGCCs in pericytes and smooth muscle cells around peripheral blood vessels, this would probably avoid the decrease in blood pressure that stems from relaxing the vasculature all over the body.



**Fig. 4** Interventions to diagnose and reduce cognitive decline at different stages of the transition from normal cognition to dementia in AD. Right third of figure: most clinical trials are initiated at relatively late stages of the disease, when cognitive decline is already apparent, and irreversible synapse or neuron loss may have taken place. This may explain why drugs that block the  $\gamma$  or  $\beta$  secretases, antibodies to different forms of A $\beta$ , and a drug that blocks tau aggregation (LMTM) have all failed (red crosses) to stop cognitive decline in AD. Left third of figure: emerging diagnostic approaches for early detection of AD include MRI assessment of white matter hyperintensities (image from Fig. 1B of [93], reproduced courtesy of Dove Medical Press) and capillary transit time heterogeneity (from Fig. 5E of [128], reproduced courtesy of John Wiley & Sons), assessment of biomark-

ers in the CSF such as PDGFR $\beta$  and neurofilament light chain (NFL), and non-invasive capillary imaging in the retina using (e.g.) optical coherence tomography angiography (OCTA). Middle third of figure: potential therapies to prevent or reverse the CBF decrease arising when  $Ca^{2+}$  activates myosin light chain kinase (MLCK) to evoke pericyte-mediated capillary constriction. These include blocking pericyte voltage-gated calcium channels to block  $Ca^{2+}$ -evoked constriction, raising pericyte cGMP level (by activating guanylate cyclase receptors, blue membrane protein) to stimulate myosin light chain phosphate (MLCP) and thus evoke dilation, disrupting neutrophil surface interactions with endothelial cells or other cells using antibodies (if this approach can be used without inducing neutropenia), or blocking thrombus formation with dabigatran [25, 26, 133]

### Preventing neutrophils occluding capillaries

As noted above, Cruz Hernández et al. [26] showed that, in AD mice, brief application of an antibody to the Ly6G protein on neutrophils increased CBF by 30% and improved memory, although in aged AD mice the cognitive effect was absent, presumably because too much synaptic damage had occurred by that stage [15]. The improvement of cognition in parallel with the increase in CBF in younger AD mice strongly supports the concept of devising interventions to preserve CBF in AD. However, prolonged application of antibody to Ly6G leads to very significant

neutropenia (a depletion of neutrophils) within hours [26], which will engender a heightened risk of infection, and thus is not suitable as a long-term therapy (this approach has not yet been used in humans). Thus, further research is needed to devise an agent which generates the blood flow increasing effect of Ly6G (which may be via more than one mechanism: see above) without causing neutropenia. As with the approach of targeting pericyte-mediated capillary constriction discussed above, it will also be necessary to consider the overall effect on blood pressure caused by a manipulation that decreases vascular resistance throughout the body.

## Use of anticoagulating agents

The prolonged use of anticoagulants to improve cerebral blood flow and outcome in patients liable to developing AD [25] might lead to an increased risk of intracranial haemorrhage. AD often coexists with cerebral amyloid angiopathy (CAA), for which asymptomatic micro-bleeds, bleeding into the cortical sulci and large symptomatic lobar cerebral haemorrhages can be complications. These are thought to be due to a breakdown in microvasculature integrity as A $\beta$  accumulates along vessel walls and injures them [50]. Criteria exist for diagnosing CAA [51], and detection of intracerebral haemorrhage (including micro-bleeds) has been greatly enhanced by T2\*-weighted MRI imaging sequences with a high sensitivity for bleeding [50]. However, further research is required to determine whether there are specific CAA-related biomarkers that would help clinicians to recognise and exclude those patients who would be put at an unacceptable risk of serious intracerebral haemorrhage from anticoagulation, before it could be adopted as a widespread prophylactic treatment for AD.

## Relevance of these approaches to other neurodegenerative disorders

The A $\beta$ -evoked constriction of capillaries by pericytes may involve ROS generation that evokes the release of endothelin-1 [133]. ROS generation also occurs when  $\alpha$ -synuclein accumulates in Parkinson's disease (PD) and Lewy body dementia (LBD) [12, 14], and may evoke ET release and constrict capillaries as for AD. Indeed, PD and LBD are associated with decreased cerebral blood flow [42, 170]. Accordingly, the therapeutic approaches outlined above may also be relevant to these disorders.

## Choice of biomarker for initiating treatment

To date, candidate treatments for AD have almost certainly been initiated too late, after irreversible damage to the brain has occurred, as a result of making treatment decisions based on significant observable cognitive decline. If we are to move towards more preventative treatments, they will need to be started as soon as the earliest changes occur in the disease, raising the question of what biomarkers to use to trigger treatment. Assuming that pericyte-mediated capillary constriction is indeed a very early event in the onset of AD (see Fig. 3) as suggested by Iturria-Medina et al. [76] and Nortley et al. [133], it will become essential to develop non-invasive tests to detect the onset of capillary constriction near pericytes. Markers of cell damage, such as CSF levels of neurofilament light chain which may indicate damage to white matter axons [35] or PDGFR $\beta$  for pericytes [118], while useful for assessing the extent

of neurologically relevant damage, may only be detectable too late for initiating a preventative drug strategy.

Techniques that look directly at deleterious decreases of CBF (which may follow a period of adaptive hyperperfusion in some brain regions [40, 53, 180]), and its capillary control, may therefore be preferable. In human patients, MRI can be used to measure CBF. Dynamic susceptibility contrast MRI with an injected tracer has been used to quantify changes of blood capillary transit time (and its heterogeneity) in early AD [38, 128], which we argue above probably reflect pericyte-mediated constriction of capillaries. If these measurements could be performed using non-invasive (i.e. without an injected tracer) arterial spin label MRI, then it would provide a method to assess changes in how pericytes control blood flow in different capillaries. An alternative, more direct, observation of pericyte-mediated capillary constriction may be possible by imaging retinal capillaries through the intact cornea, using optical coherence tomography angiography (OCTA), which has been used to detect decreases in neurovascular coupling at the arterial level [146]. OCTA could perhaps thus provide a screening method for detecting pericyte malfunction early in preclinical AD. A $\beta$  plaques are reported to be deposited in the retina before being deposited in the brain [86]. Thus, pericyte-mediated capillary constriction evoked by A $\beta$  oligomers should also be detectable early on as a focal reduction of capillary diameter around pericytes (cf [133]), although this reduction is likely to be close to the limit of resolution of the OCTA technique and this approach would require validation with post-mortem immunohistochemistry.

## Conclusions

With the discoveries that a decrease of cerebral blood flow is the earliest change to occur in AD [76], that this is generated at the capillary level [26, 38, 133] and that changes in capillary control of CBF correlate with cognitive decline [128], it is becoming impossible to ignore the vascular contribution to Alzheimer's disease. The reduction of CBF produced by pericytes constricting capillaries, along with ensuing decreases in CBF as a result of capillary occlusion by neutrophils and thrombi, is an important dysfunction in AD that potentially opens up new therapeutic approaches and new screening possibilities. Initial evidence indicates that reversing this reduction of CBF can restore cognitive function, provided that damage to synapses, neurons and circuits has not advanced significantly. Consequently, in addition to manipulation of other effects of A $\beta$  and tau, devising screening tests to allow therapeutic intervention to maintain CBF should be a key aim for the future treatment of AD.



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## References

- Abdullah L, Crawford F, Tsolaki M, Börjesson-Hanson A, Olde Rikkert M, Pasquier F et al (2020) The Influence of baseline Alzheimer's disease severity on cognitive decline and CSF biomarkers in the NILVAD trial. *Front Neurol* 11:149. <https://doi.org/10.3389/fneur.2020.00149>
- Akenhead ML, Horrall NM, Rowe D, Sethu P, Shin HY (2015) In vitro evaluation of the link between cell activation state and its rheological Impact on the microscale flow of neutrophil suspensions. *J Biomech Eng* 137:91003. <https://doi.org/10.1115/1.4030824>
- Alarcon-Martinez L, Yilmaz-Ozcan S, Yemisci M, Schallek J, Kilic K, Can A et al (2018) Capillary pericytes express  $\alpha$ -smooth muscle actin, which requires prevention of filamentous-actin depolymerization for detection. *Elife* 7:e34861. <https://doi.org/10.7554/eLife.34861.001>
- Armulik A, Genové G, Mäe M, Nisancioglu MH, Wallgard E, Niaudet C et al (2010) Pericytes regulate the blood-brain barrier. *Nature* 468:557–561. <https://doi.org/10.1038/nature09522>
- Asllani I, Habeck C, Scarmeas N, Borogovac A, Brown TR, Stern Y (2008) Multivariate and univariate analysis of continuous arterial spin labeling perfusion MRI in Alzheimer's disease. *J Cereb Blood Flow Metab* 28:725–736. <https://doi.org/10.1038/sj.jcbfm.9600570>
- Aspelund A, Antila S, Proulx ST, Karlsen TV, Karaman S, Detmar M et al (2015) A dural lymphatic vascular system that drains brain interstitial fluid and macromolecules. *J Exp Med* 212:991–999. <https://doi.org/10.1084/jem.20142290>
- Attems J, Lintner F, Jellinger KA (2004) Amyloid beta peptide 1–42 highly correlates with capillary cerebral amyloid angiopathy and Alzheimer disease pathology. *Acta Neuropathol* 107:283–291. <https://doi.org/10.1007/s00401-004-0822-6>
- Attwell D, Laughlin SB (2001) An energy budget for signaling in the grey matter of the brain. *J Cereb Blood Flow Metab* 21:1133–1145. <https://doi.org/10.1097/00004647-200110000-00001>
- Attwell D, Mishra A, Hall CN, O'Farrell FM, Dalkara T (2016) What is a pericyte? *J Cereb Blood Flow Metab* 36:451–455. <https://doi.org/10.1177/0271678X15610340>
- Barnham KJ, Masters CL, Bush CI (2004) Neurodegenerative diseases and oxidative stress. *Nat Rev Drug Discov* 3:205–214. <https://doi.org/10.1038/nrd1330>
- Beason-Held LL, Goh JO, An Y, Kraut MA, O'Brien RJ, Ferrucci L et al (2013) Changes in brain function occur years before the onset of cognitive impairment. *J Neurosci* 33:18008–18014. <https://doi.org/10.1523/JNEUROSCI.1402-13.2013>
- Belarbi K, Cuvelier E, Destée A, Gressier B, Chartier-Harlin M-C (2017) NADPH oxidases in Parkinson's disease: a systematic review. *Mol Neurodegener* 12:84. <https://doi.org/10.1186/s13024-017-0225-5>
- Bell RD, Winkler EA, Sagare AP, Singh I, LaRue B, Deane R et al (2010) Pericytes control key neurovascular functions and neuronal phenotype in the adult brain and during brain aging. *Neuron* 68:409–427. <https://doi.org/10.1016/j.neuron.2010.09.043>
- Bosco DA, Fowler DM, Zhang Q, Nieva J, Powers ET, Wentworth P et al (2006) Elevated levels of oxidized cholesterol metabolites in Lewy body disease brains accelerate alpha-synuclein fibrilization. *Nat Chem Biol* 2:249–253. <https://doi.org/10.1038/nchembio782>
- Bracko O, Njiru BN, Swallow M, Ali M, Haft-Javaherian M, Schaffer CB (2019) Increasing cerebral blood flow improves cognition into late stages in Alzheimer's disease mice. *J Cereb Blood Flow Metab* 40:1441–1452. <https://doi.org/10.1177/0271678X19873658>
- Braide M, Amundson B, Chien S, Bagge U (1984) Quantitative studies on the influence of leukocytes on the vascular resistance in a skeletal muscle preparation. *Microvasc Res* 27:331–352. [https://doi.org/10.1016/0026-2862\(84\)90064-5](https://doi.org/10.1016/0026-2862(84)90064-5)
- Bressi S, Volontè M, Alberoni M, Canal N, Franceschi M (1992) Transcranial Doppler sonography in the early phase of Alzheimer's disease. *Dement Geriatr Cogn Discord* 3:25–31. <https://doi.org/10.1177/0271678X19873658>
- Brickman AM, Provenzano FA, Muraskin J, Manly JJ, Blum S, Apa Z et al (2012) Regional white matter hyperintensity volume, not hippocampal atrophy, predicts incident Alzheimer disease in the community. *Arch Neurol* 69:1621–1627. <https://doi.org/10.1001/archneurol.2012.1527>
- Busche MA, Chen X, Henning HA, Reichwald J, Staufenbiel M, Sakmann B et al (2012) Critical role of soluble amyloid- $\beta$  for early hippocampal hyperactivity in a mouse model of Alzheimer's disease. *Proc Natl Acad Sci USA* 109:8740–8745. <https://doi.org/10.1073/pnas.1206171109>
- Busche MA, Eichhoff G, Adelsberger H, Abramowski D, Wiederhold K-H, Haass C et al (2008) Clusters of hyperactive neurons near amyloid plaques in a mouse model of Alzheimer's disease. *Science* 321:1686–1689. <https://doi.org/10.1126/science.1162844>
- Busche MA, Wegmann S, Dujardin S, Commins C, Schiantarelli J, Klickstein N et al (2019) Tau impairs neural circuits, dominating amyloid- $\beta$  effects, in Alzheimer models in vivo. *Nat Neurosci* 22:57–64. <https://doi.org/10.1038/s41593-018-0289-8>
- Casey CS, Atagi Y, Yamazaki Y, Shinohara M, Tachibana M, Fu Y et al (2015) Apolipoprotein E inhibits cerebrovascular motility through a RhoA protein-mediated pathway. *JBiol Chem* 290:14208–14217. <https://doi.org/10.1074/jbc.M114.625251>
- Cheng X, He P, Lee T, Yao H, Li R, Shen Y (2014) High activities of BACE1 in brains with mild cognitive impairment. *Am J Pathol* 184:141–147. <https://doi.org/10.1016/j.ajpat.2013.10.002>
- Collins-Praino LE, Francis YI, Griffith EY, Wiegman AF, Urbach J, Lawton A et al (2014) Soluble amyloid beta levels are elevated in the white matter of Alzheimer's patients, independent of cortical plaque severity. *Acta Neuropathol Commun* 2:83. <https://doi.org/10.1186/s40478-014-0083-0>



25. Cortes-Canteli M, Kruyer A, Fernandez-Nueda I, Marcos-Diaz A, Ceron C, Richards AT et al (2019) Long-term dabi-gatran treatment delays Alzheimer's disease pathogenesis in the TgCRND8 mouse model. *J Am Coll Cardiol* 74:1910–1923. <https://doi.org/10.1016/j.jacc.2019.07.081>
26. Cruz Hernández JC, Bracko O, Kersbergen CJ, Muse V, Haft-Javaherian M, Berg M et al (2019) Neutrophil adhesion in brain capillaries reduces cortical blood flow and impairs memory function in Alzheimer's disease mouse models. *Nat Neurosci* 22:413–420. <https://doi.org/10.1038/s41593-018-0329-4>
27. Dai W, Lopez OL, Carmichael OT, Becker JT, Kuller LH, Gach HM (2008) Abnormal regional cerebral blood flow in cognitively normal elderly subjects with hypertension. *Stroke* 39:349–354. <https://doi.org/10.1161/STROKEAHA.107.495457>
28. Da Mesquita S, Louveau A, Vaccari A, Smirnov I, Cornelison RC, Kingsmore KM et al (2018) Functional aspects of meningeal lymphatics in ageing and Alzheimer's disease. *Nature* 560:185–191. <https://doi.org/10.1038/s41586-018-0368-8>
29. de Jong DLK, de Heus RAA, Rijpma A, Donders R, Olde Rikkert MGM, Günther M et al (2019) Effects of nilvadipine on cerebral blood flow in patients With Alzheimer disease. *Hypertension* 74:413–420. <https://doi.org/10.1161/HYPERTENSIONAHA.119.12892>
30. de la Monte SM (1989) Quantitation of cerebral atrophy in pre-clinical and end-stage Alzheimer's disease. *Ann Neurol* 25:450–459. <https://doi.org/10.1002/ana.410250506>
31. de la Torre JC, Mussivand T (1993) Can disturbed brain micro-circulation cause Alzheimer's disease? *Neurol Res* 15:146–153. <https://doi.org/10.1080/01616412.1993.11740127>
32. Deane R, Du Yan S, Subramanyam RK, LaRue B, Jovanovic S, Hogg E et al (2003) RAGE mediates amyloid-beta peptide transport across the blood-brain barrier and accumulation in brain. *Nat Med* 9:907–913. <https://doi.org/10.1038/nm890>
33. Desai RA, Davies AL, Del Rossi N, Tachrount M, Dyson A, Gustavson B et al (2020) Nimodipine reduces dysfunction and demyelination in models of multiple sclerosis. *Ann Neurol* 88:123–136. <https://doi.org/10.1002/ana.25749>
34. D'haeseleer M, Beelen R, Fierens Y, Cambron M, Vanbinst AM, Verborgh C et al (2013) Cerebral hypoperfusion in multiple sclerosis is reversible and mediated by endothelin-1. *Proc Natl Acad Sci USA* 110:5654–5658. <https://doi.org/10.1073/pnas.1222560110>
35. Dhiman K, Gupta VB, Villemagne VL, Eratne D, Graham PL, Fowler C et al (2020) Cerebrospinal fluid neurofilament light concentration predicts brain atrophy and cognition in Alzheimer's disease. *Alzheimers Dement* 12:e12005. <https://doi.org/10.1002/dad2.12005>
36. Diem AK, MacGregor Sharp M, Gatherer M, Bressloff NW, Carare RO, Richardson G (2017) Arterial pulsations cannot drive intramural periarterial drainage: significance for Aβ drainage. *Front Neurosci* 11:475. <https://doi.org/10.3389/fnins.2017.00475>
37. Engler RL, Dahlgren MD, Morris DD, Peterson MA, Schmid-Schönbein GW (1986) Role of leukocytes in response to acute myocardial ischemia and reflow in dogs. *Am J Physiol* 251:H314–323. <https://doi.org/10.1152/ajpheart.1986.251.2.H314>
38. Eskildsen SF, Gyldensted L, Nagenthiraja K, Nielsen RB, Hansen MB, Dalby RB et al (2017) Increased cortical capillary transit time heterogeneity in Alzheimer's disease: a DSC-MRI perfusion study. *Neurobiol Aging* 50:107–118. <https://doi.org/10.1016/j.neurobiolaging.2016.11.004>
39. Esparza TJ, Zhao H, Cirrito JR, Cairns NJ, Bateman RJ, Holtzman DM et al (2013) Amyloid-β oligomerization in Alzheimer dementia versus high-pathology controls. *Ann Neurol* 73:104–119. <https://doi.org/10.1002/ana.23748>
40. Fazlollahi A, Calamante F, Liang X, Bourgeat P, Raniga P, Dore V et al (2020) Increased cerebral blood flow with increased amyloid burden in the preclinical phase of Alzheimer's disease. *J Magn Reson Imaging* 51:505–513. <https://doi.org/10.1002/jmri.26810>
41. Fernandez-Klett F, Potas JR, Hilpert D, Blazej K, Radke J, Huck J et al (2013) Early loss of pericytes and perivascular stromal cell-induced scar formation after stroke. *J Cereb Blood Flow Metab* 33:428–439. <https://doi.org/10.1038/jcbfm.2012.187>
42. Firbank MJ, Colloby SJ, Burn DJ, McKeith IG, O'Brien JT (2003) Regional cerebral blood flow in Parkinson's disease with and without dementia. *Neuroimage* 20:1309–1319. [https://doi.org/10.1016/S1053-8119\(03\)00364-1](https://doi.org/10.1016/S1053-8119(03)00364-1)
43. Fowler JC (1990) Adenosine antagonists alter the synaptic response to in vitro ischemia in the rat hippocampus. *Brain Res* 509:331–334. [https://doi.org/10.1016/0006-8993\(90\)90560-X](https://doi.org/10.1016/0006-8993(90)90560-X)
44. Friberg L, Rosenqvist M (2018) Less dementia with oral anticoagulation in atrial fibrillation. *Eur Heart J* 39:453–460. <https://doi.org/10.1093/eurheartj/ehx579>
45. Fulop GA, Tarantini S, Yabluchanskiy A, Molnar A, Prodan CI, Kiss T et al (2019) Role of age-related alterations of the cerebral venous circulation in the pathogenesis of vascular cognitive impairment. *Am J Physiol Heart Circ Physiol* 316:H1124–H1140. <https://doi.org/10.1152/ajpheart.00776.2018>
46. Garcia-Alloza M, Gregory J, Kuchibhotla JV, Fine S, Wei Y, Ayata C et al (2011) Cerebrovascular lesions induce transient β-amyloid deposition. *Brain* 134:3697–3707. <https://doi.org/10.1093/brain/awr300>
47. Gauthier S, Feldman HH, Schneider LS, Wilcock GK, Frisoni GB, Hardlund JH et al (2016) Efficacy and safety of tau-aggregation inhibitor therapy in patients with mild or moderate Alzheimer's disease: a randomised, controlled, double-blind, parallel-arm, phase 3 trial. *Lancet* 388:2873–2884. [https://doi.org/10.1016/S0140-6736\(16\)31275-2](https://doi.org/10.1016/S0140-6736(16)31275-2)
48. Girouard H, Iadecola C (2006) Neurovascular coupling in the normal brain and in hypertension, stroke, and Alzheimer disease. *J Appl Physiol* 100:328–335. <https://doi.org/10.1152/jappphysiol.00966.2005>
49. Gould IG, Tsai P, Kleinfeld D, Linninger A (2017) The capillary bed offers the largest hemodynamic resistance to the cortical blood supply. *J Cereb Blood Flow Metab* 37:52–68. <https://doi.org/10.1177/0271678X16671146>
50. Greenberg SM, Bacskai BJ, Hernandez-Guillamon M, Pruzin J, Sperling R, van Velew SJ (2020) Cerebral amyloid angiopathy and Alzheimer disease—one peptide, two pathways. *Nat Rev Neurol* 16:30–42. <https://doi.org/10.1038/s41582-019-0281-2>
51. Greenberg SM, Charidimou A (2018) Diagnosis of cerebral amyloid angiopathy: evolution of the Boston criteria. *Stroke* 49:491–497. <https://doi.org/10.1161/STROKEAHA.117.016990>
52. Guo T, Noble W, Hanger DP (2017) Roles of tau protein in health and disease. *Acta Neuropathol* 133:665–704. <https://doi.org/10.1007/s00401-017-1707-9>
53. Guo Y, Li X, Zhang M, Chen N, Wu S, Lei J et al (2019) Age- and brain region-associated alterations of cerebral blood flow in early Alzheimer's disease assessed in AβPPSWE/PS1ΔE9 transgenic mice using arterial spin labeling. *Mol Med Rep* 19:3045–3052. <https://doi.org/10.3892/mmr.2019.9950>
54. Gutiérrez-Jiménez E, Angley H, Rasmussen PM, West MJ, Catalini L, Iversen NK et al (2018) Disturbances in the control of capillary flow in an aged APPswe/PS1ΔE9 model of Alzheimer's disease. *Neurobiol Aging* 62:82–94. <https://doi.org/10.1016/j.neurobiolaging.2017.10.006>
55. Haft-Javaherian M, Fang L, Muse V, Schaffer CB, Nishimura N, Sabuncu MR (2019) Deep convolutional neural networks for segmenting 3D in vivo multiphoton images of vasculature in Alzheimer disease mouse models. *PLoS ONE* 14:e0213539. <https://doi.org/10.1371/journal.pone.0213539>

56. Hall CN, Reynell C, Gesslein B, Hamilton NB, Mishra A, Sutherland BA et al (2014) Capillary pericytes regulate cerebral blood flow in health and disease. *Nature* 508:55–60. <https://doi.org/10.1038/nature13165>
57. Hanseeuw BJ, Betensky RA, Jacobs HIL, Schultz AP, Sepulcre J, Becker JA et al (2019) Association of amyloid and tau with cognition in preclinical Alzheimer disease: a longitudinal study. *JAMA Neurol* 76:915–924. <https://doi.org/10.1001/jamaneurol.2019.1424>
58. Hansra GK, Popov G, Banaczek PO, Vogiatzis M, Jegathees T, Goldbury CS et al (2019) The neuritic plaque in Alzheimer's disease: perivascular degeneration of neuronal processes. *Neurobiol Aging* 82:88–101. <https://doi.org/10.1016/j.neurobiolaging.2019.06.009>
59. Hardy JA, Higgins GA (1992) Alzheimer's disease: the amyloid cascade hypothesis. *Science* 256:184–185. <https://doi.org/10.1126/science.1566067>
60. Harkany T, Abrahám I, Timmerman W, Laskay G, Tóth B, Sasvári M et al (2000) beta-amyloid neurotoxicity is mediated by a glutamate-triggered excitotoxic cascade in rat nucleus basalis. *Eur J Neurosci* 12:2735–2745. <https://doi.org/10.1046/j.1460-9568.2000.00164.x>
61. Hase Y, Ding R, Harrison G, Hawthorne E, King A, Gettings S et al (2019) White matter capillaries in vascular and neurodegenerative dementias. *Acta Neuropathol Commun* 7:16. <https://doi.org/10.1186/s40748-019-0666-x>
62. He Z, Guo JL, McBride JD, Narasimhan S, Kim H, Changolkar L et al (2018) Amyloid- $\beta$  plaques enhance Alzheimer's brain tau-seeded pathologies by facilitating neuritic plaque tau aggregation. *Nat Med* 24:29–38. <https://doi.org/10.1038/nm.4443>
63. Hijazi S, Heistek TS, Scheltens P, Neumann U, Shimshek DR, Mansvelder HD et al (2019) Early restoration of parvalbumin interneuron activity prevents memory loss and network hyperexcitability in a mouse model of Alzheimer's disease. *Mol Psychiatry*. <https://doi.org/10.1038/s41380-019-0483-4>. **(Online ahead of print)**
64. Hilal-Dandan R, Villegas S, Gonzalez A, Brunton LL (1997) The quasi-irreversible nature of endothelin binding and G protein-linked signaling in cardiac myocytes. *J Pharmacol Exp Ther* 281:267–273. <https://jpet.aspetjournals.org/content/jpet/281/1/267.full.pdf>. Accessed 20 Aug 2020
65. Hill RA, Tong L, Yuan P, Murikinati S, Gupta S, Grutzendler J (2015) Regional blood flow in the normal and ischemic brain is controlled by arteriolar smooth muscle cell contractility and not by capillary pericytes. *Neuron* 87:95–110. <https://doi.org/10.1016/j.neuron.2015.06.001>
66. Hladky SB, Barrand MA (2018) Elimination of substances from the brain parenchyma: efflux via perivascular pathways and via the blood-brain barrier. *Fluids Barriers CNS* 15:30. <https://doi.org/10.1186/s12987-018-0113-6>
67. Hoover BR, Reed MN, Su J, Penrod RD, Kotilinek LA, Grant MK et al (2010) Tau mislocalization to dendritic spines mediates synaptic dysfunction independently of neurodegeneration. *Neuron* 68:1067–1081. <https://doi.org/10.1016/j.neuron.2010.11.030>
68. Howard R, Liu KY (2020) Questions EMERGE as Biogen claims aducanumab turnaround. *Nat Rev Neurol* 16:63–64. <https://doi.org/10.1038/s41582-019-0295-9>
69. Hughes TM, Craft S, Lopez OL (2015) Review of 'the potential role of arterial stiffness in the pathogenesis of Alzheimer's disease'. *Neurodegener Dis Manag* 5:121–135. <https://doi.org/10.2217/nmt.14.53>
70. Iaccarino HF, Singer AC, Martorell AJ, Rudenko A, Gao F, Gillingham TZ et al (2016) Gamma frequency entrainment attenuates amyloid load and modifies microglia. *Nature* 540:230–235. <https://doi.org/10.1038/nature20587>
71. Iadecola C (2004) Neurovascular regulation in the normal brain and in Alzheimer's disease. *Nat Rev Neurosci* 5:347–360. <https://doi.org/10.1038/nrn1387>
72. Iadecola C, Gottesman RF (2019) Neurovascular and cognitive dysfunction in hypertension. *Circ Res* 124:1025–1044. <https://doi.org/10.1161/CIRCRESAHA.118.313260>
73. Iliff JJ, Wang M, Liao Y, Plogg BA, Peng W, Gundersen GA et al (2012) A paravascular pathway facilitates CSF flow through the brain parenchyma and the clearance of interstitial solutes, including amyloid  $\beta$ . *Sci Transl Med* 4:147ra111. <https://doi.org/10.1126/scitranslmed.3003748>
74. Iliff JJ, Wang M, Zeppenfeld DM, Venkataraman A, Plog BA, Liao Y et al (2013) Cerebral arterial pulsation drives paravascular CSF-interstitial fluid exchange in the murine brain. *J Neurosci* 33:18190–18199. <https://doi.org/10.1523/JNEUROSCI.1592-13.2013>
75. Ito S, Ueno T, Ohtsuki S, Terasaki T (2010) Lack of brain-to-blood efflux transport activity of low-density lipoprotein receptor-related protein-1 (LRP-1) for amyloid-beta peptide(1–40) in mouse: involvement of an LRP-1-independent pathway. *J Neurochem* 113:1356–1363. <https://doi.org/10.1111/j.1471-4159.2010.06708.x>
76. Iturria-Medina Y, Sotero RC, Toussaint PJ, Mateos-Pérez JM, Evans AC, Alzheimer's Disease Neuroimaging Initiative (2016) Early role of vascular dysregulation on late-onset Alzheimer's disease based on multifactorial data-driven analysis. *Nat Commun* 7:11934. <https://doi.org/10.1038/ncomms11934>
77. Ji F, Pasternak O, Ng KK, Chong JSX, Liu S, Zhang L et al (2019) White matter microstructural abnormalities and default network degeneration are associated with early memory deficit in Alzheimer's disease continuum. *Sci Rep* 9:4749. <https://doi.org/10.1038/s41598-019-41363-2>
78. Johnston JA, Liu WW, Todd SA, Coulson DT, Murphy S, Irvine GB et al (2005) Expression and activity of beta-site amyloid precursor protein cleaving enzyme in Alzheimer's disease. *Biochem Soc Trans* 33:1096–1100. <https://doi.org/10.1042/bst20051096>
79. Kanekiyo T, Cirrito JR, Liu C-C, Shinohara M, Li J, Schuler DR et al (2013) Neuronal clearance of amyloid- $\beta$  by endocytic receptor LRP1. *J Neurosci* 33:19276–19283
80. Kawamura J, Meyer JS, Terayama Y, Weathers S (1991) Cerebral white matter perfusion in dementia of Alzheimer type. *Alzheimer Dis Assoc Disord* 5:231–239. <https://doi.org/10.1097/00002093-199100540-00002>
81. Kennedy AM, Frackowiak RS, Newman SK, Bloomfield PM, Seaward J, Roques P et al (1995) Deficits in cerebral glucose metabolism demonstrated by positron emission tomography in individuals at risk of familial Alzheimer's disease. *Neurosci Lett* 186:17–20. [https://doi.org/10.1016/0304-3940\(95\)11270-7](https://doi.org/10.1016/0304-3940(95)11270-7)
82. Khennouf L, Gesslein B, Brazhe A, Oceau JC, Kutuzov N, Khakh BS et al (2018) Active role of capillary pericytes during stimulation-induced activity and spreading depolarization. *Brain* 141:2032–2046. <https://doi.org/10.1093/brain/awy143>
83. Kimura T, Hashimura T, Miyakawa T (1991) Observations of microvessels in the brain with Alzheimer's disease by the scanning electron microscopy. *Jpn J Psychiatry Neurol* 45:671–676. <https://doi.org/10.1111/j.1440-1819.1991.tb01189.x>
84. Kislser K, Nelson AR, Rege SV, Ramanathan A, Wang Y, Ahuja A et al (2017) Pericyte degeneration leads to neurovascular uncoupling and limits oxygen supply to brain. *Nat Neurosci* 20:406–416. <https://doi.org/10.1038/nn.4489>
85. Kleinberger G, Brendel M, Mracsko E, Wefers B, Groeneweg L, Xiang X et al (2017) The FTD-like syndrome causing TREM2 T66M mutation impairs microglia function, brain perfusion, and glucose metabolism. *EMBO J* 36:1837–1853. <https://doi.org/10.15252/embj.201796516>

86. Koronyo-Hamaoui M, Koronyo Y, Ljubimov AV, Miller CA, Ko MK, Black KL et al (2011) Identification of amyloid plaques in retinas from Alzheimer's patients and noninvasive in vivo optical imaging of retinal plaques in a mouse model. *Neuroimage* 54(Suppl 1):S204–217. <https://doi.org/10.1016/j.neuroimage.2010.06.020>
87. Krogh A (1920) Nobel Lecture. [www.nobelprize.org/prizes/medicine/1920/krogh/lecture/](http://www.nobelprize.org/prizes/medicine/1920/krogh/lecture/).
88. Kwon S, Moreno-Gonzalez I, Taylor-Presse K, Edwards Iii G, Gamez N, Calderon O et al (2019) Impaired peripheral lymphatic function and cerebrospinal fluid outflow in a mouse model of Alzheimer's disease. *J Alzheimers Dis* 69:585–593. <https://doi.org/10.3233/jad-190013>
89. Lambert MP, Barlow AK, Chromy BA, Edwards C, Freed R, Liosatos M et al (1998) Diffusible, nonfibrillar ligands derived from Abeta1-42 are potent central nervous system neurotoxins. *Proc Natl Acad Sci, USA* 95:6448–6453. <https://doi.org/10.1073/pnas.95.11.6448>
90. Law CSW, Yeong KY (2020) Repurposing antihypertensive drugs for the management of Alzheimer's disease. *Curr Med Chem* 27:1–15. <https://doi.org/10.2174/092986732766620031214223>
91. Lee M-S, Tsai L-H (2003) Cdk5: one of the links between senile plaques and neurofibrillary tangles? *J Alzheimers Dis* 5:127–137. <https://doi.org/10.3233/jad-2003-5207>
92. Lei M, Xu H, Li Z, Wang Z, O'Malley TT, Zhang D et al (2016) Soluble Aβ oligomers impair hippocampal LTP by disrupting glutamatergic/GABAergic balance. *Neurobiol Dis* 85:111–121. <https://doi.org/10.1016/j.nbd.2015.10.019>
93. Li R-R, He Y-S, Liu M, Nie Z-Y, Huang L-H, Lu Z et al (2019) Analysis of correlation between cerebral perfusion and KIM score of white matter lesions in patients with Alzheimer's disease. *Neuropsychiatr Dis Treat* 15:2705–2714. <https://doi.org/10.2147/ndt.s207069>
94. Li S, Hong S, Shepardson NE, Walsh DM, Shankar GM, Selkoe D (2009) Soluble oligomers of amyloid beta protein facilitate hippocampal long-term depression by disrupting neuronal glutamate uptake. *Neuron* 62:788–801. <https://doi.org/10.1016/j.neuron.2009.05.012>
95. Yanjun Li, Yongming Li, Li X, Zhang S, Zhao J, Zhu X et al (2017) Head injury as a risk factor for dementia and Alzheimer's disease: a systematic review and meta-analysis of 32 observational studies. *PLoS ONE* 12:e0169650. <https://doi.org/10.1371/journal.pone.0169650>
96. Liu C-C, Hu J, Zhao N, Wang J, Wang N, Cirrito JR et al (2017) Astrocytic LRP1 mediates brain Aβ clearance and impacts amyloid deposition. *J Neurosci* 37:4023–4031. <https://doi.org/10.1523/jneurosci.3442-16.2017>
97. Liu Q, Radwanski R, Babadjouni R, Patel A, Hodis DM, Baumbacher P et al (2019) Experimental chronic cerebral hypoperfusion results in decreased pericyte coverage and increased blood-brain barrier permeability in the corpus callosum. *J Cereb Blood Flow Metab* 39:240–250. <https://doi.org/10.1177/0271678x17743670>
98. Liu S-L, Wang C, Jiang T, Tan L, Xing A, Yu J-T (2016) The role of Cdk5 in Alzheimer's disease. *Mol Neurobiol* 53:4328–4342. <https://doi.org/10.1007/s12035-015-9369-x>
99. Long JM, Holtzman DM (2019) Alzheimer disease: an update on pathobiology and treatment strategies. *Cell* 179:312–339. <https://doi.org/10.1016/j.cell.2019.09.001>
100. Louveau A, Smirnov I, Keyes TJ, Eccles JD, Rouhani SJ, Peske JD et al (2015) Structural and functional features of central nervous system lymphatic vessels. *Nature* 523:337–341. <https://doi.org/10.1038/nature14432>
101. Love S, Miners JS (2017) Small vessel disease, neurovascular regulation and cognitive impairment: post-mortem studies reveal a complex relationship, still poorly understood. *Clin Sci* 131:1579–1589. <https://doi.org/10.1042/cs20170148>
102. Lovell MA, Abner E, Kryskio R, Xu L, Fister SX, Lynn BC (2015) Calcium channel blockers, progression to dementia, and effects on amyloid beta peptide production. *Oxid Med Cell Longev* 2015:787805. <https://doi.org/10.1155/2015/787805>
103. Ma Q, Ineichen BV, Detmar M, Proulx ST (2017) Outflow of cerebrospinal fluid is predominantly through lymphatic vessels and is reduced in aged mice. *Nat Commun* 8:1434. <https://doi.org/10.1038/s41467-017-01484-6>
104. Madry C, Kyrargyri V, Arancibia-Cárcamo IL, Jolivet R, Kohsaka S, Bryan RM et al (2018) Microglial ramification, surveillance, and interleukin-1β release are regulated by the two-pore domain K<sup>+</sup> channel THIK-1. *Neuron* 97:299–312. <https://doi.org/10.1016/j.neuron.2017.12.002>
105. Marshall RS, Lazar RM, Pile-Spellman J, Young WL, Duong DH, Joshi S et al (2001) Recovery of brain function during induced cerebral hypoperfusion. *Brain* 124:1208–1217. <https://doi.org/10.1093/brain/124.6.1208>
106. Masuda T, Croom D, Hida H, Kirov SA (2011) Capillary blood flow around microglial somata determines dynamics of microglial processes in ischemic conditions. *Glia* 59:1744–1753. <https://doi.org/10.1002/glia.21220>
107. Mattsson N, Tosun D, Insel PS, Simonson A, Jack CR, Beckett LA et al (2014) Association of brain amyloid-β with cerebral perfusion and structure in Alzheimer's disease and mild cognitive impairment. *Brain* 137:1550–1561. <https://doi.org/10.1093/brain/awu043>
108. Mawuenyega KG, Sigurdson W, Ovod V, Munsell L, Kasten T, Morris JC et al (2010) Decreased clearance of CNS beta-amyloid in Alzheimer's disease. *Science* 330:1774. <https://doi.org/10.1126/science.1197623>
109. Mehta D, Jackson R, Paul G, Shi J, Sabbagh M (2017) Why do trials for Alzheimer's disease drugs keep failing? A discontinued drug perspective for 2010–2015. *Expert Opin Investig Drugs* 26:735–739. <https://doi.org/10.1080/13543784.2017.1323868>
110. Mehta JL, Nichols WW, Mehta P (1988) Neutrophils as potential participants in acute myocardial ischemia: relevance to reperfusion. *J Am Coll Cardiol* 11:1309–1316. [https://doi.org/10.1016/0735-1097\(88\)90297-5](https://doi.org/10.1016/0735-1097(88)90297-5)
111. Michels L, Warnock G, Buck A, Macauda G, Leh SE, Kaelin AM et al (2016) Arterial spin labeling imaging reveals widespread and Aβ-independent reductions in cerebral blood flow in elderly apolipoprotein epsilon-4 carriers. *J Cereb Blood Flow Metab* 36:581–595. <https://doi.org/10.1177/0271678x15605847>
112. Mielke R, Herholz K, Grond M, Kessler J, Heiss WD (1994) Clinical deterioration in probable Alzheimer's disease correlates with progressive metabolic impairment of association areas. *Dementia* 5:36–41. <https://doi.org/10.1159/000106692>
113. Minami M, Kimura M, Iwamoto N, Arai H (1995) Endothelin-1-like immunoreactivity in cerebral cortex of Alzheimer-type dementia. *Prog Neuropsychopharmacol Biol Psychiatry* 19:509–513. [https://doi.org/10.1016/0278-5846\(95\)00031-p](https://doi.org/10.1016/0278-5846(95)00031-p)
114. Miners JS, Palmer JC, Love S (2016) Pathophysiology of hypoperfusion of the precuneus in early Alzheimer's disease. *Brain Pathol* 26:533–541. <https://doi.org/10.1111/bpa.12331>
115. Minoshima S, Giordani B, Berent S, Frey KA, Foster NL, Kuhl DE (1997) Metabolic reduction in the posterior cingulate cortex in very early Alzheimer's disease. *Ann Neurol* 42:85–94. <https://doi.org/10.1002/ana.410420114>
116. Mitew S, Kirkcaldie MTK, Halliday GM, Shepherd CE, Vickers JC, Dickson TC (2010) Focal demyelination in Alzheimer's disease and transgenic mouse models. *Acta Neuropathol* 119:567–577. <https://doi.org/10.1007/s00401-010-0657-2>



117. Mok VCT, Lam BYK, Wong A, Ko H, Markus HS, Wong LKS (2017) Early-onset and delayed-onset poststroke dementia—revisiting the mechanisms. *Nat Rev Neurol* 13:148–159. <https://doi.org/10.1038/nrneuro.2017.16>
118. Montagne A, Barnes SR, Sweeney MD, Halliday MR, Sagare AP, Zhao Z et al (2015) Blood-brain barrier breakdown in the aging human hippocampus. *Neuron* 85:296–302. <https://doi.org/10.1016/j.neuron.2014.12.032>
119. Montagne A, Nation DA, Sagare AP, Barisano G, Sweeney MD, Chakhoyan A et al (2020) APOE4 leads to blood–brain barrier dysfunction predicting cognitive decline. *Nature* 581:71–76. <https://doi.org/10.1038/s41586-020-2247-3>
120. Morizawa YM, Hirayama Y, Ohno N, Shibata S, Shigetomi E, Sui Y et al (2017) Reactive astrocytes function as phagocytes after brain ischemia via ABCA1-mediated pathway. *Nat Commun* 8:28. <https://doi.org/10.1038/s41467-017-00037-1>
121. Morrison HW, Filosa JA (2013) A quantitative spatiotemporal analysis of microglia morphology during ischemic stroke and reperfusion. *J Neuroinflammation* 10:4. <https://doi.org/10.1186/1742-2094-10-4>
122. Mottet D, Dumont V, Deccache Y, Demazy C, Ninane N, Raes M et al (2003) Regulation of hypoxia-inducible factor-1alpha protein level during hypoxic conditions by the phosphatidylinositol 3-kinase/Akt/glycogen synthase kinase 3beta pathway in HepG2 cells. *J Biol Chem* 278:31277–31285. <https://doi.org/10.1074/jbc.M300763200>
123. Mucke L, Masliah E, Yu GQ, Mallory M, Rockenstein EM, Tatsuno G et al (2000) High-level neuronal expression of abeta 1–42 in wild-type human amyloid protein precursor transgenic mice: synaptotoxicity without plaque formation. *J Neurosci* 20:4050–4058. <https://doi.org/10.1523/jneurosci.20-11-04050.2000>
124. Muller M, van der Graaf Y, Visseren FL, Mali WP, Geerlings MI, SMART Study Group (2012) Hypertension and longitudinal changes in cerebral blood flow: the SMART-MR study. *Ann Neurol* 71:825–833. <https://doi.org/10.1002/ana.23554>
125. Murray KN, Girard S, Holmes WM, Parkes LM, Williams SR, Parry-Jones AR et al (2014) Systemic inflammation impairs tissue reperfusion through endothelin-dependent mechanisms in cerebral ischemia. *Stroke* 45:3412–3419. <https://doi.org/10.1161/strokeaha.114.006613>
126. Nation DA, Sweeney MD, Montagne A, Sagare AP, D’Orazio LM, Pachicano M et al (2019) Blood-brain barrier breakdown is an early biomarker of human cognitive dysfunction. *Nat Med* 25:270–276. <https://doi.org/10.1038/s41591-018-0297-y>
127. Nelson AR, Sagare MA, Wang Y, Kisler K, Zhao Z, Zlokovic BV (2020) Channelrhodopsin excitation contracts brain pericytes and reduces blood flow in the aging mouse brain in vivo. *Front Aging Neurosci* 12:108. <https://doi.org/10.3389/fnagi.2020.00108>
128. Nielsen RB, Egefjord L, Angleys H, Mouridsen K, Gejl M, Møller A et al (2017) Capillary dysfunction is associated with symptom severity and neurodegeneration in Alzheimer’s disease. *Alzheimers Dement* 13:1143–1153. <https://doi.org/10.1016/j.jalz.2017.02.007>
129. Niwa K, Kazama K, Younkin SG, Carlson GA, Iadecola C (2002) Alterations in cerebral blood flow and glucose utilization in mice overexpressing the amyloid precursor protein. *Neurobiol Dis* 9:61–68. <https://doi.org/10.1006/mbdi.2001.0460>
130. Niwa K, Porter VA, Kazama K, Cornfield D, Carlson GA, Iadecola C (2001) A beta-peptides enhance vasoconstriction in cerebral circulation. *Am J Physiol Heart Circ Physiol* 281:H2417–2424. <https://doi.org/10.1152/ajpheart.2001.281.6.H2417>
131. Niwa K, Younkin L, Ebeling C, Turner SK, Westaway D, Younkin S et al (2000) Abeta 1–40-related reduction in functional hyperemia in mouse neocortex during somatosensory activation. *Proc Natl Acad Sci USA* 97:9735–9740. <https://doi.org/10.1073/pnas.97.17.9735>
132. Norman KE, Cotter MJ, Stewart JB, Abbitt KB, Ali M, Wagner BE et al (2003) Combined anticoagulant and antiselectin treatments prevent lethal intravascular coagulation. *Blood* 101:921–928. <https://doi.org/10.1182/blood-2001-12-0190>
133. Nortley R, Korte N, Izquierdo P, Hirunpattarasilp C, Mishra A, Jaunmuktane Z et al (2019) Amyloid  $\beta$  oligomers constrict human capillaries in Alzheimer’s disease via signaling to pericytes. *Science*. <https://doi.org/10.1126/science.aav9518>
134. Ortner M, Hauser C, Schmaderer C, Muggenthaler C, Hapfelmeier A, Sorg C et al (2019) Decreased vascular pulsatility in Alzheimer’s disease dementia measured by transcranial color-coded duplex sonography. *Neuropsychiatr Dis Treat* 15:3487–3499. <https://doi.org/10.2147/ndt.s225754>
135. Palmer JC, Barker R, Kehoe PG, Love S (2012) Endothelin-1 is elevated in Alzheimer’s disease and upregulated by amyloid- $\beta$ . *J Alzheimers Dis* 29:853–861. <https://doi.org/10.3233/JAD-2012-111760>
136. Panza F, Lozupone M, Seripa D, Imbimbo BP (2019) Amyloid- $\beta$  immunotherapy for alzheimer disease: is it now a long shot? *Ann Neurol* 85:303–315. <https://doi.org/10.1002/ana.25410>
137. Pappolla M, Sambamurti K, Vidal R, Pacheco-Quinto J, Poeggeler B, Matsubara E (2014) Evidence for lymphatic A $\beta$  clearance in Alzheimer’s transgenic mice. *Neurobiol Dis* 71:215–219. <https://doi.org/10.1016/j.nbd.2014.07.012>
138. Parhizkar S, Arzberger T, Brendel M, Kleinberger G, Deussing M, Focke C et al (2019) Loss of TREM2 function increases amyloid seeding but reduces plaque-associated ApoE. *Nat Neurosci* 22:191–204. <https://doi.org/10.1038/s41593-018-0296-9>
139. Paris D, Quadros A, Humphrey J, Patel N, Crescentini R, Crawford F et al (2004) Nilvadipine antagonizes both Abeta vasoactivity in isolated arteries, and the reduced cerebral blood flow in APPsw transgenic mice. *Brain Res* 999:53–61. <https://doi.org/10.1016/j.brainres.2003.11.061>
140. Park J-H, Hong J-H, Lee S-W, Ji HD, Jung J-A, Yoon K-W et al (2019) The effect of chronic cerebral hypoperfusion on the pathology of Alzheimer’s disease: a positron emission tomography study in rats. *Sci Rep* 9:14102. <https://doi.org/10.1038/s41598-019-50681-4>
141. Park L, Uekawa K, Garcia-Bonilla L, Koizumi K, Murphy M, Pistik R et al (2017) Brain perivascular macrophages initiate the neurovascular dysfunction of Alzheimer A $\beta$  peptides. *Circ Res* 121:258–269. <https://doi.org/10.1161/CIRCRESAHA.117.311054>
142. Peng W, Achariyar TM, Li B, Liao Y, Mestre H, Hitomi E et al (2016) Suppression of glymphatic fluid transport in a mouse model of Alzheimer’s disease. *Neurobiol Dis* 93:215–225. <https://doi.org/10.1016/j.nbd.2016.05.015>
143. Peppiatt CM, Howarth C, Mobbs P, Attwell D (2006) Bidirectional control of CNS capillary diameter by pericytes. *Nature* 443:700–704. <https://doi.org/10.1038/nature05193>
144. Prohovnik I, Mayeux R, Sackeim HA, Smith G, Stern Y, Alderson PO (1988) Cerebral perfusion as a diagnostic marker of early Alzheimer’s disease. *Neurology* 38:931–937. <https://doi.org/10.1212/wnl.38.6.931>
145. Qiu L, Ng G, Tan EK, Liao P, Kandiah N, Zeng L (2016) Chronic cerebral hypoperfusion enhances Tau hyperphosphorylation and reduces autophagy in Alzheimer’s disease mice. *Sci Rep* 6:23964. <https://doi.org/10.1038/srep23964>
146. Querques G, Borrelli E, Sacconi R, De Vitis L, Leocani L, Santangelo R et al (2019) Functional and morphological changes of the retinal vessels in Alzheimer’s disease and mild cognitive impairment. *Sci Rep* 9:63. <https://doi.org/10.1038/s41598-018-37271-6>



147. Raz L, Bhaskar K, Weaver J, Marini S, Zhang Q, Thompson JF et al (2019) Hypoxia promotes tau hyperphosphorylation with associated neuropathology in vascular dysfunction. *Neurobiol Dis* 126:124–136. <https://doi.org/10.1016/j.nbd.2018.07.009>
148. Reiman EM, Caselli RJ, Yun LS, Chen K, Bandy D, Minoshima S et al (1996) Preclinical evidence of Alzheimer's disease in persons homozygous for the epsilon 4 allele for apolipoprotein E. *N Engl J Med* 334:752–758. <https://doi.org/10.1056/NEJM199603213341202>
149. Roh M-S, Eom T-Y, Zmijewska AA, De Sarno P, Roth KA, Jope RS (2005) Hypoxia activates glycogen synthase kinase-3 in mouse brain in vivo: protection by mood stabilizers and imipramine. *Biol Psychiatry* 57:278–286. <https://doi.org/10.1016/j.biopsych.2004.10.039>
150. Rudinskiy N, Grishchuk Y, Vaslin A, Puyal J, Delacourte A, Hirling H et al (2009) Calpain hydrolysis of alpha- and beta2-adaptins decreases clathrin-dependent endocytosis and may promote neurodegeneration. *J Biol Chem* 284:12447–12458. <https://doi.org/10.1074/jbc.M804740200>
151. Ruitenbergh A, den Heijer T, Bakker SLM, van Swieten JC, Koudstaal PJ, Hofman A et al (2005) Cerebral hypoperfusion and clinical onset of dementia: the Rotterdam Study. *Ann Neurol* 57:789–794. <https://doi.org/10.1002/ana.20493>
152. Rungta RL, Chaigneau E, Osmani B-F, Charpak S (2018) Vascular compartmentalization of functional hyperemia from the synapse to the pia. *Neuron* 99:362–375. <https://doi.org/10.1016/j.neuron.2018.06.012>
153. Sagare AP, Bell RD, Zhao Z, Ma Q, Winkler EA, Ramanathan A et al (2013) Pericyte loss influences Alzheimer-like neurodegeneration in mice. *Nat Commun* 4:2932. <https://doi.org/10.1038/ncomms3932>
154. Sahathevan R, Linden T, Villemagne VL, Churilov L, Ly JV, Rowe C et al (2016) Positron emission tomographic imaging in stroke: cross-sectional and follow-up assessment of amyloid in ischemic stroke. *Stroke* 47:113–119. <https://doi.org/10.1161/strokeaha.115.010528>
155. Sandsmark DK, Bashir A, Wellington CL, Diaz-Arrastia R (2019) Cerebral microvascular injury: a potentially treatable endophenotype of traumatic brain injury-induced neurodegeneration. *Neuron* 103:367–379. <https://doi.org/10.1016/j.neuron.2019.06.002>
156. Schley D, Carare-Nnadi R, Please CP, Perry VH, Weller RO (2006) Mechanisms to explain the reverse perivascular transport of solutes out of the brain. *J Theor Biol* 238:962–974. <https://doi.org/10.1016/j.jtbi.2005.07.005>
157. Selkoe DJ (2002) Alzheimer's disease is a synaptic failure. *Science* 298:789–791. <https://doi.org/10.1126/science.1074069>
158. Seo J, Kritskiy O, Watson LA, Barker SJ, Dey D, Raja WK et al (2017) Inhibition of p25/Cdk5 attenuates tauopathy in mouse and iPSC models of frontotemporal dementia. *J Neurosci* 37:9917–9924. <https://doi.org/10.1523/JNEUROSCI.0621-17.2017>
159. Shager B, Brown CE (2020) Susceptibility to capillary plugging can predict brain region specific vessel loss with aging. *J Cereb Blood Flow Metab*. <https://doi.org/10.1177/0271678x19895245>. (Epub ahead of print)
160. Shibata M, Yamada S, Kumar SR, Calero M, Bading J, Frangione B et al (2000) Clearance of Alzheimer's amyloid-ss(1–40) peptide from brain by LDL receptor-related protein-1 at the blood-brain barrier. *J Clin Invest* 106:1489–1499. <https://doi.org/10.1172/jci10498>
161. Shinohara M, Tachibana M, Kanekiyo T, Bu G (2017) Role of LRP1 in the pathogenesis of Alzheimer's disease: evidence from clinical and preclinical studies. *J Lipid Res* 58:1267–1281. <https://doi.org/10.1194/jlr.R075796>
162. Small GW, Ercoli LM, Silverman DH, Huang SC, Komo S, Bookheimer SY et al (2000) Cerebral metabolic and cognitive decline in persons at genetic risk for Alzheimer's disease. *Proc Natl Acad Sci USA* 97:6037–6042. <https://doi.org/10.1073/pnas.090106797>
163. Small GW, Mazziotta JC, Collins MT, Baxter LR, Phelps ME, Mandelkern MA et al (1995) Apolipoprotein E type 4 allele and cerebral glucose metabolism in relatives at risk for familial Alzheimer disease. *JAMA* 273:942–947. <https://doi.org/10.1001/jama.1995.03520360056039>
164. Smith AJ, Verkman AS (2018) The “glymphatic” mechanism for solute clearance in Alzheimer's disease: game changer or unproven speculation? *FASEB J* 32:543–551. <https://doi.org/10.1096/fj.201700999>
165. Smith GS, de Leon MJ, George AE, Kluger A, Volkow ND, McRae T et al (1992) Topography of cross-sectional and longitudinal glucose metabolic deficits in Alzheimer's disease. Pathophysiologic implications. *Arch Neurol* 49:1142–1150. <https://doi.org/10.1001/archneur.1992.00530350056020>
166. Špiranec K, Chen W, Werner F, Nikolaev VO, Naruke T, Koch F et al (2018) Endothelial C-type natriuretic peptide acts on pericytes to regulate microcirculatory flow and blood pressure. *Circulation* 138:494–508. <https://doi.org/10.1161/circulationaha.117.033383>
167. Storck SE, Meister S, Nahrath J, Meißner JN, Schubert N, Spiezio A et al (2016) Endothelial LRP1 transports amyloid-β(1–42) across the blood-brain barrier. *J Clin Invest* 126:123–136. <https://doi.org/10.1172/jci81108>
168. Sun X, He G, Qing H, Zhou W, Dobie F, Cai F et al (2006) Hypoxia facilitates Alzheimer's disease pathogenesis by up-regulating BACE1 gene expression. *Proc Natl Acad Sci USA* 103:18727–18732. <https://doi.org/10.1073/pnas.0606298103>
169. Suo Z, Humphrey J, Kundtz A, Sethi F, Placzek A, Crawford F et al (1998) Soluble Alzheimer's beta-amyloid constricts the cerebral vasculature in vivo. *Neurosci Lett* 257:77–80. [https://doi.org/10.1016/s0304-3940\(98\)00814-3](https://doi.org/10.1016/s0304-3940(98)00814-3)
170. Takahashi R, Ishii K, Shimada K, Ohkawa S, Nishimura Y (2010) Hypoperfusion of the motor cortex associated with parkinsonism in dementia with Lewy bodies. *J Neurol Sci* 288:88–91. <https://doi.org/10.1016/j.jns.2009.09.033>
171. Tesco G, Koh YH, Kang EL, Cameron AN, Das S, Sena-Estevés M et al (2007) Depletion of GGA3 stabilizes BACE and enhances beta-secretase activity. *Neuron* 54:721–737. <https://doi.org/10.1016/j.neuron.2007.05.012>
172. Thambisetty M, Beason-Held L, An Y, Kraut MA, Resnick SM (2010) APOE epsilon4 genotype and longitudinal changes in cerebral blood flow in normal aging. *Arch Neurol* 67:93–98. <https://doi.org/10.1001/archneur.2009.913>
173. Thiebault AM, Hedou E, Marciniak SJ, Vivien D, Roussel BD (2019) Proteostasis during cerebral ischemia. *Front Neurosci* 13:637. <https://doi.org/10.3389/fnins.2019.00637>
174. Tong X-K, Lecrux C, Rosa-Neto P, Hamel E (2012) Age-dependent rescue by simvastatin of Alzheimer's disease cerebrovascular and memory deficits. *J Neurosci* 32:4705–4715. <https://doi.org/10.1523/JNEUROSCI.0169-12.2012>
175. van Giersbergen PLM, Dingemans J (2007) Effect of gender on the tolerability, safety and pharmacokinetics of clazosentan following long-term infusion. *Clin Drug Investig* 27:797–802. <https://doi.org/10.2165/00044011-200727110-00006>
176. Verret L, Mann EO, Hang GB, Barth AMI, Cobos I, Ho K et al (2012) Inhibitory interneuron deficit links altered network activity and cognitive dysfunction in Alzheimer model. *Cell* 149:708–721. <https://doi.org/10.1016/j.cell.2012.02.046>
177. Wang X, Xing A, Xu C, Cai Q, Liu H, Li L (2010) Cerebrovascular hypoperfusion induces spatial memory impairment, synaptic changes, and amyloid-β oligomerization in rats. *J Alzheimers Dis* 21:813–822. <https://doi.org/10.3233/JAD-2010-100216>

178. Wang Y, Nelson LD, LaRoche AA, Pfaller AY, Nencka AS, Koch KM et al (2016) Cerebral blood flow alterations in acute sport-related concussion. *J Neurotrauma* 33:1227–1236. <https://doi.org/10.1089/neu.2015.4072>
179. Weller RO, Subash M, Preston SD, Mazanti I, Carare RO (2008) Perivascular drainage of amyloid-beta peptides from the brain and its failure in cerebral amyloid angiopathy and Alzheimer's disease. *Brain Pathol* 18:253–266. <https://doi.org/10.1111/j.1750-3639.2008.00133.x>
180. Wierenga CE, Hays CC, Zlatar ZZ (2014) Cerebral blood flow measured by arterial spin labeling MRI as a preclinical marker of Alzheimer's disease. *J Alzheimers Dis* 42(Suppl 4):S411–419. <https://doi.org/10.3233/JAD-141467>
181. Wilhelmus MM, Otte-Höller I, van Triel JJ, Veerhuis R, Maat-Schieman ML, Bu G et al (2007) Lipoprotein receptor-related protein-1 mediates amyloid-beta-mediated cell death of cerebrovascular cells. *Am J Pathol* 171:1989–1999. <https://doi.org/10.2353/ajpath.2007.070050>
182. Wingo AP, Fan W, Duong DM, Gerasimov ES, Dammer EB, Liu Y et al (2020) Shared proteomic effects of cerebral atherosclerosis and Alzheimer's disease on the human brain. *Nat Neurosci* 23:696–700. <https://doi.org/10.1038/s41593-020-0635-5>
183. Wu C-L, Wen S-H (2016) A 10-year follow-up study of the association between calcium channel blocker use and the risk of dementia in elderly hypertensive patients. *Medicine* 95:e4593. <https://doi.org/10.1097/md.0000000000004593>
184. Xiong M, Zhang T, Zhang L-M, Lu S-D, Huang Y-L, Sun F-Y (2008) Caspase inhibition attenuates accumulation of beta-amyloid by reducing beta-secretase production and activity in rat brains after stroke. *Neurobiol Dis* 32:433–441. <https://doi.org/10.1016/j.nbd.2008.08.007>
185. Yamada M, Hayashi H, Suzuki K, Sato S, Inoue D, Iwatani Y et al (2019) Furin-mediated cleavage of LRP1 and increase in ICD of LRP1 after cerebral ischemia and after exposure of cultured neurons to NMDA. *Sci Rep* 9:11782. <https://doi.org/10.1038/s41598-019-48279-x>
186. Yang J, Lunde LK, Nuntagij P, Oguchi T, Camassa LMA, Nilsson LNG et al (2011) Loss of astrocyte polarization in the tg-ArcSwe mouse model of Alzheimer's disease. *J Alzheimers Dis* 27:711–722. <https://doi.org/10.3233/jad-2011-110725>
187. Yemisci M, Gursoy-Ozdemir Y, Vural A, Can A, Topalkara K, Dalkara T (2009) Pericyte contraction induced by oxidative-nitrative stress impairs capillary reflow despite successful opening of an occluded cerebral artery. *Nat Med* 15:1031–1037. <https://doi.org/10.1038/nm.2022>
188. Yew B, Nation DA, Alzheimer's Disease Neuroimaging Initiative (2017) Cerebrovascular resistance: effects on cognitive decline, cortical atrophy, and progression to dementia. *Brain* 140:1987–2001. <https://doi.org/10.1093/brain/awx112>
189. Yun C-H, Lee H-Y, Lee SK, Kim H, Seo HS, Bang SA et al (2017) Amyloid burden in obstructive sleep apnea. *J Alzheimers Dis* 59:21–29. <https://doi.org/10.3233/JAD-161047>
190. Zeisel A, Hochgerner H, Lönnerberg P, Johnsson A, Memic F, van der Zwan J et al (2018) Molecular architecture of the mouse nervous system. *Cell* 174:999–1014. <https://doi.org/10.1016/j.cell.2018.06.021>
191. Zhai P, Sciarretta S, Galeotti J, Volpe M, Sadoshima J (2011) Differential roles of GSK-3 $\beta$  during myocardial ischemia and ischemia/reperfusion. *Circ Res* 109:502–511. <https://doi.org/10.1161/CIRCRESAHA.111.249532>
192. Zhang C, Zhu Y, Wang S, Zachory WZ, Jiang MQ, Zhang Y et al (2018) Temporal gene expression profiles after focal cerebral ischemia in mice. *Aging Dis* 9:249–261. <https://doi.org/10.14336/ad.2017.0424>
193. Zhang X, Yin X, Zhang J, Li A, Gong H, Luo Q et al (2019) High resolution mapping of brain vasculature and its impairment in the hippocampus of Alzheimer's disease mice. *Natl Sci Rev* 6:1223–1238. <https://doi.org/10.1093/nsr/nwz124>
194. Zhang Y, Xiong M, Yan R-Q, Sun F-Y (2010) Mutant ubiquitin-mediated beta-secretase stability via activation of caspase-3 is related to beta-amyloid accumulation in ischemic striatum in rats. *J Cereb Blood Flow Metab* 30:566–575. <https://doi.org/10.1038/jcbfm.2009.228>
195. Zhao Y, Wu X, Li X, Jiang L-L, Gui X, Liu Y et al (2018) TREM2 Is a receptor for  $\beta$ -amyloid that mediates microglial function. *Neuron* 97:1023–1031. <https://doi.org/10.1016/j.neuron.2018.01.031>
196. Zhao Z, Sagare AP, Ma Q, Halliday MR, Kong P, Kisler K et al (2015) Central role for PICALM in amyloid- $\beta$  blood-brain barrier transcytosis and clearance. *Nat Neurosci* 18:978–987. <https://doi.org/10.1038/nn.4025>
197. Zhiyou C, Yong Y, Shanquan S, Jun Z, Liangguo H, Ling Y et al (2009) Upregulation of BACE1 and beta-amyloid protein mediated by chronic cerebral hypoperfusion contributes to cognitive impairment and pathogenesis of Alzheimer's disease. *Neurochem Res* 34:1226–1235. <https://doi.org/10.1007/s11064-008-9899-y>
198. Zlokovic BV (2005) Neurovascular mechanisms of Alzheimer's neurodegeneration. *Trends Neurosci* 28:202–208. <https://doi.org/10.1016/j.tins.2005.02.001>
199. Zott B, Simon MM, Hong W, Unger F, Chen-Engerer H-J, Frosch MP, Sakmann B et al (2019) A vicious cycle of  $\beta$  amyloid-dependent neuronal hyperactivation. *Science* 365:559–565. <https://doi.org/10.1126/science.aay0198>

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