



Targeting NK Cells for HIV-1 Treatment and Reservoir Clearance

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Combined antiretroviral therapy (cART) can inhibit the replication of human immunodeficiency virus type 1 (HIV-1) and reduce viral loads in the peripheral blood to undetectable levels. However, the presence of latent HIV-1 reservoirs prevents complete HIV-1 eradication. Several drugs and strategies targeting T cells are now in clinical trials, but their effectiveness in reducing viral reservoirs has been mixed. Interestingly, innate immune natural killer (NK) cells, which are promising targets for cancer therapy, also play an important role in HIV-1 infection. NK cells are a unique innate cell population with features of adaptive immunity that can regulate adaptive and innate immune cell populations; therefore, they can be exploited for HIV-1 immunotherapy and reservoir eradication. In this review, we highlight immunotherapy strategies for HIV infection that utilize the beneficial properties of NK cells.

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INTRODUCTION

Human immunodeficiency virus type 1 (HIV-1) can be controlled by combined antiretroviral therapy (cART) to the point at which viral loads in the peripheral blood are undetectable. However, the main obstacle to total HIV eradication and a functional cure is the presence of latent HIV-1 reservoirs, which exist in the peripheral blood and lymphoid tissues. To functionally cure HIV-1, several strategies have been proposed, including gene editing technologies based on CCR5, broadly neutralizing antibodies (bNAbs), the "shock and kill" strategy for eradication of shallow reservoirs, and the "block and lock" or "permanent silencing" approach for eradication of deep reservoirs (1). Many latency-reversing agents (LRAs) have been researched as part of the "shock and kill" approach, but only a small number of candidates have been evaluated in clinical trials. Although the majority of LRAs can activate viral transcription, they are unable to induce "killing" effectively. While preventative medications and treatments exist to reduce the disease burden of acquired immunodeficiency syndrome (AIDS), there is no effective cure for HIV; therefore, it is critical to develop useful approaches for HIV immunotherapy.

HIV infects humans through immune cells, including macrophages (2, 3), dendritic cells (DCs) (4), and especially CD4⁺ T cells; this can destroy immune function and result in severe infection, malignant tumors, autoimmune illnesses, and other complications (5). In the past years, the innate immune system has been shown to play a significant role in fighting HIV. The innate immune system is the body's initial line of defense against foreign pathogens, and it includes a variety of cells

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and cytokines that fight off pathogens in a nonspecific manner. Among innate immune cells, natural killer (NK) cells play important roles in antiviral and anti-tumor immune responses. Advances in NK cell technology have been spurred by the development of cancer immunotherapy, and these advances have the potential to enhance HIV immunotherapy and to overcome some of the problems with strategies targeting T cells. Numerous data and literature demonstrate the efficacy of NK immunotherapy against tumors (6–9) and liver inflammation (10) have been discussed elsewhere (6–9). In the present paper, we will review recent advances in NK cell technology and the potential of NK cells for use in HIV immunotherapy.

OVERVIEW OF NK CELLS

Natural killer cells are components of the innate immune system (11) that are phenotypically comparable to T cells, particularly CD8⁺ T cells (2). Human NK cells are derived from CD34⁺ lymphoid lineage cells (HPCs) (12, 13) and found in the liver, peritoneum, placenta, and other organs; they account for 5-15% of peripheral blood mononuclear cells (PBMCs). Immature NK cells in secondary lymph nodes are stimulated by cytokines such as IL-15 and IL-12 from DCs or other antigen presenting cells (APCs) (14) and are subsequently transferred to the peripheral blood, where they gradually acquire cytotoxicity during differentiation and development. NK cells are divided into three or five subgroups according to their expression of CD56 and CD16: CD3⁻CD56⁻CD16⁺, CD3⁻CD56^{dim}CD16⁺, and CD3⁻CD56^{bri}CD16^{+/-}, and the total population of NK cells included these three subsets (9, 15), with the CD56^{dim} NK cells being the most numerous in peripheral blood and the CD56^{bright} NK cells are largely enriched in the liver (16). CD56^{bright} liverresident NK cells (lrNKs) can be further defined based on the expression of CD69, CD49a, CCR5 and CXCR6 (17). Moreover, technological advances in mass cytometry revealed peripheral blood NK cell diversity with at least 6000 phenotypic populations in an individual assessed by more than 30 parameters simultaneously, which can be used for immunotherapeutic strategies for infection, reproduction, and transplantation (18).

NK cells are multifunctional natural effector cells that have antiviral and antitumor properties. They participate in killing and lysing target cells by recognizing virus-infected cells or tumor cells through their surface receptors. NK cells have diverse mechanisms of killing, including release of cytotoxic granules and apoptosis mediated by the Fas/FasL pathway or TNF-related apoptosis-inducing ligand (TRAIL) (19, 20). NK cells can also engage in antibody-dependent cell cytotoxicity (ADCC) leading to death of antibody-coated cells (21). Finally, NK cells can secrete cytokines and chemokines that contribute to their antiviral effects, including IFN- γ , GM-CSF, TNF- α , CCL3, CCL4, CCL5, XCL1, and XCL2 (14, 17, 18). In addition to their natural killing function, NK cells have significant immunomodulatory roles; for example, they regulate adaptive immune cells such as T and B cells and innate immune cells such as DCs and macrophages to impact the outcome of infections (22, 23). Multiple features of NK cells indicate that they play dynamic roles in immune-mediated protection and homeostasis (24).

NK CELL ROLES DURING HIV INFECTION

NK cells play significant roles in HIV-1 infection. In the early stages of viral infection, NK cells act more quickly than adaptive immune cells, and cytotoxic CD56^{low} NK cells expand faster than CD8⁺ T cells (25). HIV infection changes the distribution and functions of NK cell subpopulations (26-28) even after they are partially restored by antiretroviral therapy (ART) (29). Another study found that CD56^{low}CD16⁺ NK cells decreased significantly during HIV-1 infection, whereas CD56^{high}CD16⁺ NK cells populations were unchanged (26, 30), and this impacted the rapid and early progression of AIDS (31). In addition, The CD56⁻CD16⁺ NK cell subpopulation reduces spontaneous NK cytotoxicity (26). Furthermore, NK cells in HIV-1-exposed seronegative intravenous drug users (HESN-IDU) produce higher levels of IFN- γ and TNF- α (32, 33) compared to NK cells in healthy controls. Another study demonstrated that NK cells can dramatically inhibit viral entry into CD4⁺ T cells and prevent the spread of HIV-1 by producing β -chemokines following stimulation with IL-2 and IL-15 (34). NK cells are also associated with the development of bNAbs during HIV-1 infection (35). Immunogenetics, viral development, and immune escape studies showed that enhanced NK cell activity supported by cytotoxic cytokines and chemokines production are involved in delaying AIDS progression and controlling of HIV infection (36).

"Shock and kill" therapy strategies attempt to reactivate the latent HIV-1 in lymphocytes and tissues by LRAs, and this is followed by killing of the infected cells by the immune system and ART (37). Some classic LRAs have been reported to influence the function and fate of NK cells. For example, the protein kinase C (PKC) and NF-KB activator prostratin induced non-specific NK cell activation and a strong antiviral response, whereas panobinostat decreased NK cell viability, antiviral activity, and cytotoxicity; these factors should be carefully considered when evaluating strategies to eliminate HIV-1 infection (38). In a clinical trial of people living with HIV (PLWH), following analytical treatment interruption (ATI), those treated with panobinostat had decreased proviral HIV-1 DNA levels and increased viral rebound times, and this was correlated with higher frequencies of immunomodulatory CD56⁺ NK cells, CD56^{low} NK cells, and plasmacytoid dendritic cells (39). The impact of panobinostat on NK cell populations may be due to LRA-mediated alterations in germline-encoded NK receptor ligand expression on HIV-infected CD4⁺ T cells. A study of HIV-1 and Hepatitis C Virus (HCV) coinfected individuals receiving cART showed that pegylated interferon- α (PEG-IFNa) induced NK cell activation, and levels of cellassociated proviral DNA were negatively correlated with NK cell quantities (40). Peg-IFN-a2a-mediated HIV-1 suppression

was also correlated with NK cell cytotoxicity and innate immune activity (41). These findings indicate that the reduction in HIV-1 reservoirs following treatment with PEG-IFN α occurs mainly due to the antiviral properties of NK cells. It is also worth noting that targeting NK cell subsets to restore any residual dysfunction could enhance their antiviral properties and reduce related comorbidities.

NK CELL ACTIVATION

NK Cell Receptors

NK cell surface receptors are divided into two categories according to their structural features: immunoglobulin superfamily receptors (Ig-SF) and C-type lectin superfamily receptors (C1-SF). Ig-SF receptors include i) killer inhibitory receptors (KIRs) and natural cytotoxicity receptors (NCRs), which recognize human leukocyte antigen (HLA)-A, -B, and -C, KIRs are important receptors that regulate the functions of NK cells and are mainly expressed by killer cell subsets. KIRs are also major components of HLA-I binding, which is associated with disease progression (42); and ii) C-type lectin superfamily (C-agglutinin) receptors, chiefly CD94, NKG2, and NKR-P1, which recognize HLA-E (43). According to their functions, NK cell surface receptors can be divided into activating receptors and inhibitory receptors (44). The activating receptors expressed on NK cells are mainly NKG2D, NKG2C, and the NCRs (45, 46); the NCRs include NKP30, NKP44, NKP46 (47). Inhibitory NK cell receptors (iNKRs) include the inhibitory KIRs (iKIRs), LIR1/ILR2, and CD94/NKG2A or CD94/NKG2C heterodimers (Figure 1) (8, 48). These iNKRs deliver inhibitory signals to NK cells by recognizing the classical class I molecular antigenic determinants of human leukocyte antigens in "own" target cells, hence preventing attack of "self" cells (49, 50). In humans, KIR3DL1 and KIR2DL1 are two of the prominent NK cell receptors (51). Activation signals via NK receptors such as NKp30, NKp44, NKp46, NKG2D, and NKp80 mediate the activation of NK cells consistent with the loss of inhibitory signals (52). Recently a genome-wide CRISPR/Cas9 knock-out strategy identied some unconventional ligands of NK-cell receptors and uncovered a new binding of various KIRs to heparan sulfate proteoglycans that may make an effect on NK cell receptor signaling and target-cell recognition (53). Under healthy conditions, NK cells are not activated when self MHC class I molecules on the target cell surface bind to NK cell inhibitory receptors, which is known as NK cell education [100]. This allows NK cells to distinguish between self and non-self cells. Instead, NK cells are activated and may stimulate activator receptors to release signals and instigate NK cell activities (54) when self cells are transformed or infected with pathogens, and this is followed by subsequent target cell lysis (6).

Blocking Inhibitory NK Cell Receptors During HIV-1 Infection

HIV-1 infection has been shown to alter NK cell surface receptors (26, 55, 56). The activation status of NK cells is determined by the dynamic balance of activating and inhibitory signals generated by

the interactions between NK cell surface receptors and their ligands (57). It has been shown that infection with HIV-1 significantly increases the expression of KIRs on both T and NK cells (58, 59), and the activating receptors NKG2C and CD226/PTA1 on peripheral blood NK cells are abnormally expressed in PLWH (55, 60). A functionally impaired NK cell's cytotoxicity is related to the decreased expression of NCRs (56). NK cells with a specific transcriptional signature and function reportedly were protective in vivo for controlling HIV-1 in CD4⁺ T cells, and increased expression of activating receptor NKp46/NKp30 helped to contain HIV-1 reservoir size (61). The dynamic balance of NK cell activation can also be regulated by interactions with other cells such as neutrophils, macrophages, and dendritic cells, as well as other NK subsets, these cellular interactions also regulate cytokine production, initial viral loads, and CD4⁺ T cell-mediated immune responses during viral infections (62-65). This implies that monitoring, selecting, expanding, and adoptively transferring this NK cell population may be an effective strategy for HIV-1 eradication (61).

However, the quantity and functionality of NK cells in HIVinfected individuals is a matter of ongoing debate. As HIV-1 progresses, continuous stimulation of NK cell receptors may lead to continuous activation, causing chronic inflammation and damage to organs and tissues (15, 66). The expression of HLA class I ligands specific to NK cell receptors changes during HIV infection, for example, higher HLA-A levels render HIV less controllable (67). The downregulation of HLA-C expression on HIV infected cells results in impaired recognition by HLA-Crestricted CTL and a subsequent increase in viral replication in vitro (68). NKG2A and inhibitory KIR blockade may also target CD8⁺ T cells and induce a complementary treatment effect. NK cell activity can be inhibited by NKG2A recognizing and binding the ligand HLA-E on target cells (69). Thus, therapeutic blockade of NKG2A: HLA-E interactions may be an effective strategy for HIV eradication.

Motavizumab, an NKG2A blocking antibody, has been studied in various trials to enhance NK and CD8⁺ T cell functionality for cancer immunotherapy (70, 71). Interestingly, it has been shown that blocking the inhibitory receptor NKG2A in mice and patients with chronic hepatitis B virus (HBV) enhances NK cell cytotoxicity and viral clearance (72). Another study demonstrated that an HIV-1 capsid presented by HLA-E can interact with HLA-E specific NK cell inhibitory receptors (iNKRs) NKG2A/CD94, and consequently, NK cells expressing NKG2A can kill the HIVinfected cells (69). However, potential therapeutic approaches that block NK cell inhibitory receptors raise concerns about redundant negative NK cell immunomodulation, unnecessary self-reactivity, and paracellular depletion of activated uninfected T cells (73); these concerns bring challenges for HIV treatment by blocking NK cell inhibitory receptors. Recently research found that HLA-C*03: 04-presented peptides derived from noninfected CD4⁺ T cells can mediate stronger binding of inhibitory KIR2DL3 than that derived from HIV-1-infected cells, which means that HLA-Ipresented peptides alteration induced by HIV-1 infection can reduce engagement of iKIRs and provide a potencial method to activate NK cells by virus-infected cells (74). In summury, blocking

NK cells			Target cells		
	NKG2C		HLA-E		
	KIRSDS1		HLA-C2		
Activating Receptors	KIRSDS2		HLA-A, HLA-C		
	KIRSDS3		?		
	KIRSDS4		HLA-A, HLA-C, HL/	ILA-F	
	KIRSDS5		HLA-C		
	KIR3DS1		HLA-B, HLA-F		
	CD160		HLA-C	HLA-	
CD94/NKG2A			HLA-E	Ligan	
	KIR2DL1		HLA-C		
KIR2DL2/L3			HLA-C1/C2, HLA-B		
Inhibitory Receptors	KIR2DL4		HLA-G		
	KIR2DL5		?		
	KIR3DL1		HLA-A, HLA-B		
	KIR3DL2		HLA-A, HLA-F		
	KIR3DL3		?		
	L1LRB1		HLA		

iNKRs can enhance NK cell activation and could be of particular benefit for elimination of HIV.

ENHANCING NK CELL FUNCTION IN HIV-1 INFECTION

Antibody-Mediated NK Cell Responses

Freshly isolated PBMCs from HIV-infected patients can kill target cells by ADCC without additional antibodies because NK cells from HIV patients are binding with antibodies

against gp120/gp140 and can immediately kill target cells expressing gp120 (75, 76). The ADCC response is an important immune mechanism for host resistance to pathogenic microorganisms and the clearance of diseased cells, and it plays an important role in fighting HIV infection and is thought to be especially important in the small number of longterm slow-progressors (LTSP) (77). In addition to antibodies, NK cells are one of the primary mediators of ADCC because they express CD16, also known as Fc γ RIII. The NK-mediated ADCC response has been used as an indicator of HIV suppression, and Fc γ RIII binding and HIV-specific-ADCC activity can be improved by the administration of monoclonal antibodies that are regulated by single nucleotide polymorphisms (SNPs) (78).

NK cells are the main effector cells to perform ADCC, and the overall effectiveness of ADCC in both healthy and HIV-infected individuals relies on NK cells (29). Non-neutralizing functions mediated by interactions with Fc receptors, particularly those expressed by NK cells, may also be required for the eradication of virus-infected cells (79). With the advances of the monoclonal antibodis technique for anti-HIV-1, bNAbs offer stronger antiviral potential and in clearing virus-infected cells with Fcmediated clearance. In fact, Fc receptor expression levels in humanized mice and rhesus macaques appear to directly impact the ability of bNAbs to inhibit viral replication (80, 81). After passive vaccination, bNAbs have the potential to induce NK-mediated ADCC (82) and hold promise for advancing HIV treatments. Some clinical trials in PLWH have already shown the safety, tolerability, and therapeutic efficacy of bNAb 3BNC117 targeting the CD4 binding site. It has been concluded that 3BNC117-mediated immunotherapy can enhance host humoral immunity to HIV-1 infection (83). In another phase I clinical trial, a virus-like nanoparticles that present clusters of membrane-associated CD4 (CD4-VLPs) bNAbs was highly effective for reducing HIV-1 viremia (84). A separate group found that 3BNC117 has dramatic neutralizing effects in vitro and can inhibit several HIV/SHIV strains, and 3BNC117 induces humoral and cellular immune responses in vivo to control HIV-1 reservoirs (85). Nevertheless, it will be a challenge to treat 3BNC117-resistant HIV-1 strains using this approach, and the in vivo effects of bNAbs on human NK cells remain unknown. Several groups have sought to use NK cells in conjunction with bNAbs to improve targeting of virally infected cells. For example, two bNAbs (3BNC117 and 10-1074) combined with pegylated interferon $\alpha 2b$ (peg-IFN- $\alpha 2b$) were used to evaluate ADCC and NK cell activation in PLWH (NCT03588715). Strategies targeting cellular immunity that combine 3BNC117 with other pharmaceuticals (e.g. bNABs, antiretroviral medications, viral inducers) are predicted to benefit the prevention and treatment of AIDS.

BiKEs and TriKEs

The latent HIV-1 infected cells evade identification and killing by NK cells via various immune escape mechanisms. An immunomodulatory strategy has been proposed that brings killer cell engaging molecules into contact with HIV-1 infected cells to boost antigen specificity. These molecules are bispecific and trispecific killer cell engagers (BiKEs and TriKEs) (86), created by connecting an antibody single chain against NK cell CD16 to one or two fragments of a specific antibody against an antigen expressed on the target cell surface. This strategy alleviates many of the issues with NK cell therapy, including lack of specificity in NK cell targeting, limited in vivo NK cell activation, viability and proliferation, which can be overcome by higher quality TriKEs and IL-15 supplements. CD4-based BiKEs were demonstrated to activate degranulated NK cells and CD16A-expressing T cells, induce cytokine production, and kill cells expressing HIV-1 envelope glycoproteins (Envs). Due to their small molecular size, high affinity in humans, significant efficacy, and widespread binding to HIV-1 strains, CD4-based BiKEs have enormous potential for curing HIV-1 (87). Clinical trials have examined the combined effects of drugs and antibodies, and it is unclear whether these antibodies act through neutralization effects. Notably, monoclonal antibodybased therapy may facilitate the induction of antibodies directed against such humanized monoclonal antibodies (88), and thus the potential of such antibody-induced hypersensitivity reactions needs to be considered.

IMMUNE CHECKPOINT BLOCKADE

Immune checkpoint inhibition can restore the function of exhausted T cells during chronic infection and in the tumor microenvironment; this approach is widely used to treat cancer and could be an appealing adjuvant therapeutic approach for HIV. In addition to iNKRs, NK cells express immune checkpoint receptors previously found on T cells, such as programmed death receptor 1 (PD-1), lymphocyte activation gene-3 (LAG3)/ CD223, cytotoxic T lymphocyte associateprotein-4 (CTLA-4), T cell immunoglobulin-3 (TIM-3), T cell immune receptor with Ig and ITIM domains (TIGIT) and the recently identified CD276 (B7-H3) (89). Checkpoint inhibitors may strengthen NK cell activation and cytolytic effectiveness (90). Initial studies of immune checkpoint inhibitors in HIV mainly focused on CD4⁺ T cells. Although checkpoint inhibition has been shown to significantly improve NK cell activity in cancer patients, the benefits of checkpoint inhibition in PLWH on ART have yet to be determined. NK cells, like T cells, can develop an exhausted phenotype during HIV or simian immunodeficiency virus (SIV) infections (91). NK cell exhaustion can cause various problems including poor proliferative capacity, decreased expression of activating receptors (56, 92, 93), and increased expression of inhibitory receptors (26). Furthermore, exhausted NK cells are unable to degranulate, produce cytokines, and promote ADCC (93). PD-1, an key marker of T cell exhaustion during HIV-1 infection, was upregulated on NK cells from HIV-1 positive individuals following ART. Increased PD-1 expression was associated with limited NK cell proliferation, which may impact NK cell maintenance during HIV-1 infection (94). HIV infection and treatment may alter invariant natural killer T cell (iNKT) induced IFN- γ levels. iNKT dysfunction is uniquely associated with the expression of LAG-3, an immune checkpoint marker similar to PD-1, and persistent LAG-3 expression may result in deficient immune reconstitution in PLWH during ART (95).

Multiple activating and inhibitory interactions between T cells and APCs can regulate the immune response. Immune checkpoints promote suppressive interactions between immune cells to maintain homeostasis. Immune checkpoint inhibition has been consistently demonstrated to enhance not only T cell function, but also NK cell function during HIV infection. One group demonstrated that in both untreated and ART-suppressed PLWH, combined PD-1 and IL-10 blockade could enhance NK cell cytokine secretion, degranulation, and killing capacity as well as restore CD4⁺ T cell function. This implies that using immune

checkpoint inhibition to improve the cooperation between T and NK cells is a potential therapeutic approach for HIV (96). Therefore, immunotherapeutic interventions targeting immune checkpoint could augment NK cell activity and function against HIV and PLWH with cancers.

TLR AGONIST-INDUCED NK CELLS IN HIV-1 INFECTION

Toll-like receptors (TLRs) are pathogen recognition receptors (PRRs) that detect and modulate signaling responses based on molecular motifs that are conserved across pathogenic microorganisms. TLRs are mainly found on APCs and epithelial and endothelial cells (97), and they are localizated in cell surface (TLR1, 2, 4, 5, 6, 10, 11) and endosomal (TLR3, 7, 8, 9, 12, 13) (98). TLR2 heterodimerizes with TLR1 or TLR6 to function. TLR agonists have been studied as LRAs with the potential to reverse latent HIV-1 and enhance antiviral responses through immunomodulation. TLR agonists enhance immune activation, adaptive responses, and innate antiviral responses in vitro and in vivo (99). Because of unique immunomodulatory properties of TLR agonists have, further research into the approach to combine latency reversal and viral clearance strategies is undergoing in vivo (99). Different TLRs are expressed in NK cells, according to reports, TLR1-TLR9 mRNA was expressed in human NK cells, with TLR1 expression being the highest, followed by moderate levels of TLR2, TLR3, TLR5, and TLR6, and low or undetectable expression of TLR9 levels (100, 101), (reviewed in (102). TLR ligands can activate NK cells directly or indirectly, for example, a new synthetic TLR1/2 ligand named XS15 (103) can stimulate NK cells by monocytes and TLR-7/8 agonist Clo97 can activate NK cells by polymorphonuclear neutrophils (104) indirectly; NK cells also were activated and the cytotoxicity of them were enhanced by autologous DC cultured with poly (I:C) (105). In addition, TLR ligands such as TLR1/2 (Pam3CSK4 or SMU-Z1) or TLR2/6 (MALP-2) and TLR3 (poly I:C) can directly activate human NK cells (106-109). Although poly I:C treatment had no significant effect on CD4⁺ T cells and plasma viral control, the data suggested that poly I:C can induce innate immune responses in PLWH, indicating that it could be a promising adjuvant for HIV therapeutic vaccines (NCT02071095) (110). In SIV-infected rhesus monkeys, TLR7 agonist GS-9620 combined with bNAb PGT121 effectively activated NK cells and CD4⁺ T cells and delayed viral rebound after ART interruption, indicating the potential of NK cell stimulation in conjunction with bNAbs for targeting HIV reservoirs (111). GS-9620 has already been studied in clinical trials NCT03060447 and NCT02858401 in PLWH receiving ART. TLR-7 and/or TLR-8 agonists are being tested for their ability to directly activate NK cells. Although purified NK cells do not express TLR-7 or -8, TLR-7/TLR-8 agonists (R-848) enhance NK-cell cytotoxicity in vivo and control NK cell activation indirectly through immune modifiers, and TLR-8 ligands result in indirect NK-cell activation by IL-18 and IL-12p70 and possibly by other cytokines (112). Furthermore, the

TLR9 agonist MGN1703 has been shown in clinical studies to activate NK cells in PLWH (113). In summary, TLR agonists have been shown to target cellular and tissue HIV reservoirs in a variety of *in vivo* studies (114). Combined with these results, it is reasonable to expect that TLR agonists will soon become a safe and multipotent clinical drug designed to reduce the HIV reservoir.

NK CELL STIMULATION BY CYTOKINES

Many interleukins such as IL-2, -12, -15, -18 and -21 have been found to boost NK cytotoxicity and proliferation both in vitro and in vivo (7). In particular, IL-15 is known to promote the proliferation, differentiation, and maturation of NK cells and induces NK cell receptor expression, ADCC, cytotoxicity, and IFN-y production. After exposure to LRAs, NK cells stimulated with IL-15 were able to kill latent HIV-infected cells (115). Remarkably, the IL-15 superagonist ALT-803/N-803 has been identified as a LRA and can induce HIV elimination in a cooperative manner with CD8⁺ T cells (116). Moreover, in ART-treated macaques, ALT-803 could consistently reactivate SIV in cases of CD8⁺ T cell depletion (117). Furthermore, ALT-803 could be used as an immunomodulator to effectively prevent the formation HIV/SIV: it transiently increased NK cell, CD8⁺ T and memory T cell populations as well as decreased viral loads in SIV-positive rhesus macaques (118). ALT-803 has been evaluated in clinical trials for lymphoma (119), solid and hematological tumors (120, 121), and for synergy with other immunotherapies (NCT01885897). ALT-803 is also undergoing clinical trials to assess the possibility of HIV reservoir elimination (NCT02191098). A phase II, randomized, unblinded, controlled trial will be conducted to investigate the safety, tolerability, and immunomodulatory effect of combining N-803 with ART during acute HIV infection (NCT04505501). The efficacy of N-803 with or without bNAbs is being evaluated for curing HIV in PLWH receiving analytic treatment interruption (ATI) (NCT04340596). There will be more good news to report in the future. ALT-803 has shown significant results in the treatment of bladder cancer and initial success in HIV infection, however, only one drug is not enough to eliminate AIDS and more combinations need to be explored.

ADOPTIVE NK TRANSFER THERAPY

Adaptive NK Cells Responses

Classical NK cells are defined as such because they develop antigen receptors irrespective of the recombination activating gene (RAG) (122). *In vivo* studies in mice, nonhuman primates, and humans have indicated that NK cells are capable of quickly responding in an antigen specific manner (123–125). NK cells, like adaptive memory T cells, can acquire memory properties and produce stronger immune responses to previously seen antigens (126–130). Several paradigm-shifting efforts led to the acknowledgement that NK cells have features of adaptive immunity (131, 132). Studies of CD94⁺NKG2C⁺ NK cells in patients infected with cytomegalovirus (CMV) provided the first indications of adaptive-like NK cells (133–135), and memorylike NK cells that respond to CMV have since been identified in mouse models and in humans (136). Observations that NK cells can recall prior immune responses have made NK cell memory a research hotspot. However, numerous questions about NK cell memory remain. It is unclear which NK cell subsets act as "memory" initiators or which ligands induce memory, and it is uncertain if these cells can conduct effector functions immediately upon reactivation by HIV-1 (137). Despite these concerns, it is critical to understand the basic mechanisms underlying NK cell memory in HIV-1 infected patients for improving vaccine-induced cytolytic therapies and eliminating reservoirs (138).

Haploidentical and Allogeneic NK Transfer

NK cells can be obtained from bone marrow or umbilical-cord blood (UCB), as well as from human embryonic stem cells or induced pluripotent stem cells (iPSC). NK cells can be acquired from patients, which is known as the autologous setting, or from healthy donors, which is known as the allogeneic setting. There are limited circulating NK cells in the peripheral blood, and naturally-derived NK cells have low cytotoxicity; therefore, many strategies have been proposed to expand NK cells to obtain large quantities capable of antitumor and antiviral effects (139, 140), which could allow for adoptive NK cell transfer therapy. For example, PM21 particles from plasma membranes of K562mb21-41BBL cells, which express membrane bound IL-21 (mb21) and 4-1BB ligand (41BBL), can promote ex vivo specific NK-cell expansion from PBMCs in healthy donors and stimulate in vivo NK cell expansion in mice, PM21 particles may make NK cell-mediated immunotherapy more widely accessible to HIV individuales (141). Moreover, PM21-NK cells can efficiently kill oncolytic parainfluenza virus 5 (P/V virus) infected cancer cells (142), which support the potential of combining P/V virus with PM21-NK cell adoptive therapy against cancer and HIV. Otherwise, several clinical trials are underway to investigate the effectiveness of NK cell adoptive transfer for treating HIV. Two clinical trials are underway to determine whether haploidentical NK cells combined with the IL-15 agonist ALT-803 or with IL-2 are safe and well-tolerated, and if there is any measurable impact on in vivo reservoirs (NCT03899480, NCT03346499).

In addition to haploidentical NK cells, there are several allogenic sources of NK cells for adoptive transfer HIV-1 therapy. UCB, for example, is a robust source of mature, phenotypically and functionally therapeutic effector NK cells without an exact HLA match (143). UCB NK cells produce the same level of IFN- γ and higher levels of perforin and granzyme B than peripheral blood NK cells in response to several stimuli tested (144). Clinical trials NCT01619761 and NCT02280525 are actively evaluating the therapeutic efficacy of umbilical cord blood transplants for treating patients with hematological malignancies. UCB is also a rich source of hematopoietic progenitor cells (HPC)s, which may differentiate into the effector NK cells required for different phenotypes *in vitro*. The clonal NK

cell line NK-92 mediates intense cytotoxic responses against a wide range of tumor cells but not against non-malignant healthy cells, and it has been extensively studied in clinical trials for cancer immunotherapy (145–147); therefore, NK-92 cells are expected to become a future therapeutic option for HIV-1. Furthermore, independent of HLA haplotype, stem cell-derived NK cells may be employed as standardized 'off-the-shelf' treatments for all donors, allowing for the production of large quantities of homogeneous NK cells that are easier to genetically manipulate than primary NK cells (148). Stem cell-derived NK cells and iPSCs are currently being evaluated clinically in NCT04023071 and NCT04245722 for anti-lymphoma therapy [reviewed in (8)], but they may also have future uses for AIDS therapy.

CAR-NK Cells

Autologous chimeric antigen receptor (CAR) T cells, an exciting new horizon in anticancer therapy, have shown remarkable clinical efficacy for the management of different haematological malignancies (149-152). However, due to limited clinical trials of CAR-T cells for HIV treatment (117), no significant differences in HIV reservoirs were observed following CAR-T cell therapy. CAR-T cell therapy has been associated with significant side effects, including cytokine release syndrome (CRS) and neurotoxicity, offtarget effects, antigen escape, and life-threatening graft-versus-hostdisease (GVHD) (153-155). NK cells are an exciting novel candidate platform for CAR-engineering due to their increased safety, modifiability, and superior cytotoxicity. There has been no evidence of the adverse events associated with CAR-T cell therapy when CAR-NK cell therapy has been employed for hematological malignancies (156). Researchers are working to exploit CAR-NK cells derived from the NK-92 cell line, stem cells, or cord blood for the treatment of multiple tumors. In addition, humanized mice treated with both CD4ζ-modified and CD4ζ-unmodified human embryonic stem cell (hESC)-NK or iPSC-NK cells can significantly suppress HIV replication, despite the fact that CD4 ζ expression is not associated with increased suppression of HIV (157). Another study reported that CAR-modified HPCs could differentiate into functional NK cells and T cells, which can prevent HIV infection in humanized mice and pave the way for HIV clearance (158). CAR-NK cells can detect and destroy mimetic HIV-infected cell lines, especially primary CD4⁺ T cells expressing gp160 subtypes B and C, making CAR-NK cells a viable approach for HIV eradication (159). Due to their unique characteristics, NK cells are promising candidates for CAR-based cellular immunotherapy in controlling HIV and HIV associated cancers more specifically and efficiently.

CONCLUSION

In addition to T cells, NK cells are an efficient part of the immune response to HIV and act as a bridge between the adaptive and innate immune systems. Several NK cell-based therapeutics are currently used to treat cancer, and it is likely that they will be redeployed to treat HIV. Some researches has focused on natural killer cell-based HIV immunotherapeutics (53, 74, 88, 160, 161). Herein, we outline several approaches and highlight ongoing studies for treating HIV by enhancement of NK cell activation or



FIGURE 2 | Overview of strategies targeting NK cells for HIV-1 treatment. (A) NK cell activation induced by NK cell receptors. (B) Enhancing NK cell function by ADCC and specific antibodies. (C) Activating and enhancing NK cell activity by TLR agonists, IL-15 agonists, and immune checkpoint inhibitors. (D) Adoptive cell transfer, including that of haploidentical NK cells, allogeneic NK cells, and CAR-NK cells.

cytotoxicity (**Figure 2**). NK cell super-agonists may also be useful for HIV viral suppression and reservoir clearance, and NK cell transfer is another powerful weapon for fighting HIV. CAR-NK cells are advantageous because they do not require HLAmatching and are abundantly available; therefore, they have the potential to be made into off-the-shelf products that can be used immediately and extensively in the clinic. Donor-related variability in NK cell expansion makes the utilization of NK cell lines an efficient and attractive substitute for cell therapy. Strategies that combine NK cell-based immunotherapy with other therapeutic approaches have shown success in numerous of clinical trials. In conclusion, the inherent nature of NK cells will likely make them a crucial tool for future multimodal strategies to clear HIV-1.

REFERENCES

- Liu C, Ma X, Liu B, Chen C, Zhang H. HIV-1 Functional Cure: Will the Dream Come True? *BMC Med* (2015) 13:284. doi: 10.1186/s12916-015-0517-y
- Carter CA, Ehrlich LS. Cell Biology of HIV-1 Infection of Macrophages. *Annu Rev Microbiol* (2008) 62:425–43. doi: 10.1146/annurev. micro.62.081307.162758
- Aquaro S, Caliò R, Balzarini J, Bellocchi MC, Garaci E, Perno CF. Macrophages and HIV Infection: Therapeutical Approaches Toward This Strategic Virus Reservoir. *Antiviral Res* (2002) 55:209–25. doi: 10.1016/ s0166-3542(02)00052-9
- Sattentau Q. Avoiding the Void: Cell-to-Cell Spread of Human Viruses. Nat Rev Microbiol (2008) 6:815–26. doi: 10.1038/nrmicro1972
- Fauci AS, Pantaleo G, Stanley S, Weissman D. Immunopathogenic Mechanisms of HIV Infection. Ann Intern Med (1996) 124:654–63. doi: 10.7326/0003-4819-124-7-199604010-00006
- Franks SE, Wolfson B, Hodge JW. Natural Born Killers: NK Cells in Cancer Therapy. Cancers (Basel) (2020) 12(8):2131. doi: 10.3390/cancers12082131
- Hu W, Wang G, Huang D, Sui M, Xu Y. Cancer Immunotherapy Based on Natural Killer Cells: Current Progress and New Opportunities. Front Immunol (2019) 10:1205:1205. doi: 10.3389/fimmu.2019.01205
- Myers JA, Miller JS. Exploring the NK Cell Platform for Cancer Immunotherapy. Nat Rev Clin Oncol (2021) 18:85–100. doi: 10.1038/ s41571-020-0426-7
- Liu S, Galat V, Galat Y, Lee YKA, Wainwright D, Wu J. NK Cell-Based Cancer Immunotherapy: From Basic Biology to Clinical Development. *J Hematol Oncol* (2021) 14:7. doi: 10.1186/s13045-020-01014-w
- Highton AJ, Schuster IS, Degli-Esposti MA, Altfeld M. The Role of Natural Killer Cells in Liver Inflammation. *Semin Immunopathol* (2021) 43:519–33. doi: 10.1007/s00281-021-00877-6
- Robertson MJ, Ritz J. Biology and Clinical Relevance of Human Natural Killer Cells. Blood (1990) 76:2421–38. doi: 10.1182/blood.V76.12.2421.2421
- Galy A, Travis M, Cen D, Chen B. Human T, B, Natural Killer, and Dendritic Cells Arise From a Common Bone Marrow Progenitor Cell Subset. *Immunity* (1995) 3:459–73. doi: 10.1016/1074-7613(95)90175-2
- Miller JS, Alley KA, McGlave P. Differentiation of Natural Killer (NK) Cells From Human Primitive Marrow Progenitors in a Stroma-Based Long-Term Culture System: Identification of a CD34+7+ NK Progenitor. *Blood* (1994) 83:2594–601. doi: 10.1182/blood.V83.9.2594.2594
- Mattei F, Schiavoni G, Belardelli F, Tough DF. IL-15 is Expressed by Dendritic Cells in Response to Type I IFN, Double-Stranded RNA, or Lipopolysaccharide and Promotes Dendritic Cell Activation. *J Immunol* (2001) 167:1179–87. doi: 10.4049/jimmunol.167.3.1179
- Ma M, Wang Z, Chen X, Tao A, He L, Fu S, et al. NKG2C(+)NKG2A(-) Natural Killer Cells are Associated With a Lower Viral Set Point and may Predict Disease Progression in Individuals With Primary HIV Infection. *Front Immunol* (2017) 8:1176. doi: 10.3389/fimmu.2017.01176

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- Moroso V, Metselaar HJ, Mancham S, Tilanus HW, Eissens D, van der Meer A, et al. Liver Grafts Contain a Unique Subset of Natural Killer Cells That are Transferred Into the Recipient After Liver Transplantation. *Liver Transpl* (2010) 16:895–908. doi: 10.1002/lt.22080
- Hudspeth K, Donadon M, Cimino M, Pontarini E, Tentorio P, Preti M, et al. Human Liver-Resident CD56(bright)/CD16(neg) NK Cells are Retained Within Hepatic Sinusoids via the Engagement of CCR5 and CXCR6 Pathways. J Autoimmun (2016) 66:40–50. doi: 10.1016/j.jaut.2015.08.011
- Horowitz A, Strauss-Albee DM, Leipold M, Kubo J, Nemat-Gorgani N, Dogan OC, et al. Genetic and Environmental Determinants of Human NK Cell Diversity Revealed by Mass Cytometry. *Sci Transl Med* (2013) 5:208ra145. doi: 10.1126/scitranslmed.3006702
- Vivier E, Tomasello E, Baratin M, Walzer T, Ugolini S. Functions of Natural Killer Cells. Nat Immunol (2008) 9:503–10. doi: 10.1038/ni1582
- Prager I, Liesche C, van Ooijen H, Urlaub D, Verron Q, Sandström N, et al. NK Cells Switch From Granzyme B to Death Receptor-Mediated Cytotoxicity During Serial Killing. J Exp Med (2019) 216:2113–27. doi: 10.1084/jem.20181454
- Bournazos S, Wang TT, Dahan R, Maamary J, Ravetch JV. Signaling by Antibodies: Recent Progress. Annu Rev Immunol (2017) 35:285–311. doi: 10.1146/annurev-immunol-051116-052433
- Walzer T, Dalod M, Robbins SH, Zitvogel L, Vivier E. Natural-Killer Cells and Dendritic Cells: "L'union Fait La Force". *Blood* (2005) 106:2252–8. doi: 10.1182/blood-2005-03-1154
- Waggoner SN, Reighard SD, Gyurova IE, Cranert SA, Mahl SE, Karmele EP, et al. Roles of Natural Killer Cells in Antiviral Immunity. *Curr Opin Virol* (2016) 16:15–23. doi: 10.1016/j.coviro.2015.10.008
- Dogra P, Rancan C, Ma W, Toth M, Senda T, Carpenter DJ, et al. Tissue Determinants of Human NK Cell Development, Function, and Residence. *Cell* (2020) 180:749–63.e13. doi: 10.1016/j.cell.2020.01.022
- Alter G, Teigen N, Ahern R, Streeck H, Meier A, Rosenberg ES, et al. Evolution of Innate and Adaptive Effector Cell Functions During Acute HIV-1 Infection. J Infect Dis (2007) 195:1452–60. doi: 10.1086/513878
- 26. Mavilio D, Benjamin J, Daucher M, Lombardo G, Kottilil S, Planta MA, et al. Natural Killer Cells in HIV-1 Infection: Dichotomous Effects of Viremia on Inhibitory and Activating Receptors and Their Functional Correlates. *Proc Natl Acad Sci U S A* (2003) 100:15011–6. doi: 10.1073/ pnas.2336091100
- Fauci AS, Mavilio D, Kottilil S. NK Cells in HIV Infection: Paradigm for Protection or Targets for Ambush. *Nat Rev Immunol* (2005) 5:835–43. doi: 10.1038/nri1711
- Brunetta E, Hudspeth KL, Mavilio D. Pathologic Natural Killer Cell Subset Redistribution in HIV-1 Infection: New Insights in Pathophysiology and Clinical Outcomes. J Leukoc Biol (2010) 88:1119–30. doi: 10.1189/ jlb.0410225
- Mikulak J, Oriolo F, Zaghi E, Di Vito C, Mavilio D. Natural Killer Cells in HIV-1 Infection and Therapy. *Aids* (2017) 31:2317–30. doi: 10.1097/ qad.00000000001645

- 30. Tarazona R, Casado JG, Delarosa O, Torre-Cisneros J, Villanueva JL, Sanchez B, et al. Selective Depletion of CD56(dim) NK Cell Subsets and Maintenance of CD56(bright) NK Cells in Treatment-Naive HIV-1-Seropositive Individuals. J Clin Immunol (2002) 22:176–83. doi: 10.1023/ a:1015476114409
- Bruunsgaard H, Pedersen C, Skinhøj P, Pedersen BK. Clinical Progression of HIV Infection: Role of NK Cells. Scand J Immunol (1997) 46:91–5. doi: 10.1046/j.1365-3083.1997.d01-98.x
- Scott-Algara D, Truong LX, Versmisse P, David A, Luong TT, Nguyen NV, et al. Cutting Edge: Increased NK Cell Activity in HIV-1-Exposed But Uninfected Vietnamese Intravascular Drug Users. J Immunol (2003) 171:5663–7. doi: 10.4049/jimmunol.171.11.5663
- Montoya CJ, Velilla PA, Chougnet C, Landay AL, Rugeles MT. Increased IFN-Gamma Production by NK and CD3+/CD56+ Cells in Sexually HIV-1-Exposed But Uninfected Individuals. *Clin Immunol* (2006) 120:138–46. doi: 10.1016/j.clim.2006.02.008
- 34. Oliva A, Kinter AL, Vaccarezza M, Rubbert A, Catanzaro A, Moir S, et al. Natural Killer Cells From Human Immunodeficiency Virus (HIV)-Infected Individuals are an Important Source of CC-Chemokines and Suppress HIV-1 Entry and Replication *In Vitro. J Clin Invest* (1998) 102:223–31. doi: 10.1172/jci2323
- 35. Bradley T, Peppa D, Pedroza-Pacheco I, Li D, Cain DW, Henao R, et al. RAB11FIP5 Expression and Altered Natural Killer Cell Function Are Associated With Induction of HIV Broadly Neutralizing Antibody Responses. Cell (2018) 175:387–99.e17. doi: 10.1016/j.cell.2018.08.064
- Flórez-Álvarez L, Hernandez JC, Zapata W. NK Cells in HIV-1 Infection: From Basic Science to Vaccine Strategies. *Front Immunol* (2018) 9:2290:2290. doi: 10.3389/fimmu.2018.02290
- Deeks SG. HIV: Shock and Kill. Nature (2012) 487:439–40. doi: 10.1038/ 487439a
- Garrido C, Spivak AM, Soriano-Sarabia N, Checkley MA, Barker E, Karn J, et al. HIV Latency-Reversing Agents Have Diverse Effects on Natural Killer Cell Function. *Front Immunol* (2016) 7:356. doi: 10.3389/ fimmu.2016.00356
- Olesen R, Vigano S, Rasmussen TA, Søgaard OS, Ouyang Z, Buzon M, et al. Innate Immune Activity Correlates With CD4 T Cell-Associated HIV-1 DNA Decline During Latency-Reversing Treatment With Panobinostat. *J Virol* (2015) 89:10176–89. doi: 10.1128/jvi.01484-15
- Hua S, Vigano S, Tse S, Zhengyu O, Harrington S, Negron J, et al. Pegylated Interferon-α-Induced Natural Killer Cell Activation Is Associated With Human Immunodeficiency Virus-1 DNA Decline in Antiretroviral Therapy-Treated HIV-1/Hepatitis C Virus-Coinfected Patients. *Clin Infect Dis* (2018) 66:1910–7. doi: 10.1093/cid/cix1111
- Papasavvas E, Azzoni L, Kossenkov AV, Dawany N, Morales KH, Fair M, et al. NK Response Correlates With HIV Decrease in Pegylated IFN-α2a-Treated Antiretroviral Therapy-Suppressed Subjects. J Immunol (2019) 203:705–17. doi: 10.4049/jimmunol.1801511
- 42. Giavedoni LD, Velasquillo MC, Parodi LM, Hubbard GB, Hodara VL. Cytokine Expression, Natural Killer Cell Activation, and Phenotypic Changes in Lymphoid Cells From Rhesus Macaques During Acute Infection With Pathogenic Simian Immunodeficiency Virus. J Virol (2000) 74:1648–57. doi: 10.1128/jvi.74.4.1648-1657.2000
- Moretta A, Bottino C, Vitale M, Pende D, Cantoni C, Mingari MC, et al. Activating Receptors and Coreceptors Involved in Human Natural Killer Cell-Mediated Cytolysis. *Annu Rev Immunol* (2001) 19:197–223. doi: 10.1146/annurev.immunol.19.1.197
- Kiessling R, Klein E, Pross H, Wigzell H. "Natural" Killer Cells in the Mouse. II. Cytotoxic Cells With Specificity for Mouse Moloney Leukemia Cells. Characteristics of the Killer Cell. *Eur J Immunol* (1975) 5:117–21. doi: 10.1002/eji.1830050209
- 45. Parato KG, Kumar A, Badley AD, Sanchez-Dardon JL, Chambers KA, Young CD, et al. Normalization of Natural Killer Cell Function and Phenotype With Effective Anti-HIV Therapy and the Role of IL-10. *Aids* (2002) 16:1251–6. doi: 10.1097/00002030-200206140-00007
- 46. Mavilio D, Lombardo G, Kinter A, Fogli M, La Sala A, Ortolano S, et al. Characterization of the Defective Interaction Between a Subset of Natural Killer Cells and Dendritic Cells in HIV-1 Infection. J Exp Med (2006) 203:2339–50. doi: 10.1084/jem.20060894

- Barrow AD, Martin CJ, Colonna M. The Natural Cytotoxicity Receptors in Health and Disease. Front Immunol (2019) 10:909. doi: 10.3389/fimmu. 2019.00909
- Shimasaki N, Jain A, Campana D. NK Cells for Cancer Immunotherapy. Nat Rev Drug Discov (2020) 19:200-18. doi: 10.1038/s41573-019-0052-1
- López-Botet M, Llano M, Navarro F, Bellón T. NK Cell Recognition of Non-Classical HLA Class I Molecules. *Semin Immunol* (2000) 12:109–19. doi: 10.1006/smim.2000.0213
- Pende D, Falco M, Vitale M, Cantoni C, Vitale C, Munari E, et al. Killer Ig-Like Receptors (KIRs): Their Role in NK Cell Modulation and Developments Leading to Their Clinical Exploitation. *Front Immunol* (2019) 10:1179. doi: 10.3389/fimmu.2019.01179
- Fu GF, Hao S, Zhao JL, Xu XQ, Guo HX, Hu HY, et al. Changes in NK Cell Counts and Receptor Expressions and Emergence of CD3(dim)/CD56+ Cells in HIV-1 Infected Patients in China. *Viral Immunol* (2009) 22:105–16. doi: 10.1089/vim.2008.0081
- Zhang Z, Zhou Y, Lu J, Chen YF, Hu HY, Xu XQ, et al. Changes in NK Cell Subsets and Receptor Expressions in HIV-1 Infected Chronic Patients and HIV Controllers. *Front Immunol* (2021) 12:792775. doi: 10.3389/fimmu. 2021.792775
- Klein K, Hölzemer A, Wang T, Kim TE, Dugan HL, Jost S, et al. A Genome-Wide CRISPR/Cas9-Based Screen Identifies Heparan Sulfate Proteoglycans as Ligands of Killer-Cell Immunoglobulin-Like Receptors. *Front Immunol* (2021) 12:798235:798235. doi: 10.3389/fimmu.2021.798235
- Jiang YJ, Zhang ZN, Diao YY, Geng WQ, Dai D, Liu J, et al. Distribution of Natural Killer Cell Receptors in HIV Infected Individuals. *Chin Med J (Engl)* (2007) 120:1544–8. doi: 10.1097/00029330-200709010-00015
- Kottilil S, Shin K, Planta M, McLaughlin M, Hallahan CW, Ghany M, et al. Expression of Chemokine and Inhibitory Receptors on Natural Killer Cells: Effect of Immune Activation and HIV Viremia. *J Infect Dis* (2004) 189:1193– 8. doi: 10.1086/382090
- 56. De Maria A, Fogli M, Costa P, Murdaca G, Puppo F, Mavilio D, et al. The Impaired NK Cell Cytolytic Function in Viremic HIV-1 Infection is Associated With a Reduced Surface Expression of Natural Cytotoxicity Receptors (NKp46, NKp30 and Nkp44). *Eur J Immunol* (2003) 33:2410-8. doi: 10.1002/eji.200324141
- Moretta L, Moretta A. Unravelling Natural Killer Cell Function: Triggering and Inhibitory Human NK Receptors. *EMBO J* (2004) 23:255–9. doi: 10.1038/sj.emboj.7600019
- Alter G, Altfeld M. NK Cells in HIV-1 Infection: Evidence for Their Role in the Control of HIV-1 Infection. *J Intern Med* (2009) 265:29–42. doi: 10.1111/ j.1365-2796.2008.02045.x
- Flores-Villanueva PO, Yunis EJ, Delgado JC, Vittinghoff E, Buchbinder S, Leung JY, et al. Control of HIV-1 Viremia and Protection From AIDS are Associated With HLA-Bw4 Homozygosity. *Proc Natl Acad Sci U S A* (2001) 98:5140–5. doi: 10.1073/pnas.071548198
- Martin MP, Gao X, Lee JH, Nelson GW, Detels R, Goedert JJ, et al. Epistatic Interaction Between KIR3DS1 and HLA-B Delays the Progression to AIDS. *Nat Genet* (2002) 31:429–34. doi: 10.1038/ng934
- Marras F, Casabianca A, Bozzano F, Ascierto ML, Orlandi C, Di Biagio A, et al. Control of the HIV-1 DNA Reservoir Is Associated *In Vivo* and *In Vitro* With NKp46/NKp30 (CD335 CD337) Inducibility and Interferon Gamma Production by Transcriptionally Unique NK Cells. *J Virol* (2017) 91 (23):e00647–17. doi: 10.1128/jvi.00647-17
- Marcenaro E, Ferranti B, Moretta A. NK-DC Interaction: On the Usefulness of Auto-Aggression. *Autoimmun Rev* (2005) 4:520-5. doi: 10.1016/ j.autrev.2005.04.015
- Lam VC, Lanier LL. NK Cells in Host Responses to Viral Infections. Curr Opin Immunol (2017) 44:43–51. doi: 10.1016/j.coi.2016.11.003
- 64. Ghasemzadeh M, Ghasemzadeh A, Hosseini E. Exhausted NK Cells and Cytokine Storms in COVID-19: Whether NK Cell Therapy Could be a Therapeutic Choice. *Hum Immunol* (2022) 83(1):86–98. doi: 10.1016/ j.humimm.2021.09.004
- Lanier LL, Phillips JH, Hackett JJr., Tutt M, Kumar V. Natural Killer Cells: Definition of a Cell Type Rather Than a Function. J Immunol (1986) 137:2735–9.
- 66. Tomescu C, Duh FM, Hoh R, Viviani A, Harvill K, Martin MP, et al. Impact of Protective Killer Inhibitory Receptor/Human Leukocyte Antigen

Genotypes on Natural Killer Cell and T-Cell Function in HIV-1-Infected Controllers. *Aids* (2012) 26:1869–78. doi: 10.1097/QAD.0b013e32835861b0

- Ramsuran V, Naranbhai V, Horowitz A, Qi Y, Martin MP, Yuki Y, et al. Elevated HLA-A Expression Impairs HIV Control Through Inhibition of NKG2A-Expressing Cells. *Science* (2018) 359:86–90. doi: 10.1126/ science.aam8825
- Apps R, Del Prete GQ, Chatterjee P, Lara A, Brumme ZL, Brockman MA, et al. HIV-1 Vpu Mediates HLA-C Downregulation. *Cell Host Microbe* (2016) 19:686–95. doi: 10.1016/j.chom.2016.04.005
- 69. Davis ZB, Cogswell A, Scott H, Mertsching A, Boucau J, Wambua D, et al. A Conserved HIV-1-Derived Peptide Presented by HLA-E Renders Infected T-Cells Highly Susceptible to Attack by NKG2A/CD94-Bearing Natural Killer Cells. *PLoS Pathog* (2016) 12:e1005421. doi: 10.1371/journal.ppat.1005421
- McWilliams EM, Mele JM, Cheney C, Timmerman EA, Fiazuddin F, Strattan EJ, et al. Therapeutic CD94/NKG2A Blockade Improves Natural Killer Cell Dysfunction in Chronic Lymphocytic Leukemia. Oncoimmunology (2016) 5:e1226720. doi: 10.1080/2162402x.2016.1226720
- Ruggeri L, Urbani E, André P, Mancusi A, Tosti A, Topini F, et al. Effects of Anti-NKG2A Antibody Administration on Leukemia and Normal Hematopoietic Cells. *Haematologica* (2016) 101:626–33. doi: 10.3324/ haematol.2015.135301
- 72. Li F, Wei H, Wei H, Gao Y, Xu L, Yin W, et al. Blocking the Natural Killer Cell Inhibitory Receptor NKG2A Increases Activity of Human Natural Killer Cells and Clears Hepatitis B Virus Infection in Mice. *Gastroenterology* (2013) 144:392–401. doi: 10.1053/j.gastro.2012.10.039
- Alrubayyi A, Ogbe A, Moreno Cubero E, Peppa D. Harnessing Natural Killer Cell Innate and Adaptive Traits in HIV Infection. Front Cell Infect Microbiol (2020) 10:395:395. doi: 10.3389/fcimb.2020.00395
- 74. Ziegler MC, Nelde A, Weber JK, Schreitmüller CM, Martrus G, Huynh T, et al. HIV-1 Induced Changes in HLA-C*03: 04-Presented Peptide Repertoires Lead to Reduced Engagement of Inhibitory Natural Killer Cell Receptors. *Aids* (2020) 34:1713–23. doi: 10.1097/qad.00000000002596
- Battle-Miller K, Eby CA, Landay AL, Cohen MH, Sha BE, Baum LL. Antibody-Dependent Cell-Mediated Cytotoxicity in Cervical Lavage Fluids of Human Immunodeficiency Virus Type 1–Infected Women. J Infect Dis (2002) 185:439–47. doi: 10.1086/338828
- Nag P, Kim J, Sapiega V, Landay AL, Bremer JW, Mestecky J, et al. Women With Cervicovaginal Antibody-Dependent Cell-Mediated Cytotoxicity Have Lower Genital HIV-1 RNA Loads. J Infect Dis (2004) 190:1970–8. doi: 10.1086/425582
- Wren LH, Chung AW, Isitman G, Kelleher AD, Parsons MS, Amin J, et al. Specific Antibody-Dependent Cellular Cytotoxicity Responses Associated With Slow Progression of HIV Infection. *Immunology* (2013) 138:116–23. doi: 10.1111/imm.12016
- Talathi SP, Shaikh NN, Pandey SS, Saxena VA, Mamulwar MS, Thakar MR. Fcγriiia Receptor Polymorphism Influences NK Cell Mediated ADCC Activity Against HIV. BMC Infect Dis (2019) 19:1053. doi: 10.1186/ s12879-019-4674-z
- Bruel T, Guivel-Benhassine F, Amraoui S, Malbec M, Richard L, Bourdic K, et al. Elimination of HIV-1-Infected Cells by Broadly Neutralizing Antibodies. *Nat Commun* (2016) 7:10844. doi: 10.1038/ncomms10844
- Bournazos S, Klein F, Pietzsch J, Seaman MS, Nussenzweig MC, Ravetch JV. Broadly Neutralizing Anti-HIV-1 Antibodies Require Fc Effector Functions for *In Vivo* Activity. *Cell* (2014) 158:1243–53. doi: 10.1016/ j.cell.2014.08.023
- Ko SY, Pegu A, Rudicell RS, Yang ZY, Joyce MG, Chen X, et al. Enhanced Neonatal Fc Receptor Function Improves Protection Against Primate SHIV Infection. *Nature* (2014) 514:642–5. doi: 10.1038/nature13612
- Lu CL, Murakowski DK, Bournazos S, Schoofs T, Sarkar D, Halper-Stromberg A, et al. Enhanced Clearance of HIV-1-Infected Cells by Broadly Neutralizing Antibodies Against HIV-1 In Vivo. Science (2016) 352:1001–4. doi: 10.1126/science.aaf1279
- Schoofs T, Klein F, Braunschweig M, Kreider EF, Feldmann A, Nogueira L, et al. HIV-1 Therapy With Monoclonal Antibody 3BNC117 Elicits Host Immune Responses Against HIV-1. *Science* (2016) 352:997–1001. doi: 10.1126/science.aaf0972
- Hoffmann MAG, Bar-On Y, Yang Z, Gristick HB, Gnanapragasam PNP, Vielmetter J, et al. Nanoparticles Presenting Clusters of CD4 Expose a

Universal Vulnerability of HIV-1 by Mimicking Target Cells. *Proc Natl Acad Sci U S A* (2020) 117:18719–28. doi: 10.1073/pnas.2010320117

- Liu ZJ, Bai J, Liu FL, Zhang XY, Wang JZ. Focus on the Therapeutic Efficacy of 3BNC117 Against HIV-1: *In Vitro* Studies, *In Vivo* Studies, Clinical Trials and Challenges. *Int Immunopharmacol* (2017) 52:44–50. doi: 10.1016/ j.intimp.2017.08.016
- Tay SS, Carol H, Biro M. TriKEs and BiKEs Join CARs on the Cancer Immunotherapy Highway. *Hum Vaccin Immunother* (2016) 12:2790–6. doi: 10.1080/21645515.2016.1198455
- Li W, Wu Y, Kong D, Yang H, Wang Y, Shao J, et al. One-Domain CD4 Fused to Human Anti-CD16 Antibody Domain Mediates Effective Killing of HIV-1-Infected Cells. *Sci Rep* (2017) 7:9130. doi: 10.1038/s41598-017-07966-3
- Lucar O, Reeves RK, Jost S. A Natural Impact: NK Cells at the Intersection of Cancer and HIV Disease. *Front Immunol* (2019) 10:1850:1850. doi: 10.3389/ fimmu.2019.01850
- Wang C, Li Y, Jia L, Kim JK, Li J, Deng P, et al. CD276 Expression Enables Squamous Cell Carcinoma Stem Cells to Evade Immune Surveillance. *Cell Stem Cell* (2021) 28:1597–613.e7. doi: 10.1016/j.stem.2021.04.011
- Khan M, Arooj S, Wang H. NK Cell-Based Immune Checkpoint Inhibition. Front Immunol (2020) 11:167:167. doi: 10.3389/fimmu.2020.00167
- Schafer JL, Müller-Trutwin MC, Reeves RK. NK Cell Exhaustion: Bad News for Chronic Disease? Oncotarget (2015) 6:21797–8. doi: 10.18632/ oncotarget.5490
- 92. Ahmad R, Sindhu ST, Tran P, Toma E, Morisset R, Menezes J, et al. Modulation of Expression of the MHC Class I-Binding Natural Killer Cell Receptors, and NK Activity in Relation to Viral Load in HIV-Infected/AIDS Patients. J Med Virol (2001) 65:431–40. doi: 10.1002/jmv.2053
- Schafer JL, Li H, Evans TI, Estes JD, Reeves RK. Accumulation of Cytotoxic CD16+ NK Cells in Simian Immunodeficiency Virus-Infected Lymph Nodes Associated With *In Situ* Differentiation and Functional Anergy. *J Virol* (2015) 89:6887–94. doi: 10.1128/jvi.00660-15
- Norris S, Coleman A, Kuri-Cervantes L, Bower M, Nelson M, Goodier MR. PD-1 Expression on Natural Killer Cells and CD8(+) T Cells During Chronic HIV-1 Infection. *Viral Immunol* (2012) 25:329–32. doi: 10.1089/ vim.2011.0096
- 95. Juno JA, Stalker AT, Waruk JL, Oyugi J, Kimani M, Plummer FA, et al. Elevated Expression of LAG-3, But Not PD-1, Is Associated With Impaired iNKT Cytokine Production During Chronic HIV-1 Infection and Treatment. *Retrovirology* (2015) 12:17. doi: 10.1186/s12977-015-0142-z
- Porichis F, Hart MG, Massa A, Everett HL, Morou A, Richard J, et al. Immune Checkpoint Blockade Restores HIV-Specific CD4 T Cell Help for NK Cells. J Immunol (2018) 201:971–81. doi: 10.4049/jimmunol.1701551
- Macedo AB, Novis CL, Bosque A. Targeting Cellular and Tissue HIV Reservoirs With Toll-Like Receptor Agonists. *Front Immunol* (2019) 10:2450. doi: 10.3389/fimmu.2019.02450
- Akira S, Uematsu S, Takeuchi O. Pathogen Recognition and Innate Immunity. Cell (2006) 124:783–801. doi: 10.1016/j.cell.2006.02.015
- Margolis DM, Archin NM, Cohen MS, Eron JJ, Ferrari G, Garcia JV, et al. Curing HIV: Seeking to Target and Clear Persistent Infection. *Cell* (2020) 181:189–206. doi: 10.1016/j.cell.2020.03.005
- 100. Adib-Conquy M, Scott-Algara D, Cavaillon JM, Souza-Fonseca-Guimaraes F. TLR-Mediated Activation of NK Cells and Their Role in Bacterial/Viral Immune Responses in Mammals. *Immunol Cell Biol* (2014) 92:256–62. doi: 10.1038/icb.2013.99
- 101. Lauzon NM, Mian F, MacKenzie R, Ashkar AA. The Direct Effects of Toll-Like Receptor Ligands on Human NK Cell Cytokine Production and Cytotoxicity. Cell Immunol (2006) 241:102–12. doi: 10.1016/ j.cellimm.2006.08.004
- Noh JY, Yoon SR, Kim TD, Choi I, Jung H. Toll-Like Receptors in Natural Killer Cells and Their Application for Immunotherapy. J Immunol Res (2020) 2020:2045860. doi: 10.1155/2020/2045860
- 103. Rammensee H-G, Wiesmüller K-H, Chandran PA, Zelba H, Rusch E, Gouttefangeas C, et al. A New Synthetic Toll-Like Receptor 1/2 Ligand Is an Efficient Adjuvant for Peptide Vaccination in a Human Volunteer. *J Immunother Cancer* (2019) 7(1):307. doi: 10.1186/s40425-019-0796-5
- 104. Riise RE, Bernson E, Aurelius J, Martner A, Pesce S, Della Chiesa M, et al. TLR-Stimulated Neutrophils Instruct NK Cells To Trigger Dendritic Cell

Maturation and Promote Adaptive T Cell Responses. J Immunol (2015) 195 (3):1121-8. doi: 10.4049/jimmunol.1500709

- 105. van Beek JJP, Gorris MAJ, Sköld AE, Hatipoglu I, Van Acker HH, Smits EL, et al. Human Blood Myeloid and Plasmacytoid Dendritic Cells Cross Activate Each Other and Synergize in Inducing NK Cell Cytotoxicity. Oncoimmunology (2016) 5(10):e1227902. doi: 10.1080/2162402X. 2016.1227902
- 106. Chalifour A, Jeannin P, Gauchat JF, Blaecke A, Malissard M, N'Guyen T, et al. Direct Bacterial Protein PAMP Recognition by Human NK Cells Involves TLRs and Triggers Alpha-Defensin Production. *Blood* (2004) 104:1778–83. doi: 10.1182/blood-2003-08-2820
- 107. Hart OM, Athie-Morales V, O'Connor GM, Gardiner CM. TLR7/8-Mediated Activation of Human NK Cells Results in Accessory Cell-Dependent IFN-Gamma Production. J Immunol (2005) 175:1636–42. doi: 10.4049/jimmunol.175.3.1636
- Pisegna S, Pirozzi G, Piccoli M, Frati L, Santoni A, Palmieri G. P38 MAPK Activation Controls the TLR3-Mediated Up-Regulation of Cytotoxicity and Cytokine Production in Human NK Cells. *Blood* (2004) 104:4157–64. doi: 10.1182/blood-2004-05-1860
- 109. Roda JM, Parihar R, Carson WE 3rd. CpG-Containing Oligodeoxynucleotides Act Through TLR9 to Enhance the NK Cell Cytokine Response to Antibody-Coated Tumor Cells. J Immunol (2005) 175:1619–27. doi: 10.4049/jimmunol.175.3.1619
- 110. Saxena M, Sabado RL, La Mar M, Mohri H, Salazar AM, Dong H, et al. Poly-ICLC, a TLR3 Agonist, Induces Transient Innate Immune Responses in Patients With Treated HIV-Infection: A Randomized Double-Blinded Placebo Controlled Trial. *Front Immunol* (2019) 10:725:725. doi: 10.3389/ fmmu.2019.00725
- 111. Borducchi EN, Liu J, Nkolola JP, Cadena AM, Yu WH, Fischinger S, et al. Antibody and TLR7 Agonist Delay Viral Rebound in SHIV-Infected Monkeys. *Nature* (2018) 563:360–4. doi: 10.1038/s41586-018-0600-6
- 112. Gorski KS, Waller EL, Bjornton-Severson J, Hanten JA, Riter CL, Kieper WC, et al. Distinct Indirect Pathways Govern Human NK-Cell Activation by TLR-7 and TLR-8 Agonists. *Int Immunol* (2006) 18:1115–26. doi: 10.1093/ intimm/dxl046
- 113. Vibholm L, Schleimann MH, Højen JF, Benfield T, Offersen R, Rasmussen K, et al. Short-Course Toll-Like Receptor 9 Agonist Treatment Impacts Innate Immunity and Plasma Viremia in Individuals With Human Immunodeficiency Virus Infection. *Clin Infect Dis* (2017) 64:1686–95. doi: 10.1093/cid/cix201
- 114. Tsai A, Irrinki A, Kaur J, Cihlar T, Kukolj G, Sloan DD, et al. Toll-Like Receptor 7 Agonist GS-9620 Induces HIV Expression and HIV-Specific Immunity in Cells From HIV-Infected Individuals on Suppressive Antiretroviral Therapy. J Virol (2017) 91(8):e02166–16. doi: 10.1128/ jvi.02166-16
- 115. Garrido C, Abad-Fernandez M, Tuyishime M, Pollara JJ, Ferrari G, Soriano-Sarabia N, et al. Interleukin-15-Stimulated Natural Killer Cells Clear HIV-1-Infected Cells Following Latency Reversal Ex Vivo. J Virol (2018) 92(12): e00235-18. doi: 10.1128/jvi.00235-18
- 116. Jones RB, Mueller S, O'Connor R, Rimpel K, Sloan DD, Karel D, et al. A Subset of Latency-Reversing Agents Expose HIV-Infected Resting CD4+ T-Cells to Recognition by Cytotoxic T-Lymphocytes. *PLoS Pathog* (2016) 12: e1005545. doi: 10.1371/journal.ppat.1005545
- 117. McBrien JB, Mavigner M, Franchitti L, Smith SA, White E, Tharp GK, et al. Robust and Persistent Reactivation of SIV and HIV by N-803 and Depletion of CD8(+) Cells. *Nature* (2020) 578:154–9. doi: 10.1038/ s41586-020-1946-0
- 118. Ellis-Connell AL, Balgeman AJ, Zarbock KR, Barry G, Weiler A, Egan JO, et al. ALT-803 Transiently Reduces Simian Immunodeficiency Virus Replication in the Absence of Antiretroviral Treatment. *J Virol* (2018) 92 (3):e01748–17. doi: 10.1128/jvi.01748-17
- 119. Romee R, Cooley S, Berrien-Elliott MM, Westervelt P, Verneris MR, Wagner JE, et al. First-In-Human Phase 1 Clinical Study of the IL-15 Superagonist Complex ALT-803 to Treat Relapse After Transplantation. *Blood* (2018) 131:2515–27. doi: 10.1182/blood-2017-12-823757
- 120. Wrangle JM, Velcheti V, Patel MR, Garrett-Mayer E, Hill EG, Ravenel JG, et al. ALT-803, an IL-15 Superagonist, in Combination With Nivolumab in Patients With Metastatic non-Small Cell Lung Cancer: A non-Randomised,

Open-Label, Phase 1b Trial. Lancet Oncol (2018) 19:694-704. doi: 10.1016/s1470-2045(18)30148-7

- 121. Margolin K, Morishima C, Velcheti V, Miller JS, Lee SM, Silk AW, et al. Phase I Trial of ALT-803, A Novel Recombinant IL15 Complex, in Patients With Advanced Solid Tumors. *Clin Cancer Res* (2018) 24:5552–61. doi: 10.1158/1078-0432.Ccr-18-0945
- 122. Karo JM, Sun JC. Novel Molecular Mechanism for Generating NK-Cell Fitness and Memory. *Eur J Immunol* (2015) 45:1906–15. doi: 10.1002/ eji.201445339
- 123. Sun JC, Lanier LL. Is There Natural Killer Cell Memory and Can It Be Harnessed by Vaccination? NK Cell Memory and Immunization Strategies Against Infectious Diseases and Cancer. *Cold Spring Harb Perspect Biol* (2018) 10(10):a029538. doi: 10.1101/cshperspect.a029538
- Holmes TD, Bryceson YT. Natural Killer Cell Memory in Context. Semin Immunol (2016) 28:368–76. doi: 10.1016/j.smim.2016.05.008
- 125. Reeves RK, Li H, Jost S, Blass E, Li H, Schafer JL, et al. Antigen-Specific NK Cell Memory in Rhesus Macaques. Nat Immunol (2015) 16:927–32. doi: 10.1038/ni.3227
- 126. O'Leary JG, Goodarzi M, Drayton DL, von Andrian UH. T Cell- and B Cell-Independent Adaptive Immunity Mediated by Natural Killer Cells. Nat Immunol (2006) 7:507–16. doi: 10.1038/ni1332
- 127. Sun JC, Madera S, Bezman NA, Beilke JN, Kaplan MH, Lanier LL. Proinflammatory Cytokine Signaling Required for the Generation of Natural Killer Cell Memory. J Exp Med (2012) 209:947–54. doi: 10.1084/ jem.20111760
- Sun JC, Beilke JN, Lanier LL. Immune Memory Redefined: Characterizing the Longevity of Natural Killer Cells. *Immunol Rev* (2010) 236:83–94. doi: 10.1111/j.1600-065X.2010.00900.x
- Sun JC, Beilke JN, Lanier LL. Adaptive Immune Features of Natural Killer Cells. Nature (2009) 457:557–61. doi: 10.1038/nature07665
- Cooper MA, Elliott JM, Keyel PA, Yang L, Carrero JA, Yokoyama WM. Cytokine-Induced Memory-Like Natural Killer Cells. Proc Natl Acad Sci USA (2009) 106:1915–9. doi: 10.1073/pnas.0813192106
- Paust S, Blish CA, Reeves RK. Redefining Memory: Building the Case for Adaptive NK Cells. J Virol (2017) 91(20):e00169-17. doi: 10.1128/jvi.00169-17
- Geary CD, Sun JC. Memory Responses of Natural Killer Cells. Semin Immunol (2017) 31:11–9. doi: 10.1016/j.smim.2017.08.012
- 133. Lopez-Vergès S, Milush JM, Schwartz BS, Pando MJ, Jarjoura J, York VA, et al. Expansion of a Unique CD57⁺NKG2Chi Natural Killer Cell Subset During Acute Human Cytomegalovirus Infection. *Proc Natl Acad Sci U S A* (2011) 108:14725–32. doi: 10.1073/pnas.1110900108
- 134. Gumá M, Budt M, Sáez A, Brckalo T, Hengel H, Angulo A, et al. Expansion of CD94/NKG2C+ NK Cells in Response to Human Cytomegalovirus-Infected Fibroblasts. *Blood* (2006) 107:3624–31. doi: 10.1182/blood-2005-09-3682
- 135. Gumá M, Angulo A, Vilches C, Gómez-Lozano N, Malats N, López-Botet M. Imprint of Human Cytomegalovirus Infection on the NK Cell Receptor Repertoire. *Blood* (2004) 104:3664–71. doi: 10.1182/blood-2004-05-2058
- Peppa D, Pedroza-Pacheco I, Pellegrino P, Williams I, Maini MK, Borrow P. Adaptive Reconfiguration of Natural Killer Cells in HIV-1 Infection. Front Immunol (2018) 9:474:474. doi: 10.3389/fimmu.2018.00474
- Peng H, Tian Z. Natural Killer Cell Memory: Progress and Implications. Front Immunol (2017) 8:1143. doi: 10.3389/fimmu.2017.01143
- Sengupta S, Siliciano RF. Targeting the Latent Reservoir for HIV-1. Immunity (2018) 48:872–95. doi: 10.1016/j.immuni.2018.04.030
- 139. Becker PS, Suck G, Nowakowska P, Ullrich E, Seifried E, Bader P, et al. Selection and Expansion of Natural Killer Cells for NK Cell-Based Immunotherapy. *Cancer Immunol Immunother* (2016) 65:477-84. doi: 10.1007/s00262-016-1792-y
- 140. Knorr DA, Bachanova V, Verneris MR, Miller JS. Clinical Utility of Natural Killer Cells in Cancer Therapy and Transplantation. *Semin Immunol* (2014) 26:161–72. doi: 10.1016/j.smim.2014.02.002
- 141. Oyer JL, Pandey V, Igarashi RY, Somanchi SS, Zakari A, Solh M, et al. Natural Killer Cells Stimulated With PM21 Particles Expand and Biodistribute *In Vivo*: Clinical Implications for Cancer Treatment. *Cytotherapy* (2016) 18:653–63. doi: 10.1016/j.jcyt.2016.02.006
- 142. Varudkar N, Oyer JL, Copik A, Parks GD. Oncolytic Parainfluenza Virus Combines With NK Cells to Mediate Killing of Infected and non-Infected

Lung Cancer Cells Within 3D Spheroids: Role of Type I and Type III Interferon Signaling. *J Immunother Cancer* (2021) 9(6):e002373. doi: 10.1136/jitc-2021-002373

- 143. Shah N, Li L, McCarty J, Kaur I, Yvon E, Shaim H, et al. Phase I Study of Cord Blood-Derived Natural Killer Cells Combined With Autologous Stem Cell Transplantation in Multiple Myeloma. Br J Haematol (2017) 177:457– 66. doi: 10.1111/bjh.14570
- 144. Dalle JH, Menezes J, Wagner E, Blagdon M, Champagne J, Champagne MA, et al. Characterization of Cord Blood Natural Killer Cells: Implications for Transplantation and Neonatal Infections. *Pediatr Res* (2005) 57:649–55. doi: 10.1203/01.Pdr.0000156501.55431.20
- 145. Arai S, Meagher R, Swearingen M, Myint H, Rich E, Martinson J, et al. Infusion of the Allogeneic Cell Line NK-92 in Patients With Advanced Renal Cell Cancer or Melanoma: A Phase I Trial. *Cytotherapy* (2008) 10:625–32. doi: 10.1080/14653240802301872
- 146. Zhang C, Oberoi P, Oelsner S, Waldmann A, Lindner A, Tonn T, et al. Chimeric Antigen Receptor-Engineered NK-92 Cells: An Off-The-Shelf Cellular Therapeutic for Targeted Elimination of Cancer Cells and Induction of Protective Antitumor Immunity. *Front Immunol* (2017) 8:533:533. doi: 10.3389/fimmu.2017.00533
- 147. Montagner IM, Penna A, Fracasso G, Carpanese D, Dalla Pietà A, Barbieri V, et al. Anti-PSMA CAR-Engineered NK-92 Cells: An Off-The-Shelf Cell Therapy for Prostate Cancer. *Cells* (2020) 9(6):1382. doi: 10.3390/cells9061382
- 148. Zhu H, Blum RH, Bjordahl R, Gaidarova S, Rogers P, Lee TT, et al. Pluripotent Stem Cell-Derived NK Cells With High-Affinity Noncleavable CD16a Mediate Improved Antitumor Activity. *Blood* (2020) 135:399–410. doi: 10.1182/blood.2019000621
- 149. Maude SL, Laetsch TW, Buechner J, Rives S, Boyer M, Bittencourt H, et al. Tisagenlecleucel in Children and Young Adults With B-Cell Lymphoblastic Leukemia. N Engl J Med (2018) 378:439–48. doi: 10.1056/NEJMoa1709866
- 150. Park JH, Rivière I, Gonen M, Wang X, Sénéchal B, Curran KJ, et al. Long-Term Follow-Up of CD19 CAR Therapy in Acute Lymphoblastic Leukemia. *N Engl J Med* (2018) 378:449–59. doi: 10.1056/NEJMoa1709919
- 151. Porter DL, Levine BL, Kalos M, Bagg A, June CH. Chimeric Antigen Receptor-Modified T Cells in Chronic Lymphoid Leukemia. N Engl J Med (2011) 365:725–33. doi: 10.1056/NEJMoa1103849
- Halford Z, Anderson MK, Bennett LL. Axicabtagene Ciloleucel: Clinical Data for the Use of CAR T-Cell Therapy in Relapsed and Refractory Large B-Cell Lymphoma. Ann Pharmacother (2021) 55:390–405. doi: 10.1177/ 1060028020944233
- 153. Lee DW, Santomasso BD, Locke FL, Ghobadi A, Turtle CJ, Brudno JN, et al. ASTCT Consensus Grading for Cytokine Release Syndrome and Neurologic Toxicity Associated With Immune Effector Cells. *Biol Blood Marrow Transplant* (2019) 25:625–38. doi: 10.1016/j.bbmt.2018.12.758

- 154. Davila ML, Riviere I, Wang X, Bartido S, Park J, Curran K, et al. Efficacy and Toxicity Management of 19-28z CAR T Cell Therapy in B Cell Acute Lymphoblastic Leukemia. *Sci Transl Med* (2014) 6:224ra25. doi: 10.1126/ scitranslmed.3008226
- 155. Neelapu SS, Tummala S, Kebriaei P, Wierda W, Locke FL, Lin Y, et al. Toxicity Management After Chimeric Antigen Receptor T Cell Therapy: One Size Does Not Fit 'ALL'. *Nat Rev Clin Oncol* (2018) 15:218. doi: 10.1038/ nrclinonc.2018.20
- 156. Liu E, Marin D, Banerjee P, Macapinlac HA, Thompson P, Basar R, et al. Use of CAR-Transduced Natural Killer Cells in CD19-Positive Lymphoid Tumors. N Engl J Med (2020) 382:545–53. doi: 10.1056/NEJMoa1910607
- 157. Ni Z, Knorr DA, Bendzick L, Allred J, Kaufman DS. Expression of Chimeric Receptor CD4ζ by Natural Killer Cells Derived From Human Pluripotent Stem Cells Improves *In Vitro* Activity But Does Not Enhance Suppression of HIV Infection *In Vivo. Stem Cells* (2014) 32:1021–31. doi: 10.1002/stem.1611
- Zhen A, Kamata M, Rezek V, Rick J, Levin B, Kasparian S, et al. HIV-Specific Immunity Derived From Chimeric Antigen Receptor-Engineered Stem Cells. *Mol Ther* (2015) 23:1358–67. doi: 10.1038/mt.2015.102
- 159. Lim RM, Rong L, Zhen A, Xie J. A Universal CAR-NK Cell Targeting Various Epitopes of HIV-1 Gp160. ACS Chem Biol (2020) 15:2299–310. doi: 10.1021/acschembio.0c00537
- Bunders MJ, Altfeld M. Can NK Cells Purge HIV Sanctuaries? Nat Med (2017) 23:1254–5. doi: 10.1038/nm.4434
- 161. Sugawara S, Manickam C, Reeves RK. TRIGGERED: Could Refocused Cell Signaling be Key to Natural Killer Cell-Based HIV Immunotherapeutics? *Aids* (2021) 35:165–76. doi: 10.1097/qad.00000000002743

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