Caterpillar Track Complexes in Template-Directed Synthesis and Correlated Molecular Motion**

Shiqi Liu, Dmitry V. Kondratuk, Sophie A. L. Rousseaux, Guzmán Gil-Ramírez, Melanie C. O'Sullivan, Jonathan Cremers, Tim D. W. Claridge, and Harry L. Anderson*

Abstract: Small alterations to the structure of a star-shaped template totally change its mode of operation. The hexapyridyl template directs the conversion of a porphyrin dimer to the cyclic hexamer, but deleting one pyridine site changes the product to the cyclic decamer, while deleting two binding sites changes the product to the cyclic octamer. This surprising switch in selectivity is explained by the formation of 2:1 caterpillar track complexes, in which two template wheels bind inside the nanoring. Caterpillar track complexes can also be prepared by binding the hexapyridyl template inside the 8- and 10-porphyrin nanorings. NMR exchange spectroscopy (EXSY) experiments show that these complexes exhibit correlated motion, in which the conrotatory rotation of the two template wheels is coupled to rotation of the nanoring track. In the case of the 10-porphyrin system, the correlated motion can be locked by binding palladium(II) dichloride between the two templates.

emplate-directed synthesis is a powerful strategy for using reversible non-covalent interactions to control the formation of covalent bonds,^[1] and many spectacular examples have been reported recently.^[2] We have used oligo-pyridines to direct the synthesis of zinc-porphyrin nanorings,^[3] and shown that several small template molecules can cooperate to direct the formation of larger macrocycles, according to a Vernier principle.^[4] Here we demonstrate that a single template hub can be modified to direct the formation of several different

- [*] S. Liu,^[+] Dr. D. V. Kondratuk,^[+] Dr. S. A. L. Rousseaux,^[+] Dr. G. Gil-Ramírez, Dr. M. C. O'Sullivan, J. Cremers, Prof. T. D. W. Claridge, Prof. H. L. Anderson Department of Chemistry, University of Oxford Chemistry Research Laboratory Oxford, OX1 3TA (UK) E-mail: harry.anderson@chem.ox.ac.uk
- [⁺] These authors contributed equally to this work.
- [**] We thank the EPSRC, the ERC (grant 320969), the Clarendon Fund of the University of Oxford (D.V.K.), the Violette & Samuel Glasstone Research Fellowships in Science and NSERC (S.A.L.R.) for generous financial support, and Diamond Light Source for award of beam time on I22. We are grateful to Dr. Andrew Smith and Dr. Marc Malfois (Diamond Light Source) for their assistance with SAXS measurements and to Nuntaporn Kamonsutthipaijit for assistance with analysis of GPC data.

Supporting information for this article is available on the WWW under http://dx.doi.org/10.1002/anie.201412293.

ſ © 2015 The Authors. Published by Wiley-VCH Verlag GmbH & Co. KGaA. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.



Figure 1. Cartoons showing coupling of a porphyrin dimer (P2) in the presence of templates T6, T4 and T5, together with analytical GPC traces (toluene/1 % pyridine, detection at 500 nm) of the crude reaction mixtures from these three reactions. The detailed chemical structures are shown in Figure 2; the product distributions shown here were obtained using the compounds with $R = OC_8H_{17}$. The percentages are isolated yields, for the main product of each reaction, and analytical yields for byproducts.

products, simply by deleting some of the binding sites on the template (Figure 1). Coupling of linear porphyrin dimer P2 in the presence of the regular D_{6h} T6 template gives the cyclic hexamer *c*-P6 (Figure 1 a).^[3b,d] If two adjacent pyridine binding sites of the T6 template are deleted to give T4, then the main product becomes the cyclic octamer *c*-P8 (Figure 1b), or if only one binding site is deleted, the main product is the cyclic decamer c-P10 (Figure 1c). The formation of c-P10 from P2 directed by T5 is a case of Vernier templating, because 10 is the lowest common multiple of 2 and 5, whereas formation of c-P8 directed by T4, illustrates a new type of non-Vernier cooperative templating.^[5] We call the 1:2 nanoring template assemblies c-P8·(T4)₂ and c-P10·(T5)₂ "caterpillar track" complexes because of their resemblance to the wheeled treads commonly used on bulldozers, tanks and tractors. In c-P8·(T4)₂ and c-P10·(T5)₂ the template wheels do not rotate, but similar complexes $c-P8 \cdot (T6)_2$ and $c-P10 \cdot (T6)_2$ can be prepared which undergo correlated motion, like a turning caterpillar track.

Angew. Chem. Int. Ed. 2015, 54, 5355-5359

© 2015 The Authors. Published by Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim





Figure 2. Structures of T4, T5, T6, P2, c-PN.

The molecular structures of the compounds used in this study are shown in Figure 2. Despite their structural similarity, templates T4, T5 and T6 have dramatically different effects on the palladium-catalyzed oxidative coupling of porphyrin dimer P2, as seen from the gel permeation chromatography (GPC) traces of crude reaction mixtures in Figure 1. No detectable trace of *c*-P8 or *c*-P10 is formed when the reaction is carried out in the presence of T6, but these larger nanorings are isolated in yields of 14% and 20%, respectively, by using templates T4 and T5 (with $R = OC_8H_{17}$, *n*-octyloxy). Similar reactions were also carried out using the version of **P2** with R = *t*-Bu, giving *c*-**P8** and *c*-**P10** in isolated yields of 29% and 18%, respectively (see Figure S33 in the Supporting Information for GPC traces). Previously we isolated c-P10 as an unexpected byproduct during the synthesis of *c*-P30 by the Vernier coupling of a linear porphyrin decamer in the presence of T6;^[4a] the formation of a caterpillar track complex now explains how c-P10 was formed in this reaction.[6]

The discovery of "caterpillar track" template effects prompted us to investigate the structures and stabilities of the complexes c-P8·(T4)₂ and c-P10·(T5)₂. UV/Vis/NIR and ¹H NMR titrations showed that 1:2 complexes are formed in solution with high allosteric cooperativity; in both cases, the corresponding 1:1 complexes do not form in detectable concentrations (Figure S35-S37). The nanorings c-P8 and c-P10 also form similar 1:2 caterpillar track complexes with the regular T6 template. The optimized geometries of $c-P8\cdot(T6)_2$ and $c-P10\cdot(T6)_2$, from molecular mechanics calculations (Figure 3) are similar to the geometries of $c-P8\cdot(T4)_2$ and c-P10·(T5)₂ deduced from solution-phase small-angle Xray scattering (SAXS) studies (Figures S48 and S49). In both structures, all the zinc atoms lie in essentially the same plane, and in $c-P8\cdot(T6)_2$, the two template units overlap at the center.

¹H NMR exchange spectroscopy (EXSY) experiments were carried out to elucidate the dynamics of the complexes $c-P8\cdot(T4)_2$, $c-P8\cdot(T6)_2$, $c-P10\cdot(T5)_2$, and $c-P10\cdot(T6)_2$. These experiments demonstrated that the two coordinatively saturated complexes, $c-P8\cdot(T4)_2$ and $c-P10\cdot(T5)_2$, are static, with no exchange between template environments, whereas the two complexes with free pyridine sites, $c-P8\cdot(T6)_2$ and $c-P10\cdot(T6)_2$, undergo caterpillar track motion of the type shown schematically for $c-P8\cdot(T6)_2$ in Figure 4. The 1D EXSY



Figure 3. Two orthogonal views of the calculated structures of a) **c-P8·(T6)**₂ and b) **c-P10·(T6)**₂. (MM+ force field, HyperChem; *meso*aryl side groups were not included in the calculations; hydrogen atoms are omitted for clarity.)

spectra in Figure 5 demonstrate this concerted motion by revealing exchange signals only between environments that are related by a 60° rotation of the template ($m \Leftrightarrow a, a \Leftrightarrow b, l \Leftrightarrow c, and c \Leftrightarrow d$); signals corresponding to a rotation of 120° ($m \Leftrightarrow b$ and $l \Leftrightarrow d$) are absent on the time-scale of the experiment, confirming that the observed exchange peaks are due to intramolecular rotation of bound **T6**, and not due to random exchange of the template units. Under the same conditions, an identical 1D EXSY experiment on c-**P8**·(**T4**)₂ showed no exchange signals between template protons (Figure S43), confirming that the exchange peaks observed in c-**P8**·(**T6**)₂ result from intramolecular motion.

A second set of 1D EXSY NMR experiments, targeting the porphyrin nanoring protons, was performed to confirm that the nanoring and template rotation in c-**P8**·(**T6**)₂ are concerted (i.e. that the motions indicated by the blue and red arrows in Figure 4 proceed at the same rate). The *ortho* proton 7 on the porphyrin aryl side-groups and template proton *d* were chosen for irradiation in these kinetic experiments (Figure 5 and S43). Spectra were recorded under various mixing times (100–600 ms) to determine the relative rates of exchange. Aryl proton 7 generates two types of exchange signals corresponding to 1) the rotation of the aryl





Figure 4. Caterpillar track motion in *c*-**P8**-(**T6**)₂, as revealed by ¹H NMR EXSY experiments. a) Detailed molecular structure. b) Mechanism leading to a concerted 60° rotation of both template units, transforming environments **a** and **c** into **b/m** and **d/l**. The time constant for this 60° rotation is 7.7 s. According to this mechanism, both template units must rotate in the same direction, but the direction of rotation is arbitrary, and subsequent 60° rotation steps could occur in either direction with equal probability.

group $(7 \rightarrow 7';$ rate constant: $4 \text{ s}^{-1})$, and 2) the movement of c-P8 $(7 \rightarrow 5;$ rate constant: $0.13 \text{ s}^{-1})$. The rate of growth of the signals corresponding to rotation of the template $(d \rightarrow c;$ rate constant: $0.13 \text{ s}^{-1})$ is identical, within experimental error, to the rate of rotation of the porphyrin nanoring $(7 \rightarrow 5)$, whereas rotation of the aryl side-groups $(7 \rightarrow 7')$ occurs on a faster time-scale (Figure S44). An identical 1D EXSY NMR experiment using c-P8·(T4)₂ confirmed that, in this case, only exchange signals resulting from the aryl side-group rotation are observed (Figure S43). Overall, the results of these experiments provide strong evidence that the three component complex c-P8·(T6)₂ exhibits concerted "caterpillar track" molecular motion.

Similar 1D EXSY NMR experiments on c-P10·(T6)₂ showed that caterpillar track motion occurs faster in this system than in c-P8·(T6)₂, with a rate constant of $10 \pm 2 \text{ s}^{-1}$ for a 60° rotation, resulting in the observation of exchange signals corresponding to more than one 60° rotation. When the non-coordinated β -pyridyl proton (**a**' in Figure 3) is selectively



Figure 5. a) Structure of one quarter of the symmetrical $c-P8 \cdot (T6)_2$ complex. Arrows indicate the exchange signals detected between various protons in the 1D EXSY NMR experiments. b) (From top to bottom) EXSY NMR spectra (CDCl₃, 700 MHz, 298 K, 400 ms mixing time) for targeted protons *I* (black), *c* (pink), *d* (green), *m* (orange), *a* (red) and ¹H NMR spectrum of *c*-P8 \cdot (T6)₂ highlighting the region corresponding to the signals for T6.

irradiated, a gradual build-up of exchange signals corresponding to a 60° rotation of the template is observed to **b'**, followed by the appearance of exchange signals corresponding to a 120° rotation to **c'**, and finally by signals corresponding to a 180° rotation to **d'** (Figure S45). The delayed appearance of signals **c'** and **d'** is a clear signature of a sequential step-wise exchange process.^[7] This kinetic behavior demonstrates that the template undergoes a step-wise rotation within the nanoring, rather than exchanging through dissociation/association.

Several systems have been reported in which intramolecular rotary motion can be locked by binding a metal cation,^[8–11] which suggest that we might be able to control the motion in a molecular caterpillar track by coordinating a metal between the two templates (Figure 6). The distance



Figure 6. Structure of the c-P10-(T6)2-PdCl2 complex in which caterpillar track motion is locked.

between the two central nitrogen atoms in the energyminimized geometry of c-P10·(T6)₂ (Figure 3b) is 6.8 Å, which is too far to chelate a metal atom, but molecular modeling shows that these pyridine sites can easily move closer together if the complex adopts a helical twist (Figure S50). We tested this idea by carrying out a ¹H NMR titration (Figure S41). When one equivalent of [PdCl₂- $(PhCN)_2$ is added to a solution of *c*-P10·(T6)₂ in CDCl₃, new sharp signals appear corresponding to the formation of a complex of stoichiometry $c-P10\cdot(T6)_2\cdot PdCl_2$. The signal from the central α -pyridyl protons shifts from 8.74 ppm in $c-P10\cdot(T6)_2$ to 8.17 ppm in $c-P10\cdot(T6)_2\cdot PdCl_2$, while other protons are essentially unaffected (Figure S31), confirming that palladium binds between the two central nitrogens. The resonances corresponding to the T6 template protons in this palladium complex are sharper than those in c-P10·(T6)₂, suggesting that the motion has become locked. 1D EXSY NMR experiments on c-P10·(T6)₂·PdCl₂ showed no exchange signals between template protons, confirming that palladium effectively prevents the caterpillar track motion in this complex (Figure S46d).

Caterpillar tracks constitute a new type of supramolecular system exhibiting correlated motion between several movable parts.^[12] Previously studied systems include molecular gears,^[13] ball bearings,^[14] turnstiles,^[15] and rotary transduction modules.^[16] In a caterpillar track, the motion of the two wheels must be conrotatory, whereas in a gear the motion is disrotatory. The results presented here demonstrate a new approach to the transmission of molecular motion. The formation of caterpillar track complexes also provides useful access to large macrocycles, via template directed synthesis.

Keywords: molecular gears · molecular motion · porphyrinoids · supramolecular chemistry · templated synthesis

How to cite: Angew. Chem. Int. Ed. 2015, 54, 5355-5359 Angew. Chem. 2015, 127, 5445-5449

[1] a) S. Anderson, H. L. Anderson, J. K. M. Sanders, Acc. Chem. Res. 1993, 26, 469-475; b) J. E. Beves, B. A. Blight, C. J. Campbell, D. A. Leigh, R. T. McBurney, Angew. Chem. Int. Ed. 2011, 50, 9260-9327; Angew. Chem. 2011, 123, 9428-9499; c) G. T. Spence, P. D. Beer, Acc. Chem. Res. 2013, 46, 571-586.

- [2] a) B. Zhu, H. Chen, W. Lin, Y. Ye, J. Wu, S. Li, J. Am. Chem. Soc. 2014, 136, 15126-15129; b) M. J. Langton, O. A. Blackburn, T. Lang, S. Faulkner, P. D. Beer, Angew. Chem. Int. Ed. 2014, 53, 11463-11466; Angew. Chem. 2014, 126, 11647-11650; c) G. De Bo, S. Kuschel, D. A. Leigh, B. Lewandowski, M. Papmeyer, J. W. Ward, J. Am. Chem. Soc. 2014, 136, 5811-5814; d) J. R. Sanchez-Valencia, T. Dienel, O. Gröning, I. Shorubalko, A. Mueller, M. Jansen, K. Amsharov, P. Ruffieux, R. Fasel, Nature 2014, 512, 61-64; e) H. Omachi, T. Nakayama, E. Takahashi, Y. Segawa, K. Itami, Nat. Chem. 2013, 5, 572-576; f) M. E. Belowich, C. Valente, R. A. Smaldone, D. C. Friedman, J. Thiel, L. Cronin, J. F. Stoddart, J. Am. Chem. Soc. 2012, 134, 5243-5261; g) J. Guo, P. C. Mayers, G. A. Breault, C. A. Hunter, Nat. Chem. 2010, 2, 218-222.
- [3] a) P. Liu, P. Neuhaus, D. V. Kondratuk, T. S. Balaban, H. L. Anderson, Angew. Chem. Int. Ed. 2014, 53, 7770-7773; Angew. Chem. 2014, 126, 7904-7907; b) J. K. Sprafke, D. V. Kondratuk, M. Wykes, A. L. Thompson, M. Hoffmann, R. Drevinskas, W.-H. Chen, C. K. Yong, J. Kärnbratt, J. E. Bullock, M. Malfois, M. R. Wasielewski, B. Albinsson, L. M. Herz, D. Zigmantas, D. Beljonne, H. L. Anderson, J. Am. Chem. Soc. 2011, 133, 17262-17273; c) M. J. Langton, J. D. Matichak, A. L. Thompson, H. L. Anderson, Chem. Sci. 2011, 2, 1897-1901; d) M. Hoffmann, J. Kärnbratt, M.-H. Chang, L. M. Herz, B. Albinsson, H. L. Anderson, Angew. Chem. Int. Ed. 2008, 47, 4993-4996; Angew. Chem. 2008, 120, 5071-5074; e) M. Hoffmann, C. J. Wilson, B. Odell, H. L. Anderson, Angew. Chem. Int. Ed. 2007, 46, 3122-3125; Angew. Chem. 2007, 119, 3183-3186.
- [4] a) D. V. Kondratuk, L. M. A. Perigao, A. M. S. Esmail, J. N. O'Shea, P. H. Beton, H. L. Anderson, Nat. Chem. 2015, DOI: 10.1038/NCHEM.2182; b) D. V. Kondratuk, J. K. Sprafke, M. C. O'Sullivan, L. M. A. Perdigao, A. Saywell, M. Malfois, J. N. O'Shea, P. H. Beton, A. L. Thompson, H. L. Anderson, Chem. Eur. J. 2014, 20, 12826-12834; c) D. V. Kondratuk, L. M. A. Perdigao, M. C. O'Sullivan, S. Svatek, G. Smith, J. N. O'Shea, P. H. Beton, H. L. Anderson, Angew. Chem. Int. Ed. 2012, 51, 6696-6699; Angew. Chem. 2012, 124, 6800-6803; d) M. C. O'Sullivan, J. K. Sprafke, D. V. Kondratuk, C. Rinfray, T. D. W. Claridge, A. Saywell, M. O. Blunt, J. N. O'Shea, P. H. Beton, M. Malfois, H. L. Anderson, Nature 2011, 469, 72-75.
- [5] Nanoring *c*-P8 was first prepared using an octadentate template, which required a 12-step synthesis (Ref. [3e]); the preparation of c-P8 directly from P2 using T4 as a template provides a more practical route to this nanoring.
- [6] The formation of caterpillar track intermediates also explains how c-P12 was formed as an unexpected byproduct during the synthesis of c-P24 by coupling a linear porphyrin hexamer in the presence of an octadentate template (Ref [4c]).
- [7] a) C. L. Perrin, T. J. Dwyer, Chem. Rev. 1990, 90, 935-967; b) M. Pons, O. Millet, Prog. Nucl. Magn. Reson. Spectrosc. 2001, 38, 267-324.
- [8] a) N. Zigon, P. Larpent, A. Jouaiti, N. Kyritsakas, M. W. Hosseini, Chem. Commun. 2014, 50, 5040-5042; b) T. Lang, A. Guenet, E. Graf, N. Kyritsakas, M. W. Hosseini, Chem. Commun. 2010, 46, 3508-3510.
- [9] S. V. Bhosale, C. Chong, C. Forsyth, S. J. Langford, C. P. Woodward, Tetrahedron 2008, 64, 8394-8401.
- [10] S. K. Samanta, M. Schmittel, J. Am. Chem. Soc. 2013, 135, 18794-18797.
- [11] T. R. Kelly, M. C. Bowyer, K. V. Bhaskar, D. Bebbington, A. Garcia, F. Lang, M. H. Kim, M. P. Jette, J. Am. Chem. Soc. 1994, 116, 3657-3658.
- [12] E. R. Kay, D. A. Leigh, F. Zerbetto, Angew. Chem. Int. Ed. 2007, 46, 72-191; Angew. Chem. 2007, 119, 72-196.
- [13] a) H. Iwamura, K. Mislow, Acc. Chem. Res. 1988, 21, 175-182; b) S. Ogi, T. Ikeda, R. Wakabayashi, S. Shinkai, M. Takeuchi, Chem. Eur. J. 2010, 16, 8285-8290.



- [14] a) S. Hiraoka, M. Shiro, M. Shionoya, J. Am. Chem. Soc. 2004, 126, 1214–1218; b) S. Hiraoka, E. Okuno, T. Tanaka, M. Shiro, M. Shionoya, J. Am. Chem. Soc. 2008, 130, 9089–9098.
- [15] A. Carella, J. Jaud, G. Rapenne, J.-P. Launay, Chem. Commun. 2003, 2434–2435.
- [16] H. Kai, S. Nara, K. Kinbara, T. Aida, J. Am. Chem. Soc. 2008, 130, 6725-6727.

Received: December 22, 2014 Revised: January 29, 2015 Published online: February 12, 2015