

RESEARCH

Open Access



Cerebrospinal fluid level of Nogo receptor 1 antagonist lateral olfactory tract usher substance (LOTUS) correlates inversely with the extent of neuroinflammation

Keita Takahashi^{1,2}, Hideyuki Takeuchi¹, Yuji Kurihara², Hiroshi Doi¹, Misako Kunii¹, Kenichi Tanaka¹, Haruko Nakamura¹, Ryoko Fukai¹, Atsuko Tomita-Katsumoto¹, Mikiko Tada¹, Yuichi Higashiyama¹, Hideto Joki¹, Shigeru Koyano¹, Kohtaro Takei^{2*} and Fumiaki Tanaka^{1*}

Abstract

Background: Although inflammation in the central nervous system is responsible for multiple neurological diseases, the lack of appropriate biomarkers makes it difficult to evaluate inflammatory activities in these diseases. Therefore, a new biomarker reflecting neuroinflammation is required for accurate diagnosis, appropriate therapy, and comprehension of pathogenesis of these neurological disorders. We previously reported that the cerebrospinal fluid (CSF) concentration of lateral olfactory tract usher substance (LOTUS), which promotes axonal growth as a Nogo receptor 1 antagonist, negatively correlates with disease activity in multiple sclerosis, suggesting that variation in LOTUS reflects the inflammatory activities and is a useful biomarker to evaluate the disease activity. To extend this observation, we analyzed the variation of LOTUS in the CSF of patients with bacterial and viral meningitis, which are the most common neuroinflammatory diseases.

Methods: CSF samples were retrospectively obtained from patients with meningitis ($n = 40$), who were followed up by CSF study at least twice, and from healthy controls ($n = 27$). Patients were divided into bacterial ($n = 14$) and viral meningitis ($n = 18$) after exclusion of eight patients according to the criteria of this study. LOTUS concentrations, total protein levels, and CSF cell counts in the acute and recovery phases were analyzed chronologically. We also used lipopolysaccharide-injected mice as a model of neuroinflammation to evaluate LOTUS mRNA and protein expression in the brain.

Results: Regardless of whether meningitis was viral or bacterial, LOTUS concentrations in the CSF of patients in acute phase were lower than those of healthy controls. As the patients recovered from meningitis, LOTUS levels in the CSF returned to the normal range. Lipopolysaccharide-injected mice also exhibited reduced LOTUS mRNA and protein expression in the brain.

(Continued on next page)

* Correspondence: kohtaro@med.yokohama-cu.ac.jp; ftanaka@yokohama-cu.ac.jp

²Molecular Medical Bioscience Laboratory, Department of Medical Life Science, Yokohama City University Graduate School of Medical Life Science, Suehiro-cho 1-7-29, Tsurumi-Ku, Yokohama 230-0045, Japan

¹Department of Neurology and Stroke Medicine, Yokohama City University Graduate School of Medicine, 3-9 Fukuura, Kanazawa-Ku, Yokohama 236-0004, Japan



(Continued from previous page)

Conclusions: CSF levels of LOTUS correlated inversely with disease activity in both bacterial and viral meningitis, as well as in multiple sclerosis, because neuroinflammation downregulated LOTUS expression. Our data strongly suggest that variation of CSF LOTUS is associated with neuroinflammation and is useful as a biomarker for a broader range of neuroinflammatory diseases.

Keywords: Lateral olfactory tract usher substance, Neuroinflammation, Biomarker, Nogo receptor, Multiple sclerosis, Meningitis

Background

Inflammation in the central nervous system (CNS) is associated with a wide range of neurological diseases. In particular, infection and autoimmunity cause acute or chronic inflammation in the CNS [1]. However, it is difficult to evaluate neuroinflammation in these diseases, in large part because of the lack of biomarkers suitable for judging their activities [2]. Although total protein concentrations or cell counts in the cerebrospinal fluid (CSF) are the most commonly used biomarkers for neuroinflammation in CNS diseases, they do not always vary in parallel with disease activities. For example, multiple sclerosis (MS), one of the most common neuroinflammatory diseases caused by an autoimmune mechanism, exhibits poor variation in conventional CSF biomarkers [3], making precise evaluation of disease activity difficult [4]. To complement CSF biomarkers, imaging biomarkers have recently been developed, including magnetic resonance imaging with ultra-small superparamagnetic iron oxide, a novel contrast agent, and positron-emission tomography using [^{11}C] (*R*)-PK11195 ligand. These new markers now play an important role in evaluation of the activities of neuroinflammatory diseases, including MS [3, 5]. However, the availability of cutting-edge imaging techniques is generally limited, even in the advanced treatment hospitals and especially in emergency situations. Therefore, the utility of currently available biomarkers for evaluating neuroinflammation is unsatisfactory, creating a demand for new and practical biomarkers.

Recently, we showed that lateral olfactory tract usher substance (LOTUS, also called Crtac1B) is critical for axonal growth. LOTUS acts as a Nogo receptor 1 (NgR1) antagonist, preventing Nogo from binding to NgR1 [6–8]. Furthermore, we showed that LOTUS in the human CSF closely associated with the disease activity of MS, a representative neuroinflammatory disease accompanied by neurodegeneration [9–11]. Because various molecules associated with axonal growth during brain development mediate not only axonal guidance but also immune responses [12–17], our previous findings suggest that LOTUS is intimately involved in neuroinflammation, as well as axonal growth and/or degeneration in MS [18]. Therefore, we hypothesized that variations of LOTUS concentrations in the CSF may also reflect disease activity in other neuroinflammatory diseases.

In this study, to investigate the relationship between LOTUS and neuroinflammation, we analyzed the variation of LOTUS concentrations in CSF of patients with bacterial and viral meningitis. These patients' clinical courses were objectively verified using established CSF markers such as total protein concentration, cell counts, and glucose, in addition to clinical findings, and we also performed comparisons between the acute and recovery phases in the same patients. Furthermore, we investigated the mechanisms underlying the observations in human patients using lipopolysaccharide-injected mice as a disease model of neuroinflammation.

Methods

Participants

All patients enrolled in this study had undergone neurological evaluation at our university hospital and been diagnosed as meningitis. Between January 2008 and September 2017, CSF samples were obtained retrospectively from 40 patients who had been followed up for CSF study at least twice, after initial sampling in the acute phase, and 27 healthy controls. Patients with aseptic ($n = 20$), bacterial ($n = 15$), or tuberculous ($n = 5$) meningitis were classified as follows [19]. Patients with normal CSF glucose level ($\geq 40\%$ of plasma glucose level), monocyte-predominant CSF pleocytosis, or viral antigens were classified into the aseptic group ($n = 20$). Patients in the aseptic group with infectious sign ($n = 18$) were defined as viral meningitis, and two patients were ruled out due to lack of infectious sign. Patients with reduced CSF glucose level ($< 40\%$ of the plasma glucose level), neutrophil-predominant CSF pleocytosis, or positive microbiological culture were classified into the bacterial group ($n = 14$), and one patient was excluded due to complications of hydrocephalus. Patients with *Mycobacterium tuberculosis* gene-positive CSF, as confirmed by polymerase chain reaction, were classified into the tuberculous meningitis group ($n = 5$). In these patients, prolonged recovery period [20], relatively unchanged CSF total protein and cell counts over the short term made it difficult to judge disease activity. Therefore, we excluded patients with tuberculous meningitis from this study. No patients presented with *Cryptococcus* antigen or other fungal antigens in the CSF. All patients improved clinically during observation.

When the follow-up CSF was collected from the same patient twice or more, the last sample was analyzed between January 1, 2008, and January 1, 2014.

Animals and drug treatment

We used 8- to 10-week-old male C57BL/6J mice obtained from Japan SLC (Hamamatsu, Japan). A neuroinflammation response was induced by intraperitoneal injection of 40 mg/kg lipopolysaccharide (LPS; L3129, Sigma-Aldrich, St. Louis, MO, USA); phosphate-buffered saline (PBS) was administered as a negative control. Mice were analyzed 24 or 48 h after treatment.

RNA extraction and reverse transcription–polymerase chain reaction (RT-PCR)

Total RNA was extracted from the brains of LPS- or PBS-injected mice using the miRNeasy Mini Kit (Qiagen, Valencia, CA, USA) and reverse-transcribed using the SuperScript III First-Strand Synthesis SuperMix for RT-PCR (Life Technologies, Rockville, MD, USA). Expression levels of the genes encoding LOTUS (*Lotus*) and hypoxanthine phosphoribosyltransferase 1 (*Hprt1*) were evaluated by quantitative PCR (qPCR) using Power SYBR Green PCR Master Mix (Thermo Fisher Scientific, Wilmington, DE, USA) on a LightCycler® 96 SW 1.1. (Roche, Basel, Switzerland). Relative expression levels were determined using the $\Delta\Delta C_T$ method; the levels of the mRNAs of interest were normalized against the geometric mean level of *Hprt1* mRNA. The following specific primer sets were used:

Mouse *Lotus* sense: 5'-CATGTTCACTGCAGTCAC CAA-3';

Mouse *Lotus* antisense: 5'-TTATTGGTGTGAGAAAG TAGAT-3';

Mouse *Hprt1* sense: 5'-CCTAAGATGAGCGCAAGTT GAA-3';

Mouse *Hprt1* antisense: 5'-CCACAGGACTAGAA CACCTGCTAA-3'.

Assessments of LOTUS protein

LOTUS protein levels in patients' CSF and mouse brains were analyzed by immunoblotting using a

commercially available specific antibody (AF5234, R&D System, Minneapolis, MN, USA). The detailed methodology of this assay is described elsewhere [9]. Experiments using CSF were performed in triplicate. The mean value and coefficient of variation were calculated to certificate intra-assay reproducibility. Relative levels of LOTUS protein in mouse brains were quantified relative to β -actin, used as a loading control. The experiments were carried out in five independent trials.

Statistical analysis

Statistical analysis was performed with Wilcoxon matched-pairs signed-rank test or Student's *t* test as appropriate, using GraphPad Prism version 6.0 (GraphPad Software, La Jolla, CA, USA). $P < 0.05$ was considered statistically significant.

Results

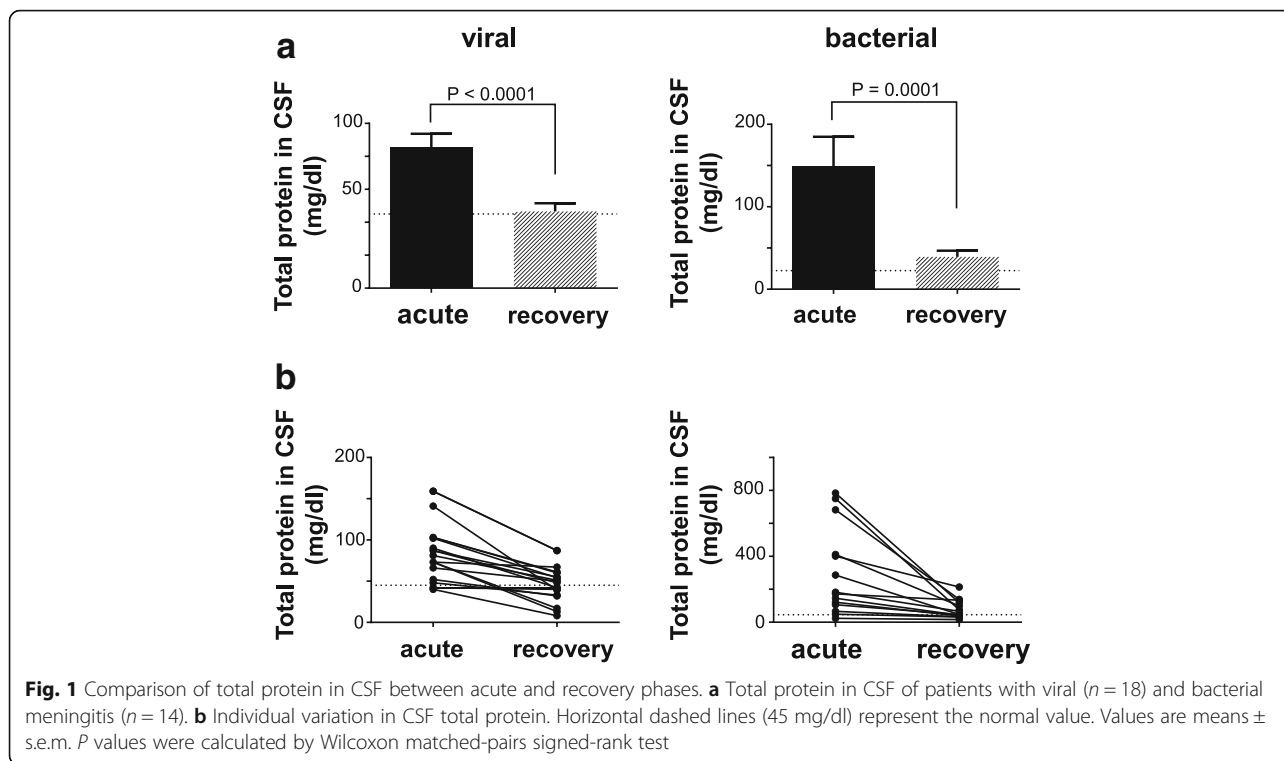
The characteristics of the patients enrolled in this study are summarized in Table 1. As shown in Table 1, the timing of sample collection did not differ significantly between viral and bacterial meningitis in either the acute or recovery phases ($p = 0.31$ and 0.53 , respectively).

First, we examined conventional CSF biomarkers of meningitis, such as total protein and cell counts, in the acute and recovery phases in the same patients. Objective improvements in these markers and clinical recovery were confirmed in all patients with meningitis (Figs. 1 and 2).

We next assessed the CSF LOTUS concentration in the acute and recovery phases by immunoblot analysis (Fig. 3a). The standard value for CSF LOTUS concentration was defined as ≥ 192.1 ng/ml, based on the results in healthy controls (Table 1). Significant differences in LOTUS concentrations were observed between the acute and recovery phases regardless of whether the meningitis was viral or bacterial. The CSF levels of LOTUS significantly decreased in the acute phase of viral and bacterial meningitis, but returned to the normal level in the recovery phase

Table 1 Demographics and clinical characteristics of patients and controls

	Viral meningitis		Bacterial meningitis		Control
Number of patients	18		14		27
Age (year)	37.6 ± 14.9		64.1 ± 21.5		44.4 ± 20.5
Sex (male/female)	6/12		5/9		15/12
	Acute phase	Recovery phase	Acute phase	Recovery phase	–
Time of CSF sampling (days from onset)	3.4 ± 2.2	12.4 ± 4.8	2.5 ± 2.7	13.9 ± 7.9	–
CSF total protein (mg/dl)	85.3 ± 36.6	46.6 ± 21.4	298.5 ± 266.5	78.6 ± 55.8	30.8 ± 12.4
CSF cell count (μ l)	185.9 ± 161.9	78.0 ± 90.0	1017.0 ± 1592.0	37.6 ± 45.0	2.4 ± 2.7
CSF LOTUS level (ng/ml)	133.5 ± 44.2	193.2 ± 83.5	114.2 ± 46.6	187.5 ± 92.1	192.1 ± 47.1

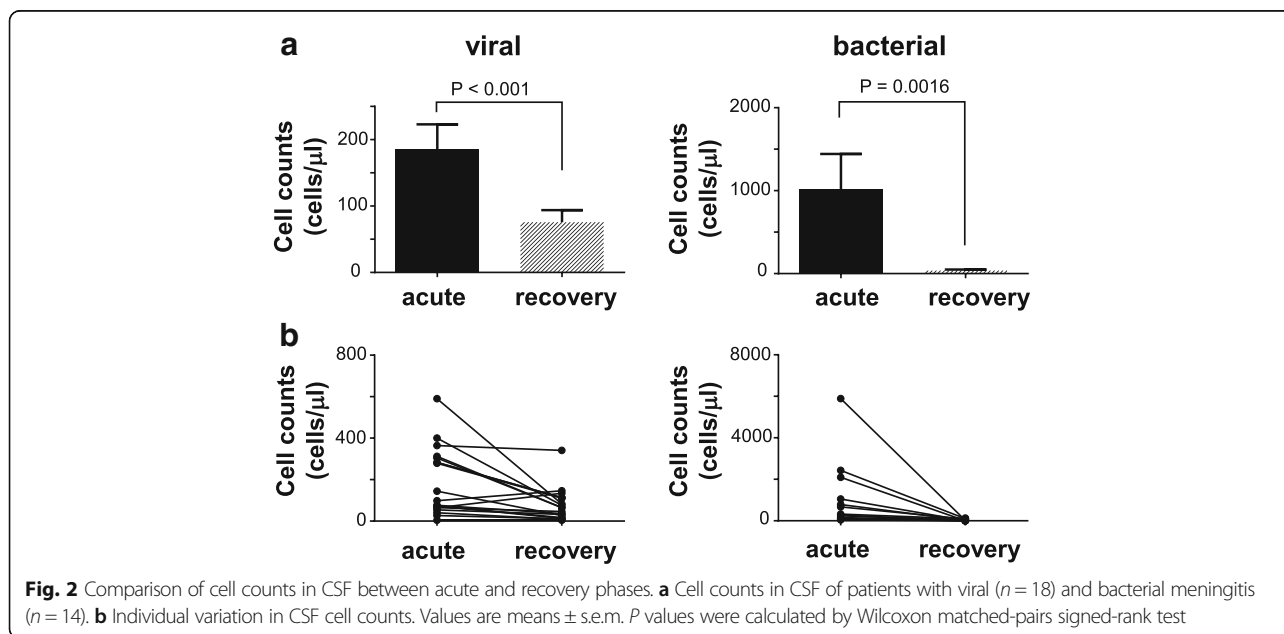


(Fig. 3b). Individual analysis also revealed an increase in LOTUS concentration in all patients with both viral and bacterial meningitis (Fig. 3c).

Finally, we asked whether neuroinflammation downregulates *Lotus* mRNA expression in the brain, using LPS-injected mice as a model. *Lotus* mRNA expression was downregulated in LPS-inflamed brains, followed by a decrease in LOTUS protein levels (Fig. 4).

Discussion

Myelin components in the CNS are important environmental factors that exert an inhibitory effect on axonal growth. Nogo, myelin-associated glycoprotein (MAG), and oligodendrocyte myelin glycoprotein (OMgp) are representative myelin components [21]. These molecules bind to a common receptor, NgR1, and prevent axonal growth, thereby leading to neuronal degeneration [22].



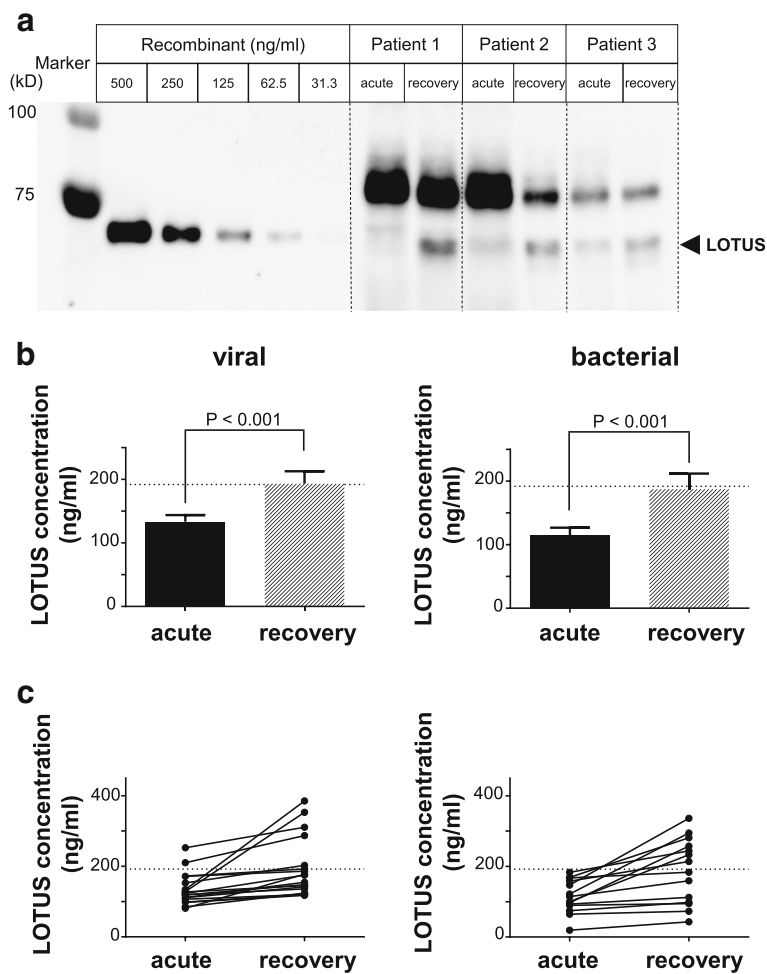
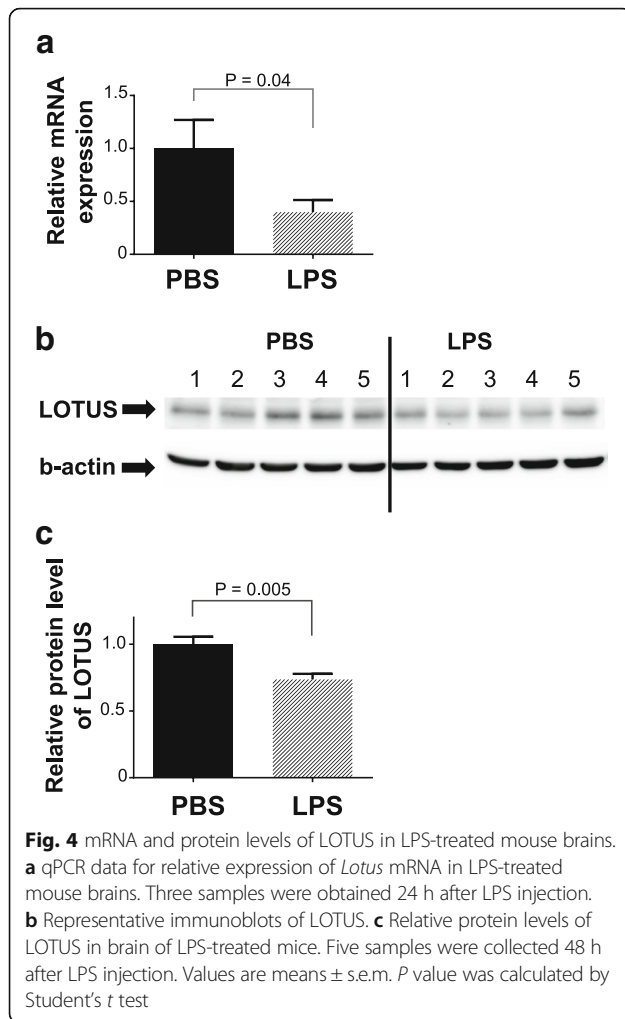


Fig. 3 Comparison of LOTUS concentrations in CSF between acute and recovery phases. **a** Representative immunoblots of CSF LOTUS in acute and recovery phase of three patients with meningitis. The concentration of LOTUS in the CSF was calculated using a standard curve generated from blot band intensities of recombinant protein. **b** LOTUS concentration in CSF of patients with viral ($n = 18$) and bacterial meningitis ($n = 14$). Horizontal dashed lines (192.1 ng/ml) represent mean LOTUS concentration in healthy controls. Values are means \pm s.e.m. **c** Individual variation in CSF LOTUS concentration. Horizontal dashed lines (192.1 ng/ml) represent mean LOTUS concentration in healthy controls. P values were calculated by Wilcoxon matched-pairs signed-rank test

Because LOTUS endogenously functions as an NgR1 antagonist, preventing Nogo from binding to NgR1, it can promote axonal growth and is thus a promising therapeutic target for neural regeneration [6]. These molecules have attracted attention due to their relationship with MS pathogenesis and also as therapeutic targets. In particular, recent studies have demonstrated that the activation of NgR1-mediated signaling plays a substantial role in axonal degeneration in MS. Furthermore, a previous study by our group showed that LOTUS levels in the CSF correlated inversely with disease activity of MS, suggesting that its reduction during the active phase may be involved in the pathogenesis of MS. Interestingly, a recent study suggested that Nogo and NgR1, as well as other molecules, are related not only to axonal guidance but also to the

immune system. This observation demonstrated that lymphocytes express NgR1 and that Nogo–NgR1 signaling can alter lymphocyte phenotypes [17]. Accordingly, we hypothesized that LOTUS is also involved in the immune system and thus represents a candidate novel biomarker for disease activity associated with neuroinflammation. In this study, we elucidated the correlation between the CSF LOTUS level and neuroinflammation in meningitis, one of the most common neuroinflammatory diseases. Our results indicated that, similar to Nogo, LOTUS is involved in the immune system as well as playing a role in axonal guidance.

Currently, total protein concentration and cell counts in the CSF are widely used as biomarkers to evaluate disease activity, although they are not necessarily appropriate for monitoring neuroinflammation in most CNS



diseases. For evaluation of meningitis, these conventional markers are well established; therefore, we first confirmed improvement of meningitis in reference to these CSF biomarkers, in addition to the clinical neurological findings. Then, we chronologically compared the CSF LOTUS concentration between acute and recovery phases. We observed that the LOTUS level was reduced in acute phase, followed by recovery to the healthy control level, accompanied by clinical improvement. These data demonstrated that the CSF LOTUS concentration is inversely correlated with the disease activity of meningitis, as we previously reported in MS.

We then considered two possible mechanisms for a decrease in LOTUS: (1) downregulation of *Lotus* mRNA expression by neuroinflammation, and (2) enhanced degradation of LOTUS protein. Accordingly, we investigated whether *Lotus* mRNA levels decreased with the severity of neuroinflammation in LPS-injected mice, which is widely used as an animal model for neuroinflammation [23–25]. As expected, LPS-treated mice exhibited a significant downregulation of *Lotus* mRNA,

along with a reduced level of LOTUS protein, in the brain. These results indicated that the mechanism underlying reduced CSF LOTUS concentration is downregulation of *Lotus* mRNA expression in response to neuroinflammation.

Previous proteomic studies reported LOTUS in CSF from normal subjects [26], and another group reported that LOTUS was not detectable in the CSF of patients with viral meningitis and CNS sarcoidosis [27]. Indeed, we confirmed a reduction in CSF concentration of LOTUS in acute phase of three patients with CNS sarcoidosis (61.6 ± 21.1 ng/ml). Thus, these findings support our results that reduced LOTUS concentration is associated with neuroinflammation in the acute phase of the diseases.

Interleukin-6 (IL-6) concentration in the CSF is often used as a biomarker of neuroinflammation in bacterial meningitis and progressive neuro-Behçet's disease [28–31]. However, the sensitivity of IL-6 is too low to estimate disease activity in non-progressive neuro-Behçet's disease or MS [32–34], indicating that IL-6 is of limited utility as a biomarker of neuroinflammation.

In contrast to IL-6, LOTUS may reflect disease activity of wider range of CNS diseases such as viral and bacterial meningitis, CNS sarcoidosis, and MS. Thus, the CSF LOTUS level is potentially of great diagnostic value for estimating neuroinflammation in a variety of CNS diseases, and we hypothesize that LOTUS might be critical to the pathogenesis of these diseases.

Conclusion

CSF levels of LOTUS correlated inversely with disease activity both in bacterial and viral meningitis, as in multiple sclerosis. Therefore, variation of CSF LOTUS is associated with neuroinflammation and is useful as a biomarker, leading accurate diagnosis, appropriate therapy, and comprehension of pathogenesis for a broader range of neuroinflammatory diseases.

Abbreviations

CNS: Central nervous system; IL-6: Interleukin-6; LOTUS: Lateral olfactory tract usher substance; LPS: Lipopolysaccharide; MAG: Myelin-associated glycoprotein; MS: Multiple sclerosis; NgR1: Nogo receptor 1; OMgp: Oligodendrocyte myelin glycoprotein; PBS: Phosphate-buffered saline; PCR: Polymerase chain reaction; RT: Reverse transcription

Acknowledgements

Not applicable.

Funding

This study was supported by Grants-in-Aid from the Ministry of Education, Culture, Sport, Science and Technology of Japan (to K. Takahashi, K. Takei, F. Tanaka); Grants-in Aid from the Ministry of Health, Labour and Welfare of Japan (to F. Tanaka); a grant for Research and Development Project III of Yokohama City University, Japan (to K. Takei); and grants for medical research from Takeda Science Foundation for Visionary Research and Astellas Foundation for Research on Metabolic Disorders (to K. Takei).

Availability of data and materials

All data generated or analyzed during this study are included in this published article.

Authors' contributions

KoT and FT designed the study. KeT, HD, SK, KoT, HT, and FT supervised the experiments, data analysis, and interpretation. KeT, MK, KT, HN, RF, AK, MT, YH, HJ, and FT managed and provided samples and evaluated clinical data. KeT analyzed the LOTUS concentration in CSF. YK collaborated on the detection of LOTUS. KeT wrote the paper. HT, FT, and KoT edited the manuscript. KoT and FT had full access to all data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. All authors read and approved the final manuscript.

Ethics approval and consent to participate

This study was approved by the ethics committee of the Yokohama City University Hospital (approval number: B121101045). Informed consent was obtained from each participant according to the research ethics of our university hospital.

Consent for publication

Not applicable.

Competing interests

Research funds were obtained from the Cosmic Corporation. The sponsors had no role in the design and conduct of the study; collection, management, analysis, or interpretation of the data; preparation, review, or approval of the manuscript; and decision to submit the manuscript for publication. No other conflict of interest exists.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 12 July 2017 Accepted: 1 February 2018

Published online: 17 February 2018

References

- Singhal G, Jaehne EJ, Corrigan F, Toben C, Baune BT. Inflammasomes in neuroinflammation and changes in brain function: a focused review. *Front Neurosci.* 2014;8:315.
- Bielekova B, Komori M, Xu Q, Reich DS, Wu T. Cerebrospinal fluid IL-12p40, CXCL13 and IL-8 as a combinatorial biomarker of active intrathecal inflammation. *PLoS One.* 2012;7:e48370.
- Polman CH, Reingold SC, Banwell B, Clanet M, Cohen JA, Filippi M, Fujihara K, Havrdova E, Hutchinson M, Kappos L, et al. Diagnostic criteria for multiple sclerosis: 2010 revisions to the McDonald criteria. *Ann Neurol.* 2011;69:292–302.
- Rotstein DL, Healy BC, Malik MT, Chitnis T, Weiner HL. Evaluation of no evidence of disease activity in a 7-year longitudinal multiple sclerosis cohort. *Jama Neurology.* 2015;72:152–8.
- Jacobs AH, Tavitian B, Consortium IN. Noninvasive molecular imaging of neuroinflammation. *J Cereb Blood Flow Metab.* 2012;32:1393–415.
- Sato Y, Iketani M, Kurihara Y, Yamaguchi M, Yamashita N, Nakamura F, Arie Y, Kawasaki T, Hirata T, Abe T, et al. Cartilage acidic protein-1B (LOTUS), an endogenous Nogo receptor antagonist for axon tract formation. *Science.* 2011;333:769–73.
- Kurihara Y, Takei K. LOTUS, a potent blocker of Nogo receptor-1 causing inhibition of axonal growth. *Neural Regen Res.* 2015;10:46–8.
- Kurihara Y, Iketani M, Ito H, Nishiyama K, Sakakibara Y, Goshima Y, Takei K. LOTUS suppresses axon growth inhibition by blocking interaction between Nogo receptor-1 and all four types of its ligand. *Mol Cell Neurosci.* 2014;61:211–8.
- Takahashi K, Kurihara Y, Suzuki Y, Goshima Y, Tanaka F, Takei K. Association of cerebrospinal fluid levels of lateral olfactory tract usher substance (LOTUS) with disease activity in multiple sclerosis. *JAMA Neurol.* 2015;72:176–9.
- Takahashi K, Tanaka F, Takei K. LOTUS, a possible endogenous inhibitor of axonal degeneration, as a new biomarker for multiple sclerosis. *Neurodegener Dis Manag.* 2015;5:469–72.
- Takahashi K, Tanaka F, Takei K. Axonal regeneration-related molecules as biomarkers for multiple sclerosis. *Brain Nerve.* 2016;68:82–9.
- Frischer JM, Bramow S, Dal-Bianco A, Lucchinetti CF, Rauschka H, Schmidbauer M, Laursen H, Sorensen PS, Lassmann H. The relation between inflammation and neurodegeneration in multiple sclerosis brains. *Brain.* 2009;132:1175–89.
- Yamashita T, Mueller BK, Hata K. Neogenin and repulsive guidance molecule signaling in the central nervous system. *Curr Opin Neurobiol.* 2007;17:29–34.
- Muramatsu R, Kubo T, Mori M, Nakamura Y, Fujita Y, Akutsu T, Okuno T, Taniguchi J, Kumanogoh A, Yoshida M, et al. RGMa modulates T cell responses and is involved in autoimmune encephalomyelitis. *Nat Med.* 2011;17:488–94.
- Yu HH, Kolodkin AL. Semaphorin signaling: a little less per-plexin. *Neuron.* 1999;22:11–4.
- Suzuki K, Kumanogoh A, Kikutani H. Semaphorins and their receptors in immune cell interactions. *Nat Immunol.* 2008;9:17–23.
- Pool M, Niino M, Rambaldi I, Robson K, Bar-Or A, Fournier AE. Myelin regulates immune cell adhesion and motility. *Exp Neurol.* 2009;217:371–7.
- Petratos S, Ozturk E, Azari MF, Kenny R, Lee JY, Magee KA, Harvey AR, McDonald C, Taghian K, Moussa L, et al. Limiting multiple sclerosis related axonopathy by blocking Nogo receptor and CRMP-2 phosphorylation. *Brain.* 2012;135:1794–818.
- Tunkel AR, Hartman BJ, Kaplan SL, Kaufman BA, Roos KL, Scheld WM, Whitley RJ. Practice guidelines for the management of bacterial meningitis. *Clin Infect Dis.* 2004;39:1267–84.
- Thwaites GE, Hien TT. Tuberculous meningitis: many questions, too few answers. *Lancet Neurol.* 2005;4:160–70.
- Chen MS, Huber AB, van der Haar ME, Frank M, Schnell L, Spillmann AA, Christ F, Schwab ME. Nogo-A is a myelin-associated neurite outgrowth inhibitor and an antigen for monoclonal antibody IN-1. *Nature.* 2000;403:434–9.
- Fournier AE, GrandPre T, Strittmatter SM. Identification of a receptor mediating Nogo-66 inhibition of axonal regeneration. *Nature.* 2001;409:341–6.
- Grabner GF, Eichmann TO, Wagner B, Gao Y, Farzi A, Taschler U, Radner FP, Schweiger M, Lass A, Holzer P, et al. Deletion of monoglyceride lipase in astrocytes attenuates lipopolysaccharide-induced neuroinflammation. *J Biol Chem.* 2016;291:913–23.
- Behairi N, Belkhef M, Rafa H, Labsi M, Deghbar N, Bouzid N, Mesbah-Amroun H, Touil-Boukoffa C. All-trans retinoic acid (ATRA) prevents lipopolysaccharide-induced neuroinflammation, amyloidogenesis and memory impairment in aged rats. *J Neuroimmunol.* 2016;300:21–9.
- Qin L, Wu X, Block ML, Liu Y, Breese GR, Hong JS, Knapp DJ, Crews FT. Systemic LPS causes chronic neuroinflammation and progressive neurodegeneration. *Glia.* 2007;55:453–62.
- Zhang J, Goodlett DR, Peskind ER, Quinn JF, Zhou Y, Wang Q, Pan C, Yi E, Eng J, Aebersold RH, Montine TJ. Quantitative proteomic analysis of age-related changes in human cerebrospinal fluid. *Neurobiol Aging.* 2005;26:207–27.
- Hammack BN, Fung KY, Hunsucker SW, Duncan MW, Burgoon MP, Owens GP, Gilden DH. Proteomic analysis of multiple sclerosis cerebrospinal fluid. *Mult Scler.* 2004;10:245–60.
- Akman-Demir G, Tuzun E, Icoz S, Yesilot N, Yentur SP, Kurtuncu M, Mutlu M, Saruhan-Direskeneli G. Interleukin-6 in neuro-Behcet's disease: association with disease subsets and long-term outcome. *Cytokine.* 2008;44:373–6.
- Hirohata S, Isshi K, Oguchi H, Ohse T, Haraoka H, Takeuchi A, Hashimoto T. Cerebrospinal fluid interleukin-6 in progressive neuro-Behcet's syndrome. *Clin Immunol Immunopathol.* 1997;82:12–7.
- Matsuzono Y, Narita M, Akutsu Y, Togashi T. Interleukin-6 in cerebrospinal fluid of patients with central nervous system infections. *Acta Paediatr.* 1995;84:879–83.
- Houssiau FA, Bukasa K, Sindic CJ, Van Damme J, Van Snick J. Elevated levels of the 26K human hybridoma growth factor (interleukin 6) in cerebrospinal fluid of patients with acute infection of the central nervous system. *Clin Exp Immunol.* 1988;71:320–3.
- Uzawa A, Mori M, Ito M, Uchida T, Hayakawa S, Masuda S, Kuwabara S. Markedly increased CSF interleukin-6 levels in neuromyelitis optica, but not in multiple sclerosis. *J Neurol.* 2009;256:2082–4.
- Miljkovic D, Drulovic J, Trajkovic V, Mesaros S, Dujmovic I, Maksimovic D, Samardzic T, Stojsavljevic N, Levic Z, Mostarica SM. Nitric oxide metabolites and interleukin-6 in cerebrospinal fluid from multiple sclerosis patients. *Eur J Neurol.* 2002;9:413–8.
- Ireland SJ, Monson NL, Davis LS. Seeking balance: potentiation and inhibition of multiple sclerosis autoimmune responses by IL-6 and IL-10. *Cytokine.* 2015;73:236–44.