

Evaluation of the Steelex M600H coagulometer prothrombin time-international normalized ratio assay with Steelex test reagents

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Abstract

Introduction: The aim of the present study was to validate prothrombin time (PT) international normalized ratio (INR) results obtained using Steelex test reagents and a Steelex coagulometer (Steelex Scientific Instrument Company, Beijing, China), in comparison with use of a well-established standard test employing Pacific Hemostasis reagents (Fisher Diagnostics, Middletown, VA, USA) and Teco Coatron A4 coagulometer (Teco Medical Instruments GmbH, Neufahrn, Germany).

Materials and methods: Between- and within-day coefficients of variation (CVs) of both assays were calculated using control samples provided by the test manufacturers. Samples from 90 subjects were collected and INR values were determined in a double-blind parallel manner employing both systems.

Results: The within-day coefficients of variation (CVs) in INR estimates ranged from 2.6% (INR = 1.12) to 3.1% (INR = 2.51) for the Steelex system and from 2.1% (INR = 1.09) to 1.8% (INR = 2.8) for the Pacific test; the between-day values ran from 3.4% (INR = 1.16) to 7.9% (INR = 2.64) and from 3.3% (INR = 1.1) to 2.3% (INR = 2.7), respectively. Passing-Bablok fit of the of the Steelex and Pacific methods yielded the equation: Steelex INR = 0.85 (0.79-0.91) × Pacific INR + 0.12 (-0.02-0.21), whereas the CUSUM linearity P value was < 0.01. The mean bias as determined by the Bland-Altman test was -0.156 (-0.912-0.600).

Conclusion: The results obtained using Steelex reagents and the M600H coagulometer are not equivalent to those obtained using Pacific Hemostasis reagents and a Teco Coatron A4 coagulometer, at least in the therapeutic range.

Key words: Steelex coagulometer; prothrombin time, INR, evaluation

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Introduction

Prothrombin time (PT) is the most common coagulation test performed in the clinical laboratory. PT is used to detect abnormalities in the extrinsic coagulation pathway. PT measurement is critical when thrombotic patients are on oral anticoagulant therapy. However, it can be difficult to compare PT test results from different laboratories (1); standardization is essential. Thromboplastin, the principal reagent of the PT assay, is commercially available in different forms. To achieve the required standardization, all thromboplastin preparations should be calibrated using the International

al Sensitivity Index (ISI) and PT results should be presented in International Normalized Ratio (INR) units (2). The WHO Expert Committee on Biological Standardization has decreed that standardization requires a comparison of results yielded by the test thromboplastin with those afforded when a reference thromboplastin is employed (2,3). However, it is clear that standardization is not universally performed (4-7).

In Turkey, public procurement law requires that the lowest bid be accepted when equipment or reagents are ordered. Thus, the choice of a coagu-

lometer is price-dependent, even though the accuracy and reliability of INR measurement is essential. The Steelex M600H (Steelex Scientific Instrument Company, Beijing, China), approved for use in our laboratory because the device is inexpensive, is a new coagulometer that measures PT and INR.

The aim of the present study was to validate INR results obtained using the Steelex M600H coagulometer and Steelex reagents (the PT kit) by comparing such data with the results yielded by a Teco Coatron A4 coagulometer (Teco Medical Instruments GmbH, Neufahrn, Germany) employing a Pacific Hemostasis test reagent (Thromboplastin-DS, Fisher Diagnostics, VA, USA). The latter test is standard in the clinical laboratory.

Materials and methods

The present study was conducted in the period from June to July 2011 in the clinical laboratory of the Şevket Yılmaz Research and Education Hospital. The study was approved by our institutional Ethics Committee and all participants gave written informed consent. All procedures were in accordance with the Second Declaration of Helsinki.

PT and INR determination methods

As a reference method, plasma PT was analyzed in an automated manner using a Teco Coatron A4 coagulometer. The PT of citrated plasma was determined photometrically employing a high-resolution four-channel optic system (400 nm) and a suspension of rabbit brain thromboplastin (Thromboplastin-DS) prepared in calcium chloride solution. The instrument was calibrated employing calibration reference plasmas with manufacturer-assigned INR values (TECal N calibration plasma cat. no. P800-010, Teco GmbH, Germany) (8). The thromboplastin ISI value assigned by the manufacturer was 1.03. This is the method by which PT is routinely measured, in INR, in our hospital.

Steelex M600H

PT coagulation time was measured employing a new fully automated Steelex M600H coagulometer and Steelex test reagents, and a freeze-dried extract of rabbit brain thromboplastin (product

no. SS00110001; lot no. STG20101-LG4) prepared in a buffer containing calcium chloride. The ISI assigned to this preparation by the manufacturer was 1.18. In the Steelex system, light passing through a medium in which fibrinogen has been converted to fibrin is absorbed by the fibrin stands. The instrument identifies a coagulation endpoint by measuring the change in optical density at 470 nm, and this information is processed, using in-built software, to determine a clot point. The instrument was calibrated employing calibration reference plasmas with manufacturer-assigned INR values (Unicalibrator cat. no. 00675 Diagnostica Stago, Inc., France) (8). The INR is calculated and automatically displayed by the instruments from the prothrombin time using the formula (3):

$$\text{INR} = (\text{PT}_{\text{sample}} / \text{PT}_{\text{normal}})^{\text{ISI}}$$

The coefficients of variation of the regression line slopes of calibration curves of both instruments were less than 3%, in accordance with the WHO requirements (9). All INRs were calculated using a local prothrombin time (PT); this was the mean normal prothrombin time (MNPT) obtained using 20 fresh normal samples from healthy individuals (10).

Analytical precision of PT and INR determinations

Before specimen analysis, we performed two-level quality-control (QC) tests using materials supplied by either manufacturer (Steelex ANCP, catalog no. SS005711001 and NCP, catalog no. SS000500007; Pacific Hemostasis INR Control Plasma catalog nos. 100595 and 100596). Between-day differences in QC data were determined via analysis of duplicates on each of 20 successive days. Within-day precision was calculated by conducting 20 replicate analyses of the QC materials in a single run. Between-day precision is a measure of random analytical error (11).

Patients and blood sampling

Venous blood samples were obtained from 90 patients for whom PT (INR) tests were scheduled as part of routine preoperative check-up (N = 14) or for monitoring of oral anticoagulant therapy (OAT) (N = 76). Patients with atrial fibrillation and/or a prosthet-

ic heart valve, and those prescribed long-term OAT were accepted to the study. Of all patients, 35 were in their first 3 months of treatment (the dose-adjustment phase) whereas 41 were in a later treatment stage (the maintenance phase). Of all patients, the INR values of 48 were 1-1.09 units, those of 27 2-2.7 units, and those of 15 3-4.9 units. Hence, all possible anticoagulation phases (pretreatment, dose-adjustment, and steady-state) were included.

Blood samples (each 1.8 mL) were collected into citrate coagulation tubes (Vacuette; Greiner Labortechnik GmbH, Krems- münster, Austria; catalog no. 454322 9NC) each containing 0.2 mL 3.2% (w/v) sodium citrate (0.109 M). Both the WHO and the NCCLS state that 3.2% (w/v) citrate is the appropriate anticoagulant level when coagulation is to be tested (2,12). Plasma was obtained via centrifugation for 15 min at 1,500 x g. All samples were assayed using the two systems within 1 hour of blood collection.

Statistical analysis

Data were evaluated using SPSS version 13.0 (SPSS Inc., Chicago, IL, USA) and Analyse-It version 2.04 (Analyse-It Software, Leeds, UK). Means, standard deviations, and coefficients of variation were calculated. (13-15).

Results

The coefficients of variation (CVs) of within- and between-day QC data are presented in Table 1.

Passing and Bablok regression analysis yielded an intercept of 0.12 units (95% confidence interval (CI)

= -0.02-0.21) and a slope of 0.85 (95% CI = 0.79-0.91), suggesting that, a proportional difference existed between the two methods (Figure 1) (14). The cumulative sum linearity test (CUSUM test) revealed significant deviation from linearity (P < 0.01) (14).

The Bland–Altman difference plot showed that the mean bias was -0.156 units and the 95% limits of agreement were - 0.912 to 0.600 when the Steelex and Pacific test data were compared (Figure 2). The bias was lower in the sub-therapeutic range (INR < 2.0) and higher in the therapeutic range (INR = 2.0-4.5) (Figure 1).

Discussion

Extensive international collaboration has allowed laboratory INR measurement to become standardized worldwide; this is an essential feature of quality control (16,17). To ensure test result accuracy, clinical laboratories are required to conduct internal quality control tests and to participate in external quality assurance (EQA) networks. Our laboratory is a member of the national external quality assessment scheme (KBUDEK, Istanbul, Turkey) that evaluates coagulation test data (PT/INR; and assays of APTT and fibrinogen). We routinely employ Pacific Hemostasis test reagents. At the time of the present study, the average values of the Standard Deviation Index (SDI; Mean minus Peer Group Mean/SD of Peer Group Mean) obtained from 134 laboratories using Pacific test reagents were 0% for an INR of 1.04 and minus 0.15% for an INR of 1.95. The SDI is a parameter for evaluating

TABLE 1: Within-day imprecision data and between-day imprecision data for Steelex test reagent on Steelex M600H coagulometer and Pacific Hemostasis test reagents on Teco Coatron A4 coagulometer.

		Steelex test reagents				Pacific Hemostasis test reagents			
		Within-Day Mean (SD) (N = 20)	CV (%)	Between-Day Mean (SD) (N = 20)	CV (%)	Within-Day Mean (SD) (N = 20)	CV (%)	Between-Day Mean (SD) (N = 20)	CV (%)
QC1*	PT(sec)	13.8 (0.36)	2.6	14.2 (0.49)	3.4	15.1 (0.32)	2.1	15.2 (0.49)	3.2
	INR	1.12 (0.03)	2.6	1.16 (0.04)	3.4	1.09 (0.02)	2.1	1.1 (0.03)	3.3
QC2**	PT(sec)	27.2 (0.71)	2.6	28.8 (2.19)	7.6	37.6 (0.68)	1.8	37.3 (0.86)	2.3
	INR	2.51 (0.08)	3.1	2.64 (0.21)	7.9	2.8 (0.05)	1.8	2.7 (0.07)	2.5

*QC1, **QC2 = quality control level 1 and level 2

bias relative to the consensus group. It provides a good measure of accuracy because each laboratory's results are compared to the mean value for all laboratories, which is assumed to be the true value.

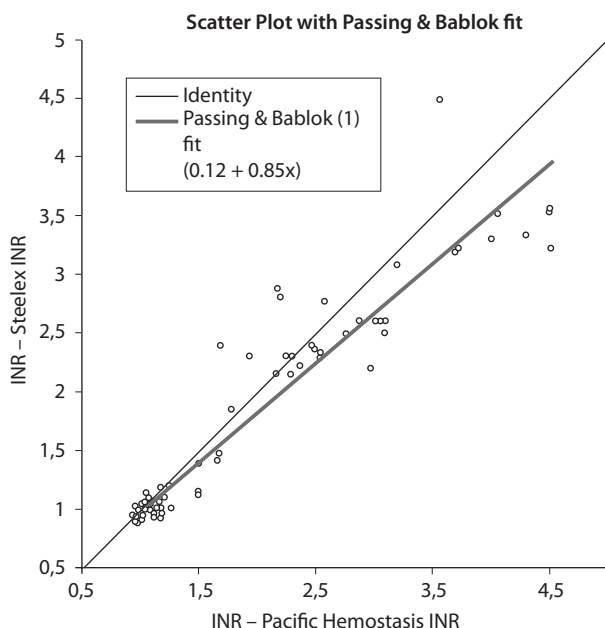


FIGURE 1. Passing-Bablok regression analysis for the Steelex INR results vs. laboratory (Pacific) INR results. The solid line indicates the regression line.

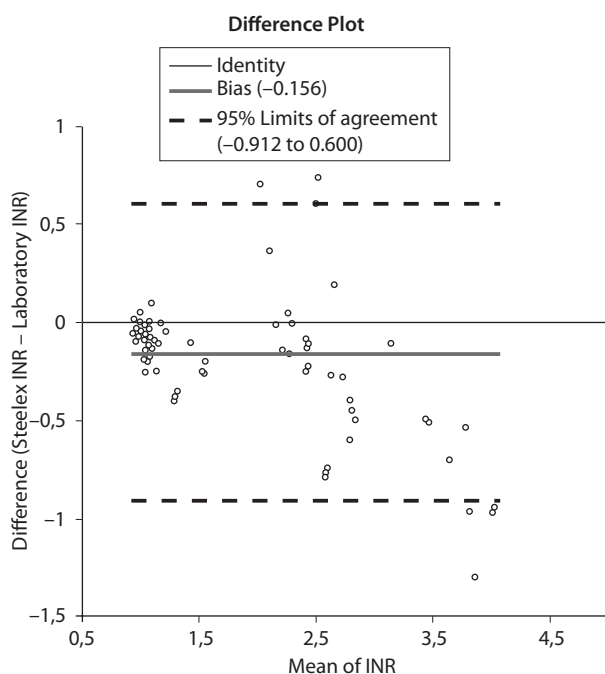


FIGURE 2. Bland-Altman plot analysis between compared methods.

We consider that the Pacific system is a suitable standard for use in validation studies as this method yielded acceptable quality control data during our study period and employs highly sensitive human thromboplastin (ISI = 1.03), as recommended by the College of American Pathologists (12).

Within-run imprecision was satisfactory for both systems (CV ≤ 3.1%). The Steelex system had a higher between-run CV than did the Pacific method, explained by the fact that use of reagents of higher ISI is associated with greater interlaboratory INR CVs (18). The level of between-run imprecision using the Steelex system (Level 2; CV = 7.9%) was unacceptable; the value should be less than 5% when an INR assay is performed (19).

Monitoring of PT via calculation of INRs is important for patients on oral anticoagulation therapy; appropriate doses can thus be prescribed. Accurate INR estimation is crucial (20). Global harmonization of INR results, and agreement on the therapeutic ranges associated with varying clinical indications, is important (21). Many factors influence INR values; these include the levels of various coagulation factors present and their interaction with thromboplastin reagents differing in composition and sensitivity to the actions of such factors (22). It can be difficult to obtain accurate and precise INR measurements. Both PT and INR data should be identical regardless of the reagents, instrument, or test method used. However, agreement among results yielded by many commercial INR methods is poor. Such variations may compromise patient care (1,7).

In our present work; Passing and Bablok regression analysis yielded the appropriate y-intercept value of zero but the slope of the regression line did not approximate unity, suggesting that, in addition to the presence of a small constant bias, a proportional difference existed between the two methods (14). Our results are in agreement with previous findings of poor agreement among data obtained using different methods to calculate PT and INR (7,23). The variation noted in the present work may be attributable to differences between any or all of methodology, the coagulometers used, and thromboplastin source. Many recent reports have shown that coagulometer choice has a marked

and unpredictable effect on INR value (24). The precision of PT testing tends to be greater when ISI is lower (24) but end-point reliability is also affected by reagent quality (24).

Quality control is essential; it is recommended that analytical imprecision be less than or equal to half of intra-individual biological variation (25-27). Ricos *et al.* (28) found intra-individual INR variation of 4% and inter-individual variation of 6.8%. Kjeldsen (29) recorded in-treatment intra-individual INR variation of 10.1%. Via computer simulation of serial INR measurements within the generally accepted therapeutic range (INR 2.0-3.0), the cited author concluded that analytical imprecision should be less than 5% and analytical bias less than ± 0.2 INR units (29). In the present study, the overall analytical bias was indeed less than ± 0.2 INR units, but, in the therapeutic range, the bias was greater than ± 0.2 INR units, thus significantly higher than current recommendations (28,29). It is evident that ISI strongly influences INR inter-laboratory variability (30). Although the ISI assigned by the manufacturer of the Steelex M600H coagulometer was relatively low (1.18), a major limitation of our study is that local ISI calibration using WHO-certified plasma was not performed. Incorrect assignment of ISI values by manufacturers may cause among-method differences in test results.

References

1. Jackson CM, Esnouf MP. Has the time arrived to replace the quick prothrombin time test for monitoring oral anticoagulant therapy? *Clin Chem* 2005;51:483-5.
2. WHO Expert Committee on Biological Standardisation. Thirty-third Report. Technical Report Series 687. Vol. 81. Geneva: WHO; 1983.
3. International Committee for Standardisation in Haematology. International Committee On Thrombosis and Haemostasis. ICISH/ICTH recommendations for reporting prothrombin time in oral anticoagulant control. *Thromb Haemost* 1985;53:155.
4. Cunningham MT, Johnson GF, Pennell BJ, Olson JD. The reliability of manufacturer-determined, instrument-specific international sensitivity index values for calculating the international normalized ratio. *Am J Clin Pathol* 1994;102:128-33.
5. van Rijn JL, Schmidt NA, Rutten WP. Correction of instrument and reagent based differences in determination of the international normalized ratio (INR) for monitoring anticoagulant therapy. *Clin Chem* 1989;35:840-3.
6. Ng VL, Levin J, Corash L, Gottfried EL. Failure of the International Normalized Ratio to generate consistent results within a local medical community. *Am J Clin Pathol* 1993;99:689-94.
7. Horsti J, Uppa H, Vilpo J. Poor agreement between different prothrombin time International Normalized Ratio (INR) methods: comparison of seven commercial reagents. *Clin Chem* 2005;51:553-60.
8. Houbouyan LI, Goguel AF. Long-term French experience in INR standardization by a procedure using plasma calibrants. *Am J Clin Pathol* 1997;108:83-9.
9. WHO Expert Committee on Biological Standardization. Annex 3: guidelines for thromboplastins and plasma used to control oral anticoagulant therapy. In: World Health Organization Technical Report Series. 48th report. No. 889. Geneva, Switzerland: World Health Organization; 1999:64-93.
10. Peters RH, van den Besselaar AM, Olthuis FM. Determination of the mean normal prothrombin time for assessment of international normalized ratios. Usefulness of lyophilized plasma. *Thromb Haemost* 1991;66:442-5.

Conclusion

In conclusion, reliable PT, INR test data are important for proper management of patients on OAT. Our results indicate that results obtained from Steelex M600H coagulometer using Steelex reagents kit are not interchangeable with Teco Coatron A4 coagulation coagulometer using Pacific Hemostasis reagent kit, in the therapeutic range.

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Potential conflict of interest

None declared.

11. Westgard JO, Carey RN, Wold S. Criteria for judging precision and accuracy in method development and evaluation. *Clin Chem* 1974;20:825-33.
12. Fairweather RB, Ansell J, van den Besselaar AM, Brandt JT, Bussey HI, Poller L, et al. College of American Pathologists Conference XXXI on laboratory monitoring of anticoagulant therapy: laboratory monitoring of oral anticoagulant therapy. *Arch Pathol Lab Med* 1998;122:768-81.
13. Bland JM, Altman DG. Statistical methods for assessing agreement between two methods of clinical measurement. *Lancet* 1986;i:307-10.
14. Bilic-Zulle L. Comparison of methods: Passing and Bablok regression. *Biochem Med* 2011;21:49-52.
15. Sonntag O. Quality in the analytical phase. *Biochem Med* 2010;20:147-53.
16. Van den Besselaar AM, Barrowcliffe TW, Houbouyan- Réveillard LL, Johnston M, Poller L, Tripodi A. and On behalf of the Subcommittee on Control of Anticoagulation of the Scientific and Standardization Committee of the ISTH. Guidelines on preparation, certification, and use of certified plasmas for ISI calibration and INR determination. *J Thromb Haemost* 2004;2:1946-53.
17. Olson JD, Brandt JT, Chandler WL, Van Cott EM, Cunningham MT, Hayes TE, et al. Laboratory reporting of the international normalized ratio: progress and problems. *Arch Pathol Lab Med* 2007;131:1641-7.
18. Poller L, Triplett DA, Hirsh J, Carroll J, Clarke K. The value of plasma calibrants in correcting coagulometer effects on international normalized ratios— an international multicenter study. *Am J Clin Pathol* 1995;103:358-65.
19. Arkin CF, Adcock DM, Day HJ, Carroll JJ, Joist JH, Lenahan JG, et al. One-stage prothrombin time (PT) Test and activated partial thromboplastin time (aPTT) test: Approved Guideline. *NCCLS*: 1996;12:1-26.
20. Christensen TD. Self-management of oral anticoagulation therapy—methodological and clinical aspects. *Dan Med Bull* 2011;58:B4284.
21. Horsti J. Comparison of Quick and Owren prothrombin time with regard to the harmonization of the International Normalized Ratio (INR) system. *Clin Chem Lab Med* 2002;40:399-403.
22. Favaloro EJ, Adcock DM. Standardization of the INR: how good is your laboratory's INR and can it be improved? *Semin Thromb Hemost* 2008;34:593-603.
23. Jackson CM, Esnouf MP, Lindahl TL. A critical evaluation of the prothrombin time for monitoring oral anticoagulant therapy. *Pathophysiol Haemost Thrombosis* 2003;33:43-51.
24. Poller L. International Normalized Ratios (INR): the first 20 years. *J Thromb Haemost* 2004;2:849-60.
25. Lassen JF, Kjeldsen J, Antonsen S, Petersen PH, Brandslund I. Interpretation of serial measurements of international normalized ratio for prothrombin times in monitoring oral anticoagulant therapy. *Clin Chem* 1995;41:1171-6.
26. Lassen JF, Brandslund I, Antonsen S. Determination of critical difference and probability of significant change in consecutive measurements of International Normalized Ratio (INR) from patients on oral anticoagulant therapy. *Clin Chem* 1995;41:444-7.
27. Ricos C, Perich C, Minchinella J, Alvarez V, Simon M, Biosca C, et al. Application of biological variation - a review. *Biochem Med* 2009;19:250-9.
28. Ricos C, Alvarez V, Cava F, García-Lario JV, Hernández A, Jiménez CV, et al. Current databases on biological variation: pros, cons and progress. *Scand J Clin Lab Invest* 1999;66:337-49.
29. Kjeldsen J, Lassen JF, Petersen PH, Brandslund I. Biological variation of International Normalised Ratio for prothrombin times, and consequences in monitoring oral anticoagulant therapy: computer simulation of serial measurements with goal-setting for analytical quality. *Clin Chem* 1997;43:2175-82.
30. Tabernar DA, Poller L, Thomson JM, Darby KV. Effect of international sensitivity index (ISI) of thromboplastins on precision of international normalised ratios (INR). *J Clin Pathol* 1989;42:92-6.

Evaluacija rezultata dobivenih koagulometrom M600H tvrtke Steelex pri određivanju protrombinskog vremena INR Steelexovim reagensima

Sažetak

Uvod: Cilj ovog istraživanja je validirati rezultate određivanja protrombinskog vremena (PV) međunarodnim normaliziranim omjerom (engl. *international normalized ratio*, INR) dobivene reagensima tvrtke Steelex na njihovom koagulometru (Steelex Scientific Instrument Company, Peking, Kina) u usporedbi s često primjenjivanim standardnim testom tvrtke Pacific Hemostasis (Fisher Diagnostics, Middletown, Virginia, SAD) na koagulometru proizvođača Teco Coatron A4 (Teco Medical Instruments GmbH, Neufahrn, Njemačka).

Materijali i metode: Za oba su testa izračunati koeficijenti varijacije (CV) iz dana u dan i u seriji primjenjujući kontrolne uzorke od proizvođača testa. Sakupljeni su uzorci od 90 ispitanika te su im određene INR vrijednosti dvostruko slijepim određivanjem primjenjujući oba sustava.

Rezultati: Koeficijenti varijacije u seriji (CV) kod određivanja INR varirali su kod Steelexovog sustava od 2,6% (INR = 1,12) do 3,1% (INR = 2,51) i kod testa tvrtke Pacific od 2,1% (INR = 1,09) do 1,8% (INR = 2,8); vrijednosti iz dana u dan su za Steelexov sustav sezale od 3,4% (INR = 1,16) to 7,9% (INR = 2,64) a za test tvrtke Pacific od 3,3% (INR = 1,1) do 2,3% (INR = 2,7). Passing Bablokovom regresijom Steelexove metode i metode proizvođača Pacific dobivena je jednadžba: $\text{Steelex INR} = 0,85 (0,79-0,91) \times \text{Pacific INR} + 0,12 (-0,02-0,21)$, gdje je P vrijednost CUSUM-ovog testa linearnosti bila $< 0,01$, a srednja sustavna pogreška određena Bland-Altmanovim testom iznosila je $-0,156$ jedinica ($-0,912-0,600$).

Zaključak: Rezultati dobiveni primjenom Steelexovih reagensa na koagulometru M600H nisu, što se terapijskog raspona tiče, ekvivalentni rezultatima dobivenim primjenom reagensa tvrtke Pacific Hemostasis na koagulometru Teco Coatron A4.

Cljučne riječi: Steelexov koagulometar; protrombinsko vrijeme; INR; evaluacija