

Supplemental information

**Identification of a Kupffer cell subset
capable of reverting the T cell dysfunction
induced by hepatocellular priming**

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Figure S1

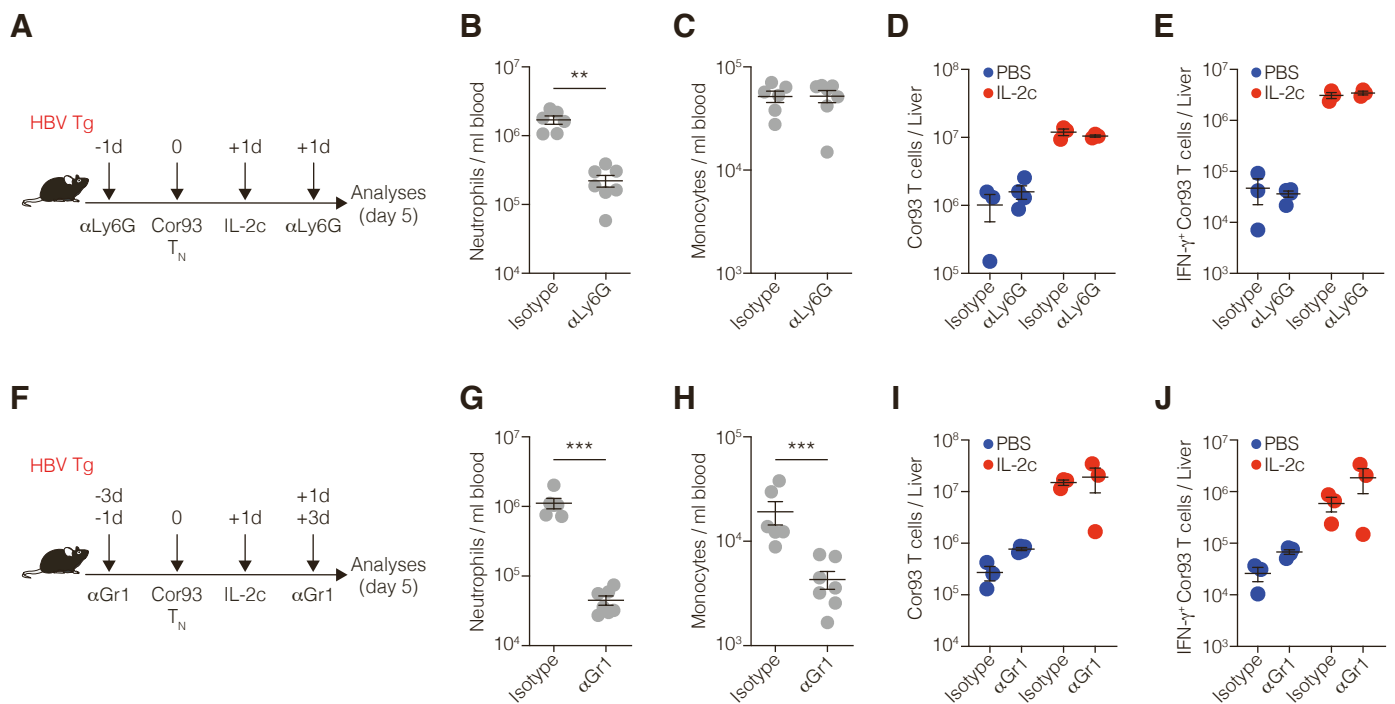


Figure S1. Neutrophils and monocytes are dispensable for T cell reinvigoration by IL-2. Related to Figure 1

(**A**) Schematic representation of the experimental setup. 1×10^6 Cor93 T_N were transferred into HBV transgenic (HBV Tg) recipients. Mice were injected with anti-Ly6G depleting antibody or the isotype control one day before and one day after T cell injection. Indicated mice received IL-2c one day after Cor93 T_N transfer. Livers were collected and analyzed five days after T cell transfer.

(**B-C**) Numbers of neutrophils (**B**) and monocytes (**C**) in the blood in the indicated mice at the time of Cor93 T_N injection (Isotype control $n=6$, anti-Ly6G $n=8$). ** p value < 0.01 , one-tailed Mann-Whitney U-test.

(**D-E**) Total numbers (**D**) and numbers of IFN- γ -producing (**E**) Cor93 T cells in the livers of the indicated mice (PBS: isotype control $n=3$, anti-Ly6G $n=4$; IL-2c: isotype control $n=3$, anti-Ly6G $n=3$).

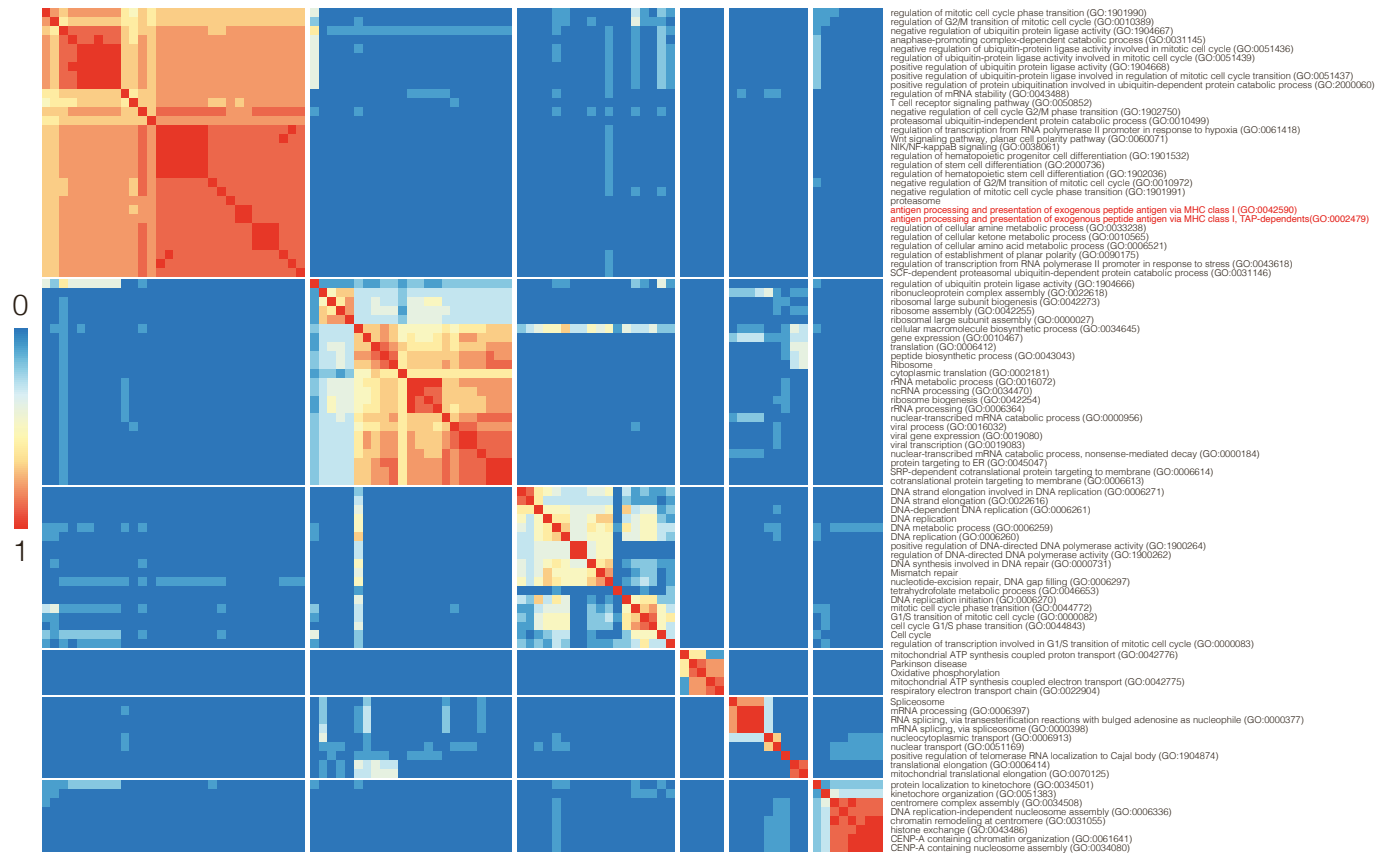
(**F**) Schematic representation of the experimental setup. 1×10^6 Cor93 T_N were transferred into HBV Tg recipients. Mice were injected with anti-Gr1 depleting antibody or isotype control every 48h starting 3 days before T cell injection. Indicated mice received IL-2c one day after Cor93 T_N cell transfer. Livers were collected and analyzed five days after T cell transfer.

(**G-H**) Numbers of neutrophils (**G**) and monocytes (**H**) in the blood of the indicated mice at the time of T cell injection (Isotype control $n=6$, anti-Gr1 $n=7$). *** p value < 0.001 , one-tailed Mann-Whitney U-test.

(**I-J**) Total numbers (**I**) and numbers of IFN- γ -producing (**J**) Cor93 T cells in the livers of the indicated mice (PBS: isotype control $n=3$, anti-Gr1 $n=4$; IL-2c: isotype control $n=3$ anti-Gr1, $n=3$). Data are representative of at least 3 independent experiments.

Figure S2

A



B

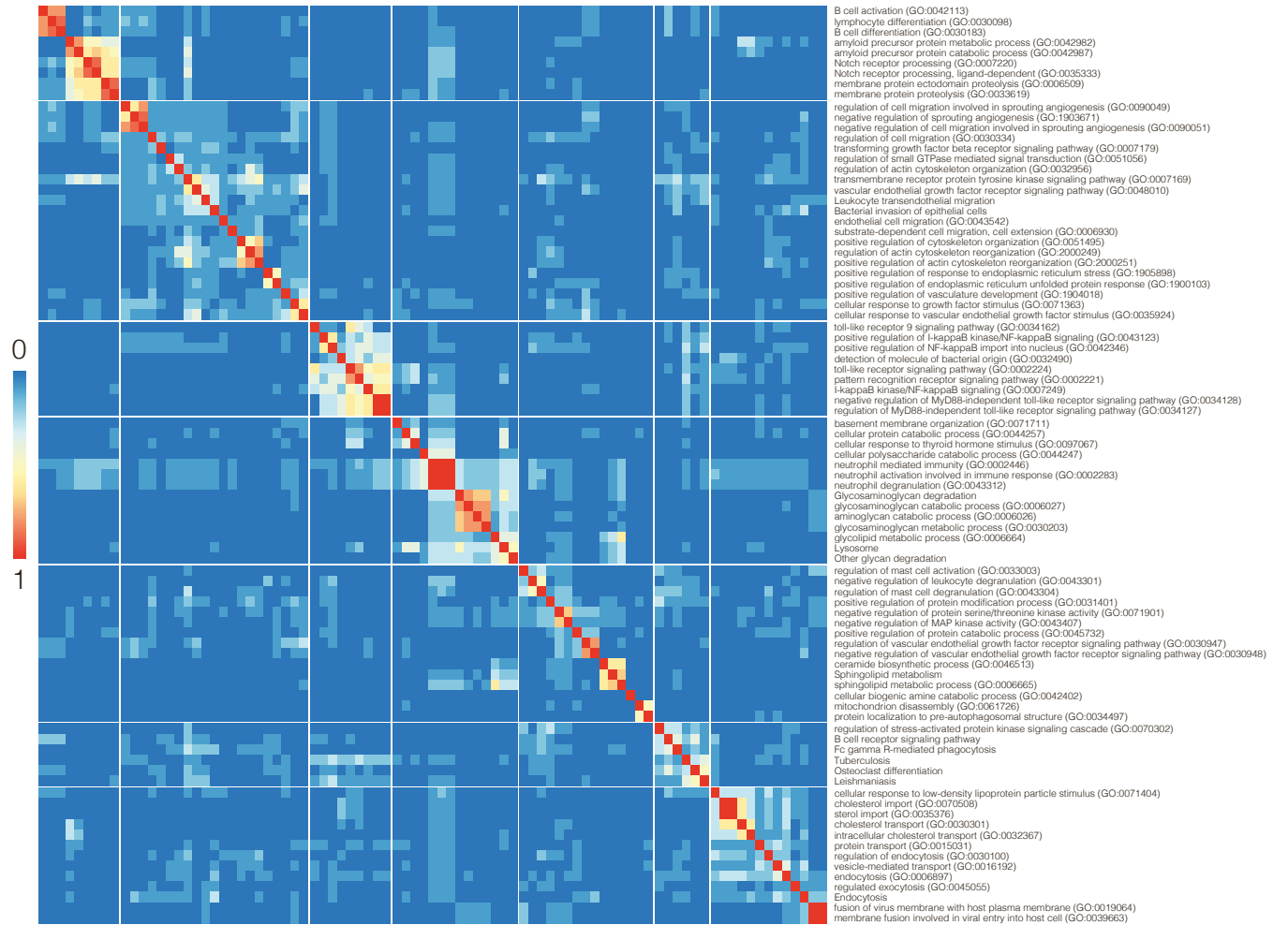


Figure S2. Regulated processes in KCs upon in vivo IL-2c treatment. Related to Figure 2

(A) Clustering of top significant (EnrichR Combined Score > 100, FDR < 0.05) Gene Ontology Biological Processes and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways of biological processes up-regulated in KCs upon in vivo IL-2c treatment. The thermal scale represents the Jaccard Similarity Coefficient between every gene set pair (blue representing a 0 Similarity Coefficient, red a 1 Similarity Coefficient).

(B) Clustering of top significant (EnrichR Combined Score > 30, FDR<0.05) Gene Ontology Biological Processes and KEGG pathways of biological processes down-regulated in KCs upon in vivo IL-2c treatment. The thermal scale represents the Jaccard Similarity Coefficient between every gene set pair (blue representing a 0 Similarity Coefficient, red a 1 Similarity Coefficient).

Figure S3

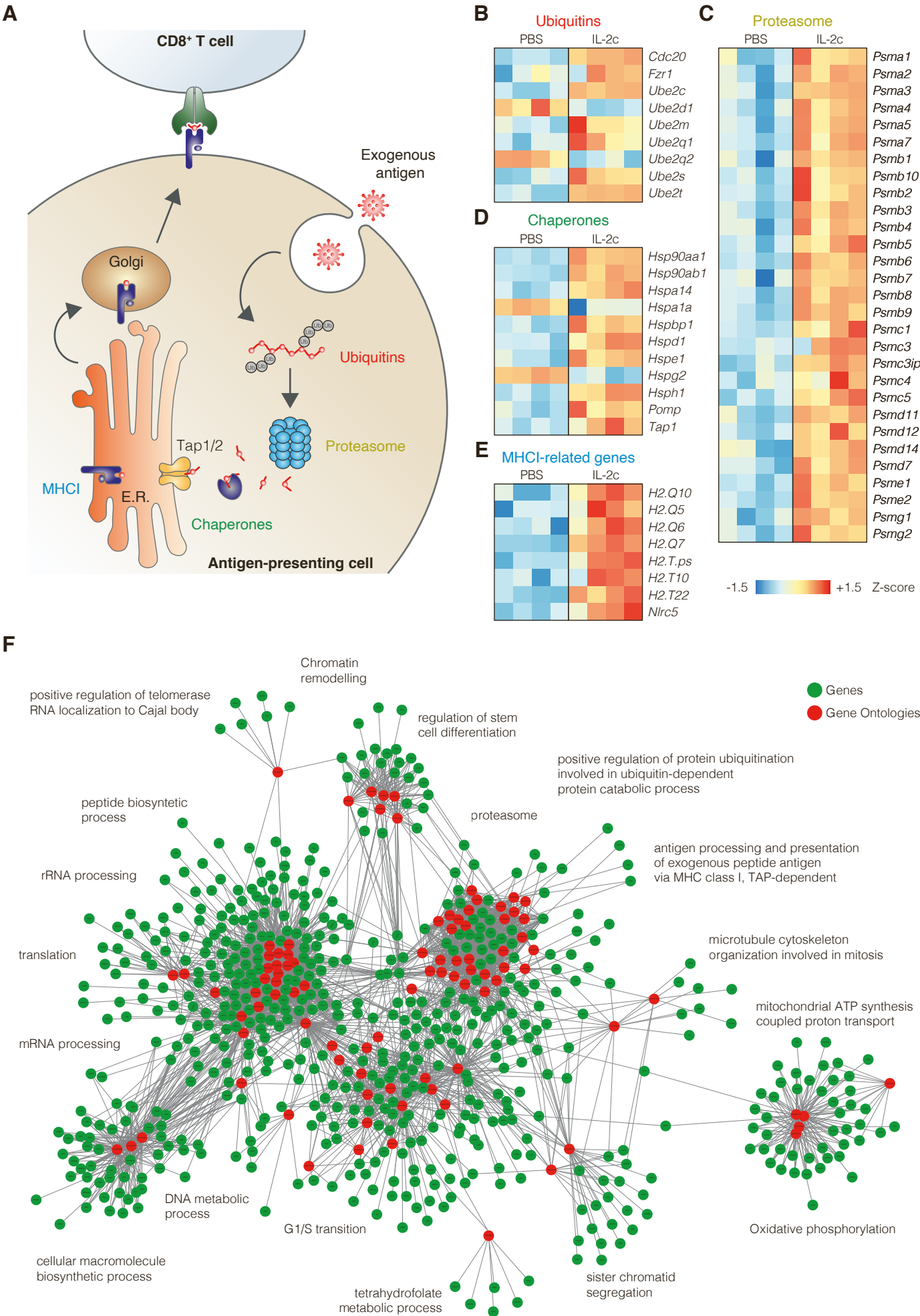


Figure S3. Genes associated to cross-presentation are upregulated in KCs upon *in vivo* IL-2c treatment. Related to Figure 2

(**A-E**) Schematic representation (**A**) and expression heatmap (**B-E**) of selected genes belonging to biological processes implicated in antigen cross-presentation, upregulated in KCs after IL-2c treatment. Values are in Z-score, calculated from scaling by row the Log2(TPM) values.

(**F**) Cytoscape network of top significant (EnrichR Combined Score > 100, FDR < 0.05) Gene Ontology Biological Processes and KEGG pathways of up-regulated processes. Red dots indicate enriched terms, green dots indicate the relative enriched genes.

Figure S4

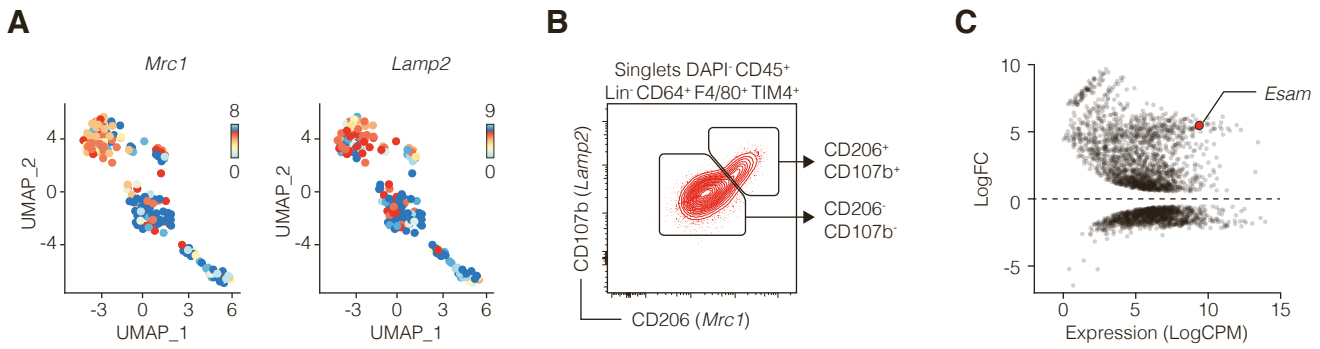


Figure S4. *Ad interim* bulk RNA sequencing on sorted KC subpopulations for the identification of suitable markers. Related to Figure 3

(A) Feature plot representation of the normalized expression level of *Mrc1* (CD206) and *Lamp2* (CD107b) on the scRNAseq dataset described in Fig. 3. The expression is measured as the $\ln(\text{TPM}+1)$.

(B) Sorting strategy for ad interim bulk RNA sequencing.

(C) MA plot showing differentially expressed genes (DEGs, p value < 0.001) between CD206⁻ CD107b⁻ KCs and CD206⁺ CD107b⁺ KCs. Each dot denotes a differentially expressed gene. The y-axis represents the logarithmic Fold Change (FC) between CD206⁻ CD107b⁻ KCs and CD206⁺ CD107b⁺ KCs, while the x-axis shows the expression level of each DEG in logarithmic Counts Per Million (CPM). *Esam* is highlighted as one of the most over-expressed gene in CD206⁺ CD107b⁺ KCs compared to CD206⁻ CD107b⁻ KCs.

Figure S5

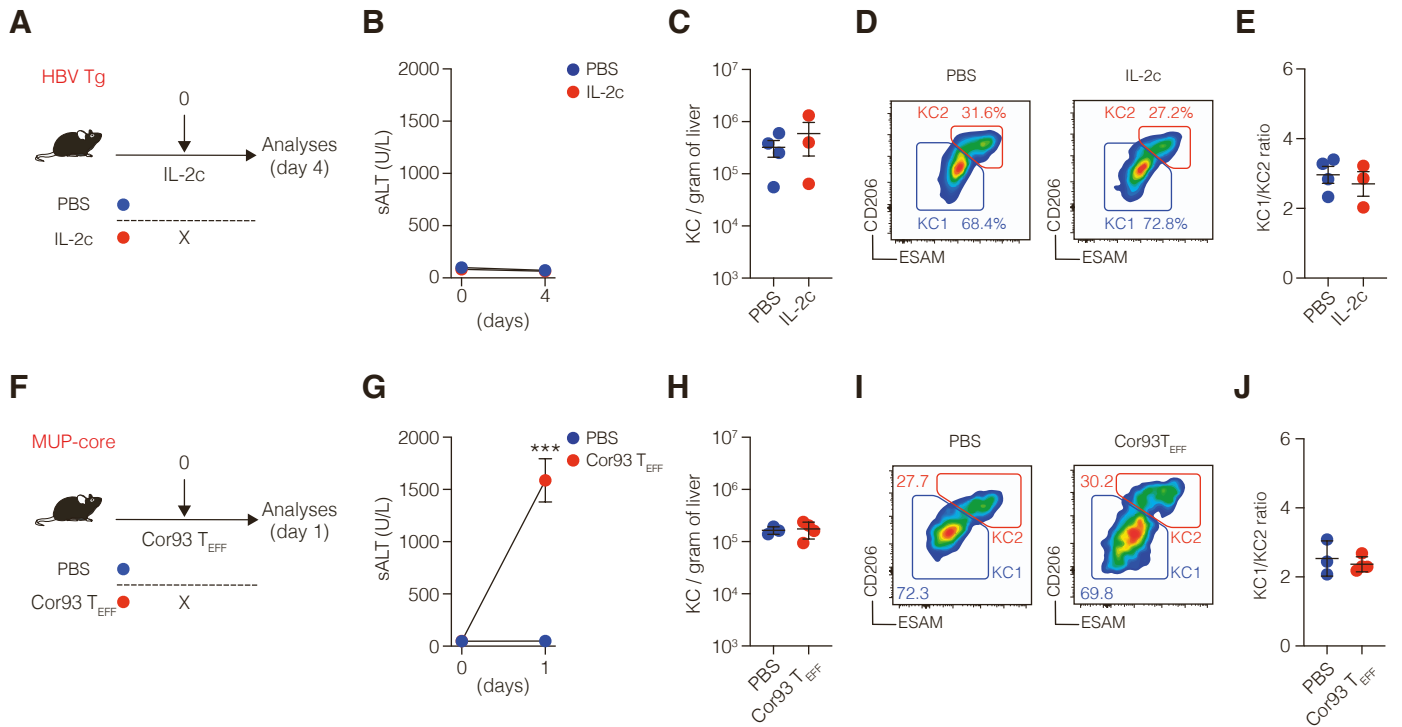


Figure S5. IL-2c treatment alone or liver inflammation have no impact on KC1/KC2 ratio. Related to Figure 4

(A) Schematic representation of the experimental setup. HBV Tg mice were treated with PBS or IL-2c and livers were collected and analyzed four days after treatment.

(B) Amount of ALT in the serum of the indicated mice at the indicated timepoints.

(C) Numbers of KCs (identified as live, CD45⁺, F4/80⁺, TIM4⁺ cells) per gram of liver in the indicated mice.

(D) Representative flow cytometry plots of KC1 (CD206⁻ ESAM⁻) and KC2 (CD206⁺ ESAM⁺) in the indicated mice.

(E) KC1/KC2 ratio in the indicated mice (PBS, n=4; IL-2c, n=3).

(F) Schematic representation of the experimental setup. MUP-core mice were injected with PBS or Cor93 T_{EFF}. Livers were collected and analyzed one day after T cell transfer.

(G) Amount of ALT in the serum of indicated mice at the indicated timepoints.

(H) Numbers of KCs per gram of liver in the indicated mice.

*** p value < 0.001, two-way ANOVA with Sidak's multiple comparison test.

(I) Representative flow cytometry plots of KC1 (CD206⁻ ESAM⁻) and KC2 (CD206⁺ ESAM⁺) in the indicated mice.

(J) KC1/KC2 ratio in the indicated mice (PBS, n=3; Cor93 T_{eff}, n=3). Data are representative of at least 2 independent experiments.