

The “*Sharp*” blade against HIF-mediated metastasis

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Hypoxia-inducible factors (HIFs) control cellular adaptation to oxygen deprivation. Cancer cells engage HIFs to sustain their growth in adverse conditions, thus promoting a cellular reprogramming that includes metabolism, proliferation, survival and mobility. HIFs overexpression in human cancer biopsies correlates with high metastasis and mortality. A recent report has elucidated a novel mechanism for HIFs regulation in triple-negative breast cancer. Specifically, the basic helix-loop-helix (bHLH), Sharp-1, serves HIF1 α to the proteasome and promotes its O₂-indendpendet degradation, counteracting HIF-mediated metastasis. These findings shed light on how HIFs are manipulated during cancer pathogenesis.

Metastasis dissemination is the main determinant for cancer-related deaths.^{1–4} The molecular basis to acquire ability to colonize other organs by cancer cells has been long studied, but it is still poorly understood.⁵ Hypoxia-inducible factors (HIFs) play a key role in different aspects of cancer biology, including angiogenesis,^{6–13} metabolic reprogramming,^{14–20} drug resistance,^{21–24} epithelial-mesenchymal transition (EMT)^{25–29} and metastasis.^{30–33} A recent paper reports novel data, elucidating novel aspects of the molecular basis of HIFs’ regulation in metastasization of human breast cancer.³⁴

The hypoxia-inducible factors 1 and 2 (HIF1 and HIF2, together referred to as HIFs) mediate physiological functions in cellular adaptation to hypoxic stress, modulating O₂-dependent transcriptional responses.^{35–44} HIFs are heterodimeric

transcriptional factors composed by the O₂-labile α -subunits (HIF1 α and HIF2 α) and the stable β -subunits (HIF1 β and HIF2 β). HIFs activity, hence, is primarily controlled by intracellular oxygen level. In normoxia condition, high O₂ concentration (20% O₂) confers to specific prolyl-hydroxilases (PHDs)⁴⁵ the ability to modify two proline residues on HIFs α -subunits, promoting their ubiquitin-dependent proteosomal degradation, mainly mediated by Von Hippel-Lindau E3 ubiquitin-ligase (VHL).^{46–51} When intracellular oxygen concentration falls down to hypoxia (less than 2% O₂), unhydroxylated status of prolines impairs VHL recognition, resulting in HIFs intracellular accumulation.^{45,52–54} HIFs activation has the ability to widely modify the cell transcriptional profile.^{55,56} HIFs, indeed, mediate expression of several genes related to metabolism (GLUT1, GLUT3, ALDOA, ENO1, GAPDH, HK1, HK2, PFKL, PGK1, PKM2, LDHA),^{57–61} proliferation (IGF-2, TGFA, VEGFA),^{12,62–64} survival (TERT, NANOG, OCT4)^{65–71} and cell migration-invasion (ZEB1, ZEB2, SNAI2, MMP14, MMP9, AMF, MET, PTHrP)^{72–74} (for review, Keith et al.³⁵). Thus, HIFs support the adaptation to cell growth in oxygen deprivation.^{35,41,75–79} Upregulation of HIF α -subunits has been reported in a wide range of human cancers and is often associated with poor prognosis.^{80–85} Ability to adapt to hypoxia, indeed, has long been correlated to cancer progression, metastasis and bad prognosis in human cancers.^{57,61,75,86–88} Multiple animal cancer models sustain a role for HIFs in these processes.^{42,89–94} For example, conditional deletion of HIF-1 α in

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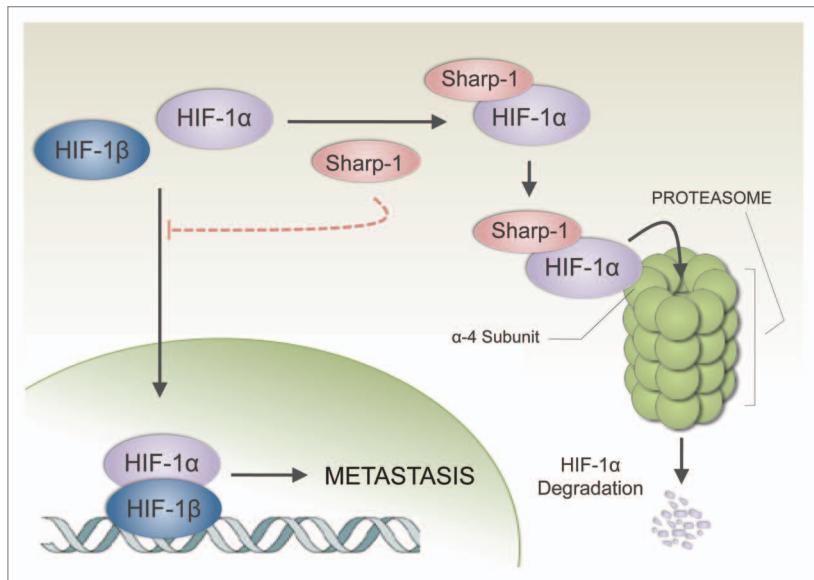


Figure 1. Sharp-1 counteracts metastasis promoting HIF-1 α degradation. HIF-1 α and HIF-1 β form an active heterodimer able to regulate expression of several genes required by cancer cells for acquisition of metastasis propensity. Upon physical interaction, Sharp-1 drives HIF-1 α on the $\alpha 4$ subunit of 20 S proteasome, promoting its ubiquitin-independent degradation.

breast epithelium of a transgenic model for metastatic mammary tumor (mouse mammary tumor virus, MMTV) reduced initial growth of primary tumors and significantly repressed pulmonary metastasis.⁹⁵ Moreover, HIFs also support cancer progression in tumor-associated cells.⁹⁶⁻¹⁰¹ Indeed, endothelial-specific deletion of HIF1 α or HIF2 α in recipient mice reduced tumor expansion in xeno-graft experiments.¹⁰² However, very often HIFs regulation in cancer cells appears to exempt from the control exerted by O₂ level.¹⁰³

In a recent *Nature* letter, Montagner et al., in Piccolo's lab, elucidated a novel mechanism for O₂-independent degradation of HIFs, mediated by Sharp-1, a rising relevant implications for invasiveness and metastatic ability of human triple-negative breast cancers (TNBC).³⁴ TNBC includes a heterogeneous subset of breast cancers, carrying negative expression for three clinical parameters: estrogen receptor (ER), progesterone receptor (PR) and epithelial receptor B2 (ERBB2, also known as HER2).¹⁰⁴⁻¹⁰⁸ This subset is generally identified with an aggressive phenotype, with short periods of disease-free, high propensity for visceral or central nervous system metastases and poor overall survival. Moreover, this hormonal

status (ER; PR; HER2⁺) drastically limits the therapeutic chances.¹⁰⁹⁻¹¹²

Sharp-1 is a basic helix-loop-helix (bHLH) transcription factor involved in different cellular processes, including proliferation, differentiation and regulation of circadian rhythm.¹¹³ Sharp-1 has also been proposed as a metastatic suppressor candidate implicated in mutant p53-mediated metastasis.¹¹⁴⁻¹²³ Mutant-p53, indeed, promotes TGF β -dependent migration by repressing transcriptional ability of the metastatic suppressor TAp63,¹²⁴⁻¹³¹ resulting in a downstream Sharp-1 downregulation.^{115,132-136} Knockdown of Sharp-1 resembled mutant-p53 overexpression, increasing migration ability and metastasis propensity of different breast cancer cell lines.¹³⁷ Montagner and colleagues identified a significant association between HIFs activity and Sharp-1 signature in a cohort of TNBC patients. Low Sharp-1 expression, indeed, associated with high HIF activity and with low metastasis-free survival. Moreover, HIF-1 α physically interacts with Sharp-1 in different cellular contexts. In a previous report, overexpressed HIF-1 α and Sharp-1 co-immunoprecipitated in Cos7 cells, and Sharp-1 overexpression repressed HIF-1 α -dependent control of VEGF-A promoter.¹³⁸ In Montagner's

paper, authors showed physical interaction between endogenous HIF-1 α and Sharp-1 in different TNBC cell lines (MDA-231, Hs578T and SUM159). Upon this interaction, HIF-1 α protein level was proved to be reduced and HIF's targets inhibited in an O₂-independent manner. Interestingly, HIF-1 α protein reduction was a consequence of proteasome-dependent degradation: Sharp-1, indeed, led to HIF-1 α direct interaction with 20 S $\alpha 4$ subunit of proteasome with an ubiquitin-independent mechanism, promoting HIF-1 α degradation (Fig. 1). Therefore, according to this model, Sharp-1 represents a determinant for HIFs stability that, alternative to VHL, acts in both normoxic and hypoxic cells, promoting ubiquitin-independent HIFs' proteosomal degradation. This pathway holds a high clinical relevance. Montagner et al. performed a genome-wide analysis on TNBC cell line after Sharp-1 overexpression or HIF-1 α and HIF-2 α stable depletion. The two independent lists of genes obtained from this experiment showed a highly statistically significant overlap (Fisher test, $p < 10^{-73}$). This leads to the hypothesis that Sharp-1 acts as a global inhibitor of HIFs activity. Interestingly, from these lists of genes, authors obtained a signature with prognostic value for TNBC: low Sharp-1 signature displayed high propensity to metastasis formation. Moreover, multivariate analysis showed that signature for Sharp-1-repressed genes did not add any prognostic information if combined with high HIFs activity signature. This supported the idea that the prognostic value of Sharp-1 signature is completely contained in the prognostic value of HIFs signature. In addition to clinical data, authors also performed elegant *in vivo* experiments. Both overexpression of Sharp-1 and depletion of HIFs in TNBC cell line MDA-231 comparably abolished the ability of lung colonization after tail vein injection in mice. On the contrary, contextual overexpression of a constitutive active form of HIF-1 α partially rescued the effect of Sharp-1 overexpression.

HIFs represent a key determinant for cancer pathogenesis. Altered control of HIFs stabilization and activation involves different mechanisms that together contribute to pathologic engagement of HIFs

in cancer cell reprogramming to sustain adverse growth condition. In this scenario, the novel findings about Sharp/HIFs axis contribute to the elucidation of HIFs' regulation in human disease. Sharp-1, as potential biomarker or therapeutic target, opens alternative chances for human cancer diagnosis and treatments.¹³⁹⁻¹⁴¹

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References

- Klein CA. Parallel progression of primary tumours and metastases. *Nat Rev Cancer* 2009; 9:302-12; PMID:19308069; <http://dx.doi.org/10.1038/nrc2627>.
- Steeg PS, Theodorescu D. Metastasis: a therapeutic target for cancer. *Nat Clin Pract Oncol* 2008; 5:206-19; PMID:18253104; <http://dx.doi.org/10.1038/ncponc1066>.
- Royer C, Lu X. Epithelial cell polarity: a major gatekeeper against cancer? *Cell Death Differ* 2011; 18:1470-7; PMID:21617693; <http://dx.doi.org/10.1038/cdd.2011.60>.
- Eckert MA, Yang J. Targeting invadopodia to block breast cancer metastasis. *Oncotarget* 2011; 2:562-8; PMID:21725138.
- Iaccarino I, Martins LM. Therapeutic targets in cancer cell metabolism and death. *Cell Death Differ* 2011; 18:565-70; PMID:21212794; <http://dx.doi.org/10.1038/cdd.2010.174>.
- Carmeliet P, Dor Y, Herbert JM, Fukumura D, Brusselmans K, Dewerchin M, et al. Role of HIF-1alpha in hypoxia-mediated apoptosis, cell proliferation and tumour angiogenesis. *Nature* 1998; 394:485-90; PMID:9697772; <http://dx.doi.org/10.1038/28867>.
- Lee K, Qian DZ, Rey S, Wei H, Liu JO, Semenza GL. Anthracycline chemotherapy inhibits HIF-1 transcriptional activity and tumor-induced mobilization of circulating angiogenic cells. *Proc Natl Acad Sci USA* 2009; 106:2353-8; PMID:19168635; <http://dx.doi.org/10.1073/pnas.0812801106>.
- Lee K, Zhang H, Qian DZ, Rey S, Liu JO, Semenza GL. Acriflavine inhibits HIF-1 dimerization, tumor growth, and vascularization. *Proc Natl Acad Sci USA* 2009; 106:17910-5; PMID:19805192; <http://dx.doi.org/10.1073/pnas.0909353106>.
- Ebos JM, Lee CR, Cruz-Munoz W, Bjarnason GA, Christensen JG, Kerbel RS. Accelerated metastasis after short-term treatment with a potent inhibitor of tumor angiogenesis. *Cancer Cell* 2009; 15:232-9; PMID:19249681; <http://dx.doi.org/10.1016/j.ccr.2009.01.021>.
- Loges S, Mazzone M, Hohensinner P, Carmeliet P. Silencing or fueling metastasis with VEGF inhibitors: antiangiogenesis revisited. *Cancer Cell* 2009; 15:167-70; PMID:19249675; <http://dx.doi.org/10.1016/j.ccr.2009.02.007>.
- Zhang Z, Neiva KG, Lingen MW, Ellis LM, Nör JE. VEGF-dependent tumor angiogenesis requires inverse and reciprocal regulation of VEGFR1 and VEGFR2. *Cell Death Differ* 2010; 17:499-512; PMID:19834490; <http://dx.doi.org/10.1038/cdd.2009.152>.
- Trisciuglio D, Gabellini C, Desideri M, Ragazzoni Y, De Luca T, Ziparo E, et al. Involvement of BH4 domain of bcl-2 in the regulation of HIF-1-mediated VEGF expression in hypoxic tumor cells. *Cell Death Differ* 2011; 18:1024-35; PMID:21233846; <http://dx.doi.org/10.1038/cdd.2010.175>.
- Zhang Y, Chen M, Venugopal S, Zhou Y, Xiang W, Li YH, et al. Isthmin exerts pro-survival and death-promoting effect on endothelial cells through alphav-beta5 integrin depending on its physical state. *Cell Death Dis* 2011; 2:e153; PMID:21544092; <http://dx.doi.org/10.1038/cddis.2011.37>.
- Zhang H, Gao P, Fukuda R, Kumar G, Krishnamachary B, Zeller KI, et al. HIF-1 inhibits mitochondrial biogenesis and cellular respiration in VHL-deficient renal cell carcinoma by repression of C-MYC activity. *Cancer Cell* 2007; 11:407-20; PMID:17482131; <http://dx.doi.org/10.1016/j.ccr.2007.04.001>.
- Gao P, Zhang H, Dinavahi R, Li F, Xiang Y, Raman V, et al. HIF-dependent antitumorigenic effect of antioxidants in vivo. *Cancer Cell* 2007; 12:230-8; PMID:17785024; <http://dx.doi.org/10.1016/j.ccr.2007.08.004>.
- Luo W, Hu H, Chang R, Zhong J, Knabel M, O'Meally R, et al. Pyruvate kinase M2 is a PHD3-stimulated coactivator for hypoxia-inducible factor 1. *Cell* 2011; 145:732-44; PMID:21620138; <http://dx.doi.org/10.1016/j.cell.2011.03.054>.
- Zhang N, Fu Z, Linke S, Chicher J, Gorman JJ, Visk D, et al. The asparaginyl hydroxylase factor inhibiting HIF-1alpha is an essential regulator of metabolism. *Cell Metab* 2010; 11:364-78; PMID:20399150; <http://dx.doi.org/10.1016/j.cmet.2010.03.001>.
- Puisségur MP, Mazure NM, Bertero T, Pradelli L, Grossi S, Robbe-Sermesant K, et al. miR-210 is over-expressed in late stages of lung cancer and mediates mitochondrial alterations associated with modulation of HIF-1 activity. *Cell Death Differ* 2011; 18:465-78; PMID:20885442; <http://dx.doi.org/10.1038/cdd.2010.119>.
- Liu X, Hajnóczky G. Altered fusion dynamics underlie unique morphological changes in mitochondria during hypoxia-reoxygenation stress. *Cell Death Differ* 2011; 18:1561-72; PMID:21372848; <http://dx.doi.org/10.1038/cdd.2011.13>.
- Ben Mosbah I, Alfany-Fernández I, Martel C, Zaouali MA, Bintanel-Morcillo M, Rimola A, et al. Endoplasmic reticulum stress inhibition protects steatotic and non-steatotic livers in partial hepatectomy under ischemia-reperfusion. *Cell Death Dis* 2010; 1:e52; PMID:21364657; <http://dx.doi.org/10.1038/cddis.2010.29>.
- Comerford KM, Wallace TJ, Karhausen J, Louis NA, Montalvo MC, Colgan SP. Hypoxia-inducible factor-1-dependent regulation of the multidrug resistance (MDR1) gene. *Cancer Res* 2002; 62:3387-94; PMID:12067980.
- Erler JT, Cawthorne CJ, Williams KJ, Koritzinsky M, Wouters BG, Wilson C, et al. Hypoxia-mediated down-regulation of Bid and Bax in tumors occurs via hypoxia-inducible factor 1-dependent and -independent mechanisms and contributes to drug resistance. *Mol Cell Biol* 2004; 24:2875-89; PMID:15024076; <http://dx.doi.org/10.1128/MCB.24.7.2875-2889.2004>.
- Verbrugge I, Maas C, Heijkoop M, Verheij M, Borst J. Radiation and anticancer drugs can facilitate mitochondrial bypass by CD95/Fas via c-FLIP downregulation. *Cell Death Differ* 2010; 17:551-61; PMID:19798106; <http://dx.doi.org/10.1038/cdd.2009.141>.
- Morizot A, Mérimo D, Lalaoui N, Jacquemin G, Granci V, Iessi E, et al. Chemotherapy overcomes TRAIL-R4-mediated TRAIL resistance at the DISC level. *Cell Death Differ* 2011; 18:700-11; PMID:21072058; <http://dx.doi.org/10.1038/cdd.2010.144>.
- Esteban MA, Tran MG, Harten SK, Hill P, Castellanos MC, Chandra A, et al. Regulation of E-cadherin expression by VHL and hypoxia-inducible factor. *Cancer Res* 2006; 66:3567-75; PMID:16585181; <http://dx.doi.org/10.1158/0008-5472.CAN-05-2670>.
- Mak P, Leav I, Pursell B, Bae D, Yang X, Taglienti CA, et al. ERbeta impedes prostate cancer EMT by destabilizing HIF-1alpha and inhibiting VEGF-mediated snail nuclear localization: implications for Gleason grading. *Cancer Cell* 2010; 17:319-32; PMID:20385358; <http://dx.doi.org/10.1016/j.ccr.2010.02.030>.
- Wu MZ, Tsai YP, Yang MH, Huang CH, Chang SY, Chang CC, et al. Interplay between HDAC3 and WDR5 is essential for hypoxia-induced epithelial-mesenchymal transition. *Mol Cell* 2011; 43:811-22; PMID:21884981; <http://dx.doi.org/10.1016/j.molcel.2011.07.012>.
- Saxena M, Stephens MA, Pathak H, Rangarajan A. Transcription factors that mediate epithelial-mesenchymal transition lead to multidrug resistance by upregulating ABC transporters. *Cell Death Dis* 2011; 2:e179; PMID:21734725; <http://dx.doi.org/10.1038/cddis.2011.61>.
- Smit MA, Peepre DS. Epithelial-mesenchymal transition and senescence: two cancer-related processes are crossing paths. *Aging (Albany NY)* 2010; 2:735-41; PMID:20975209.
- Erler JT, Bennewith KL, Nicolau M, Dornhöfer N, Kong C, Le QT, et al. Lysyl oxidase is essential for hypoxia-induced metastasis. *Nature* 2006; 440:1222-6; PMID:16642001; <http://dx.doi.org/10.1038/nature04695>.
- Wong CC, Gilkes DM, Zhang H, Chen J, Wei H, Chaturvedi P, et al. Hypoxia-inducible factor 1 is a master regulator of breast cancer metastatic niche formation. *Proc Natl Acad Sci USA* 2011; 108:16369-74; PMID:21911388; <http://dx.doi.org/10.1073/pnas.1113483108>.
- Zhang H, Wong CC, Wei H, Gilkes DM, Korangath P, Chaturvedi P, et al. HIF-1-dependent expression of angiopoietin-like 4 and L1CAM mediates vascular metastasis of hypoxic breast cancer cells to the lungs. *Oncogene* 2012; 31:1757-70; PMID:21860410; <http://dx.doi.org/10.1038/onc.2011.365>.
- Carbonaro M, O'Brate A, Giannakakou P. Microtubule disruption targets HIF-1alpha mRNA to cytoplasmic P-bodies for translational repression. *J Cell Biol* 2011; 192:83-99; PMID:21220510; <http://dx.doi.org/10.1083/jcb.201004145>.
- Montagner M, Enzo E, Forcato M, Zanconato F, Parenti A, Rampazzo E, et al. SHARP1 suppresses breast cancer metastasis by promoting degradation of hypoxia-inducible factors. *Nature* 2012; 487:380-4; PMID:22801492; <http://dx.doi.org/10.1038/nature11207>.
- Keith B, Johnson RS, Simon MC. HIF1α and HIF2α: sibling rivalry in hypoxic tumour growth and progression. *Nat Rev Cancer* 2012; 12:9-22; PMID:22169972.
- Wang GL, Jiang BH, Rue EA, Semenza GL. Hypoxia-inducible factor 1 is a basic-helix-loop-helix-PAS heterodimer regulated by cellular O2 tension. *Proc Natl Acad Sci USA* 1995; 92:5510-4; PMID:7539918; <http://dx.doi.org/10.1073/pnas.92.12.5510>.
- Tian H, McKnight SL, Russell DW. Endothelial PAS domain protein 1 (EPAS1), a transcription factor selectively expressed in endothelial cells. *Genes Dev* 1997; 11:72-82; PMID:9000051; <http://dx.doi.org/10.1101/gad.11.1.72>.
- Flamme I, Fröhlich T, von Reutern M, Kappel A, Damert A, Risau W, HRF, a putative basic helix-loop-helix-PAS-domain transcription factor is closely related to hypoxia-inducible factor-1 alpha and developmentally expressed in blood vessels. *Mech Dev* 1997; 63:51-60; PMID:9178256; [http://dx.doi.org/10.1016/S0925-4773\(97\)00674-6](http://dx.doi.org/10.1016/S0925-4773(97)00674-6).

39. Ema M, Taya S, Yokotani N, Sogawa K, Matsuda Y, Fuji-Kuriyama Y. A novel bHLH-PAS factor with close sequence similarity to hypoxia-inducible factor 1alpha regulates the VEGF expression and is potentially involved in lung and vascular development. *Proc Natl Acad Sci USA* 1997; 94:4273-8; PMID:9113979; <http://dx.doi.org/10.1073/pnas.94.9.4273>.
40. Hogenesch JB, Chan WK, Jackiw VH, Brown RC, Gu YZ, Pray-Grant M, et al. Characterization of a subset of the basic-helix-loop-helix-PAS superfamily that interacts with components of the dioxin signaling pathway. *J Biol Chem* 1997; 272:8581-93; PMID:9079689; <http://dx.doi.org/10.1074/jbc.272.13.8581>.
41. Acker T, Diez-Juan A, Aragones J, Tjwa M, Brusselmans K, Moons L, et al. Genetic evidence for a tumor suppressor role of HIF-2alpha. *Cancer Cell* 2005; 8:131-41; PMID:16098466; <http://dx.doi.org/10.1016/j.ccr.2005.07.003>.
42. Ousset M, Bouquet F, Fallone F, Biard D, Dray C, Valet P, et al. Loss of ATM positively regulates the expression of hypoxia inducible factor 1 (HIF-1) through oxidative stress: Role in the physiopathology of the disease. *Cell Cycle* 2010; 9:2814-22; PMID:20676049; <http://dx.doi.org/10.4161/cc.9.14.12253>.
43. Franovic A, Lee S. HIF-2alpha: many cancers, one engine? *Cell Cycle* 2010; 9:859-60; PMID:20348840; <http://dx.doi.org/10.4161/cc.9.5.11183>.
44. Hwang AB, Lee SJ. Regulation of life span by mitochondrial respiration: the HIF-1 and ROS connection. *Aging (Albany NY)* 2011; 3:304-10; PMID:21389351.
45. Kaelin WG Jr., Ratcliffe PJ. Oxygen sensing by metazoans: the central role of the HIF hydroxylase pathway. *Mol Cell* 2008; 30:393-402; PMID:18498744; <http://dx.doi.org/10.1016/j.molcel.2008.04.009>.
46. Qian DZ, Kachhap SK, Collis SJ, Verheul HM, Carducci MA, Atadja P, et al. Class II histone deacetylases are associated with VHL-independent regulation of hypoxia-inducible factor 1 alpha. *Cancer Res* 2006; 66:8814-21; PMID:16951198; <http://dx.doi.org/10.1158/0008-5472.CAN-05-4598>.
47. Kaelin WG Jr. The von Hippel-Lindau tumour suppressor protein: O₂ sensing and cancer. *Nat Rev Cancer* 2008; 8:865-73; PMID:18923434; <http://dx.doi.org/10.1038/nrc2502>.
48. Maranchie JK, Vasselli JR, Riss J, Bonifacino JS, Linehan WM, Klausner RD. The contribution of VHL substrate binding and HIF1-alpha to the phenotype of VHL loss in renal cell carcinoma. *Cancer Cell* 2002; 1:247-55; PMID:12086861; [http://dx.doi.org/10.1016/S1535-6108\(02\)00044-2](http://dx.doi.org/10.1016/S1535-6108(02)00044-2).
49. Kondo K, Klco J, Nakamura E, Lechpammer M, Kaelin WG Jr. Inhibition of HIF is necessary for tumor suppression by the von Hippel-Lindau protein. *Cancer Cell* 2002; 1:237-46; PMID:12086860; [http://dx.doi.org/10.1016/S1535-6108\(02\)00043-0](http://dx.doi.org/10.1016/S1535-6108(02)00043-0).
50. Badiola N, Penas C, Mifano-Molina A, Barneda-Zahonero B, Fadó R, Sánchez-Opazo G, et al. Induction of ER stress in response to oxygen-glucose deprivation of cortical cultures involves the activation of the PERK and IRE-1 pathways and of caspase-12. *Cell Death Dis* 2011; 2:e149; PMID:21525936; <http://dx.doi.org/10.1038/cddis.2011.31>.
51. Roe JS, Kim HR, Hwang IY, Ha NC, Kim ST, Cho EJ, et al. Phosphorylation of von Hippel-Lindau protein by checkpoint kinase 2 regulates p53 transactivation. *Cell Cycle* 2011; 10:3920-8; PMID:22071692; <http://dx.doi.org/10.4161/cc.10.22.18096>.
52. Majmundar AJ, Wong WJ, Simon MC. Hypoxia-inducible factors and the response to hypoxic stress. *Mol Cell* 2010; 40:294-309; PMID:20965423; <http://dx.doi.org/10.1016/j.molcel.2010.09.022>.
53. Guitart AV, Debeissat C, Hermite F, Villacreses A, Ivanovic Z, Boeuf H, et al. Very low oxygen concentration (0.1%) reveals two FDCP-Mix cell subpopulations that differ by their cell cycling, differentiation and p27kip1 expression. *Cell Death Differ* 2011; 18:174-82; PMID:20671746; <http://dx.doi.org/10.1038/cdd.2010.85>.
54. Tiede LM, Cook EA, Morsey B, Fox HS. Oxygen matters: tissue culture oxygen levels affect mitochondrial function and structure as well as responses to HIV viroproteins. *Cell Death Dis* 2011; 2:e246; PMID:22190005; <http://dx.doi.org/10.1038/cddis.2011.128>.
55. Sheffer M, Simon AJ, Jacob-Hirsch J, Rechavi G, Domany E, Givol D, et al. Genome-wide analysis discloses reversal of the hypoxia-induced changes of gene expression in colon cancer cells by zinc supplementation. *Oncotarget* 2011; 2:1191-202; PMID:22202117.
56. Nardinocchi L, Puca R, D'Orazi G. HIF-1α antagonizes p53-mediated apoptosis by triggering HIPK2 degradation. *Aging (Albany NY)* 2011; 3:33-43; PMID:21248371.
57. Chen ZX, Pervaiz S. Involvement of cytochrome c oxidase subunits Va and Vb in the regulation of cancer cell metabolism by Bcl-2. *Cell Death Differ* 2010; 17:408-20; PMID:19834492; <http://dx.doi.org/10.1038/cdd.2009.132>.
58. Russo R, Berliocchi L, Adornetto A, Varano GP, Cavaliere F, Nucci C, et al. Calpain-mediated cleavage of Beclin-1 and autophagy deregulation following retinal ischemic injury *in vivo*. *Cell Death Dis* 2011; 2:e144; PMID:21490676; <http://dx.doi.org/10.1038/cddis.2011.29>.
59. Darnell JE Jr. STAT3, HIF-1, glucose addiction and Warburg effect. *Aging (Albany NY)* 2010; 2:890-1; PMID:21149895.
60. Demaria M, Giorgi C, Lebiedzinska M, Esposito G, D'Angeli L, Bartoli A, et al. A STAT3-mediated metabolic switch is involved in tumour transformation and STAT3 addiction. *Aging (Albany NY)* 2010; 2:823-42; PMID:21084727.
61. Pavlides S, Tsirigos A, Vera I, Flomenberg N, Frank PG, Casimiro MC, et al. Transcriptional evidence for the "Reverse Warburg Effect" in human breast cancer tumor stroma and metastasis: similarities with oxidative stress, inflammation, Alzheimer's disease, and "Neuron-Glia Metabolic Coupling". *Aging (Albany NY)* 2010; 2:185-99; PMID:20442453.
62. Cam H, Easton JB, High A, Houghton PJ. mTORC1 signaling under hypoxic conditions is controlled by ATM-dependent phosphorylation of HIF-1α. *Mol Cell* 2010; 40:509-20; PMID:21095582; <http://dx.doi.org/10.1016/j.molcel.2010.10.030>.
63. Mitchell GC, Fillinger JL, Sittadjody S, Avila JL, Burd R, Limesand KH. IGF1 activates cell cycle arrest following irradiation by reducing binding of ΔNp63 to the p21 promoter. *Cell Death Dis* 2010; 1:e50; PMID:21480565; <http://dx.doi.org/10.1038/cddis.2010.28>.
64. Li X, Kumar A, Zhang F, Lee C, Li Y, Tang Z, et al. VEGF-independent angiogenic pathways induced by PDGF-C. *Oncotarget* 2010; 1:309-14; PMID:20871734.
65. Mathieu J, Zhang Z, Zhou W, Wang AJ, Heddleston JM, Pinna CM, et al. HIF induces human embryonic stem cell markers in cancer cells. *Cancer Res* 2011; 71:4640-52; PMID:21712410; <http://dx.doi.org/10.1158/0008-5472.CAN-10-3320>.
66. Mazumdar J, O'Brien WT, Johnson RS, LaManna JC, Chavez JC, Klein PS, et al. O₂ regulates stem cells through Wnt/β-catenin signalling. *Nat Cell Biol* 2010; 12:1007-13; PMID:20852629; <http://dx.doi.org/10.1038/ncb2102>.
67. Ko CY, Tsai MY, Tseng WF, Cheng CH, Huang CR, Wu JS, et al. Integration of CNS survival and differentiation by HIF2α. *Cell Death Differ* 2011; 18:1757-70; PMID:21546908; <http://dx.doi.org/10.1038/cdd.2011.44>.
68. Torii S, Goto Y, Ishizawa T, Hoshi H, Goryo K, Yasumoto K, et al. Pro-apoptotic activity of inhibitory PAS domain protein (IPAS), a negative regulator of HIF-1, through binding to pro-survival Bcl-2 family proteins. *Cell Death Differ* 2011; 18:1711-25; PMID:21546903; <http://dx.doi.org/10.1038/cdd.2011.47>.
69. Mora-Castilla S, Tejedo JR, Hmadcha A, Cahuana GM, Martín F, Soria B, et al. Nitric oxide repression of Nanog promotes mouse embryonic stem cell differentiation. *Cell Death Differ* 2010; 17:1025-33; PMID:20075941; <http://dx.doi.org/10.1038/cdd.2009.204>.
70. Stacpoole SR, Bilican B, Webber DJ, Luzhynskaya A, He XL, Compston A, et al. Derivation of neural precursor cells from human ES cells at 3% O₂ is efficient, enhances survival and presents no barrier to regional specification and functional differentiation. *Cell Death Differ* 2011; 18:1016-23; PMID:21274009; <http://dx.doi.org/10.1038/cdd.2010.171>.
71. Casati A, Frascoli M, Traggiai E, Proietti M, Schenk U, Grassi F. Cell-autonomous regulation of hematopoietic stem cell cycling activity by ATP. *Cell Death Differ* 2011; 18:396-404; PMID:20798687; <http://dx.doi.org/10.1038/cdd.2010.107>.
72. Bracken CP, Gregory PA, Kolesnikoff N, Bert AG, Wang J, Shannon MF, et al. A double-negative feedback loop between ZEB1-SIP1 and the microRNA-200 family regulates epithelial-mesenchymal transition. *Cancer Res* 2008; 68:7846-54; PMID:18829540; <http://dx.doi.org/10.1158/0008-5472.CAN-08-1942>.
73. Brabetz S, Brabetz T. The ZEB/miR-200 feedback loop—a motor of cellular plasticity in development and cancer? *EMBO Rep* 2010; 11:670-7; PMID:20706219; <http://dx.doi.org/10.1038/embor.2010.117>.
74. Manisterski M, Golan M, Amir S, Weisman Y, Mabjeesh NJ. Hypoxia induces PTHrP gene transcription in human cancer cells through the HIF-2α. *Cell Cycle* 2010; 9:3723-9; PMID:20890122; <http://dx.doi.org/10.4161/cc.9.18.12931>.
75. Bertout JA, Patel SA, Simon MC. The impact of O₂ availability on human cancer. *Nat Rev Cancer* 2008; 8:967-75; PMID:18987634; <http://dx.doi.org/10.1038/nrc2540>.
76. Li Q, Li H, Roughton K, Wang X, Kroemer G, Blomgren K, et al. Lithium reduces apoptosis and autophagy after neonatal hypoxia-ischemia. *Cell Death Dis* 2010; 1:e56; PMID:21364661; <http://dx.doi.org/10.1038/cddis.2010.33>.
77. Lemarie A, Huc L, Pazarentzos E, Mahul-Mellier AL, Grimm S. Specific disintegration of complex II succinate:ubiquinone oxidoreductase links pH changes to oxidative stress for apoptosis induction. *Cell Death Differ* 2011; 18:338-49; PMID:20706275; <http://dx.doi.org/10.1038/cdd.2010.93>.
78. Mongiardi MP, Stagni V, Natoli M, Giaccari D, D'Agnano I, Falchetti ML, et al. Oxygen sensing is impaired in ATM-defective cells. *Cell Cycle* 2011; 10:4311-20; PMID:22134239; <http://dx.doi.org/10.4161/cc.10.24.18663>.
79. Whelan KA, Regnato MJ. Surviving without oxygen: hypoxia regulation of mammary morphogenesis and anoikis. *Cell Cycle* 2011; 10:2287-94; PMID:21670595; <http://dx.doi.org/10.4161/cc.10.14.16532>.
80. Yoshimura H, Dhar DK, Kohno H, Kubota H, Fujii T, Ueda S, et al. Prognostic impact of hypoxia-inducible factors 1alpha and 2alpha in colorectal cancer patients: correlation with tumor angiogenesis and cyclooxygenase-2 expression. *Clin Cancer Res* 2004; 10:8554-60; PMID:15623639; <http://dx.doi.org/10.1158/1078-0432.CCR-0946-03>.

81. Noguera R, Fredlund E, Piqueras M, Pietras A, Beckman S, Navarro S, et al. HIF-1alpha and HIF-2alpha are differentially regulated in vivo in neuroblastoma: high HIF-1alpha correlates negatively to advanced clinical stage and tumor vascularization. *Clin Cancer Res* 2009; 15:7130-6; PMID:19903792; <http://dx.doi.org/10.1158/1078-0432.CCR-09-0223>.
82. Lidgren A, Hedberg Y, Grankvist K, Rasmussen T, Vasko J, Ljungberg B. The expression of hypoxia-inducible factor 1alpha is a favorable independent prognostic factor in renal cell carcinoma. *Clin Cancer Res* 2005; 11:1129-35; PMID:15709180.
83. Klatte T, Seligson DB, Riggs SB, Leppert JT, Berkman MK, Kleid MD, et al. Hypoxia-inducible factor 1 alpha in clear cell renal cell carcinoma. *Clin Cancer Res* 2007; 13:7388-93; PMID:18094421; <http://dx.doi.org/10.1158/1078-0432.CCR-07-0411>.
84. Tomé M, López-Romero P, Albo C, Sepúlveda JC, Fernández-Gutiérrez B, Dopazo A, et al. miR-335 orchestrates cell proliferation, migration and differentiation in human mesenchymal stem cells. *Cell Death Differ* 2011; 18:985-95; PMID:21164520; <http://dx.doi.org/10.1038/cdd.2010.167>.
85. Adam J, Ratcliffe PJ, Pollard PJ. Novel insights into FH-associated disease are KEAPing the lid on oncogenic HIF signalling. *Oncotarget* 2011; 2:820-1; PMID:22064867.
86. Kim WY, Oh SH, Woo JK, Hong WK, Lee HY. Targeting heat shock protein 90 overrides the resistance of lung cancer cells by blocking radiation-induced stabilization of hypoxia-inducible factor 1alpha. *Cancer Res* 2009; 69:1624-32; PMID:19176399; <http://dx.doi.org/10.1158/0008-5472.CAN-08-0505>.
87. Marcel V, Dichtel-Danjoy ML, Sagne C, Hafsi H, Ma D, Ortiz-Cuaran S, et al. Biological functions of p53 isoforms through evolution: lessons from animal and cellular models. *Cell Death Differ* 2011; 18:1815-24; PMID:21941372; <http://dx.doi.org/10.1038/cdd.2011.120>.
88. Takeuchi M, Kimura S, Kuroda J, Ashihara E, Kawatani M, Osada H, et al. Glyoxalase-I is a novel target against Bcr-Abl+ leukemic cells acquiring stem-like characteristics in a hypoxic environment. *Cell Death Differ* 2010; 17:1211-20; PMID:20139893; <http://dx.doi.org/10.1038/cdd.2010.6>.
89. Semenza GL. Targeting HIF-1 for cancer therapy. *Nat Rev Cancer* 2003; 3:721-32; PMID:13130303; <http://dx.doi.org/10.1038/nrc1187>.
90. Rankin EB, Giaccia AJ. The role of hypoxia-inducible factors in tumorigenesis. *Cell Death Differ* 2008; 15:678-85; PMID:18259193; <http://dx.doi.org/10.1038/cdd.2008.21>.
91. Mazumdar J, Hickey MM, Pant DK, Durham AC, Sweet-Cordero A, Vachani A, et al. HIF-2alpha deletion promotes Kris-driven lung tumor development. *Proc Natl Acad Sci USA* 2010; 107:14182-7; PMID:20660313; <http://dx.doi.org/10.1073/pnas.1001296107>.
92. Martinez-Outschoorn UE, Trimmer C, Lin Z, Whitaker-Menezes D, Chiavarina B, Zhou J, et al. Autophagy in cancer associated fibroblasts promotes tumor cell survival: Role of hypoxia, HIF1 induction and NF κ B activation in the tumor stromal microenvironment. *Cell Cycle* 2010; 9:3515-33; PMID:20855962; <http://dx.doi.org/10.4161/cc.9.17.12928>.
93. Thangarajah H, Vial IN, Grogan RH, Yao D, Shi Y, Januszyk M, et al. HIF-1alpha dysfunction in diabetes. *Cell Cycle* 2010; 9:75-9; PMID:20016290; <http://dx.doi.org/10.4161/cc.9.1.10371>.
94. Fer N, Melillo G. The HIF-1 α -c-Myc pathway and tumorigenesis: evading the apoptotic gate-keeper. *Cell Cycle* 2011; 10:3228; PMID:21971179; <http://dx.doi.org/10.4161/cc.10.19.17049>.
95. Liao D, Corle C, Seagroves TN, Johnson RS. Hypoxia-inducible factor-1alpha is a key regulator of metastasis in a transgenic model of cancer initiation and progression. *Cancer Res* 2007; 67:563-72; PMID:17234764; <http://dx.doi.org/10.1158/0008-5472.CAN-06-2701>.
96. Takeda N, O'Dea EL, Doedens A, Kim JW, Weidemann A, Stockmann C, et al. Differential activation and antagonistic function of HIF-alpha isoforms in macrophages are essential for NO homeostasis. *Genes Dev* 2010; 24:491-501; PMID:20194441; <http://dx.doi.org/10.1101/gad.188140>.
97. Weidemann A, Krohne TU, Aguirre E, Kurihara T, Takeda N, Dorrell MI, et al. Astrocyte hypoxic response is essential for pathological but not developmental angiogenesis of the retina. *Glia* 2010; 58:1177-85; PMID:20544853.
98. Stockmann C, Kerdiles Y, Nomaksteinsky M, Weidemann A, Takeda N, Doedens A, et al. Loss of myeloid cell-derived vascular endothelial growth factor accelerates fibrosis. *Proc Natl Acad Sci USA* 2010; 107:4329-34; PMID:20142499; <http://dx.doi.org/10.1073/pnas.0912766107>.
99. Magenta A, Cencioni C, Fasanaro P, Zaccagnini G, Greco S, Sarra-Ferraris G, et al. miR-200c is upregulated by oxidative stress and induces endothelial cell apoptosis and senescence via ZEB1 inhibition. *Cell Death Differ* 2011; 18:1628-39; PMID:21527937; <http://dx.doi.org/10.1038/cdd.2011.42>.
100. Qi J, Pellecchia M, Ronai ZA. The Siah2-HIF-FoxA2 axis in prostate cancer – new markers and therapeutic opportunities. *Oncotarget* 2010; 1:379-85; PMID:21037926.
101. Kuo MT, Savaraj N, Feun LG. Targeted cellular metabolism for cancer chemotherapy with recombinant arginine-degrading enzymes. *Oncotarget* 2010; 1:246-51; PMID:21152246.
102. Tang N, Wang L, Esko J, Giordano FJ, Huang Y, Gerber HP, et al. Loss of HIF-1alpha in endothelial cells disrupts a hypoxia-driven VEGF autocrine loop necessary for tumorigenesis. *Cancer Cell* 2004; 6:485-95; PMID:15542432; <http://dx.doi.org/10.1016/j.ccr.2004.09.026>.
103. Pouysségur J, Dayan F, Mazure NM. Hypoxia signalling in cancer and approaches to enforce tumour regression. *Nature* 2006; 441:437-43; PMID:16724055; <http://dx.doi.org/10.1038/nature04871>.
104. Buzdar AU. Medical oncology: Endocrine-therapy-related symptoms and breast cancer. *Nature reviews. Clin Oncol* 2009; 6:309-10.
105. Yalcin-Ozysiz O, Fiche M, Gutierrez M, Wagner KU, Raffoul W, Briskin C. Antagonistic roles of Notch and p63 in controlling mammary epithelial cell fates. *Cell Death Differ* 2010; 17:1600-12; PMID:20379195; <http://dx.doi.org/10.1038/cdd.2010.37>.
106. Spizzo R, Nicolo MS, Lupini L, Lu Y, Fogarty J, Rossi S, et al. miR-145 participates with TP53 in a death-promoting regulatory loop and targets estrogen receptor-alpha in human breast cancer cells. *Cell Death Differ* 2010; 17:246-54; PMID:19730444; <http://dx.doi.org/10.1038/cdd.2009.117>.
107. Dever SM, Golding SE, Rosenberg E, Adams BR, Idowu MO, Quillin JM, et al. Mutations in the BRCT binding site of BRCA1 result in hyper-recombination. *Aging (Albany NY)* 2011; 3:515-32; PMID:21666281.
108. Harris JL, Khanna KK. BRCA1 A-complex fine tunes repair functions of BRCA1. *Aging (Albany NY)* 2011; 3:461-3; PMID:21805697.
109. Rieserter O, Milas L, Ang KK. Use of molecular biomarkers for predicting the response to radiotherapy with or without chemotherapy. *J Clin Oncol* 2007; 25:4075-83; PMID:17827456; <http://dx.doi.org/10.1200/JCO.2007.11.8497>.
110. Kutuk O, Letai A. Displacement of Bim by Bmf and Puma rather than increase in Bim level mediates paclitaxel-induced apoptosis in breast cancer cells. *Cell Death Differ* 2010; 17:1624-35; PMID:20431602; <http://dx.doi.org/10.1038/cdd.2010.41>.
111. Li QQ, Chen ZQ, Cao XX, Xu JD, Xu JW, Chen YY, et al. Involvement of NF- κ B/miR-448 regulatory feedback loop in chemotherapy-induced epithelial-mesenchymal transition of breast cancer cells. *Cell Death Differ* 2011; 18:16-25; PMID:20798686; <http://dx.doi.org/10.1038/cdd.2010.103>.
112. Liu B, Wen JK, Li BH, Fang XM, Wang JJ, Zhang YP, et al. Celecoxib and acetylbutrilannactone interact synergistically to suppress breast cancer cell growth via COX-2-dependent and -independent mechanisms. *Cell Death Dis* 2011; 2:e185; PMID:21796157; <http://dx.doi.org/10.1038/cddis.2011.64>.
113. Rossner MJ, Oster H, Wichert SP, Reinecke L, Wehr MC, Reinecke J, et al. Disturbed clockwork resetting in Sharp-1 and Sharp-2 single and double mutant mice. *PLoS One* 2008; 3:e2762; PMID:18648504; <http://dx.doi.org/10.1371/journal.pone.0002762>.
114. Muller PA, Vousden KH, Norman JC. p53 and its mutants in tumor cell migration and invasion. *J Cell Biol* 2011; 192:209-18; PMID:21263025; <http://dx.doi.org/10.1083/jcb.20100959>.
115. Kogan-Sakin I, Tabach Y, Buganim Y, Molchadsky A, Solomon H, Madar S, et al. Mutant p53(R175H) upregulates Twist1 expression and promotes epithelial-mesenchymal transition in immortalized prostate cells. *Cell Death Differ* 2011; 18:271-81; PMID:20689556; <http://dx.doi.org/10.1038/cdd.2010.94>.
116. Sermeus A, Michiels C. Reciprocal influence of the p53 and the hypoxic pathways. *Cell Death Dis* 2011; 2:e164; PMID:21614094; <http://dx.doi.org/10.1038/cddis.2011.48>.
117. Michaelis M, Rothweiler F, Barth S, Cinatl J, van Rikxoort M, Löschmann N, et al. Adaptation of cancer cells from different entities to the MDM2 inhibitor nutlin-3 results in the emergence of p53-mutated multi-drug-resistant cancer cells. *Cell Death Dis* 2011; 2:e243; PMID:22170099; <http://dx.doi.org/10.1038/cddis.2011.129>.
118. Gatta R, Dolfini D, Mantovani R. NF-Y joins E2Fs, p53 and other stress transcription factors at the apoptosis table. *Cell Death Dis* 2011; 2:e162; PMID:21614092; <http://dx.doi.org/10.1038/cddis.2011.45>.
119. Jiang M, Chiu SY, Hsu W. SUMO-specific protease 2 in Mdm2-mediated regulation of p53. *Cell Death Differ* 2011; 18:1005-15; PMID:21183956; <http://dx.doi.org/10.1038/cdd.2010.168>.
120. Antico Arciuch VG, Russo MA, Dima M, Kang KS, Dasrath F, Liao XH, et al. Thyrocyte-specific inactivation of p53 and Pten results in anaplastic thyroid carcinomas faithfully recapitulating human tumors. *Oncotarget* 2011; 2:1109-26; PMID:22190384.
121. Hill R, Madureira PA, Waismann DM, Lee PW. DNA-PKcs binding to p53 on the p21WAF1/CIP1 promoter blocks transcription resulting in cell death. *Oncotarget* 2011; 2:1094-108; PMID:22190353.
122. Napoli M, Girardini JE, Piazza S, Del Sal G. Wiring the oncogenic circuitry: Pin1 unleashes mutant p53. *Oncotarget* 2011; 2:654-6; PMID:21926448.
123. Leonieva OV, Blagosklonny MV. DNA damaging agents and p53 do not cause senescence in quiescent cells, while consecutive re-activation of mTOR is associated with conversion to senescence. *Aging (Albany NY)* 2010; 2:924-35; PMID:21212465.
124. Su X, Chakravarti D, Cho MS, Liu L, Gi YJ, Lin YL, et al. TAp63 suppresses metastasis through coordinate regulation of Dicer and miRNAs. *Nature* 2010; 467:986-90; PMID:20962848; <http://dx.doi.org/10.1038/nature09459>.

125. Beaudry VG, Jiang D, Dusek RL, Park EJ, Knezevich S, Ridd K, et al. Loss of the p53/p63 regulated desmosomal protein Perp promotes tumorigenesis. *PLoS Genet* 2010; 6:e1001168; PMID:20975948; <http://dx.doi.org/10.1371/journal.pgen.1001168>.
126. Aoubala M, Murray-Zmijewski F, Khouri MP, Fernandes K, Perrier S, Bernard H, et al. p53 directly transactivates Δ 133p53 α , regulating cell fate outcome in response to DNA damage. *Cell Death Differ* 2011; 18:248-58; PMID:20689555; <http://dx.doi.org/10.1038/cdd.2010.91>.
127. Barton CE, Johnson KN, Mays DM, Boehnke K, Shyr Y, Boukamp P, et al. Novel p63 target genes involved in paracrine signaling and keratinocyte differentiation. *Cell Death Dis* 2010; 1:e74; PMID:21151771; <http://dx.doi.org/10.1038/cddis.2010.49>.
128. Borrelli S, Candi E, Hu B, Dolfini D, Ravo M, Grober OM, et al. The p63 target HBP1 is required for skin differentiation and stratification. *Cell Death Differ* 2010; 17:1896-907; PMID:20523354; <http://dx.doi.org/10.1038/cdd.2010.59>.
129. Lena AM, Cipollone R, Amelio I, Catani MV, Ramadan S, Brown G, et al. Skn-1a/Oct-11 and Δ Np63 α exert antagonizing effects on human keratin expression. *Biochem Biophys Res Commun* 2010; 401:568-73; PMID:20888799; <http://dx.doi.org/10.1016/j.bbrc.2010.09.102>.
130. Neilsen PM, Noll JE, Suetani RJ, Schulz RB, Al-Ejeh F, Evdokiou A, et al. Mutant p53 uses p63 as a molecular chaperone to alter gene expression and induce a pro-invasive secretome. *Oncotarget* 2011; 2:1203-17; PMID:22203497.
131. Lane DP, Verma C, Fang CC. The p53 inducing drug dosage may determine quiescence or senescence. *Aging (Albany NY)* 2010; 2:748; PMID:21068468.
132. Xu J, Reumers J, Couceiro JR, De Smet F, Gallardo R, Rudyak S, et al. Gain of function of mutant p53 by coaggregation with multiple tumor suppressors. *Nat Chem Biol* 2011; 7:285-95; PMID:21445056; <http://dx.doi.org/10.1038/nchembio.546>.
133. Melino G. p63 is a suppressor of tumorigenesis and metastasis interacting with mutant p53. *Cell Death Differ* 2011; 18:1487-99; PMID:21760596; <http://dx.doi.org/10.1038/cdd.2011.81>.
134. Collavin L, Lunardi A, Del Sal G. p53-family proteins and their regulators: hubs and spokes in tumor suppression. *Cell Death Differ* 2010; 17:901-11; PMID:20379196; <http://dx.doi.org/10.1038/cdd.2010.35>.
135. Tucci P, Agostini M, Grespi F, Markert EK, Terrinoni A, Vousden KH, et al. Loss of p63 and its microRNA-205 target results in enhanced cell migration and metastasis in prostate cancer. *Proc Natl Acad Sci USA* 2012; 109:15312-7; PMID:22949650; <http://dx.doi.org/10.1073/pnas.110977109>.
136. Sayeed A, Meng Z, Luciani G, Chen LC, Bennington JL, Dairkee SH. Negative regulation of UCP2 by TGF β signaling characterizes low and intermediate-grade primary breast cancer. *Cell Death Dis* 2010; 1:e53; PMID:21364658; <http://dx.doi.org/10.1038/cddis.2010.30>.
137. Adorno M, Cordenonsi M, Montagner M, Dupont S, Wong C, Hann B, et al. A Mutant-p53/Smad complex opposes p63 to empower TGFbeta-induced metastasis. *Cell* 2009; 137:87-98; PMID:19345189; <http://dx.doi.org/10.1016/j.cell.2009.01.039>.
138. Sato F, Bhawal UK, Kawamoto T, Fujimoto K, Imaizumi T, Imanaka T, et al. Basic-helix-loop-helix (bHLH) transcription factor DEC2 negatively regulates vascular endothelial growth factor expression. *Genes Cells* 2008; 13:131-44; PMID:18233956; <http://dx.doi.org/10.1111/j.1365-2443.2007.01153.x>.
139. Podar K, Anderson KC. A therapeutic role for targeting c-Myc/Hif-1-dependent signaling pathways. *Cell Cycle* 2010; 9:1722-8; PMID:20404562; <http://dx.doi.org/10.4161/cc.9.9.11358>.
140. Agostini M, Tucci P, Melino G. Cell death pathology: perspective for human diseases. *Biochem Biophys Res Commun* 2011; 414:451-5; PMID:21964290; <http://dx.doi.org/10.1016/j.bbrc.2011.09.081>.
141. Amelio I, Melino G, Knight RA. Cell death pathology: cross-talk with autophagy and its clinical implications. *Biochem Biophys Res Commun* 2011; 414:277-81; PMID:21963447; <http://dx.doi.org/10.1016/j.bbrc.2011.09.080>.