REVIEW

PAX8 as a Potential Target for Ovarian Cancer: What We Know so Far

Tina Di Palma, Mariastella Zannini

IEOS - Institute of Experimental Endocrinology and Oncology 'G. Salvatore', National Research Council, Napoli, 80131, Italy

Correspondence: Mariastella Zannini, IEOS - Institute of Experimental Endocrinology and Oncology 'G. Salvatore', National Research Council, via S. Pansini 5, Napoli, 80131, Italy, Tel +39-081-5465530, Fax +39-081-2296674, Email s.zannini@ieos.cnr.it

Abstract: The Fallopian tube epithelium harbors the origin cells for the majority of high-grade serous ovarian carcinomas (HGSCs), the most lethal form of gynecologic malignancies. PAX8 belongs to the paired-box gene family of transcription factors and it is a marker of the FTE secretory cell lineage. Its role has been investigated in migration, invasion, proliferation, cell survival, stem cell maintenance, angiogenesis and tumor growth. In this review, we focus on the pro-tumorigenic role of PAX8 in ovarian cancer; in this context, PAX8 possibly continues to exert its transcriptional activity on its physiological targets but may also function on newly available targets after the tumorigenic hits. Acquiring new insights into the different PAX8 mechanism(s) of action in the tumor microenvironment could uncover new viable therapeutic targets and thus improve the current treatment regimen. **Keywords:** PAX8, ovarian cancer, transcription factor, Fallopian tube, STICs

Introduction

Ovarian cancer (OC) represents the fifth leading cause of cancer-related death in women and, like other cancers, is a spectrum of diseases.¹ OC includes epithelial, germ cell and sex cord-stromal tumors originating from surface epithelial cells, from the germ cell or the oocyte and from stromal and primitive ovarian sex cord cells, as shown in Figure 1. The sex cord-stromal and germ cells types contribute to 8% and 3-7% of the malignant ovarian tumors, respectively; and given their histological and molecular heterogeneity, it is not easy to establish criteria for making the distinction between benign and malignant tumors. Most stromal tumors and germ cell tumors have a good prognosis.² The epithelial tumors (90%) can be divided in four major histotypes: serous, mucinous, endometrioid and clear cell³ (Figure 1). Of these, HGSC is the most common and lethal histotype and is associated with mutations in BRCA1 or BRCA2 and p53.^{4,5} Although it was originally believed that HGSC arises from the ovarian surface epithelium (OSE), it has been recently suggested by various experimental evidences that it may instead arise from the Fallopian tube fimbria.⁶ In particular, the serous tubal intraepithelial carcinomas (STICs), which are identified in the distal end (fimbria) of the Fallopian tube and arise from p53 mutations, have been indicated as the primary lesions that evolve through subsequent oncogenic events into HGSC^{6,7} (Figure 1). This hypothesis is further supported by genetically engineered murine models that mimic the transformation from Fallopian tubal secretory epithelial cells to HGSC.^{8,9} In addition, gene profiling studies showing that the gene expression profile of HGSC is more closely related to the Fallopian tubes than to the ovarian surface epithelium supported the Fallopian tubes rather than ovarian surface epithelium as the site of origin of HGSC.¹⁰ It was indeed observed that STICs and HGSC share similar histomorphological and expression profiles and 92% of the STICs express TP53 mutations that are identical between these precursor lesions and the matched invasive ovarian carcinomas.¹¹ Among the evidence that reinforce the tubal origin of the HGSC, there is also the transcription factor PAired boX8 (PAX8) that is expressed in the Fallopian tube secretory cells but not in OSE and is conserved in HGSC.^{9,12} This transcription factor is a marker of the Fallopian tube secretory cell lineage and its expression is retained in 96% of the serous ovarian carcinomas, in 89% of the endometrioid and 100% of the clear cell carcinomas, whilst it is not detected in the mucinous carcinoma.¹³ PAX8 is crucial for the organogenesis of the thyroid gland, kidney, nervous system and



Figure I The origin of OCs. OCs are classified according to the type of cells from which they originate. Among EOC, the HGSC arises from Fallopian tube lesions including STICs and p53 signature at the end of the fimbriae. As STICs progress, exfoliation of malignant cells from STICs and/or fimbriae may spread and invade rapidly to the surface of the ovary and peritoneum.

Müllerian system.^{14,15} In previous studies, we have shown that PAX8 plays a critical role in cell cycle progression and cell survival of thyroid differentiated epithelial cells.¹⁶ In tumors, PAX8 positivity has been displayed in gliomas, renal, thyroid, ovarian, Wilms and well-differentiated pancreatic neuroendocrine cancers.^{17–21} Moreover, PAX8 represents a useful marker for the diagnosis of primary or metastatic neoplasms, given its lack of expression in mammary, lung and mesothelial primary tumors.^{22–24} In the scenario of OC, the Cancer Genome Atlas (TGCA) Project indicated *PAX8* as a survival gene essential for the proliferation of ovarian cancer cells.¹³ Overall, *PAX8* belongs to a class of lineage-survival genes that are required for both normal development of specific tissues and for cancer cell proliferation/survival. In this review, we outline PAX8's function and its role in OC; moreover, we discuss PAX8 regulatory mechanism(s) and possible therapeutic approaches.

PAX8 Structure and Function

PAX8 is a member of the PAired boX (PAX) gene family that is composed of nine genes in mammals tightly regulated in both temporal and spatial expression patterns.²⁵ These genes, as described in details in Table 1, have been sub-classified into four subgroups based on shared structural motifs and similar expression pattern during embryogenesis.²⁶ The *PAX8* gene is located on chromosome 2 at position 2q12-14. Alternative splicing of *PAX8* generates five different transcripts that give rise to equally distinct isoforms that are distinguished in their functional properties.^{27,28} PAX8A with 450aa is the longest and most common isoform that includes all the codons from exon 2 to 12. PAX8 as the other members of subgroup II is characterized by the presence of a Paired-domain (PD), an octapeptide (OP) and a partial homeodomain (HD) (Table 1). The PD is a conserved 128 amino-acid DNA-binding domain composed by two helix-turn-helix motif subdomains, PAI and RED.²⁹ The OP and HD exert positive or negative regulation of transcription depending on the cellular context and availability of cofactors. Most of the current insight regarding PAX8 function during embryonic development and in adults came from studies in the thyroid. PAX8 is expressed during thyroid development and it is involved in sporadic and familial cases of CH with TD.^{30,31} Its expression plays a key role in determining the thyroid

Group	Gene	Structural Domain	Chromosomal Location	Embryonic Expression Domain
Ι	PAXI	PD	20p I I	Skeleton, thymus, pharyngeal
	PAX9	OP	14q12	Skeleton, thymus, teeth
II	PAX2 PAX5 PAX8	PD OP Partial HD	10q24 9p13 2q12	CNS, kidney CNS, B-cells CNS, kidney, thyroid, Müllerian system
III	PAX3 PAX7	PD OP HD	2q35 1p36	Neural crest, CNS, skeletal muscle Neural crest, CNS, skeletal muscle
IV	PAX4 PAX6	PD HD	7q32 p 3	Pancreas, gut CNS, pancreas, gut, eye

Table I Overview of PAX Transcription Factors

Abbreviations: PD, paired domain; OP, octapeptide; HD, homeodomain; CNS, central nervous system.

differentiated phenotype,³² typically seen in adults. *Pax8* knockout mice show hypoplastic thyroid deprived of the follicular cells and die within 2–3 weeks after weaning.³³ Exogenous thyroid hormone replacement allow female *Pax8* null mice to survive, but they are infertile due to atresia of the uterus, closed vaginal openings and hydrosalpinx.³⁴ In the male reproductive tissues, seminal vesicle and epididymis are PAX8 positive. The same treatment with exogenous thyroxine in male *Pax8* null mice restores the general deficits of CH, but also these mice are infertile due to abnormal development of the efferent ducts, epididymis and complete absence of spermatozoa.³⁵ PAX8 is also involved in the organogenesis of the kidney, but because of co-expression of PAX2, a severe phenotype is obvious only in the double mutant mice.³⁶ Summarizing, in line with its expression pattern, PAX8 is essential for proper development of different body organ systems, especially the thyroid gland, the renal system and the Müllerian system.

Biological Activity of PAX8 in Cancers

It is well known that *PAX* genes are responsible for specifying positional information during embryonic development; in cancers, their over-expression or aberrant expression confers a strong advantage to cancer cell growth, tumor progression and maintenance, but they are not sufficient to initiate tumor.³⁷ Based on this, the PAX8 transcription factor seems to be crucial for the proliferation/survival of thyroid differentiated epithelial cells¹⁶ and thyroid cancer cells.³⁸

Kang et al³⁹ evidenced that transforming growth factor-β1 (TGF-β1) exerts its function through PAX8, regulating normal and pathological processes in thyroid. Moreover, several studies reported that in a subset of human follicular thyroid carcinomas (FTC) the PAX8/PPARγ fusion protein (PPFP), composed of the paired and partial homeobox DNA binding domains of PAX8 fused to the coding exons of PPARγ, caused by a chromosomal translocation, represents a plausible early event in the development or progression of the tumor.^{40–43} The chimeric PPFP, whose expression is under the transcriptional regulation of the *PAX8* promoter,⁴⁴ exerts a dominant-negative effect on wild-type PPARγ and stimulates transcription of PAX8-responsive promoters.⁴³ PAX8 is also expressed in papillary thyroid carcinoma (PTC) and anaplastic thyroid carcinoma (ATC) with ranging positive expression.^{45–47} Since *PAX8* acts as a master gene for thyroid differentiation³² and represents an essential player for function and survival of adult thyroid cells,⁴⁸ it is likely that variations in its expression level may affect the downstream molecular cascades (ie target genes) and in turn crucial pathways in thyroid tumorigenesis.⁴⁹ In addition to the thyroid, the expression of PAX8 in neoplastic tissues has been analyzed in several cancers, including Wilms' tumor, renal carcinomas,^{17,50} well-differentiated pancreatic neuroendocrine tumors,¹⁸ gliomas,¹⁹ endometrial and ovarian carcinomas.^{51–53} According to its expression pattern, PAX8 is involved in numerous pathways that contribute to carcinogenesis. In glioma cell lines, it plays a critical role for maintenance of

telomeres, regulating the expression of telomerase RNA (hTR).¹⁹ Recently, it has been shown that PAX8 plays an oncogenic role in renal cell carcinoma (RCC).⁵⁴ To date, several immunohistochemical analyses denote PAX8 expression in RCC, endometrial, cervical and various epithelial cancers.^{50,55–57} In 2021, numerous and representative studies on PAX8 expression and role in epithelial carcinomas were thoroughly and accurately reviewed by Khizer et al.⁵⁸ Overall, PAX8 emerges as an important factor for cancer cell growth and viability/survival through transcriptional regulation of key factors such as Bcl2,⁵⁹ E2F1,⁶⁰ p53 and p21.⁶¹ However, to improve our knowledge on the role of this transcription factor in malignancies, it will be necessary to define the molecular pathways regulated by PAX8 and the multiprotein complexes it is part of in different contexts.

PAX8 Mechanism of Action in HGSC

In OC, PAX8 was initially used as a histological marker but Bowen et al in 2007⁵² suggested a functional role of this transcription factor in the uncontrolled growth of epithelial ovarian cancer (EOC) and in the physiological development of the oviduct. Few years later, in 2011 Cheung et al¹³ using a genome-wide approach identified PAX8 as one of 54 genes specifically essential for proliferation and viability of ovarian cancer cells. Subsequently, our studies provided strong evidence of PAX8 involvement in OC tumorigenicity in vitro and in vivo.⁶² We demonstrated that PAX8 affects cell growth, migration and invasion of ovarian cancer cells and tumor growth in nude mice. Moreover, we suggested a possible role of PAX8 in the peritoneal dissemination of ovarian cancer cells by modulating the susceptibility to anoikis of cancer cells and their interaction with the extracellular matrix.⁶³ In HGSC PAX8 acts as positive regulator of mutated TP53, that in turns induces p21 to promote proliferative and anti-apoptotic effect.⁶⁴ On the contrary, in endometrial carcinoma mutated p53 and cytoplasmic p21 can independently mediate the pro-proliferative role of PAX8.⁶¹ To better understand PAX8 function and to gain new insights on the transcriptional mechanism(s) regulated by PAX8 leading to OC, RNA-seq analysis before and after PAX8 knockdown were performed in Fallopian tube and ovarian cancer cell lines by several research groups. In 2016, our group compared the transcriptome of normal Fallopian tube secretory epithelial cells (FT-194) with that of ovarian cancer cells (SKOV-3) before and after PAX8 silencing, to identify genes and pathways modified during the transformation process and regulated by this transcription factor.⁶⁵ We highlighted that about 60% of the genes were differentially expressed between FT-194 and SKOV-3 and upon PAX8 silencing; the pathways most affected in both FT194 and SKOV-3 cells were TNFa signaling, inflammatory response, apoptosis, UV response and epithelial mesenchymal transition, estrogen response and p53 pathway. Elias et al⁶⁶ demonstrated that the gene expression changes between Fallopian tube and serous ovarian cancer cells were due to PAX8 cistrome reprogramming. RNA-Seq and ChIP-Seq analysis were used to determine that the genes that were differentially expressed between the cell lines above mentioned were located and clustered around the PAX8 binding sites. Epigenetic remodeling of PAX8 cistrome may be responsible for gene expression alterations leading HGSC from Fallopian tube; in fact, only in tumor the interaction between PAX8 and YAP1, important regulator of Hippo pathway, was observed. The majority of the PAX8 binding sites were enriched in intergenic regions and super-enhancers (SEs); this might explain the low phenotypic effects observed in normal Fallopian tube following PAX8 knockdown, also consistent with the results in the murine tissue.⁶⁷ In tumor context, PAX8 cistrome alterations may enrich the binding sites for PAX8 in intragenic regions and/or SEs, to ensure the presence in the PAX8 network of co-regulators and target genes that encompass differentiation, development and tumorigenesis. Reddy et al⁶⁸ developed the Cancer Core Transcription Factor Specificity (CaCTS) algorithm, to identify tumor-driving master Transcription Factors (MTFs), that is a set of SEs-associated transcription factors whose predicted binding motifs are enriched at SEs, that exhibit tumor type-specific expression. By this approach it has been shown that PAX8, SOX17 and MECOM form a core regulatory circuit driving tumor cell survival in high-grade serous ovarian carcinoma cell lines. Recently, it has been demonstrated that the protein complex PAX8/SOX17 promotes angiogenesis in OC through inhibition of SERPINE1, that is a potent suppressor of VEGF pathway.⁶⁹ Bleu et al⁷⁰ have examined in detail the mechanistic interaction between PAX8 and MECOM. They demonstrated that PAX8 and MECOM are part of the same transcriptional complex and determined a PAX8-MECOM gene signature featuring patients of OC with poor prognosis. Studies based on transcriptional signatures allowed to distinguish epithelial subpopulation during Fallopian tube differentiation and to identify an early secretory cluster enriched in HGSC.⁷¹ PAX8 role in non-HGSC is similar to that in HGSC. Cheung et al¹³

indicated PAX8 as a lineage-specific survival gene, essential for ovarian cancer cells' proliferation. Another study reported that PAX8 knockdown reduces anchorage dependent and independent growth and impairs in vivo tumorigenesis.⁷² Furthermore, ovarian lineage-specific PAX8 regulon (regulatory network) is involved in decisive events that determine cell fate and phenotype during Müllerian duct organogenesis, and if dysregulated aids ovarian carcinogenesis.⁷³ PAX8 also is involved with RB in a reciprocal relation wherein PAX8 regulates *E2F1* promoter and stabilizes RB helping in tumor cell growth.⁶⁰ All studies above mentioned point to PAX8 as an important regulator of signaling pathways in the context of OC. Its ability to associate with different partners in normal versus tumor cells suggests that the PAX8 protein complexes could be targeted specifically in tumor cells, leaving normal cells and tissues completely unaffected.

PAX8 as a Target for Ovarian Cancer Therapy

HGSC is still one of the most lethal malignancies that affect women worldwide because the majority of the cases are diagnosed after intraperitoneal spread, leading to a poor prognosis. Since PAX8 is expressed in Fallopian tube secretory cells and is retained in HGSC and mediates several tumorigenic effects in serous carcinoma, it can be considered a promising drug target. As already mentioned, the altered PAX8 cistrome in HGSC offers the opportunity to target PAX8 pharmacologically only in the transformed tissue. Recently, it has been demonstrated that PAX8 regulon in OC is susceptible to HDAC inhibitors.⁷³ Briefly, operative PAX8 regulon composed by 9 genes is epigenetically disabled by HDAC inhibitors, resulting in downregulation of PAX8 and its entire responsive network and suppression of ovarian cancer growth. In the same year, Hardy et al⁷⁴ demonstrated that PAX8 targeting either through CRISPR genomic alteration or by drug treatment with thiostrepton leads to a reduction of tumor growth and metastasis. Nowadays, it is clear that strategies aimed at reducing PAX8 levels or inhibiting binding to crucial interacting proteins may impair tumor progression and modulate the aggressive properties of cancer cells. As already cited, SOX17 functions as driving-tumor MTF together with PAX8⁶⁸; along the same line Lin et al⁷⁵ showed that both transcription factors display Müllerian lineage-specific co-expression and co-regulate a broad range of downstream genes, including those involved in cell cycle and tissue morphogenesis. Moreover, they uncover that transcriptional CDK inhibitors efficiently reduce SOX17 and PAX8 protein levels and also putative PAX8 regulon genes was appreciably abolished. These orally bioavailable CDK inhibitors are able to efficiently suppress tumorigenesis in vivo, in xenograft models. Another strategy employed to target PAX factors is to inhibit their binding activity. Grimley et al⁷⁶ identified among 227 potential PAX2 inhibitors a small molecule, termed EG1 that effectively blocked PAX2-5-8 DNA binding activity and decreased the viability of cancer cells. Recently, the same research group used an unbiased, cell-based, high-throughput screen to isolate compounds that inhibit PAX2-5-8 transactivation ability.⁷⁷ These small molecules, BG, do not interfere with PAX2-5-8 DNA binding activity, whereas exert inhibitory effect on PAX2-5-8 transcription activation and consequently on cancer cells proliferation. Given that EG1 and BG compounds exert their action through two different mechanisms, the authors suggest the possibility that the combining of both molecules may add therapeutic benefit in the future.

Conclusion

Epithelial ovarian cancers, shown to develop from both OSE and FTE, are the most common and aggressive form of OC. About 80% of these are attributed to HGSC, that is the deadliest histological subtype of EOC. This is due to the fact that at an early-stage HGSC is asymptomatic and specific biomarkers still do not exist, consequently the majority of the cases are detected only at an advanced stage often with distant metastases (FIGO stage III–IV). The primary line of therapy includes surgical debulking followed by taxanes and platinum-based chemotherapies. HGSCs are initially responsive to drugs treatment, but usually very soon become chemoresistant and highly metastatic with subsequent patient relapse. For many years, the ovary was considered the site of origin of HGSC development; however, this theory fails to explain the presence of STICs in 60% of the patients with HGSC.⁷⁸ Non-invasive STICs derived by secretory Fallopian tube epithelial cells that undergo *p53* mutations acquire invasive malignant behavior by accumulating genomic instability and being exposed to paracrine factors of the ovarian microenvironment reach the ovary and peritoneum with all the signatures of HGSC.

It is relevant to underline that PAX8 is expressed in normal Fallopian tube secretory cells and is preserved through every stage of transformation until HGSC. Such retention is one of the evidences supporting the tubal origin of HGSC,⁹ thus investigating PAX8 function in normal Fallopian tube cells could shed light on its role in tumor progression and provide critical clues in the understanding of its exact niche in ovarian cancer development. Unpublished data from our laboratory indicate that PAX8 silencing affects stemness in 3D spheroids of Fallopian tube cell line, suggesting a critical role of PAX8 in the regulation of the secretory cells multipotency.

Given its cell-type and tissue type-specificity, limited expression in healthy adults and the dependency of OC to its expression and activity, PAX8 is a very promising diagnostic and therapeutic target for HGSC. It is known that PAX8 is associated with secretory pathways in thyroid and it is most likely that it has a similar role in the Müllerian tract, as recently described for SERPINE1.⁷⁹ Hence, it is reasonable to suppose that its putative targets could have an important role as secretory factors and serve both as suitable biomarkers for the early diagnosis of HGSC and as useful therapeutic target against this malignancy. In parallel, the early evidences on PAX8 involvement in the metabolic reprogramming^{54,61} could indicate new candidates in terms of novel targets and predictive biomarkers. In conclusion, we think that the combined therapeutic approaches of PAX inhibitors with other chemotherapeutic drugs may enhance their efficacy and in particular improve the survival of patients with a worse prognosis.

Disclosure

The authors report no conflicts of interest in this work.

References

- 1. Saad AF, Hu W, Sood AK. Microenvironment and pathogenesis of epithelial ovarian cancer. *Horm Cancer*. 2010;1:277–290. doi:10.1007/s12672-010-0054-2
- Chen VW, Ruiz B, Killeen JL, Cotè TR, Wu XC, Correa CN. Pathology and classification of ovarian tumors. *Cancer*. 2003;97(10 Suppl):2631–2642. doi:10.1002/cncr.11345
- 3. Gilks CB. Molecular abnormalities in ovarian cancer subtypes other than high-grade serous carcinoma. J Oncol. 2010;2010:740968. doi:10.1155/2010/740968
- 4. Press JZ, De Luca A, Boyd N, et al. Ovarian carcinomas with genetic and epigenetic BRCA1 loss have distinct molecular abnormalities. *BMC Cancer*. 2008;22(8):17. doi:10.1186/1471-2407-8-17
- 5. Shaw PA, McLaughlin JR, Zweemer RP, et al. Histopathologic features of genetically determined ovarian cancer. Int J Gynecol Pathol. 2002;21:407-411. doi:10.1097/00004347-200210000-00011
- 6. Nik NN, Vang R, Shih M, Kurman RJ. Origin and pathogenesis of pelvic (ovarian, tubal, and primary peritoneal) serous carcinoma. *Annu Rev Pathol.* 2014;9:27–45. doi:10.1146/annurev-pathol-020712-163949
- 7. Kurman RJ, Shihle M. The origin and pathogenesis of epithelial ovarian cancer: a proposed unifying theory. *Am J Surg Pathol.* 2010;34 (3):433–443. doi:10.1097/PAS.0b013e3181cf3d79.
- Kim J, Coffey DM, Creighton CJ, Yu Z, Hawkins SM, Matzuk MM. High-grade serous ovarian cancer arises from fallopian tube in a mouse model. Proc Natl Acad Sci USA. 2012;109:3921–3926. doi:10.1073/pnas.1117135109
- 9. Perets R, Wyant GA, Muto KW, et al. Transformation of the fallopian tube secretory epithelium leads to high-grade serous ovarian cancer in Brca; Tp53;Pten models. *Cancer Cell*. 2013;24(6):751–765. doi:10.1016/j.ccr.2013.10.013.
- 10. Beirne JP, McArt DG, Roddy A, et al. Defining the molecular evolution of extrauterine high grade serous carcinoma. *Gynecol Oncol.* 2019;155 (2):305–317. doi:10.1016/j.ygyno.2019.08.029
- 11. Merritt MA, Bentink S, Schwede M, et al. Gene expression signature of normal cell-of-origin predicts ovarian tumor outcomes. *PLoS One*. 2013;8 (11):e80314. doi:10.1371/journal.pone.0080314
- 12. Xiang L, Rong G, Zhao J, Wang Z, Shi F. Identification of candidate genes associated with tubal origin of high-grade serous ovarian cancer. *Oncol Lett.* 2018;15(5):7769–7775. doi:10.3892/ol.2018.8346
- Cheung HW, Cowley GS, Weir BA, et al. Systematic investigation of genetic vulnerabilities across cancer cell lines reveals lineage-specific dependencies in ovarian cancer. Proc Natl Acad Sci U S A. 2011;108:12372–12377. doi:10.1073/pnas.1109363108
- 14. Bouchard M, Souabni A, Mandler M, Neubuser A, Busslinger M. Nephric lineage specification by Pax2 and Pax8. Genes Dev. 2002;16 (22):2958-2970. doi:10.1101/gad.240102
- 15. Plachov D, Chowdhury K, Walther C, Simon D, Guenet JL, Gruss P. Pax8, a murine paired box gene expressed in the developing excretory system and thyroid gland. *Development*. 1990;110(2):643–651. doi:10.1242/dev.110.2.643
- Di Palma T, Filippone MG, Pierantoni GM, Fusco A, Soddu S, Zannini M. Pax8 has a critical role in epithelial cell survival and proliferation. *Cell Death Dis.* 2013;4(7):e729. doi:10.1038/cddis.2013.262
- 17. Poleev A, Fickenscher H, Mundlos S, et al. PAX8, a human paired box gene: isolation and expression in developing thyroid, kidney and Wilms' tumors. *Development*. 1992;116(3):611–623. doi:10.1242/dev.116.3.611
- Sangoi AR, Ohgami RS, Pai RK, Beck AH, McKenney JK, Pai RK. PAX8 expression reliably distinguishes pancreatic well-differentiated neuroendocrine tumors from ileal and pulmonary well-differentiated neuroendocrine tumors and pancreatic acinar cell carcinoma. *Mod Pathol.* 2011;24(3):412–424. doi:10.1038/modpathol.2010.176

- Chen YJ, Campbell HG, Wiles AK, et al. PAX8 regulates telomerase reverse transcriptase and telomerase RNA component in glioma. *Cancer Res.* 2008;68(14):5724–5732. doi:10.1158/0008-5472.CAN-08-0058
- Ozcan A, de la Roza G, Ro JY, Shen SS, Truong LD. PAX2 and PAX8 expression in primary and metastatic renal tumors: a comprehensive comparison. Arch Pathol Lab Med. 2012;136(12):1541–1551. doi:10.5858/arpa.2012-0072-OA
- Hardy LR, Salvi A, Burdette JE. UnPAXing the divergent roles of PAX2 and PAX8 in high-grade serous ovarian cancer. Cancers. 2018;10(8):262. doi:10.3390/cancers10080262
- 22. Nonaka D, Chiriboga L, Soslow RA. Expression of pax8 as a useful marker in distinguishing ovarian carcinomas from mammary carcinomas. *Am J Surg Pathol.* 2008;32(10):1566–1571. doi:10.1097/PAS.0b013e31816d71ad
- Laury AR, Hornick JL, Perets R, et al. PAX8 reliably distinguishes ovarian serous tumors from malignant mesothelioma. Am J Surg Pathol. 2010;34(5):627–635. doi:10.1097/PAS.0b013e3181da7687
- 24. Ye J, Hameed O, Findeis-Hosey JJ, et al. Diagnostic utility of PAX8, TTF-1 and napsin A for discriminating metastatic carcinoma from primary adenocarcinoma of the lung. *Biotech Histochem*. 2012;87(1):30–34. doi:10.3109/10520295.2011.591838
- 25. Dahl E, Koseki H, Balling R. Pax genes and organogenesis. Bioessays. 1997;19(9):755-765. doi:10.1002/bies.950190905
- 26. Mansouri A, Hallonet M, Gruss P. Pax genes and their roles in cell differentiation and development. *Current Opinion Cell Biol.* 1996;8(6):851–857. doi:10.1016/s0955-0674(96)80087-1
- 27. Poleev A, Wendler F, Fickenscher H, et al. Distinct functional properties of three human paired-box-protein, PAX8, isoforms generated by alternative splicing in thyroid, kidney and Wilms' tumors. *European J of Biochem*. 1995;228(3):899–911. doi:10.1111/j.1432-1033.1995.tb20338.x
- Kozmik Z, Kurzbauer R, Dörfler P, Busslinger M. Alternative splicing of Pax-8 gene transcripts is developmentally regulated and generates isoforms with different transactivation properties. *Mol Cell Biol.* 1993;13(10):6024–6035. doi:10.1128/mcb.13.10.6024-6035.1993
- 29. Czerny T, Schaffner G, Busslinger M. DNA sequence recognition by Pax proteins: bipartite structure of the paired domain and its binding site. *Genes Dev.* 1993;7(10):2048–2061. doi:10.1101/gad.7.10.2048
- 30. De Felice M, Di Lauro R. Thyroid development and its disorders: genetics and molecular mechanisms. *Endocrine Rev.* 2004;25(5):722–746. doi:10.1210/er.2003-0028
- 31. De Felice M, Di Lauro R. Minireview: intrinsic and extrinsic factors in thyroid gland development: an update. *Endocrinology*. 2011;152 (8):2948–2956. doi:10.1210/en.2011-0204
- 32. Pasca Di Magliano M, Di Lauro R, Zannini M. Pax8 has a key role in thyroid cell differentiation. Proc Natl Acad Sci U S A. 2000;97 (24):13144–13149. doi:10.1073/pnas.240336397
- 33. Mansouri A, Chowdhury K, Gruss P. Follicular cells of the thyroid gland require Pax8 gene function. Nat Genet. 1998;19(1):87–90. doi:10.1038/ ng0598-87
- 34. Mittag J, Winterhager E, Bauer K, Grümmer R. Congenital hypothyroid female pax8-deficient mice are infertile despite thyroid hormone replacement therapy. *Endocrinology*. 2007;148(2):719–725. doi:10.1210/en.2006-1054
- 35. Wistuba J, Mittag J, Luetjens CM, et al. Male congenital hypothyroid Pax8-/- mice are infertile despite adequate treatment with thyroid hormone. *J Endocrinol.* 2007;192(1):99–109. doi:10.1677/JOE-06-0054
- Narlis M, Grote D, Gaitan Y, Boualia SK, Bouchard M. Pax2 and pax8 regulate branching morphogenesis and nephron differentiation in the developing kidney. J Am Soc Nephrol. 2007;18(4):1121–1129. doi:10.1681/ASN.2006070739
- Muratovska A, Zhou C, He S, Goodyer P, Eccles MR. Paired-box genes are frequently expressed in cancer and often required for cancer cell survival. Oncogene. 2003;22(39):7989–7997. doi:10.1038/sj.onc.1206766
- Credendino SC, Bellone ML, Lewin N, et al. A ceRNA circuitry involving the long noncoding RNA Klhl14-AS, Pax8, and Bcl2 drives thyroid carcinogenesis. *Cancer Res.* 2019;79(22):5746–5757. doi:10.1158/0008-5472.CAN-19-0039
- 39. Kang HC, Ohmori M, Harii N, Endo T, Onaya T. Pax-8 is essential for regulation of the thyroglobulin gene by transforming growth factor-β1. Endocrinology. 2001;142(1):267–275. doi:10.1210/endo.142.1.7918
- Eberhardt NL, Grebe SK, McIver B, Reddi HV. The role of the PAX8/PPARgamma fusion oncogene in the pathogenesis of follicular thyroid cancer. *Mol Cell Endocrinol.* 2010;321(1):50–56. doi:10.1016/j.mce.2009.10.013
- Marques AR, Espadinha C, Catarino AL, et al. Expression of PAX8-PPAR gamma 1 rearrangements in both follicular thyroid carcinomas and adenomas. J Clin Endocrinol Metab. 2002;87(8):3947–3952. doi:10.1210/jcem.87.8.8756
- 42. Raman P, Koenig RJ. Pax-8-PPAR-γ fusion protein in thyroid carcinoma. Nat Rev Endocrinol. 2014;10(10):616–623. doi:10.1038/nrendo.2014.115
- 43. Placzkowski KA, Reddi HV, Grebe SK, Eberhardt NL, McIver B. The role of the PAX8/PPARgamma fusion oncogene in thyroid cancer. *PPAR Res.* 2008;2008:672829. doi:10.1155/2008/672829
- 44. Kroll TG, Sarraf P, Pecciarini L, et al. PAX8-PPARgamma1 fusion oncogene in human thyroid carcinoma. Science. 2000;289(5483):1357–1360. doi:10.1126/science.289.5483.1357
- 45. Nonaka D, Tang Y, Chiriboga L, Rivera M, Ghossein R. Diagnostic utility of thyroid transcription factors Pax8 and TTF-2 (FoxE1) in thyroid epithelial neoplasms. *Mod Pathol*. 2008;21(2):192–200. doi:10.1038/modpathol.3801002
- 46. Puglisi F, Cesselli D, Damante G, Pellizzari L, Beltrami CA, Di Loreto C. Expression of Pax-8, p53 and bcl-2 in human benign and malignant thyroid diseases. *Anticancer Res.* 2000;20(1A):311–316.
- 47. Bishop JA, Sharma R, Westra WH. PAX8 immunostaining of anaplastic thyroid carcinoma: a reliable means of discerning thyroid origin for undifferentiated tumors of the head and neck. *Hum Pathol.* 2011;42:1873–1877. doi:10.1016/j.humpath.2011.02.004
- Marotta P, Amendola E, Scarfò M, et al. The paired box transcription factor Pax8 is essential for function and survival of adult thyroid cells. *Mol Cell Endocrinol.* 2014;396(1–2):26–36. doi:10.1016/j.mce.2014.08.004
- Rosignolo F, Sponziello M, Durante C, et al. Expression of PAX8 target genes in papillary thyroid carcinoma. PLoS One. 2016;11(6):e0156658. doi:10.1371/journal.pone.0156658
- 50. Tacha D, Zhou D, Cheng L. Expression of PAX8 in Normal and Neoplastic Tissues A Comprehensive Immunohistochemical Study. *Appl Immunohistochem Mol Morphol.* 2011;19(4):293–299. doi:10.1097/PAI.0b013e3182025f66
- 51. Yemelyanova A, Gown AM, Wu LS, Holmes BJ, Ronnett BM, Vang R. PAX8 expression in uterine adenocarcinomas and mesonephric proliferations. *Int J Gynecol Pathol*. 2014;33(5):492–499. doi:10.1097/PGP.0b013e3182a54afa
- 52. Bowen NJ, Logani S, Dickerson EB, et al. Emerging roles for PAX8 in ovarian cancer and endosalpingeal development. *Gynecol Oncol*. 2007;104 (2):331–337. doi:10.1016/j.ygyno.2006.08.052

- 53. Chai HJ, Ren Q, Fan Q, et al. PAX8 is a potential marker for the diagnosis of primary epithelial ovarian cancer. *Oncol Lett.* 2017;14(5):5871–5875. doi:10.3892/ol.2017.6949
- 54. Bleu M, Gaulis S, Lopes R, et al. PAX8 activates metabolic genes via enhancer elements in renal cell carcinoma. *Nat Commun.* 2019;10(1):3739. doi:10.1038/s41467-019-11672-1
- 55. Barr ML, Jilaveanu LB, Camp RL, Adeniran AJ, Kluger HM, Shuch B. PAX-8 expression in renal tumours and distant sites: a useful marker of primary and metastatic renal cell carcinoma? J Clin Pathol. 2015;68(1):12–17. doi:10.1136/jclinpath-2014-202259
- 56. Ozcan A, Shen SS, Hamilton C, et al. PAX 8 expression in non-neoplastic tissues, primary tumors, and metastatic tumors: a comprehensive immunohistochemical study. *Mod Pathol*. 2011;24(6):751–764. doi:10.1038/modpathol.2011.3
- 57. Laury AR, Perets R, Piao H, et al. A comprehensive analysis of PAX8 expression in human epithelial tumors. *Am J Surg Pathol.* 2011;35 (6):816–826. doi:10.1097/PAS.0b013e318216c112
- 58. Khizer K, Padda J, Khedr A, et al. Paired-box gene 8 (PAX8) and its association with epithelial carcinomas. Cureus. 2021;13(8):e17208. doi:10.7759/cureus.17208
- Hewitt SM, Hamada S, Monarres A, Kottical LV, Saunders GF, McDonnell TJ. Transcriptional activation of the bcl-2 apoptosis suppressor gene by the paired box transcription factor PAX8. *Anticancer Res.* 1997;17(5A):3211–3215.
- 60. Li CG, Nyman JE, Braithwaite AW, Eccles MR. PAX8 promotes tumor cell growth by transcriptionally regulating E2F1 and stabilizing RB protein. Oncogene. 2011;30(48):4824–4834. doi:10.1038/onc.2011.190
- 61. Fares B, Berger L, Bangiev-Girsh E, et al. PAX8 plays an essential antiapoptotic role in uterine serous papillary cancer. *Oncogene*. 2021;40 (34):5275–5285. doi:10.1038/s41388-021-01925-z
- 62. Di Palma T, Lucci V, de Cristofaro T, Filippone MG, Zannini M. A role for PAX8 in the tumorigenic phenotype of ovarian cancer cells. *BMC Cancer*. 2014;14:292. doi:10.1186/1471-2407-14-292
- 63. Soriano AA, de Cristofaro T, Di Palma T, et al. PAX8 expression in high-grade serous ovarian cancer positively regulates attachment to ECM via Integrin β3. Cancer Cell Int. 2019;19:303. doi:10.1186/s12935-019-1022-8
- 64. Ghannam-Shahbari D, Jacob E, Kakun RR, et al. PAX8 activates a p53-p21-dependent pro-proliferative effect in high grade serous ovarian carcinoma. *Oncogene*. 2018;37(17):2213-2224. doi:10.1038/s41388-017-0040-z
- 65. de Cristofaro T, Di Palma T, Soriano AA, et al. Candidate genes and pathways downstream of PAX8 involved in ovarian high-grade serous carcinoma. *Oncotarget*. 2016;7(27):41929–41947. doi:10.18632/oncotarget.9740
- 66. Elias KM, Emori MM, Westerling T, et al. Epigenetic remodeling regulates transcriptional changes between ovarian cancer and benign precursors. JCI Insight. 2016;1(13):e87988. doi:10.1172/jci.insight.87988
- 67. Rodgers LH. Loss of PAX8 in high-grade serous ovarian cancer reduces cell survival despite unique modes of action in the fallopian tube and ovarian surface epithelium. *Oncotarget*. 2016;7(22):32785–32795. doi:10.18632/oncotarget.9051
- Reddy J, Fonseca MAS, Corona RI, et al. Predicting master transcription factors from pan-cancer expression data. Sci Adv. 2021;7(48):eabf6123. doi:10.1126/sciadv.abf6123
- Chaves-Moreira D, Mitchell MA, Arruza C, et al. The transcription factor PAX8 promotes angiogenesis in ovarian cancer through interaction with SOX17. Sci Signal. 2022;15(728):eabm249. doi:10.1126/scisignal.abm2496
- 70. Bleu M, Mermet-Meillon F, Apfel V, et al. PAX8 and MECOM are interaction partners driving ovarian cancer. *Nat Commun.* 2021;12(1):2442. doi:10.1038/s41467-021-22708-w
- Dinh HQ, Lin X, Abbasi F, et al. Single-cell transcriptomics identifies gene expression networks driving differentiation and tumorigenesis in the human fallopian tube. Cell Rep. 2021;35(2):108978. doi:10.1016/j.celrep.2021.108978
- Adler EK, Corona RI, Lee JM, et al. The PAX8 cistrome in epithelial ovarian cancer. Oncotarget. 2017;8(65):108316–108332. doi:10.18632/ oncotarget.22718
- 73. Shi K, Yin X, Cai MC, et al. PAX8 regulon in human ovarian cancer links lineage dependency with epigenetic vulnerability to HDAC inhibitors. *Elife.* 2019;8:e44306. doi:10.7554/eLife.44306
- 74. Hardy LR, Pergande MR, Esparza K, et al. Proteomic analysis reveals a role for PAX8 in peritoneal colonization of high grade serous ovarian cancer that can be targeted with micelle encapsulated thiostrepton. *Oncogene*. 2019;38(32):6003–6016. doi:10.1038/s41388-019-0842-2
- 75. Lin L, Shi K, Zhou S, et al. SOX17 and PAX8 constitute an actionable lineage-survival transcriptional complex in ovarian cancer. *Oncogene*. 2022;41(12):1767–1779. doi:10.1038/s41388-022-02210-3
- 76. Grimley E, Liao C, Ranghini EJ, Nikolovska-Coleska Z, Dressler GR. Inhibition of Pax2 Transcription Activation with a Small Molecule that Targets the DNA Binding Domain. ACS Chem Biol. 2017;12(3):724–734. doi:10.1021/acschembio.6b00782
- 77. Bradford STJ, Grimley E, Laszczyk AM, Lee PH, Patel SR, Dressler GR. Identification of Pax protein inhibitors that suppress target gene expression and cancer cell proliferation. Cell Chem Biol. 2022;29(3):412–422.e4. doi:10.1016/j.chembiol.2021.11.003
- 78. Chen F, Gaitskell K, Garcia MJ, Albukhari A, Tsaltas J, Ahmed AA. Serous tubal intraepithelial carcinomas associated with high-grade serous ovarian carcinomas: a systematic review. *BJOG*. 2017;124(6):872–878. doi:10.1111/1471-0528.14543
- 79. Chaves-Moreira D, Mitchell MA, Arruza C, et al. The transcription factor PAX8 promotes angiogenesis in ovarian cancer through interaction with SOX17. *Sci Signal*. 2022;15(728):eabm2496. doi:10.1126/scisignal.abm2496

OncoTargets and Therapy

Dovepress

Publish your work in this journal

OncoTargets and Therapy is an international, peer-reviewed, open access journal focusing on the pathological basis of all cancers, potential targets for therapy and treatment protocols employed to improve the management of cancer patients. The journal also focuses on the impact of management programs and new therapeutic agents and protocols on patient perspectives such as quality of life, adherence and satisfaction. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit http://www.dovepress.com/testimonials.php to read real quotes from published authors.

Submit your manuscript here: https://www.dovepress.com/oncotargets-and-therapy-journal