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Exome sequencing of Filaggrin and related genes in African-American children with atopic dermatitis

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Letter

Atopic dermatitis (AD) is a common chronic relapsing disease. There is a considerable body of evidence supporting a genetic basis for AD (Bussman *et al.*, 2011;Ellinghaus *et al.*, 2013). Mutations in the Filaggrin (*FLG*) gene have been consistently found to be associated with AD in people of European and Asian ancestry (Brown and McLean, 2012). More than 40 *FLG* loss-of-function mutations have been described in Europeans and Asians, (Brown and McLean, 2012).However, *FLG* loss-of-function mutations have not commonly been found in Africans or African-Americans (Margolis *et al.*, 2012;Brown and McLean, 2012;Winge *et al.*, 2011a). Loss-of-function mutations in exon 3 of *FLG* result in diminished or absent filaggrin protein, most often due to a premature stop codon or a frameshift mutation resulting in a stop codon further downstream. Interestingly, the absence of profilaggrin protein (precursor of filaggrin) has also been noted in keratohyalin granules in the majority of those with ichthyosis vulgaris (IV) of European and Asian ancestry (Perusquia-Ortiz.A.M. *et al.*, 2013;Thyssen *et al.*, 2013;Fleckman and Brumbaugh, 2002).

FLG is located on chromosome 1q21 in a region called the epidermal differentiation complex (EDC). It is part of a family of genes that code for S100-fused like proteins (SFTP). The SFTPs include the proteins profilaggrin (coded by *FLG*), hornerin (*HRNR*),

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filaggrin-2 (*FLG2*), repetin (*RPTN*), cornulin (*CRNN*), trichohyalin (*TCHH*), and trichohyalin-like 1(*TCHHL1*)(Henry *et al.*, 2012). These genes are very similar to one another with respect to structure and function, and lie in close proximity to each other in the EDC (Marenholz *et al.*, 2011;Henry *et al.*, 2012;Pellerin *et al.*, 2013). Based on prior experience with *FLG*, it has been hypothesized that a stop-gain (null) mutation in exon 3 of any of the SFTP genes will result in decreased or absent protein production (Henry *et al.*, 2012;Marenholz *et al.*, 2011;Margolis *et al.*, 2014).

The goal of this study was to identify stop-gain variants in FLG and closely related genes in African-Americans with AD from the Pediatric Eczema Elective Registry (PEER)(Margolis et al., 2012). From this cohort we randomly selected 60 subjects for whole exome sequencing to ensure sufficient power to detect variants with a minor allelic frequency (MAF) of greater than 3%. Sequencing was performed by Ambry Genetics (Aliso Viejo, California) using whole exome targeted enrichment by Agilent SureSelectXT Human All Exon 50Mb kit. Quality assessment revealed that most samples were above 50% on target and mean coverage per gene was excellent. The libraries were indexed using 100 base paired ends and processed using Illumina HiSeq2000 at 100x coverage per exon. Data was assessed utilizing a pipeline generated at the University of Pennsylvania based on the best practices protocol from the Broad Institute (Cambridge, MA). This report focused on stop-gain mutations of exon 3 (i.e., loss-of-function mutations) in the SFTP genes because of their likely functional relevance (Brown and McLean, 2012;Henry et al., 2012;Marenholz et al., 2011). Taqman allelic discrimination assays were created for any newly identified FLG lossof-function mutations, which were then used to genotype an additional random sample of 100 African-American PEER children.

Sequencing of the SFTP genes in 60 self-reported (ancestry previously confirmed with ancestral informative markers(Margolis *et al.*, 2012)) African-American children with AD revealed a total of 289 variants in *FLG*, 107 variants in *FLG2*, 339 variants in *HRNR*, 4 variants in *RPTN*, 37 variants in *CRNN*, 88 variants in *TCHH* and 14 variants in *TCHHL1*. However, very few variants resulted in a premature stop codon in exon 3 (Table 1). Each of the three newly identified *FLG* stop-gain mutations, Q570X, R3409X and S3707X, were observed only once. S2392X and S2377X in *FLG2* were noted 1 and 16 times respectively. In *TCHHL1*, the variant Q294X was noted twice. All subjects were heterozygous for the mutations. The MAF for variants noted once, twice and 16 times were 0.008, 0.017 and 0.133, respectively. Next, we used Taqman based allelic discrimination assays to evaluate the three *FLG* mutations, Q570X, R3409X, and S3707X, in an additional 100 African-American PEER children. However, none of these variants could be detected in other members of our cohort

This report is from the largest whole exome sequencing study of African-Americans with AD performed to date. Here we have reported results specific to the SFTP genes. We identified a few new null mutations, albeit the ones in *FLG* had low MAFs. The MAFs noted for S2377X (*FLG2*) and Q294X (*TCHHL1*) vary from the healthy subjects in the 1000 Genomes database (The 1000 Genomes Project Consortium, 2012) (Table 1). S2377X was seen about half as frequently in our cohort as compared to the healthy 1000 Genomes African population; while Q294X, though more common in our cohort, still had a low MAF

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Our findings are in agreement with that of Winge et al., who also failed to detect common *FLG* loss-of-function mutations in people of African ancestry with AD (Winge *et al.*, 2011a). Our study does have limitations in that we focused only on exon 3 stop-gain mutations in genes. We did not assess copy number variations. We also did not assay protein function. Another point to be noted is that since most African-Americans have their origins in West Africa; our findings may not generalize to everyone with African ancestry. However, based on the experience of others as well as our study, which is the largest whole exome study of African-Americans with AD, it seems unlikely that *FLG* stop-gain mutations have a prominent role with respect to **incident** AD in African-American children(Thaswer-Esmail *et al.*, 2014;Winge *et al.*, 2011b).

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References

- 1. Brown SJ, McLean WH. One remarkable molecule: filaggrin. Journal of Investigative Dermatology. 2012; 132:751–762. [PubMed: 22158554]
- Bussman C, Weidinger S, Novak N. Genetics of atopic dermatitis. Journal of German Society of Dermatology. 2011; 9:670–679.
- 3. Ellinghaus D, Baurecht H, Esparza-Gordillo J, Rodriguez E, Matanovic A, Marenholz I, Hubner N, Schaarschmidt H, Novak N, Michel S, Maintz L, Werfel T, Meyer-Hoffert U, Hotze M, Prokish H, Heim K, Herder C, Hirota T, Tamari M, Kubo M, Takahashi A, Nakamura Y, Tsoi LC, Stuart P, Elder JT, Sun L, Zuo X, Yang S, Zhang X, Hoffman P, Nothen MM, Folster-Holst R, Winkelmann J, Illig T, Boehm BO, Duerr RH, Buning C, Brand S, Glas J, McAleer MA, Fahy CM, Kabesch M, Brown S, McLean WH, Irvine AD, Schreiber S, Lee YA, Franke A, Weidinger S. High-density genotyping study indentifies four new susceptibility loci for atopic dermatitis. Nature Genetics. 2013; 45:808–812. [PubMed: 23727859]
- Fleckman P, Brumbaugh S. Absence of granular layer and keratohylin define a morphologically distinct subset of individuals with ichthyosis vulgaris. Experimental Dermatology. 2002; 11:327– 336. [PubMed: 12190941]
- Henry J, Toulza E, Hsu CY, Pellerin L, Balica S, Mazereeuw-Hautier J, Paul C, Serre G, Jonca N, Simon M. Update on the epidermal differentiation complex. Frontiers in Bioscience. 2012; 17:1517–1532.
- 6. Marenholz I, Rivera VA, Esparza-Gordillo J, Bauerfeind A, Lee-Kirsch MA, Ciechanowicz A, Kurek M, Piskackova T, Macek M, Lee YA. Association screening in the Epidermal Differentiation Complex (EDC) identifies an SPRR3 repeat number variant as a risk factor for eczema. Journal of Investigative Dermatology. 2011; 131:1644–1649. [PubMed: 21490620]
- Margolis DJ, Apter AJ, Gupta J, Hoffstad O, Papadopoulos M, Campbell LE, Sandilands A, McLean WHI, Rebbeck TR, Mitra N. The persistence of atopic dermatitis and Filaggrin mutations in a US longitudinal cohort. Journal of Allergy & Clinical Immunology. 2012; 130:912–917. [PubMed: 22951058]
- Margolis DJ, Gupta J, Apter AJ, Ganguly T, Hoffstad O, Papadopoulos M, Rebbeck TR, Mitra N. Filaggrin-2 variation is associated with more persistent atopic dermatitis in Arican Americans subjects. Journal of Allergy & Clinical Immunology. 2014 on line.

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- Pellerin L, Henry J, Hsu CY, Balica S, Jean-Decoster C, Mechin MC, Hansmann B, Rodriguez E, Weidinger S, Schmitt AM, Serre G, Paul C, Simon M. Defects in filaggrin-like proteins in both lesional and nonlesional atopic skin. Journal of Allergy & Clinical Immunology. 2013; 131:1094– 1102. [PubMed: 23403047]
- Perusquia-Ortiz AM, Oji V, Sauerland MC, Tarinski T, Zaraeva I, Sellar N, Meteze D, Aufenvenne K, Hausser I, Traupe H. Complete filaggrin deficiency in ichthyosis vulgaris is associated with only moderate changes in epidermal permeability barrier function profile. JEADV. 2013; 27:1552–1558. [PubMed: 23297869]
- Thaswer-Esmail F, Jakasa I, Todd G, Wen Y, Brown SJ, Krobach K, Campbell LE, O'Regan GM, McLean WHI, Irvine AD, Kezic S, Sandilands A. South African amaXhosa patient with atopic dermatitis have decreased levels of filaggrin breakdown products but no loss-of-function mutations in filaggrin. Journal of Allergy & Clinical Immunology. 2014; 133:280–282. [PubMed: 24369804]
- The 1000 Genomes Project Consortium. An integrated map of genetic variation from 1,092 human genomes. Nature. 2012; 491:56–65. [PubMed: 23128226]
- Thyssen JP, Godoy-Gijon E, Elias PM. Ichthyosis vulgaris: the filaggrin mutation disease. British Journal of Dermatology. 2013; 168:155–158.
- Winge MC, Bilcha KD, Lieden A, Shibeshi D, Sandilands A, Wahlgren CF, McLean WH, Nordenskjold M, Bradley M. Novel filaggrin mutation but no other loss-of-function variants found in Ethiopian patients with atopic dermatitis. British Journal of Dermatology. 2011a; 165:1074– 1080. [PubMed: 21692775]
- 15. Winge MC, Bilcha KD, Lieden A, Shibeshi D, Sandilands A, Wahlgren CF, McLean WH, Nordenskjold M, Bradley M, Winge MCG, Bilcha KD, Lieden A, Shibeshi D, Sandilands A, Wahlgren CF, McLean WHI, Nordenskjold M, Bradley M. Novel filaggrin mutation but no other loss-of-function variants found in Ethiopian patients with atopic dermatitis. British Journal of Dermatology. 2011b; 165:1074–1080. [PubMed: 21692775]

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Table 1

Null mutations identified in genes belonging to S100-fused like protein family.

| Gene | Variant | Variant Reference Allele Alternat | - e | allele Location of amino acid change | MAF Current study | MAF Current study MAF African ancestry (Source: 1000 Genomes) | dbSNP Designation |
|--------|-------------------------|-----------------------------------|-----|--|-------------------|--|-------------------|
| FLG | Stop-gain | G | А | FLG:NM_002016:exon3:c.1708C>T:p.Q570X | 0.008 | - | - |
| FLG | Stop-gain | G | Υ | FLG:NM_002016:exon3:c.10225C>T:p.R3409X | 0.008 | - | I |
| FLG | Stop-gain | G | Т | FLG:NM_002016:exon3:11120C>A:p.S3707X | 0.008 | - | I |
| FLG2 | Stop-gain | G | Т | FLG2:NM_001014342:exon3:c.7130C>A:p.S2377X | 0.133 | 0.29 | rs12568784 |
| FLG2 | Stop-loss | С | G | FLG2:NM_001014342:exon3:c.7175G>C:p.S2392X | 0.008 | 0.01 | rs150529054 |
| TCHHLI | TCHHLI Stop-gain | G | V | TCHHL1:NM_001008536:exon3:c.880C>T:p.Q294X | 0.017 | 0.01 | rs61749316 |
| | | | | | | | |

MAF= Minor allele frequency