### MITOGENOME ANNOUNCEMENT

OPEN ACCESS Check for updates

Taylor & Francis

Taylor & Francis Group

# The complete mitochondrial genome of *Aconitum kusnezoffii* Rchb. (Ranales, Ranunculaceae)

Sheng-Nan Li<sup>a</sup>\*, Yan-Yun Yang<sup>a</sup>\*, Liang Xu<sup>a</sup>, Yan-Ping Xing<sup>a</sup>, Rong Zhao<sup>a</sup>, Wu-Liji Ao<sup>b</sup>, Ting-Ting Zhang<sup>a</sup>, Da-Chuan Zhang<sup>a</sup>, Yue-Yue Song<sup>a</sup>, Gui-Hua Bao<sup>b</sup> and Ting-Guo Kang<sup>a</sup>

<sup>a</sup>School of Pharmacy, Liaoning University of Traditional Chinese Medicine, Dalian, China; <sup>b</sup>School of Mongol Medicine, Inner Mongolia University for Nationalities, Tongliao, China

#### ABSTRACT

Aconitum kusnezoffii Rchb. is a medicinal plant in the Ranunculaceae family. In this study, we report the first complete mitochondrial genome of *A. kusnezoffii*. The total length of the mitochondrial genome of *A. kusnezoffii* is 440,720 bp and the GC content of 46.85%. The mitochondrial genome contained 37 protein-coding genes, 29 tRNAs, and three rRNAs. These data will provide the basis for the systematic evolutionary analysis of Ranunculaceae.

#### **ARTICLE HISTORY**

Received 28 September 2020 Accepted 24 January 2021

**KEYWORDS** Mitochondrial genome; *Aconitum kusnezoffii*; Ranunculaceae

There are more than 2000 species of Ranunculaceae recorded in the world (Cossard et al. 2016). They are distributed on all continents of the world except Antarctica, especially in temperate, cold temperate, and alpine regions. Ranunculaceae plants contain a variety of chemical components, many of which can be used as medicinal plants. *Aconitum kusnezoffii* Rchb. is a poisonous medicinal plant and has a long history of clinical application. At present, the research on *A. kusnezoffii* mainly focuses on chemical composition (Zan et al. 2018), efficacy–toxicity relation (Zhang et al. 2018), and safety dose research (Kim et al. 2012). However, the genetics and molecular biology of *A. kusnezoffii* are poorly understood, which has hindered research on the phylogenetic and molecular mechanism research.

Genomic DNA was obtained from fresh leaves of A. kusnezoffii collected from Dalian, China (E 121°87'63.24", N 39°06′18.72″) using the Illumina TruSeq<sup>™</sup> Nano DNA Sample Prep Kit method. The voucher specimen (A. kusnezoffii number: 10162200520005LY) and genomic DNA were deposited at the herbarium of Liaoning University of Traditional Chinese Medicine. Mitochondrial genome sequencing was performed using Illumina NovaSeq platform (Illumina, San Diego, CA) and the Nanopore platform (Oxford Nanopore Technologies, Oxford Park, UK). First, ABySS v2.0.2 (Simpson et al. 2009) was used for genome assembly of multiple-Kmer parameters, and an optimal assembly result was obtained. Second, BLASR (Chaisson and Tesler 2012) was used to map the preliminary assembly results to the Nanopore long reads. Then, SPAdes v3.10.1 (Bankevich et al. 2012) was used to assemble them together to construct contigs (scaffolds). Finally, all aligned Nanopore reads were extracted to perform self-correction and mt genome de novo assembly using the Canu v2.0 (Koren et al. 2017) package, followed by error correction using the Pilon v1.21. The Nanopore assembled sequences were then checked if the sequences have overlap and connection between them.

The paired-end (PE) libraries with an insert size of 450 bp were subjected to the Illumina platform and Nanopore platform, which generated 24.9 million reads. The length of PE was 150 bp  $\times$  2. The assembled mitochondrial genome was a closed circular molecule with the length of 440,720 bp and the GC content of 46.85%. The mitochondrial genome contained 37 protein-coding genes, 29 tRNAs, and three rRNAs (*rrn18, rrn26, rrn5*). The total length of the protein-coding genes was 31,836 bp, accounting for 7.22% of the total length of the mitochondrial genome. The average length of tRNAs was 73 bp, while the average length of rRNAs was 2120 bp. In addition, we found seven genes (*nad1, nad7, nad4, ccmFC, nad2, nad5, rps3*) containing 16 introns in total.

The MUMmer (Marcais et al. 2018) and BLAT software (Kent 2002) were used to do global alignment and local alignment between sample sequence and the reference genome under default parameters, and then manually optimized. The maximum-likelihood (ML) methods were performed for the genome-wide phylogenetic analyses using PhyML 3.0, respectively. Nucleotide substitution model selection was estimated with jModelTest 2.1.10 and Smart Model Selection in PhyML 3.0. The model GTR + I+G was selected for the ML analyses with 1000 bootstrap (BS) replicates to calculate the

CONTACT Liang Xu 861364054@qq.com School of Pharmacy, Liaoning University of Traditional Chinese Medicine, Dalian 116600, China; Gui-Hua Bao 836788623@qq.com School of Mongol Medicine, Inner Mongolia University for Nationalities, Tongliao 028000, China \*Both authors contributed equally to this work.

<sup>© 2021</sup> The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



Figure 1. Maximum-likelihood (ML) tree based on the mitogenome sequence of Aconitum kusnezoffii with 23 other species, the bootstrap supports are shown on each node.

BS values. The results tree was treated with iTOL 3.4.3. We modeled the ML tree with 23 other species (Figure 1). It was found that *A. kusnezoffii* and *Anemone maxima* (Park and Park 2020) from the order Ranunculales clustered closely together into one branch, which had 100% BS values. These data provide references for further study of the evolutionary history of *A. kusnezoffii*.

## **Disclosure statement**

No potential conflict of interest was reported by the author(s).

## Funding

This research was funded by 2019 Liaoning Provincial Department of Education Scientific Research Project, China [No. L201942], National Key Research and Development in the 13th Five-Year Plan [No. 2018YFC1708200], Major Special Fund for Science and Technology of Inner Mongolia Autonomous Region [No. 2019ZD004], Natural Science Fund Project of Liaoning Province [No. 2020-MS-224], and Mongolian Medicine R&D National Local Joint Engineering Research Center Open Fund Project, China [No. MDK2019047].

## Data availability statement

The genome sequence data that support the findings of this study are openly available in GenBank of NCBI at (https://www.ncbi.nlm.nih.gov/) under the accession no. MW013323. The associated BioProject, SRA, and

Bio-Sample numbers are PRJNA679444, SRX9535583, and SRS7742053, respectively.

#### References

- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, et al. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol. 19(5):455–477.
- Chaisson MJ, Tesler G. 2012. Mapping single molecule sequencing reads using basic local alignment with successive refinement (BLASR): application and theory. BMC Bioinformatics. 13:238.
- Cossard G, Sannier J, Sauquet H, Damerval C, Craene LR, Jabbour F, Nadot S. 2016. Subfamilial and tribal relationships of Ranunculaceae: evidence from eight molecular markers. Plant Syst Evol. 302(4): 419–431.
- Kent WJ. 2002. BLAT—the BLAST-like alignment tool. Genome Res. 12(4): 656–664.
- Kim J, Kim S, Lee S, Jeong H, Park M, Kim DW, Song BK, Lee JD, Kim S. 2012. Study of single-dose toxicity of *Aconitum Kusnezoffii* Reichb. Pharmacopuncture in rats. J Pharmacopuncture. 15(3):48–52.
- Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive kmer weighting and repeat separation. Genome Res. 27(5): 722–736.
- Marcais G, Delcher AL, Phillippy AM, Coston R, Salzberg SL, Zimin A. 2018. MUMmer4: a fast and versatile genome alignment system. PLoS Comput Biol. 14(1):e1005944.
- Park S, Park SJ. 2020. Large-scale phylogenomics reveals ancient introgression in Asian Hepatica and new insights into the origin of the insular endemic *Hepatica maxima*. Sci Rep. 10(1):16288.

- Simpson JT, Wong K, Jackman SD, Schein JE, Jones SJ, Birol I. 2009. ABySS: a parallel assembler for short read sequence data. Genome Res. 19(6):1117–1123.
- Zan K, Wangjie CR, Lu J, Guo LN, Zheng J, Ma SC. 2018. Content determination of four diester diterpenoid alkaloids in leaves of

Aconitum kusnezoffii by HPLC. China J Chin Mater Med. 43(4): 766–771.

Zhang XM, Lin ZJ, Li F, Zhang B, Guo XX, Yang L. 2018. Study on dosage rules of *Aconitum* herbs in oral prescriptions based on efficacy–toxicity relation. China J Chin Mater Med. 43(2):205.