

Increased Basal and Alum-Induced Interleukin-6 Levels in Geriatric Patients Are Associated with Cardiovascular Morbidity

Nathalie Compté¹, Karim Zouaoui Boudjeltia², Michel Vanhaeverbeek², Sandra De Breucker³, Thierry Pepersack³, Joel Tassinon⁴, Anne Trelcat⁴, Stanislas Goriely^{1*}

1 Institute for Medical Immunology (IMI), Université Libre de Bruxelles, Charleroi, Belgium, **2** Experimental Medicine Laboratory (Unit 222), Université Libre de Bruxelles, Hôpital A. Vésale, Montigny-Le-Tilleul, Belgium, **3** Service de Gériatrie, Hôpital Erasme, Bruxelles, Belgium, **4** ImmuneHealth, Charleroi, Belgium

Abstract

Background/Aim of the study: Low-grade systemic inflammation was suggested to participate to the decline of physiological functions and increased vulnerability encountered in older patients. Geriatric syndromes encompass various features such as functional dependence, polymorbidity, depression and malnutrition. There is a strong prevalence of cardiovascular diseases and related risk factors and chronic cytomegalovirus infections in the geriatric population. As these underlying conditions were proposed to influence the inflammatory state, the aim of this study was to assess their potential contribution to the association of geriatric syndromes with inflammatory parameters.

Methodology: We recruited 100 subjects in the general population or hospitalized for chronic medical conditions (age, 23–96 years). We collected information on clinical status (medical history, ongoing comorbidities, treatments and geriatric scales), biological parameters (hematological tests, cytomegalovirus serology) and cytokines production (basal and alum-induced interleukin (IL)-1 β and IL-6 levels). Using stepwise backward multivariate analyses, we defined which set of clinical and biological variables could be predictive for increased inflammatory markers.

Principal Findings: We confirmed the age-associated increase of circulating IL-6 levels. In contrast to geriatric scales, we found history of cardiovascular diseases to be strongly associated for this parameter as for high IL-6 production upon *ex vivo* stimulation with alum.

Conclusions: Association between low-grade inflammation and geriatric conditions could be linked to underlying cardiovascular diseases.

Citation: Compté N, Boudjeltia KZ, Vanhaeverbeek M, De Breucker S, Pepersack T, et al. (2013) Increased Basal and Alum-Induced Interleukin-6 Levels in Geriatric Patients Are Associated with Cardiovascular Morbidity. PLoS ONE 8(11): e81911. doi:10.1371/journal.pone.0081911

Editor: Sunil K Ahuja, South Texas Veterans Health Care System and University Health Science Center San Antonio, United States of America

Received: November 6, 2012; **Accepted:** October 28, 2013; **Published:** November 14, 2013

Copyright: © 2013 Compté et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: The Institute for Medical Immunology is sponsored by the government of the Walloon Region and GlaxoSmithKline Biologicals. This study was supported by the Fonds National de la Recherche Scientifique (FRS-FNRS, Belgium) and an Interuniversity Attraction Pole of the Belgian Federal Science Policy. NC was supported by the FRS-FNRS and Fonds Erasme. SG is a research associate of the FRS-FNRS. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The study was partly supported GlaxoSmithKline Biologicals. Authors JT and AT are employed by ImmuneHealth, Charleroi, Belgium. There are no patents, products in development or marketed products to declare. This does not alter the authors' adherence to all the PLOS ONE policies on sharing data and materials, as detailed online in the guide for authors.

* E-mail: stgoriel@ulb.ac.be

Introduction

Geriatric patients are often affected by low-grade, chronic systemic inflammatory state. This process referred to as “inflamm-aging” was suggested to participate to the decline of physiological functions and increased vulnerability encountered in older patients [1]. Indeed, inflammatory markers, such as serum interleukin (IL)-6, C-reactive protein (CRP) or TNF-alpha are powerful predictors of morbidity and mortality in very old humans [2–5]. Furthermore, several reports indicate an association between inflammatory markers and geriatric

conditions such as frailty, functional decline or depression [6–8].

Because of the overlap between geriatric syndromes and comorbidities, it is extremely difficult from the current literature to identify which clinical parameters are associated with increased inflammatory markers. Cardiovascular (CV) diseases are among the most prevalent comorbidities in geriatric patients [9]. CV diseases and related risk factors were proposed to influence the inflammatory state [10]. Other factors such as chronic cytomegalovirus (CMV) infections could also contribute to the association between geriatric conditions and

low-grade inflammation [11,12]. The aim of this study was to assess the contribution of these underlying conditions in the association between inflammatory markers and common geriatric conditions (such as comorbidities, functional dependence, cognitive disorders, depression and malnutrition). For this purpose, we recruited old subjects in the general population or hospitalized for chronic medical conditions. We performed a comprehensive geriatric assessment (CGA) to identify comorbidities and common geriatric conditions. In this cross-sectional study, we also included younger subjects (with or without CV diseases) to assess the contribution of ongoing chronic co-morbidities in low-grade inflammation independently of age or geriatric conditions.

We focused our analysis on plasmatic IL-6, a classical systemic inflammatory marker. In order to more specifically address the functional status of blood innate immune cells, we also monitored cytokine production upon *ex vivo* stimulation with Alum. This vaccine adjuvant is a classical activator of the NLRP3-dependent inflammasome [13]. On its own, it does not directly support transcriptional activation of pro-inflammatory genes [13]. However, if cells are in a pre-activated state that leads to accumulation of pro-IL1 β (e.g. in response to Toll-like receptor (TLR) ligands), alum will induce the secretion of mature IL-1 β and downstream IL-6 production. Hence, cytokine production in these conditions will indirectly reflect the global activation status of blood innate immune cells.

Materials and Methods

Subjects

Between 2010 and 2012, 108 subjects aged between 23 to 93 years (65 women and 43 men) were enrolled in this cross-sectional study. The exclusion criteria were: CRP \geq 1 mg/dl, hepatic disturbance, presence of active cancer, autoimmune disease or infection, immunosuppression state, use of glucocorticoids, immunosuppressors or non steroid anti-inflammatory drugs (NSAID); advanced dementia (MMSE below 23 points[14]) was also excluded. Healthy young and old volunteers were recruited at the geriatric day ward of Erasme hospital among hospital and laboratory employees, volunteers of a non-profit seniors association ("Association pour le Soutien de l'Etude du Vieillessement") or through public solicitation. Hospitalized volunteers were recruited from cardiology, neurology, rehabilitation or endocrinology units. For patients > 75 years, we assessed the risk of frailty (ISAR score >1 point) to recruit patients with geriatric conditions in geriatric unit (tertiary care at Erasme hospital, Brussels). The "Identification of Senior At Risk" (ISAR) score is a rapid scale performed at the emergency department which evaluates frailty and the risk of functional decline during hospitalization with 6 questions about dependence, previous hospitalization, eye troubles, memory problems and number of medications [15].

Ethics statement

All subject signed an informed consent and the study received approval from Erasme hospital Ethics Committee (808 route de Lennik, B-1070 Brussels, Belgium, agreement n°OM021).

Determination of clinical characteristics

All subjects were screened for underlying illnesses by direct questioning, medical archives and blood sampling. Social evaluation included determination of age, gender, home (private versus institution), and marital status. Clinical data comprised: smoking and alcohol habits, pneumococcal and influenza vaccine status, allergy, Body mass index (BMI), medical history, current treatment and reasons for hospitalization. Cardiovascular (CV) diseases were defined as history of stroke, myocardial infarct, cardiac insufficiency, cerebral vascular disease or atheromatosis assessed by carotid or leg Doppler echography and ischemic symptoms. CV risk factors were defined by the presence in the anamnesis of hypertension, type 2 diabetes, hypercholesterolemia or statin intake, infarct history or smoking. Osteoporosis was defined by self report.

For subjects > 75 years, we performed a CGA to identify comorbidity and common geriatric conditions. The poly pathology and the severity of the medical problems were scored using the "Cumulative Illness Rating Scale-Geriatric" (CIRS-G). It is an instrument to quantify disease burden. It differentiates older adults with the highest risk and severity of infection with a markedly impaired vaccine response [16–18]. It comprises a comprehensive review of medical problems of 14 organ systems. It is based on a 0 to 4 rating of each organ system [19–21]. The "Geriatric Depression Scale" was used to assess the probability of depressed mood (GDS-15) in 15 questions [22–22]. The assessment of "Activities of Daily Living" (ADL) was made by using Katz's scale. It includes the following items: bathing, dressing, transfer, toilet, continence and eating. Each task is graded on a 3-level scale (1 to 3 for Katz's scale), where lower levels represent the absence of dependence and upper level the maximal dependence for the task [23]. Cognitive functions were assessed using the "Mini Mental State Examination" (MMSE). Possible scores range from 0 to 30 points, with lower scores indicating impaired cognitive function [14]. Nutritional status was assessed using the "Mini Nutritional Assessment" (MNA) [24,25]. A score \geq 24 identifies patients with a good nutritional status. Scores between 17 and 23.5 identify patients at risk of malnutrition. These latter patients have not yet started to lose weight and do not show low plasma albumin levels but have lower protein-caloric intakes than recommended. A score < 17 indicates protein-caloric malnutrition. Pain was assessed using a visual analogical scale from 0 to 10 points. Maximal grip strength and fatigue resistance were measured using a Martin vigorimeter (Elmed Inc., Addison, USA) as described previously [26,27]. Briefly, the shoulder is adducted and neutrally rotated, elbow flexed at 90°, forearm in neutral position and wrist in slight extension (0 to 30°). The subject is then asked to squeeze the large bulb of the vigorimeter as hard as possible. The highest of three attempts is noted as the maximal grip strength (in KPa). Afterwards, the subject is instructed to squeeze again the bulb of the vigorimeter as hard as possible and to maintain this maximal pressure; the time (in seconds) during which grip strength dropped to 50% of its maximum is recorded as fatigue resistance. This fatigue resistance test is highly reproducible in elderly subjects with ICC-values ranging respectively from 0.91

to 0.94 and from 0.88 to 0.91 for intra- and inter-observer reliability. An estimate of the total effort produced during the fatigue resistance test, defined as Grip Work, can be calculated as $\text{Grip Work} = (\text{Grip Strength} \times 0.75) \times \text{Fatigue Resistance}$. This parameter represents the physiologic work delivered by the handgrip muscles during the fatigue resistance test. When graphically represented, grip work is the area under the curve with grip strength in the vertical and time in the horizontal axis. All handgrip performance tests are executed with the dominant hand.

We performed routine biochemical assessment to identify potential exclusion factors. CMV-specific IgG levels were determined by ELISA (ETI-CYTOK-GLPUS; Diasorin, P002033)

Blood sample collection, management and ex vivo stimulation

Venous blood samples (55ml) from all subjects were collected in pyrogen-free, heparinized tubes between 11.00 and 13.00. After 2 h (because of transport), blood samples were centrifuged; plasma were collected and stored at -80° . Basal IL-6 was measured by ELISA (quantikine immunoassay, R&D systems).

Blood was diluted 2-fold with sterile RPMI-1640 and incubated in the presence or not of alum (Alum hydroxide and magnesium hydroxide, 77161, Pierce/Thermo Fischer, 500 μ g/ml) at 37°C and 5% CO_2 . After 18h, cell-free supernatants were collected and stored at -80° to measure cytokine production. IL-1 β and IL-6 levels were determined using ELISA kits (duoset, R&D systems). A control sample was placed in all ELISA plates to assess the internal quality of our tests.

Statistics

The SigmaStat[®] software package version 3.5 (Jandel Scientific) was used for multivariate analyses and GraphPad prism 5[®] software for univariate analyses and Mann-Whitney rank sum test. For statistical analyses, cytokine concentrations were \log_{10} -transformed in view of their non parametric distribution.

In order to identify clinical and biological factors that could be significantly associated with cytokine levels, we performed univariate analyses (depicted by Pearson's coefficient). For all volunteers, these parameters comprised: age, gender, CV diseases and risk factors (smoking, arterial hypertension, type 2 diabetes, cholesterol levels, BMI), monocytes and white blood cell counts (WBC), depression, creatinin clearance and CMV status. For individuals > 75 years, geriatric evaluations (MNA, KATZ, GDS, MMSE, CIRS scales, grip strength and osteoporosis) were also included in this analysis.

In order to define independent predictive factors of inflammation, we tested several models by stepwise backward multi-linear regression analyses. To decrease the risk of over-fitting, no more than 8 or 5 variables were included at a time for the entire study group (n=100) or for older patients (n=52), respectively.

Katz and MNA (cut off values: ≥ 8 , < 23.5 , > 1 respectively) and clinical variables were treated as dichotomous variables

Table 1. Main characteristics of the study group.

	Entire group
N	100
Age (year)	66.4 (23 - 93)*
Gender M/F	43/65
BMI (kg/m ³)	24.9 (17-46) [†]
Active smokers (%)	11
Hypertension (%)	50
Type 2 diabetes (%)	12
Hypercholesterolemia (%)	56
Cardiovascular diseases (%)	29
CMV seropositivity (%)	55

*. Median (range)

doi: 10.1371/journal.pone.0081911.t001

while other data were continuous. A probability level of $p < 0.05$ was considered to be significant.

Results

Characteristics of the enrolled individuals

In the entire study population, 8 subjects were excluded because CRP values exceeded 1mg/dl and 100 subjects were included in the analyses. The demographic, clinical and biochemical characteristics of the entire group are presented in table 1. Fifty two subjects were older than 75 years. We performed a comprehensive geriatric assessment in this subgroup. The characteristics of old individuals with ISAR >1 (n=25) or ISAR ≤ 1 point (n=27) are presented in tables 2-4. As expected, in patients with ISAR >1 point, decreased grip strength, depression state (GDS), comorbidity burden (CIRS-G), dependence (Katz scale), cognitive troubles (MMSE) and impaired nutritional status (MNA) were more prevalent in comparison to individuals with ISAR ≤ 1 point. However, 4/25 individuals in the group ISAR >1 point did not present any geriatric conditions (Katz > 8 points, MNA ≤ 23.5 points or GDS > 5 points). In the group ISAR ≤ 1 point, one patient had an MNA score at 18.5 points and two patients had a GDS score at 6 points.

Occurrence of cardiovascular diseases is independently associated with high circulating IL-6 levels

We analyzed basal plasma IL-6 levels rather than spontaneous secretion by isolated PBMC or monocytes as it also reflects the contribution of other potential sources such as adipocytes, muscle or endothelial cells.

Using univariate analyses, we first attempted to identify relevant associations between clinical and biological parameters on one hand and basal IL-6 levels on the other. We found a significant association for monocytes ($R^2=0.8$; $p=0.01$) and WBC counts ($R^2=0.12$; $p=0.001$), age ($R^2=0.05$; $p=0.03$), CV risk factors ($R^2=0.08$; $p=0.01$) and diseases ($R^2=0.14$; $p=0.0007$), type 2 diabetes ($R^2=0.13$; $p=0.013$) and a trend for hypertension ($R^2=0.08$; $p=0.06$).

Table 2. Demographic characteristics and comorbidities of old individuals with ISAR score ≤ 1 point and > 1 point.

	ISAR ≤ 1	ISAR > 1
N	27	25
Recruitment type	Ambulatory	Hospitalized
Age (years)	80 (76-88)*	83 (73-90)*
Gender (M/F)	9/18	8/17
BMI (kg/m ²)	25.4 (19-32)*	24.7 (18-34)*
Active smokers (%)	0	8
Hypertension (%)	48	28
Type 2 diabetes (%)	0	28
Hypercholesterolemia (%)	74	60
Cardiovascular diseases (%)	3.7	60
Osteoporosis (%)	14.8	40

*. Median (range)

doi: 10.1371/journal.pone.0081911.t002

Table 3. Geriatric characteristics of old individuals with ISAR score ≤ 1 point and > 1 point.

	ISAR ≤ 1	ISAR > 1
ISAR (score)	0 (0-1)	3 (2- 4)
GDS (score)	1 (0- 2)	5 (2-5)
Katz (score)	6 (6-7)	9 (7-12)
MMSE (score)	29 (28- 29)	26 (24- 28)
MNA (score)	26.75 (25.6-27.8)	20 (17.5- 22.5)
Vigrometer Force (kPa)	30 (17- 45.5)	10 (0-24)
Vigrometer ; fatigue resistance (sec)	20 (10- 40)	0 (0-10)
CIRS-G (category number)	6 (3.5 -6.5)	9 (7-10)
CIRS-G (global score)	9 (6.5-11)	19 (15-22)
CIRS-G (severity index)	1.6 (1.4-2)	2.2 (1.72-2.33)

Median (range)

doi: 10.1371/journal.pone.0081911.t003

Table 4. Biochemical characteristics of old individuals with ISAR score ≤ 1 point and > 1 point.

	ISAR ≤ 1	ISAR > 1
Cholesterol (mg/dl)*	202 (182 -223)	187 (173 -206)
Ferritin (mg/dl)*	107 (60-184)	176 (112 -275)
Prealbumin (mg/dl)*	25 (22-28)	19 (17-24)
CMV seropositivity (%)	48	64

*. Median (range)

doi: 10.1371/journal.pone.0081911.t004

To assess the relative contribution of these variables, we next performed multivariate analysis (summarized in Table 5) including age, gender, cardiovascular risk factors and diseases, WBC and monocytes counts, creatinin clearance and CMV status. Age, CV diseases and WBC counts appeared to be associated with basal IL-6 levels. IL-6 values were then normalized to WBC counts and we performed the same

Table 5. Multivariate analyses for basal IL-6 levels in the whole study group.

N=100	R ² ; F value	Standardized Coefficient	p value
Log₁₀ IL-6¹	R ² =0.31; F=10.93		
WBC		0.363	<0.001
CV diseases		0.319	0.002
Age		0.255	0.0013
Log₁₀ IL-6/WBC ratio²	R ² =0.31; F=10.93		
CV diseases		0.35	0.001
Age		0.232	0.03

1. adjusted for age, gender, CV risk factors and diseases, WBC and monocytes counts, creatinin clearance, CMV status.

2. adjusted for age, gender, CV risk factors and diseases, creatinin clearance, CMV status.

doi: 10.1371/journal.pone.0081911.t005

multivariate analysis. Once again, age and CV diseases were found to be significantly associated with basal IL-6 levels/WBC ratio.

Secondly, we assessed the association between geriatric conditions and low-grade inflammation. Geriatric scales are only validated in aged individuals. Hence, to assess the association between inflammatory state and geriatric conditions, we restricted our analyses to subjects > 75 years of age. We compared plasma IL-6 levels between older individuals with ISAR > 1 and ISAR ≤ 1 point. IL-6 levels were significantly increased in this second group (Figure 1). To take into account possible confounding variables, we performed univariate analyses. No geriatric conditions (MMSE, CIRS-G and grip strength) were significantly correlated with this inflammatory parameter but there was a trend for Katz, MNA and GDS. As for the entire study group, we observed a significant association between CV diseases (R²=0.1; p=0.04), type 2 diabetes (R²=0.13; p=0.02) and plasma IL-6 as well as a trend for CV risk factors (R²=0.08; p=0.07). We performed multivariate analyses with age, CV risk factors, gender and geriatric scales. As geriatric scales influence each other, we analyzed separately MNA, GDS or MMSE scores in multivariate analysis. No geriatric score appeared as a significant predictive factor (cfr table 6).

CV diseases rather than geriatric conditions are associated with cytokine production upon alum stimulation

Incubation of whole blood with alum led to the induction of IL-1 β and downstream IL-6 production in most donors. In univariate analyses, CV risk factors (R²=0.06, p=0.003) and diseases (R²=0.06; p=0.01), hypertension (R²=0.07; p=0.007), WBC counts (R²=0.07; p=0.01) were significantly associated with IL-1 β production post-alum and there was a trend for type 2 diabetes (R²=0.03; p=0.07) and for monocytes counts (R²=0.03; p=0.07). For IL-6 production upon alum stimulation, CV risk factors (R²=0.06; p=0.02), type 2 diabetes (R²=0.04; p=0.048), CV diseases (R²=0.05; p=0.02), monocytes counts (R²=0.06; p=0.02) were significant predictive factors. For this

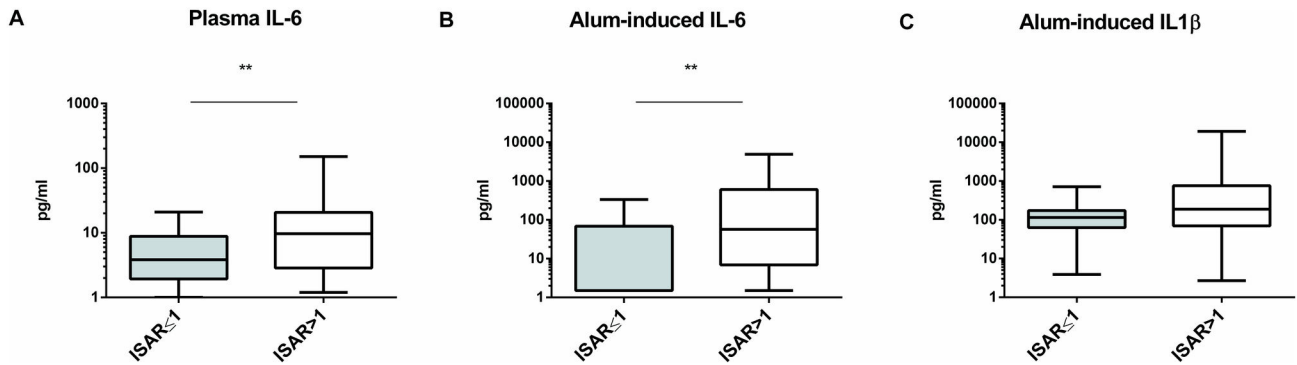


Figure 1. Inflammatory markers in individuals with ISAR. ≤ 1 and > 1 point.

(A) Plasmatic IL-6 levels were analyzed by ELISA. (B-C) Whole blood samples were incubated with alum (500 μ g/ml) for 18h. Cell-free supernatants were collected and analyzed for IL-6 and IL-1 β levels by ELISA. ISAR > 1 point, n=27; ISAR ≤ 1 point, n=25. For statistical analysis, we used the Mann-Whitney rank sum test *p<0.05; **p<0.01.

doi: 10.1371/journal.pone.0081911.g001

parameter, we also observed a trend for CMV status ($R^2=0.03$; $p=0.09$).

We then performed multivariate analyses to assess the relative contribution of these factors. CV diseases were significantly associated with both alum-induced IL-1 β and IL-6 levels (table 7). The same analyses were performed after normalization of cytokine levels with WBC counts. Once again, CV diseases were found to be significantly associated with alum-induced IL-1 β and IL-6 levels/WBC ratio. CMV status was also found to be associated with alum-induced IL-6/WBC ratio. We confirmed that alum-induced IL-1 β /WBC ratio was associated with CV risk factors. We next compared the levels of inflammatory markers between older subjects with ISAR score ≤ 1 point and > 1 point. Alum-induced IL-6 levels but not IL-1 β levels were significantly increased in this second group (Figure 1). In these older subjects, univariate analyses confirmed that CV risk factors ($R^2=0.19$; $p=0.001$) and CV diseases ($R^2=0.17$; $p=0.002$) were significant predictive factors for IL-1 β production. Similarly, for IL-6 production upon alum stimulation, CV risk factors ($R^2=0.17$; $p=0.002$), CV diseases ($R^2=0.19$; $p=0.001$) and CMV status ($R^2=0.08$; $p=0.03$) were significant predictive factors. We found a positive trend for MNA, MMSE and GDS for IL-6 upon alum stimulation.

We performed multivariate analyses with age, gender, CV risk factors and diseases, CMV status, WBC counts and geriatric scales (table 8). As geriatric scales influence each other, we performed the same multivariate analyses replacing MNA score by GDS or MMSE score for IL-6 upon alum stimulation. In contrast to geriatric scales, CV risk factors were found to be significant predictive factors for IL-6 and IL-1 β levels. Taken together, we observed that association between low-grade inflammation and geriatric conditions could be linked to the presence of cardiovascular diseases.

Discussion

Previous studies on inflammation in older individuals and geriatric patients yielded conflicting conclusions. The main

Table 6. Multivariate analyses for basal IL-6 levels in subjects

N=52	R ² ; F value	Standardized Coefficient	p value
Log₁₀ IL-6¹	R ² =0.13; F=6.5		
Age		0.03	0.014
Log₁₀ IL-6²	R ² =0.14; F=7		
Age		0.03	0.011
Log₁₀ IL-6³	R ² =0.13; F=6.5		
Age		0.03	0.014
Log₁₀ IL-6/WBC ratio⁴	R ² =0.15; F=7.7		
Age		0.04	0.008
Log₁₀ IL-6/WBC ratio⁵	R ² =0.16; F=8.1		
Age		0.04	0.007
Log₁₀ IL-6/WBC ratio⁶	R ² =0.15; F=7.7		
Age		0.04	0.008

1. adjusted for Katz, age, gender, CV risk factors and WBC counts.
2. adjusted for MNA, age, gender, CV risk factors and WBC counts.
3. adjusted for GDS, age, gender, CV risk factors and WBC counts.
4. adjusted for Katz, age, gender, CV risk factors and diseases.
5. adjusted for MNA, age, gender, CV risk factors and diseases.
6. adjusted for GDS, age, gender, CV risk factors and diseases.

doi: 10.1371/journal.pone.0081911.t006

difficulty in the interpretation of the results comes from the heterogeneity of the population and differences in the classification, definitions and inclusion/exclusion criteria used. A summary of such studies is shown in Table S1. We included main study parameters (size of the group, age groups, inclusion criteria, clinical data, design and statistical methods used) and their conclusions. Increased inflammatory markers are generally associated with age, CV comorbidities and geriatric conditions. However, when confounding factors are taken into account, the relative contribution of these parameters is less clear. Herein, we took particular care to characterize the clinical parameters of the enrolled subjects. As

Table 7. Multivariate analyses of alum-induced cytokine levels in the whole study group.

N=100	R ² ; F value	Standardized Coefficient	p value
Log₁₀ Alum-IL-6¹	R ² =0.08; F=9		
CV diseases		0.29	0.003
Log₁₀ Alum-IL-1¹	R ² =0.13; F=7.1		
CV risk factors		0.26	0.01
WBC		0.21	0.03
Log₁₀ Alum-IL-6/WBC ratio²	R ² =0.12 ; F=6.3		
CV diseases		0.29	0.004
CMV		0.19	0.048
Log₁₀ Alum-IL-1/WBC ratio²	R ² =0.07 ; F=7.5		
CV risk factors		0.27	0.007

1. adjusted for age, gender, CV risk factors and diseases, WBC and monocytes counts, creatinin clearance, CMV status.

2. adjusted for age, gender, CV risk factors and diseases, creatinin clearance, CMV status.

doi: 10.1371/journal.pone.0081911.t007

Table 8. Multivariate analyses of alum-induced cytokine levels in subjects > 75 years.

N=52	R ² ; F value	Standardized Coefficient	p value
Log₁₀ Alum-IL-1/WBC ratio¹	R ² =0.18 ; F=10.6		
CV risk factors		0.42	0.002
Log₁₀ Alum-IL-6/WBC ratio¹	R ² =0.35 ; F=8.7		
CV risk factors		0.45	<0.001
CMV		0.34	0.005
Gender		0.26	0.03

1. adjusted for age, gender, CV risk factors, CMV and GDS or MNA or MMSE.

doi: 10.1371/journal.pone.0081911.t008

CV disorders are associated with low-grade inflammation and as healthy old individuals might present underlying CV disorders without any symptoms, we included younger subjects (with or without CV diseases) to assess the contribution of ongoing chronic co-morbidities in low-grade inflammation independently of age or geriatric conditions. Indeed, in enrolled younger subjects (mean age: 49 years), proportion of CV diseases or risk factors (hypertension: 38%, type 2 diabetes: 10%, hypercholesterolemia: 63% and ongoing CV diseases: 27%) was comparable to that observed in older individuals. We observed that the strongest predictive factors for increased IL-6 plasmatic levels were the history of CV diseases or risk factors and age. A previous study indicated that IL-6 levels were associated with age but not with CV risk factors (assessed by BMI, blood pressure or lipidemia) in a 20-84 year old healthy population. However, the authors did not report the occurrence of CV diseases in their study population [7,28]. Forsey et al. also observed an increase of IL-6 with age and poor health status (using SENIEUR, OCTO and NONA criteria) but they did not characterize their group for ongoing comorbidities [29].

Hence, as suggested previously [10], our findings support the notion that part of age-associated inflammatory state could be linked to accumulation of CV risk factors and morbidities in the geriatric population. In another study, high inflammatory markers (in particular basal IL-6) were found to be predictive for the development of CV events within a 3-year period in older well-functioning individuals [30]. In people without known CV diseases, CRP level is also an important predictive factor for CV events [31]. Furthermore, two large mendelian randomized studies showed that polymorphisms of the IL-6R are associated with reduced CRP levels and the risk of coronary artery diseases. However, both studies showed increased concentrations of IL-6 levels that seem in contradiction with a causal role of IL-6. This paradox might be resolved by the fact that both IL-6R polymorphisms seem to be associated with downregulation of IL-6R signaling and that increased IL-6 levels could be a result of a feedback loop to increase IL-6 signaling [32,33]. A meta-analysis of prospective studies involving white populations confirmed that polymorphisms of the IL-6 gene were also associated with the development of coronary artery diseases [34].

We observed that IL-1 β and IL-6 production by blood cells in response to alum was also strongly associated with CV diseases and risk factors. This result suggests that in these patients, circulating innate immune cells could be in a pre-activated state. Indeed, CV risk factors such as hypercholesterolemia, high saturated fat diet, obesity and hyperglycemia are known to promote inflammation through different pathways such as the endoplasmic reticulum (ER) stress, the inflammasomes and the TLRs [3,35,36]. ER stress and TLR pathways both lead to transcriptional activation of inflammatory genes such as pro-IL1 β that could account for the association between high IL-1 β and IL-6 production in response to alum and CV diseases that we observed. However, the design of our study does not allow us to establish a causal links between CV diseases and low-grade inflammation. Coronary artery diseases, high BMI and atherosclerosis are associated with an increase in circulating inflammatory monocytes (CD14^{dim}CD16⁺). Furthermore, the proportion of intermediate monocytes (CD14⁺⁺CD16⁺) is also associated with increased CV risk factors and is predictive of CV events and poor outcome [37,38]. In comparison to classical monocytes (CD14⁺⁺CD16), this subset displays an activated phenotype, reduced telomere length and produces high amounts of proinflammatory cytokines [39,40]. It would therefore be important to determine the responsiveness of these cells to alum in the context of CV diseases.

Along this line, chronic CMV infection was also associated with modulation of monocyte functions [41] that could account for the association between CMV status and IL-6 levels upon alum that we observed. Chronic infection with CMV is seen as a major factor that could influence exhaustion of memory T lymphocytes in older individuals. CMV seropositivity is part of the "immune risk phenotype" (IRP) and is associated with increased all-cause mortality [42]. It has been suggested that immune response to persistent CMV infection could also participate to the establishment or maintenance of inflammaging processes [2,42]. Several studies previously reported an

association between IL-6 levels and CMV status. In patients with coronary artery diseases, Blanckenberg et al. observed that CMV titers were correlated to IL-6 levels and together could predict cardiac mortality [43]. In community-dwelling post-menopausal women, mean IL-6 levels were found to be higher in CMV+ subjects but significance was lost after adjustment for confounding factors [11,44]. Interestingly, in the “Hertfordshire Ageing study” group, increase in plasma IL-6 levels upon aging was similar in seronegative, seropositive volunteers and volunteers who became seropositive for CMV, arguing against a central role of CMV infection in this process [45]. We identified CMV status as an independent predictor of IL-6 production upon alum stimulation but not for other inflammatory parameters, suggesting that CMV could directly or indirectly influence the capacity of the immune cells to produce this cytokine. It would be important to define the potential mechanistic links responsible for these observations.

In our study, geriatric patients with ISAR>1 point display increased basal and alum-induced IL-6 production compared to old individuals with ISAR<1 point. However, we did not find any significant association between geriatric conditions (assessed by Katz, MNA, GDS, CIRS-G, MMSE, Grip strength) and low-grade inflammation. Studies that have identified association between frailty and high IL-6 levels generally used criteria developed by Fried et al. (slow gait speed, low physical activity, unintentional weight loss, self-reported exhaustion and muscle weakness) [46]. Initial studies indicated an association between elevated CRP levels and frailty after exclusion of CV diseases and diabetes [47]. However, association between increased IL-6 and frailty was less clear in other studies. For example, in the “Longitudinal Aging Study Amsterdam” (LASA), IL-6 was not associated with frailty parameters [48]. While IL-6 could be predictive for frailty in a cohort of women aged 70 to 79 [11], increased IL-6 levels were not found to be predictive for the later development of frailty in the “Hertfordshire Ageing study” (HAS, 10 year follow-up, age between 65-70 years) [49]. The use of NSAID and corticoids, the heterogeneity of the population and follow-up bias because of mortality and various frailty criteria could partly explain these discrepancies.

Dependency is a direct expression of frailty. In institutionalized subjects older than 65 years old selected for cerebral vascular diseases, a decline in activities of daily living (Katz ADL and Barthel index) was associated with high IL-6 levels [8]. Gonzalo-Calvo et al. showed the same results in institutionalized volunteers [8,50]. However, variability in interpretation of Katz score hampers interpretation and comparison of these studies with the present one [51].

Muscle strength and walking speed are major phenotypic criteria of frailty. Several studies used this parameter to assess the link between frailty and inflammation. Indeed, Schaap et al. observed that higher IL-6 and CRP levels were associated with decreased muscle strength in a cohort of independent individuals over 55 years after adjustment for confounders such as age, depressive symptoms, chronic diseases and cognitive troubles [52]. Similar conclusions were also previously reached by Ferruci et al. in community-dwelling women (above 65) [53]. In a cohort of healthy subjects between 20 to 72 year old, Blain and colleagues observed a similar association between IL-6

levels and walking performance but not with muscle strength [54]. Interestingly, Bautmans et al. showed that while better grip strength and fatigue resistance were associated with higher IL-6 levels in well-functioning subjects, this was not the case for nursing home patients and hospitalized patients [26,55,56]. Taken together, these data indicate that association between inflammation and grip strength is highly dependent on the population under study and might reflect variations in important confounding factors. In particular, CV diseases and underlying pathological conditions (such as atherosclerosis) are likely to participate to the development of frailty and reduced multi-organ physiological reserve in old age. Hence, CV diseases, frailty and geriatric conditions are probably deeply intertwined.

Cognitive and psychological factors are other important aspects of the geriatric patient. As these factors also participate to the general health status of the patients and could impact their immunological profile, we used a CGA to characterize our patients [57,58]. Some studies found an association between depressive or stress symptoms and IL-6 levels [6,7,59]. Meta-analysis confirmed the association between IL-6, CRP and IL-1 β levels and depression. However, it was not significant anymore once age or medication intake were taken into account [60]. In the context of this study, for ethical reasons and accuracy of the other geriatric scales, we did not recruit patients presenting with strong cognitive impairment. The potential association of these clinical parameters and inflammation is therefore probably underestimated. Several studies have shown an association with cognitive decline and IL-6 [61,62] but most studies observed an association with vascular dementia but not Alzheimer’s dementia [63,64], suggesting that underlying CV conditions could be responsible for such association. However, other studies that included patients with low MMSE scores did not detect an association for TNF α and IL-6 levels [65–67].

One limit of our study is the small size of the group given its heterogeneity. We performed numerous comparisons that increase α error of the statistical analyses. As we recruited patients that were hospitalized, acute medical conditions could bias inflammatory markers. To reduce this potential effect, data were collected just before discharge to home. Although conclusions were similar when the whole group was analyzed (not shown), we limited evaluation of geriatric scales to subjects above 75 years of age, as these clinical parameters might not reflect the same underlying conditions in younger patients. As we included younger and older individuals with chronic diseases, we cannot fully assume that the impact of these diseases on physiological and immune functions will be the same in both age groups. Transversal studies do not take into account the evolution of clinical and immunological parameters. We did not specifically look at periodontal diseases known to be associated with chronic inflammation [68].

Despite these limitations, we identified a clear association between CV diseases and inflammatory parameters suggesting that among clinical parameters, these factors have the largest impact on the development of low-grade inflammation observed in the geriatric population.

Supporting Information

Table S1. Summary of studies that looked at inflammatory markers in old age. A) Correlation of plasma IL-6 levels with age, comorbidities and frailty. B) Inflammatory markers as predictive factor for mortality in old age. (DOC)

Acknowledgements

The authors would like to thank Arnaud Marchant, Muriel Nguyen, Julie Callenaere, Yves Delmarcelle, Thierry Honorez,

References

- Fulop T, Larbi A, Witkowski JM, McElhane J, Loeb M et al. (2010) Aging, frailty and age-related diseases. *Biogerontology* 11: 547-563. doi:10.1007/s10522-010-9287-2. PubMed: 20559726.
- Wikby A, Nilsson BO, Forsey R, Thompson J, Strindhall J et al. (2006) The immune risk phenotype is associated with IL-6 in the terminal decline stage: findings from the Swedish NONA immune longitudinal study of very late life functioning. *Mech Ageing Dev* 127: 695-704. doi: 10.1016/j.mad.2006.04.003. PubMed: 16750842.
- Krabbe KS, Pedersen M, Bruunsgaard H (2004) Inflammatory mediators in the elderly. *Exp Gerontol* 39: 687-699. doi:10.1016/j.exger.2004.01.009. PubMed: 15130663.
- Bruunsgaard H, Ladelund S, Pedersen AN, Schroll M, Jørgensen T et al. (2003) Predicting death from tumour necrosis factor-alpha and interleukin-6 in 80-year-old people. *Clin Exp Immunol* 132: 24-31. doi: 10.1046/j.1365-2249.2003.02137.x. PubMed: 12653832.
- van den Biggelaar AH, Huizinga TW, de Craen AJ, Gussekloo J, Heijmans BT et al. (2004) Impaired innate immunity predicts frailty in old age. The Leiden 85-plus study. *Exp Gerontol* 39: 1407-1414. doi: 10.1016/j.exger.2004.06.009. PubMed: 15489064.
- Penninx BW, Kritchevsky SB, Yaffe K, Newman AB, Simonsick EM et al. (2003) Inflammatory markers and depressed mood in older persons: results from the Health, Aging and Body Composition study. *Biol Psychiatry* 54: 566-572. doi:10.1016/S0006-3223(02)01811-5. PubMed: 12946885.
- Kiecolt-Glaser JK, Preacher KJ, MacCallum RC, Atkinson C, Malarkey WB et al. (2003) Chronic stress and age-related increases in the proinflammatory cytokine IL-6. *Proc Natl Acad Sci U S A* 100: 9090-9095. doi:10.1073/pnas.1531903100. PubMed: 12840146.
- Zuliani G, Guerra G, Ranzini M, Rossi L, Munari MR et al. (2007) High interleukin-6 plasma levels are associated with functional impairment in older patients with vascular dementia. *Int J Geriatr Psychiatry* 22: 305-311. doi:10.1002/gps.1674. PubMed: 17022108.
- Chang SS, Weiss CO, Xue QL, Fried LP (2012) Association between inflammatory-related disease burden and frailty: results from the Women's Health and Aging Studies (WHAS) I and II. *Arch Gerontol Geriatr* 54: 9-15. doi:10.1016/j.archger.2011.05.020. PubMed: 21763008.
- Ferrucci L, Corsi A, Lauretani F, Bandinelli S, Bartali B et al. (2005) The origins of age-related proinflammatory state. *Blood* 105: 2294-2299. doi:10.1182/blood-2004-07-2599. PubMed: 15572589.
- Schmaltz HN, Fried LP, Xue QL, Walston J, Leng SX et al. (2005) Chronic cytomegalovirus infection and inflammation are associated with prevalent frailty in community-dwelling older women. *J Am Geriatr Soc* 53: 747-754. doi:10.1111/j.1532-5415.2005.53250.x. PubMed: 15877548.
- Wang GC, Kao WH, Murakami P, Xue QL, Chiou RB et al. (2010) Cytomegalovirus infection and the risk of mortality and frailty in older women: a prospective observational cohort study. *Am J Epidemiol* 171: 1144-1152. doi:10.1093/aje/kwq062. PubMed: 20400465.
- Li H, Nookala S, Re F (2007) Aluminum hydroxide adjuvants activate caspase-1 and induce IL-1beta and IL-18 release. *J Immunol* 178: 5271-5276. PubMed: 17404311.
- Folstein MF, Folstein SE, McHugh PR (1975) "Mini-mental state". A practical method for grading the cognitive state of patients for the clinician. *J Psychiatr Res* 12: 189-198. doi: 10.1016/0022-3956(75)90026-6. PubMed: 1202204.
- McCusker J, Bellavance F, Cardin S, Trépanier S (1998) Screening for geriatric problems in the emergency department: reliability and validity. Identification of Seniors at Risk (ISAR) Steering Committee. *Acad Emerg Med* 5: 883-893. doi:10.1111/j.1553-2712.1998.tb02818.x. PubMed: 9754501.
- Gravenstein S, Drinka P, Duthie EH, Miller BA, Brown CS et al. (1994) Efficacy of an influenza hemagglutinin-diphtheria toxoid conjugate vaccine in elderly nursing home subjects during an influenza outbreak. *J Am Geriatr Soc* 42: 245-251. PubMed: 8120307.
- Castle SC (2000) Clinical relevance of age-related immune dysfunction. *Clin Infect Dis* 31: 578-585. doi:10.1086/313947. PubMed: 10987724.
- Nagaratnam N, Gayagay G Jr. (2007) Validation of the Cumulative Illness Rating Scale (CIRS) in hospitalized nonagenarians. *Arch Gerontol Geriatr* 44: 29-36. doi:10.1016/j.archger.2006.02.002. PubMed: 16621072.
- Linn BS, Linn MW, Gurel L (1968) Cumulative illness rating scale. *J Am Geriatr Soc* 16: 622-626. PubMed: 5646906.
- Parmelee PA, Thuras PD, Katz IR, Lawton MP (1995) Validation of the Cumulative Illness Rating Scale in a geriatric residential population. *J Am Geriatr Soc* 43: 130-137. PubMed: 7836636.
- Miller MD, Paradis CF, Houck PR, Mazumdar S, Stack JA et al. (1992) Rating chronic medical illness burden in geropsychiatric practice and research: application of the Cumulative Illness Rating Scale. *Psychiatry Res* 41: 237-248. doi:10.1016/0165-1781(92)90005-N. PubMed: 1594710.
- Yesavage JA (1988) Geriatric Depression Scale. *Psychopharmacol Bull* 24: 709-711. PubMed: 3249773.
- Katz S, Ford AB, Moskowitz RW, Jackson BA, Jaffe MW (1963) Studies of illness in the aged. The index of ADL: a standardized measure of biological and psychosocial function. *JAMA* 185: 914-919. doi:10.1001/jama.1963.03060120024016. PubMed: 14044222.
- Guigoz Y, Vellas B, Garry PJ (1996) Assessing the nutritional status of the elderly: The Mini Nutritional Assessment as part of the geriatric evaluation. *Nutr Rev* 54: S59-S65. PubMed: 8919685.
- Vellas B, Villars H, Abellan G, Soto ME, Rolland Y et al. (2006) Overview of the MNA—Its history and challenges. *J Nutr Health Aging* 10: 456-463. PubMed: 17183418.
- Bautmans I, Gorus E, Njemini R, Mets T (2007) Handgrip performance in relation to self-perceived fatigue, physical functioning and circulating IL-6 in elderly persons without inflammation. *BMC Geriatr* 7: 5. doi: 10.1186/1471-2318-7-5. PubMed: 17331228.
- Bautmans I, Mets T (2005) A fatigue resistance test for elderly persons based on grip strength: reliability and comparison with healthy young subjects. *Aging Clin Exp Res* 17: 217-222. doi:10.1007/BF03324600. PubMed: 16110735.
- Miles EA, Rees D, Banerjee T, Cazzola R, Lewis S et al. (2008) Age-related increases in circulating inflammatory markers in men are independent of BMI, blood pressure and blood lipid concentrations. *Atherosclerosis* 196: 298-305. doi:10.1016/j.atherosclerosis.2006.11.002. PubMed: 17118371.
- Forsey RJ, Thompson JM, Ernerudh J, Hurst TL, Strindhall J et al. (2003) Plasma cytokine profiles in elderly humans. *Mech Ageing Dev* 124: 487-493. doi:10.1016/S0047-6374(03)00025-3. PubMed: 12714257.
- Cesari M, Penninx BW, Newman AB, Kritchevsky SB, Nicklas BJ et al. (2003) Inflammatory markers and onset of cardiovascular events: results from the Health ABC study. *Circulation* 108: 2317-2322. doi: 10.1161/01.CIR.0000097109.90783.FC. PubMed: 14568895.
- Risk Emerging Factors Collaboration, Kaptoge S, Di Angelantonio E, Pennells L (2012) C-reactive protein, fibrinogen, and cardiovascular

- disease prediction. *N Engl J Med* 367: 1310-1320. doi:10.1056/NEJMoa1107477. PubMed: 23034020.
32. Interleukin-6 Receptor Mendelian Randomisation Analysis (IL6R MR) Consortium, Hingorani AD (2012) The interleukin-6 receptor as a target for prevention of coronary heart disease: a mendelian randomisation analysis. *Lancet* 379: 1214-1224. doi:10.1016/S0140-6736(12)60110-X. PubMed: 22421340.
 33. Niu W, Liu Y, Qi Y, Wu Z, Zhu D et al. (2012) Association of interleukin-6 circulating levels with coronary artery disease: a meta-analysis implementing mendelian randomization approach. *Int J Cardiol* 157: 243-252. doi:10.1016/j.ijcard.2011.12.098. PubMed: 22261689.
 34. Zheng GH, Chen HY, Xiong SQ (2012) Polymorphisms of -174G>C and -572G>C in the interleukin 6 (IL-6) gene and coronary heart disease risk: a meta-analysis of 27 research studies. *PLOS ONE* 7: e34839. doi:10.1371/journal.pone.0034839. PubMed: 22509361.
 35. Hotamisligil GS (2010) Endoplasmic reticulum stress and atherosclerosis. *Nat Med* 16: 396-399. doi:10.1038/nm0410-396. PubMed: 20376052.
 36. Tedgui A, Mallat Z (2006) Cytokines in atherosclerosis: pathogenic and regulatory pathways. *Physiol Rev* 86: 515-581. doi:10.1152/physrev.00024.2005. PubMed: 16601268.
 37. Hristov M, Weber C (2011) Differential role of monocyte subsets in atherosclerosis. *Thromb Haemostasis* 106: 757-762. doi:10.1160/TH11-07-0500. PubMed: 21901241.
 38. Hristov M, Leyendecker T, Schuhmann C, von Hundelshausen P, Heussen N et al. (2010) Circulating monocyte subsets and cardiovascular risk factors in coronary artery disease. *Thromb Haemostasis* 104: 412-414. doi:10.1160/TH10-01-0069. PubMed: 20431842.
 39. Merino A, Buendia P, Martin-Malo A, Aljama P, Ramirez R et al. (2011) Senescent CD14+CD16+ monocytes exhibit proinflammatory and proatherosclerotic activity. *J Immunol* 186: 1809-1815. doi:10.4049/jimmunol.1001866. PubMed: 21191073.
 40. Cros J, Cagnard N, Woollard K, Patey N, Zhang SY et al. (2010) Human CD14dim monocytes patrol and sense nucleic acids and viruses via TLR7 and TLR8 receptors. *Immunity* 33: 375-386. doi:10.1016/j.immuni.2010.08.012. PubMed: 20832340.
 41. Chan G, Bivins-Smith ER, Smith MS, Yurochko AD (2009) NF-kappaB and phosphatidylinositol 3-kinase activity mediates the HCMV-induced atypical M1/M2 polarization of monocytes. *Virus Res* 144: 329-333. doi:10.1016/j.virusres.2009.04.026. PubMed: 19427341.
 42. Pawelec G, Derhovanessian E (2011) Role of CMV in immune senescence. *Virus Res* 157: 175-179. doi:10.1016/j.virusres.2010.09.010. PubMed: 20869407.
 43. Blankenberg S, Rupprecht HJ, Bickel C, Espinola-Klein C, Rippon G et al. (2001) Cytomegalovirus infection with interleukin-6 response predicts cardiac mortality in patients with coronary artery disease. *Circulation* 103: 2915-2921. doi:10.1161/01.CIR.103.24.2915. PubMed: 11413080.
 44. Mathei C, Vaes B, Wallemacq P, Degryse J (2011) Associations between cytomegalovirus infection and functional impairment and frailty in the BELFRAIL Cohort. *J Am Geriatr Soc* 59: 2201-2208. doi:10.1111/j.1532-5415.2011.03719.x. PubMed: 22092044.
 45. Bartlett DB, Firth CM, Phillips AC, Moss P, Baylis D et al. (2012) The age-related increase in low-grade systemic inflammation (Inflammaging) is not driven by cytomegalovirus infection. *Aging Cell* 11(5):912-5.
 46. Fried LP, Tangen CM, Walston J, Newman AB, Hirsch C et al. (2001) Frailty in older adults: evidence for a phenotype. *J Gerontol A Biol Sci Med Sci* 56: M146-M156. doi:10.1093/geron/56.3.M146. PubMed: 11253156.
 47. Walston J, McBurnie MA, Newman A, Tracy RP, Kop WJ et al. (2002) Frailty and activation of the inflammation and coagulation systems with and without clinical comorbidities: results from the Cardiovascular Health Study. *Arch Intern Med* 162: 2333-2341. doi:10.1001/archinte.162.20.2333. PubMed: 12418947.
 48. Puts MT, Visser M, Twisk JW, Deeg DJ, Lips P (2005) Endocrine and inflammatory markers as predictors of frailty. *Clin Endocrinol (Oxf)* 63: 403-411. doi:10.1111/j.1365-2265.2005.02355.x. PubMed: 16181232.
 49. Baylis D, Bartlett DB, Syddall HE, Ntani G, Gale CR et al. (2012) Immune-endocrine biomarkers as predictors of frailty and mortality: a 10-year longitudinal study in community-dwelling older people. *Age (Dordr)* 35(3):963-71.
 50. de Gonzalo-Calvo D, de Luxán-Delgado B, Rodríguez-González S, García-Macia M, Suárez FM et al. (2012) Interleukin 6, soluble tumor necrosis factor receptor I and red blood cell distribution width as biological markers of functional dependence in an elderly population: a translational approach. *Cytokine* 58: 193-198. doi:10.1016/j.cyto.2012.01.005. PubMed: 22309694.
 51. Buurman BM, van Munster BC, Korevaar JC, de Haan RJ, de Rooij SE (2011) Variability in measuring (instrumental) activities of daily living functioning and functional decline in hospitalized older medical patients: a systematic review. *J Clin Epidemiol* 64: 619-627. doi:10.1016/j.jclinepi.2010.07.005. PubMed: 21074969.
 52. Schaap LA, Pluijm SM, Deeg DJ, Visser M (2006) Inflammatory markers and loss of muscle mass (sarcopenia) and strength. *Am J Med* 119: 526-517. PubMed: 16750969.
 53. Ferrucci L, Penninx BW, Volpato S, Harris TB, Bandeen-Roche K et al. (2002) Change in muscle strength explains accelerated decline of physical function in older women with high interleukin-6 serum levels. *J Am Geriatr Soc* 50: 1947-1954. doi:10.1046/j.1532-5415.2002.50605.x. PubMed: 12473005.
 54. Blain H, Jausset A, Béziat S, Dupuy AM, Bernard PL et al. (2012) Low serum IL-6 is associated with high 6-minute walking performance in asymptomatic women aged 20 to 70years. *Exp Gerontol* 47: 143-148. doi:10.1016/j.exger.2011.11.008. PubMed: 22123428.
 55. Bautmans I, Njemini R, Predom H, Lemper JC, Mets T (2008) Muscle endurance in elderly nursing home residents is related to fatigue perception, mobility, and circulating tumor necrosis factor-alpha, interleukin-6, and heat shock protein 70. *J Am Geriatr Soc* 56: 389-396. doi:10.1111/j.1532-5415.2007.01571.x. PubMed: 18179479.
 56. Bautmans I, Onyema O, Van Puyvelde K, Pleck S, Mets T (2011) Grip work estimation during sustained maximal contraction: validity and relationship with dependency and inflammation in elderly persons. *J Nutr Health Aging* 15: 731-736. doi:10.1007/s12603-010-0317-1. PubMed: 21968873.
 57. Bauer ME, Jeckel CM, Luz C (2009) The role of stress factors during aging of the immune system. *Ann N Y Acad Sci* 1153: 139-152. doi:10.1111/j.1749-6632.2008.03966.x. PubMed: 19236337.
 58. Gouin JP, Hantsoo L, Kiecolt-Glaser JK (2008) Immune dysregulation and chronic stress among older adults: a review. *Neuroimmunomodulation* 15: 251-259. doi:10.1159/000156468. PubMed: 19047802.
 59. Baune BT, Smith E, Reppermund S, Air T, Samaras K et al. (2012) Inflammatory biomarkers predict depressive, but not anxiety symptoms during aging: The prospective Sydney Memory and Aging Study. *Psychoneuroendocrinology* 37(9):1521-30.
 60. Howren MB, Lamkin DM, Suls J (2009) Associations of depression with C-reactive protein, IL-1, and IL-6: a meta-analysis. *Psychosom Med* 71: 171-186. doi:10.1097/PSY.0b013e3181907c1b. PubMed: 19188531.
 61. Wright CB, Sacco RL, Rundek T, Delman J, Rabbani L et al. (2006) Interleukin-6 is associated with cognitive function: the Northern Manhattan Study. *J Stroke Cerebrovasc Dis* 15: 34-38. doi:10.1016/j.jstrokecerebrovasdis.2005.08.009. PubMed: 16501663.
 62. Licastro F, Pedrini S, Caputo L, Annoni G, Davis LJ et al. (2000) Increased plasma levels of interleukin-1, interleukin-6 and alpha-1-antichymotrypsin in patients with Alzheimer's disease: peripheral inflammation or signals from the brain? *J Neuroimmunol* 103: 97-102. doi:10.1016/S0165-5728(99)00226-X. PubMed: 10674995.
 63. Ravaglia G, Forti P, Maioli F, Chiappelli M, Montesi F et al. (2007) Blood inflammatory markers and risk of dementia: The ConSelice Study of Brain Aging. *Neurobiol Aging* 28: 1810-1820. doi:10.1016/j.neurobiolaging.2006.08.012. PubMed: 17011077.
 64. Zuliani G, Ranzini M, Guerra G, Rossi L, Munari MR et al. (2007) Plasma cytokines profile in older subjects with late onset Alzheimer's disease or vascular dementia. *J Psychiatr Res* 41: 686-693. doi:10.1016/j.psychires.2006.02.008. PubMed: 16600299.
 65. Uslu S, Akarkarasu ZE, Ozbabalik D, Ozkan S, Colak O et al. (2012) Levels of amyloid beta-42, interleukin-6 and tumor necrosis factor-alpha in Alzheimer's disease and vascular dementia. *Neurochem Res* 37: 1554-1559. doi:10.1007/s11064-012-0750-0. PubMed: 22437436.
 66. Dik MG, Jonker C, Hack CE, Smit JH, Comijs HC, Eikelenboom P (2005) Serum inflammatory proteins and cognitive decline in older persons. *Neurology* 64: 1371-1377. doi:10.1212/01.WNL.0000158281.08946.68. PubMed: 15851726.
 67. Bonotis K, Krikki E, Holeva V, Aggouridaki C, Costa V et al. (2008) Systemic immune aberrations in Alzheimer's disease patients. *J Neuroimmunol* 193: 183-187. doi:10.1016/j.jneuroim.2007.10.020. PubMed: 18037502.
 68. Nakajima T, Honda T, Domon H, Okui T, Kajita K et al. (2010) Periodontitis-associated up-regulation of systemic inflammatory mediator level may increase the risk of coronary heart disease. *J Periodontol Res* 45: 116-122. doi:10.1111/j.1600-0765.2009.01209.x. PubMed: 19602107.