



Effect of Saengshik Supplementation on the Gut Microbial Composition of Healthy Korean Adults: A Single-Group Pilot Study

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Saengshik is a type of meal-replacement product or dietary supplement comprising an uncooked and dried plant-based food mixture with various health-promoting properties, such as antidiabetic, anti-dyslipidemic, antioxidant, and anticancer properties. Although these properties are considered attributable to the various bioactive components absorbed through the intestine and its remolding effect on intestinal microorganisms, the effect of Saengshik supplementation on gut microbiota profiles has not yet been studied. In this study, we investigated the effect of Saengshik administration on the composition of gut microbiota. This single-group design trial was conducted on 102 healthy men and women who received 40 g/day of Saengshik powder for 8 weeks, during which stool samples were collected at two fixed time points (baseline and the endpoint) for gut microbiota-profiling analysis. We observed a significant decrease in the α -diversity of gut microbiota after Saengshik consumption (P < 0.05), with significant changes identified in the composition of major microbial taxa, such as *Bacteroidetes* (P < 0.0001), Proteobacteria, Actinobacteria, and Verrucomicrobia (P < 0.0001). Notably, the gut microbial response was related to the inter-individual variability of habitual dietary intake and enterotype at baseline. To the best of our knowledge, this is the first study investigating the effects of Saengshik intake on changes in gut microbiota, with the results suggesting that individual habitual diet patterns and gut microbial shapes should be considered key aspects in Saengshik-mediated health-promotion effects.

Keywords: gut microbiota, raw-food formula, plant-based dietary supplement, enterotype, Saengshik

INTRODUCTION

According to the Korea Food Standards Codex, *Saengshik* refers to a product processed from plant- or animal-derived materials by drying and can be consumed with beverages, such as water and milk (1). Commercially-available *Saengshik* mainly comprises 30–60 types of whole grains, vegetables, seaweeds, mushrooms, and other minor plant-based ingredients. Due to the process of freeze-drying or low-temperature drying of its ingredients, *Saengshik* preserves various nutrients, including minerals, vitamins, and phytochemicals in raw materials (2–9). Additionally, *Saengshik*

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consumption has increased based on its advantage of allowing convenient consumption of various plant-derived raw materials.

To date, several studies have reported the beneficial role of *Saengshik*. Given that *Saengshik* is generally a low-calorie meal replacement, anti-obesity effects have been reported through weight-loss studies in obese adults (2, 3). *Saengshik* also exhibits various bioactive functions, including antidiabetic (4, 5), anti-dyslipidemic, antioxidant (4, 6), and anticancer (7, 8) activities. Furthermore, Shin et al. (9) reported that *Seangshik* supplementation attenuated colitis in murine models of inflammatory bowel disease. These beneficial effects might be attributed to the functional ingredients of plant-based foods, such as phytochemicals, minerals, vitamins, and dietary fibers. However, despite evidence of a link between *Saengshik* consumption and health benefits, no intervention studies have assessed its modulation of the gut microbiome.

The role of gut microbiota cannot be overlooked in acquiring an understanding of the physiological effects of specific food components, because microorganisms are an important factor that influences nutrient utilization. The human gut harbors a vast community of microbes that include ~ 100 trillion bacterial cells, with these microorganisms playing pivotal roles in human health improvement, as well as the pathogenesis of diseases (10, 11). Diet is a major external factor in the variability of gut microbiota composition (12). Specifically, plant bioactive compounds, such as polyphenols and fibers, are transformed by gut microbiota into a bioactive form responsible for human health effects (13, 14). Thus, plant-based dietary modulation of the gut microbiome has been continuously reported in human and animal studies (15-18). Furthermore, emerging evidence suggests that discrepancies in gut microbiota responses to dietary interventions among individuals might be related to differences in both baseline gut microbiota composition and habitual dietary intake (19-21). Individual baseline composition of intestinal microflora is referred to as an enterotype, with this used to classify gut microbiota according to the proportion of dominant bacteria, such as Bacteroides, Prevotella, and Ruminococcus. A previous study reported that enterotypes are generally affected by individual long-term diet patterns, Bacteroides-dominated enterotypes associated with animal fat and protein intake, and Prevotelladominated enterotypes linked to a carbohydrate-rich diet (22). Therefore, individual gut microbial responses to Saengshik intake could be influenced by indigenous gut microbiota and habitual dietary intake.

The objectives of this study were to confirm changes in intestinal microflora after *Saengshik* intake and verify the hypothesis that these changes might be related to the status of habitual dietary intake and baseline gut microbiota composition.

MATERIALS AND METHODS

Study Participants

The participants were recruited using posters at Theragen Bio (Seongnam, Gyeonggi, South Korea). The eligible subjects were healthy men and women between 20 and 65 years of age.

Individuals who consumed pro-, pre-, or antibiotics within 3 months before the study were excluded. All participants provided written informed consent, and the study protocol was approved by the Institutional Review Board (IRB) of Theragen Bio (IRB No. 700062-20180905-JR-005-01).

Preparation of the Experimental Formula

Saengshik (Eromplus Cellfood Goodmorning Saengshik Premium) was provided by Erom Co., Ltd. (Seongnam, Gyeonggi, South Korea) and prepared by mixing freeze-dried and powdered ingredients and transferring them into plastic packaging. The formula contained grains, beans, vegetables, fruits, mushrooms, seaweed, and other diverse raw materials (**Supplementary Table 1**). The nutrient contents were measured at SGS Korea Co., Ltd. (Uiwang-si, Gyeonggi-do, Korea) and are summarized in **Supplementary Table 2**.

Experimental Design and Intervention

This study was a single-group trial conducted at Theragen Bio from October 2018 to February 2019. On day 1 of subject arrival, they were provided enough Saengshik powder in 40-g packages and enough for 8 weeks of consumption. Subjects were instructed to consume 40 g of Saengshik powder with 100-150 mL of water daily over the 8-week treatment period. Although not assigned a time to eat Saengshik, subjects were asked to report whether they ate Saengshik daily. After 8 weeks, subjects that failed consume Saengshik more than once were excluded from the analysis due to poor compliance. Additionally, all participants were instructed to maintain their regular diet patterns and avoid any pro- and prebiotic supplements during the study period. Fecal samples from the participants were collected using a commercial fecal sampling kit (OMNIgene GUT; OMR-200; DNA Genotek, ON, Canada) at two fixed time points (at baseline and the study endpoint) for gut microbiota-profiling analysis (Supplementary Figure 1). The instructions specified in the stool sampling kit were explained, with each subject instructed to collect a specific amount of stool.

Habitual Dietary Assessment

Habitual dietary intake was assessed at baseline using the food frequency questionnaire (FFQ) method, which was developed and validated by the Korea Centers for Disease Control and Prevention (23). The FFQ included 103 food or beverage items with three serving sizes and nine intake-frequency questions, with the survey used to assess the items consumed over the previous year. The validity and reproducibility of the FFQ has previously been verified (23). According to the food intake measured by the FFQ, the amount of calories and nutrients consumed daily by each individual was calculated and corrected to a level of 1000 kcal for use in creating subgroups based on nutrient intake.

16S rRNA Sequencing and Data Analysis

The metagenomic DNA of each stool sample was extracted with a QIAcube instrument using a QIAamp DNA stool mini kit (Qiagen, Hilden, Germany) according to manufacturer instructions along with an additional bead-beating procedure. Briefly, 250 aliquots of stool samples were transferred to a tube together with 1.2 mL of ASL lysis buffer and 0.3 g of sterile 0.1 mm zirconia beads (BioSpec, Bartlesville, OK, USA) and homogenized twice at 30 Hz for 1 min on the TissueLyser system (Qiagen) (24). The third to fourth hypervariable region (V3–V4) of the 16S rRNA gene from the extracted DNA sample was amplified and sequenced using the Illumina MiSeq 2×300 System (Illumina, San Diego, CA, USA).

The DADA2 pipeline (25) of the QIIME2 package (v.2019.1; https://qiime2.org) (26) was used to generate unique sequence variants by filtering low-quality sequences and chimeras. Taxonomy was assigned using a pre-trained Naïve Bayes classifier based on the Greengenes 13_8 (27) 99% operational taxonomic units (OTUs), which had been trimmed to include the V3–V4 region. Chimeric reads were filtered from the sequencing data using UCHIME software (28). After excluding samples with low sequencing depths (<10,000 reads), the gut microbial compositions were analyzed in 66 paired samples (29 men and 37 women) and compared before and after *Saengshik* supplementation. The relative abundance of bacterial genera was calculated from sequencing data at baseline, with this data used to establish the enterotype of the subject.

Statistical Analyses

Statistical analysis was performed using GraphPad Prism software (v.8.0.1; GraphPad Software Inc., San Diego, CA, USA) and R studio (v.1.1.447) (29). Differences in genera and phyla between before and after *Saengshik* supplementation were assessed using a Wilcox test in R studio using the "wilcox.test (paired=TRUE)" function from the "stats" R package. Significant changes in bacterial taxa between groups were calculated using a two-tailed Mann–Whitney *U* test (GraphPad Prism v.8.0). To cluster the gut microbial community of each subject, enterotype

assignment was performed based on a published tutorial (http://enterotype.embl.de/enterotypes.html) (22). Briefly, the 66 samples were clustered using the PAM (partitioning around medoids) clustering algorithm in the R package "cluster," and the optimal number of clusters was chosen based on the Calinski–Harabasz index using the "clusterSim" R-package. The result of enterotype clustering was visualized in a principal component analysis plot using the R package "ade4." The level of statistical significance was set at P < 0.05.

RESULTS

Baseline Characteristics

A total of 102 individuals who were assessed for eligibility were included in this study, with 11 participants withdrawing before the end of the study. After an 8-week treatment period, we performed 16S rRNA sequencing of fecal samples before and after intervention from 78 participants that completed the study. In total, 12 subjects (15.4%) withdrew from the data analysis owing to an insufficient number of sequencing reads (<10,000 reads) and low compliance. Therefore, the gut microbial composition of 66 participants at baseline was compared with that at post-intervention, as presented in the schematic procedure of the intervention study (Figure 1). Table 1 shows the daily energy and nutrient intake of participants at baseline. At baseline, participants demonstrated a lower intake of energy, carbohydrates, proteins, fats, vitamin B₂, folate, calcium, phosphorus, potassium, iron, and zinc than that of an average Korean, although intake of vitamins A and C were higher than that of the average Korean. Daily intake of vitamin B1 and niacin were similar to the Korean average (Table 1). The baseline characteristics of the participants are shown in Table 2. The 66 participants comprised 29 men and 37 women, with a mean age of 40.89 \pm 9.94 years.

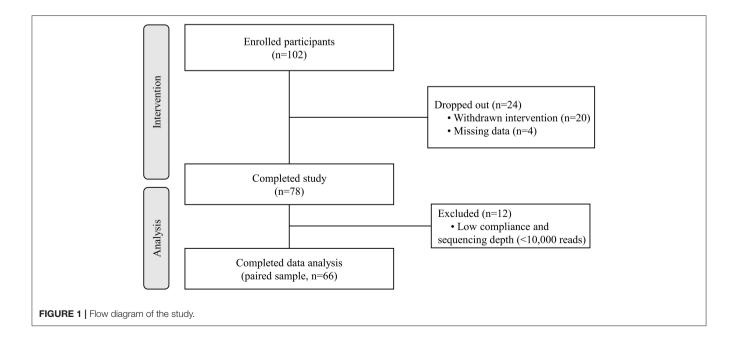


TABLE 1 | The nutrient intake (g/day) of participants at baseline.

	All
	(<i>n</i> = 66)
Energy (Kcal)	1560.25 ± 703.22
Carbohydrate (g/d)	250.1 ± 128.40
Protein (g/d)	55.93 ± 22.59
Fat (g/d)	36.26 ± 18.48
Cholesterol (mg/d)	206.2 ± 102.40
Fiber (g/d)	4.237 ± 2.93
Vitamin B1 (mg/d)	1.035 ± 0.51
Vitamin B2 (mg/d)	0.8499 ± 0.38
Vitamin B6 (mg/d)	1.316 ± 0.68
Folate (ug/d)	177.1 ± 121.20
Niacin (mg/d)	12.83 ± 5.32
Vitamin C (mg/d)	79.82 ± 74.95
Vitamin A (R.E/d)	403.3 ± 277.80
Retinol (ug/d)	82.39 ± 49.32
Carotene (ug/d)	1810 ± 1533
Vitamin E (mg/d)	7.682 ± 4.12
Calcium (mg/d)	381.5 ± 213.90
Phosphorus (mg/d)	776.3 ± 305.70
Potassium (mg/d)	1813 ± 934.30
Iron (mg/d)	9.183 ± 4.66
Sodium (mg/d)	1982 ± 1160
Zinc (mg/d)	6.797 ± 2.73
Ash (mg/d)	12.74 ± 7.00

Values are means and standard deviations.

TABLE 2 | General characteristics of participants at baseline.

	All (n = 66)
Characteristics	
Men; n (%)	29 (43.94)
Women; n (%)	37 (56.06)
Age (yr)	40.89 ± 9.94
Age group (yr)	
20–29; n (%)	7 (10.61)
30–39; n (%)	28 (42.42)
40–49; n (%)	19 (28.79)
50–59; n (%)	8 (12.12)
60–69; n (%)	4 (6.06)
Height (cm)	166.65 ± 7.59
Weight (kg)	64.86 ± 11.77
BMI (kg/m ²)	23.22 ± 2.96

Values are means and standard deviations.

Effect of *Saengshik* Consumption on Gut Microbial Diversity and Composition

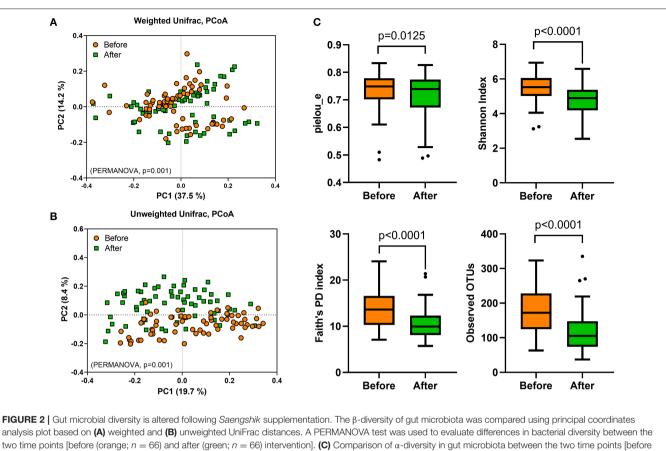
To evaluate the effect of *Saengshik* administration on gut microbiota profiles, we performed 16S rRNA sequencing analysis of stool samples collected at baseline and post-intervention. First, we evaluated β -diversity by calculating the weighted and

unweighted unique fraction metric (UniFrac) distances, finding that the overall gut microbial community differed significantly between baseline and post-intervention (weighted F = 6.365, P = 0.001; unweighted F = 7.489, P = 0.001; PERMANOVA) (**Figures 2A,B**). To measure differences in diversity and evenness between baseline and post-intervention samples, we calculated Pielou's evenness (pielou_e), Shannon, Faith's phylogenetic diversity (Faith's PD), and observed OTU indices, which revealed significant decreases in α -diversity following intervention as compared with that at baseline (pielou_e, P = 0.0125; Shannon, P < 0.0001; Faith's PD, P < 0.0001; observed OTUs, P < 0.0001; Wilcoxon signed-rank test) (**Figure 2C**).

To detect the specific bacterial taxa that were significantly affected by Saengshik consumption, we compared the composition of gut microbiota at the phylum and genus levels before and after intervention. Figure 3A shows the relative abundance (%) of each bacterial phylum of participants as different color bars. The distribution of Bacteroidetes increased after Saengshik consumption, whereas that of Verrucomicrobia, Proteobacteria, and Actinobacteria decreased, with statistically significant changes observed in the relative abundance of all phyla [Bacteroidetes (P = 0.0002), Proteobacteria (P <0.0001), Actinobacteria (P < 0.0001), and Verucomicrobia (P = 0.0093)] (Figure 3B). At the genus level, we identified 23 genera showing significant differences in abundance between baseline and post-intervention (P < 0.01, Wilcoxon signed-rank test), with Figure 3C showing fold changes for each genus. The relative abundance of Faecalibacterium, Bacteroides, and Bifidobacterium significantly increased after Saengshik consumption, whereas that of Alistipes, Megamnas, Odoribacter, Oscillospira, Anaerotruncus, Sutterella, Dialister, Butyricicoccus, Bilophila, Phascolarctobacterium, (Eubacterium), Fusobacterium, Veillonella, Holdemania, Butyricimonas, Akkermansia, Streptococcus, Collinsella, Weissella, and Clostridium significantly decreased (Figure 3C). Collectively, these data demonstrate that daily intake of a plant-based food powder altered gut microbial communities and decreased gut microbial diversity. Furthermore, we identified specific bacterial phyla and genera that were significantly affected by Saengshik consumption.

Effect of Individual Enterotype on Changes in the Gut Microbial Composition After *Saengshik* Consumption

The human gut microbiome reportedly comprises two or three strains of major indigenous bacteria, which are referred to as the enterotype. Enterotypes reportedly reflect the unique gut microbial characteristics of individuals and are unaffected by body mass index (BMI), sex, or race. To verify whether enterotypes can affect compositional changes in gut microbiota after *Seangshik* consumption, we classified participant enterotypes and compared changes in gut microbiota among enterotypes. Following identification of a Calinski-Harabasz index of k = 2 as an optimal number of clusters, we confirmed that 66 participants were separated into two distinct clusters (**Figures 4A,B**), of which 43 belonged to the



two time points [before (orange; n = 66) and after (green; n = 66) intervention]. (C) Comparison of α -diversity in gut microbiota between the two time points [before (orange; n = 66) and after (green; n = 66) intervention]. Tukey's box-and-whisker plots show the median (horizontal line) and interquartile range (IQR) (box), with additional data within 1.5 IQR shown as whiskers. The Pielou's evenness, Shannon, Faith's PD, and observed OTU indices of the samples were statistically compared between the two time points using the Wilcoxon rank-sum test.

Bacteroides-rich enterotype (B-type) and 23 to the Prevotellarich enterotype (P-type). The relative abundance of Bacteroides was significantly greater in B-type than that in P-type (P < P0.0001), whereas Prevotella was more abundant in P-type than in B-type (*P* < 0.0001) (Supplementary Figure 2). Additionally, we observed no statistically significant differences in age, height, weight, or BMI between individuals characterized as B- or P-type (all P > 0.05) (Supplementary Table 3). We then examined whether changes in the relative abundance of genera Figure 3C differed significantly according to enterotype before and after Saengshik intake. Figure 4C shows that the relative abundance of *Bacteroides* (P = 0.028) was significantly higher in B-type individuals relative to P-type individuals after Seangshik consumption, whereas the relative abundance of Eggerthella (P = 0.010) and Fusobacterium (P = 0.021) decreased significantly decreased in B-type relative to P-type individuals. However, Dialister (P = 0.020), Anaerostipes (P = 0.022), and Catenibacterium (P = 0.035) decreased significantly in P-type relative to B-type individuals after Seangshik consumption (Figure 4C). These observations suggest that enterotypes affected the response of gut microbial composition to Saengshik consumption.

Effect of Habitual Dietary Patterns on Changes in Gut Microbial Composition After Saengshik Consumption

To investigate whether habitual dietary patterns affect changes in gut microbial composition after Saengshik consumption, we compared changes in gut microbiota among subgroups classified by habitual dietary patterns. Figure 5A shows the results of clustering each subject based on the amount of micronutrient intake per 1000 kcal/day at baseline and according to FFQ data. We specified these two subgroups as dietary pattern 1 (DP1; n = 31) and DP2 (n = 35). There were no significant differences in age, height, weight, or BMI between DP1and DP2 (all P > 0.05) (**Supplementary Table 4**). Those in DP1consumed higher amounts of carbohydrates, fiber, vitamin C, and vitamin E, whereas those in the DP2consumed higher amounts of protein, fat, cholesterol, vitamin B₁, retinol, carotene, and zinc (all P < 0.05) (Supplementary Table 5). We then determined whether changes in the relative abundance of genera (Figure 3C) differed significantly according to individual dietary pattern in each subgroup and compared these changes before and after Saengshik intake. The results showed that the relative abundance of Faecalibacterium (P = 0.043), Clostridium (P = 0.015), and

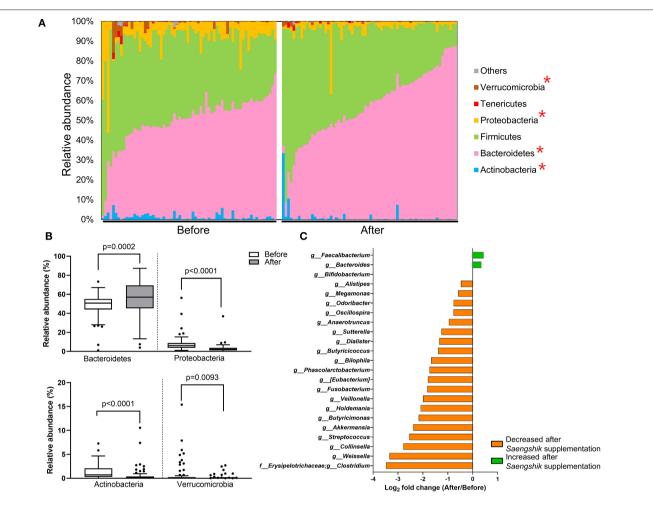
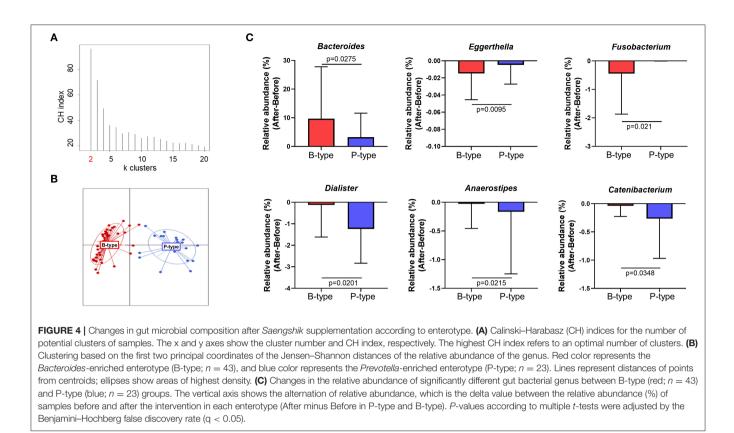


FIGURE 3 | The relative abundance of bacterial taxa is affected by *Saengshik* intervention. (A) Distribution of gut microbial phyla before (n = 66) and after (n = 66) intervention. Each bar represents an individual participant. The proportion of the various taxa in each phylum is presented as a percentage of relative abundance with respect to total bacterial sequences. Different color schemes represent different phyla (blue: Actinobacteria, pink: Bacteroidetes, green: Firmicutes, yellow: Proteobacteria, red: Tenericutes, brown: Verrucomicrobia, gray: others). (B) Differences in the mean relative abundance of phyla before (n = 66) and after (n = 66) intervention. Tukey's box-and-whisker plots show the median (horizontal line) and interquartile range (IQR) (box), with additional data within 1.5 IQR shown as whiskers. The white and gray box plots represent the mean relative abundance of phyla before and after (n = 66) intervention. Tukey's box-and-whisker plots show the median (horizontal line) and interquartile range (IQR) (box), with additional data within 1.5 IQR shown as whiskers. The white and gray box plots represent the mean relative abundance of phyla before and after the intervention, respectively. (C) List of significantly changed genera according to genera found before (n = 66) and after (n = 66) intervention. Box plots of relative abundance show the fold change after vs. before intervention on the log₂ scale. P < 0.01, Wilcoxon signed-rank test. The green and orange bars represent the bacterial genera that increased and decreased in abundance after *Saengshik* consumption, respectively.

Coprobacillus (P = 0.036) was significantly higher in DP2 than in DP1 after *Seangshik* consumption (**Figure 5B**). Moreover, the relative abundance of *Oscillospira* decreased significantly in DP2 as compared with that in DP1 (P = 0.023); however, *Veillonella* decreased significantly in DP1 as compared with that in DP2 (P = 0.037) (**Figure 5B**). These results indicate that compositional changes in gut microbiota after *Saengshik* consumption were differentially affected by individual dietary patterns.

Collective Effect of Individual Enterotype and Habitual Dietary Patterns on Changes in Specific Genera Compositions

To determine whether enterotype and habitual dietary patterns can collectively influence changes in gut microbial genera after Saengshik consumption, we compared changes in 23 genera (**Figure 3C**) between subjects with different dietary patterns in the same enterotype. In the B-type, 20 and 23 individuals belonged to the DP1 and DP2 clusters, respectively, whereas in the P-type, 11 and 12 individuals belonged to each respective cluster (**Supplementary Figure 3**). Notably, three of 29 genera were affected by enterotypes and habitual dietary patterns. After intervention and among B-type individuals, the relative abundance of *Faecalibacterium* (P = 0.018, Mann–Whitney U test) and Veillonella (P = 0.005, Mann–Whitney U test) increased in DP2 individuals but not in DP1 individuals. There was no difference in changes in the abundance of these two genera between DP1 and DP2 P-type individuals (**Figures 6A,B**). Among P-type individuals, DP2 showed a larger decrease in the



relative abundance of *Bilophila* than DP1 individuals (P = 0.042, Mann–Whitney *U* test) (**Figure 6C**). Collectively, these results indicate that *Saengshik* consumption changed gut microbial communities, which were influenced by not only individual habitual dietary patterns but also enterotype.

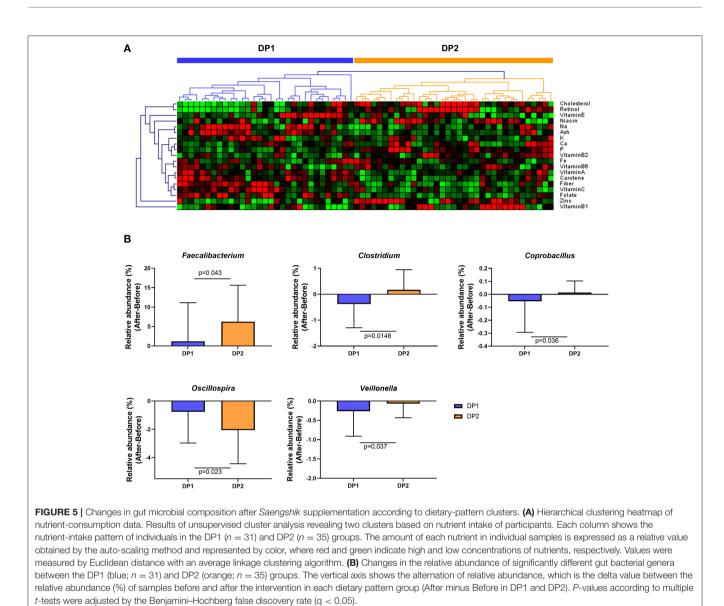
DISCUSSION

This study provides new insights into the effects of *Saengshik*, a plant-based commercial powder, in altering the composition of fecal microbiota in healthy adults. The results confirmed the hypothesis that *Saengshik* can directly alter specific bacterial taxa, and that these changes depend on individual baseline gut microbiome and diet patterns. Moreover, bacterial diversity after intervention was significantly lower than that before, and importantly, the relative abundance of *Faecalibacterium*, *Veillonella*, and *Bilophila* was differentially influenced by a combination of baseline gut microbiota and diet patterns. Thus, these findings suggest that baseline gut microbiota and diet patterns can promote modifications in gut microbiota composition following supplementation.

Saengshik Consumption and Gut Microbial Diversity

Previous studies revealed that plant-rich diets are related to increased bacterial diversity because of their capability to provide a source of microbiota-accessible carbohydrates (30, 31). Compared with plant-rich diets, a Western-style diet

is characterized by high intake of saturated fat and simple carbohydrates, whereas low intake of fiber is associated with reduced bacterial diversity (32). Additionally, microbial diversity is inversely associated with chronic diseases and metabolic dysfunction (33, 34). Therefore, we expected that Saengshik intake would improve gut microbial diversity; however, contrary to our expectation, we observed significant decreases in gut microbial diversity after 8 weeks of Saengshik consumption relative to that at baseline. Given that the average daily fiber intake of Korean adults is 24.3 g (35), we speculated that the amount of fiber contained in the product used in this study (Supplementary Table 2) would not contribute to increasing the diversity of intestinal microbes. Multiple in vivo and in vitro studies have revealed that phenolic compounds and flavonoids exhibit antibacterial activity against several pathogenic bacteria (36, 37). However, discrepancies in the antibacterial properties of even the same phenolic compounds have been reported among bacterial strains (38). Therefore, the antimicrobial activity of various phenolic compounds and flavonoids present in Saengshik might selectively affect different strains of gut microbiota, which might have contributed to the reduced diversity observed in the present study. Another possibility is that the various food components consumed as complex combinations in daily meals during the intervention period might have had an interactive effect (39), although this factor was not ruled out in the present study. Furthermore, the present study was conducted in healthy adults who already showed relatively high gut microbial diversity. The possibility of individuals with low bacterial diversity being



more responsive to *Saengshik* intake should be considered. Additionally, some interventions or cross-sectional studies report contradictory findings, including cases in which changes in diversity were not observed (40–42) or those in which diversity decreased after a plant-based diet (43, 44). Thus, to confirm the effect of *Saengshik* intake on gut microbial diversity, well-controlled and designed clinical trials are needed for individuals with decreased microbial diversity.

The Physiological Role of Specific Gut Microbial Taxa Which Changed by Saengshik Intake

We found that the population of *Faecalibacterium* increased significantly after *Saengshik* consumption, which might be related to positive effects on health. This result is consistent with the correlation between *Faecalibacterium* and plant-food intake

revealed reported in previous studies (40, 45). Faecalibacterium can utilize non-digestible carbohydrates (dietary fiber) from host mucus and concomitantly produce butyrate, which is associated with several beneficial health properties (46). This might be a factor related to the beneficial effects of Saengshik, including its anti-obesity and anti-inflammatory effects. Moreover, recent studies demonstrated an inverse association between Faecalibacterium and the risk of diseases, such as obesity (47), inflammatory colitis (48), and diabetes (49). In the present study, we also showed that the abundance of Akkermansia and Collinsella decreased after Seangshik consumption. Akkermansia exhibits probiotic potential that has been linked to maintaining intestinal barrier integrity, antiinflammatory properties, and immune function (50). By contrast, other studies reported that Akkermansia spp. exacerbated gut inflammation in an animal infection model based on their ability to degrade mucins (51). Furthermore, elevated Collinsella

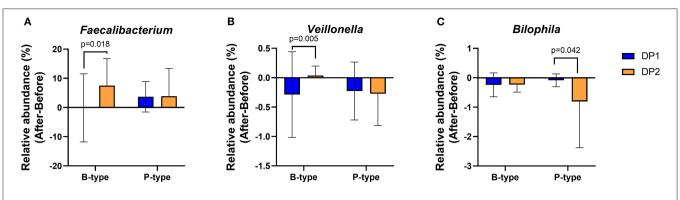


FIGURE 6 | Changes in gut microbial composition after *Saengshik* supplementation according to enterotype and dietary-pattern clusters. Changes in the relative abundance of **(A)** *Faecalibacterium*, **(B)** *Veillonella*, and **(C)** *Bilophila* according to enterotype and habitual dietary-pattern group. The vertical axis shows the alternation of relative abundance, which is the delta value between the relative abundance (%) of samples before and after the intervention in each dietary pattern and enterotype group (After minus Before in B-type with DP1, B-type with DP2, P-type with DP1, and P-type with DP2). *P*-values according to multiple *t*-tests were adjusted by the Benjamini–Hochberg false discovery rate (q < 0.05). B-type: DP1 (n = 20) and DP2 (n = 23). P-type: DP1 (n = 11) and DP2 (n = 12).

abundance is reportedly positively associated with increased levels of insulin, cholesterol, and triglycerides (52) and negatively correlated with vegetable intake (53, 54). In line with previous studies, the present results suggest that a lower abundance of *Collinsella* might be mediated by *Saengshik* consumption. Nevertheless, the reduced abundance of beneficial microbes, including *Butyricimonas, Megamonas, Phascolarctobacterium,* and *Butirycicoccus*, should be verified through further studies. Collectively, although it is difficult to conclude that the changes in intestinal microflora caused by *Saengshik* would have a positive effect on host health, these findings show a possible link between gut microbiota, *Saengshik* consumption, and physiological properties.

The Relationship Between Nutrients Contained in *Saengshik* and Intestinal Microflora

Saengshik contains many micronutrients and compounds, including phenolics and flavonoids. Studies suggest that the gut microbiome-altering effects of Saengshik might originate from a variety of these ingredients. Indeed, several clinical trials report that polyphenol consumption can modulate gut microbial composition, anthropometric variables, and clinical markers (55). In the present study, we found that Saengshik consumption increased the abundance of Bacteroidetes, Bifidobacterium, and Bacteroides. Consistent with these findings, an intervention study reported increases in these microbes following intake of polyphenol-rich foods, such as red wine, freeze-dried cranberry powder, fruits, and vegetables (15, 56-58). Moreover, Bacteroides and Bifidobacterium are negatively correlated with various clinical markers, such as tumor necrosis factor- α , cholesterol, blood pressure, and lipopolysaccharide (55) levels. These results support our findings that Saengshik might be involved in beneficial changes in biological markers. In addition to being a good source of flavonoids and phenolic compounds, Saengshik is high in folate. The serving size of Saengshik (40 g) used in this study contained folate at a level of 83% of the recommended dietary allowance for Korean adults (400 μ g) (59); therefore, the benefit of a sufficient folate intake could be expected. Mammals cannot synthesize their own folate *de novo*; therefore, they need to obtain folate via the diet and gut microbes, such as *Lactobacillus* and *Bifidobacterium* (60). Mice with folate deficiency reportedly show a significant increase in the *Firmicutes:Bacteroidetes* ratio (61), and cross-talk between gut microbiota and folate can modulate the regeneration of intestinal epithelial cells through stem cells by epigenetic modification (62). Although it remains unclear whether dietary folate intake is directly attributable to the composition and metabolism of gut microbiota (63), the consumption of sufficient folate (i.e., >3-fold greater than the habitual intake of the study participants) might contribute to microbial changes.

Effect of Individual Enterotype and Dietary Pattern on Changes in Intestinal Microbiota Caused by Saengshik Consumption

A notable finding in the present study was the inter-individual variability in response to the intervention, which was similar to results from clinical trials investigating the effect of diet on gut microbiota. Here, we showed that B-type individuals in the DP2 group showed a stronger increase in Faecalibacterium levels than those with DP1 group. Faecalibacterium reportedly plays a crucial role in maintaining human health owing to its production of anti-inflammatory metabolites or short-chain fatty acids. Indeed, Faecalibacterium is less abundant in patients with many diseases relative to healthy populations (47-49). Additionally, we showed that Veillonella abundance increased in individuals with a B-type enterotype and those with unhealthy dietary habits, whereas it decreased in individuals belonging to the B-type and DP1 groups. A recent study on healthy nonobese adults reported that increased abundance of Veillonella was negatively associated with saturated fatty acid consumption and body fat percentage (64). Furthermore, Veillonella abundance increases in women with high fiber intake, with its abundance associated with improved insulin sensitivity (65). Based on these findings, we suggest that adults with "less healthy" diet patterns and a *Bacteroides*-dominant enterotype might experience greater *Saengshik*-mediated positive health benefits. Moreover, these results suggest that further investigation of the role of habitual dietary patterns or enterotypes in evaluating dietary intervention is warranted.

Study Limitations

This study has several limitations. First, given to the descriptive nature of the study and the absence of related clinical outcomes, we cannot provide insights into the beneficial properties of changed bacterial taxa related to *Saengshik* consumption. Second, the study lacked use of a placebo or blank control and presented a small sample size. Third, given that we did not collect 3-day or 24-h diet records of participants, we cannot completely rule out the possibility that differences in the usual dietary intake before and after intervention could have affected the outcomes. Finally, pre-selected participants with dysbiosis or low bacterial diversity rather than heathy individuals might be more suitable for identifying the effect of *Saengshik* on gut microbiota. Therefore, well-designed and controlled clinical trials with a larger sample size are necessary to determine the biological effect of *Saengshik* on the gut microbial community.

Despite these limitations, this represents the first study providing evidence that *Saengshik* consumption by healthy adults can influence gut microbial composition and diversity. Furthermore, the observed compositional changes were influenced by not only baseline gut microbial composition (enterotype) but also habitual dietary intake. We believe that this study underlines the need to investigate both baseline gut microbiota composition and diet patterns to allow the design of personalized interventions for diet-induced changes in gut microbiota and human health.

CONCLUSION

In summary, we showed that *Saengshik* consumption altered gut microbial composition and diversity. The results demonstrated that in healthy Korean adults, the effect of *Saengshik* intake on gut microbial composition was dependent on individual

REFERENCES

- 1. *The Korea Food Standards Codex*. Cheongju: Korea Ministry of Food Drug Safety (2018).
- Chang Y-K, Park Y-S, Park M-H, Lee J-H, Kim S-H. The effects of low-calorie diet with raw-food formula on obesity and its complications in the obese premenopausal women. *J Commun Nutr.* (2002) 4:99–108. Available online at: https://www.koreascience.or.kr/article/JAKO200211921404809.page
- 3. Ha TY, Kim NY. The effects of uncooked grains and vegetables with mainly brown rice on weight control and serum components in Korean overweight/obese female. *Korean J Nutr.* (2003) 36:183–90.
- 4. Jang Y, Lee JH, Kim OY, Park HY, Lee SY. Consumption of whole grain and legume powder reduces insulin demand, lipid peroxidation, and plasma homocysteine concentrations in patients with coronary artery disease:

enterotype and dietary patterns at baseline. Although the precise mechanisms responsible for this outcome remain unclear, their identification could potentially yield new approaches for maintaining overall health linked to gut microbiota.

DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found at: https://dataview.ncbi.nlm. nih.gov/object/PRJNA747737.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Institutional Review Board (IRB) of Theragen Bio (IRB No. 700062-20180905-JR-005-01). The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

Y-DN and YJA conceived and designed the project. J-HK and MHP provided an experimental supplement and technical support. MYL and YJA collected the samples and managed the intervention study. J-HS, W-HC, and SPH conducted data analysis. J-HS, W-HC, and Y-DN wrote the manuscript. All authors read and approved the final version of the manuscript.

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SUPPLEMENTARY MATERIAL

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randomized controlled clinical trial. Arterioscler Thromb Vasc Biol. (2001) 21:2065–71. doi: 10.1161/hq1201.100258

- Kim M, Kim ES, Park MH, Hwang SJ, Jeong Y. Saengshik, a formulated health food, decreases blood glucose and increases survival rate in streptozotocin-induced diabetic rats. J Med Food. (2004) 7:162–7. doi: 10.1089/1096620041224030
- Kim HY, Kim JH, Lee SA, Chang HE, Park MH, Hwang SJ, et al. Saengshik, a formulated health food, prevents liver damage in CCl4-induced mice and increases antioxidant activity in elderly women. *J Med Food*. (2008) 11:323–30. doi: 10.1089/jmf.2007.549
- Lee S-A, Kim J-H, Kim W-Y, Hwang S-J, Park M-H. Saengshik administration reduced the side effects of chemotherapy in chemotherapeutic agent injected mice. *Cancer Prev Res.* (2007) 12:319–28. Available online at: https://www. jcpjournal.org/journal/view.html?spage=319&volume=12&number=4

- Kim H-Y, Lee S-A, Kim J-H, Chung J-S, Park M-H, Hwang S-J, et al. Effect of saengshik on azoxymethane-induced colon carcinogenesis and colonic mucosa inflammatory responses in F344 rats. *Cancer Prev Res.* (2009) 14:248–55. Available online at: https://www.jcpjournal.org/journal/ view.html?volume=14&number=3&spage=248&vmd=A
- Shin SH, Song JL, Park MG, Park MH, Hwang SJ, Park KY. Effects of natural raw meal (NRM) on high-fat diet and dextran sulfate sodium (DSS)induced ulcerative colitis in C57BL/6J mice. *Nutr Res Pract.* (2015) 9:619–27. doi: 10.4162/nrp.2015.9.6.619
- Ley RE, Peterson DA, Gordon JI. Ecological and evolutionary forces shaping microbial diversity in the human intestine. *Cell.* (2006) 124:837–48. doi: 10.1016/j.cell.2006.02.017
- Wu GD, Lewis JD. Analysis of the human gut microbiome and association with disease. *Clin Gastroenterol Hepatol.* (2013) 11:774–7. doi: 10.1016/j.cgh.2013.03.038
- Scott KP, Gratz SW, Sheridan PO, Flint HJ, Duncan SH. The influence of diet on the gut microbiota. *Pharmacol Res.* (2013) 69:52–60. doi: 10.1016/j.phrs.2012.10.020
- Kumar Singh A, Cabral C, Kumar R, Ganguly R, Kumar Rana H, Gupta A, et al. Beneficial effects of dietary polyphenols on gut microbiota and strategies to improve delivery efficiency. *Nutrients.* (2019) 11:2216. doi: 10.3390/nu11092216
- Bayram B, González-Sarrías A, Istas G, Garcia-Aloy M, Morand C, Tuohy K, et al. Breakthroughs in the health effects of plant food bioactives: a perspective on microbiomics, nutri (epi) genomics, and metabolomics. *J Agric Food Chem.* (2018) 66:10686–92. doi: 10.1021/acs.jafc.8b03385
- Klinder A, Shen Q, Heppel S, Lovegrove JA, Rowland I, Tuohy KM. Impact of increasing fruit and vegetables and flavonoid intake on the human gut microbiota. *Food Funct.* (2016) 7:1788–96. doi: 10.1039/C5FO01096A
- David LA, Maurice CF, Carmody RN, Gootenberg DB, Button JE, Wolfe BE, et al. Diet rapidly and reproducibly alters the human gut microbiome. *Nature*. (2014) 505:559–63. doi: 10.1038/nature12820
- Rodríguez-Daza MC, Roquim M, Dudonné S, Pilon G, Levy E, Marette A, et al. Berry polyphenols and fibers modulate distinct microbial metabolic functions and gut microbiota enterotype-like clustering in obese mice. *Front Microbiol.* (2020) 11:2032. doi: 10.3389/fmicb.2020.02032
- Nagano T, Yano H. Dietary cellulose nanofiber modulates obesity and gut microbiota in high-fat-fed mice. *Bioact Carbohydr Diet Fibre*. (2020) 22:100214. doi: 10.1016/j.bcdf.2020.100214
- Healey GR, Murphy R, Brough L, Butts CA, Coad J. Interindividual variability in gut microbiota and host response to dietary interventions. *Nutr Rev.* (2017) 75:1059–80. doi: 10.1093/nutrit/nux062
- Shin JH, Jung S, Kim SA, Kang MS, Kim MS, Joung H, et al. Differential effects of typical Korean versus American-style diets on gut microbial composition and metabolic profile in healthy overweight Koreans: a randomized crossover trial. *Nutrients.* (2019) 11:2450. doi: 10.3390/nu11102450
- 21. Healey G, Murphy R, Butts C, Brough L, Whelan K, Coad J. Habitual dietary fibre intake influences gut microbiota response to an inulin-type fructan prebiotic: a randomised, double-blind, placebo-controlled, cross-over, human intervention study. Br J Nutr. (2018) 119:176–89. doi: 10.1017/S0007114517003440
- Arumugam M, Raes J, Pelletier E, Paslier DL, Yamada T, Mende DR, et al. Enterotypes of the human gut microbiome. *Nature*. (2011) 473.7346: 174–80. doi: 10.1038/nature09944
- Ahn Y, Kwon E, Shim JE, Park MK, Joo Y, Kimm K, et al. Validation and reproducibility of food frequency questionnaire for Korean genome epidemiologic study. *Eur J Clin Nutr.* (2007) 61:1435–41. doi: 10.1038/sj.ejcn.1602657
- Lim MY, Song EJ, Kim SH, Lee J, Nam YD. Comparison of DNA extraction methods for human gut microbial community profiling. *Syst Appl Microbiol.* (2018) 41:151–7. doi: 10.1016/j.syapm.2017.11.008
- Callahan BJ, McMurdie PJ, Rosen MJ, Han AW, Johnson AJA, Holmes SP. DADA2: high-resolution sample inference from Illumina amplicon data. *Nat Methods*. (2016) 13:581–3. doi: 10.1038/nmeth.3869
- Bolyen E, Rideout JR, Dillon MR, Bokulich NA, Abnet CC, Al-Ghalith GA, et al. Reproducible, interactive, scalable and extensible microbiome data science using QIIME 2. Nat Biotechnol. (2019) 37:852–7. doi: 10.1038/s41587-019-0209-9

- McDonald D, Price MN, Goodrich J, Nawrocki EP, DeSantis TZ, Probst A, et al. An improved Greengenes taxonomy with explicit ranks for ecological and evolutionary analyses of bacteria and archaea. *ISME J.* (2012) 6:610–8. doi: 10.1038/ismej.2011.139
- Edgar RC, Haas BJ, Clemente JC, Quince C, Knight R. UCHIME improves sensitivity and speed of chimera detection. *Bioinformatics*. (2011) 27:2194– 200. doi: 10.1093/bioinformatics/btr381
- RStudio Team. RStudio: Integrated Development Environment for R. Boston, MA: R Studio Inc. (2020). Available online at: http://www.rstudio.com/2019 (accessed October 5, 2021).
- Sonnenburg ED, Smits SA, Tikhonov M, Higginbottom SK, Wingreen NS, Sonnenburg JL. Diet-induced extinctions in the gut microbiota compound over generations. *Nature*. (2016) 529:212–5. doi: 10.1038/nature16504
- Gentile CL, Weir TL. The gut microbiota at the intersection of diet and human health. *Science*. (2018) 362:776–80. doi: 10.1126/science.aau5812
- Nagpal R, Shively CA, Appt SA, Register TC, Michalson KT, Vitolins MZ, et al. Gut microbiome composition in non-human primates consuming a Western or Mediterranean diet. *Front Nutr.* (2018) 5:28. doi: 10.3389/fnut.2018.00028
- Huttenhower C, Gevers D, Knight R, Abubucker S, Badger JH, Chinwalla AT, et al. Structure, function and diversity of the healthy human microbiome. *Nature*. (2012) 486:207–14. doi: 10.1038/nature11234
- Lozupone CA, Stombaugh JI, Gordon JI, Jansson JK, Knight R. Diversity, stability and resilience of the human gut microbiota. *Nature*. (2012) 489:220– 30. doi: 10.1038/nature11550
- Song S, Song Y. Dietary fiber and its source are associated with cardiovascular risk factors in Korean adults. *Nutrients*. (2021) 13:160. doi: 10.3390/nu13010160
- 36. Cardoso RR, Neto RO, dos Santos D'Almeida CT, do Nascimento TP, Pressete CG, Azevedo L, et al. Kombuchas from green and black teas have different phenolic profile, which impacts their antioxidant capacities, antibacterial and antiproliferative activities. *Food Res Int.* (2020) 128:108782. doi: 10.1016/j.foodres.2019.108782
- 37. Righi N, Boumerfeg S, Fernandes PA, Deghima A, Baali F, Coelho E, et al. Thymus algeriensis Bioss & Reut: Relationship of phenolic compounds composition with *in vitro/in vivo* antioxidant and antibacterial activity. *Food Res Int.* (2020) 136:109500. doi: 10.1016/j.foodres.2020.109500
- Puupponen-Pimiä R, Nohynek L, Meier C, Kähkönen M, Heinonen M, Hopia A, et al. Antimicrobial properties of phenolic compounds from berries. J Appl Microbiol. (2001) 90:494–507. doi: 10.1046/j.1365-2672.2001.01271.x
- Jacobs DR Jr, Steffen LM. Nutrients, foods, and dietary patterns as exposures in research: a framework for food synergy. *Am J Clin Nutr.* (2003) 78(Suppl.):508S-13. doi: 10.1093/ajcn/78.3.508S
- Roager HM, Vogt JK, Kristensen M, Hansen LBS, Ibrügger S, Mærkedahl RB, et al. Whole grain-rich diet reduces body weight and systemic low-grade inflammation without inducing major changes of the gut microbiome: a randomised cross-over trial. *Gut.* (2019) 68:83–93. doi: 10.1136/gutjnl-2017-314786
- 41. Vanegas SM, Meydani M, Barnett JB, Goldin B, Kane A, Rasmussen H, et al. Substituting whole grains for refined grains in a 6-wk randomized trial has a modest effect on gut microbiota and immune and inflammatory markers of healthy adults. *Am J Clin Nutr.* (2017) 105:635–50. doi: 10.3945/ajcn.116.146928
- Barrett HL, Gomez-Arango LF, Wilkinson SA, McIntyre HD, Callaway LK, Morrison M, et al. A vegetarian diet is a major determinant of gut microbiota composition in early pregnancy. *Nutrients*. (2018) 10:890. doi: 10.3390/nu10070890
- Brahe LK, Le Chatelier E, Prifti E, Pons N, Kennedy S, Blædel T, et al. Dietary modulation of the gut microbiota–a randomised controlled trial in obese postmenopausal women. Br J Nutr. (2015) 114:406–17. doi: 10.1017/S0007114515001786
- 44. Alfa MJ, Strang D, Tappia PS, Graham M, Van Domselaar G, Forbes JD, et al. A randomized trial to determine the impact of a digestion resistant starch composition on the gut microbiome in older and mid-age adults. *Clin Nutr.* (2018) 37:797–807. doi: 10.1016/j.clnu.2017.03.025
- Holscher HD, Taylor AM, Swanson KS, Novotny JA, Baer DJ. Almond consumption and processing affects the composition of the gastrointestinal microbiota of healthy adult men and women: a randomized controlled trial. *Nutrients*. (2018) 10:126. doi: 10.3390/nu10020126

- Hold GL, Schwiertz A, Aminov RI, Blaut M, Flint HJ. Oligonucleotide probes that detect quantitatively significant groups of butyrate-producing bacteria in human feces. *Appl Environ Microbiol.* (2003) 69:4320–4. doi: 10.1128/AEM.69.7.4320-4324.2003
- Remely M, Hippe B, Geretschlaeger I, Stegmayer S, Hoefinger I, Haslberger A. Increased gut microbiota diversity and abundance of Faecalibacterium prausnitzii and Akkermansia after fasting: a pilot study. Wien klin Wochenschr. (2015) 127:394–8. doi: 10.1007/s00508-015-0755-1
- Miquel S, Leclerc M, Martin R, Chain F, Lenoir M, Raguideau S, et al. Identification of metabolic signatures linked to anti-inflammatory effects of Faecalibacterium prausnitzii. *mBio.* (2015) 6:e00300–15. doi: 10.1128/mBio.00300-15
- Karlsson FH, Tremaroli V, Nookaew I, Bergström G, Behre CJ, Fagerberg B, et al. Gut metagenome in European women with normal, impaired and diabetic glucose control. *Nature*. (2013) 498:99–103. doi: 10.1038/nature12198
- Earley H, Lennon G, Balfe Á, Coffey JC, Winter DC, O'Connell PR. The abundance of Akkermansia muciniphila and its relationship with sulphated colonic mucins in health and ulcerative colitis. *Sci Rep.* (2019) 9:15683. doi: 10.1038/s41598-019-51878-3
- Ganesh BP, Klopfleisch R, Loh G, Blaut M. Commensal Akkermansia muciniphila exacerbates gut inflammation in Salmonella Typhimurium-infected gnotobiotic mice. *PLoS ONE.* (2013) 8:e74963. doi: 10.1371/journal.pone.0074963
- Frost F, Storck LJ, Kacprowski T, Gärtner S, Rühlemann M, Bang C, et al. A structured weight loss program increases gut microbiota phylogenetic diversity and reduces levels of Collinsella in obese type 2 diabetics: a pilot study. *PLoS ONE*. (2019) 14:e0219489. doi: 10.1371/journal.pone.0219489
- 53. Menni C, Louca P, Berry SE, Vijay A, Astbury S, Leeming ER, et al. High intake of vegetables is linked to lower white blood cell profile and the effect is mediated by the gut microbiome. *BMC Med.* (2021) 19:37. doi: 10.1186/s12916-021-01913-w
- 54. Ghosh TS, Rampelli S, Jeffery IB, Santoro A, Neto M, Capri M, et al. Mediterranean diet intervention alters the gut microbiome in older people reducing frailty and improving health status: the NU-AGE 1-year dietary intervention across five European countries. *Gut.* (2020) 69:1218–28. doi: 10.1136/gutjnl-2019-319654
- Moorthy M, Chaiyakunapruk N, Jacob SA, Palanisamy UD. Prebiotic potential of polyphenols, its effect on gut microbiota and anthropometric/clinical markers: a systematic review of randomised controlled trials. *Trends Food Sci Technol.* (2020) 99:634–49. doi: 10.1016/j.tifs.2020.03.036
- Queipo-Ortuño MI, Boto-Ordóñez M, Murri M, Gomez-Zumaquero JM, Clemente-Postigo M, Estruch R, et al. Influence of red wine polyphenols and ethanol on the gut microbiota ecology and biochemical biomarkers. Am J Clin Nutr. (2012) 95:1323–34. doi: 10.3945/ajcn.111. 027847
- Rodríguez-Morató J, Matthan NR, Liu J, de la Torre R, Chen C-YO. Cranberries attenuate animal-based diet-induced changes in microbiota composition and functionality: a randomized crossover controlled feeding trial. J Nutr Biochem. (2018) 62:76–86. doi: 10.1016/j.jnutbio.2018. 08.019

- Istas G, Wood E, Le Sayec M, Rawlings C, Yoon J, Dandavate V, et al. Effects of aronia berry (poly) phenols on vascular function and gut microbiota: a double-blind randomized controlled trial in adult men. *Am J Clin Nutr.* (2019) 110:316–29. doi: 10.1093/ajcn/nqz075
- Lee J, Oh KH, Park SK. Dietary micronutrients and risk of chronic kidney disease: a cohort study with 12 year follow-up. *Nutrients*. (2021) 13:1517. doi: 10.3390/nu13051517
- Rossi M, Amaretti A, Raimondi S. Folate production by probiotic bacteria. Nutrients. (2011) 3:118–34. doi: 10.3390/nu3010118
- 61. Park S, Kang S, Sol Kim DS. Folate and vitamin B12 deficiencies additively impaired memory function and disturbed the gut microbiota in amyloid- β infused rats. *Int J Vitam Nutr Res.* (2019) 16:1–13. doi: 10.1024/0300-9831/a000624
- Nagy-Szakal D, Kellermayer R. The remarkable capacity for gut microbial and host interactions. *Gut Microbes*. (2011) 2:178–82. doi: 10.4161/gmic.2.3.16107
- Maynard C, Weinkove D. Bacteria increase host micronutrient availability: mechanisms revealed by studies in C. elegans. *Genes Nutr.* (2020) 15:4. doi: 10.1186/s12263-020-00662-4
- Bailén M, Bressa C, Martínez-López S, González-Soltero R, Montalvo Lominchar MGM, San Juan C, et al. Microbiota features associated with a high-fat/low-fiber diet in healthy adults. *Front Nutr.* (2020) 7:583608. doi: 10.3389/fnut.2020.583608
- 65. Tomsett KI, Barrett HL, Dekker EE, Callaway LK, McIntyre DH, Dekker Nitert M. Dietary fiber intake alters gut microbiota composition but does not improve gut wall barrier function in women with future hypertensive disorders of pregnancy. *Nutrients*. (2020) 12:3862. doi: 10.3390/nu12123862

Conflict of Interest: YJA was employed by the company Theragen Bio Co., Ltd. MHP and J-HK were employed by the company Erom Co. Ltd.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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