



Pathophysiological, Cellular, and Molecular Events of the Vascular System in Anaphylaxis

Emilio Nuñez-Borque^{1†}, Sergio Fernandez-Bravo^{1†}, Alma Yuste-Montalvo^{1†} and Vanesa Esteban^{1,2*}

OPEN ACCESS

Edited by:

Vijay Kumar,
Duke University, United States

Reviewed by:

Claudia Gonzalez-Espinosa,
Centro de Investigación y de Estudios
Avanzados del Instituto Politécnico
Nacional, Mexico
Kazushige Obata-Ninomiya,
Benaroya Research Institute,
United States
Sylvie Chollet-Martin,
INSERM U996 Inflammation,
Chimiokines et Immunopathologie,
France

*Correspondence:

Vanesa Esteban
vesteban@fjd.es

[†]These authors have contributed
equally to this work

Specialty section:

This article was submitted to
Inflammation,
a section of the journal
Frontiers in Immunology

Received: 15 December 2021

Accepted: 07 February 2022

Published: 08 March 2022

Citation:

Nuñez-Borque E, Fernandez-Bravo S,
Yuste-Montalvo A and Esteban V
(2022) Pathophysiological, Cellular,
and Molecular Events of the Vascular
System in Anaphylaxis.
Front. Immunol. 13:836222.
doi: 10.3389/fimmu.2022.836222

¹ Department of Allergy and Immunology, Instituto en Investigación Sanitaria - Fundación Jiménez Díaz (IIS-FJD), Universidad Autónoma de Madrid (UAM), Madrid, Spain, ² Faculty of Medicine and Biomedicine, Alfonso X El Sabio University, Madrid, Spain

Anaphylaxis is a systemic hypersensitivity reaction that can be life threatening. Mechanistically, it results from the immune activation and release of a variety of mediators that give rise to the signs and symptoms of this pathological event. For years, most of the research in anaphylaxis has focused on the contribution of the immune component. However, approaches that shed light on the participation of other cellular and molecular agents are necessary. Among them, the vascular niche receives the various signals (e.g., histamine) that elicit the range of anaphylactic events. Cardiovascular manifestations such as increased vascular permeability, vasodilation, hypotension, vasoconstriction, and cardiac alterations are crucial in the pathophysiology of anaphylaxis and are highly involved to the development of the most severe cases. Specifically, the endothelium, vascular smooth muscle cells, and their molecular signaling outcomes play an essential role downstream of the immune reaction. Therefore, in this review, we synthesized the vascular changes observed during anaphylaxis as well as its cellular and molecular components. As the risk of anaphylaxis exists both in clinical procedures and in routine life, increasing our knowledge of the vascular physiology and their molecular mechanism will enable us to improve the clinical management and how to treat or prevent anaphylaxis.

Key Message: Anaphylaxis, the most severe allergic reaction, involves a variety of immune and non-immune molecular signals that give rise to its pathophysiological manifestations. Importantly, the vascular system is engaged in processes relevant to anaphylactic events such as increased vascular permeability, vasodilation, hypotension, vasoconstriction, and decreased cardiac output. The novelty of this review focuses on the fact that new studies will greatly improve the understanding of anaphylaxis when viewed from a vascular molecular angle and specifically from the endothelium. This knowledge will improve therapeutic options to treat or prevent anaphylaxis.

Keywords: anaphylaxis, immune system, vascular system, endothelium, vasodilation, vascular permeability

INTRODUCTION

Anaphylaxis is a systemic reaction with a range of clinical manifestations due to the varying involvement of several organs and systems. At the cellular and molecular levels, most existing research focuses exclusively on the immune component. However, interactions between vascular and immunological microenvironments are responsible for most manifestations of anaphylaxis, including the most severe. Therefore, precise understanding of the cellular communication between immune and resident cells within the complex pathophysiology of this pathological event is necessary. This review provides an overview of the human vascular system in anaphylaxis and seeks further understanding of the dysregulation of its underlying processes.

ANAPHYLAXIS

Anaphylaxis is considered the most serious manifestation of allergic disorders. The World Health Organization defines it as “a severe, life-threatening systemic hypersensitivity reaction characterized by being rapid in onset with potentially life-threatening airway, breathing, or circulatory problems and is usually, although not always, associated with skin and mucosal changes” (1). In the United States of America (USA) alone, annual expenditures due to anaphylaxis total 1.8 billion dollars between direct and indirect costs (2). The World Allergy Organization has found that its incidence and prevalence have increased over the last decade. In 2020, global incidence was 50–112 episodes per 100,000 people, and its lifetime prevalence was between 0.3% and 5.1%. Moreover, although the mortality rate remains low (0.05–0.51, 0.03–0.32, and 0.09–0.13 per million people/year for drug-, food-, and venom-induced lethal reactions, respectively), the rate of recurrence was 26.5%–54.0% over a follow-up time of 1.5–25 years (1, 3). However, current data underestimate the true actual rate due to factors such as the lack of a common definition of anaphylaxis or discrepancies in the methodologies used in epidemiological studies (1, 4). In addition, diagnosis of this pathologic event is based on clinical symptoms that are shared with other diseases, causing anaphylactic reactions to be underdiagnosed (5). Thus, in order to complement diagnosis, molecular markers are also evaluated. Currently, the main biomarker used in clinical practice is serum tryptase, a molecule released by the effector cells of anaphylaxis. The diagnostic reference threshold for this measurement has undergone considerable modifications to the current value of 11.4 µg/l. However, its clinical utility has several drawbacks since serum tryptase is also elevated in other conditions such as mast cell disorders (6). In addition, it is not increased in most patients who develop an anaphylactic reaction (7). This could be due to the sample collection as the peak of this protein is obtained 2 h after the reaction (8). Moreover, most studies have now demonstrated the importance of considering the basal value of serum tryptase at least 24 h after the episode (9, 10). Specifically, the increase of 20% plus 2 µg/l in the acute

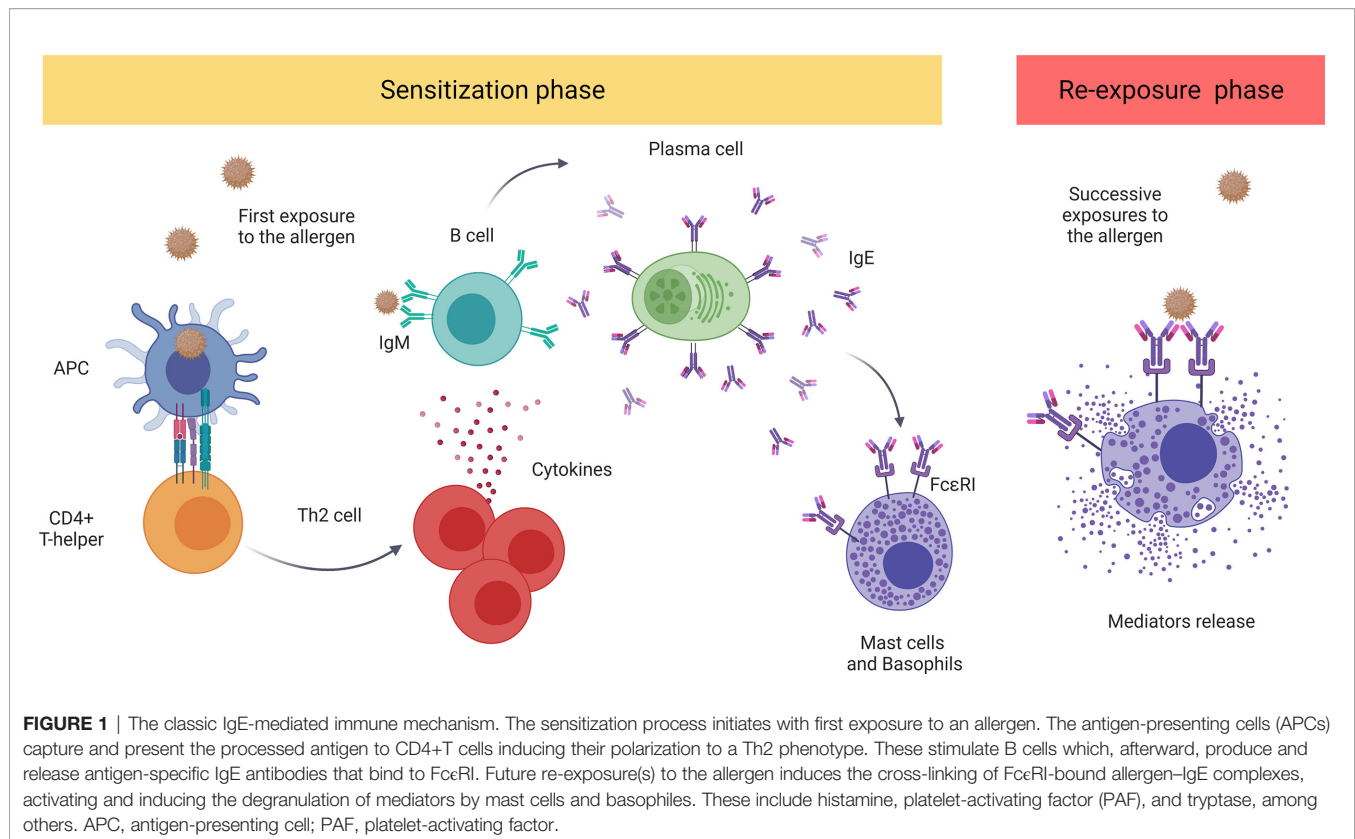
condition with respect to baseline revealed a substantial improvement in the diagnosis of anaphylaxis (11). Therefore, correct sampling and personalized diagnosis of each individual, as well as the need to find new biomarkers, are crucial for the correct management of this life-threatening reaction.

Several triggers can induce anaphylaxis, the most common being foods, drugs, and Hymenoptera venoms, although in some cases the cause of the reaction is unknown (idiopathic) (12–14). In particular, the etiological distribution of anaphylaxis differs with age since the most common triggers are drugs and insect stings in adults, while food is the main culprit in children (5, 15). However, regardless of the allergen, the signs and symptoms of anaphylaxis are the same and affect several systems. Cutaneous symptoms (e.g., localized urticaria, erythema, angioedema, pruritus) are the most common manifestations and are present in 80% of cases. Other systems may be affected, such as the gastrointestinal (e.g., emesis, diarrhea), nervous (e.g., confusion, drowsiness, seizure), respiratory (e.g., dyspnea, cough, wheezing, bronchoconstriction), and circulatory (e.g., palpitations, tachycardia, hypotension) (16). The cardiovascular system is highly involved during mild anaphylactic reactions and plays a key role in the most severe cases, in which anaphylactic shock could take place (17, 18). Deterioration of the circulatory system due to age, other diseases, and treatments administered such as angiotensin-converting enzyme inhibitors are some of the main risk factors associated with fatal reactions (19–21). Therefore, the cardiovascular system is essential in the development of the reaction and a key target to developing future therapeutic strategies.

IMMUNE SYSTEM IN ANAPHYLAXIS

Cellular and Molecular Mechanisms Mediated by IgE

The major molecular mechanism underlying anaphylaxis is the classic allergic IgE-mediated reaction involving mast cells and basophils (22). Mast cells reside in all vascularized tissues, and studies have shown a correlation among the severity of the reaction, early degranulation of mast cells, and release of mediators (23–25). On the other hand, basophils are also granular immune cells, although they are blood-circulating leukocytes and not tissue-resident cells, unlike mast cells (26, 27). Recent studies suggest that basophils play a key role in food-mediated anaphylaxis. However, since the activation of these cells is complementary to that of mast cells, their contribution to human anaphylaxis remains a pivotal point of study (28, 29). Both cell types present the high-affinity IgE receptor (FcεRI) on their surface and are considered the main effectors in this pathologic event (22). Mechanistically, when the individual is first exposed to an allergen, the immune sensitization process is initiated, triggering the production of antigen-specific IgE antibodies. In successive reexposures to the antigen, there is cross-linking of FcεRI-bound allergen-IgE complexes, activating effector cells and giving rise to the release of a large number of mediators (12, 13, 30) (**Figure 1**).



IgE-Independent Cellular and Molecular Mechanisms

Interestingly, after experiencing an anaphylactic reaction, some patients have no detectable levels of allergen-specific IgE (9, 31, 32). This happening means that a considerable percentage of subjects do not show evidence of IgE-dependent immune activation, so other cells and processes must be involved (33–35). Due to the similarity between mouse and human immune systems, the use of murine models has been essential in elucidating the role of other mechanisms in anaphylaxis. Among them, the IgG pathway appeared as the main alternative immune process described in anaphylactic reactions (5, 35, 36). Unlike the IgE mechanism, the mechanism guided by IgGs seems to require higher levels of specific IgG and antigens, presumably due to the lower affinity of FcγR compared with FcεRI (33). Nowadays, it is well established that IgG antibodies can induce anaphylaxis by binding to their different receptors (FcγR) (37). These ones are found in mast cells, basophils, neutrophils, monocytes, and macrophages conforming to the major cellular types activated by this alternative pathway. The cellular consequence is elicited reactions due to the release of abundant mediators (33, 38–41). One of the common mediators released, in response to both IgE and IgG molecules, is the platelet-activating factor (PAF) which is released from all the subsets coming from a myeloid progenitor (**Table 1**).

Specifically, the implication of both monocytes and macrophages has been demonstrated in passive and active

systemic anaphylaxis (12, 89, 90). However, the role of neutrophils has gained much relevance as key cellular players eliciting anaphylactic reactions (43). Their contribution was firstly observed in experimental mouse models which suggested that a similar pathway could be operative in human reactions (43, 56). In fact, evidence in patients has shown elevated circulating serum levels of neutrophil elastase and myeloperoxidase (the major mediators stored in their granules). These results supported the existence of a neutrophil-associated IgG molecular mechanism associated with drug-induced anaphylaxis (86, 91). Therefore, neutrophils and IgG arise as important factors in the etiopathogenesis of the reaction but also lead to the possibility of constituting new biomarkers of anaphylaxis.

Other reactions triggered by drugs (e.g., opioids, vancomycin) are also capable of activating mast cells and basophils as well as degranulation led by the Mas-related G-protein coupled receptor member X2 (MRGPRX2) (92–95). This relevant pathway would explain those reactions absent of immunoglobulins, and therefore huge effort is being realized to elucidate its relevance in the setting of anaphylaxis (35, 96, 97). Even further, external factors (e.g., physical exercise, exposure to cold, or ultraviolet radiation) contribute as cofactors in the reactions (12, 34, 41, 98).

Concerning the participation of other immune cells in these events, accumulation of eosinophils was detected in passive cutaneous anaphylactic reactions in guinea pigs, as well as in spleens and coronary arteries from anaphylactic human cadavers (99, 100). Specifically, these cells express on their surface both

TABLE 1 | Anaphylaxis-associated products and their main cellular sources are listed.

	Molecules	Source	References	
Vasoactive agents	Histamine	MC, BAS, NEUT	(42, 43)	
	Bradykinin	P	(44, 45)	
	NO	EC	(46, 47)	
	Endothelin-1	EC, SMC, hMC, MAC, MON	(47–50)	
Proteases	Trypsin	MC, BAS	(46, 51)	
	Carboxypeptidase A	MC, BAS	(21, 51, 52)	
	Chymase	MC	(9, 42, 46)	
	Plasminogen activator	MC	(9)	
	Cathepsin G	MC, NEUT, EOS, BAS	(9, 26, 46, 53, 54)	
	Elastase	NEUT	(55)	
Lipidic molecules	LTB4, LTC4, LTD4, LTE4	MC, BAS, NEUT	(9, 26, 42, 46, 51, 56)	
	PGD2	MC, NEUT, MON, MAC, PLAT	(51, 56–58)	
	PGE2	MC, BAS, MAC, EOS, PLAT, SMC	(26, 58, 59)	
	PGF2	MC, PLAT	(26, 58)	
	TXA2	MC, EOS, PLAT,	(26, 42, 58)	
	TXB2	MC	(60, 61)	
	Prostacyclin	MC, ECs, SMC, PLAT	(48, 58, 62, 63)	
	PAF	MC, BAS, NEUT, EOS, MON, MAC, PLAT, EC	(43, 52, 64–67)	
	S1P	MC, PLAT, ERY, EC	(68–70)	
	Derived from	FXII, PK, HK Thrombin	P	(44)
			P	(44)
	Contact, Coagulation and Complement systems activation	C3a	P	(44)
		C5a	P	(44)
Heparine		MC, BAS	(9)	
Chondroitin sulfate		MC, BAS	(9)	
Cytokines and chemokines	Renin	hMC	(9)	
	TNF α	MC, MAC, NEUT, BAS	(26, 42, 56, 71, 72)	
	TGF- β	MC, EOS	(22, 26, 46)	
	IFN- γ	NEUT, LT, NK	(56, 73, 74)	
	G-CSF	MC, MAC, MONO, NEUT, EC	(42, 75)	
	M-CSF	MC, MONO, EC	(42, 76)	
	GM-CSF	MC, BAS, EC	(26, 42, 77)	
	CCL-2	MC, EC, NEUT, MAC, MON	(22, 46, 78–82)	
	CCL-5	MC, EOS, MON	(26, 46, 83)	
	Stem cell factor	MC	(9, 46)	
	TWEAK	EC	(84)	
	IgE-dependent histamine releasing factor	BAS	(9)	
	MIP-1 α	BAS	(9)	
Interleukins	Platelet factor 4 (PF4)	PLAT	(64, 85)	
	IL-1 β	MC, NEUT	(42, 56)	
	IL-3	MC, EOS	(26, 42)	
	IL-4	MC, BAS, EOS	(9, 22, 42, 46)	
	IL-5	MC, EOS	(9, 22, 26, 46)	
	IL-6	MC, MAC, NEUT, BAS	(9, 21, 42, 46, 72, 86)	
	IL-8	MC, NEUT, EOS	(9, 22, 26, 42, 56)	
	IL-10	MC, EOS	(22, 26, 42, 46)	
	IL-13	MC, BAS, EOS	(21, 26, 42, 46)	
	IL-16	MC, EOS	(26, 42)	

(Continued)

TABLE 1 | Continued

Molecules	Source	References
IL-18	MC, EOS	(26, 42)
IL-22	MC	(42)
IL-33	MC, BAS	(21)
VEGF	MC, MAC, PLAT	(46, 71, 87)
Basogranulin	BAS	(9, 52)
L-Selectin	NEUT	(86)
MPO	NEUT, MON	(56, 86, 88)
MBP	EOS	(26)
β-TG	PLAT	(64)

NO, nitric oxide; LTB4, leukotriene B4; LTC4, leukotriene C4; LTD4, leukotriene D4; LTE4, leukotriene E4; PGD2, prostaglandin D2; PGE2, prostaglandin E2; PGF2, prostaglandin F2; TXA2, thromboxane A2; TXB2, thromboxane B2; PAF, platelet activating factor; S1P, sphingosine 1 phosphate; FXII, coagulation factor XII; PK, plasma kallikrein; HK, kininogen; TNF, tumor necrosis factor; TGF, transforming growth factor; IFN, interferon; G-CSF, granulocyte colony-stimulating factor; M-CSF, macrophage colony-stimulating factor; GM-CSF, granulocyte-macrophage colony-stimulating factor; TWEAK, TNF-related weak inducer of apoptosis; MIP, macrophage inflammatory proteins; PF4, platelet factor 4; IL, interleukin; VEGF, vascular endothelial growth factor; MPO, myeloperoxidase; MBP, major basic protein; β-TG, beta-thromboglobulin; MC, mast cells; nMC, mast cells; EOS, eosinophils; BAS, basophils; MAC, macrophages; MON, monocytes; EC, endothelial cells; PLAT, platelets; ERY, erythrocytes; SMC, smooth muscle cells; NK, natural killer cells; LT, T lymphocytes; P, plasma.

receptor types: FcγR and FcεR (26). Moreover, the contribution of platelets to the reaction by the release of important mediators has been also evidenced through IgE- and IgG-dependent mechanisms (64, 101). Therefore, it seems that a big portion of myeloid cells activated in anaphylaxis (endotypes) exists, which are associated with different anaphylactic phenotypes and biomarkers (5).

SOLUBLE MEDIATORS AND CASCADES IN ANAPHYLAXIS

Overall, a large number of anaphylactic mediators coming from different cellular sources are released both into the bloodstream and in local microenvironments (Table 1). The nature of these molecules is very diverse: vasoactive agents, proteases, lipidic particles, cytokines, chemokines, interleukins, hormones, and neurotransmitters, among others. In addition, due to the homeostatic imbalance and acute inflammatory state characteristic of anaphylaxis, complement and contact/coagulation cascades are involved in the pathophysiology of this event leading to the release of numerous intermediate products (42, 44, 102).

Classically, the best-characterized mediators have been classified into preformed and newly synthesized. Those stored in cytoplasmic granules contain highly sulfated polysaccharides (heparin or other proteoglycans), tryptase, chymase, histamine, and PAF, being mainly released from mast cells and basophils (22, 24). However, other cellular providers, such as neutrophils or platelets, also supply key molecular effectors as PAF to the reaction (103). Molecules derived from arachidonic acid, interleukins, chemokines, and/or cytokines are also massively released (42, 104, 105). In summary, the diversity of triggers and/or signals described upstream to the cellular activation induces different degranulation strategies in mast cells (106). Probably, it also occurs with other participating cells of the anaphylactic reaction. Into the downstream signaling following cellular activation, the influx of mediators causes the recruitment of other immune cells but also the activation of resident cells that contribute, in turn, with a multitude of other anaphylactic products amplifying the molecular and cellular signaling (107, 108).

In relation, multitudes of soluble products from both the contact (bradykinin) and the complement system (C3a, C4a, C5a) are also released in the pool of mediators (109, 110). The activation of the coagulation, fibrinolytic, contact, and complement system pathways is highly involved in anaphylaxis. Peptides C3a, C4a, and C5a derived from the proteolysis of the C3, C4 and C5 components of the complement system are considered, along IgG molecules, the main elicitors of non-IgE anaphylactic reactions. These molecules, often known as anaphylatoxins, induce mast cell degranulation through its specific receptors on their surface (35). On the other hand, contact and coagulation/fibrinolytic systems are closely related to each other and are involved in the so-called non-immunologic anaphylactic reactions. Coagulation factors such as kininogen, fibrinogen, and factors V and VII are decreased in anaphylactic patients (44).

In addition, other molecular players such as extracellular vesicles (EVs), microRNAs, and metabolites are just starting to

gain relevance as surrogate biomarkers in anaphylaxis but also participating in their underlying molecular pathways (13, 111–113).

Altogether, this large variety of molecular signaling pathways contributes to amplify the number and heterogeneity of processes occurring in these pathological events.

THE CARDIOVASCULAR SYSTEM

The cellular components of the vascular compartment are both target and effector resident cells in anaphylaxis. Thus, the following section expands on the morphological and functional particularities of the cardiovascular system in order to broaden its knowledge in anaphylactic reactions. The circulatory system is composed of a set of interconnected tubular organs that form a closed circuit in the human body. It is divided into the blood vascular system, which carries blood, and the lymphoid vascular system, which transports lymph (114). However, in this review, we will only focus on the former.

The functions of the blood vascular system are as follows: transportation of substances entering the body from the external environment (mainly nutrients and oxygen), transfer of molecules such as hormones, hydraulic force generation, regulation of heat, ultrafiltration in the kidney, and defense through the transport of immune cells and mediators (115). This circuit is made up of the heart and a number of different types of vessels that distribute blood to the organs. Elastic arteries emerge from the heart and branch out to the muscular arteries and arterioles. The latter give rise to the capillaries, and from these, venules and veins return blood to the heart (114, 116, 117) (**Figure 2** and **Box 1**).

All blood vessels have a common structure that is mainly composed of three layers, i.e., *tunica intima*, *tunica media*, and *tunica adventitia* (*ti*, *tm*, and *ta*, respectively), but morphological and functional differences exist between them. The *ti* consists of a single and extensive layer of endothelial cells (ECs) that forms a physical barrier between blood and tissues, allowing the selective transport of molecules through it (132). This morphological and functional structure is named the endothelium, and it participates in a multitude of vital functions such as modulating coagulation/fibrinolysis, regulating transportation of inflammatory cells, controlling cellular metabolism, sustaining homeostasis of resident stem cells, guiding organ repair, and releasing inflammatory and angiocrine factors (133, 134). The lumen of the endothelium is covered by a set of proteoglycans and glycoproteins (glycocalyx) which also confer multifunctional and dynamic properties to this layer (135). Thus, *ti* plays a critical role in many physiologic and pathologic processes and as a result the endothelium is considered an organ in itself (136). At the cellular level, ECs are very heterogeneous (137). Their morphology, function, and gene and antigen composition vary between different organs and sections of the vasculature (138, 139). For instance, the capacity of EC to quickly contract in response to inflammatory and vasoactive mediators leads up to endothelial breakdown, resulting in increased vascular permeability, which occurs mainly

in the microvasculature (37, 140–143). At the molecular level, two main canonical pathways regulate endothelium stability: the actin-myosin cytoskeleton and those molecular processes related to the tight and adherent junctions (144–147).

The next tunica, *tm*, is located in the middle area of the vessels determining their diameter and is usually composed of several layers of vascular smooth muscle cells (VSMCs) and/or elastin fibers supported mainly by a collagen matrix. VSMCs are fundamental homeostatic players in different diseases such as hypertension and atherosclerosis (148, 149). These cells contain the main molecular machinery to directly regulate the vascular tone (dilatation and constriction) mainly by changes in their intracellular Ca^{2+} (iCa^{2+}) levels. Indirectly and with the main purpose of maintaining homeostasis, ECs also contribute to vascular tone modulation *via* synthesis and release of vasoactive substances (150, 151). Specifically, the main relaxant product released by ECs is nitric oxide (NO) (152–154).

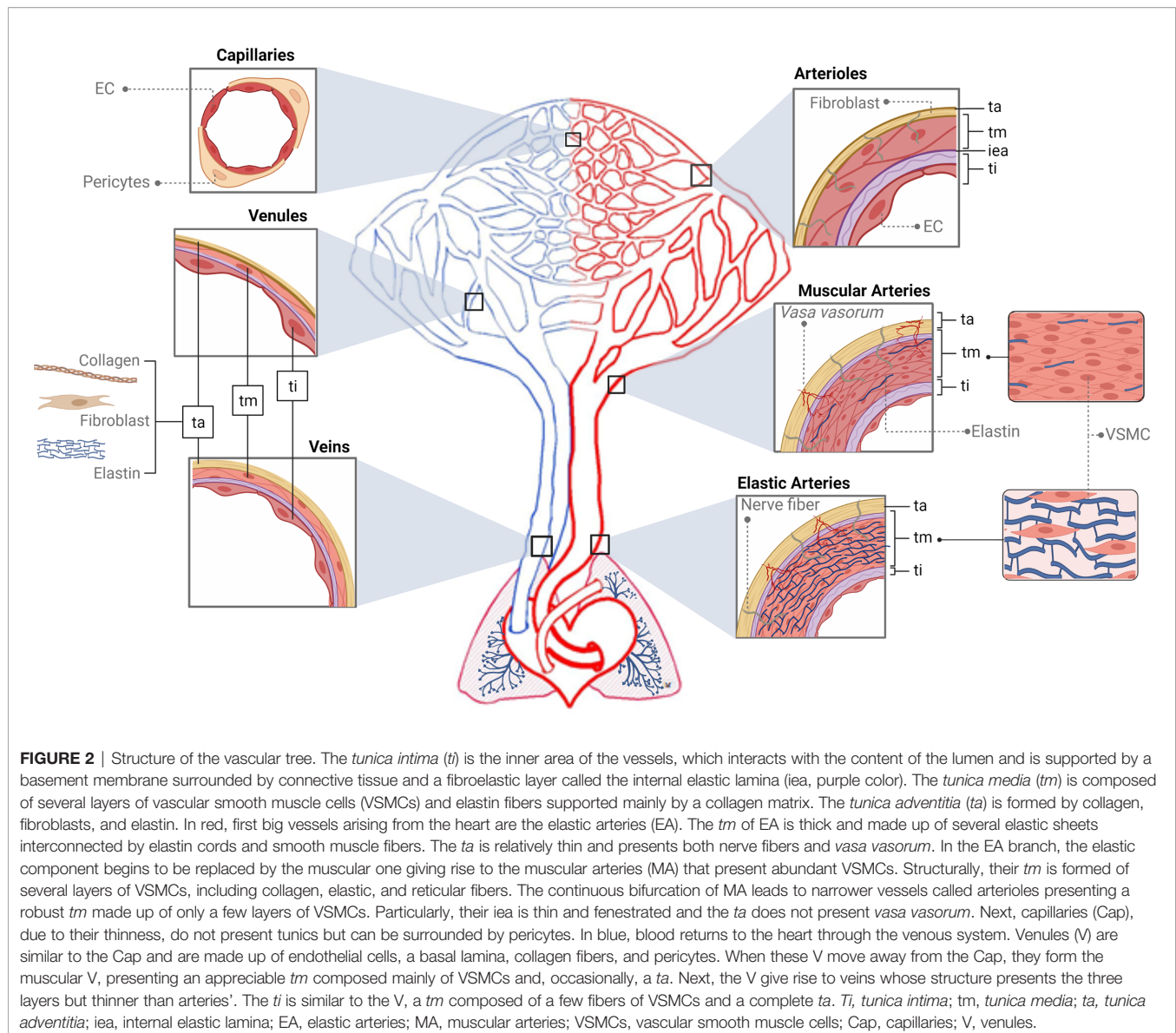
The outer layer of the vessel is the *ta*, which is responsible for the integrity and resistance to physical stress of the vascular wall. This layer consists mainly of connective tissue and contains *vasa vasorum*. These small vessels irrigate the whole wall of the large vessel since nutrients and oxygen cannot reach all cells by diffusion (114, 116, 155–157). In addition, this layer might present nerve fibers which regulate vascular tone and which are also altered during pathological processes (158).

CARDIOVASCULAR PATHOPHYSIOLOGY OF ANAPHYLAXIS

Decades of investigations have demonstrated the importance of specific interactions between immune and vascular cells in different diseases (159). In anaphylaxis, the participation of immune cells leading to the final release of mediators is very well defined (42). Nevertheless, their impact throughout the vascular niche is less clear. Both the rupture of the endothelial barrier and vascular tone disturbances are the essential anomalies observed during this pathologic event. In addition, phenomena occurring in the chest cavity (e.g., cardiac arrest, decreased cardiac output, hypoxia) are present in anaphylactic reactions (**Figure 3**). Molecularly, the EC- and VSMC-signaling pathways govern these processes (**Figure 4**). A dysfunctional endothelium is the cause of important cardiovascular diseases such as thrombosis, atherosclerosis, or hypertension. Even a damaged endothelium has been observed in patients with mastocytosis (160). Additionally, in acute inflammatory situations, as it has been observed in COVID-19 disease, ECs contribute as effector cells to the cytokine storm (161). In addition, the endothelium actively participates in the activation of the coagulation, contact, and complement in anaphylaxis (44, 162, 163). Therefore, its study is a hot topic and of great importance for the treatment of several pathologies such as anaphylaxis.

Increased Vascular Permeability and Endothelial Barrier Breakdown

For years, anaphylaxis has been associated with a concomitant leakage of fluids (164). A highly relevant investigation employing



indirect measurements of changes in hemoglobin concentration demonstrated that a transfer of up to 35% of the fluid to the extracellular space can occur within 10 min of allergen exposure (165). Another interesting study carried out in mice determined that histamine-induced vascular permeability associated with anaphylaxis occurs in postcapillary venules (166). Furthermore, a recent investigation focusing on humans has shown the relevance of micro-ECs as the sole responders to anaphylactic mediators in *in vitro* vascular permeability (167). The high extravasation of fluid contributes to edema of the airways and pulmonary emphysema (168). Specifically, a study including 50 idiopathic anaphylaxis subjects showed that upper airway obstruction was strongly associated with laryngeal edema, possibly leading to asphyxia (169). In addition, investigations based on systemic anaphylaxis using allergic rats observed an association between interstitial airway vascular leakage and

severe bronchoconstriction (170). On the other hand, angioedema and urticaria, the most common symptoms of anaphylaxis, involve vascular fluid extravasation and are characterized by temporary localized swelling (171, 172). Angioedema can affect all layers of the skin and visceral walls, such as the respiratory system and the gastrointestinal tract (171, 173, 174). Furthermore, angioedema of the upper airways or pulmonary edema has been observed in one out of two human anaphylactic biopsies indicating the necessity to control vascular permeability (18). Importantly, the endothelial IL-4 receptor α chain (IL-4R α) and its underlying signaling appears as relevant in the gastrointestinal extravasation disturbances associated with food allergic reactions (175). A recent study carried out in peanut-challenged anaphylactic patients has shown that most leakage volume takes place in the gut, contributing to the appearance of gastrointestinal symptoms (176). Furthermore,

BOX 1 | Understanding the structure and function of the main vascular regions.

The heart is an organ located in the central part of the thoracic cavity and made up of two thin-walled atria and two thick-walled ventricles. The right and left parts of the heart are separated by a septum, so blood from one side does not mix with the other (118). However, the atria and ventricles in both areas contract in a coordinated manner. The function of this organ is to pump blood so that it may be distributed throughout the vessels in the rest of the body. For this to happen, a sequence of muscular contractions (systole) and relaxations (diastole) is carried out due to changes in the levels of intracellular calcium in cardiac myocytes, giving rise to the cardiac cycle (119–121). Increasing evidence indicates that the human heart is a target of cardiac anaphylaxis, and in which human heart mast cells (HHMC) play a key role (122).

Elastic arteries (e.g., aorta, pulmonary artery, carotids) arise from the heart and transport blood to the muscular arteries. In each ventricular systole, the heart pumps a very large volume of blood, causing distension of the elastic walls, which accumulate the potential energy released during the diastole. Therefore, they push the blood when the heart is relaxed, acting as a secondary propulsion pump complementing the heart and resulting in a continuous flow (114, 117).

Muscular arteries (e.g., brachial artery, femoral artery) are the most abundant arteries in the human body. They usually appear near the organs, and their function is to distribute the blood through the different regions of the body since, by contracting their muscular component, they can regulate the size of their lumen (114, 116, 117, 123).

Arterioles are the vessels responsible for regulating blood flow since their muscle fibers modulate the lumen of the vessel (114, 116, 117, 124). Specifically, arterioles have a precapillary sphincter with a conical shape that also allows blood flow regulation (125).

Capillaries appear behind the sphincters of the arterioles and are the smallest blood vessels (5–10 μM). Due to their thinness, they do not present tunics and are the organ of the circulatory system with the highest level of gas and nutrient exchange. There are three types of capillaries: continuous, fenestrated, and sinusoids according to the specialized subpopulations of EC (114, 116, 126).

Venules are structurally and functionally similar to the capillaries. The postcapillary venules are the main vessel involved in the extravasation of leukocytes to the tissues. Here, endothelial junctions are very labile and sensitive to inflammatory mediators, which makes hyperpermeability processes possible (114, 117, 127).

Veins accumulate a larger volume of blood as they have lower pressure than arteries, and the blood is moved by the skeletal muscular system rather than by the heart (117, 128). They are classified into three types depending on their diameter, which increases as they approach the heart: small (0.1–1 mm), medium (1 mm–1 cm), or large (>1 cm) (116, 129, 130). Therefore, the venous system has additional venous valves formed by folds of the *tunica intima* and whose function is to prevent blood reflux (116, 117, 129, 131).

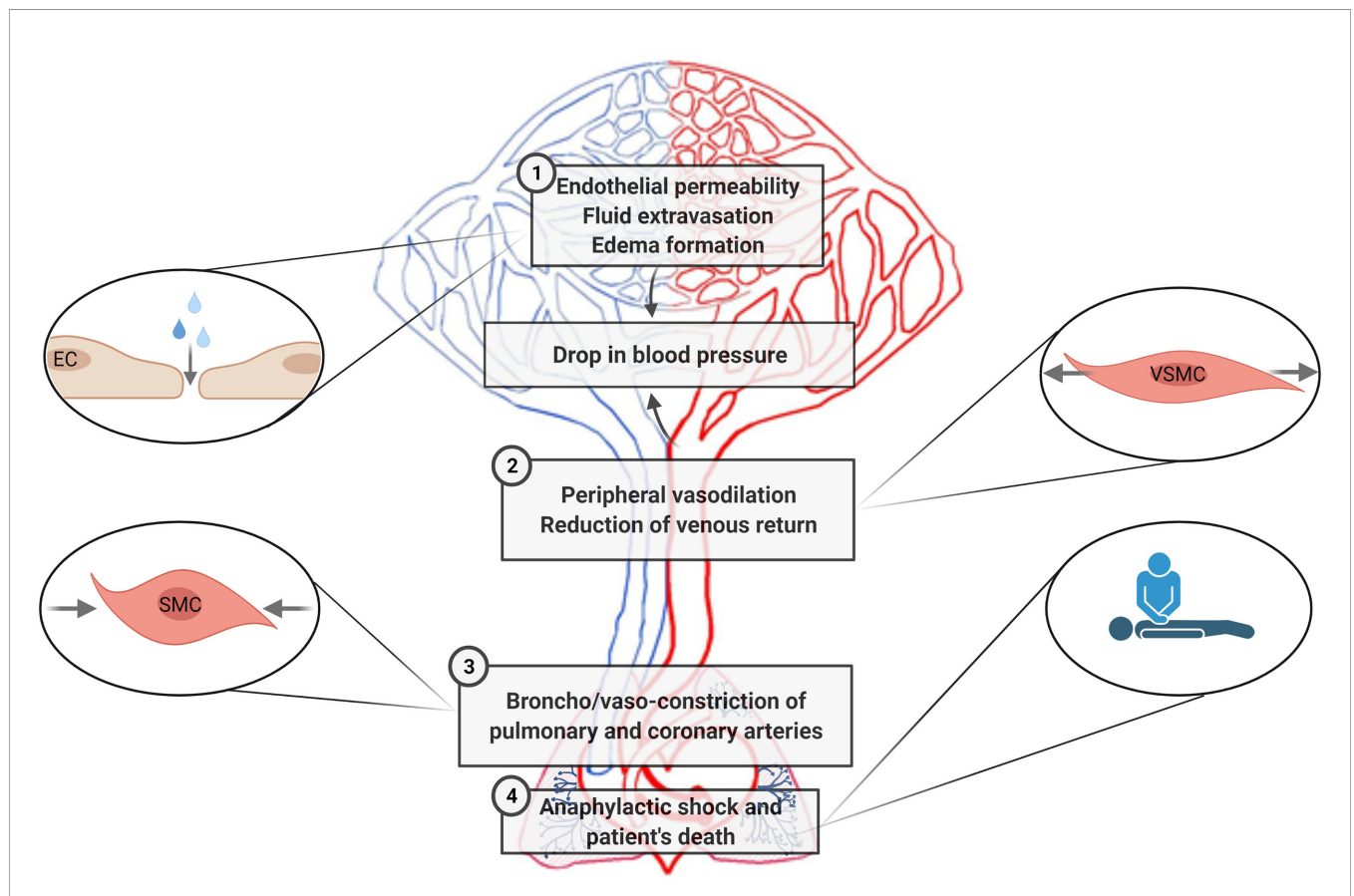
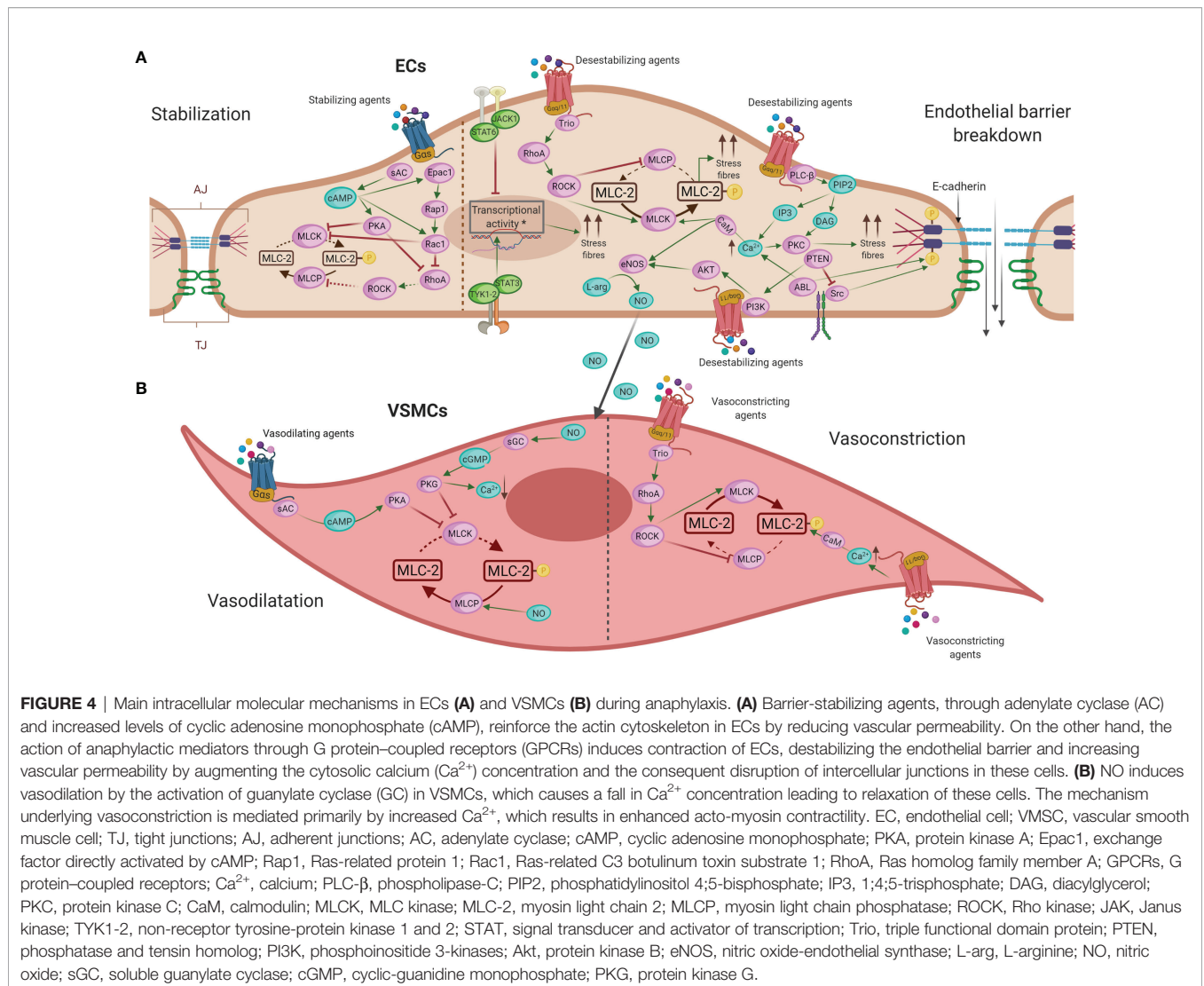


FIGURE 3 | Cardiovascular pathophysiological manifestations in anaphylaxis are sketched according to the main areas/type of vessels/organs corresponding to the processes affected. 1. Increased fluid extravasation (vascular permeability/leakiness)—microcirculation (capillaries and venules). 2. Profound systemic vasodilation (decreased peripheral vascular resistance)—peripheral vessels. 3. Vasoconstriction of the thoracic cavity—coronary and pulmonary arteries. 4. Tachycardia, reduced myocardial contractility and decreased cardiac output—heart. EC, endothelial cells; SMC, smooth muscle cells; VSMCs, vascular smooth muscle cells.



intestinal extravasation has also been determined in a drug-induced anaphylactic patient in whom a diffuse edema was observed (177).

Molecularly, stabilization of the endothelial barrier depends on a series of connections, such as tight junctions (TJ) and adherent junctions (AJ). TJs are mainly composed of occludins and claudins that bind to the actin cytoskeleton and α -catenin. In AJs, VE-cadherin is the major structural protein, and they contribute to barrier stabilization by providing mechanical cohesive strength between ECs (178, 179). Different known anaphylactic mediators such as histamine and PAF are widely described as inducers of vascular permeability (46). Their action through its endothelial receptors activates phospholipase C (PLC- β) which hydrolyzes phosphatidylinositol 4,5-bisphosphate (PIP2) to produce two second messengers: inositol 1,4,5-trisphosphate (IP3) and diacylglycerol (DAG). IP3 raises intracellular iCa^{2+} levels stimulating calcium-regulated mechanisms. This iCa^{2+} acts, together with DAG, to activate protein kinase C (PKC) that contributes to the

disruption of junctional protein complexes. In addition, iCa^{2+} /calmodulin (CaM) activates myosin light chain kinase (MLCK) leading to the phosphorylation of myosin light chain 2 (MLC-2) and resulting in increased acto-myosin contractility, stress fibers, and the disruption of the endothelial barrier. In turn, myosin light chain phosphatase (MLCP) dephosphorylates MLC-2 producing cellular stability. MLCP inhibition is achieved by Rho kinase (ROCK) activation downstream of the guanine nucleotide exchange factor, Trio, which initiates the activation of RhoA (46, 178, 180, 181). In addition, the non-receptor tyrosine kinases, ABL and Src, are activated in response to anaphylactic stimuli contributing also to iCa^{2+} mobilization and VE-cadherin dissociation (182, 183). Moreover, other novel endothelial molecules also participate in leakage processes. The fibroblast growth factor-inducible 14 (Fn14) receptor, the signal transducer and activator of transcription 3 (STAT3), the peroxisome proliferator-activated receptor (PPAR β/δ), and MALT1 protease activity have been proposed as therapeutic strategies in allergy and anaphylaxis (84, 175, 183–

185). In addition, recently, a challenge study has demonstrated the capacity of EVs purified from plasma of anaphylactic patients to interact with ECs, eliciting vascular permeability and thereby suggesting that cellular communication between microenvironments may also be established in anaphylaxis (186).

In contrast, other mediators stabilize the endothelial barrier, mostly to maintain homeostasis. These barrier-stabilizing agents, through adenylate cyclase (AC), increase cyclic adenosine monophosphate (cAMP) levels in ECs. It activates the protein kinase A (PKA) and guanine nucleotide exchange factors such as Rac1 which lead to inhibition of the small GTPase RhoA and the strengthening of the actin cytoskeleton stabilizing the endothelial barrier (46, 178). Specifically, sphingosine-1-phosphate (S1P) and prostaglandin D2 (PGD2) are some of these soluble stabilizing molecules while intracellularly phosphatidylinositol 3-kinase (PI3K-C2 α) and the regulator of calcineurin 1 (Rcan1) have been argued to strengthen the endothelium (187–193).

Vasodilation and Hypotension

Adequate control of peripheral vascular pressure is essential, as it provides the driving force to pump blood to the organs (194). Vasodilation is predominantly controlled by the autonomic nervous response (195). However, anaphylactic mediators can directly contribute to the loss of the vascular resistance even in the absence of neural input. Vasodilation has been observed in anaphylactic patients with severe cardiovascular manifestations such as hypotension (16, 165, 196). The drop of blood pressure has been attributed both to fluid extravasation and to loss of vascular resistance, rather than a direct effect of the myocardium (165). Specifically, a study points to the action of mediators leading to vasodilation and increased vascular permeability (197). Therefore, it remains unclear whether vasodilation is derived from or leads to fluid leakage in human anaphylaxis. To shed light on this, an interesting study using an experimental murine model found a correlation between the increase in vascular permeability and a fall in blood pressure during early anaphylaxis. This fact suggests that fluid leakage may act as a precursor of hypotension and have shared molecular mechanisms (198). On the other hand, vasodilation causes excessive venous blood accumulation due to the significant reduction in venous return that occurs during anaphylaxis (199). It is consequent to think that veins, which are the reservoirs of blood in the body, undergo serious functional impairment during this event. Specifically, hemodynamic monitoring in a clinical case of anaphylactic reaction to penicillin indicated a reduction in cardiac output owing to this decrease in venous return (200). Therefore, the impact of the altered flow leads to cardiac compromise and suggests that veins are an important niche to study (199, 201).

In anaphylaxis, the main anaphylactic mediators not only play a role in vascular permeability but also regulate the tone of the vessel. At the cellular and molecular levels, the activation of the endothelial nitric synthase (eNOS) and the underlying production of NO have been probably characterized as the most harmful vasodilator and hypotensive factor (187, 202–204). This molecule induces vasodilation by the activation of soluble guanylate cyclase (sGC) in VSMCs, causing increased formation of cyclic-guanidine monophosphate (cGMP). Subsequently, it activates protein kinase

G (PKG), which causes a fall in iCa^{2+} concentration, ensuring that MLCK can no longer phosphorylate MLC-2 and leading to relaxation of the VSMCs and vasodilation. Furthermore, NO activates MLCP and, by dephosphorylating MLC-2, causes additional relaxing effects (46, 204). Anaphylactic patients with respiratory manifestations associate with increased levels of exhaled NO (205). In addition, this mediator could act as a regulator of microvascular permeability (152–154). Therefore, the endothelium participates in modulating the resistance of the vessels and VSMCs contribute to maintaining a correct vascular tone and the consequent homeostasis. Furthermore, other mediators with vasodilator capacity have been described in this context. In the case of histamine, its contractile and dilating effects depend on the expression and activity of the histamine receptors located on both ECs and VSMCs (206, 207). Tryptase modulates vasodilation through calcitonin genes and by the release of neuromodulators such as substance P (208–210). Bradykinin, a hypotensive factor released during anaphylaxis as a result of contact system activation, leads to vasodilation and associates with laryngeal edema (109).

Vasoconstriction

The regulation of the vascular system presents a dilemma because a third block of important circulatory disturbances may also appear in anaphylaxis. Severe reactions seem to present vasoconstriction of the vessels in the thoracic cavity leading to cardiac arrest (211). The coronary and pulmonary arteries are the primary vessels susceptible to contraction (150, 212). However, constrictive processes carried out by the main underlying anaphylactic mediators are neither clearly described nor studied in human anaphylaxis (213–216), except increased serum troponin that is observed in Kounis syndrome and Takotsubo cardiomyopathy (211). Therefore, due to the obvious complications of studying these processes in emergency situations, most of the available evidence is based on animal studies. Vasoconstriction of the pulmonary artery has been associated with right ventricular failure in animal models, favoring the circulatory collapse seen in anaphylaxis (217). On the other hand, left ventricular dysfunction has been related to coronary vasoconstriction in a rat model (218). Additionally, another rat model of anaphylaxis revealed increased portal venous resistance and, therefore, hepatic vasoconstriction (219). There is also a contraction of the bronchial tree that hinders gas exchange in the alveoli, generating hypoxia. This fact, in turn, supports the constriction of the pulmonary circulation compromising the entire vascular system and has been associated with bronchospasm and death in guinea pigs (220, 221). Therefore, in anaphylaxis, vasoconstriction of pulmonary and coronary arteries contributes to the reduction of myocardial contractility and may be a major cause of death (196, 211, 212).

Underlying mechanisms to vasoconstriction are mostly mediated by increased iCa^{2+} which activates MLCK leading to the phosphorylation of MLC-2 and resulting in increased actomyosin contractility (148).

Anaphylactic Shock

The previous described pathophysiological manifestations (increased endothelial permeability, peripheral vasodilation,

and thoracic cavity constriction) alter blood flow, resulting in impaired vascular homeostasis, which may lead to anaphylactic shock (171, 197).

The exacerbated fluid extravasation, in combination with the loss of peripheral vascular resistance, reduces venous return to the heart. In addition, the blood flow is removed from surface areas, reducing oxygen demands and delivering it to vital organs, thereby decreasing body temperature. However, it is insufficient to satisfy the metabolic demands of the human organism. On the other hand, constriction of pulmonary arteries, together with the effect of tightened airway smooth muscle, leads to reduced oxygen uptake. Therefore, to make up for this lack of oxygen in the body, breathing becomes rapid and deep and the heart rate increases (tachycardia). However, a second phase characterized by bradycardia occurs because this organ cannot remedy the influence of the previous manifestations of this pathological event, causing cardiac collapse and death of the patient (171, 201, 222, 223).

MEMBRANE RECEPTORS IN ANAPHYLAXIS

A wide variety of soluble mediators have been proposed as participants of the anaphylaxis pathophysiology (46, 71). The majority of these molecules are ligands for the largest group of membrane receptors which are those linked to heterotrimeric G proteins (GPCRs). Additionally, they have been implicated in a variety of diseases (224–226). However, GPCR downstream signaling pathways are not well characterized for every anaphylactic mediator, although their impact on vascular permeability and vasodilation is well established (Table 2).

In this sense, most of the studies have mainly been carried out in experimental models (37). Specifically, anaphylactic shock depending on endothelial Gq/G11 was characterized in mice (247).

TREATMENT

To date, the first-line and most effective anaphylaxis treatment is intramuscular administration of adrenaline/epinephrine (245). It is indicated after recognition of symptoms due to its ability to directly remedy alterations in the cardiovascular system caused by the reaction. Its administration prevents cardiovascular collapse and enhances blood flow during anaphylactic shock (12, 248, 249). Mechanistically, this drug resolves anaphylaxis *via* its different adrenergic receptors (Figure 5). Epinephrine exerts its vasoconstrictor action through α -adrenergic receptors on VSMCs. Its activation causes stimulation of PLC- β , which cleaves PIP₂ into DAG and IP₃. This increased intracellular IP₃ binds to its receptors, leading to higher iCa^{2+} levels and subsequent myosin phosphorylation (250, 251). The result of this pathway is a peripheral vasoconstriction that reverses peripheral vasodilation alleviating hypotension and reducing edema (252, 253). On the other hand, epinephrine, through β -adrenergic receptors and subsequent G α (s) activation, results in enhanced

AC activity, which promotes cAMP formation and the stability of the ECs barrier (254). In addition, through β ₁ receptors it increases heart rate and its force of contraction, while β ₂ receptor-mediated action restores bronchoconstriction and reduces the release of inflammatory mediators by the main immune effector cells, mast cells, and basophils (251, 252, 255). In bronchial smooth muscle cells (BSMCs), epinephrine restores its constriction through the β ₂-adrenergic receptor. cAMP production activates PKA, which phosphorylates and inactivates the MLCK. These facts stop the downstream signal for contraction and thus relax BSMCs (251). However, this drug administration is not always effective and patients may still die.

Epinephrine would be ineffective in cardiopathy patients receiving β -blockers, and glucagon is occasionally administered. The action of this drug is not mediated by adrenergic receptors and allows reversal of refractory hypotension and bronchospasm (1, 249). Moreover, there are other complementary therapies which may be useful but should never replace the first-line treatment. These include supplemental oxygen, β ₂-agonists to reverse bronchospasm, intravenous crystalloid solutions to restore circulatory volume, antihistamines to control cutaneous symptoms, and glucocorticosteroids to prevent biphasic reactions and reduce inflammation (256).

Currently, various investigations on potential alternative drugs for the treatment of anaphylaxis are being conducted, although they are not implemented in clinical routine yet. Methylene blue is an inhibitor of NO pathways and has been reported to be a safe medication for refractory hypotension underlying anaphylactic shock (257). On the other hand, sugammadex, a γ -cyclodextrin, has been shown to encapsulate and inactivate neuromuscular blocking agents, which may be triggers of this pathological event. However, its usefulness remains controversial since a recent study of perioperative anaphylaxis treated with this drug concluded that it did not modify the reaction (258). In addition, promising results have been obtained from mouse models where PAF antagonists have been shown to mitigate the severity and duration of anaphylaxis (259).

Otherwise, desensitization interventions have been proposed for the long-term treatment and prevention of anaphylaxis (260). Specifically, biological agents are emerging as a potential adjuvant for this process (36, 261). Among them, omalizumab, a monoclonal antibody against IgE, has been shown to be a successful treatment in reducing the number and severity of anaphylactic reactions (262). In addition, several therapies are appearing for the prevention of food-induced anaphylaxis. These are aimed to modulate specific immune pathways in different ways, such as antibodies against the main cytokines involved in the response (anti-IL-33, anti-IL-5, etc.) or immunotherapies to polarize the Th₂ response characteristic of allergic reactions to Th₁ and T regulatory (Treg) ones through toll-like receptor (TLR) agonists (TLR4 and TLR9), nanoparticles, probiotics, or IFN γ , among others (263–265).

CONCLUSIONS

Understanding the extent of human vessels in anaphylaxis is the challenge we have taken on in this study. Although the

TABLE 2 | Main vascular GPCRs in anaphylaxis.

Mediators	Receptors	Vascular permeability	Vasodilation/ Hypotension	Vasoconstriction	References
Histamine	HR1 (G $\alpha_{q/11}$)	X	X	X	(181, 206, 207, 227–229)
	HR2 (G α_s)	X	X	X	
PAF	PAF-R (G $\alpha_{q/11}$)	X	X	X	(24, 230, 231)
Tryptase	PAR2 (G $\alpha_{q/11}$)	X	X	X	(208, 232, 233)
LTB4, LTC4, LTD4, LTE4	CysLT1R (G $\alpha_{q/11}$)	X	X	X	(234–236)
	CysLT2R (G $\alpha_{q/11}$)	X	X	X	
PGD2	DP (G α_s)	X	X		(71, 192)
PGE2	EP1 (G $\alpha_{q/11}$)		X	X	(237)
	EP2 (G α_s)		X	X	
	EP4 (G α_s)		X	X	
PGF2	FP (G $\alpha_{q/11}$)			X	(238)
TXA2	TP (G $\alpha_{q/11}$)			X	(238)
Bradykinin	BR1 (G $\alpha_{q/11}$)	X	X	X	(109, 163, 239)
	BR2 (G α_i)	X	X	X	
S1P	S1PR1 (G α_i)		X	X	(187, 189, 190, 240, 241)
	S1PR2 (G $\alpha_{i/o}$, G $\alpha_{q/11}$, G $\alpha_{12/13}$)		X	X	
	S1PR3 (G $\alpha_{i/o}$, G $\alpha_{q/11}$, G $\alpha_{12/13}$)		X	X	
C3a	C3aR (G $\alpha_{q/11}$)	X	X	X	(242–244)
C5a	C5aR (G α_i)	X	X	X	
Epinephrine	α 1AR (G $\alpha_{q/11}$)		X	X	(245, 246)
	α 2AR (G α_i)		X	X	
	β 2AR (G α_s)		X	X	

The processes affected by those most described anaphylactic mediators are listed. They signal through different G protein-coupled receptors (GPCRs) which address their intracellular signaling through different pathways that ultimately lead to the main cardiovascular alterations in anaphylaxis (vascular permeability, vasodilation/hypotension, or constrictive processes).

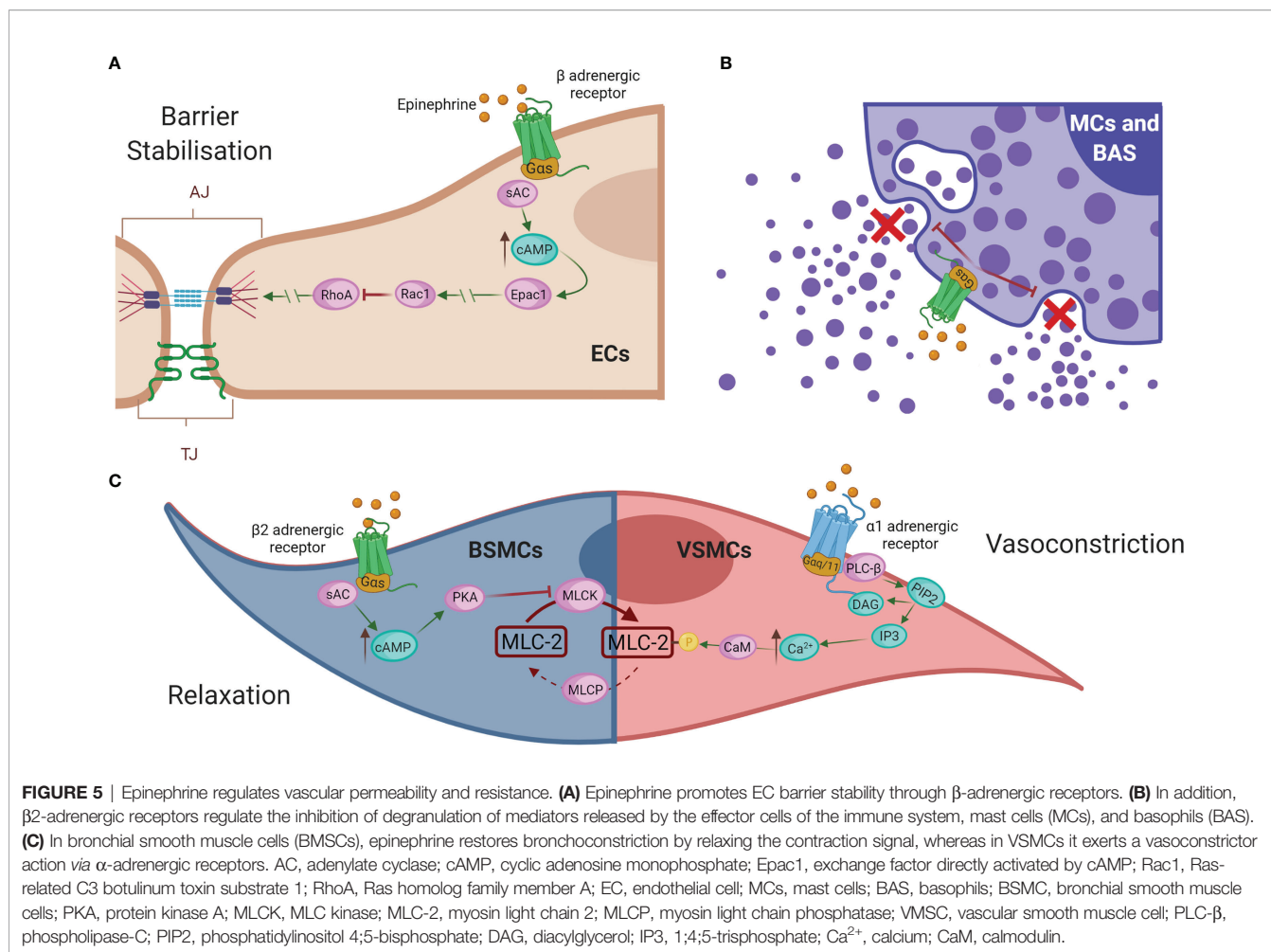
vascular system functions as a whole, specific features are identified in the different types of vessels. Their specialization is based on the morphology, functionality, and molecular signaling pathways of the vascular cells composing those regions (138). Therefore, expanding the knowledge of human vessels in anaphylaxis is crucial to implementing tools based on their underlying molecular mechanisms. It would help to achieve better quality of acute and long-term clinical management of anaphylactic patients.

For years, the classic cellular and molecular mechanism described for anaphylaxis has been the cross-linking of Fc ϵ RI-bound allergen–IgE complexes activating mast cells/basophils and the consequent release of mediators (5). More recent studies shed light about the relevance of other immune effector cells in anaphylaxis such as monocytes, macrophages, neutrophils, eosinophils, and platelets (266). Unfortunately, most of this evidence is based on murine models but knowledge about its involvement in human anaphylaxis still remains scarce (12). Due to the features of these innate immune cells, the main research focus in molecular anaphylaxis is on the immune system. However, anaphylaxis is a systemic reaction in which several organs and systems are involved. Therefore, simultaneously to the inflammatory response, the vasculature actively participates in such pathophysiological process. Epinephrine is the treatment of choice to palliate anaphylactic symptoms accordingly to

guidelines (245). It is the “safeguard” molecule which triggers both endogenous and exogenous mechanisms through its adrenergic receptors (1). Therefore, the downstream signaling pathways of these receptors involve a dual behavior (constriction and relaxation processes) in vascular and bronchial smooth muscle cells. In addition, both cell types, either located within the vessel wall or conforming to organs, would relate to the variety of clinical consequences and the severity grade of the reactions.

In this context, the wide endothelium surface would also be contributing differentially to the pathophysiology of the anaphylaxis. The endothelium arises as an important *organ-cell like* in anaphylaxis playing a role not only in the control of fluids and the vascular tone but also as an activation surface for the coagulation, contact, and complement systems (44, 162, 163). Moreover, ECs release relevant anaphylactic mediators such as NO, although it is likely that other molecules (presumably cytokines or interleukins) could also be released contributing to the pool of mediators in the reaction. However, the exact contribution of these cells as provider of mediators to the anaphylactic cellular microenvironments existing between immune and resident niches is unknown.

Altogether, we can summarize that the permeable and/or vasomotor capacity of the different anaphylactic mediators (NO, tryptase, histamine, bradykinin, or a multitude of other scarce or unknown ones) would determine the magnitude of



the anaphylactic events. Furthermore, the role of their receptors (GPCRs) in human anaphylaxis is not yet fully understood, but they result to be essential in preventing or inducing vascular effects. Therefore, GPCRs are the most promising therapeutic targets of study nowadays and the degree of responsiveness of ECs and VSMCs is an important factor to determine in anaphylaxis.

Due to the complexity to investigate in human subjects the cellular and molecular mechanisms of anaphylaxis, the improvement and reproducibility of animal—or *in vitro* human—models is necessary. This fact is one of the major limitations to study this pathological event. A big piece of the literature around it derives from animal models. For that reason, conclusions must be taken cautious when translated at human reactions. Strategies based on functional *in vitro* studies by using human cell cultures combined with sera would provide clues about cellular and molecular happenings occurring in specific anaphylactic microenvironments. Therefore, abundant investigations are aimed at finding vascular targets revealing future alternative strategies to treat or prevent anaphylaxis. Efforts must be focused on alternative therapies called to specifically modulate the whole or parts of the vascular system

that benefit the clinical management of these life-threatening events in the near future.

AUTHOR CONTRIBUTIONS

Conceptualization and—original draft preparation, VE. Writing—review and editing, EN-B, SF-B, AY-M, and VE. Funding acquisition, VE. All authors contributed to the article and approved the submitted version.

FUNDING

This research was supported by grants from the Instituto de Salud Carlos III (PI18/00348 and PI21/00158) and FEDER Thematic Networks and Cooperative Research Centers RETICS ARADyAL RD16/0006/0013. This work was also supported by the SEAIC (19_A08) and Alfonso X el Sabio University Foundations. EN-B was granted by funding from the Community of Madrid included in the project FOOD-AL (CM_P2018/BAAA-4574).

REFERENCES

- Cardona V, Ansotegui IJ, Ebisawa M, El-Gamal Y, Fernandez Rivas M, Fineman S, et al. World Allergy Organization Anaphylaxis Guidance 2020. *World Allergy Organ J* (2020) 13(10):100472. doi: 10.1016/j.waojou.2020.100472
- Fromer L. Prevention of Anaphylaxis: The Role of the Epinephrine Auto-Injector. *Am J Med* (2016) 129(12):1244–50. doi: 10.1016/j.amjmed.2016.07.018
- Tejedor-Alonso MA, Moro-Moro M, Múgica-García MV. Epidemiology of Anaphylaxis: Contributions From the Last 10 Years. *J Investig Allergol Clin Immunol* (2015) 25(3):163–175; quiz follow 174–175.
- Tanno L, Bierrenbach A, Simons F, Cardona V, Thong B, Molinari N, et al. Critical View of Anaphylaxis Epidemiology: Open Questions and New Perspectives. *Allergy Asthma Clin Immunol* (2018) 14:12. doi: 10.1186/s13223-018-0234-0
- Castells M. Diagnosis and Management of Anaphylaxis in Precision Medicine. *J Allergy Clin Immunol* (2017) 140(2):321–33. doi: 10.1016/j.jaci.2017.06.012
- Platzgummer S, Bizzaro N, Bilò MB, Pravettoni V, Cecchi L, Sargentini V, et al. Recommendations for the Use of Tryptase in the Diagnosis of Anaphylaxis and Clonal Mastcell Disorders. *Eur Ann Allergy Clin Immunol* (2020) 52(2):51–61. doi: 10.23822/EurAnnACI.1764-1489.133
- Sala-Cunill A, Cardona V, Labrador-Horrillo M, Luengo O, Esteso O, Garriga T, et al. Usefulness and Limitations of Sequential Serum Tryptase for the Diagnosis of Anaphylaxis in 102 Patients. *Int Arch Allergy Immunol* (2013) 160(2):192–9. doi: 10.1159/000339749
- Dua S, Dowe J, Foley L, Islam S, King Y, Ewan P, et al. Diagnostic Value of Tryptase in Food Allergic Reactions: A Prospective Study of 160 Adult Peanut Challenges. *J Allergy Clin Immunol Pract* (2018) 6(5):1692–8.e1. doi: 10.1016/j.jaip.2018.01.006
- Simons F, Frew A, Ansotegui I, Bochner B, Golden D, Finkelman F, et al. Risk Assessment in Anaphylaxis: Current and Future Approaches. *J Allergy Clin Immunol* (2007) 120(1 Suppl):S2–24. doi: 10.1016/j.jaci.2007.05.001
- Vitte J, Sabato V, Tacquard C, Garvey L, Michel M, Mertes P, et al. Use and Interpretation of Acute and Baseline Tryptase in Perioperative Hypersensitivity and Anaphylaxis. *J Allergy Clin Immunol Pract* (2021) 9(8):2994–3005. doi: 10.1016/j.jaip.2021.03.011
- Mateja A, Wang Q, Chovanec J, et al. Defining baseline variability of serum tryptase levels improves accuracy in identifying anaphylaxis. *J Allergy Clin Immunol* (2021) 149(2):S0091-6749(21)01295-1. doi: 10.1016/j.jaci.2021.08.007
- Reber LL, Hernandez JD, Galli SJ. The Pathophysiology of Anaphylaxis. *J Allergy Clin Immunol* (2017) 140(2):335–48. doi: 10.1016/j.jaci.2017.06.003
- Nuñez-Borque E, Fernandez-Bravo S, Rodriguez Del Rio P, Alwashali E, Lopez-Dominguez D, Gutierrez-Blazquez M, et al. Increased miR-21-3p and miR-487b-3p Serum Levels During Anaphylactic Reaction in Food Allergic Children. *Pediatr Allergy Immunol* (2021) 32(6):1296–306. doi: 10.1111/pai.13518
- Fischer D, Vander Leek TK, Ellis AK, Kim H. Anaphylaxis. *Allergy Asthma Clin Immunol* (2018) 14(Suppl 2):54. doi: 10.1186/s13223-018-0283-4
- Braganza SC, Acworth JP, Mckinnon DRL, Peake JE, Brown AFT. Paediatric Emergency Department Anaphylaxis: Different Patterns From Adults. *Arch Dis Child* (2006) 91(2):159–63. doi: 10.1136/adc.2004.069914
- Dribin T, Schnadower D, Spergel J, Campbell R, Shaker M, Neuman M, et al. Severity Grading System for Acute Allergic Reactions: A Multidisciplinary Delphi Study. *J Allergy Clin Immunol* (2021) 148(1):173–81. doi: 10.1016/j.jaci.2021.01.003
- Khan BQ, Kemp SF. Pathophysiology of Anaphylaxis. *Curr Opin Allergy Clin Immunol* (2011) 11(4):319–25. doi: 10.1097/ACI.0b013e3283481ab6
- Martínez-Fernández P, Vallejo-de-Torres G, Sánchez-de-León-Robles MS, Navarro-Escayola E, Moro-Moro M, Alberti-Masgrau N, et al. Medical and Pathologic Characteristics of Fatal Anaphylaxis: A Spanish Nationwide 17-Year Series. *Forensic Sci Med Pathol* (2019) 15(3):369–81. doi: 10.1007/s12024-019-00134-1
- Mullins RJ, Wainstein BK, Barnes EH, Liew WK, Campbell DE. Increases in Anaphylaxis Fatalities in Australia From 1997 to 2013. *Clin Exp Allergy* (2016) 46(8):1099–110. doi: 10.1111/cea.12748
- Turner PJ, Jerschow E, Umasunthar T, Lin R, Campbell DE, Boyle RJ. Fatal Anaphylaxis: Mortality Rate and Risk Factors. *J Allergy Clin Immunol Pract* (2017) 5(5):1169–78. doi: 10.1016/j.jaip.2017.06.031
- Simons FER. Anaphylaxis. *J Allergy Clin Immunol* (2010) 125(2 Suppl 2):S161–181. doi: 10.1016/j.jaci.2009.12.981
- Elieh Ali Komi D, Wöhrl S, Bielory L. Mast Cell Biology at Molecular Level: A Comprehensive Review. *Clin Rev Allerg Immunol* (2020) 58(3):342–65. doi: 10.1007/s12016-019-08769-2
- van der Linden PW, Hack CE, Poortman J, Vivié-Kipp YC, Struyvenberg A, van der Zwan JK. Insect-Sting Challenge in 138 Patients: Relation Between Clinical Severity of Anaphylaxis and Mast Cell Activation. *J Allergy Clin Immunol* (1992) 90(1):110–8. doi: 10.1016/s0091-6749(06)80017-5
- Vadas P, Perelman B, Liss G. Platelet-Activating Factor, Histamine, and Tryptase Levels in Human Anaphylaxis. *J Allergy Clin Immunol* (2013) 131(1):144–9. doi: 10.1016/j.jaci.2012.08.016
- Galli SJ. The Mast Cell-IgE Paradox: From Homeostasis to Anaphylaxis. *Am J Pathol* (2016) 186(2):212–24. doi: 10.1016/j.ajpath.2015.07.025
- Stone KD, Prussin C, Metcalfe DD. IgE, Mast Cells, Basophils, and Eosinophils. *J Allergy Clin Immunol* (2010) 125(2):S73–80. doi: 10.1016/j.jaci.2009.11.017
- Siracusa MC, Kim BS, Spergel JM, Artis D. Basophils and Allergic Inflammation. *J Allergy Clin Immunol* (2013) 132(4):789–8. doi: 10.1016/j.jaci.2013.07.046
- Savage JH, Courneya JP, Sterba PM, Macglashan DW, Saini SS, Wood RA. Kinetics of Mast Cell, Basophil, and Oral Food Challenge Responses in Omalizumab-Treated Adults With Peanut Allergy. *J Allergy Clin Immunol* (2012) 130(5):1123–29.e2. doi: 10.1016/j.jaci.2012.05.039
- Korosec P, Turner PJ, Silar M, Kopac P, Kosnik M, Gibbs B, et al. Basophils, High-Affinity IgE Receptors, and CCL2 in Human Anaphylaxis. *J Allergy Clin Immunol* (2017) 140(3):750–8.e15. doi: 10.1016/j.jaci.2016.12.989
- Galli SJ, Tsai M. IgE and Mast Cells in Allergic Disease. *Nat Med* (2012) 18(5):693–704. doi: 10.1038/nm.2755
- Golden DBK, Tracy JM, Freeman TM, Hoffman DR. Insect Committee of the American Academy of Allergy, Asthma and Immunology. Negative Venom Skin Test Results in Patients With Histories of Systemic Reaction to a Sting. *J Allergy Clin Immunol* (2003) 112(3):495–8. doi: 10.1016/s0091-6749(03)01537-9
- Hoffman DR. Fatal Reactions to Hymenoptera Stings. *Allergy Asthma Proc* (2003) 24(2):73–7.
- Finkelman FD, Khodoun MV, Strait R. Human IgE-Independent Systemic Anaphylaxis. *J Allergy Clin Immunol* (2016) 137(6):1674–80. doi: 10.1016/j.jaci.2016.02.015
- Muñoz-Cano R, Picado C, Valero A, Bartra J. Mechanisms of Anaphylaxis Beyond IgE. *J Investig Allergol Clin Immunol* (2016) 26(2):73–82. doi: 10.18176/jiaci.0046
- Cianferoni A. Non-IgE-Mediated Anaphylaxis. *J Allergy Clin Immunol* (2021) 147(4):1123–31. doi: 10.1016/j.jaci.2021.02.012
- Jimenez-Rodriguez TW, Garcia-Neuer M, Alenazy LA, Castells M. Anaphylaxis in the 21st Century: Phenotypes, Endotypes, and Biomarkers. *J Asthma Allergy* (2018) 11:121–42. doi: 10.2147/JAA.S159411
- Finkelman FD. Anaphylaxis: Lessons From Mouse Models. *J Allergy Clin Immunol* (2007) 120(3):506–515; quiz 516–517. doi: 10.1016/j.jaci.2007.07.033
- Gillis CM, Jönsson F, Mancardi DA, et al. Mechanisms of Anaphylaxis in Human Low-Affinity IgG Receptor Locus Knock-in Mice. *J Allergy Clin Immunol* (2017) 139(4):1253–65.e14. doi: 10.1016/j.jaci.2016.06.058
- Galli SJ, Franco CB. Basophils are Back! *Immunity* (2008) 28(4):495–7. doi: 10.1016/j.immuni.2008.03.010
- Karasuyama H, Tsujimura Y, Obata K, Mukai K. Role for Basophils in Systemic Anaphylaxis. *Chem Immunol Allergy* (2010) 95:85–97. doi: 10.1159/000315939
- Kow ASF, Chik A, Soo KM, Khoo LW, Abas F, Tham CL. Identification of Soluble Mediators in IgG-Mediated Anaphylaxis. *via Fcγ Receptor: A Meta-Analysis Front Immunol* (2019) 10:190. doi: 10.3389/fimmu.2019.00190
- Ogawa Y, Grant JA. Mediators of Anaphylaxis. *Immunol Allergy Clin North Am* (2007) 27(2):249–260, vii. doi: 10.1016/j.jiac.2007.03.013
- Jönsson F, Mancardi DA, Kita Y, Karasuyama H, Iannascoli B, Van Rooijen N, et al. Mouse and Human Neutrophils Induce Anaphylaxis. *J Clin Invest* (2011) 121(4):1484–96. doi: 10.1172/JCI45232

44. Guilarte M, Sala-Cunill A, Luengo O, Labrador-Horrillo M, Cardona V. The Mast Cell, Contact, and Coagulation System Connection in Anaphylaxis. *Front Immunol* (2017) 8:846. doi: 10.3389/fimmu.2017.00846
45. Rubin LE, Levi R. Protective Role of Bradykinin in Cardiac Anaphylaxis. Coronary-Vasodilating and Antiarrhythmic Activities Mediated by Autocrine/Paracrine Mechanisms. *Circ Res* (1995) 76(3):434–40. doi: 10.1161/01.res.76.3.434
46. Nguyen SMT, Rupprecht CP, Haque A, Pattanaik D, Yusin J, Krishnaswamy G. Mechanisms Governing Anaphylaxis: Inflammatory Cells, Mediators, Endothelial Gap Junctions and Beyond. *Int J Mol Sci* (2021) 22(15):7785. doi: 10.3390/ijms22157785
47. Russo D, Minutolo R, Clienti C, De Nicola L, Iodice C, Savino FA, et al. Endothelin-1 Released by Vascular Smooth Muscle Cells Enhances Vascular Responsiveness of Rat Mesenteric Arterial Bed Exposed to High Perfusion Flow. *Am J Hypertens* (1999) 12(11 Pt 1):1119–23. doi: 10.1016/s0895-7061(99)00085-0
48. Lambden S, Creagh-Brown BC, Hunt J, Summers C, Furni LG. Definitions and Pathophysiology of Vasoplegic Shock. *Crit Care* (2018) 22(1):174. doi: 10.1186/s13054-018-2102-1
49. Reid AC, Silver RB, Levi R. Renin: At the Heart of the Mast Cell. *Immunol Rev* (2007) 217:123–40. doi: 10.1111/j.1600-065X.2007.00514.x
50. Kawakami T, Mitsuhashi H, Saitoh J, Takeuchi H, Hasome N, Hiruta M, et al. Hypotension Associated With Systemic Aggregated Anaphylaxis is Not Attenuated by a Selective Endothelin-A Receptor Antagonist, BQ 610, in Rabbits. *vivo J Anesth* (2003) 17(1):22–9. doi: 10.1007/s005400300004
51. Watts MM, Marie Ditto A. Anaphylaxis. *Allergy Asthma Proc* (2019) 40(6):453–6. doi: 10.2500/aap.2019.40.4270
52. Beck SC, Wilding T, Buka RJ, Baretto RL, Huissoon AP, Krishna MT. Biomarkers in Human Anaphylaxis: A Critical Appraisal of Current Evidence and Perspectives. *Front Immunol* (2019) 10:494. doi: 10.3389/fimmu.2019.00494
53. Burster T, Gärtner F, Knippschild U, Zhanapiya A. Activity-Based Probes to Utilize the Proteolytic Activity of Cathepsin G in Biological Samples. *Front Chem* (2021) 9:628295. doi: 10.3389/fchem.2021.628295
54. Tralau T, Meyer-Hoffert U, Schröder JM, Wiedow O. Human Leukocyte Elastase and Cathepsin G are Specific Inhibitors of C5a-Dependent Neutrophil Enzyme Release and Chemotaxis. *Exp Dermatol* (2004) 13(5):316–25. doi: 10.1111/j.0906-6705.2004.00145.x
55. van der Heijden J, Geissler J, van Mirre E, van Deuren M, van der Meer JW, Salama A, et al. A Novel Splice Variant of FcγRIIa: A Risk Factor for Anaphylaxis in Patients With Hypogammaglobulinemia. *J Allergy Clin Immunol* (2013) 131(5):1408–16.e5. doi: 10.1016/j.jaci.2013.02.009
56. Jönsson F, Mancardi DA, Albanesi M, Bruhns P. Neutrophils in Local and Systemic Antibody-Dependent Inflammatory and Anaphylactic Reactions. *J Leukoc Biol* (2013) 94(4):643–56. doi: 10.1189/jlb.1212623
57. Ono E, Taniguchi M, Mita H, Fukutomi Y, Higashi N, Miyazaki E, et al. Increased Production of Cysteinyl Leukotrienes and Prostaglandin D2 During Human Anaphylaxis. *Clin Exp Allergy* (2009) 39(1):72–80. doi: 10.1111/j.1365-2222.2008.03104.x
58. Braune S, Küpper JH, Jung F. Effect of Prostanoids on Human Platelet Function: An Overview. *Int J Mol Sci* (2020) 21(23):E9020. doi: 10.3390/ijms21239020
59. Muñoz-Cano RM, Casas R, Araujo G, de la Cruz C, Martín M, Roca-Ferrer J, et al. Prostaglandin E2 Decreases Basophil Activation in Patients With Food-Induced Anaphylaxis. *Allergy* (2021) 76(5):1556–9. doi: 10.1111/all.14615
60. Schulman ES, Newball HH, Demers LM, Fitzpatrick FA, Adkinson NF. Anaphylactic Release of Thromboxane A2, Prostaglandin D2, and Prostacyclin From Human Lung Parenchyma. *Am Rev Respir Dis* (1981) 124(4):402–6. doi: 10.1164/arrd.1981.124.4.402
61. McManus LM, Shaw JO, Pinckard RN. Thromboxane B2 (TxB2) Release During IgE Anaphylaxis in the Rabbit. *J Immunol* (1980) 125(5):1950–4.
62. Peters SP, Schleimer RP, Naclerio RM, MacGlashan DW Jr, Togias AG, Proud D, et al. The Pathophysiology of Human Mast Cells. *In Vitro and In Vivo Function*. *Am Rev Respir Dis* (1987) 135(5):1196–200. doi: 10.1164/arrd.1987.135.5.1196
63. Burka JF, Garland LG. A Possible Modulatory Role for Prostacyclin (PGI2) INIGa-Induced Release of Slow-Reacting Substance of Anaphylaxis in Rats. *Br J Pharmacol* (1977) 61(4):697–9. doi: 10.1111/j.1476-5381.1977.tb07564.x
64. Kasperska-Zajac A, Rogala B. Platelet Function in Anaphylaxis. *J Investig Allergol Clin Immunol* (2006) 16(1):1–4.
65. Prescott SM, Zimmerman GA, McIntyre TM. Human Endothelial Cells in Culture Produce Platelet-Activating Factor (1-Alkyl-2-Acetyl-Sn-Glycero-3-Phosphocholine) When Stimulated With Thrombin. *Proc Natl Acad Sci U S A* (1984) 81(11):3534–8. doi: 10.1073/pnas.81.11.3534
66. Triggiani M, Schleimer RP, Warner JA, Chilton FH. Differential Synthesis of 1-Acyl-2-Acetyl-Sn-Glycero-3-Phosphocholine and Platelet-Activating Factor by Human Inflammatory Cells. *J Immunol* (1991) 147(2):660–6.
67. Tsujimura Y, Obata K, Mukai K, Shindou H, Yoshida M, Nishikado H, et al. Basophils Play a Pivotal Role in Immunoglobulin-G-Mediated But Not Immunoglobulin-E-Mediated Systemic Anaphylaxis. *Immunity* (2008) 28(4):581–9. doi: 10.1016/j.immuni.2008.02.008
68. Weth D, Benetti C, Rauch C, Gstraunthaler G, Schmidt H, Geisslinger G, et al. Activated Platelets Release Sphingosine 1-Phosphate and Induce Hypersensitivity to Noxious Heat Stimuli *In Vivo*. *Front Neurosci* (2015) 9:140. doi: 10.3389/fnins.2015.00140
69. Incorvaia C, Mauro M, Pravettoni V, Incorvaia S, Riario-Sforza GG. Anaphylaxis: An Update on its Understanding and Management. *Recent Pat Inflammation Allergy Drug Discovery* (2010) 4(2):124–9. doi: 10.2174/187221310791163107
70. Oskeritzian CA, Milstien S, Spiegel S. Sphingosine-1-Phosphate in Allergic Responses, Asthma and Anaphylaxis. *Pharmacol Ther* (2007) 115(3):390–9. doi: 10.1016/j.pharmthera.2007.05.011
71. Nakamura T, Murata T. Regulation of Vascular Permeability in Anaphylaxis. *Br J Pharmacol* (2018) 175(13):2538–42. doi: 10.1111/bph.14332
72. Mican JA, Arora N, Burd PR, Metcalfe DD. Passive Cutaneous Anaphylaxis in Mouse Skin is Associated With Local Accumulation of Interleukin-6 mRNA and Immunoreactive Interleukin-6 Protein. *J Allergy Clin Immunol* (1992) 90(5):815–24. doi: 10.1016/0091-6749(92)90107-d
73. Kubota Y, Koga T, Nakayama J. *In Vitro* Released Interferon-Gamma in the Diagnosis of Drug-Induced Anaphylaxis. *Eur J Dermatol* (1999) 9(7):559–60.
74. Castro F, Cardoso AP, Gonçalves RM, Serre K, Oliveira MJ. Interferon-Gamma at the Crossroads of Tumor Immune Surveillance or Evasion. *Front Immunol* (2018) 9:847. doi: 10.3389/fimmu.2018.00847
75. Tabbara IA. Granulocyte Colony-Stimulating Factor. *South Med J* (1993) 86(3):350–5. doi: 10.1097/00007611-199303000-00020
76. Chockalingam S, Ghosh SS. Macrophage Colony-Stimulating Factor and Cancer: A Review. *Tumour Biol* (2014) 35(11):10635–44. doi: 10.1007/s13277-014-2627-0
77. Schuett J, Schuett H, Oberoi R, Koch AK, Pretzer S, Luchtefeld M, et al. NADPH Oxidase NOX2 Mediates TLR2/6-Dependent Release of GM-CSF From Endothelial Cells. *FASEB J* (2017) 31(6):2612–24. doi: 10.1096/fj.201600729R
78. Zyrianova T, Lopez B, Liao A, Gu C, Wong L, Ottolia M, et al. BK Channels Regulate LPS-Induced CCL-2 Release From Human Pulmonary Endothelial Cells. *Am J Respir Cell Mol Biol* (2021) 64(2):224–34. doi: 10.1165/rcmb.2020-0228OC
79. Blidberg K, Palmberg L, Dahlén B, Lantz AS, Larsson K. Chemokine Release by Neutrophils in Chronic Obstructive Pulmonary Disease. *Innate Immun* (2012) 18(3):503–10. doi: 10.1177/1753425911423270
80. Gerlach K, Köhler-Bachmann S, Jungck D, Körber S, Yanik S, Knoop H, et al. Endothelin Receptor-Antagonists Suppress Lipopolysaccharide-Induced Cytokine Release From Alveolar Macrophages of Non-Smokers, Smokers and COPD Subjects. *Eur J Pharmacol* (2015) 768:123–30. doi: 10.1016/j.ejphar.2015.10.040
81. Vantur R, Rihar M, Koren A, Rijavec M, Kopac P, Bidovec-Stojkovic U, et al. Chemokines During Anaphylaxis: The Importance of CCL2 and CCL2-Dependent Chemotactic Activity for Basophils. *Clin Transl Allergy* (2020) 10(1):63. doi: 10.1186/s13601-020-00367-2
82. Wahlund C, Gucluler Akpinar G, Steiner L, Ibrahim A, Bandeira E, Lepzien R, et al. Sarcoidosis Exosomes Stimulate Monocytes to Produce Pro-Inflammatory Cytokines and CCL2. *Sci Rep* (2020) 10(1):15328. doi: 10.1038/s41598-020-72067-7
83. Persaud AT, Bennett SA, Thaya L, Burnie J, Guzzo C. Human Monocytes Store and Secrete Preformed CCL5, Independent of De Novo Protein Synthesis. *J Leukoc Biol* (2021). doi: 10.1002/JLB.3A0820-522RR

84. Mendez-Barbero N, Yuste-Montalvo A, Nuñez-Borque E, Jensen BM, Gutiérrez-Muñoz C, Tome-Amat J, et al. The TNF-Like Weak Inducer of the Apoptosis/Fibroblast Growth Factor-Inducible Molecule 14 Axis Mediates Histamine and Platelet-Activating Factor-Induced Subcutaneous Vascular Leakage and Anaphylactic Shock. *J Allergy Clin Immunol* (2020) 145(2):583–96.e6. doi: 10.1016/j.jaci.2019.09.019
85. McManus LM, Morley CA, Levine SP, Pinckard RN. Platelet Activating Factor (PAF) Induced Release of Platelet Factor 4 (PF4) *In Vitro* and During IgE Anaphylaxis in the Rabbit. *J Immunol* (1979) 123(6):2835–41.
86. Francis A, Bosio E, Stone SF, Fatovich DM, Arendts G, Nagree Y, et al. Neutrophil Activation During Acute Human Anaphylaxis: Analysis of MPO and Scd62l. *Clin Exp Allergy* (2017) 47(3):361–70. doi: 10.1111/cea.12868
87. Sunderkötter C, Steinbrink K, Goebeler M, Bhardwaj R, Sorg C. Macrophages and Angiogenesis. *J Leukoc Biol* (1994) 55(3):410–22. doi: 10.1002/jlb.55.3.410
88. Aratani Y. Myeloperoxidase: Its Role for Host Defense, Inflammation, and Neutrophil Function. *Arch Biochem Biophys* (2018) 640:47–52. doi: 10.1016/j.jabb.2018.01.004
89. Escribese MM, Rosace D, Chivato T, Fernández TD, Corbí AL, Barber D. Alternative Anaphylactic Routes: The Potential Role of Macrophages. *Front Immunol* (2017) 8:515. doi: 10.3389/fimmu.2017.00515
90. Jiao D, Liu Y, Lu X, Liu B, Pan Q, Liu Y, et al. Macrophages are the Dominant Effector Cells Responsible for IgG-Mediated Passive Systemic Anaphylaxis Challenged by Natural Protein Antigen in BALB/c and C57BL/6 Mice. *Cell Immunol* (2014) 289(1–2):97–105. doi: 10.1016/j.cellimm.2014.03.018
91. Jönsson F, de Chaisemartin L, Granger V, et al. An IgG-Induced Neutrophil Activation Pathway Contributes to Human Drug-Induced Anaphylaxis. *Sci Trans Med* (2019) 11(500). doi: 10.1126/scitranslmed.aat1479
92. Spoerl D, Nigolian H, Czarnetzki C, Harr T. Reclassifying Anaphylaxis to Neuromuscular Blocking Agents Based on the Presumed Patho-Mechanism: IgE-Mediated, Pharmacological Adverse Reaction or “Innate Hypersensitivity”? *Int J Mol Sci* (2017) 18(6):E1223. doi: 10.3390/ijms18061223
93. Subramanian H, Gupta K, Ali H. Roles of Mas-Related G Protein-Coupled Receptor X2 on Mast Cell-Mediated Host Defense, Pseudoallergic Drug Reactions, and Chronic Inflammatory Diseases. *J Allergy Clin Immunol* (2016) 138(3):700–10. doi: 10.1016/j.jaci.2016.04.051
94. Lieberman P, Garvey LH. Mast Cells and Anaphylaxis. *Curr Allergy Asthma Rep* (2016) 16(3):20. doi: 10.1007/s11882-016-0598-5
95. Ali H. Revisiting the Role of MRGPRX2 on Hypersensitivity Reactions to Neuromuscular Blocking Drugs. *Curr Opin Immunol* (2021) 72:65–71. doi: 10.1016/j.coi.2021.03.011
96. Elst J, Maurer M, Sabato V, Faber MA, Bridts CH, Mertens C, et al. Novel Insights on MRGPRX2-Mediated Hypersensitivity to Neuromuscular Blocking Agents And Fluoroquinolones. *Front Immunol* (2021) 12:668962. doi: 10.3389/fimmu.2021.668962
97. Liu R, Che D, Zhao T, Pundir P, Cao J, Lv Y, et al. MRGPRX2 is Essential for Sinomenine Hydrochloride Induced Anaphylactoid Reactions. *Biochem Pharmacol* (2017) 146:214–23. doi: 10.1016/j.bcp.2017.09.017
98. Gao Y, Han Y, Zhang X, Fei Q, Qi R, Hou R, et al. Penicillin Causes Non-Allergic Anaphylaxis by Activating the Contact System. *Sci Rep* (2020) 10(1):14160. doi: 10.1038/s41598-020-71083-x
99. Weg VB, Watson ML, Faccioli LH, Williams TJ. Investigation of the Endogenous Chemoattractants Involved in 111In-Eosinophil Accumulation in Passive Cutaneous Anaphylactic Reactions in the Guinea-Pig. *Br J Pharmacol* (1994) 113(1):35–42. doi: 10.1111/j.1476-5381.1994.tb16170.x
100. Edston E. Accumulation of Eosinophils, Mast Cells, and Basophils in the Spleen in Anaphylactic Deaths. *Forensic Sci Med Pathol* (2013) 9(4):496–500. doi: 10.1007/s12024-013-9468-9
101. Beutier H, Hechler B, Godon O, Wang Y, Gillis CM, de Chaisemartin L, et al. Platelets Expressing IgG Receptor Fcγriia/CD32A Determine the Severity of Experimental Anaphylaxis. *Sci Immunol* (2018) 3(22):ean5997. doi: 10.1126/sciimmunol.aan5997
102. Li QJ, Duan JH, Hu GL, et al. [Epidemiological Characteristics and Control of Filariasis in Hunan Province]. *Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi* (1990) 8(2):134–7.
103. Gill P, Jindal NL, Jagdis A, Vadas P. Platelets in the Immune Response: Revisiting Platelet-Activating Factor in Anaphylaxis. *J Allergy Clin Immunol* (2015) 135(6):1424–32. doi: 10.1016/j.jaci.2015.04.019
104. Stone SF, Cotterell C, Isbister GK, Holdgate A, Brown SGA. Emergency Department Anaphylaxis Investigators. Elevated Serum Cytokines During Human Anaphylaxis: Identification of Potential Mediators of Acute Allergic Reactions. *J Allergy Clin Immunol* (2009) 124(4):786–92.e4. doi: 10.1016/j.jaci.2009.07.055
105. Stone SF, Bosco A, Jones A, Cotterell CL, van Eeden PE, Arendts G, et al. Genomic Responses During Acute Human Anaphylaxis are Characterized by Upregulation of Innate Inflammatory Gene Networks. *PLoS One* (2014) 9(7):e101409. doi: 10.1371/journal.pone.0101409
106. Gaudenzio N, Sibilano R, Marichal T, Starkl P, Reber LL, Cenac N, et al. Different Activation Signals Induce Distinct Mast Cell Degranulation Strategies. *J Clin Invest* (2016) 126(10):3981–98. doi: 10.1172/JCI85538
107. Metcalfe DD, Peavy RD, Gilfillan AM. Mechanisms of Mast Cell Signaling in Anaphylaxis. *J Allergy Clin Immunol* (2009) 124(4):639–46. doi: 10.1016/j.jaci.2009.08.035
108. Ben-Shoshan M, Clarke AE. Anaphylaxis: Past, Present and Future. *Allergy* (2011) 66(1):1–14. doi: 10.1111/j.1398-9995.2010.02422.x
109. Kaplan AP. Preventing Anaphylaxis Fatalities: Should We Target Bradykinin? *J Allergy Clin Immunol* (2020) 145(5):1365–6. doi: 10.1016/j.jaci.2020.01.043
110. Kodama T, Sekine H, Takahashi M, Iwaki D, Machida T, Kanno K, et al. Role of Complement in a Murine Model of Peanut-Induced Anaphylaxis. *Immunobiology* (2013) 218(6):844–50. doi: 10.1016/j.imbio.2012.10.003
111. Biethahn K, Orinska Z, Vigorito E, Goyeneche-Patino DA, Mirghomizadeh F, Föger N, et al. miRNA-155 Controls Mast Cell Activation by Regulating the PI3Ky Pathway and Anaphylaxis in a Mouse Model. *Allergy* (2014) 69(6):752–62. doi: 10.1111/all.12407
112. Perales-Chorda C, Obeso D, Twomey L, Rojas-Benedicto A, Puchades-Carrasco L, Roca M, et al. Characterization of Anaphylaxis Reveals Different Metabolic Changes Depending on Severity and Triggers. *Clin Exp Allergy* (2021) 51(10):1295–309. doi: 10.1111/cea.13991
113. Kim M, Kwon Y, Jung HS, Kim Y, Jeoung D. Fcεr1-HDAC3-MCP1 Signaling Axis Promotes Passive Anaphylaxis Mediated by Cellular Interactions. *Int J Mol Sci* (2019) 20(19):E4964. doi: 10.3390/ijms20194964
114. Pugsley MK, Tabrizchi R. The Vascular System. An Overview of Structure and Function. *J Pharmacol Toxicol Methods* (2000) 44(2):333–40. doi: 10.1016/s1056-8719(00)00125-8
115. Monahan-Earley R, Dvorak AM, Aird WC. Evolutionary Origins of the Blood Vascular System and Endothelium. *J Thromb Haemost* (2013) 11 Suppl 1:46–66. doi: 10.1111/jth.12253
116. Taylor AM, Bordoni B. Histology, Blood Vascular System, in: *StatPearls* (2021). StatPearls Publishing. Available at: <http://www.ncbi.nlm.nih.gov/books/NBK553217/> (Accessed October 12, 2021).
117. Tucker WD, Arora Y, Mahajan K. Anatomy, Blood Vessels, in: *StatPearls* (2021). StatPearls Publishing. Available at: <http://www.ncbi.nlm.nih.gov/books/NBK470401/> (Accessed October 12, 2021).
118. Mori S, Tretter JT, Spicer DE, Bolender DL, Anderson RH. What is the Real Cardiac Anatomy? *Clin Anat* (2019) 32(3):288–309. doi: 10.1002/ca.23340
119. Chaudhry R, Miao JH, Rehman A. Physiology, Cardiovascular, in: *StatPearls* (2021). StatPearls Publishing. Available at: <http://www.ncbi.nlm.nih.gov/books/NBK493197/> (Accessed October 12, 2021).
120. Dornbush S, Turnquest AE. Physiology, Heart Sounds, in: *StatPearls* (2021). StatPearls Publishing. Available at: <http://www.ncbi.nlm.nih.gov/books/NBK541010/> (Accessed October 12, 2021).
121. Pollock JD, Makaryus AN. Physiology, Cardiac Cycle, in: *StatPearls* (2021). StatPearls Publishing. Available at: <http://www.ncbi.nlm.nih.gov/books/NBK459327/> (Accessed October 12, 2021).
122. Marone G, Genovese A, Varricchi G, Granata F. Human Heart as a Shock Organ in Anaphylaxis. *Allergo J Int* (2014) 23(2):60–6. doi: 10.1007/s40629-014-0007-3
123. Jadidi M, Razian SA, Habibnezhad M, Anttila E, Kamenskiy A. Mechanical, Structural, and Physiologic Differences in Human Elastic and Muscular Arteries of Different Ages: Comparison of the Descending Thoracic Aorta to the Superficial Femoral Artery. *Acta Biomater* (2021) 119:268–83. doi: 10.1016/j.actbio.2020.10.035
124. Martinez-Lemus LA. The Dynamic Structure of Arterioles. *Basic Clin Pharmacol Toxicol* (2012) 110(1):5–11. doi: 10.1111/j.1742-7843.2011.00813.x

125. Grubb S, Cai C, Hald BO, Khennouf L, Murmu RP, Jensen A, et al. Precapillary Sphincters Maintain Perfusion in the Cerebral Cortex. *Nat Commun* (2020) 11(1):395. doi: 10.1038/s41467-020-14330-z
126. Augustin HG, Koh GY. Organotypic Vasculature: From Descriptive Heterogeneity to Functional Pathophysiology. *Science* (2017) 357(6353):eaal2379. doi: 10.1126/science.aal2379
127. Lauridsen HM, Pober JS, Gonzalez AL. A Composite Model of the Human Postcapillary Venule for Investigation of Microvascular Leukocyte Recruitment. *FASEB J* (2014) 28(3):1166–80. doi: 10.1096/fj.13-240986
128. Cox JL, Chiasson DA, Gotlieb AI. Stranger in a Strange Land: The Pathogenesis of Saphenous Vein Graft Stenosis With Emphasis on Structural and Functional Differences Between Veins and Arteries. *Prog Cardiovasc Dis* (1991) 34(1):45–68. doi: 10.1016/0033-0620(91)90019-i
129. MacColl E, Khalil RA. Matrix Metalloproteinases as Regulators of Vein Structure and Function: Implications in Chronic Venous Disease. *J Pharmacol Exp Ther* (2015) 355(3):410–28. doi: 10.1124/jpet.115.227330
130. Sansilvestri-Morel P, Fioretti F, Rupin A, Senni K, Fabiani JN, Godeau G, et al. Comparison of Extracellular Matrix in Skin and Saphenous Veins From Patients With Varicose Veins: Does the Skin Reflect Venous Matrix Changes? *Clin Sci (Lond)* (2007) 112(4):229–39. doi: 10.1042/CS20060170
131. Caggiati A, Phillips M, Lametschwandtner A, Allegra C. Valves in Small Veins and Venules. *Eur J Vasc Endovasc Surg* (2006) 32(4):447–52. doi: 10.1016/j.ejvs.2006.04.021
132. Jourde-Chiche N, Fakhouri F, Dou L, Bellien J, Burtey S, Frimat M, et al. Endothelium Structure and Function in Kidney Health and Disease. *Nat Rev Nephrol* (2019) 15(2):87–108. doi: 10.1038/s41581-018-0098-z
133. Rafii S, Butler JM, Ding BS. Angiocrine Functions of Organ-Specific Endothelial Cells. *Nature* (2016) 529(7586):316–25. doi: 10.1038/nature17040
134. Schmaier AH. The Contact Activation and Kallikrein/Kinin Systems: Pathophysiologic and Physiologic Activities. *J Thromb Haemost* (2016) 14(1):28–39. doi: 10.1111/jth.13194
135. Moore KH, Murphy HA, George EM. The Glycocalyx: A Central Regulator of Vascular Function. *Am J Physiol Regul Integr Comp Physiol* (2021) 320(4):R508–18. doi: 10.1152/ajpregu.00340.2020
136. Aird WC. Endothelial Cell Heterogeneity. *Cold Spring Harb Perspect Med* (2012) 2(1):a006429. doi: 10.1101/cshperspect.a006429
137. Jambusaria A, Hong Z, Zhang L, Srivastava S, Jana A, Toth PT, et al. Endothelial Heterogeneity Across Distinct Vascular Beds During Homeostasis and Inflammation. *Elife* (2020) 9:e51413. doi: 10.7554/eLife.51413
138. Aird WC. Phenotypic Heterogeneity of the Endothelium: I. Structure, Function, and Mechanisms. *Circ Res* (2007) 100(2):158–73. doi: 10.1161/01.RES.0000255691.76142.4a
139. Heltianu C, Simionescu M, Simionescu N. Histamine Receptors of the Microvascular Endothelium Revealed *In Situ* With a Histamine-Ferritin Conjugate: Characteristic High-Affinity Binding Sites in Venules. *J Cell Biol* (1982) 93(2):357–64. doi: 10.1083/jcb.93.2.357
140. Uhlig S, Yang Y, Waade J, Wittenberg C, Babendreyer A, Kuebler WM. Differential Regulation of Lung Endothelial Permeability. *Vitro situ Cell Physiol Biochem* (2014) 34(1):1–19. doi: 10.1159/000362980
141. Michel CC, Curry FE. Microvascular Permeability. *Physiol Rev* (1999) 79(3):703–61. doi: 10.1152/physrev.1999.79.3.703
142. Rho SS, Ando K, Fukuhara S. Dynamic Regulation of Vascular Permeability by Vascular Endothelial Cadherin-Mediated Endothelial Cell-Cell Junctions. *J Nippon Med Sch* (2017) 84(4):148–59. doi: 10.1272/jnms.84.148
143. Mehta D, Malik AB. Signaling Mechanisms Regulating Endothelial Permeability. *Physiol Rev* (2006) 86(1):279–367. doi: 10.1152/physrev.00012.2005
144. Radeva MY, Waschke J. Mind the Gap: Mechanisms Regulating the Endothelial Barrier. *Acta Physiol (Oxf)* (2018) 222(1). doi: 10.1111/apha.12860
145. Spindler V, Schlegel N, Waschke J. Role of GTPases in Control of Microvascular Permeability. *Cardiovasc Res* (2010) 87(2):243–53. doi: 10.1093/cvr/cvq086
146. Dejana E, Orsenigo F. Endothelial Adherens Junctions at a Glance. *J Cell Sci* (2013) 126(Pt 12):2545–9. doi: 10.1242/jcs.124529
147. Vestweber D, Winderlich M, Cagna G, Nottebaum AF. Cell Adhesion Dynamics at Endothelial Junctions: VE-Cadherin as a Major Player. *Trends Cell Biol* (2009) 19(1):8–15. doi: 10.1016/j.tcb.2008.10.001
148. Touyz RM, Alves-Lopes R, Rios FJ, Camargo LL, Anagnostopoulou A, Arner A, et al. Vascular Smooth Muscle Contraction in Hypertension. *Cardiovasc Res* (2018) 114(4):529–39. doi: 10.1093/cvr/cvy023
149. Shi J, Yang Y, Cheng A, Xu G, He F. Metabolism of Vascular Smooth Muscle Cells in Vascular Diseases. *Am J Physiol Heart Circ Physiol* (2020) 319(3):H613–31. doi: 10.1152/ajpheart.00220.2020
150. Bochaton-Piallat ML, Bäck M. Novel Concepts for the Role of Smooth Muscle Cells in Vascular Disease: Towards a New Smooth Muscle Cell Classification. *Cardiovasc Res* (2018) 114(4):477–80. doi: 10.1093/cvr/cvy031
151. Godo S, Shimokawa H. Endothelial Functions. *Arterioscler Thromb Vasc Biol* (2017) 37(9):e108–14. doi: 10.1161/ATVBAHA.117.309813
152. Cyr AR, Huckaby LV, Shiva SS, Zuckerbraun BS. Nitric Oxide and Endothelial Dysfunction. *Crit Care Clin* (2020) 36(2):307–21. doi: 10.1016/j.ccc.2019.12.009
153. Hatakeyama T, Pappas PJ, Hobson RW, Boric MP, Sessa WC, Durán WN. Endothelial Nitric Oxide Synthase Regulates Microvascular Hyperpermeability. *in vivo J Physiol* (2006) 574(Pt 1):275–81. doi: 10.1113/jphysiol.2006.108175
154. Durán WN, Breslin JW, Sánchez FA. The NO Cascade, eNOS Location, and Microvascular Permeability. *Cardiovasc Res* (2010) 87(2):254–61. doi: 10.1093/cvr/cvq139
155. Tennant M, McGeachie JK. Blood Vessel Structure and Function: A Brief Update on Recent Advances. *Aust N Z J Surg* (1990) 60(10):747–53. doi: 10.1111/j.1445-2197.1990.tb07468.x
156. Halper J. Basic Components of Vascular Connective Tissue and Extracellular Matrix. *Adv Pharmacol* (2018) 81:95–127. doi: 10.1016/bs.apha.2017.08.012
157. Munjal A, Bordoni B. Histology, Vascular, in: *StatPearls* (2021). StatPearls Publishing. Available at: <http://www.ncbi.nlm.nih.gov/books/NBK554407/> (Accessed October 12, 2021).
158. Sheng Y, Zhu L. The Crosstalk Between Autonomic Nervous System and Blood Vessels. *Int J Physiol Pathophysiol Pharmacol* (2018) 10(1):17–28.
159. Shao Y, Saredy J, Yang WY, Sun Y, Lu Y, Saaoud F, et al. Vascular Endothelial Cells and Innate Immunity. *Arterioscler Thromb Vasc Biol* (2020) 40(6):e138–52. doi: 10.1161/ATVBAHA.120.314330
160. Bucci T, Parente R, De Feo G, Cardamone C, Triggiani M. Flow-Mediated Dilation Shows Impaired Endothelial Function in Patients With Mastocytosis. *J Allergy Clin Immunol* (2019) 144(4):1106–11. doi: 10.1016/j.jaci.2019.05.037
161. Libby P, Lüscher T. COVID-19 is, in the End, an Endothelial Disease. *Eur Heart J* (2020) 41(32):3038–44. doi: 10.1093/eurheartj/ehaa623
162. Yuste-Montalvo A, Fernandez-Bravo S, Oliva T, Pastor-Vargas C, Betancor D, Goikoetxea MJ, et al. Proteomic and Biological Analysis of an *In Vitro* Human Endothelial System in Response to Drug Anaphylaxis. *Front Immunol* (2021) 12:692569. doi: 10.3389/fimmu.2021.692569
163. Sala-Cunill A, Björkqvist J, Senter R, Guilarde M, Cardona V, Labrador M, et al. Plasma Contact System Activation Drives Anaphylaxis in Severe Mast Cell-Mediated Allergic Reactions. *J Allergy Clin Immunol* (2015) 135(4):1031–43.e6. doi: 10.1016/j.jaci.2014.07.057
164. Fisher M. Blood Volume Replacement in Acute Anaphylactic Cardiovascular Collapse Related to Anaesthesia. *Br J Anaesth* (1977) 49(10):1023–6. doi: 10.1093/bja/49.10.1023
165. Fisher MM. Clinical Observations on the Pathophysiology and Treatment of Anaphylactic Cardiovascular Collapse. *Anaesth Intensive Care* (1986) 14(1):17–21. doi: 10.1177/0310057X8601400105
166. Egawa G, Nakamizo S, Natsuaki Y, Doi H, Miyachi Y, Kabashima K. Intravital Analysis of Vascular Permeability in Mice Using Two-Photon Microscopy. *Sci Rep* (2013) 3:1932. doi: 10.1038/srep01932
167. Callesen KT, Yuste-Montalvo A, Poulsen LK, Jensen BM, Esteban V. *In Vitro* Investigation of Vascular Permeability in Endothelial Cells From Human Artery, Vein and Lung Microvessels at Steady-State and Anaphylactic Conditions. *Biomedicines* (2021) 9(4):439. doi: 10.3390/biomedicines9040439
168. James LP, Austen KF. FATAL SYSTEMIC ANAPHYLAXIS IN MAN. *N Engl J Med* (1964) 270:597–603. doi: 10.1056/NEJM196403192701202
169. Sonin L, Grammer LC, Greenberger PA, Patterson R. Idiopathic Anaphylaxis: A Clinical Summary. *Ann Intern Med* (1983) 99(5):634–5. doi: 10.7326/0003-4819-99-5-634

170. Barthel G, Zheng F, Demoulin B, Davidson J, Montémont C, Gaburro J, et al. Biphasic Airway-Lung Response to Anaphylactic Shock in Brown Norway Rats. *Respir Physiol Neurobiol* (2013) 189(1):47–51. doi: 10.1016/j.resp.2013.07.003
171. Ebo DG, Clarke RC, Mertes PM, Platt PR, Sabato V, Sadleir PHM. Molecular Mechanisms and Pathophysiology of Perioperative Hypersensitivity and Anaphylaxis: A Narrative Review. *Br J Anaesth* (2019) 123(1):e38–49. doi: 10.1016/j.bja.2019.01.031
172. Miller RL, Shtessel M, Robinson LB, Banerji A. Advances in Drug Allergy, Urticaria, Angioedema, and Anaphylaxis in 2018. *J Allergy Clin Immunol* (2019) 144(2):381–92. doi: 10.1016/j.jaci.2019.06.010
173. Shirai T, Mori M, Uotani T, Chida K. Gastrointestinal Disorders in Anaphylaxis. *Intern Med* (2007) 46(6):315–6. doi: 10.2169/INTERNALMED.46.6126
174. Kaplan AP. Kinins, Airway Obstruction, and Anaphylaxis. *Chem Immunol Allergy* (2010) 95:67–84. doi: 10.1159/000315938
175. Yamani A, Wu D, Waggoner L, Noah T, Koleske AJ, Finkelman F, et al. The Vascular Endothelial Specific IL-4 Receptor Alpha-ABL1 Kinase Signaling Axis Regulates the Severity of IgE-Mediated Anaphylactic Reactions. *J Allergy Clin Immunol* (2018) 142(4):1159–72.e5. doi: 10.1016/j.jaci.2017.08.046
176. Ruiz-Garcia M, Bartra J, Alvarez O, Lakhani A, Patel S, Tang A, et al. Cardiovascular Changes During Peanut-Induced Allergic Reactions in Human Subjects. *J Allergy Clin Immunol* (2021) 147(2):633–42. doi: 10.1016/j.jaci.2020.06.033
177. Jacobsen RC, Gratton MC. A Case of Unrecognized Prehospital Anaphylactic Shock. *Prehosp Emerg Care* (2011) 15(1):61–6. doi: 10.3109/10903127.2010.519823
178. Wetschurck N, Strlic B, Offermanns S. Passing the Vascular Barrier: Endothelial Signaling Processes Controlling Extravasation. *Physiol Rev* (2019) 99(3):1467–525. doi: 10.1152/physrev.00037.2018
179. Bazzoni G, Dejana E. Endothelial Cell-to-Cell Junctions: Molecular Organization and Role in Vascular Homeostasis. *Physiol Rev* (2004) 84(3):869–901. doi: 10.1152/physrev.00035.2003
180. Somlyo AP, Somlyo AV. Ca²⁺ Sensitivity of Smooth Muscle and Nonmuscle Myosin II: Modulated by G Proteins, Kinases, and Myosin Phosphatase. *Physiol Rev* (2003) 83(4):1325–58. doi: 10.1152/physrev.00023.2003
181. Mikelis CM, Simaan M, Ando K, Fukuhara S, Sakurai A, Amornphimoltham P, et al. RhoA and ROCK Mediate Histamine-Induced Vascular Leakage and Anaphylactic Shock. *Nat Commun* (2015) 6:6725. doi: 10.1038/ncomms7725
182. Chislock EM, Pendergast AM. Abl Family Kinases Regulate Endothelial Barrier Function *In Vitro* and in Mice. *PLoS One* (2013) 8(12):e85231. doi: 10.1371/journal.pone.0085231
183. Hox V, O'Connell MP, Lyons JJ, Sackstein P, Dimaggio T, Jones N, et al. Diminution of Signal Transducer and Activator of Transcription 3 Signaling Inhibits Vascular Permeability and Anaphylaxis. *J Allergy Clin Immunol* (2016) 138(1):187–99. doi: 10.1016/j.jaci.2015.11.024
184. Wawrzyniak M, Pich C, Gross B, Schütz F, Fleury S, Quemener S, et al. Endothelial, But Not Smooth Muscle, Peroxisome Proliferator-Activated Receptor β/δ Regulates Vascular Permeability and Anaphylaxis. *J Allergy Clin Immunol* (2015) 135(6):1625–35.e5. doi: 10.1016/j.jaci.2014.11.006
185. Alfano DN, Klei LR, Klei HB, Trotta M, Gough PJ, Foley KP, et al. MALT1 Protease Plays a Dual Role in the Allergic Response by Acting in Both Mast Cells and Endothelial Cells. *J Immunol* (2020) 204(9):2337–48. doi: 10.4049/jimmunol.1900281
186. Nuñez-Borque E, Fernandez-Bravo S, Pastor-Vargas C, Alvarez-Llamas G, Gutierrez-Blazquez MD, Alwashali E, et al. Proteomic Profile of Extracellular Vesicles in Anaphylaxis and Their Role in Vascular Permeability. *Allergy* (2021) 76(7):2276–9. doi: 10.1111/all.14792
187. Cui H, Okamoto Y, Yoshioka K, Du W, Takuwa N, Zhang W, et al. Sphingosine-1-Phosphate Receptor 2 Protects Against Anaphylactic Shock Through Suppression of Endothelial Nitric Oxide Synthase in Mice. *J Allergy Clin Immunol* (2013) 132(5):1205–14.e9. doi: 10.1016/j.jaci.2013.07.026
188. Gazit SL, Mariko B, Thérond P, Decouture B, Xiong Y, Couty L, et al. Platelet and Erythrocyte Sources of S1P Are Redundant for Vascular Development and Homeostasis, But Both Rendered Essential After Plasma S1P Depletion in Anaphylactic Shock. *Circ Res* (2016) 119(8):e110–126. doi: 10.1161/CIRCRESAHA.116.308929
189. Wilkerson BA, Argraves KM. The Role of Sphingosine-1-Phosphate in Endothelial Barrier Function. *Biochim Biophys Acta* (2014) 1841(10):1403–12. doi: 10.1016/j.bbali.2014.06.012
190. Camerer E, Regard JB, Cornelissen I, Srinivasan Y, Duong DN, Palmer D, et al. Sphingosine-1-Phosphate in the Plasma Compartment Regulates Basal and Inflammation-Induced Vascular Leak in Mice. *J Clin Invest* (2009) 119(7):1871–9. doi: 10.1172/jci38575
191. Yoshioka K, Yoshida K, Cui H, Wakayama T, Takuwa N, Okamoto Y, et al. Endothelial PI3K-C2 α , a Class II PI3K, has an Essential Role in Angiogenesis and Vascular Barrier Function. *Nat Med* (2012) 18(10):1560–9. doi: 10.1038/nm.2928
192. Nakamura T, Fujiwara Y, Yamada R, Fujii W, Hamabata T, Lee MY, et al. Mast Cell-Derived Prostaglandin D2 Attenuates Anaphylactic Reactions in Mice. *J Allergy Clin Immunol* (2017) 140(2):630–2.e9. doi: 10.1016/j.jaci.2017.02.030
193. Ballesteros-Martinez C, Mendez-Barbero N, Montalvo-Yuste A, Jensen BM, Gomez-Cardenosa A, Klitfod L, et al. Endothelial Regulator of Calcineurin 1 Promotes Barrier Integrity and Modulates Histamine-Induced Barrier Dysfunction in Anaphylaxis. *Front Immunol* (2017) 8:1323. doi: 10.3389/fimmu.2017.01323
194. Smith N, Lopez RA, Silberman M. Distributive Shock, in: *StatPearls* (2021). StatPearls Publishing. Available at: <http://www.ncbi.nlm.nih.gov/books/NBK470316/> (Accessed October 12, 2021).
195. Burnstock G. Purinergic Signaling in the Cardiovascular System. *Circ Res* (2017) 120(1):207–28. doi: 10.1161/CIRCRESAHA.116.309726
196. Smith PL, Kagey-Sobotka A, Bleecker ER, Traystman R, Kaplan AP, Gralnick H, et al. Physiologic Manifestations of Human Anaphylaxis. *J Clin Invest* (1980) 66(5):1072–80. doi: 10.1172/JCI109936
197. Brown SGA. Cardiovascular Aspects of Anaphylaxis: Implications for Treatment and Diagnosis. *Curr Opin Allergy Clin Immunol* (2005) 5(4):359–64. doi: 10.1097/01.all.0000174158.78626.35
198. Faye N, Fournier L, Balvay D, Thiam R, Orliaguet G, Clément O, et al. Macromolecular Capillary Leakage is Involved in the Onset of Anaphylactic Hypotension. *Anesthesiology* (2012) 117(5):1072–9. doi: 10.1097/ALN.0b013e31826d3dc5
199. Rothe CF. Physiology of Venous Return. An Unappreciated Boost to the Heart. *Arch Intern Med* (1986) 146(5):977–82. doi: 10.1001/archinte.1986.00360170223028
200. Silverman HJ, Van Hook C, Haponik EF. Hemodynamic Changes in Human Anaphylaxis. *Am J Med* (1984) 77(2):341–4. doi: 10.1016/0002-9343(84)90717-4
201. Brown SGA. The Pathophysiology of Shock in Anaphylaxis. *Immunol Allergy Clin North Am* (2007) 27(2):165–75. doi: 10.1016/j.iac.2007.03.003
202. Osada S, Ichiki H, Oku H, Ishiguro K, Kunitomo M, Semma M. Participation of Nitric Oxide in Mouse Anaphylactic Hypotension. *Eur J Pharmacol* (1994) 252(3):347–50. doi: 10.1016/0014-2999(94)90185-6
203. Cauwels A. Nitric Oxide in Shock. *Kidney Int* (2007) 72(5):557–65. doi: 10.1038/sj.ki.5002340
204. Cauwels A, Janssen B, Buys E, Sips P, Brouckaert P. Anaphylactic Shock Depends on PI3K and eNOS-Derived NO. *J Clin Invest* (2006) 116(8):2244–51. doi: 10.1172/JCI25426
205. Nakamura Y, Hashiba Y, Endo J, Furuie M, Isozaki A, Yagi K. Elevated Exhaled Nitric Oxide in Anaphylaxis With Respiratory Symptoms. *Allergol Int* (2015) 64(4):359–63. doi: 10.1016/j.alit.2015.05.005
206. O'Mahony L, Akdis M, Akdis CA. Regulation of the Immune Response and Inflammation by Histamine and Histamine Receptors. *J Allergy Clin Immunol* (2011) 128(6):1153–62. doi: 10.1016/j.jaci.2011.06.051
207. Jin H, Koyama T, Hatanaka Y, Akiyama S, Takayama F, Kawasaki H. Histamine-Induced Vasodilation and Vasoconstriction in the Mesenteric Resistance Artery of the Rat. *Eur J Pharmacol* (2006) 529(1–3):136–44. doi: 10.1016/j.ejphar.2005.10.060
208. Payne V, Kam PCA. Mast Cell Tryptase: A Review of Its Physiology and Clinical Significance. *Anaesthesia* (2004) 59(7):695–703. doi: 10.1111/j.1365-2044.2004.03757.x
209. Udem BJ, Taylor-Clark T. Mechanisms Underlying the Neuronal-Based Symptoms of Allergy. *J Allergy Clin Immunol* (2014) 133(6):1521–34. doi: 10.1016/j.jaci.2013.11.027
210. Ng MFY. The Role of Mast Cells in Wound Healing. *Int Wound J* (2010) 7(1):55–61. doi: 10.1111/j.1742-481X.2009.00651.x
211. Kounis NG, Cervellini G, Koniari I, Bonfanti L, Dousdampanis P, Charokopos N, et al. Anaphylactic Cardiovascular Collapse and Kounis

- Syndrome: Systemic Vasodilation or Coronary Vasoconstriction? *Ann Transl Med* (2018) 6(17):332. doi: 10.21037/atm.2018.09.05
212. Felix SB, Baumann G, Berdel WE. Systemic Anaphylaxis—Separation of Cardiac Reactions From Respiratory and Peripheral Vascular Events. *Res Exp Med (Berl)* (1990) 190(4):239–52. doi: 10.1007/BF00000029
 213. Lieberman P. The Use of Antihistamines in the Prevention and Treatment of Anaphylaxis and Anaphylactoid Reactions. *J Allergy Clin Immunol* (1990) 86(4 Pt 2):684–6. doi: 10.1016/s0091-6749(05)80241-6
 214. Liu M, Yokomizo T. The Role of Leukotrienes in Allergic Diseases. *Allergol Int* (2015) 64(1):17–26. doi: 10.1016/j.alit.2014.09.001
 215. Smyth EM. Thromboxane and the Thromboxane Receptor in Cardiovascular Disease. *Clin Lipidol* (2010) 5(2):209–19. doi: 10.2217/clp.10.11
 216. Dell'Italia LJ, Collawn JF, Ferrario CM. Multifunctional Role of Chymase in Acute and Chronic Tissue Injury and Remodeling. *Circ Res* (2018) 122(2):319–36. doi: 10.1161/CIRCRESAHA.117.310978
 217. Shinomiya S, Shibamoto T, Kurata Y, Kuda Y, Zhang W, Tanida M, et al. Nitric Oxide and $\beta(2)$ -Adrenoceptor Activation Attenuate Pulmonary Vasoconstriction During Anaphylactic Hypotension in Anesthetized BALB/c Mice. *Exp Lung Res* (2013) 39(3):119–29. doi: 10.3109/01902148.2013.768720
 218. Kuda Y, Kurata Y, Wang M, Tanida M, Shibamoto T. Major Contribution of Vasospasm-Induced Coronary Blood Flow Reduction to Anaphylactic Ventricular Dysfunction Assessed in Isolated Blood-Perfused Rat Heart. *Cardiol J* (2014) 21(1):11–7. doi: 10.5603/CJ.a2013.0047
 219. Cui S, Shibamoto T, Takano H, Zhang W, Kurata Y. Leukotrienes and Cyclooxygenase Products Mediate Anaphylactic Venoconstriction in Ovalbumin Sensitized Rat Livers. *Eur J Pharmacol* (2007) 576(1-3):99–106. doi: 10.1016/j.ejphar.2007.07.046
 220. Melli G, Folli G, Mazzei D, Vitolo E, Sacchi A. SHOCK ORGAN AND SHOCK TISSUE IN VARIOUS ANIMAL SPECIES. *Acta Allergol* (1963) 18:188–210. doi: 10.1111/j.1398-9995.1963.tb03176.x
 221. Auer J, Lewis PA. THE PHYSIOLOGY OF THE IMMEDIATE REACTION OF ANAPHYLAXIS IN THE GUINEA-PIG. *J Exp Med* (1910) 12(2):151–75. doi: 10.1084/jem.12.2.151
 222. LoVerde D, Iweala OI, Eginli A, Krishnaswamy G. Anaphylaxis. *Chest* (2018) 153(2):528–43. doi: 10.1016/j.chest.2017.07.033
 223. Schadt JC, Ludbrook J. Hemodynamic and Neurohumoral Responses to Acute Hypovolemia in Conscious Mammals. *Am J Physiol* (1991) 260(2 Pt 2):H305–18. doi: 10.1152/ajpheart.1991.260.2.H305
 224. Wetschurck N, Offermanns S. Mammalian G Proteins and Their Cell Type Specific Functions. *Physiol Rev* (2005) 85(4):1159–204. doi: 10.1152/physrev.00003.2005
 225. Pierce KL, Premont RT, Lefkowitz RJ. Seven-Transmembrane Receptors. *Nat Rev Mol Cell Biol* (2002) 3(9):639–50. doi: 10.1038/nrm908
 226. Syrovatkin V, Alegre KO, Dey R, Huang XY. Regulation, Signaling, and Physiological Functions of G-Proteins. *J Mol Biol* (2016) 428(19):3850–68. doi: 10.1016/j.jmb.2016.08.002
 227. White MV. The Role of Histamine in Allergic Diseases. *J Allergy Clin Immunol* (1990) 86(4 Pt 2):599–605. doi: 10.1016/s0091-6749(05)80223-4
 228. Wechsler JB, Schroeder HA, Byrne AJ, Chien KB, Bryce PJ. Anaphylactic Responses to Histamine in Mice Utilize Both Histamine Receptors 1 and 2. *Allergy* (2013) 68(10):1338–40. doi: 10.1111/all.12227
 229. Vigorito C, Russo P, Picotti GB, Chiariello M, Poto S, Marone G. Cardiovascular Effects of Histamine Infusion in Man. *J Cardiovasc Pharmacol* (1983) 5(4):531–7. doi: 10.1097/00005344-198307000-00004
 230. Vadas P. The Platelet-Activating Factor Pathway in Food Allergy and Anaphylaxis. *Ann Allergy Asthma Immunol* (2016) 117(5):455–7. doi: 10.1016/j.anai.2016.05.003
 231. Montrucchio G, Alloati G, Camussi G. Role of Platelet-Activating Factor in Cardiovascular Pathophysiology. *Physiol Rev* (2000) 80(4):1669–99. doi: 10.1152/physrev.2000.80.4.1669
 232. Lyons JJ, Chovanec J, O'Connell MP, Liu Y, Šelj J, Zanotti R, et al. Heritable Risk for Severe Anaphylaxis Associated With Increased α -Trypsin-Encoding Germline Copy Number at TPSAB1. *J Allergy Clin Immunol* (2021) 147(2):622–32. doi: 10.1016/j.jaci.2020.06.035
 233. Kempkes C, Buddenkotte J, Cevikbas F, Buhl T, Steinhoff M. PAR-2 in Neuroimmune Communication and Itch, in: *Itch: Mechanisms and Treatment*. *Frontiers in Neuroscience* (2014). CRC Press/Taylor & Francis. Available at: <http://www.ncbi.nlm.nih.gov/books/NBK200911/> (Accessed November 30, 2021).
 234. Walch L, Norel X, Gascard JP, Brink C. Functional Studies of Leukotriene Receptors in Vascular Tissues(2000) (Accessed 16(2 Pt 2)).
 235. Hui Y, Cheng Y, Smalera I, Jian W, Goldhahn L, Fitzgerald GA, et al. Directed Vascular Expression of Human Cysteinyl Leukotriene 2 Receptor Modulates Endothelial Permeability and Systemic Blood Pressure. *Circulation* (2004) 110(21):3360–6. doi: 10.1161/01.CIR.0000147775.50954.AA
 236. Maekawa A, Austen KF, Kanaoka Y. Targeted Gene Disruption Reveals the Role of Cysteinyl Leukotriene 1 Receptor in the Enhanced Vascular Permeability of Mice Undergoing Acute Inflammatory Responses. *J Biol Chem* (2002) 277(23):20820–4. doi: 10.1074/jbc.M203163200
 237. Rastogi S, Willmes DM, Nassiri M, Babina M, Worm M. PGE2 Deficiency Predisposes to Anaphylaxis by Causing Mast Cell Hyperresponsiveness. *J Allergy Clin Immunol* (2020) 146(6):1387–96.e13. doi: 10.1016/j.jaci.2020.03.046
 238. Rasković S, Bogić M, Perić-Popadić A, Arandjelović S, Jovčić Z, Tomić-Spirić V. [The Role of Prostaglandins in Allergic Inflammation]. *Srp Arh Celok Lek* (1998) 126(9-10):388–93.
 239. Bender L, Weidmann H, Rose-John S, Renné T, Long AT. Factor XII-Driven Inflammatory Reactions With Implications for Anaphylaxis. *Front Immunol* (2017) 8:1115. doi: 10.3389/fimmu.2017.01115
 240. Olivera A, Eisner C, Kitamura Y, Dillahunt S, Allende L, Tuymetova G, et al. Sphingosine Kinase 1 and Sphingosine-1-Phosphate Receptor 2 are Vital to Recovery From Anaphylactic Shock in Mice. *J Clin Invest* (2010) 120(5):1429–40. doi: 10.1172/JCI40659
 241. Olivera A, Dillahunt SE, Rivera J. Interrogation of Sphingosine-1-Phosphate Receptor 2 Function In Vivo Reveals a Prominent Role in the Recovery From IgE and IgG-Mediated Anaphylaxis With Minimal Effect on Its Onset. *Immunol Lett* (2013) 150(1-2):89–96. doi: 10.1016/j.imlet.2013.01.005
 242. Schäfer B, Piliponsky AM, Oka T, Song CH, Gerard NP, Gerard C, et al. Mast Cell Anaphylatoxin Receptor Expression can Enhance IgE-Dependent Skin Inflammation in Mice. *J Allergy Clin Immunol* (2013) 131(2):541–8.e1-9. doi: 10.1016/j.jaci.2012.05.009
 243. Tőro K, Borka K, Kardos M, Kristóf I, Sótónyi P. Expression and Function of C5a Receptor in a Fatal Anaphylaxis After Honey Bee Sting. *J Forensic Sci* (2011) 56(2):526–8. doi: 10.1111/j.1556-4029.2010.01681.x
 244. Schraufstatter IU, Trieu K, Sikora L, Sriramarao P, DiScipio R. Complement C3a and C5a Induce Different Signal Transduction Cascades in Endothelial Cells. *J Immunol* (2002) 169(4):2102–10. doi: 10.4049/jimmunol.169.4.2102
 245. Kemp SF, Lockey RF, Simons FER. World Allergy Organization ad hoc Committee on Epinephrine in Anaphylaxis. Epinephrine: The Drug of Choice for Anaphylaxis. A Statement of the World Allergy Organization. *Allergy* (2008) 63(8):1061–70. doi: 10.1111/j.1398-9995.2008.01733.x
 246. Simons KJ, Simons FER. Epinephrine and Its Use in Anaphylaxis: Current Issues. *Curr Opin Allergy Clin Immunol* (2010) 10(4):354–61. doi: 10.1097/ACI.0b013e32833bc670
 247. Korhonen H, Fisslthaler B, Moers A, Wirth A, Habermehl D, Wieland T, et al. Anaphylactic Shock Depends on Endothelial Gq/G11. *J Exp Med* (2009) 206(2):411–20. doi: 10.1084/jem.20082150
 248. Simons FER, Ebisawa M, Sanchez-Borges M, Thong BY, Worm M, Tanno LK, et al. 2015 Update of the Evidence Base: World Allergy Organization Anaphylaxis Guidelines. *World Allergy Organ J* (2015) 8(1):32. doi: 10.1186/s40413-015-0080-1
 249. Kemp AM, Kemp SF. Pharmacotherapy in Refractory Anaphylaxis: When Intramuscular Epinephrine Fails. *Curr Opin Allergy Clin Immunol* (2014) 14(4):371–8. doi: 10.1097/ACI.0000000000000080
 250. Nagano K, Kwon C, Ishida J, Hashimoto T, Kim JD, Kishikawa N, et al. Cooperative Action of APJ and α 1a-Adrenergic Receptor in Vascular Smooth Muscle Cells Induces Vasoconstriction. *J Biochem* (2019) 166(5):383–92. doi: 10.1093/jb/mvz071
 251. Motiejunaite J, Amar L, Vidal-Petiot E. Adrenergic Receptors and Cardiovascular Effects of Catecholamines. *Ann Endocrinol (Paris)* (2021) 82(3-4):193–7. doi: 10.1016/j.ando.2020.03.012
 252. Ring J, Klimek L, Worm M. Adrenaline in the Acute Treatment of Anaphylaxis. *Dtsch Arztebl Int* (2018) 115(31-32):528–34. doi: 10.3238/arztebl.2018.0528

253. Lieberman P, Simons FER. Anaphylaxis and Cardiovascular Disease: Therapeutic Dilemmas. *Clin Exp Allergy* (2015) 45(8):1288–95. doi: 10.1111/cea.12520
254. Spindler V, Waschke J. Beta-Adrenergic Stimulation Contributes to Maintenance of Endothelial Barrier Functions Under Baseline Conditions. *Microcirculation* (2011) 18(2):118–27. doi: 10.1111/j.1549-8719.2010.00072.x
255. Simons FER. First-Aid Treatment of Anaphylaxis to Food: Focus on Epinephrine. *J Allergy Clin Immunol* (2004) 113(5):837–44. doi: 10.1016/j.jaci.2004.01.769
256. Muraro A, Roberts G, Worm M, Bilò MB, Brockow K, Fernández-Rivas M, et al. Anaphylaxis: Guidelines From the European Academy of Allergy and Clinical Immunology. *Allergy* (2014) 69(8):1026–45. doi: 10.1111/all.12437
257. McCartney SL, Duce L, Ghadimi K. Intraoperative Vasoplegia: Methylene Blue to the Rescue! *Curr Opin Anaesthesiol* (2018) 31(1):43–9. doi: 10.1097/ACO.0000000000000548
258. Platt PR, Clarke RC, Johnson GH, Sadleir PHM. Efficacy of Sugammadex in Rocuronium-Induced or Antibiotic-Induced Anaphylaxis. A Case-Control Study. *Anaesthesia* (2015) 70(11):1264–7. doi: 10.1111/anae.13178
259. Arias K, Baig M, Colangelo M, Chu D, Walker T, Goncharova S, et al. Concurrent Blockade of Platelet-Activating Factor and Histamine Prevents Life-Threatening Peanut-Induced Anaphylactic Reactions. *J Allergy Clin Immunol* (2009) 124(2):307–314, 314. doi: 10.1016/j.jaci.2009.03.012
260. Labella M, Garcia-Neuer M, Castells M. Application of Precision Medicine to the Treatment of Anaphylaxis. *Curr Opin Allergy Clin Immunol* (2018) 18(3):190–7. doi: 10.1097/ACI.0000000000000435
261. Tanno LK, Martin B. Biologic Agents for the Treatment of Anaphylaxis. *Immunol Allergy Clin North Am* (2020) 40(4):625–33. doi: 10.1016/j.iac.2020.06.006
262. Ricciardi L. Omalizumab: A Useful Tool for Inducing Tolerance to Bee Venom Immunotherapy. *Int J Immunopathol Pharmacol* (2016) 29(4):726–8. doi: 10.1177/0394632016670920
263. Tanno LK, Alvarez-Perea A, Pouessel G. Therapeutic Approach of Anaphylaxis. *Curr Opin Allergy Clin Immunol* (2019) 19(4):393–401. doi: 10.1097/ACI.0000000000000539
264. Vickery BP, Ebisawa M, Shreffler WG, Wood RA. Current and Future Treatment of Peanut Allergy. *J Allergy Clin Immunol Pract* (2019) 7(2):357–65. doi: 10.1016/j.jaip.2018.11.049
265. Virkud YV, Wang J, Shreffler WG. Enhancing the Safety and Efficacy of Food Allergy Immunotherapy: A Review of Adjunctive Therapies. *Clin Rev Allergy Immunol* (2018) 55(2):172–89. doi: 10.1007/s12016-018-8694-z
266. Bruhns P, Chollet-Martin S. Mechanisms of Human Drug-Induced Anaphylaxis. *J Allergy Clin Immunol* (2021) 147(4):1133–42. doi: 10.1016/j.jaci.2021.02.013

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Nuñez-Borque, Fernandez-Bravo, Yuste-Montalvo and Esteban. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.