## Research Article

# A Case-Control Study between Gene Polymorphisms of Polyunsaturated Fatty Acid Metabolic Rate-Limiting Enzymes and Acute Coronary Syndrome in Chinese Han Population

#### Zikai Song, Hongyan Cao, Ling Qin, and Yanfang Jiang

Department of Cardiology, the Second Division of the First Hospital, Jilin University, 3302 JiLin Street, Changchun 130031, China

Correspondence should be addressed to Ling Qin; qinling@medmail.com.cn and Yanfang Jiang; yanfangjiang@hotmail.com

Received 5 September 2012; Accepted 14 January 2013

Academic Editor: Joseph Fomusi Ndisang

Copyright © 2013 Zikai Song et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The purpose of this study is to analyze the relationship between the polymorphisms of fatty acid desaturase 1 (*FADS*1), fatty acid desaturase 2 (*FADS*2), and elongation of very long-chain fatty acids-like 2 (*ELOVL2*) and acute coronary syndrome (ACS) in Chinese Han population. Therefore, we selected three single nucleotide polymorphisms (SNPs) from these candidate genes and genotyped them using PCR-based restriction fragment length polymorphism analysis in 249 ACS patients and 240 non-ACS subjects, as were Han Chinese ancestry. The results showed that rs174556 in the *FADS*1 gene is found to be in allelic association (P = 0.003) and genotypic association (P = 0.036) with ACS. The frequencies of rs174556 minor allele (T) in case group were obviously higher than in control group. The trans-phase gene-gene interaction analysis showed that the combined genotype of rs174556 (T/T) and rs3756963 (T/T) was associated with ACS (P = 0.031). And the results suggest that, for rs174556 C>T, the CT/TT genotypes were more likely to lead in ACS in subjects with hypertension after correction of all risk factors (OR = 4.236, 95% CI, 2.216–7.126). These findings suggest that the polymorphisms of rs174556 in the *FADS*1 gene are very likely to be associated with ACS in Chinese Han population, especially in subjects with hypertension.

## 1. Introduction

Acute coronary syndrome (ACS) is a common disease, and a major determinant of morbidity and mortality in all races, ethnicities, and cultures [1], which is caused by a combination of genetic background and environmental factors. The epidemic of coronary artery disease (CAD), especially its manifestation as ACS, is a global issue that accounts for more than 80% of the burden of this disease in developing countries [2] and results in approximately 30% of all deaths worldwide each year [3]. The spectrum of ACS ranges from unstable angina pectoris (UAP) to acute myocardial infarction (AMI), including ST-segment elevation myocardial infarction (STEMI) and non-STEMI (NSTEMI) [4]. Atherosclerotic plaque instability is the main feature of ACS pathogenesis.

Blood and tissue contents of polyunsaturated fatty acid (PUFA) and long-chain PUFA (LC-PUFA) are related to numerous health outcomes including cardiovascular health, allergies, mental health, and cognitive development [5]. There

are two families of PUFA, and they are classified as omega-3 (n-3) and omega-6 (n-6) based on the location of the last double bond relative to the terminal methyl end of the molecule [6]. Evidence from various research paradigms supports the cardiovascular benefits of a high intake of n-3 polyunsaturated fatty acids (PUFAs), especially the longchain, marine-derived n-3 PUFA, eicosapentaenoic acids, and docosahexaenoic acids [7, 8]. And n-6 PUFA are well known for their critical role in many physiological functions and seem to reduce risks of CAD [9].

Both families of fatty acids, n-3 and n-6, share and compete for the same enzymes ( $\Delta$ 6-desaturase,  $\Delta$ 5-desaturase, and elongases) in their biosynthesis, and  $\Delta$ 6-desaturase is the rate-limiting step [10–12].  $\Delta$ 5-desaturase (D5D) and  $\Delta$ 6-desaturase (D6D) are encoded by the genes *FADS1* and *FADS2*, respectively, which form a gene cluster jointly with the gene for fatty acid desaturase 3 (*FADS3*) on the human chromosome 11q12-q13.1 [12, 13]. And D5D and D6D mainly involved in regulating the levels of proinflammatory and anti-inflammatory eicosanoids derived from PUFAs [14]. Additionally, another essential enzyme, elongase, is involved in the homeostasis of longer chain n-3 fatty acids, which is encoded by elongation of very long-chain fatty acids-like 2 (FEN1/Elo2, SUR4/Elo3, yeast) (*ELOVL2*) gene [15].

Polymorphisms in the genes FADS1 and FADS2 are associated with n-3 and n-6 fatty acid levels and especially with arachidonic acid (ARA) amounts in blood and several tissues [16–22]. The presence of a variant T to deletion (T-del) in the promoter of the  $\Delta 6$ -desaturase gene (FADS2) leads to reduced timnodonic acid (EPA) concentrations in plasma and adipose tissue [18], suggesting that this variant decreases enzyme activity and therefore conversion from ALA. The presence of the FADS2 T-del variant is also associated with higher plasma triglyceride concentrations [18]. Additionally, a number of studies have reported significant associations between FADS genotypes and the risk of CAD [23, 24]. In CHIANTI study, the genome-wide association study (GWAS) showed that the strongest evidence for association was observed in a region of chromosome 11 that encodes three fatty acid desaturases (FADS1, FADS2, and FADS3) [25]. In 2010, another large GWAS repeated the strongest association between FADS1 and ELOVL2 genes and the ratio of product to precursor fatty acids [26]. Several studies, including a recent meta-analysis of genome-wide association (GWA) scans, confirmed that polymorphisms in the FADS gene cluster were associated with PUFA concentrations in serum phospholipids and erythrocyte cell membranes in several populations, including Caucasians, East Asians, and African Americans [19, 23, 25, 27-29].

Until now, it is unknown whether or not SNPs in the *FADS1/FADS2* gene cluster and *ELOVL2* gene are associated with ACS in Chinese Han population. The aim of this study was to investigate the possible association between the gene SNPs of PUFA metabolic rate-limiting enzymes and risk of ACS in Chinese Han population through the case-control study containing *FADS1/FADS2* and *ELOVL2* genes.

#### 2. Subjects and Methods

2.1. Study Subjects. This case-control study included 249 ACS patients and 240 controls in order to undertake a genetic analysis for association between the PUFA rate-limiting enzymes gene polymorphisms and ACS. All patients used for this study were Chinese of Han descent. Patients with ACS were admitted to the First Hospital of Jilin University, Changchun, China, in the period between 2008 and 2010. All subjects gave written informed consent for the study. The study was approved by the ethics committee of Jilin University, Changchun, Changchun, China.

Diagnosis was carried out independently by at least two well-trained physicians based on the following criteria. All patients (143 males and 106 females) were identified with ACS by coronary computed tomographic angiography (SIEMNS Somatom Definition AS + 128 row spiral CT). ACS was defined by  $\geq$ 75% stenosis in any major coronary artery. Acute coronary syndrome encompasses three clinical diagnoses: unstable angina, non-ST-segment elevation myocardial infarction, and ST-segment elevation myocardial infarction. Myocardial infarction with cardiac chest pain, serologic evidence of myonecrosis, and persistent (>20 min) ST-segment elevation was confirmed as WHO criteria issued in 1979 [30]; the definition of UAP is when cardiac chest pain was new or worsening without serologic evidence of myonecrosis (i.e., no elevation of serum troponin or creatine kinase MB isoenzyme concentration), or dynamic electrocardiographic (ECG) changes (i.e., ST depression and/or T wave inversion); the definition of NSTEMI is when the patient had cardiac chest pain and serologic evidence of myocardial necrosis in the absence of ST-segment elevation on the ECG [31], and patients with nonatherosclerotic vascular diseases, congenital heart disease, cardiomyopathy, valvular disease, renal or hepatic disease, and cancer were excluded. Based on the principle of epidemiology for setting control group [32], control group contained 240 healthy patients (126 males and 114 females) randomly selected from the same geographical area who were undergoing a routine checkup as part of annual physical examination, which included an ECG, chest X-ray, and serum analysis. The control group was formed on the basis of their unremarkable physical examination, as well as the absence of personal or family history and reasons to suspect ACS.

The presence of cardiovascular risk factors, including diabetes mellitus, blood pressure, and cigarette smoking, was obtained from all participants. Diagnosis of hypertension and diabetes mellitus was performed according to World Health Organization criteria. In this study, hypercholesterolemia was defined as a serum total cholesterol level of 200 mg/dL or more, and a smoking habit was defined as a daily intake of >10 cigarettes [4]. Overnight fasting venous blood samples were collected from all subjects for genomic DNA extraction.

2.2. Genotyping. Three SNPs rs174556 (Mbo I site) in the FADS1, rs174617 (Msp I site) in the FADS2, and rs3756963 (Hha I site) in the ELOVL2 were selected as genetic markers. They were all C to T base change present in intron. SNP information was obtained from NCBI dbSNP Build 132 (http://www.Ncbi.nlm.nih.gov/SNP/). The candidates SNPs were restricted to minor allele frequency bigger than 15% in HAPMAP-CHB database (http://www.hapmap.org/). Genomic DNA used for PCR amplification was extracted from the whole blood sample using a DNA extraction kit (Promega, Beijing, China). SNPs were genotyped using standard polymerase chain reaction and restriction fragment length polymorphism (RFLP) analysis. The sequences of primers for amplification are available as follows (Table 1). PCR conditions included predenaturation at 94°C for 5 minutes followed by 35 cycles of 95°C for 35-40 seconds, 63–57°C for 1 minutes, and 72°C for 1 minutes, and a final extension at 72°C for 10 minutes.

2.3. Statistical Analysis. Data were expressed as percentages of total categorical variables, or mean  $\pm$  SD. The statistical analyses on the characteristics of the subjects were performed with Pearson  $\chi^2$  test for the categorical variables such as sex, smokers, and nonnormal distribution variable age was compared by Mann-Whitney rank sum test.

TABLE 1: The sequences of primers for amplification.

Genes	SNPs	Primers	Sequences
FADS1	ro174556	Forward	5'AAGCAGGGACCTCAAGAC3'
	181/4550	Reverse	5'AGCCCACCAAGAATGTAA3'
FADS2	ma174617	Forward	5'GAACTGTCAGAGGCAACG3'
	181/401/	Reverse	5'CTGGGCAATAAAGCAAGA3'
ELOVL2	m2756062	Forward	5'CCCTTTGTGCGAGAACCT3'
	185/36965	Reverse	5'ATCCCAAGCGACAGACCC3'

TABLE 2: General characteristics of patients and controls included in our study.

Subject characteristics	Case	Control	P value	
Subject characteristics	(n = 249)	(n = 240)		
Age (years)	$62.00 \pm 17.00$	$63.00 \pm 23.70^{a}$	0.148	
Male, <i>n</i> (%)	143 (57.4)	126 (52.5)	0.273	
Diabetes, n (%)	86 (34.5)	30 (14.4)	< 0.001	
Hypertension, n (%)	154 (61.8)	96 (40.0)	< 0.001	
Smokers, n (%)	162 (65.1)	86 (35.8)	< 0.001	
a				

 $Median \pm QR.$ 

Age was compared by using Mann-Whitney rank sum test.

Male, diabetes, hypertension, and smokers were compared by using Pearson's Chi-square test.

The Hardy-Weinberg equilibrium for the genotypic distributions of SNPs was tested using the Chi-square  $(\chi^2)$ goodness-of-fit test. The Haploview program (version 4.1) was applied to estimate the linkage disequilibrium (LD) measures  $(D' \text{ and } r^2)$  between paired SNPs. Allelic association, genotypic association, and analysis for gene-gene interaction were performed with the UNPHASED program, which is an application for performing genetic association analysis in nuclear families and unrelated subjects [21]. Results are expressed as odds ratio (OR) and 95% confidence intervals (CI). P values < 0.05 were considered significant. With regard to the gene-gene interaction tests, the genotypes with relative frequencies of less than 3% were not considered for analysis. We also applied the permutation test (1000 times) performed with UNPHASED to correct the final P values for the markers used.

#### 3. Results

3.1. Characteristics of Study Subjects. Table 2 lists the demographic and clinical characteristics of the 249 ACS patients and 240 control subjects. Compared with control group, ACS group had more males, more smokers, and more individuals with diabetes. However, there was no significant difference of the mean age and proportion of hypertension between case and control groups.

3.2. Allele and Genotype Analysis. Rs174556 of FADS1, rs174617 of FADS2, and rs3756963 of ELOVL2 were genotyped, and they all lie within intron. Rs174556 and rs174617 locate in different LD block on 11q12-q13.1 region (D' = 0.57,  $r^2 = 0.52$ ). Rs3756963 locates on 6p24.2. The  $\chi^2$  3

goodness-of-fit test showed that the genotypic distributions of rs174556, rs174617, and rs3756963 were not deviated from Hardy-Weinberg equilibrium in both case and control groups (P > 0.05). Tables 3 and 4 present the distributions of alleles and genotypes of 3 SNPs among participants, respectively. For rs174556, C was major allele, frequency was 60.6% and 69.6% in case and control group, respectively, but for rs174617 and rs3756963, T was major allele.

Analysis with the UNPHASED software showed that rs174556 was associated with ACS before ( $\chi^2 = 8.592$ , P = 0.003) and after 1000 permutation test (P = 0.003996), and frequency of minor allele T of rs174556 was significantly higher in case than control subjects (Table 3). However, rs174617 and rs3756963 were not associated with ACS. As shown in Table 4, the logistic regression analysis test revealed genotypic association between rs174556 and ACS after being adjusted for confounding factors ( $\chi^2 = 6.084$ , P = 0.036), but not the genotypic association was observed between the other 2 SNPs and ACS.

We further analyzed the associations between the polymorphisms of three SNPs and ACS for subgroups with or without hypertension, DM, and smoking. The results suggest that, for rs174556 C>T, compared with the CC genotype, the CT/TT genotypes were more likely to result in ACS in subjects with hypertension after correction of all risk factors (OR = 4.236, 95% CI, 2.216–7.126) (Table 5). Whereas, another two SNPs, rs174556 and rs3756963, were not associated with ACS after correction of all risk factors (Table 6).

3.3. Trans-Phase Gene-Gene Interaction Analysis. In Table 7, the combined genotypes for rs174556 and rs3756963 with frequency more than 3% are presented. We used the most commonly combined genotype major homozygote as a reference, and the results showed that *ELOVL2* gene had a combined effect with *FADS1* gene ( $\chi^2 = 14.112$ , df = 6, *P* = 0.028). Rs174556 (C/C)-rs3756963 (T/T) and rs174556 (T/T)-rs3756963 (T/T) were associated with ACS ( $\chi^2 = 4.478$ , *P* = 0.034,  $\chi^2 = 4.656$ , *P* = 0.031).

*3.4. Logistic Regression Analysis.* As shown in Table 8, according to a multivariate logistic regression analysis, the most predictive risk factor for ACS was hypertension, followed by smoking, diabetes, and the T allele in rs174556. The T allele in rs174556 may be a risk factor for ACS (OR = 1.791, 95% CI, 1.088–2.951).

TABLE 3: Distribution of allele frequencies of SNPs in case and control groups.

SNPs	Allele	Case (%)	Control (%)	$\chi^2$	Р	OR	95% CI
rs174556	С	302 (60.6)	334 (69.6)	8 502	0.003 <sup>a</sup>	1 495	1 130 _ 1 035
181/4550	Т	196 (39.4)	96 (39.4) 146 (30.4)	0.392	0.005	1.405	1.139-1.933
rc174617	Т	386 (77.5)	380 (79.2)	0 305	0.520	1 102	0 813 1 405
FAD52 IS1/401/	С	112 (22.5)	100 (20.8)	0.395	0.550	1.105	0.813-1.495
ro3756063	Т	368 (73.9)	372 (77.5)	1 725	0 190	1 217	0.008 1.631
1357 50905	С	130 (26.1)	108 (22.5)	1.723	0.109	1.21/	0.908-1.031
	SNPs rs174556 rs174617 rs3756963	SNPs Allele   rs174556 C   T T   rs174617 C   rs3756963 T   C C	SNPs Allele Case (%)   rs174556 C 302 (60.6)   T 196 (39.4) 196 (39.4)   rs174617 T 386 (77.5)   C 112 (22.5) 112 (22.5)   rs3756963 T 368 (73.9)   C 130 (26.1) 130 (26.1)	SNPs Allele Case (%) Control (%)   rs174556 C 302 (60.6) 334 (69.6)   T 196 (39.4) 146 (30.4)   rs174617 T 386 (77.5) 380 (79.2)   C 112 (22.5) 100 (20.8)   rs3756963 T 368 (73.9) 372 (77.5)   C 130 (26.1) 108 (22.5)	$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	$ \begin{array}{c c c c c c c c c c c c c c c c c c c $

<sup>®</sup> The adjusted *P* value was 0.003996 from 1000 permutations.

TABLE 4: Distribution of genotype frequencies of SNPs in case and control groups.

Genes	SNPs	Genotype	Case (%)	Control (%)	$\chi^2$	Р	OR	95% CI
		C/C	96 (38.6)	119 (49.6)				
FADS1	rs174556	C/T	110 (44.2)	96 (40.0)	6.084	0.036 <sup>a</sup>	0.826 <sup>a</sup>	$0.418 - 1.459^{a}$
		T/T	43 (17.3)	25 (10.4)			0.324 <sup>a</sup>	$0.126 - 0.864^{a}$
FADS2 rs		T/T	148 (59.4)	149 (62.1)				
	rs174617	C/T	90 (36.1)	82 (34.2)	0.410	0.815	1.105	0.759-1.609
		C/C	11 (4.4)	9 (3.8)			1.230	0.495-3.056
		T/T	141 (56.6)	145 (60.4)				
ELOVL2	rs3756963	C/T	86 (34.5)	82 (34.2)	2.301	0.317	1.079	0.737-1.579
		C/C	22 (8.8)	13 (5.4)			1.740	0.844-3.589

<sup>a</sup> Adjustment for age, sex, and the presence of diabetes, hypertension, and smoking by forward logistic regression analysis.

TABLE 5: Stratified analysis between the rs174556 C>T polymorphisms and risk of ACS by hypertension, DM, and smoking.

	Case ( <i>n</i> = 249)		Control $(n = 240)$		Adjusted OR (95% CI) <sup>a</sup>	
	CC (%)	CT + TT (%)	CC (%)	CT + TT (%)	CC	CT + TT
		R	s174556 C>T genot	ypes		
Hypertension						
No	24 (25.3)	71 (74.7)	43 (29.9)	101 (70.1)	1.00	2.640 (1.410-4.560)
Yes	72 (46.8)	82 (53.2)	76 (79.2)	20 (20.8)	1.00	4.236 (2.216-7.126)
DM						
No	26 (16.0)	137 (84.0)	101 (48.1)	109 (51.9)	1.00	4.120 (2.326-7.824)
Yes	70 (81.4)	16 (18.6)	18 (60.0)	12 (40.0)	1.00	0.424 (0.182-0.926)
Smoking						
No	10 (11.5)	77 (88.5)	49 (31.8)	105 (68.2)	1.00	3.624 (1.842-7.636)
Yes	86 (53.1)	76 (46.9)	70 (81.4)	16 (18.6)	1.00	0.820 (0.326-1.726)
No 3 risk	10 (16.1)	52 (83.9)	36 (32.1)	76 (67.9)	1.00	2.016 (1.046-5.236)

Ajusted for age, sex, and the presence of diabetes, hypertension, and smoking status except the stratified factor at each stratum.

## 4. Discussion

*FADS1*, *FADS2*, and *ELOVL2* all encode rate-limiting enzymes in PUFA metabolism. And many studies have confirmed that high levels of PUFA in plasma phospholipids, cell membranes, or whole blood were associated with lower risk of multiple diseases, including metabolic syndrome, CAD, et al. Therefore, we investigate the association between the common variants of the three genes and ACS.

Our results show that the frequency of minor allele T of rs174556 was remarkably higher in case than control group (P = 0.003), as the frequency of TT genotype of rs174556 in case group was also markedly higher than in control group

(P = 0.036). And for rs174556 C>T, compared with the CC genotype, the CT/TT genotypes were more likely to lead to ACS in subjects with hypertension after correction of all factors. Rs174617 in the *FADS2* was not associated with ACS. Based on our results, therefore, rs174556 in the *FADS1* has a significant role in the development of ACS, especially in subjects with hypertension. But the locus rs174556 is not a functional SNP, which located in intron of *FADS1* gene. As a result, we infer that the *FADS1* gene may confer susceptibility of ACS through affecting nearby gene. Many studies also have proved that SNPs in *FADS1/FADS2* were associated with plasma lipid concentrations in adult populations and children [33, 34]. However, in contrast with our results, Aslibekyan

TABLE 6: Stratified analysis between the rs174617	and rs3756963 T>C poly	morphisms and risk of ACS by	hypertension, DM, and smoking.

	Case	(n = 249)	Contro	n = 240	Adjusted OR (95% CI) <sup>a</sup>	
	TT (%)	CC + CT (%)	TT (%)	CC + CT (%)	TT	CC + CT
		R	s174617 T>C genoty	ypes		
Hypertension						
No	51 (58.0)	37 (42.0)	109 (75.7)	35 (24.3)	1.00	2.326 (1.327-4.120)
Yes	90 (58.4)	64 (41.6)	40 (41.7)	56 (58.3)	1.00	0.492 (0.262-0.812)
DM						
No	88 (54.0)	75 (46.0)	135 (64.3)	75 (35.7)	1.00	1.242 (1.006-2.012)
Yes	60 (69.8)	26 (30.2)	14 (46.7)	16 (53.3)	1.00	0.326 (0.126-0.801)
Smoking						
No	58 (82.8)	29 (17.2)	113 (73.4)	41 (26.6)	1.00	0.524 (0.326-0.884)
Yes	90 (46.9)	72 (53.1)	36 (41.9)	50 (58.1)	1.00	0.628 (0.412-1.021)
No 3 risk	50 (71.4)	20 (28.6)	60 (65.2)	32 (34.8)	1.00	0.782 (0.382-1.410)
		Rs	3756963 T>C geno	types		
Hypertension						
No	67 (70.5)	28 (29.5)	115 (79.9)	29 (20.1)	1.00	1.542 (0.743-2.732)
Yes	74 (48.1)	80 (51.9)	30 (31.2)	66 (68.8)	1.00	0.524 (0.301-0.980)
DM						
No	89 (16.0)	74 (84.0)	125 (59.5)	85 (40.5)	1.00	1.364 (0.896-1.868)
Yes	52 (60.5)	34 (39.5)	20 (66.7)	10 (33.3)	1.00	0.692 (0.284-1.790)
Smoking						
No	59 (11.5)	28 (88.5)	105 (31.8)	49 (68.2)	1.00	0.423 (0.298-0.792)
Yes	82 (53.1)	80 (46.9)	40 (81.4)	46 (18.6)	1.00	0.920 (0.583-1.492)
No 3 risk	60 (85.7)	10 (14.3)	80 (74.1)	28 (25.9)	1.00	0.524 (0.301-1.162)

<sup>4</sup> Ajusted for age, sex, and the presence of diabetes, hypertension, and smoking status except the stratified factor at each stratum.

<b>— — — —</b>	1 1 .	C		1 • 1 0		1	. 1	
ADIE / Ibotrone	nhace analycu	e tor aona	stunic com	binod off	oct in	cace and	control	aroune
IADLE /. IIIC (Iallo-	Dilase allarysis	5 101 2010		ionica ch		case and	control	groups.
	I							0

Combined genotype <sup>a</sup>	Case	Control	$\chi^2$	Р	OR (95% CI)
rs174556 (C/C)-rs3756963 (T/T)	52	70	4.478	0.034	referent
rs174556 (C/C)-rs3756963 (C/T)	38	46	1.310	0.252	1.112 (0.635–1.946)
rs174556 (C/T)-rs3756963 (C/T)	30	34	0.482	0.487	1.188 (0.806–2.891)
rs174556 (C/T)-rs3756963 (T/T)	70	66	0.023	0.880	1.428 (0.873-2.335)
rs174556 (T/T)-rs3756963 (T/T)	21	9	4.656	0.031	3.141 (1.330-7.418)

Test of overall association:  $\chi^2 = 14.112$ , df = 6, P = 0.028.

<sup>a</sup>Only listed the combined genotypes with frequencies >3%.

TABLE 8: The logistic regression analysis for the relation between the risk factors and ACS.

Risk	Р	OR value	95% CI
Hypertension	0.000	4.114	2.224-7.611
Smoking	0.000	2.444	1.563-3.823
Diabetes	0.035	2.243	1.052-4.798
rs174556 T allele	0.022	1.791	1.088-2.951

Smoking versus nonsmoking, hypertension versus nonhypertension, DM versus non-DM, rs174556 T allele versus rs174556 C allele.

et al. [35] failed to show relationship between rs174556 and MI in the Costa Rica Study. Firstly, this discrepancy may be due to racial differences in two studies. Secondly, the research

objects are different. Finally, the difference of sample size also may be another possible reason.

*FADS2* is located in chromosome 11 and expressed in almost all human tissues, especially in the liver, heart, and brain [36]. Some studies have report the metabolic effects of polymorphisms in this gene or their effects on the risk of CAD [24]. But the locus rs174617 of *FADS2* is rarely reported. In 2011, our previous study showed that this locus was not associated with CAD [37], which is consistent with our present study. The reason may be that a large proportion of regulatory SNPs, which affect gene expression, are located in the promoter regions [38].

At present, rs174537 near *FADS1* is considered to be the most relevant locus with ARA. GWAS in the In CHIANTI Study showed that minor allele homozygotes rs174537 (TT)

had lower plasma concentrations of ARA compared to major allele homozygotes [25]. And a study in Korea population confirmed that rs174537 T had lower proportions of ARA in serum phospholipids and reduced CAD risk [24]. Many studies suggested the minor allele carriers including rs174556 T may have lower desaturase activity [25, 37]. Therefore, based on above results, a possible causality link between lower desaturase activity and vascular disease has been suggested [39]. However, at present, the further research of gene function is still lacking.

ELOVL2 is a member of the mammalian microsomal ELOVL fatty acid enzyme family, involved in the elongation of very long-chain fatty acids including PUFAs required for various cellular functions in mammals [40]. A GWAS study found that an association of EPA with variants in the FADS1 gene reached genome-wide significance level, and independent follow-up investigation showed associations of a selected FADS1 variant with erythrocyte membrane levels of EPA, ALA, and DPA and of an ELOVL2 variant with DPA and DHA [25]. These findings confirm an influence of FADS1 and ELOVL2 on selected n-3 PUFAs. In contrast, we do not found the association between rs3756593 of ELOVL2 gene and ACS. Both allele and genotype analysis all have no statistical significance, but the trans-phase gene-gene interaction test revealed that the ELOVL2 gene combined with FADS1 gene had an effect on ACS. Above result implies that the FADS1 gene-ELOVL2 gene interaction may affect PUFA concentrations and afford susceptibility to ACS. But the exact mechanisms for the interaction are currently unknown.

In conclusion, this case-control study preliminary indicates that variations in *FADS1* may affect the risk of ACS and provide a genetic basis of molecular biology. We will continue to analyze the gene functional and the serum levels of phospholipids fatty acid in ACS.

## **Conflict of Interests**

All authors have no conflict of interests.

### **Authors' Contribution**

L. Qin and Y.-F. Jiang participated in the design and conduct of the study, data collection and analysis, data interpretation, and paper writing. Z.-K. Song and H.-Y. Cao participated in data collection and analysis. All authors read and approved the final paper. Z. Song contributed to this work.

#### Acknowledgments

The authors thank the participants for their support and participation. This study was sponsored by the Central Lab, the Second Division of the First hospital, Jilin University, Jilin Province, China. They gratefully acknowledge Dr. Lin Xie who helps in designing the primers for the PCR processing.

## References

 C. T. Ruff and E. Braunwald, "The evolving epidemiology of acute coronary syndromes," *Nature Reviews Cardiology*, vol. 8, no. 3, pp. 140–147, 2011.

- [2] S. Yusuf, S. Reddy, S. Ôunpuu, and S. Anand, "Global burden of cardiovascular diseases: part I: general considerations, the epidemiologic transition, risk factors, and impact of urbanization," *Circulation*, vol. 104, no. 22, pp. 2746–2753, 2001.
- [3] A. D. Lopez, C. D. Mathers, M. Ezzati, D. T. Jamison, and C. J. Murray, "Global and regional burden of disease and risk factors, 2001: systematic analysis of population health data," *The Lancet*, vol. 367, no. 9524, pp. 1747–1757, 2006.
- [4] Y. H. Chen, J. M. Liu, R. J. Hsu et al., "Angiotensin converting enzyme DD genotype is associated with acute coronary syndrome severity and sudden cardiac death in Taiwan: a casecontrol emergency room study," *BMC Cardiovascular Disorders*, vol. 12, p. 6, 2012.
- [5] C. Glaser, E. Lattka, P. Rzehak, C. Steer, and B. Koletzko, "Genetic variation in polyunsaturated fatty acid metabolism and its potential relevance for human development and health," *Maternal & Child Nutrition*, vol. 7, no. supplement 2, pp. 27–40, 2011.
- [6] R. Wall, R. P. Ross, G. F. Fitzgerald, and C. Stanton, "Fatty acids from fish: the anti-inflammatory potential of long-chain ω-3 fatty acids," *Nutrition Reviews*, vol. 68, no. 5, pp. 280–289, 2010.
- [7] C. M. Albert, K. Oh, W. Whang et al., "Dietary α-linolenic acid intake and risk of sudden cardiac death and coronary heart disease," *Circulation*, vol. 112, no. 21, pp. 3232–3238, 2005.
- [8] Q. Sun, J. Ma, H. Campos et al., "A prospective study of trans fatty acids in erythrocytes and risk of coronary heart disease," *Circulation*, vol. 115, no. 14, pp. 1858–1865, 2007.
- [9] S. Czernichow, D. Thomas, and E. Bruckert, "N-6 fatty acids and cardiovascular health: a review of the evidence for dietary intake recommendations," *British Journal of Nutrition*, vol. 104, no. 6, pp. 788–796, 2010.
- [10] M. Geiger, B. S. Mohammed, S. Sankarappa, and H. Sprecher, "Studies to determine if rat liver contains chain-length-specific acyl-CoA 6-desaturases," *Biochimica et Biophysica Acta*, vol. 1170, no. 2, pp. 137–142, 1993.
- H. Sprecher, "Metabolism of highly unsaturated *n*-3 and *n*-6 fatty acids," *Biochimica et Biophysica Acta*, vol. 1486, no. 2-3, pp. 219–231, 2000.
- [12] M. T. Nakamura and T. Y. Nara, "Structure, function, and dietary regulation of Δ6, Δ5, and Δ9 desaturases," *Annual Review of Nutrition*, vol. 24, pp. 345–376, 2004.
- [13] A. Marquardt, H. Stöhr, K. White, and B. H. F. Weber, "cDNA cloning, genomic structure, and chromosomal localization of three members of the human fatty acid desaturase family," *Genomics*, vol. 66, no. 2, pp. 175–183, 2000.
- [14] E. Lattka, S. Eggers, G. Moeller et al., "A common FADS2 promoter polymorphism increases promoter activity and facilitates binding of transcription factor ELK1," *Journal of Lipid Research*, vol. 51, no. 1, pp. 182–191, 2010.
- [15] E. A. Emken, R. O. Adlof, and R. M. Gulley, "Dietary linoleic acid influences desaturation and acylation of deuterium-labeled linoleic and linolenic acids in young adult males," *Biochimica et Biophysica Acta*, vol. 1213, no. 3, pp. 277–288, 1994.
- [16] L. Schaeffer, H. Gohlke, M. Müller et al., "Common genetic variants of the FADS1 FADS2 gene cluster and their reconstructed haplotypes are associated with the fatty acid composition in phospholipids," *Human Molecular Genetics*, vol. 15, no. 11, pp. 1745–1756, 2006.
- [17] L. Xie and S. M. Innis, "Genetic variants of the FADS1 FADS2 gene cluster are associated with altered (n-6) and (n-3) essential fatty acids in plasma and erythrocyte phospholipids in women

during pregnancy and in breast milk during lactation," *Journal of Nutrition*, vol. 138, no. 11, pp. 2222–2228, 2008.

- [18] A. Baylin, E. Ruiz-Narvaez, P. Kraft, and H. Campos, "αlinolenic acid, Δ6-desaturase gene polymorphism, and the risk of nonfatal myocardial infarction," *American Journal of Clinical Nutrition*, vol. 85, no. 2, pp. 554–560, 2007.
- [19] G. Malerba, L. Schaeffer, L. Xumerle et al., "SNPs of the FADS gene cluster are associated with polyunsaturated fatty acids in a cohort of patients with cardiovascular disease," *Lipids*, vol. 43, no. 4, pp. 289–299, 2008.
- [20] E. Lattka, P. Rzehak, É. Szabó et al., "Genetic variants in the FADS gene cluster are associated with arachidonic acid concentrations of human breast milk at 1.5 and 6 mo postpartum and influence the course of milk dodecanoic, tetracosenoic, and *trans*-9-octadecenoic acid concentrations over the duration of lactation," American Journal of Clinical Nutrition, vol. 93, no. 2, pp. 382–391, 2011.
- [21] C. Moltó-Puigmartí, J. Plat, R. P. Mensink et al., "FADS1 FADS2 gene variants modify the association between fish intake and the docosahexaenoic acid proportions in human milk," American Journal of Clinical Nutrition, vol. 91, no. 5, pp. 1368–1376, 2010.
- [22] B. Koletzko, E. Lattka, S. Zeilinger, T. Illig, and C. Steer, "Genetic variants of the fatty acid desaturase gene cluster predict amounts of red blood cell docosahexaenoic and other polyunsaturated fatty acids in pregnant women: findings from the avon longitudinal study of parents and children," *American Journal of Clinical Nutrition*, vol. 93, no. 1, pp. 211–219, 2011.
- [23] N. Martinelli, D. Girelli, G. Malerba et al., "FADS genotypes and desaturase activity estimated by the ratio of arachidonic acid to linoleic acid are associated with inflammation and coronary artery disease," American Journal of Clinical Nutrition, vol. 88, no. 4, pp. 941–949, 2008.
- [24] J. H. Kwak, J. K. Paik, O. Y. Kim et al., "FADS gene polymorphisms in Koreans: association with ω6 polyunsaturated fatty acids in serum phospholipids, lipid peroxides, and coronary artery disease," Atherosclerosis, vol. 214, no. 1, pp. 94–100, 2011.
- [25] T. Tanaka, J. Shen, G. R. Abecasis et al., "Genome-wide association study of plasma polyunsaturated fatty acids in the InCHI-ANTI study," *PLoS Genetics*, vol. 5, no. 1, Article ID e1000338, 2009.
- [26] T. Illig, C. Gieger, G. Zhai et al., "A genome-wide perspective of genetic variation in human metabolism," *Nature Genetics*, vol. 42, no. 2, pp. 137–141, 2010.
- [27] P. Rzehak, J. Heinrich, N. Klopp et al., "Evidence for an association between genetic variants of the *fatty acid desaturase* 1 *fatty acid desaturase* 2 (FADS1 FADS2) gene cluster and the fatty acid composition of erythrocyte membranes," British Journal of Nutrition, vol. 101, no. 1, pp. 20–26, 2009.
- [28] R. N. Lemaitre, T. Tanaka, W. Tang et al., "Genetic loci associated with plasma phospholipid N-3 fatty acids: a meta-analysis of genome-wide association studies from the charge consortium," *PLoS Genetics*, vol. 7, no. 7, Article ID e1002193, 2011.
- [29] R. A. Mathias, S. Sergeant, I. Ruczinski et al., "The impact of *FADS* genetic variants on ω6 polyunsaturated fatty acid metabolism in African Americans," *BMC Genetics*, vol. 12, article 50, 2011.
- [30] "Nomenclature and criteria for diagnosis of ischemic heart disease. Report of the Joint International Society and Federation of Cardiology/World Health Organization Task Force on standardization of clinical nomenclature," *Circulation*, vol. 59, no. 3, pp. 607–609, 1979.

- [31] J. C. Trost and R. A. Lange, "Treatment of acute coronary syndrome: part 1: non-ST-segment acute coronary syndrome," *Critical Care Medicine*, vol. 39, no. 10, pp. 2346–2353, 2011.
- [32] Clinical Epidemeoligy, 3rd edition, 2002.
- [33] L. Schaeffer, H. Gohlke, M. Müller et al., "Common genetic variants of the FADS1 FADS2 gene cluster and their reconstructed haplotypes are associated with the fatty acid composition in phospholipids," *Human Molecular Genetics*, vol. 15, no. 11, pp. 1745–1756, 2006.
- [34] M. Standl, E. Lattka, B. Stach et al., "FADS1 FADS2 gene cluster, PUFA intake and blood lipids in children: results from the GINIplus and LISAplus studies," PLoS ONE, vol. 7, no. 5, Article ID e37780, 2012.
- [35] S. Aslibekyan, M. K. Jensen, H. Campos et al., "Fatty Acid desaturase gene variants, cardiovascular risk factors, and myocardial infarction in the costa rica study," *Front Genet*, vol. 3, p. 72, 2012.
- [36] H. P. Cho, M. T. Nakamura, and S. D. Clarke, "Cloning, expression, and nutritional regulation of the mammalian Δ-6 desaturase," *Journal of Biological Chemistry*, vol. 274, no. 1, pp. 471– 477, 1999.
- [37] L. Qin, L. Sun, L. Ye et al., "A case-control study between the gene polymorphisms of polyunsaturated fatty acids metabolic rate-limiting enzymes and coronary artery disease in Chinese Han population," *Prostaglandins, Leukotrienes and Essential Fatty Acids*, vol. 85, no. 6, pp. 329–333, 2011.
- [38] P. R. Buckland, "The importance and identification of regulatory polymorphisms and their mechanisms of action," *Biochimica et Biophysica Acta*, vol. 1762, no. 1, pp. 17–28, 2006.
- [39] U. N. Das, "A defect in the activity of Δ6 and Δ5 desaturases may be a factor in the initiation and progression of atherosclerosis," *Prostaglandins, Leukotrienes and Essential Fatty Acids*, vol. 76, no. 5, pp. 251–268, 2007.
- [40] D. Zadravec, P. Tvrdik, H. Guillou et al., "ELOVL2 controls the level of n-6 28:5 and 30:5 fatty acids in testis, a prerequisite for male fertility and sperm maturation in mice," *Journal of Lipid Research*, vol. 52, no. 2, pp. 245–255, 2011.