

REVIEW OPEN ACCESS

The Critical Role of Inhibitor of Differentiation 4 in Breast Cancer: From Mammary Gland Development to Tumor Progression

Yuhang Song¹  | Panshi Zhang² | Sudhanshu Bhushan³ | Xinhong Wu¹ | Hongmei Zheng¹ | Yalong Yang¹

¹Department of Breast Surgery, Hubei Cancer Hospital, Tongji Medical College, Huazhong University of Science and Technology, Hubei Provincial Clinical Research Center for Breast Cancer, Wuhan Clinical Research Center for Breast Cancer; National key clinical specialty construction discipline, Wuhan, Hubei, China | ²Department of Thyroid and Breast Surgery, Tongji Hospital of Huazhong University of Science and Technology, Wuhan, Hubei, China | ³Department of Anatomy and Cell Biology, Unit of Reproductive Biology, Justus-Liebig-University Giessen, Giessen, Germany

Correspondence: Hongmei Zheng (zhenghongmeicj@sina.com) | Yalong Yang (yalong.yang@whu.edu.cn)

Received: 13 November 2024 | **Revised:** 12 March 2025 | **Accepted:** 26 March 2025

Funding: Supported by grants from Hubei Province Medical Youth Top Talent Project (EWT[2023] 65, Oncology), Hubei Cancer Hospital Biomedical Center Research Project (Grant Nos. 2022SWZX03, 2022SWZX09), Hubei Province Natural Science Foundation of Innovation and development joint fund projects (Grant No. 2024AFD454), Talent Project of Hubei Cancer Hospital (2025HBCHLHRC002, 2025HBCHHHRC005, 2025HBCHQHRC018) and Hubei Chen Xiaoping Science and Technology Development Foundation (Grant No. CXPJH123001-2320).

Keywords: breast cancer | development | ID4 | mammary gland

ABSTRACT

Inhibitor of differentiation 4 (ID4) is a highly conserved DNA-binding inhibitory protein of mammals, and its main role is to bind basic helix–loop–helix (b-HLH) so that it loses its DNA-binding activity, which in turn regulates the transcription of key genes, regulating cell differentiation and proliferation as the physiological function. Breast tissue is a highly heterogeneous tissue organ with a strong capacity for remodeling and differentiation, and studies of breast carcinogenesis suggest that the mechanisms regulating the differentiation of breast tissue interact critically with tumorigenesis. The expression level of ID4 and its regulatory mechanism play a crucial role in the study of breast cancer, but its oncogenic or oncostatic role has not yet been unanimously identified, and its regulatory mechanism in breast cancer still needs to be further elucidated. This review summarizes and analyzes the relevant studies of ID4 and the research progress in breast cancer, integrating the development of breast tissue and tumorigenesis with the regulatory role of ID4, to provide some insights into develop new treatment strategies and diagnostic biomarkers.

1 | Introduction

ID4 (inhibitor of differentiation 4, also known as an inhibitor of DNA-binding 4) is a transcriptional regulator that plays a critical role in various cellular processes, including cell differentiation, proliferation, and apoptosis [1]. ID4 was found to be

mostly expressed in cells with stem properties or low degrees of differentiation, especially in embryonic tissues, which decreases with the differentiation of organ tissues [2–4]. In many disease models, including tumors, researchers have found that the expression level of ID4 is closely associated with tumor incidence, prognosis, and other important indicators, especially in prostate

Abbreviations: BC, breast cancer; BLBC, basal-like breast cancer; CBF1, C-promoter binding factor-1; CSC, cancer stem cell; DFS, disease-free survival; ER, estrogen receptor; FOXA1, Forkhead Box Protein A1; ID4, inhibitor of differentiation 4; KO, knock out; MDC1, mediator of DNA damage checkpoint protein 1; OS, overall survival; PR, progesterone receptor; TEBS, terminal end buds; TNBC, triple-negative breast cancer; WT, wild-type.

The first two authors contributed equally to this article.

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2025 The Author(s). *Cancer Medicine* published by John Wiley & Sons Ltd.

cancer, where *ID4* is considered to be an independent and crucial oncogene [4–8].

Breast cancer (BC) has surpassed lung cancer to become the most prevalent malignant tumor affecting human health [9]. With advances in molecular phenotyping, a better understanding of the regulatory mechanisms of the immune microenvironment, and comprehensive analyses of breast cancer genomics, researchers have gradually uncovered key aspects of breast cancer development [10–13]. However, the key mechanisms of BC are still to be elucidated, especially in triple-negative breast cancer (TNBC), and accumulating data indicate that the development of BC is, to a certain extent, related to the dysregulation of stem cells in the mammary gland [14–19].

This review provides insights into the research progress of *ID4* in mammary gland development and breast carcinogenesis from the perspective of development and further discusses the potential of *ID4* as a target for intervention of key factors in BC pathogenesis.

2 | Basic Physiological Functions of *ID4*

*ID4*s induce cell proliferation, and *ID1* antagonizes the action of E proteins, whereas *ID2* and *ID3* act by binding to pRB (in retinoblastoma) and related proteins, p107 and p130 [20–22]. These

three ID proteins have been described in other reviews [23–25] and will not be listed here. *ID4* is a transcriptional regulator that belongs to the helix–loop–helix (HLH) family, characterized by a typical HLH domain comprising two α -helices connected by a loop. The two helices generally twist in the same direction and form a specific spatial configuration. The basic-HLH (b-HLH) transcription factors contain two highly conserved and functionally distinct structural domains: the basic region and the HLH region [2]. The basic region consists of 10–20 amino acids and is located at the N-terminal end, which serves as a DNA-binding region that recognizes the E-box and the G-box. The HLH region located at the C-terminal end relies on the interactions of hydrophobic amino acids to form a homo- or heterodimer of two HLH proteins, which then regulate the expression of downstream target genes [2, 26]. *ID4* does not contain the basic DNA-binding domain and inactivates the transcriptional activity of b-HLH proteins by forming inactive heterodimeric complexes with their b-HLH partners [4, 27]. This property has led them to be called DNA-binding inhibitors (shown in Figure 1A).

This review focuses primarily on *ID4*, not only because *ID4* is regulated in a very different manner from the other three IDs, but also because the expression of *ID4* shows a close correlation with diseases and also displays a unique negative regulation on *ID1/2/3* [28, 29]. Unlike *ID1/2/3*, which have overlapping expression in embryonic tissues, *ID4* is predominantly expressed in the mesodermal part of the embryo [28]. At a particular time point in

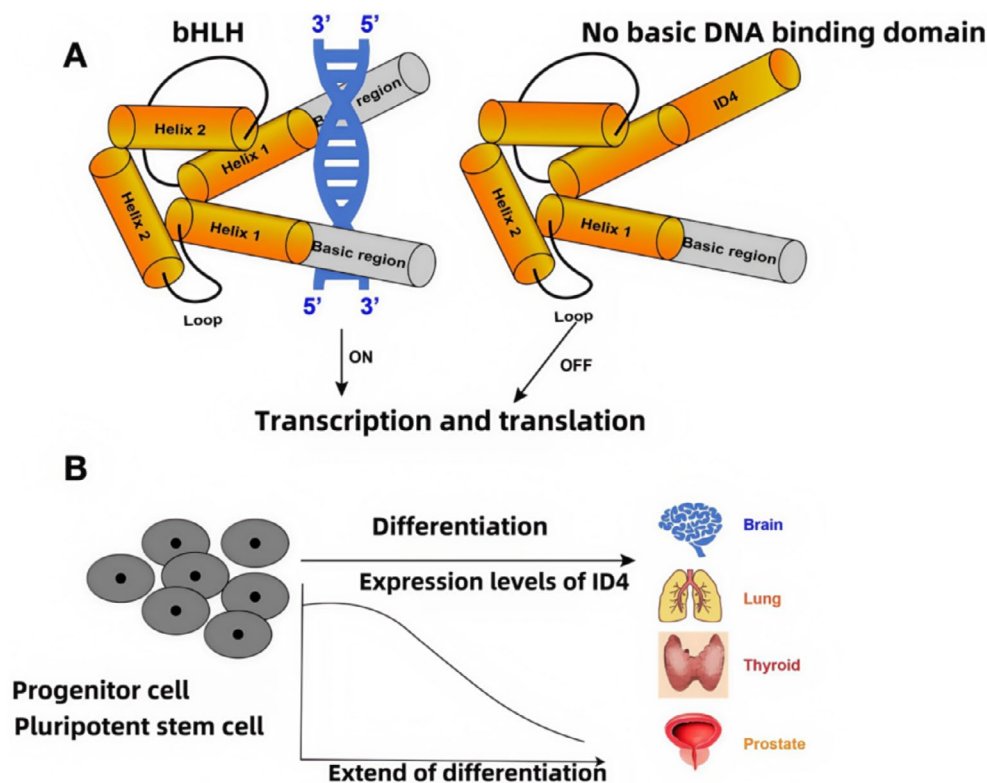


FIGURE 1 | (A) The structure and basic physiological functions of b-HLH and *ID4*. The left part of the figure shows the structure of b-HLH, which consists of the basic region and the HLH region. The basic region serves as a DNA-binding region that recognizes the E-box and the G-box. The right part of the figure shows that *ID4* is characterized by a typical HLH domain comprising two α -helices connected by a loop. *ID4* does not contain the basic DNA binding domain and inactivates the transcriptional activity of b-HLH proteins by forming inactive heterodimeric complexes with their b-HLH partners. (B) The expression level of *ID4* gradually decreases as the differentiation of embryonic cells increases. The expression level of *ID4* gradually decreases as the differentiation of embryonic cells increases. Created in <https://BioRender.com>.

embryonic development, ID proteins are expressed in all cell lines, and the expression level of ID4 gradually decreases as the differentiation of embryonic cells increases [23, 24, 28]. In human fetal tissues, ID4 is highly expressed in the brain, lung, and kidney, but not in the liver [4, 28]. In adult tissues, it is expressed predominantly in the thyroid gland and, to a lesser extent, but also highly expressed in the brain and testis [28]. Studies indicated that ID4 is highly expressed in osteoblasts, prostate epithelial cells, neurons in the central nervous system, sertoli cells in the testis, and glial cells in the brain, supporting its role as a pro-differentiation factor [20, 28, 30] (shown in Figure 1B).

3 | The Roles of ID4 in Normal Tissues

By developing a mouse global *Id4*-KO (knock out) model, the researchers found at least 50% embryonic lethality, significant weight loss in born purebred mice, and less than 20% survived to adulthood, and all surviving mice developed adipose dysplasia and osteoporosis [31, 32]. Depletion of *Id4* also induced slower proliferation of neural precursor cells than wild-type (WT) counterparts, and spermatogenesis was significantly impaired in adulthood [30, 32]. Mice with *Id4*-KO exhibited obvious osteoporosis, as evidenced by a significant reduction in osteoblast differentiation, the main mechanism being that normal *Id4* promotes osteoblast differentiation by releasing *Hes1* through the promotion of the *Hes1*-*Hey2* complex [33] (Table 1). In addition to adjunctive validation of the physiological role of *Id4* in promoting tissue differentiation through gene knockdown, researchers have also found, through functional experiments, that *Id4* inhibits oligodendrocyte differentiation by inhibiting the transcriptional activity of *Olig1/2* through binding to b-HLH [34]. An in vivo test also showed that one of the mechanisms by which *Id4* may regulate normal prostate development is through regulating androgen receptor binding to respective response elements such as those on NKX3.1 promoter [35]. *Nkx3.1* regulates early postnatal ductal morphogenesis and maintains normal differentiation of the prostate epithelium, including the production of secretory proteins [39, 40]. Taken together with the accumulating evidence described above, we can see that *Id4* has important regulatory roles in tissue and cellular differentiation.

4 | The Roles of ID4 in Cancers

The *Id4*-KO mouse model is theoretically capable of successfully inducing a wide range of tumor models based on the indicative role of the results of a large number of retrospective studies; however, to date, only prostate carcinogenesis has been confirmed in the *Id4*-KO mouse model [31]. The perspective of *Id4*'s tumor suppressor activity is based on evidence that *Id4* undergoes epigenetic gene silencing expression in mouse and human tumors including leukemia, rectal and gastric cancers [41, 42]. *P53* mutations are seen in about half of all tumors, and a large body of data suggests that about one-third of prostate carcinogenesis is associated with *p53* mutations and aberrant transcriptional activity, whereas *Id4* has been found to have a significant role in regulating *p53* transcriptional activity [37] (Table 1). In vitro assays with the WT-prostate cancer cell line (LNCaP) and the *p53*-mutant-cell line (DU145) showed that *Id4* significantly modulated *p53* transcriptional activity by inducing acetylation at the K373 position, thereby activating its downstream transcriptional activity [37].

However, some studies have noted that ID4 exhibited aberrantly activated pro-oncogenic effects in bladder cancer and B-cell acute lymphoblastic leukemia [36, 38, 43, 44]. Aberrant expression of ID4 blocks the cell cycle and proliferation, which correlates with increased expression of the cell cycle protein-dependent kinase inhibitors p21 and p27 [36]. According to the above studies on the role of ID4 in various disease models, it can be seen that the process of cellular tissue differentiation is regulated by the expression level of ID4 and, remarkably, bone dysplasia and central nervous system developmental abnormalities are closely related to the abnormal expression of ID4. Meanwhile, studies on various tumor models have pointed to the tumor-regulating role of ID4. Although some studies have not reached a consensus on whether *ID4* acts as an oncogene or suppressor gene, the relevance of some tumors, such as prostate cancer and leukemia, to the regulatory mechanism of ID4 has been supported by a large amount of data and interventions to modulate the expression of ID4 have been applied in clinical experiments. The mechanism of ID4 functions in other diseases needs to be further elucidated with the application of genomics,

TABLE 1 | The different roles of ID4 in normal tissues and cancer cells.

Normal tissues or cancer cells	Role	Mechanism	References
Bone	Promotes osteoblast differentiation	Releasing <i>Hes1</i> through the promotion of the <i>Hes1</i> - <i>Hey2</i> complex	[33]
Nervous system	Inhibits oligodendrocyte differentiation	Inhibiting the transcriptional activity of <i>Olig1/2</i> by binding to b-HLH	[34]
Prostate	Regulates normal prostate development	Regulating androgen receptor binding to response elements on the NKX3.1 promoter	[35]
Gastric cancer	Tumor suppressor	Aberrant methylation of the ID4 promoter	[36]
Prostate cancer	Tumor suppressor	Modulating <i>p53</i> transcriptional activity	[37]
B-cell acute lymphoblastic leukemia	Oncogenic effects	Blocking the cell cycle and proliferation	[36]
Bladder cancer	Oncogenic effects	Blocking the cell cycle and proliferation	[38]

transcriptomics, and metabolomics. As shown in Figure 2, we analyzed the expression of ID4 in different cancers and normal tissues using the TCGA database. We found that the expression of ID4 was significantly lower in breast cancer than in normal tissues.

5 | ID4 Regulates Mammary Gland Development

The role of the Id proteins has been studied to a limited extent in mammary gland development. Id1 is unnecessary for mammary gland development [45], whereas Id2 is necessary for normal RANK signaling within the mammary gland [46]. Id4 may play a more significant role in mammary gland development compared to other Id proteins. By observing the global *Id4*-KO mouse model, researchers found that although the phenotype of abnormal mammary tissue development in mice can be observed, the effect of Id4 on mammary tissue in developing mice is difficult to observe dynamically in the knockout model due to its high mortality rate [31]. However, by histological observation of mammary structures in surviving 6-week-old mice, researchers found that ductal branching in the *Id4*-null mice was significantly reduced until approximately 25 weeks old to fill the fat pad with an obvious reduction in branching, whereas there was no significant difference in mammary development between *Id4*-null heterogeneous and WT mice, suggesting that a single copy of the *Id4* gene is sufficient to maintain mammary development in mice [47]. Researchers also found that Id4 was mainly expressed in the cap cells (specialized cells located at the tip of terminal end buds in the developing mammary gland) and basal cells of the mammary gland, and Id4 maintains mammary cell survival by inhibiting p38MAPK [47]. The mammary gland developmental abnormalities in *Id4*-null mice were most notable

for the abnormalities in the branching of the milk ducts as well as the number and structure of terminal end buds (TEBs): disorganization of the capsule cell layer, separation from the somatic cells, and abnormalities in the number and distribution of stroma, collagens, smooth muscle proteins, and keratins [47]. Although the mammary tissue development of *Id4*-null mice was slowed down with reduced branching and incomplete degeneration of TEBs, the mammary gland had no impairment of functional differentiation, which manifested as normal lactation [47]. These early findings using *Id4*-KO mice to study mammary gland development suggested that Id4 has a critical regulatory role in the development of mammary glands.

Several studies have further found that the *Id4*-KO mice may compensatorily up-regulate the expression levels of FOXA1 (Forkhead Box Protein A1) and estrogen receptor (ER)/progesterone receptor (PR), showing that the *Id4*-KO mice's mammary gland showed significantly higher expression of ER/PR and FOXA1 in basal and luminal cells [48]. However, in the same model, the researchers found that the ovaries showed a reduction in granulosa cells and significant estrogen synthesis deficits, and both ovarian dysfunction and reduced estrogen could significantly affect the development of mammary tissue and the expression of ER/PR in mice [48]. This hypothesis was indirectly explained by Dong et al., who found that mammary cells from *Id4*-null mice exhibited normal responses to estrogen and progesterone stimulation, including cell proliferation capacity [47]. According to the morphology of breast development, intraglandular lumen formation is a characteristic structural alteration of breast development, whereas degeneration or absence of lumen formation occurs in breast carcinogenesis, and researchers found that overexpression of ID4 can replace the CEACAM1 protein to repromote the formation of lumen in BC cells, and

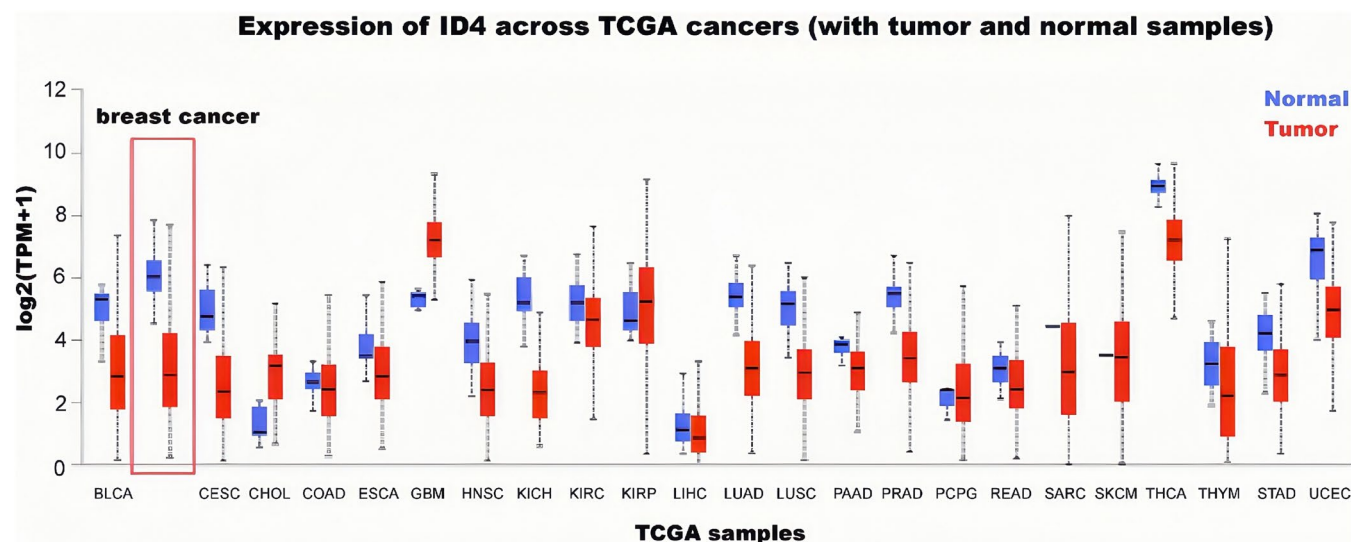


FIGURE 2 | Expression of ID4 across TCGA cancers (with tumor and normal samples). The red rectangular box shows that the expression of ID4 was significantly lower in breast cancer than in normal tissues. BLCA, Bladder Urothelial Carcinoma; CESC, Cervical Squamous Cell Carcinoma and Endocervical Adenocarcinoma; CHOL, Cholangiocarcinoma; COAD, Colon Adenocarcinoma; ESCA, Esophageal Carcinoma; GBM, Glioblastoma Multiforme; HNSC, Head and Neck Squamous Cell Carcinoma; KICH, Kidney Chromophobe; KIRC, Kidney Renal Clear Cell Carcinoma; KIRP, Kidney Renal Papillary Cell Carcinoma; LIHC, Liver Hepatocellular Carcinoma; LUAD, Lung Adenocarcinoma; LUSC, Lung Squamous Cell Carcinoma; PAAD, Pancreatic Adenocarcinoma; PRAD, Prostate Adenocarcinoma; PCPG, Pheochromocytoma and Paraganglioma; READ, Rectum Adenocarcinoma; SARC, Sarcoma; SKCM, Skin Cutaneous Melanoma; THCA, Thyroid Carcinoma; THYM, Thymoma; STAD, Stomach Adenocarcinoma; UCEC, Uterine Corpus Endometrial Carcinoma.

this evidence suggests that the role of ID4 in breast tissue has a correlative connection in mammary gland development and breast carcinogenesis [49] (shown in Figure 3).

Based on the above findings from a large number of mouse models of mammary development, it can be consistently concluded that Id4 expression is indispensable in mammary differentiation and development and primarily regulates the proliferation and differentiation of mammary basal cells and luminal epithelium, which are critical links in breast carcinogenesis.

6 | ID4 Regulates Breast Carcinogenesis

The mammalian mammary gland develops not only during the embryonic period, but also during postnatal with significant differentiation, development, and remodeling. Many studies have demonstrated that multiple cross-cutting regulatory mechanisms regulate breast carcinogenesis and mammary development, such as the NOTCH and Wnt signaling pathways [17, 18]. The review indicated that the aberrant expression of several

signaling pathways that regulate mammary development, including the Notch signaling pathway, can cause mammary developmental disorders and BC, including silencing and over-activation [17]. Of note, the regulation of the NOTCH pathway was found to interact with ID4 expression with the underlying mechanism to be uncovered [50]. One of the confirmed mechanisms [51] might be that ID4 contributes to breast carcinogenesis and chemotherapy resistance via the CBF1 (C-promoter binding factor-1, a critical downstream transcriptional factor in NOTCH1 signal pathway) and MRP1 (a chemo-resistance related protein) signaling axis (shown in Figure 3). Early in 2009, researchers noted that while molecular phenotyping was useful in predicting the prognosis of BC subtypes and appropriate therapeutic strategies, detecting the origin of the tumor cells within the BC and targeting the developmental stage of these “cells of origin” was more instructive, based on the discovery of the biology of stem and progenitor cells within the mammary tissue [52]. Therefore, exploring key genes or proteins with critical regulatory roles under different models of both mammary gland development and breast carcinogenesis becomes a potentially effective means of intervening in BC.

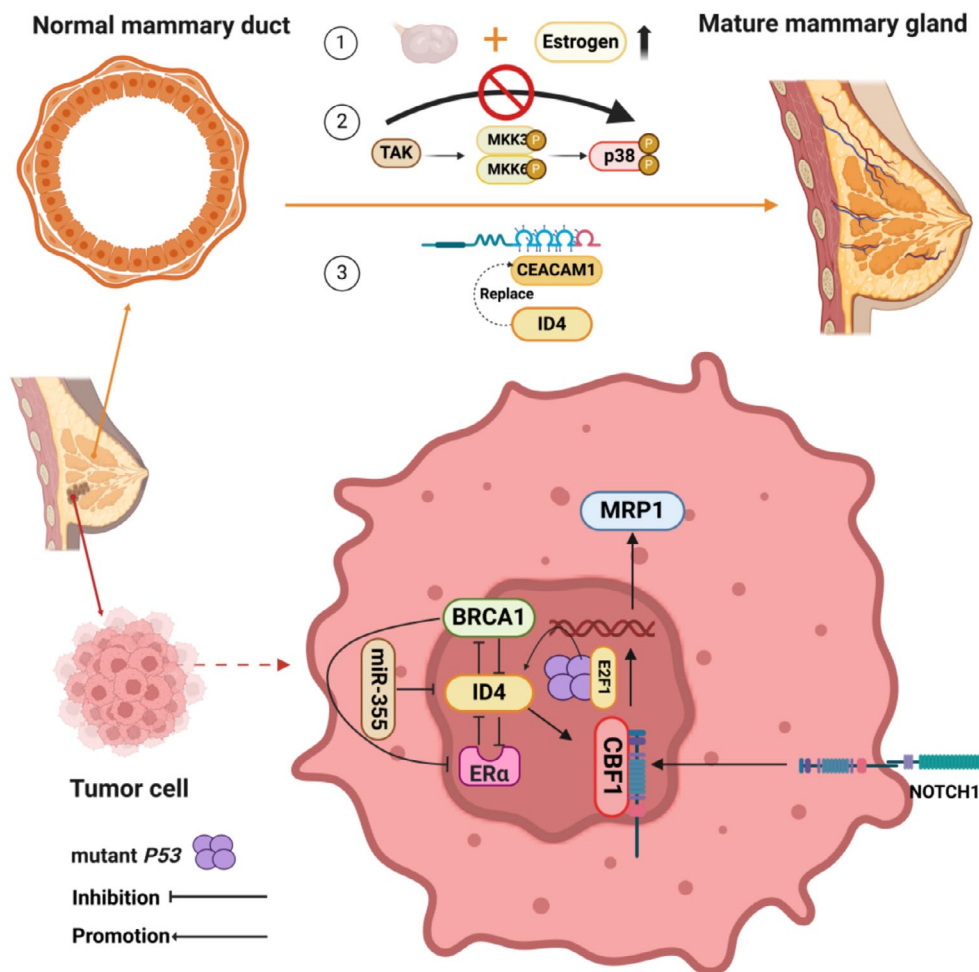


FIGURE 3 | Diagram of the function of ID4 as a bridge linking mammary gland development and breast tumors. The top part of the figure represents that ID4 promotes mammary gland development through three major pathways. Among them, ①: ID4 can maintain ovarian function and promote estrogen expression; ②: ID4 can inhibit the p38MAPK pathway to maintain mammary cell survival; ③: ID4 can replace the CEACAM1 protein to repromote the formation of lumen in BC cells. The lower part of the figure illustrates the mechanism by which ID4 promotes breast carcinogenesis. These possible mechanisms include the interaction between ID4 and BRCA1, ERα, and participation in the NOTCH1 pathway. The protein complex of mutant p53-E2F1 binds to a specific region of the ID4 promoter and promotes ID4 expression. Created in <https://BioRender.com>.

BRAC1 was the first oncogene identified in the breast and ovary, showing inherited mutations of *BRAC1* in the hereditary BC population and significantly reduced *BRAC1* expression levels in sporadic BC patients [53], and as early as 2001, Beger et al. screened out the gene *ID4* as a key upstream regulator of *BRAC1* by using a ribozyme-library-based inverse genomics approach [54]. Subsequently, in 2003, using the rat carcinogen-induced BC model, researchers found that BC-inducing chemicals significantly increased the expression level of *ID4* in mammary epithelial cells [55]. This evidence strongly suggested that breast carcinogenesis was accompanied by significant changes in *ID4* expression level and, of particular importance, that the BC geographic oncogene *BRAC1* was strongly associated with *ID4* expression. While *BRAC1* genetic mutations are not significantly associated with nonhereditary BC, data suggested reduced or undetectable levels of *BRAC1* mRNA in sporadic BC, ovarian cancer, and BC cell lines [54]. Analysis of clinical samples [56] demonstrated that *ID4* is amplified and overexpressed at a higher frequency in *BRAC1*-mutant basal-like breast cancer (BLBC) compared with sporadic BLBC. A study suggested that *BRAC1*-loss BLBC originated from luminal epithelial progenitor cells rather than from basal stem cells. BC caused by specific knockdown of *Brac1* from mammary basal stem cells did not behave in the same way as human *BRAC1*-associated BC or sporadic BLBC, whereas BC caused by specific knockdown of *Brac1* from luminal epithelial cells behaved in a typical BLBC [57]. In the context of BC, overexpression of *ID4* is strongly associated with the TNBC [58, 59] and is negatively correlated with the expression of *BRAC1* and ER α [60]. In 2012, researchers suggested that high expression of *ID4* protein may be related to *BRAC1* downregulation in TNBC, and TNBC with *ID4* overexpression showed a basal-like phenotype, consistent with the fact that BLBC is associated with low-*BRAC1* expression [59]. In a large number of subsequent studies, the reciprocal regulatory mechanisms of *ID4* and *BRAC1* have also been gradually revealed, such as *ID4* negatively controlling the expression of *Brca1* [54], and the *BRAC1*-positive regulator miR-335 downregulating the expression of *ID4* (Figure 3) [61]. Furthermore, silencing *ID4* led to an increase in *BRAC1* expression and resulted in a less aggressive phenotype in the MDA-MB231 cell lines. The result of a study [56] from proteogenomic analysis of *ID4* in BLBC indicated that *ID4* localizes DNA at sites of active transcription and DNA damage, bridged through its biochemical interaction with the DNA damage response machinery, namely MDC1 (mediator of DNA damage checkpoint protein 1). Through MDC1, *ID4* interacts with other DNA repair proteins (γ H2AX and *BRAC1*) at fragile chromatin sites [56]. These results suggested a role for *ID4* in the DNA damage response rather than regulation of transcription at these sites. However, the mechanism of interaction between *ID4* and *BRAC1* has not been fully elucidated, and even conflicting conclusions between some studies have been observed; thus, more in-depth studies are urgently needed.

In BC cell lines SKBR3 and MDA-MB-231, *ID4* is found to interact with the *p53* mutants R175H and R280K, facilitating angiogenesis [62]. Conversely, in MCF7 cells with wild-type *p53*, this interaction is absent [62]. The underlying mechanism is that the protein complex of mutant *p53*-E2F1 binds to a specific region of the *ID4* promoter and promotes *ID4* expression [62] (Figure 3). In highly aggressive TNBC, MALAT1 is associated with the degree of malignancy, and the researchers noted that

in *p53*-mutant TNBC, the cancer cells gain-of-function express VEGFA isoforms, and the *ID4*-*p53* complex has a role in regulating the recruitment of lncRNA MALAT1, which in turn regulates the degree of malignancy of TNBC [63]. Subsequent research [64] revealed that the lncRNA MALAT1 and the *ID4* protein promote the back-splicing of VEGFA exon 7, resulting in the formation of the circular RNA circ_0076611. This circular RNA interacts with various proliferation-related transcripts, including MYC and VEGFA mRNAs, thereby enhancing the proliferation and migration of TNBC cells. However, a meta-analysis showed that there was no significant correlation between *ID4* expression and *p53* mutation in BC, and this conclusion was validated in the TCGA data [65] (Figure 4).

In addition to the correlation between *ID4* and the oncogene *BRAC1* and the tumor suppressor *p53*, which has been studied in depth, the correlation between *ID4* expression status and the ER in BC has also been emphasized. Early in 2006, Paola de Candia et al. detected *ID4* mRNA in BC tissues of different pathological types and found that *ID4* mRNA levels were related to the distribution and density of ER, independently of Her-2 status, and that *ID4* mRNA was present in nontumor breast tissues and ER-negative mammary epithelium, whereas ER-positive cells were *ID4* negative [58]. This evidence suggested that *ID4* may act as a tumor suppressor gene in ER-positive BC and may be regulated by estrogen. *ID4* is not expressed in ER α -positive atypical ductal hyperplasia, ductal carcinoma in situ, and invasive carcinoma, but is present in ER α -negative mammary epithelial cells, suggesting that ER α might negatively regulate *ID4* (Figure 3) [58, 60]. *ID4* expression is increased in BLBC/TNBC, but not in non-TNBC [66–68]. However, in a meta-analysis, there was no significant correlation between *ID4* status and BC subtypes [69]. TCGA database analysis showed that *ID4* expression was different in different molecular and pathological subtypes (Figure 5). Further comprehensive studies are needed to reveal the correlation between ER status and *ID4* expression.

The latest research [70] suggested that *ID4* expression in BC cells is linked with the activation of motility pathways and enhances the production of VEGFA. This enhancement facilitates

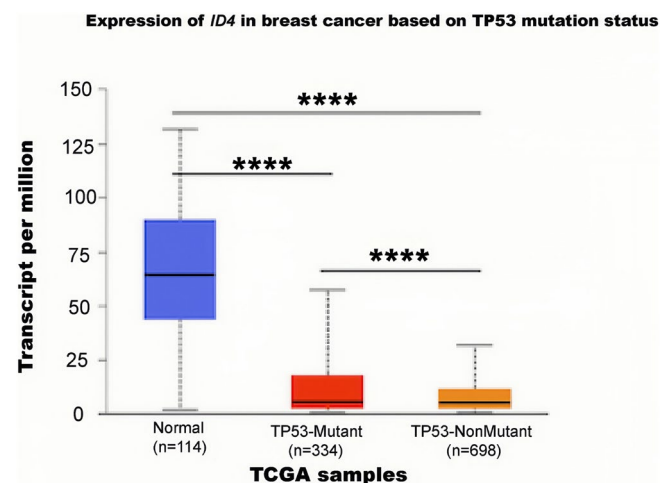


FIGURE 4 | Expression of *ID4* in breast cancer based on TP53 mutation status. Data were derived from breast cancer samples in The Cancer Genome Atlas (TCGA).

a paracrine interaction between VEGFR2 and integrin $\beta 3$. Such interactions trigger the focal adhesion pathway downstream, promoting cell migration, invasion, and the formation of stress fibers [70]. This pathway has also been verified in other studies [71, 72], such as miR342 downregulating the expression of its target gene *Id4*, which further leads to the reduction of VEGF and Bcl2 (anti-apoptotic)/Bax (pro-apoptotic) ratio, thereby inhibiting the progression of TNBC.

Meanwhile, researchers found that *ID4* is frequently silenced by promoter methylation in ER+ BC and functions as a tumor suppressor gene in these tumors [73], and we also demonstrated that the luminal BC has a higher level of *ID4* promoter methylation using the TCGA database (Figure 6). Furthermore, researchers found that ER+ BC with *ID4* hypomethylation were more likely to develop tamoxifen resistance [74], so interventions targeting the regulation of *ID4* methylation levels have potential function for treating endocrine-resistant BC. In addition, *ID4* methylation status could serve as a prognostic biomarker in BC [75].

Using a melanoma model, the researchers found that the regulation of TGF- β in the tumor immuno-microenvironment is closely related to the expression of *ID4* and that the expression level of *ID4* significantly modulates the degree of immune infiltration, altering the tumor responsiveness to immunosuppressive therapy [76]. Donzelli et al. [77] showed a significant correlation between ID4 and macrophage marker CD68 protein expression in a range of triple-negative breast tumors. Their further research [78] showed that activation of ID4 expression in tumor-associated macrophages is observed as a consequence of BC cell paracrine activity and could participate in macrophage reprogramming in BC. Intervention in the immuno-microenvironment of BC has been shown to have significant efficacy, so whether ID4 can similarly modulate the immuno-microenvironment of BC needs to be further elucidated and is expected to be a means of combination immunotherapy.

Although we only discuss the role of ID4 in the occurrence and development of BC in this article, other ID proteins are also inevitably involved in this process, and understanding their roles

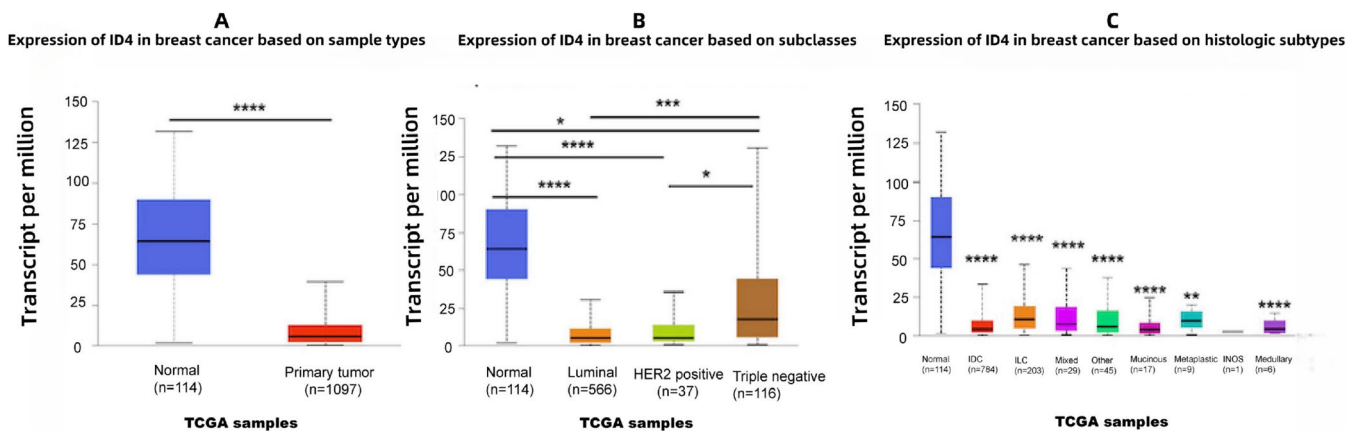


FIGURE 5 | Analysis of ID4 expression in breast cancer. It illustrates the expression levels of the ID4 gene across distinct feature-based classifications of breast cancer: (A) sample types (e.g., tumor tissue vs. normal tissue), (B) molecular subtypes (e.g., Luminal A, Luminal B, HER2+, Triple negative), and (C) histologic subtypes (e.g., ductal carcinoma, lobular carcinoma). The vertical axis represents transcript abundance in transcripts per million (TPM). Data were derived from breast cancer samples in The Cancer Genome Atlas (TCGA). IDC, invasive ductal carcinoma; ILC, invasive lobular carcinoma; INOS, invasive carcinoma not otherwise specified. * $p < 0.05$, ** $p < 0.01$, *** or **** $p < 0.0001$.

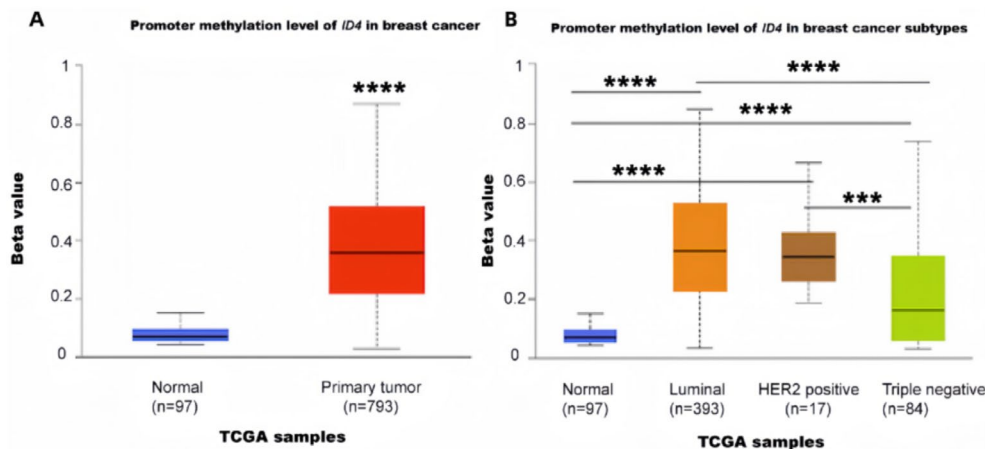


FIGURE 6 | (A) Promoter methylation β values of ID4 in normal breast tissue ($n = 97$) versus primary breast tumors ($n = 793$). (B) Methylation β -values of ID4 across breast cancer subtypes: Normal tissue ($n = 97$), luminal ($n = 393$), HER2-positive ($n = 17$), and triple-negative ($n = 84$). β -values range from 0 (unmethylated) to 1 (fully methylated). Data derived from TCGA samples. *** or **** means $p < 0.0001$.

may find new breakthroughs for ID4. For example, Id1 and Id3 have an important role in maintaining the cancer stem cell (CSC) phenotype in the TNBC subtype [79–81]. Similarly, ID4 is a key regulator of CSC self-renewal and marks a subset of BLBC with a putative mammary basal cell of origin [82]. Not only that, Id1 may be a general negative regulator of anticancer drug-induced apoptosis [83, 84], which revealed a joint role of Id4 and Id1 in BC development.

A study [85] showed that breast tumors with *ID4* overexpression presented significantly decreased rates of overall survival (OS, 69.4% vs. 82.2%; $p=0.013$) and disease-free survival (DFS, 61.2% vs. 86.3%; $p<0.001$). *ID1* also presented markedly shorter OS and DFS rates. On the contrary, neither *ID2* nor *ID3* overexpression was associated with a worse prognosis in this subgroup of BC patients. Surprisingly, *ID4* did not significantly correlate with overall survival for these patients in the public database [85]. Similarly, we did not see a significant survival difference in the TCGA database (Figure 7).

7 | Summary and Prospect

ID4 plays a pivotal role in various biological processes, particularly in cellular differentiation and cancer progression [1]. As a member of the transcriptional regulator family, ID4 lacks a DNA-binding domain and functions primarily as a negative regulator of other b-HLH proteins, leading to the formation of inactive heterodimers [4, 27]. This unique mechanism makes ID4 a critical player in embryonic development, where its expression decreases with cell differentiation [2–4]. ID4 is highly expressed in specific tissues, including the brain, thyroid, and mammary gland, and is implicated in osteoblast differentiation, neural precursor cell proliferation, and the regulation of several signaling pathways [20, 28, 30].

ID4 plays a crucial role in breast carcinogenesis by regulating the NOTCH1 signaling pathway and exhibiting reciprocal negative regulation with *BRCAl*, particularly in BLBC and TNBC [51, 54, 56]. Additionally, ID4 interacts with mutant *p53* to promote angiogenesis and is influenced by ER status, suggesting its

potential function as a tumor suppressor in ER + BC [58]. Recent studies indicate that ID4 expression in BC activates motility pathways and enhances VEGFA production, promoting cell migration and invasion [70–72]. In ER + BC, ID4 is frequently silenced by methylation, functioning as a tumor suppressor and correlating with tamoxifen resistance, thereby highlighting its potential as a target for therapeutic intervention [74, 75]. Moreover, ID4 shows a strong correlation with macrophage marker CD68 expression, suggesting its role in macrophage reprogramming driven by paracrine signaling from BC cells [77, 78]. Notably, ID4's expression has prognostic implications, with overexpression associated with poorer survival outcomes in BC patients [85]. However, findings regarding its prognostic value remain inconsistent across studies and databases, necessitating further exploration.

The targeted inhibition of ID4 in breast cancer therapy remains an evolving field, with potential adverse effects largely dependent on the specific mechanisms of action of these agents. Given ID4's essential role in cellular differentiation and proliferation, its inhibition may disrupt tissue homeostasis, particularly in organs where it is highly expressed, such as the brain, thyroid, and reproductive system, potentially leading to neurodevelopmental abnormalities, thyroid dysfunction, or reproductive disturbances. As a key regulator of mammary gland development and estrogen receptor (ER) signaling, ID4 suppression may interfere with endocrine homeostasis, increasing the risk of menstrual irregularities, altered estrogen signaling, and other hormone-related dysfunctions. Additionally, ID4 plays a critical role in shaping the tumor microenvironment, particularly in macrophage reprogramming and immune cell infiltration, suggesting that its inhibition could impair antitumor immunity or increase susceptibility to infections due to immune dysregulation. Moreover, ID4 exhibits a dual role in tumorigenesis, functioning as an oncogene in certain malignancies such as bladder cancer and leukemia while acting as a tumor suppressor in others, including ER-positive breast cancer. Consequently, its targeted inhibition may lead to unintended oncogenic effects, particularly in patients with multiple cancer predispositions, highlighting the need for careful risk assessment. Furthermore, given its established role in chemotherapy resistance via the CBF1-MRP1

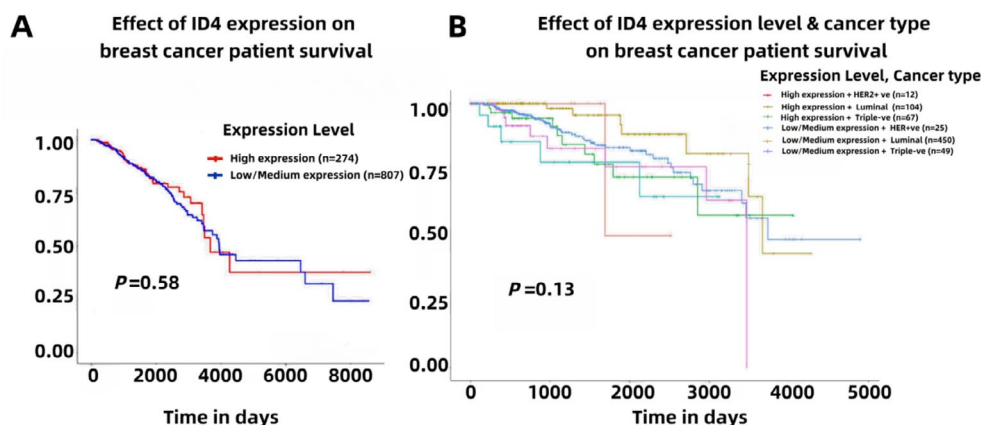


FIGURE 7 | (A) Kaplan–Meier survival analysis comparing breast cancer patients with high ID4 expression ($n=274$) versus low/medium ID4 expression ($n=807$). (B) Survival analysis stratified by ID4 expression levels and cancer subtypes: High expression combined with HER2-positive ($n=12$), luminal ($n=109$), or triple-negative ($n=27$), and low/medium expression combined with HER2-positive ($n=35$), luminal ($n=450$), or triple-negative ($n=49$). Time is shown in days. Data derived from TCGA samples.

pathway, ID4 inhibition may enhance chemosensitivity; however, it could also disrupt apoptotic pathways, resulting in unpredictable cellular responses. These potential risks underscore the necessity for a highly stratified approach in the clinical application of ID4-targeted therapies, integrating molecular profiling to identify suitable patient populations and ensuring precise drug delivery to mitigate adverse effects.

Future research should focus on elucidating the multifaceted roles of ID4 in cancer biology and tissue differentiation, particularly in BC. Investigating the molecular mechanisms underlying ID4's regulatory functions could provide insights into its dual role as a tumor suppressor and an oncogene. Employing advanced genomic, epigenomics, transcriptomics, metabolomics, and lipidomics approaches will enhance our understanding of ID4's interactions with various signaling pathways and transcription factors, particularly in the context of tumor microenvironments [76–78].

Additionally, exploring the potential of ID4 as a therapeutic target in cancer treatment may yield significant clinical implications. The impact of ID4 on immune cell infiltration and its role in modulating the tumor immune microenvironment offer promising avenues for combination immunotherapy strategies. Understanding the mechanisms driving ID4 methylation and expression in BC could lead to novel biomarkers for treatment resistance and recurrence [86, 87].

In summary, the continued investigation of ID4's biological functions and its role in BC will be essential for developing targeted therapies and improving prognostic accuracy in clinical settings.

Author Contributions

Conceptualization: Y.S., P.Z., and S.B.; Writing – original draft preparation: Y.S., Y.Y., P.Z., and S.B. Writing – review and editing: Y.Y., Y.S., X.W., and H.Z. All authors have read and agreed to the published version of the manuscript.

Acknowledgments

The authors have nothing to report.

Consent

All authors read and approved the submission and final publication.

Conflicts of Interest

The authors declare no conflicts of interest.

Data Availability Statement

The data sets used and analyzed in this study are available from the corresponding author upon reasonable request.

References

1. C. Roschger and C. Cabrele, “The id-Protein Family in Developmental and Cancer-Associated Pathways,” *Cell Communication and Signaling: CCS* 15, no. 1 (2017): 7, <https://doi.org/10.1186/s12964-016-0161-y>.

2. C. Murre, “Helix-Loop-Helix Proteins and the Advent of Cellular Diversity: 30 Years of Discovery,” *Genes & Development* 33, no. 1–2 (2019): 6–25, <https://doi.org/10.1101/gad.320663.118>.
3. F. Ling, B. Kang, and X. H. Sun, “Id Proteins: Small Molecules, Mighty Regulators,” *Current Topics in Developmental Biology* 110 (2014): 189–216, <https://doi.org/10.1016/b978-0-12-405943-6.00005-1>.
4. J. P. Coppé, A. P. Smith, and P. Y. Desprez, “Id Proteins in Epithelial Cells,” *Experimental Cell Research* 285, no. 1 (2003): 131–145, [https://doi.org/10.1016/s0014-4827\(03\)00014-4](https://doi.org/10.1016/s0014-4827(03)00014-4).
5. S. K. Komaragiri, D. H. Bostanthirige, D. J. Morton, et al., “ID4 Promotes AR Expression and Blocks Tumorigenicity of PC3 Prostate Cancer Cells,” *Biochemical and Biophysical Research Communications* 478, no. 1 (2016): 60–66, <https://doi.org/10.1016/j.bbrc.2016.07.092>.
6. A. Tian, M. Schepers, R. Riemens, et al., “DNA Methylation Regulates the Expression of the Negative Transcriptional Regulators ID2 and ID4 During OPC Differentiation,” *Cellular and Molecular Life Sciences* 78, no. 19–20 (2021): 6631–6644, <https://doi.org/10.1007/s00018-021-03927-2>.
7. M. Y. Li, Y. Y. Xu, H. Y. Kang, et al., “Quantitative Detection of ID4 Gene Aberrant Methylation in the Differentiation of Myelodysplastic Syndrome From Aplastic Anemia,” *Chinese Medical Journal* 128, no. 15 (2015): 2019–2025, <https://doi.org/10.4103/0366-6999.161351>.
8. K. O. Uhm, E. S. Lee, Y. M. Lee, et al., “Differential Methylation Pattern of ID4, SFRP1, and SHP1 Between Acute Myeloid Leukemia and Chronic Myeloid Leukemia,” *Journal of Korean Medical Science* 24, no. 3 (2009): 493–497, <https://doi.org/10.3346/jkms.2009.24.3.493>.
9. R. L. Siegel, K. D. Miller, and A. Jemal, “Cancer Statistics, 2020,” *CA: A Cancer Journal for Clinicians* 70, no. 1 (2020): 7–30, <https://doi.org/10.3322/caac.21590>.
10. J. Y. S. Tsang and G. M. Tse, “Molecular Classification of Breast Cancer,” *Advances in Anatomic Pathology* 27, no. 1 (2020): 27–35, <https://doi.org/10.1097/pap.0000000000000232>.
11. N. F. Pondé, D. Zardavas, and M. Piccart, “Progress in Adjuvant Systemic Therapy for Breast Cancer,” *Nature Reviews. Clinical Oncology* 16, no. 1 (2019): 27–44, <https://doi.org/10.1038/s41571-018-0089-9>.
12. A. G. Waks and E. P. Winer, “Breast Cancer Treatment: A Review,” *JAMA* 321, no. 3 (2019): 288–300, <https://doi.org/10.1001/jama.2018.19323>.
13. Y. Z. Jiang, D. Ma, C. Suo, et al., “Genomic and Transcriptomic Landscape of Triple-Negative Breast Cancers: Subtypes and Treatment Strategies,” *Cancer Cell* 35, no. 3 (2019): 428–440, <https://doi.org/10.1016/j.ccell.2019.02.001>.
14. Y. R. Seo, J. Lee, H. S. Ryu, et al., “Lateral Interactions Between CD276 and CD147 Are Essential for Stemness in Breast Cancer: A Novel Insight From Proximal Proteome Analysis,” *Scientific Reports* 13, no. 1 (2023): 14242, <https://doi.org/10.1038/s41598-023-41416-7>.
15. N. Zakiyah, S. I. Wanandi, R. D. Antariantio, R. A. Syahrani, and S. Arumsari, “Mesenchymal Stem Cell-Derived Extracellular Vesicles Increase Human MCF7 Breast Cancer Cell Proliferation Associated With OCT4 Expression and ALDH Activity,” *Asian Pacific Journal of Cancer Prevention* 24, no. 8 (2023): 2781–2789, <https://doi.org/10.31557/apjcp.2023.24.8.2781>.
16. X. Bai, J. Ni, J. Beretov, P. Graham, and Y. Li, “Cancer Stem Cell in Breast Cancer Therapeutic Resistance,” *Cancer Treatment Reviews* 69 (2018): 152–163, <https://doi.org/10.1016/j.ctrv.2018.07.004>.
17. W. Chen, W. Wei, L. Yu, et al., “Mammary Development and Breast Cancer: A Notch Perspective,” *Journal of Mammary Gland Biology and Neoplasia* 26, no. 3 (2021): 309–320, <https://doi.org/10.1007/s10911-021-09496-1>.
18. C. Ercan, P. J. van Diest, and M. Vooijs, “Mammary Development and Breast Cancer: The Role of Stem Cells,” *Current Molecular Medicine* 11, no. 4 (2011): 270–285, <https://doi.org/10.2174/156652411795678007>.

19. X. Yang, H. Wang, and B. Jiao, "Mammary Gland Stem Cells and Their Application in Breast Cancer," *Oncotarget* 8, no. 6 (2017): 10675–10691, <https://doi.org/10.18632/oncotarget.12893>.
20. Z. Zebedee and E. Hara, "Id Proteins in Cell Cycle Control and Cellular Senescence," *Oncogene* 20, no. 58 (2001): 8317–8325, <https://doi.org/10.1038/sj.onc.1205092>.
21. A. E. Schade, M. Fischer, and J. A. DeCaprio, "RB, p130 and p107 Differentially Repress G1/S and G2/M Genes After p53 Activation," *Nucleic Acids Research* 47, no. 21 (2019): 11197–11208, <https://doi.org/10.1093/nar/gkz961>.
22. R. G. Russell, A. Lasorella, L. E. Dettin, and A. Iavarone, "Id2 Drives Differentiation and Suppresses Tumor Formation in the Intestinal Epithelium," *Cancer Research* 64, no. 20 (2004): 7220–7225, <https://doi.org/10.1158/0008-5472.Can-04-2095>.
23. Z. Zhao, Z. Bo, W. Gong, and Y. Guo, "Inhibitor of Differentiation 1 (Id1) in Cancer and Cancer Therapy," *International Journal of Medical Sciences* 17, no. 8 (2020): 995–1005, <https://doi.org/10.7150/ijms.42805>.
24. X. S. Chen, Y. H. Zhang, Q. Y. Cai, and Z. X. Yao, "ID2: A Negative Transcription Factor Regulating Oligodendroglia Differentiation," *Journal of Neuroscience Research* 90, no. 5 (2012): 925–932, <https://doi.org/10.1002/jnr.22826>.
25. K. Rahmouni and C. D. Sigmund, "Id3, E47, and SREBP-1c: Fat Factors Controlling Adiponectin Expression," *Circulation Research* 103, no. 6 (2008): 565–567, <https://doi.org/10.1161/circresaha.108.184366>.
26. M. Rigolet, T. Rich, M. S. Gross-Morand, D. Molina-Gomes, E. Viegas-Pequignot, and C. Junien, "cDNA Cloning, Tissue Distribution and Chromosomal Localization of the Human ID4 Gene," *DNA Research* 5, no. 5 (1998): 309–313, <https://doi.org/10.1093/dnares/5.5.309>.
27. M. V. Barone, R. Pepperkok, F. A. Peverali, and L. Philipson, "Id Proteins Control Growth Induction in Mammalian Cells," *Proceedings of the National Academy of Sciences of the United States of America* 91, no. 11 (1994): 4985–4988, <https://doi.org/10.1073/pnas.91.11.4985>.
28. D. Patel, D. J. Morton, J. Carey, M. C. Havrda, and J. Chaudhary, "Inhibitor of Differentiation 4 (ID4): From Development to Cancer," *Biochimica et Biophysica Acta* 1855, no. 1 (2015): 92–103, <https://doi.org/10.1016/j.bbcan.2014.12.002>.
29. P. Sharma, S. Chinaranagari, and J. Chaudhary, "Inhibitor of Differentiation 4 (ID4) Acts as an Inhibitor of ID-1, -2 and -3 and Promotes Basic Helix Loop Helix (bHLH) E47 DNA Binding and Transcriptional Activity," *Biochimie* 112 (2015): 139–150, <https://doi.org/10.1016/j.biochi.2015.03.006>.
30. C. Sachs, B. D. Robinson, L. Andres Martin, et al., "Evaluation of Candidate Spermatogonial Markers ID4 and GPR125 in Testes of Adult Human Cadaveric Organ Donors," *Andrology* 2, no. 4 (2014): 607–614, <https://doi.org/10.1111/j.2047-2927.2014.00226.x>.
31. J. A. Blake, C. J. Bult, J. T. Eppig, J. A. Kadin, and J. E. Richardson, "The Mouse Genome Database: Integration of and Access to Knowledge About the Laboratory Mouse," *Nucleic Acids Research* 42, no. Database issue (2014): D810–D817, <https://doi.org/10.1093/nar/gkt1225>.
32. L. Bedford, R. Walker, T. Kondo, I. van Cruchten, E. R. King, and F. Sablitzky, "Id4 is Required for the Correct Timing of Neural Differentiation," *Developmental Biology* 280, no. 2 (2005): 386–395, <https://doi.org/10.1016/j.ydbio.2005.02.001>.
33. Y. Tokuzawa, K. Yagi, Y. Yamashita, et al., "Id4, a New Candidate Gene for Senile Osteoporosis, Acts as a Molecular Switch Promoting Osteoblast Differentiation," *PLoS Genetics* 6, no. 7 (2010): e1001019, <https://doi.org/10.1371/journal.pgen.1001019>.
34. M. Marin-Husstege, Y. He, J. Li, T. Kondo, F. Sablitzky, and P. Casaccia-Bonnel, "Multiple Roles of Id4 in Developmental Myelination: Predicted Outcomes and Unexpected Findings," *Glia* 54, no. 4 (2006): 285–296, <https://doi.org/10.1002/glia.20385>.
35. P. Sharma, A. E. Knowell, S. Chinaranagari, et al., "Id4 Deficiency Attenuates Prostate Development and Promotes PIN-Like Lesions by Regulating Androgen Receptor Activity and Expression of NKX3.1 and PTEN," *Molecular Cancer* 12 (2013): 67, <https://doi.org/10.1186/1476-4598-12-67>.
36. J. P. Carey, A. J. Asirvatham, O. Galm, T. A. Ghogomu, and J. Chaudhary, "Inhibitor of Differentiation 4 (Id4) is a Potential Tumor Suppressor in Prostate Cancer," *BMC Cancer* 9, no. 1 (2009): 173, <https://doi.org/10.1186/1471-2407-9-173>.
37. A. E. Knowell, D. Patel, D. J. Morton, P. Sharma, S. Glymph, and J. Chaudhary, "Id4 Dependent Acetylation Restores Mutant-p53 Transcriptional Activity," *Molecular Cancer* 12 (2013): 161, <https://doi.org/10.1186/1476-4598-12-161>.
38. Q. Wu, M. J. Hoffmann, F. H. Hartmann, and W. A. Schulz, "Amplification and Overexpression of the ID4 Gene at 6p22.3 in Bladder Cancer," *Molecular Cancer* 4, no. 1 (2005): 16, <https://doi.org/10.1186/1476-4598-4-16>.
39. C. J. Bieberich, K. Fujita, W. W. He, and G. Jay, "Prostate-Specific and Androgen-Dependent Expression of a Novel Homeobox Gene," *Journal of Biological Chemistry* 271, no. 50 (1996): 31779–31782, <https://doi.org/10.1074/jbc.271.50.31779>.
40. R. Bhatia-Gaur, A. A. Donjacour, P. J. Scivolino, et al., "Roles for Nkx3.1 in Prostate Development and Cancer," *Genes & Development* 13, no. 8 (1999): 966–977, <https://doi.org/10.1101/gad.13.8.966>.
41. J. D. Zhou, X. X. Li, T. J. Zhang, et al., "MicroRNA-335/ID4 Dysregulation Predicts Clinical Outcome and Facilitates Leukemogenesis by Activating PI3K/Akt Signaling Pathway in Acute Myeloid Leukemia," *Aging (Albany NY)* 11, no. 10 (2019): 3376–3391, <https://doi.org/10.18632/aging.101991>.
42. A. S. Chan, W. Y. Tsui, X. Chen, et al., "Downregulation of ID4 by Promoter Hypermethylation in Gastric Adenocarcinoma," *Oncogene* 22, no. 44 (2003): 6946–6953, <https://doi.org/10.1038/sj.onc.1206799>.
43. M. Bellido, A. Aventín, A. Lasa, et al., "Id4 Is Deregulated by a t(6;14)(p22;q32) Chromosomal Translocation in a B-Cell Lineage Acute Lymphoblastic Leukemia," *Haematologica* 88, no. 9 (2003): 994–1001.
44. X. Z. Gao, W. G. Zhao, G. N. Wang, M. Y. Cui, Y. R. Zhang, and W. C. Li, "Inhibitor of DNA Binding 4 Functions as a Tumor Suppressor and Is Targetable by 5-Aza-2'-Deoxycytosine With Potential Therapeutic Significance in Burkitt's Lymphoma," *Molecular Medicine Reports* 13, no. 2 (2016): 1269–1274, <https://doi.org/10.3892/mmr.2015.4640>.
45. R. Nair, S. Junankar, S. O'Toole, et al., "Redefining the Expression and Function of the Inhibitor of Differentiation 1 in Mammary Gland Development," *PLoS One* 5, no. 8 (2010): e11947, <https://doi.org/10.1371/journal.pone.0011947>.
46. N. S. Kim, H. T. Kim, M. C. Kwon, et al., "Survival and Differentiation of Mammary Epithelial Cells in Mammary Gland Development Require Nuclear Retention of Id2 due to RANK Signaling," *Molecular and Cellular Biology* 31, no. 23 (2011): 4775–4788, <https://doi.org/10.1128/mcb.05646-11>.
47. J. Dong, S. Huang, M. Caikovski, et al., "ID4 Regulates Mammary Gland Development by Suppressing p38MAPK Activity," *Development* 138, no. 23 (2011): 5247–5256, <https://doi.org/10.1242/dev.069203>.
48. S. A. Best, K. J. Hutt, N. Y. Fu, et al., "Dual Roles for Id4 in the Regulation of Estrogen Signaling in the Mammary Gland and Ovary," *Development* 141, no. 16 (2014): 3159–3164, <https://doi.org/10.1242/dev.108498>.
49. T. Nguyen and J. E. Shively, "Induction of Lumen Formation in a Three-Dimensional Model of Mammary Morphogenesis by Transcriptional Regulator ID4: ROLE OF CaMK2D IN THE EPIGENETIC REGULATION OF ID4 GENE EXPRESSION," *Journal of Biological Chemistry* 291, no. 32 (2016): 16766–16776, <https://doi.org/10.1074/jbc.M115.710160>.

50. L. Choy, T. J. Hagenbeek, M. Solon, et al., "Constitutive NOTCH3 Signaling Promotes the Growth of Basal Breast Cancers," *Cancer Res* 77, no. 6 (2017): 1439–1452, <https://doi.org/10.1158/0008-5472.Can-16-1022>.
51. X. Zhang, G. Gu, L. Song, et al., "ID4 Promotes Breast Cancer Chemotherapy Resistance via CBF1-MRP1 Pathway," *Journal of Cancer* 11, no. 13 (2020): 3846–3857, <https://doi.org/10.7150/jca.31988>.
52. J. E. Visvader, "Keeping Abreast of the Mammary Epithelial Hierarchy and Breast Tumorigenesis," *Genes & Development* 23, no. 22 (2009): 2563–2577, <https://doi.org/10.1101/gad.1849509>.
53. G. Sourvinos and D. A. Spandidos, "Decreased BRCA1 Expression Levels May Arrest the Cell Cycle Through Activation of p53 Checkpoint in Human Sporadic Breast Tumors," *Biochemical and Biophysical Research Communications* 245, no. 1 (1998): 75–80, <https://doi.org/10.1006/bbrc.1998.8379>.
54. C. Beger, L. N. Pierce, M. Kruger, et al., "Identification of Id4 as a Regulator of BRCA1 Expression by Using a Ribozyme-Library-Based Inverse Genomics Approach," *Proceedings of the National Academy of Sciences of the United States of America* 98, no. 1 (2001): 130–135, <https://doi.org/10.1073/pnas.98.1.130>.
55. L. Shan, M. Yu, C. Qiu, and E. G. Snyderwine, "Id4 Regulates Mammary Epithelial Cell Growth and Differentiation and Is Overexpressed in Rat Mammary Gland Carcinomas," *American Journal of Pathology* 163, no. 6 (2003): 2495–2502, [https://doi.org/10.1016/s0002-9440\(10\)63604-8](https://doi.org/10.1016/s0002-9440(10)63604-8).
56. L. A. Baker, H. Holliday, D. Roden, et al., "Proteogenomic Analysis of Inhibitor of Differentiation 4 (ID4) in Basal-Like Breast Cancer," *Breast Cancer Research* 22, no. 1 (2020): 63, <https://doi.org/10.1186/s13058-020-01306-6>.
57. G. Molyneux, F. C. Geyer, F. A. Magnay, et al., "BRCA1 Basal-Like Breast Cancers Originate From Luminal Epithelial Progenitors and Not From Basal Stem Cells," *Cell Stem Cell* 7, no. 3 (2010): 403–417, <https://doi.org/10.1016/j.stem.2010.07.010>.
58. P. de Candia, M. Akram, R. Benezra, and E. Brogi, "Id4 Messenger RNA and Estrogen Receptor Expression: Inverse Correlation in Human Normal Breast Epithelium and Carcinoma," *Human Pathology* 37, no. 8 (2006): 1032–1041, <https://doi.org/10.1016/j.humpath.2006.03.004>.
59. Y. H. Wen, A. Ho, S. Patil, et al., "Id4 Protein Is Highly Expressed in Triple-Negative Breast Carcinomas: Possible Implications for BRCA1 Downregulation," *Breast Cancer Research and Treatment* 135, no. 1 (2012): 93–102, <https://doi.org/10.1007/s10549-012-2070-0>.
60. G. Roldán, L. Delgado, and I. M. Musé, "Tumoral Expression of BRCA1, Estrogen Receptor Alpha and ID4 Protein in Patients With Sporadic Breast Cancer," *Cancer Biology & Therapy* 5, no. 5 (2006): 505–510, <https://doi.org/10.4161/cbt.5.5.2597>.
61. H. Heyn, M. Engelmann, S. Schreek, et al., "MicroRNA miR-335 Is Crucial for the BRCA1 Regulatory Cascade in Breast Cancer Development," *International Journal of Cancer* 129, no. 12 (2011): 2797–2806, <https://doi.org/10.1002/ijc.25962>.
62. G. Fontemaggi, S. Dell'Orso, D. Trisciuglio, et al., "The Execution of the Transcriptional Axis Mutant p53, E2F1 and ID4 Promotes Tumor Neo-Angiogenesis," *Nature Structural & Molecular Biology* 16, no. 10 (2009): 1086–1093, <https://doi.org/10.1038/nsmb.1669>.
63. M. Pruszek, E. Milano, M. Forcato, et al., "The Mutant p53-ID4 Complex Controls VEGFA Isoforms by Recruiting lncRNA MALAT1," *EMBO Reports* 18, no. 8 (2017): 1331–1351, <https://doi.org/10.15252/embr.201643370>.
64. C. Turco, G. Esposito, A. Izaia, et al., "MALAT1-Dependent hsa_circ_0076611 Regulates Translation Rate in Triple-Negative Breast Cancer," *Communications Biology* 5, no. 1 (2022): 598, <https://doi.org/10.1038/s42003-022-03539-x>.
65. D. Coradini, M. Fornili, F. Ambrogi, P. Boracchi, and E. Biganzoli, "TP53 Mutation, Epithelial-Mesenchymal Transition, and Stemlike Features in Breast Cancer Subtypes," *Journal of Biomedicine & Biotechnology* 2012 (2012): 254085, <https://doi.org/10.1155/2012/254085>.
66. N. C. Turner, J. S. Reis-Filho, A. M. Russell, et al., "BRCA1 Dysfunction in Sporadic Basal-Like Breast Cancer," *Oncogene* 26, no. 14 (2007): 2126–2132, <https://doi.org/10.1038/sj.onc.1210014>.
67. S. J. Park, R. J. Kim, and J. S. Nam, "Inhibitor of DNA-Binding 4 Contributes to the Maintenance and Expansion of Cancer Stem Cells in 4T1 Mouse Mammary Cancer Cell Line," *Laboratory Animal Research* 27, no. 4 (2011): 333–338, <https://doi.org/10.5625/lar.2011.27.4.333>.
68. M. T. Branham, D. M. Marzese, S. R. Laurito, et al., "Methylation Profile of Triple-Negative Breast Carcinomas," *Oncogene* 1, no. 7 (2012): e17, <https://doi.org/10.1038/oncsis.2012.17>.
69. A. Graudenzi, C. Cava, G. Bertoli, et al., "Pathway-Based Classification of Breast Cancer Subtypes," *Frontiers in Bioscience* 22, no. 10 (2017): 1697–1712, <https://doi.org/10.2741/4566>.
70. A. Benedetti, C. Turco, E. Gallo, et al., "ID4-Dependent Secretion of VEGFA Enhances the Invasion Capability of Breast Cancer Cells and Activates YAP/TAZ via Integrin β 3-VEGFR2 Interaction," *Cell Death & Disease* 15, no. 2 (2024): 113, <https://doi.org/10.1038/s41419-024-06491-2>.
71. Z. Alidoost, F. Attari, F. Saadatpour, and E. Arefian, "Inhibitory Effect of miR342 on the Progression of Triple-Negative Breast Cancer Cells In Vitro and in the Mice Model," *BioImpacts: BI* 14, no. 1 (2024): 27758, <https://doi.org/10.34172/bi.2023.27758>.
72. S. Yu, Y. Zhou, L. Niu, Y. Qiao, and Y. Yan, "Mesenchymal Stem Cell-Derived Exosome Mir-342-3p Inhibits Metastasis and Chemo-Resistance of Breast Cancer Through Regulating ID4," *Genes & Genomics* 44, no. 5 (2022): 539–550, <https://doi.org/10.1007/s13258-021-01200-1>.
73. D. Nasif, E. Campoy, S. Laurito, et al., "Epigenetic Regulation of ID4 in Breast Cancer: Tumor Suppressor or Oncogene? Clin," *Epigenetics* 10, no. 1 (2018): 111, <https://doi.org/10.1186/s13148-018-0542-8>.
74. Y. Zhang, B. Zhang, J. Fang, and X. Cao, "Hypomethylation of DNA-Binding Inhibitor 4 Serves as a Potential Biomarker in Distinguishing Acquired Tamoxifen-Refractory Breast Cancer," *International Journal of Clinical and Experimental Pathology* 8, no. 8 (2015): 9500–9505.
75. E. Noetzel, J. Veeck, D. Niederacher, et al., "Promoter Methylation-Associated Loss of ID4 Expression Is a Marker of Tumour Recurrence in Human Breast Cancer," *BMC Cancer* 8 (2008): 154, <https://doi.org/10.1186/1471-2407-8-154>.
76. K. A. DiVito, V. A. Trabosh, Y. S. Chen, C. M. Simbulan-Rosenthal, and D. S. Rosenthal, "Inhibitor of Differentiation-4 (Id4) Stimulates Pigmentation in Melanoma Leading to Histocyte Infiltration," *Experimental Dermatology* 24, no. 2 (2015): 101–107, <https://doi.org/10.1111/exd.12582>.
77. S. Donzelli, E. Milano, M. Pruszek, et al., "Expression of ID4 Protein in Breast Cancer Cells Induces Reprogramming of Tumour-Associated Macrophages," *Breast Cancer Research* 20, no. 1 (2018): 59, <https://doi.org/10.1186/s13058-018-0990-2>.
78. S. Donzelli, A. Sacconi, C. Turco, et al., "Paracrine Signaling From Breast Cancer Cells Causes Activation of ID4 Expression in Tumor-Associated Macrophages," *Cells* 9, no. 2 (2020): 418, <https://doi.org/10.3390/cells9020418>.
79. A. P. Thankamony, R. Murali, N. Karthikeyan, et al., "Targeting the Id1-Kif11 Axis in Triple-Negative Breast Cancer Using Combination Therapy," *Biomolecules* 10, no. 9 (2020): 1295, <https://doi.org/10.3390/biom10091295>.
80. W. S. Teo, H. Holliday, N. Karthikeyan, et al., "Id Proteins Promote a Cancer Stem Cell Phenotype in Mouse Models of Triple Negative Breast Cancer via Negative Regulation of Robo1," *Frontiers in Cell and Development Biology* 8 (2020): 552, <https://doi.org/10.3389/fcell.2020.00552>.
81. D. H. Shin, J. H. Park, J. Y. Lee, et al., "Overexpression of Id1 in Transgenic Mice Promotes Mammary Basal Stem Cell Activity and

Breast Tumorigenesis,” *Oncotarget* 6, no. 19 (2015): 17276–17290, <https://doi.org/10.18632/oncotarget.3640>.

82. S. Junankar, L. A. Baker, D. L. Roden, et al., “ID4 Controls Mammary Stem Cells and Marks Breast Cancers With a Stem Cell-Like Phenotype,” *Nature Communications* Mar 27 2015; 6 (2015): 6548, <https://doi.org/10.1038/ncomms7548>.

83. X. Zhang, M. T. Ling, Y. C. Wong, and X. Wang, “Evidence of a Novel Antiapoptotic Factor: Role of Inhibitor of Differentiation or DNA Binding (Id-1) in Anticancer Drug-Induced Apoptosis,” *Cancer Science* 98, no. 3 (2007): 308–314, <https://doi.org/10.1111/j.1349-7006.2007.00400.x>.

84. H. Kim, H. Chung, H. J. Kim, et al., “Id-1 Regulates Bcl-2 and Bax Expression Through p53 and NF-kappaB in MCF-7 Breast Cancer Cells,” *Breast Cancer Research and Treatment* 112, no. 2 (2008): 287–296, <https://doi.org/10.1007/s10549-007-9871-6>.

85. M. Garcia-Escolano, Y. G. Montoyo-Pujol, F. Ortiz-Martinez, et al., “ID1 and ID4 Are Biomarkers of Tumor Aggressiveness and Poor Outcome in Immunophenotypes of Breast Cancer,” *Cancers* 13, no. 3 (2021): 492, <https://doi.org/10.3390/cancers13030492>.

86. D. Nasif, S. Real, M. Roqué, and M. T. Branham, “CDC42 as an Epigenetic Regulator of ID4 in Triple-Negative Breast Tumors,” *Breast Cancer* 29, no. 3 (2022): 562–573, <https://doi.org/10.1007/s12282-022-01334-4>.

87. H. Kang, X. Wang, L. Gao, et al., “Clinical Implications of the Quantitative Detection of ID4 Gene Methylation in Myelodysplastic Syndrome,” *European Journal of Medical Research* 20, no. 1 (2015): 16, <https://doi.org/10.1186/s40001-015-0092-x>.