

Pollen Alone or a Mixture of Pollen Types? Assessing Their Suitability for Mass Rearing of *Neoseiulus cucumeris* (Acari: Phytoseiidae) Over 20 Generations

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Subject Editor: Muhammad Chaudhury

Received 30 April 2022; Editorial decision 21 June 2022.

Abstract

The generalist predatory mite, *Neoseiulus cucumeris* (Oudemans) is known as one of the most effective natural enemies on many pests. This economically important biocontrol agent was reared for 20 generations on date palm and castor bean pollen as well as a mixture of pollen types, including date palm, castor bean, and almond. The performance of this predator was evaluated by comparing its life table parameters after different generations (G1–G20) fed on each diet in a laboratory at $25 \pm 1^{\circ}$ C, $60 \pm 5\%$ RH, and a photoperiod of 16:8 (L: D) h. The development time of the predator reared on all tested diets had no significant difference in G20. The intrinsic rate of increase (*r*) of *N. cucumeris* by feeding on the mixed pollen (0.197 day⁻¹) was significantly higher than that on castor bean, or date palm pollen (0.146 or 0.152 day⁻¹ in G1, respectively). Our results indicated that the predator's performance was not affected by long-term feeding on the pollen diets, as well as there was no considerable difference between pollen alone and pollen mixture diets. Furthermore, mites reared on pollen diets had higher quality than those reared on natural prey, *Tetranychus urticae* Koch.

Key words: Phytoseiidae, alternative food, life table, main prey, mass rearing

Biological control has long been known as a highly efficient and environmentally safe pest management procedure, and predatory mites as biocontrol agents are increasingly used in pest control programs across the globe (van Lenteren 2012). The phytoseiid predatory mites can control tiny insect and mite pests, which have been classified as specialists and generalists. The generalist predatory mites can feed on different food and diets, including small arthropods, nematodes, fungi, and pollen (McMurtry et al. 2013).

Neoseiulus cucumeris (Oudemans) is a generalist phytoseiid predator known because of its success in controlling a wide range of pests in greenhouses (Zhang et al. 2011, Delisle et al. 2015). Feeding and reproducing on a pollen diet is a considerable characteristic of this predatory mite that facilitates the cost-effective production of large numbers of this predator so-called mass rearing. Since the performance of a predator is conditioned by long-term mass rearing (Nemati and Riahi 2020), the potential of *N. cucumeris* has already been evaluated on *Tetranychus urticae* Koch (Acari: Tetranychidae) as main prey and *Tyrophagus putrescentiae* (Schrank) (Acari: Acaridae) as factitious prey for 30 generations (Yazdanpanah et al. 2022).

It is generally accepted that both inbreeding-mediated reduction in fitness and loss of genetic variation induced during long-term rearing can lead to poor quality natural enemies, inappropriate control results, and unpredictable consequence of augmentative biological control (van Lenteren 2003, Cônsoli et al. 2010). Therefore, the quality of mass-reared organisms should be evaluated during mass rearing to monitor their quality to be sure they are still in a condition to be reared and appropriately control the target pest after release (van Lenteren 1991, Bellutti 2011, Sørensen et al. 2012).

Although several pollen grains such as castor bean and date palm are suitable alternative diets for *N. cucumeris* (Yazdanpanah et al. 2021a), they have not been tested as a long time food source. Therefore, in the current study, we investigated the multigenerational impacts of date palm pollen, castor bean pollen, and a mixture of two mentioned pollen as well as almond pollen on *N. cucumeris* over 20 consecutive generations. Furthermore, the data were compared with the previously obtained data on a long time rearing of the predator on the main prey, *T. urticae* (Yazdanpanah et al. 2022).

Materials and Methods

Pollen Collection

Pollen of castor bean (*Ricinus communis* L.) was collected from the plants grown at the campus of the Faculty of Agriculture, Tarbiat Modares University, Tehran, Iran. Pollen of date palm (*Phoenix*

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dactylifera L.) was collected from trees planted in Bandar-Abbas, Hormozgan province, Iran. Almond pollen (*Prunus amygdalus* Batsch) was collected from trees in Shiraz city, Fars province, Iran. The pollen grains were oven-dried at 25°C for 24 h and then frozen at -18°C for long-term storage or refrigerated at 4°C for up to 2 wk for early use in the experiments. For preparing the mixed pollen diet, three mentioned pollen grains were mixed in equal volume ratios.

Stock culture of N. cucumeris

To establish the stock culture of *N. cucumeris*, a package contains individuals of this predator was received from Bio-Planet, Italy. To constitute the laboratory colony of *N. cucumeris*, the individuals were transferred to a Plexiglas box $(30 \times 15 \times 12 \text{ cm})$ half-filled with water with a water-soaked sponge inside and a green plastic sheet $(16 \times 11 \times 0.1 \text{ cm})$ on it. All plastic sheet edges were covered using moist tissue paper which acted as a barrier prohibiting mites escaping (Walzer and Schausberger 1999). The stock culture of *N. cucumeris* was kept in a growth chamber at $25 \pm 1^{\circ}$ C, $60 \pm 5\%$ RH, and a photoperiod of 16:8 (L:D) h. The fresh pollen was offered at 4-day intervals as food over 20 generations.

Experimental Setup

Life Table Construction

The experimental units were quite similar to the stock culture rearing unit, but their size was smaller. In other words, 3-cm-diameter rearing units consisted of green plastic sheets $(3 \times 3 \times 0.1 \text{ cm})$, plastic trays $(7 \times 5 \times 4 \text{ cm})$, and wet sponges were used for designing the experimental units. Some cotton fibers were stuck on the plastic sheet to provide a suitable place for oviposition. To supply the moisture needed for the mites, the edges of the sheet were covered with moist tissue paper, and water was added daily to prevent the strips from drying out.

To have the same-aged eggs of N. cucumeris, more than 30 pairs of the predator were selected randomly from the stock culture and kept on the new experimental units for less than 24 h. The eggs laid by females were transferred to the experimental units individually. After larval emergence, the pollen grains were offered as a food source. The presence of exuvial as moulting evidence was used for determining the duration of each life stage of N. cucumeris. Immature development and survival were recorded daily until they reached adulthood. After adult emergence, females were coupled with the males of the same treatment. In total, 40 replicates per treatment were prepared and checked daily. Daily monitoring was continued until the adults' death. Oviposition, survival, adult longevity, and oviposition periods of the mites were recorded. The predators were reared on the respective pollen diet (date palm, castor bean, and the mixture of pollen types) for 20 generations. For each pollen diet, three generations, including the first generation (G1), 10th generation (G10), and 20th generation (G20) were evaluated. All experiments were conducted in the laboratory condition at $25 \pm 1^{\circ}$ C, $60 \pm 5\%$ RH, and a photoperiod of 16:8 (L:D) h. Fresh pollen was added at 4-day intervals after removing the old pollen in all treatments and replicates. In order to compare the performance of N. cucumeris on pollen diets and the main prey T. urticae, we used the data of 20 generations on this prey for multiple comparisons that was originated from Yazdanpanah et al. (2022).

Data Analysis

The values of all life table parameters, including the net reproductive rate (R_0), gross reproductive rate (*GRR*), finite rate of increase (λ),

intrinsic rate of increase (*r*), and mean generation time (*T*), as well as age-stage-specific survival rate (s_{xi}), age-specific survival rate (l_x) (the probability that a newborn will survive to age *x*, calculated by pooling all of the surviving individuals of different stages), age-stagespecific fecundity (f_{xi}), and age-specific fecundity (m_x) were calculated according to the age-stage, two-sex life table procedure (Chi and Liu 1985, Chi 1988) using the TWOSEX-MSChart software (Chi 2021) at different generations on different diets. The variances and standard errors of all life table parameters were estimated using the bootstrap procedure with 100,000 samples. Multiple comparisons among different generations of each diet and the corresponding generation among different diets were carried out using the paired bootstrap test (Reddy and Chi 2015, Bahari et al. 2018).

Results

Effects of Pollen Diets on Life Table Parameters of *N. cucumeris*

The deutonymphal durations did not differ among generations on all diets except for castor bean pollen. Overall, no significant difference in the total pre-adult duration was observed between G1 and G20, whereas the shortest developmental time was recorded in G10 when pollens were offered separately. Furthermore, when the mixed pollen was a diet, the developmental time was longest in G20, while there was no significant difference between G1 and G10 (Table 1). Adult pre-oviposition period (APOP) of females on castor bean and mixed diet did not differ among three generations, whereas it was longest in G20 on date palm pollen. When the date palm pollen was the food source, the total pre-oviposition period (TPOP) was longest in G20 and shortest in G10, while on the castor bean pollen, this parameter was not affected by the generation. The oviposition period and fecundity of N. cucumeris did not differ among generations on all diets tested, except for date palm pollen. The mentioned parameters in G10 were lower than those in G1 and G20 on date palm pollen. Male longevity had no significant difference among tested generations on all diets except for castor bean pollen which males in G10 had shorter longevity than G10 and G20. Female longevity and the total life span (from birth to death) of N. cucumeris were not significantly different among different generations. When date palm pollen was offered, the female longevity and total life span in G20 was longer than G10, while on the mixture of pollen grains, the mentioned parameters were not different between G10 and G20. Whereas the immature survival of N. cucmeris did not differ among three generations on all diets; survival in all tested generations was 100% when the mixed diet was offered (Table 1).

Age-stage survival rate (s_{ri}) of *N. cucumeris* fed in the first, 10th, and 20th generation on different diets are shown in Fig. 1. The adult stage began around the age of 6, 5, and 6 d in G1, G10, and G20, respectively, when the diet was date palm pollen. Furthermore, females of the first generation and males of the 20th generation lived more (Fig. 1). The adult stage began around the age of 5, 4, and 6 d on castor bean pollen in G1, G10, and G20, respectively. In addition, females in G1 and G20 lived more, whereas males of G10 had the shortest lifetime (Fig. 1). The beginning of the adult stage on the mixed diet was around the age of 4, 6, and 6 d in G1, G10, and G20, respectively, and females in G10 lived more (Fig. 1). Based on the fecundity curves, the peak of fecundity when date palm pollen was offered was observed in G10 (2.32 eggs/female) at the age of 10 d, followed by G20 at the age of 13 d (2.17 eggs/female) (Fig. 2). It was highest in G10 (1.77 eggs) at the age of 30 d, followed by G20 (1.72 eggs) at the age of 20 d when castor bean was offered, while in G1 (2.85 eggs) at the age of 18 d on the mixed diet (Fig. 2).

		Date palm			Castor bean			Mixed pollen		- ·	Tetranychus urticae ^a	<i>b</i> ¢
	61	G10	G20	61	G10	G20	G1	G10	G20	G1	G10	G20
Egg	2.39 ± 0.121^{aA}	$1.52 \pm 0.149^{\text{bB}}$	2.190 ± 0.145^{aA}	2.12 ± 0.124^{aAB}	$1.48 \pm 0.161^{\text{bB}}$	$1.74 \pm 0.146^{\text{bB}}$	$1.64 \pm 0.217^{\text{bB}}$	$2.05 \pm 0.158^{\rm bA}$	2.45 ± 0.107^{aA}	2.42 ± 0.116^{A}	$2.10 \pm 0.177^{\text{A}}$	$2.38 \pm 0.142^{\text{A}}$
Larva	1 ± 0.00^{aA}	1 ± 0.00^{aA}	1 ± 0.00^{aA}	1 ± 0.00^{aA}	1 ± 0.00^{aA}	1 ± 0.00^{aA}	1 ± 0.000^{aA}	1 ± 0.000^{aA}	1 ± 0.000^{aA}	$1 \pm 0.000^{\text{A}}$	$1.47 \pm 0.462^{\text{A}}$	1 ± 0.000^{A}
Protonymph	$1.88 \pm 0.057^{\rm bC}$	$1.96 \pm 0.042^{\text{bB}}$	2.23 ± 0.093^{aA}	2.37 ± 0.076^{aA}	1.91 ± 0.06^{bB}	2.16 ± 0.083^{aA}	2.00 ± 0.000^{aB}	1.91 ± 0.092^{aB}	2.13 ± 0.073^{aA}	2.10 ± 0.064^{B}	$2.32 \pm 0.131^{\text{A}}$	$1.83 \pm 0.100^{\text{B}}$
Deutonymph	1.94 ± 0.087^{aA}	1.82 ± 0.104^{aA}	2.05 ± 0.082^{aA}	$1.65 \pm 0.083^{\text{bB}}$	1.91 ± 0.089^{aA}	1.89 ± 0.103^{abAB}	1.86 ± 0.094^{aAB}	2.00 ± 0.067^{aA}	1.82 ± 0.083^{aB}	$1.7 \pm 0.103^{\mathrm{AB}}$	$2.17\pm0.181^{\rm A}$	1.86 ± 0.097^{AB}
Pre-adult	7.19 ± 0.121^{aA}	$6.27 \pm 0.159^{\text{bB}}$	7.47 ± 0.160^{aA}	7.08 ± 0.134^{aA}	$6.23 \pm 0.192^{\text{bB}}$	6.79 ± 0.159^{ab}	$6.50 \pm 0.242^{\text{bB}}$	$6.95 \pm 0.158^{\rm bA}$	7.41 ± 0.105^{aA}	$7.25 \pm 0.157^{\rm A}$	$7.5 \pm 0.240^{\text{A}}$	7.09 ± 0.191^{AB}
APOP	$2.466 \pm 0.162^{\text{bB}}$	$2.369 \pm 0.111^{\text{bB}}$	2.94 ± 0.177^{aA}	2.68 ± 0.200^{aAB}	2.79 ± 0.176^{aA}	2.36 ± 0.165^{aB}	2.22 ± 0.215^{aB}	2.36 ± 0.194^{aAB}	2.67 ± 0.278^{aAB}	$3.4 \pm 0.424^{\text{A}}$	2.75 ± 0.446^{AB}	2.62 ± 0.208^{AB}
TPOP	$9.73 \pm 0.244^{\text{bA}}$	8.63 ± 0.238^{cB}	10.53 ± 0.190^{aA}	9.82 ± 0.296^{aA}	9.05 ± 0.338^{aAB}	9.07 ± 0.279^{aC}	$8.89 \pm 0.340^{\text{bB}}$	9.50 ± 0.335^{abA}	10.00 ± 0.241^{aAB}	$10.8 \pm 0.506^{\rm A}$	$10.25 \pm 0.637^{\rm A}$	$9.69 \pm 0.171^{\rm BC}$
Oviposition	$20.67 \pm 1.790^{\mathrm{aBC}}$	20.67 ± 1.790^{aBC} 15.79 ± 1.06^{bB}	21.88 ± 0.946^{aA}	27.92 ± 1.892^{aA}	25.53 ± 2.173^{aA}	23.64 ± 1.657^{aA}	$23.55 \pm 3.125^{\text{aAB}}$	27.57 ± 0.993^{aA}	23.84 ± 2.382^{aA}	$15.2 \pm 2.390^{\circ}$	$9.75 \pm 1.797^{\circ}$	$12.07\pm1.60^{\rm B}$
days												
Male longevity	45.41 ± 3.391^{aB}	44.46 ± 9.952^{aAB}	Male longevity $45.41 \pm 3.391^{a B}$ $44.46 \pm 9.952^{a A B}$ $52.34 \pm 8.586^{a A}$ $64.94 \pm 5.733^{a A}$	64.94 ± 5.733^{aA}	20.17 ± 5.586^{bC}	70.18 ± 14.557^{aA}		50.83 ± 12.967^{aAB} 55.15 ± 8.159^{aA}	51.42 ± 5.655^{aA}	$22.23 \pm 4.95^{\circ}$	25.51 ± 2.594^{BC} 26.33 ± 3.064^{B}	$26.33 \pm 3.064^{\text{B}}$
Female lon-	48.4 ± 5.861^{abA}	$39.10 \pm 3.649^{\text{bB}}$	$48.4 \pm 5.861^{\rm abA} 39.10 \pm 3.649^{\rm bB} 54.64 \pm 3.041^{\rm aA} 56.69 \pm 4.974^{\rm aA}$	56.69 ± 4.974^{aA}	59.96 ± 4.547^{aA}	63.37 ± 6.229^{aA}	$48.86 \pm 7.634^{\rm bA}$	72.46 ± 6.976^{aA}	54.01 ± 6.835^{abA}	28.70 ± 3.396^{B}	$25 \pm 2.567^{\circ}$	$25.22 \pm 2.485^{\text{B}}$
gevity												
Total life span	$54.0 \pm 3.213^{ab B}$	$46.36 \pm 3.230^{\text{bB}}$	$54.0 \pm 3.213^{ab B}$ $46.36 \pm 3.230^{b B}$ $61.80 \pm 2.728^{a A}$ $67.48 \pm 3.802^{a A}$	67.48 ± 3.802^{aA}	60.89 ± 4.921^{aA}	71.76 ± 5.540^{aA}	$56.08 \pm 6.439^{\text{bA AB}}$	73.64 ± 5.594^{aA}	60.24 ± 4.404^{abA} 27.46 $\pm 2.862^{C}$	$27.46 \pm 2.862^{\circ}$	$25.17 \pm 1.915^{\circ}$	$25.67 \pm 1.885^{\text{B}}$
Fecundity	32.88 ± 2.411^{aAB}	32.88 ± 2.411^{aAB} 25.37 ± 1.831^{bC}	$36.40 \pm 1.690^{\mathrm{aB}}$	38.20 ± 2.910^{aA}	43.32 ± 3.766^{aB}	40.35 ± 4.114^{aAB}	46.54 ± 6.538^{aA}	55.79 ± 1.856^{aA}	47.58 ± 4.784^{aA}	27.57 ± 4.366^{B}	$9.93 \pm 2.811^{\text{D}}$	22.76 ± 2.897^{C}
Immature sur-	97 ± 0.030^{aB}	96 ± 0.042^{aA}	95 ± 0.044^{aA}	93 ± 0.039^{aAB}	95 ± 0.042^{aA}	100 ± 0.000^{aA}	100 ± 0.000^{aA}	100 ± 0.000^{aA}	100 ± 0.000^{aA}	$83 \pm 0.076^{\circ}$	85 ± 0.076^{A}	91 ± 0.056^{A}
vival (%)												

Table 1. Long-term effects of feeding on date palm pollen, castor bean pollen, and mixture of date palm, castor bean and almond pollen types on duration of different life stages (days), fecundity (eggs/female), and immature survival (%) (mean ± SE) of Neoseiulus cucumeris compared with a natural prey. Tetranychus urticae are comparisons between the MO letters tollowed by different capital The means followed by different lowercase letters in the same row are comparisons among different generations of each diet. The corresponding generations of four diets (P < 0.05, paired-bootstrap).

" For more details on comparisons among tested generations when T. intricae was the diet, please see Yazdanpanah et al. (2022).

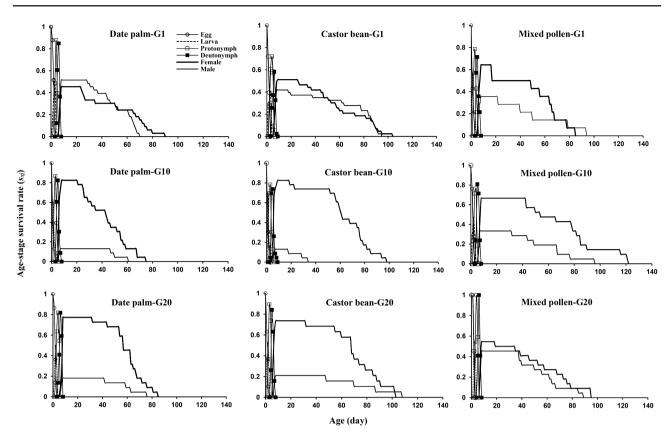


Fig. 1. Age-stage survival rate (sx) of sequential generations (G) of Neoseiulus cucumeris reared on date palm pollen, castor bean pollen, and mixed pollen diet.

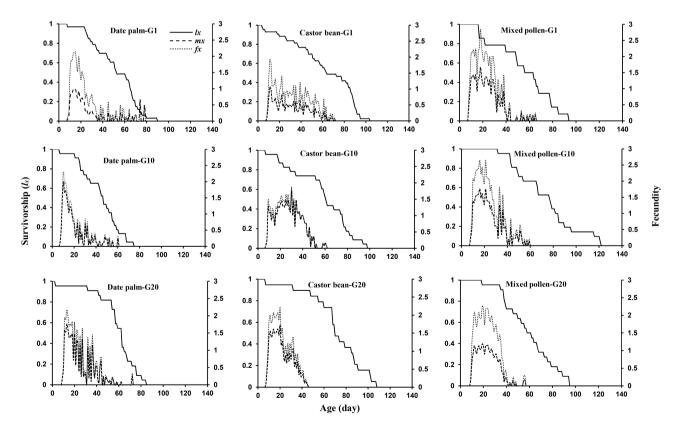


Fig. 2. Age-specific survivorship (I_x) , age-specific fecundity (m_x) , and age-stage-specific fecundity (f_{xj}) of sequential generations (G) of *Neoseiulus cucumeris* reared on date palm pollen, castor bean pollen, and mixed pollen diet.

The highest gross reproductive rate (GRR) and net reproductive rate (R_0) when the food was date palm pollen were observed in G20; however, there was no significant difference between G20 and G10 (Table 2). In addition, the highest values of the intrinsic and finite rate of increase (r and λ , respectively) were observed in G10, whereas there was no significant difference in the mentioned parameters between G1 and G20 (Table 2). When castor bean was the diet, R_0 and r in G10 and G20 were significantly higher than G1. The GRR, R_{o} , and r parameters were not influenced by the generation when the mixed diet was offered. N. cucumeris individuals reared on the mixed diet for one generation had the shortest mean generation time. In contrast, they had no significant difference with those fed on the mentioned diet for 10 generations (Table 2).

Comparing Life Table Parameters of N. cucumeris Reared on Pollen Diets and T. urticae

Although the pre-adult duration of N. cucumeris was shortest in G1 when the mixed diet was offered, developmental time was significantly shorter in G10 when date palm pollen and castor bean pollen were offered compared with both the mixed pollen and the main prey (Table 1). TPOP was shortest on a mixed diet in G1, while it was the fastest in G20 on T. uricae and castor bean pollen. The predator had the shortest oviposition days and lowest fecundity by feeding on the prey in all generations tested; however, no significant difference was observed between predators reared on date palm pollen and the prey in G1 (Table 1). In addition, in all generations tested, the shortest total life span was recorded when the predator individuals fed on T. urticae. In G1, immature survival rate differed significantly among different treatments, while in other generations, there was no significant difference among food diets (Table 1).

Females reread on the prey had the highest GRR in G1; however, it did not differ from those fed on the mixed diet, but the mentioned treatment showed the lowest value in the other generations (Table 2). In G1, R_0 did not differ among treatments, while it was highest on the mixed diet and castor bean pollen in G20. In G1, the females reared on the mixed diet showed the highest growth rate, similar to those raised on the prey. No differences were observed among different pollen diets in terms of the intrinsic and finite rate of increase in G10 and G20. In G1, the females supplied with castor bean pollen had the longest mean generation time than others. In contrast, feeding on castor bean pollen and the mixed diet resulted in the longest T in G10, while it was longest on the mixed diet and date palm pollen in G20 (Table 2).

Discussion

Since pollen from different plant species may differ in their nutritional value and affect the predatory mites' performance (McMurtry and Scriven 1966), the current study investigated the potential of three types of pollen diets for N. cucumeris during 20 generations. Additionally, the results of long-term rearing on these alternative diets were compared to the effects of long-term rearing of the predator on T. urticae as a main prey.

The long-term effects of feeding on pollen grains on a predator's performance must be evaluated if such a diet is used for mass rearing (Bellutti 2011, Sørensen et al. 2012). The results of the present study revealed that N. cucumeris could be reared for 20 generations on castor bean, date palm pollen, and the mixture of pollen types. Similarly, this predator was able to develop and reproduce on almond pollen for 50 generations (Yazdanpanah et al. 2021b) and

For more details on comparisons among tested generations when T. *urticae* was the diet, please see Yazdanpanah et al. (2022)

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Table 2. Life table parameters (mean ± SE) of *Neoseiulus cucumeris* reared on date palm pollen, castor bean pollen, and mixture of date palm, castor bean and almond pollen types for

den-

20

		Date parti			Castor bean			Mixed pollen		L	Tetranychusurticae ^a	
Parameter	G1	G10	G20	G1	G10	G20	G1	G10	G20	G1	G10	G20
GRR (eggs/ indi-	$^{\prime}$ 18.12 ± 3.984 ^{bB}	24.10 ± 2.793^{abb}	$\frac{1}{\text{GRR}} \left(\frac{1}{\text{eggs}} 18.12 \pm 3.984^{\text{fb}} 24.10 \pm 2.793^{\text{ab}} 30.35 \pm 3.477^{\text{ah}} 23.85 \pm 3.865^{\text{bb}} 43.21 \pm 3.419^{\text{ah}} 33.94 \pm 5.00^{\text{ab}} 35.08 \pm 7.838^{\text{a}} 8.24 \pm 5.887^{\text{ah}} 27.18 \pm 5.805^{\text{ab}} 45.70 \pm 8.960^{\text{b}} \text{ab} 10^{\text{ab}} 18.12 \pm 3.410^{\text{ab}} 35.08 \pm 7.838^{\text{ab}} 38.24 \pm 5.887^{\text{ab}} 27.18 \pm 5.805^{\text{ab}} 45.70 \pm 8.960^{\text{ab}} \text{ab} 10^{\text{ab}} 18.12 \pm 3.410^{\text{ab}} 18.12 \pm 3.410^{\text{ab}} 18.12 \pm 3.410^{\text{ab}} 18.12 \pm 3.410^{\text{ab}} 18.12 \pm 5.805^{\text{ab}} 18.12 \pm 5.12^{\text{ab}} 18.12 \pm 5.805^{\text{ab}} 18.12 \pm 5.12^{\text{ab}} 18.12 \pm 5.12^{ab$	23.85 ± 3.865^{bb}	43.21 ± 3.419^{aA}	33.94 ± 5.00^{abA}	35.08 ± 7.838^{aAB}	38.24 ± 5.887^{aA}	27.18 ± 5.805^{aAB}	$45.70 \pm 8.960^{\text{A}}$	8.87 ± 2.772 ^c	$8.87 \pm 2.772^{\text{C}}$ $18.87 \pm 4.327^{\text{B}}$
R_0 (eggs/ indi-	14.93 ± 3.039^{bA}	20.96 ± 2.513^{abB}	vidual) $ \begin{array}{lllllllllllllllllllllllllllllllllll$	$19.55 \pm 3.267^{\rm bh}$	35.79 ± 4.626ª ^A	35.79 ± 4.626^{4A} 31.86 ± 4.962^{ab}	29.92 ± 7.237^{aA}	37.19 ± 5.854^{aA}	25.98 ± 5.659^{aA}	$18.38 \pm 3.910^{\text{A}}$	$6.15 \pm 2.014^{\circ}$	$6.15 \pm 2.014^{\circ}$ 12.34 $\pm 2.774^{\circ}$
$r (day^{-1})$		0.208 ± 0.010^{aA}	$0.152 \pm 0.013^{\text{hb}} 0.208 \pm 0.010^{\text{ah}} 0.180 \pm 0.008^{\text{hb}} 0.146 \pm 0.011^{\text{hb}}$	$0.146 \pm 0.011^{\text{bB}}$	0.186 ± 0.009^{aA}	0.200 ± 0.010^{aA}	$0.186 \pm 0.009^{\rm aA} 0.200 \pm 0.010^{\rm aA} 0.197 \pm 0.017^{\rm aA}$	0.199 ± 0.012^{aA}	$0.199 \pm 0.012^{ah} 0.167 \pm 0.014^{ahB} 0.164 \pm 0.011^{AB} 0.109 \pm 0.023^{B} 0.158 \pm 0.015^{B}$	0.164 ± 0.011^{AB}	0.109 ± 0.023^{B}	$0.158 \pm 0.015^{\text{B}}$
۸ (day ⁻) T (day)	1.165 ± 0.013^{22} 17.62 ± 0.473^{aB}	17.62 ± 0.473^{ab} 14.61 ± 0.349 ^{bb} 18.46 ± 0.332 ^{ab}	1.198 ± 0.010^{m} 18.46 ± 0.332^{aA}	20.29 ± 0.637^{aA}	1.203 ± 0.011^{-1} 19.16 ± 0.586^{aA}	1.222 ± 0.012^{m} 17.25 ± 0.515^{bb}	1.218 ± 0.021^{m} 17.13 ± 0.715^{bB}	1.220 ± 0.014^{m} 18.13 ± 0.419^{abA}	$1.10^{-4} \times 1.10^{-4} \times 1.10^{-4} \times 1.12^{-4} \times 1.12^{-4} \times 1.12^{-4} \times 1.21^{-4} \times 1.12^{-4} \times 1.12^{-4} \times 1.10^{-4} \times 1.10$	$1.1/8 \pm 0.014^{10}$ 17.58 ± 0.659^{B}	$1.116 \pm 0.026^{\circ}$ $16.04 \pm 0.699^{\circ}$	$1.1/2 \pm 0.01/2$ $15.69 \pm 0.420^{\circ}$

cattail pollen for 25 generations (Gravandian et al. 2022). Successful long-term rearing on pollen grains for other phytoseiid mites such as *Neoseiulus californicus* (McGregor) (Acari: Phytoseiidae) (Khanamani et al. 2017) and *Amblyseius swirskii* Athias-Henriot (Acari: Phytoseiidae) (Nemati and Riahi 2020) have been reported beyond 20 and 6 generations, respectively when fed on almond pollen.

Our results showed that the performance of *N. cucumeris* had no reduction during long-term feeding on date palm pollen. In addition, the highest value of *r* in G10 showed the most adaptation of the predator after ten generations of rearing on date palm pollen, which this value (0.208 day⁻¹) was close to the value obtained on tomato russet mite *Aculops lycopersici* (Massee) (Acari: Eriophyideae) (0.211 day⁻¹) (Al-Azzazy and Alhewairini 2018). Date palm pollen has been recommended as a favorable diet for mass rearing of other phytoseiid mites such as *A. swirskii* (Riahi et al. 2017), and *Typhlodromus bagdasarjani* Wainstein & Arutunjan (Acari: Phytoseiidae) (Riahi et al. 2016).

Although some researchers indicated low quality or even toxicity of castor bean pollen for phytoseiid species (Ranabhat et al. 2014, Khanamani et al. 2016), we found that not only this pollen is not toxic for this predator, but also it could support its long-term rearing at least for 20 generations. Actually, the nutritional quality of pollen which can be influenced by environmental conditions, method of pollen preparation (Bogdanov 2006, Dabija 2010), plant genotype (Linskens and Pfahler 1977), and plant age (Brodbeck et al. 2001) may be a reason for different performance of predators on them. The high value of R_0 and r in G10 and G20 indicated that the castor bean pollen could be a suitable diet for the long-term rearing of this predator. Similarly, the suitability of this pollen for N. cucumeris (van Rijn and Tanigoshi 1999, Yazdanpanah et al. 2021a), Amblyseius zaheri Yousef and El-Borolossy (Acari: Phytoseiidae), and Euseius yousefi Zaher & El-Borolossy (Acari: Phytoseiidae) (Momen 2004) has been recorded previously. In addition, the value of the intrinsic rate of increase on castor bean in G1 (0.146 day-1) was close to the value of this parameter by feeding on apple pollen (0.149 day⁻¹), Christmas cactus (0.156 day⁻¹) (Ranabhat et al. 2014), and T. urticae (0.131 day⁻¹) (Al-Azzazy et al. 2018).

Some important parameters such as total life span, oviposition days, fecundity, R_0 , and r had no significant difference among all tested generations by feeding on the mixed pollen, which means that this diet is a suitable food for long-term rearing of *N. cucumeris*. The estimated values of r in the current study at the first generation were higher than the value of r when the predator fed on cattail pollen (0.120 day⁻¹) (Gravandian et al. 2022) or fed on almond pollen (0.128 day⁻¹) (Yazdanpanah et al. 2021b) in G1.

Based on Yazdanpanah et al. (2022), long-term feeding on *T. urticae* caused no decline in *N. cucumeris* performance, and its predation potential remained high; therefore, *T. urticae* can be one of its main prey species. Comparison of the life table parameters between the reared predators on the pollen diets and natural prey (*T. urticae*) during 20 generations revealed that although pre-adult duration, especially incubation and larval periods were affected by maternal diets, developmental time in G20 had no significant difference in all pollen diets versus prey (*T. urticae*) diet. Consequently, long-term rearing and adaptation to the diet have a similar effect on the pre-adult period in all diets tested, including alternative or main food.

The values of R_0 , r, and λ had no significant difference among all tested diets at the first generation, but the values of the fecundity, oviposition days, and total life span were the lowest in all tested generations by feeding on *T. urticae*, while all mentioned parameters were

the highest when the predator fed on castor bean or mixed pollen diets (especially in the later generations). This may be due to prey movement and the time spent subduing a prey individual (Hassell 1978, Yazdanpanah et al. 2022). Actually, pollen grains as immotile food sources are more available than prey, conserving energy for the rest of the predator's life. Although *N. cucumeris* was successfully reared on *T. urticae* for up to 30 generations (Yazdanpanah et al. 2022), rearing the phytoseiid mites on natural prey gives rise to high labor costs.

In conclusion, mass rearing of *N. cucumeris* on castor bean, date palm, and mixed pollen (castor bean + date palm + almond) did not decrease its performance during 20 generations of rearing. Furthermore, mites reared on pollen diets had higher quality than those reared on natural prey (*T. urticae*); therefore, it can be concluded that castor bean or date palm lonely, and mixed pollen are good candidates for the mass rearing of *N. cucumeris* for use in augmentative biological control programs.

Author Contributions

S.Y.: conceptualization; investigation; data curation; formal analysis; methodology; writing original draft. Y.F.: project administration; funding acquisition; supervision; methodology, validation; review and edit. E.R: methodology; validation; review and editing. M.P.Z.: validation; methodology; review and editing.

Acknowledgments

This research was partly supported by a grant (4003128) from Iran National Science Foundation (INSF) and partly by Tarbiat Modares University, Tehran, Iran, which is greatly appreciated.

References Cited

- Al-Azzazy, M., and S. Alhewairini. 2018. Relationship between temperature and developmental rate of tomato russet mite Aculops lycopersici (Massee) (Acari: Eriophyideae) on tomato. JFAE. 16: 18–23.
- Bahari, F., Y. Fathipour, A. A. Talebi, and Z. Alipour. 2018. Long-term feeding on greenhouse cucumber affects life table parameters of two-spotted spider mite and its predator *Phytoseiulus persimilis*. Sys. Appl. Acarol. 23: 2304–2316.
- Bellutti, N. 2011. Effects of mass rearing on life-history traits of an invasive fruit moth species, Grapholita molesta (Busck). Master Thesis, Institute of Forest Entomology, Forest Pathology and Forest Protection, BOKU Vienna, p. 35.
- Bogdanov, S. 2006. Contaminants of bee products. Apidologie. 37: 1-18.
- Brodbeck, B. V., J. Stavisky, J. E. Funderburk, P. C. Andersen, and S. M. Olson. 2001. Flower nitrogen status and population of *Frankliniella occidentalis* feeding on *Lycopersicon esculentum*. *Entomol. Exp. Appl.* 99: 165–172.
- Chi, H. 1988. Life-table analysis incorporating both sexes and variable development rate among individuals. *Environ. Entomol.* 17: 26–34.
- Chi, H. 2021. TWO SEX-MSChart: a computer program for the age-stage, two-sex life table analysis. Available: http://140.120.197.173/ecology/
- Chi, H., and H. Liu. 1985. Two new methods for the study of insect population ecology. Bull. Inst. Zool. Academia Sinica. 24: 225–240.
- Cônsoli, F., J. R. Parra, and R. Zucchi. 2010. Egg parasitoids in agroecosystems with emphasis on Trichogramma. Springer Netherlands, Dordrecht.
- Dabija, T. 2010. Study of amino acids in pollen's composition. Bull UASVM Anim Biol. 67: 1–2.
- Delisle, J. F., J. Brodeur, and L. Shipp. 2015. Evaluation of various types of supplemental food for two species of predatory mites, *Amblyseius swirskii* and *Neoseiulus cucumeris* (Acari: Phytoseiidae). *Exp. Appl. Acarol.* 65: 483–494.
- Gravandian, M., Y. Fathipour, H. Hajiqanbar, E. Riahi, and E. W. Riddick. 2022. Long-term effects of cattail *Typha latifolia* pollen on development,

reproduction, and predation capacity of *Neoseiulus cucumeris*, a predator of *Tetranychus urticae*. *BioControl*. 67: 149–160.

- Hassell, M. P. 1978. The dynamics of arthropod predator-prey systems. Princeton University Press, Princeton, NJ, 248 pp.
- Khanamani, M., Y. Fathipour, A.A. Talebi, and M. Mehrabadi. 2016. Linking pollen quality and performance of *Neoseiulus californicus* (Acari: Phytoseiidae) in two-spotted spider mite management programmes. *Pest Manag. Sci.* 73: 452–461.
- Khanamani, M., Y. Fathipour, A. A. Talebi, and M. Mehrabadi. 2017. Quantitative analysis of long-term mass rearing of *Neoseiulus californicus* (Acari: Phytoseiidae) on almond pollen. J. Econ. Entomol. 110: 1442–1450.
- van Lenteren, J. C. 1991. Success in biological control of arthropods by augmentation of natural enemies, pp. 77–103. In G. Gurr and S. Wratten (eds.), Biological control: measures of success. Springer, Dordrecht, The Netherlands.
- van Lenteren, J. C. 2003. *Quality control and production of biological control agents theory and testing procedures*. CABI, Wallingford, UK.
- van Lenteren, J. C. 2012. IOBC Internet book of biological control, version 6, pp. 93–103. https://www.iobc global.org/publications_iobc_internet_ book_of_biological_control.html. Accessed 17 Sept 2019.
- Linskens, H. F., and P. L. Pfahler. 1977. Genotypic effects on the amino acid relationships in maize (*Zea mays L.*) pollen and style. *Theor. Appl. Genet.* 50: 173–177.
- McMurtry, J. A., and G. T. Scriven. 1966. The influence of pollen and prey density on the numbers of prey consumed by *Amblyseius hibisci* (Acarina: Phytoseiidae). Ann. Entomol. Soc. Am. 59: 149–157.
- McMurtry, J. A., G. J. de Moraes, and N. F. Sourassou. 2013. Revision of the lifestyles of phytoseiid mites (Acari: Phytoseiidae) and implications for biological control strategies. *Syst. Appl. Acarol.* 18: 297–320.
- Momen, F. M. 2004. Suitability of the pollen grains, *Ricinus communis* and *Helianthus annuus*, as food for six species of phytoseiid mites (Acari: Phytoseiidae). *Acta Phytopathol. Entomol. Hung.* 39: 415–422.
- Nemati, A. R., and E. Riahi. 2020. Does feeding on pollen grains affect the performance of *Amblyseius swirskii* (Acari: Phytoseiidae) during subsequent generations? *Bull. Entomol. Res.* 110: 449–456.
- Ranabhat, N. B., I. Goleva, and C. P. W. Zebitz. 2014. Life tables of Neoseiulus cucumeris exclusively fed with seven different pollens. BioControl. 59: 195–203.

- Reddy, G. V., and H. Chi. 2015. Demographic comparison of sweet potato weevil reared on a major host, *Ipomoea batatas*, and an alternative host, I. *triloba. Sci. Reports.* 5: 11871–11879.
- Riahi, E., Y. Fathipour, A. A. Talebi, and M. Mehrabadi. 2016. Pollen quality and predator viability: Life table of *Typhlodromus bagdasarjani* on seven different plant pollens and two-spotted spider mite. *Syst. Appl. Acarol.* 21: 1399–1412.
- Riahi, E., Y. Fathipour, A. A. Talebi, and M. Mehrabadi. 2017. Natural diets vs factitious prey: comparative effects on development, fecundity and life table of *Amblyseius swirskii* (Acari: Phytoseiidae). *Syst. Appl. Acarol.* 22: 711–723.
- van Rijn, P. C., and L. K. Tanigoshi. 1999. Pollen as food for the predatory mites *Iphiseius degenerans* and *Neoseiulus cucumeris* (Acari: Phytoseiidae): dietary range and life history. *Exp. Appl. Acarol* 23: 785–802.
- Sørensen, J. G., M. F. Addison, and J. S. Terblanche. 2012. Mass rearing of insects for pest management: Challenges, synergies and advances from evolutionary physiology. *Crop Protec.* 38: 87–94.
- Walzer, A., and P. Schausberger. 1999. Cannibalism and interspecific predation in the phytoseiid mites *Phytoseiulus persimilis* and *Neoseiulus californicus*: predation rates and effects on reproduction and juvenile development. *BioControl*. 43: 457–468.
- Yazdanpanah, S., Y. Fathipour, and E. Riahi. 2021a. Pollen grains are suitable alternative food for rearing the commercially used predatory mite *Neoseiulus cucumeris* (Acari: Phytoseiidae). Sys. Appl. Acarol. 26: 1009–1020.
- Yazdanpanah, S., Y. Fathipour, E. Riahi, M. P. Zalucki. 2021b. Mass production of *Neoseiulus cucumeris* (Phytoseiidae): an assessment of 50 generations reared on almond pollen. *J. Econ. Entomol.* 114: 2255–2263.
- Yazdanpanah, S., Y. Fathipour, and E. Riahi. 2022. Cost-effective and efficient factitious prey for mass production of *Neoseiulus cucumeris* (Acari: Phytoseiidae): assessing its quality compared with natural prey. Egypt. J. Biol. *Pest Control.* 32: 16.
- Zhang, Y. X., J. Z. Lin, G. Q. Zhang, C. Xia, and J. J. Q. Tang. 2011. Research and application of *Neoseiulus cucumeris* (Oudemans) for control of *Bemisia tabaci* (Gennadius) on sweet pepper in plastic greenhouse. *Fuj. J. Agric. Sci.* 26: 91–97.