

## Supplementary information

### Cancer-cell-secreted miR-204-5p induces leptin signaling pathway in white adipose tissue to promote cancer-associated cachexia

Yong Hu<sup>1,2,5</sup>, Liu Liu<sup>1,5</sup>, Yong Chen<sup>1</sup>, Xiaohui Zhang<sup>1</sup>, Haifeng Zhou<sup>1</sup>, Sheng Hu<sup>1</sup>, Xu Li<sup>1</sup>, Meixin Li<sup>1</sup>, Juanjuan Li<sup>3</sup>, Siyuan Cheng<sup>2</sup>, Yong Liu<sup>1,4</sup>, Yancheng Xu<sup>2\*</sup> and Wei Yan<sup>1\*</sup>

**Supplementary information includes nine Supplementary Figures and four Supplementary Tables.**

**Supplementary Figure 1.** Cancer-derived sEVs induces white adipose tissue fat loss.

**Supplementary Figure 2.** Cancer cell-secreted miR-204 promotes hypoxia signaling by targeting *Vhl*.

**Supplementary Figure 3.** Cancer-secreted miR-204 induces leptin signaling pathway in white adipose tissue.

**Supplementary Figure 4.** Leptin receptor deficiency blocks miR-204 mediated white adipose tissue browning.

**Supplementary Figure 5.** Circulating miR-204 enhances phosphatidylethanolamines production to promote cancer-associated cachexia.

**Supplementary Figure 6.** Exogenous VHL expression suppresses white adipose tissue browning.

**Supplementary Figure 7.** Exosomal miR-204 promotes thermogenesis and lipolysis in LLC-xenografted tumour mice.

**Supplementary Figure 8.** sEV characterization and the effect of miR-204 sEVs on the tumour metastatic.

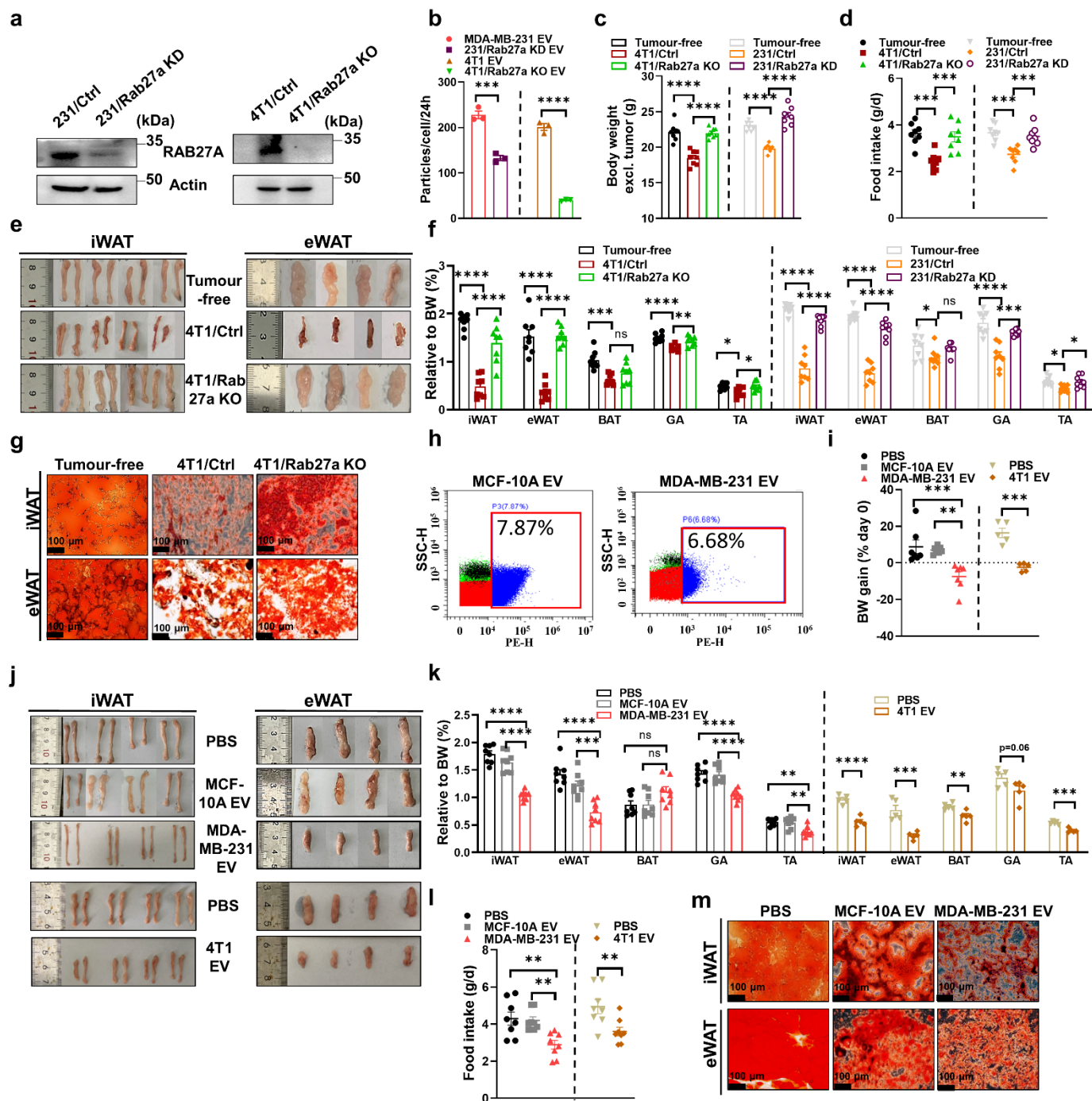
**Supplementary Figure 9.** Nanoflow cytometry gating strategy for MDA-MB-231 sEVs and MCF-10A sEVs.

**Supplementary Table 1.** Predicted miRNA regulators of *VHL*.

**Supplementary Table 2.** Oligonucleotide primers for qRT-PCR analysis.

**Supplementary Table 3.** Antibodies used in this study.

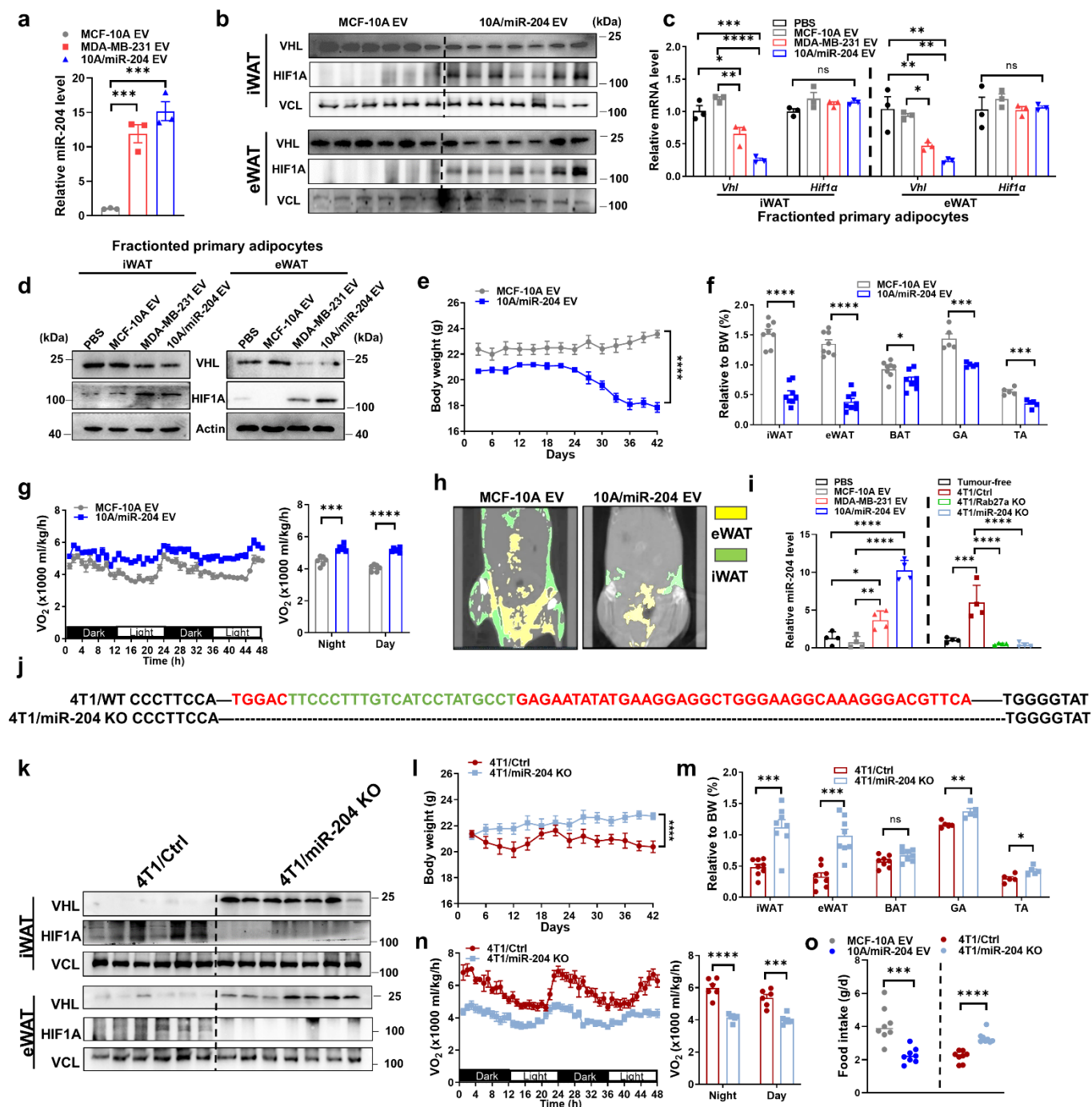
**Supplementary Table 4.** Clinical information included in this study.



**Supplementary Figure 1. Cancer-derived sEVs induces white adipose tissue fat loss.**

**(a)** RAB27A level in MDA-MB-231/Rab27a KD cells and 4T1/Rab27a KO cells. **(b)** The particles of 231/Rab27a KD sEVs and 4T1/Rab27a KO sEVs ( $n=3$  biologically independent samples per group). **(c)** Body weight (calculated by subtracting tumour weight from the total weight) of mice bearing indicated tumours ( $n=8$  mice for Balb/c mice;  $n=7$  mice for NSG mice), results of two independent experiments were combined). For the body weight changes of BALB/c mice ( $n=7$  mice per group):  $11.77 \pm 1.63\%$  (tumour-free group),  $-6.41 \pm 1.36\%$  (4T1/Ctrl group) and  $12.19 \pm 0.48\%$  (4T1/Rab27a KO group); For the body weight change of NSG mice ( $n=5$  per group):  $7.05 \pm 1.16\%$  (tumour-free group),  $-7.57 \pm 1.38\%$  (231/Ctrl group) and  $9.47 \pm 3.52\%$  (231/Rab27a KD group). **(d)** Daily food intake of mice in indicated groups ( $n=8$  mice per group). **(e)** Representative adipose tissue (iWAT, eWAT) images in tumour group mice ( $n=4$  mice in each group). **(f)** Relative tissues weight normalized to BW (body weight).  $n=8$  mice in each group. **(g)** Oil Red O staining in adipose tissues of tumour group mice. Scale bar, 100  $\mu\text{m}$ . **(h)** Nanoflow cytometry showing PE conjugated

CD9 expression on the sEVs membranes. **(i)** Body weight gain after 5 weeks sEV injection. n=7 mice for each group on the left; n=5 mice for each group on the right. **(j)** Representative adipose tissue (iWAT, eWAT) images in sEVs group mice (n=4 mice in each group). **(k)** Relative tissues weight (normalized to body weight). n=8 mice for each group on the left; n=5 mice for each group on the right. **(l)** Daily food intake of mice in indicated groups (n=8 mice per group). **(m)** Oil Red O staining in adipose tissues of indicated groups of mice. Scale bar, 100  $\mu$ m. Oil Red O staining experiments were repeated three times independently with similar results obtained. Data are presented as mean  $\pm$  s.e.m; \*P<0.05, \*\*P<0.01, \*\*\*P<0.001, \*\*\*\*P<0.0001, ns: not significant. One-way ANOVA followed by Dunnett multiple comparison test was used for c, d, f, i, k, l. Unpaired two-tailed t-test was used for b. Source data and exact *P* value are provided as a Source data file.

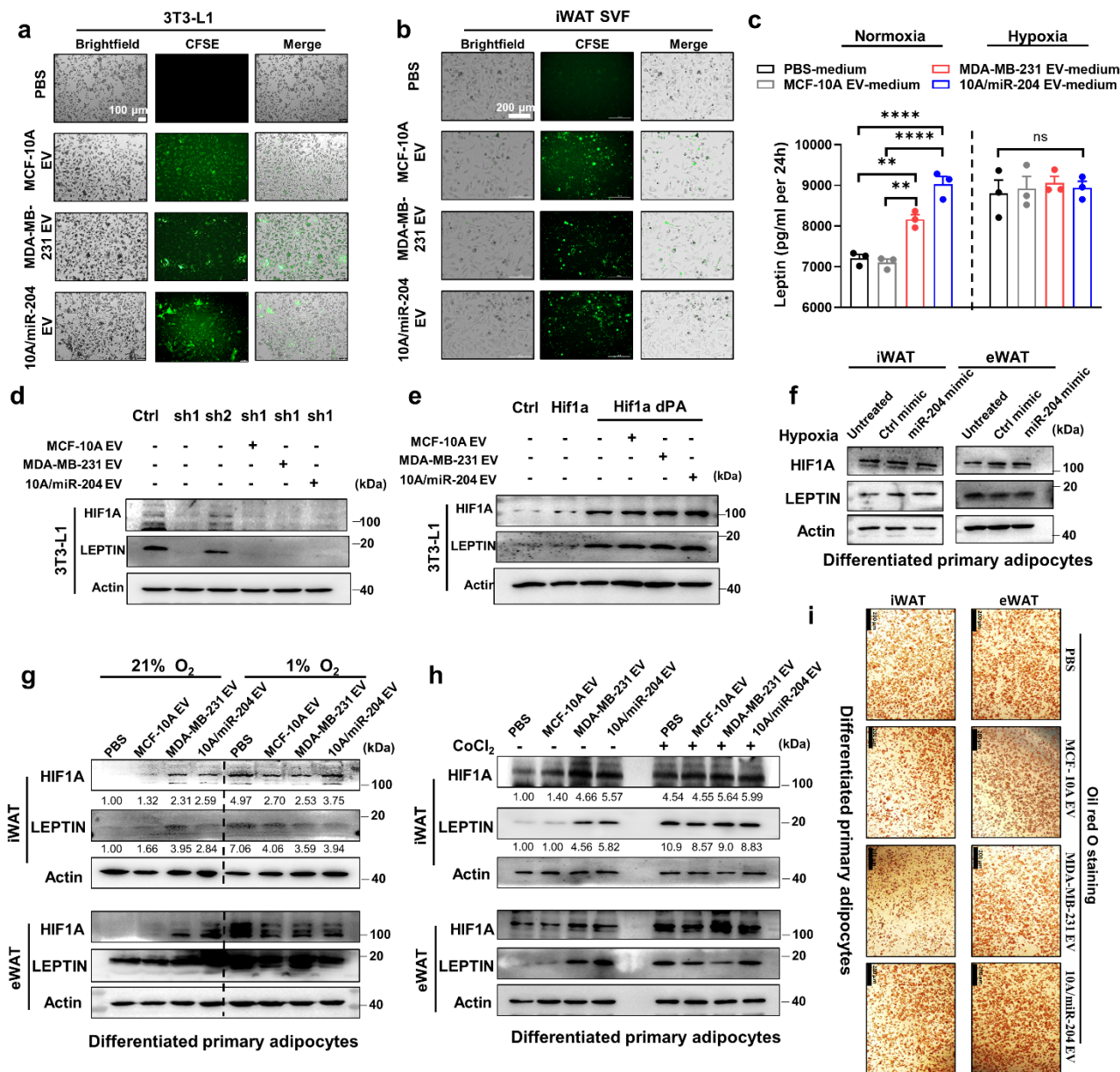


**Supplementary Figure 2. Cancer cell-secreted miR-204 promotes hypoxia signaling by targeting Vhl.**

(a) The expression level of miR-204 in indicated sEVs (n=3 biologically independent samples per group). (b) Western blots showing indicated proteins levels in iWAT and eWAT from sEVs injected NSG mice in indicated groups. (n=6 mice for MCF-10A sEVs; n=7 mice for 10A/miR-204 sEVs). (c) The qRT-PCR analysis of mRNA abundance of *Vhl* and *Hif1a* in fractionated primary adipocytes from each group mice (n=3 biologically independent samples per group). (d) Western blots showing indicated proteins in fractionated primary adipocytes from each group mice. (e) Body weight (n=7 mice per group) and (f) relative eWAT, iWAT, BAT (n=8 mice per group), gastrocnemius (GA) and tibial anterior (TA) skeletal muscle weight (n=5 mice per group) (normalized to body weight) of mice in indicated groups. (g) Oxygen consumption ( $VO_2$ ) in sEV injected mice. Dot plots represent the 48-hour average values (n=6 mice per group). (h) eWAT (yellow) and iWAT (green) distribution in sEV injected NSG mice, as visualized by micro-CT (n=3 mice per group). (i) The qRT-PCR analysis of miR-204 abundance of adipose tissue from each group mice (n=4 mice per group). (j)



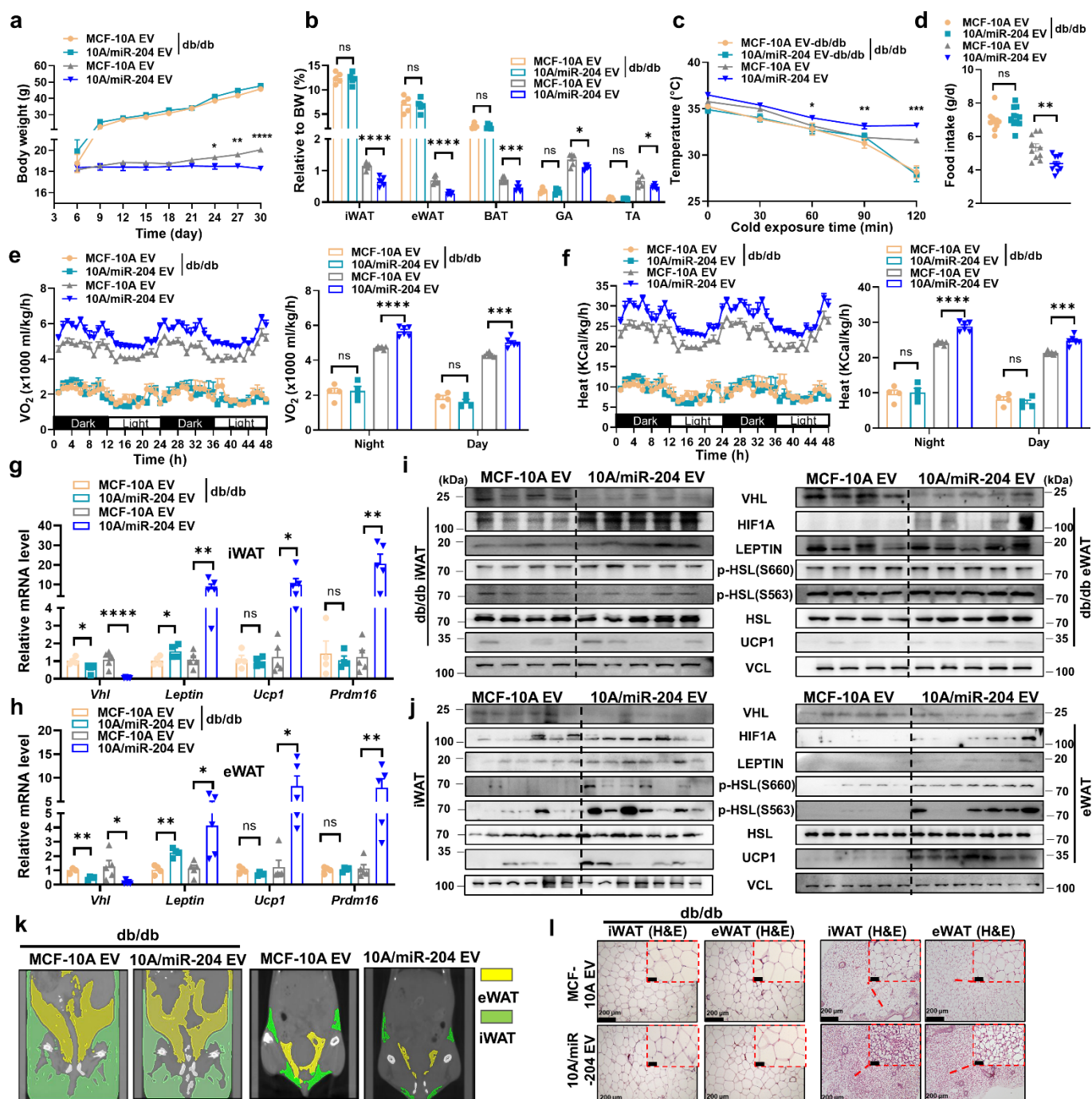
Sanger sequencing showing genetic KO of *mmu*-miR-204 gene in 4T1/miR-204 KO cells. Dashes indicated the deleted region. **(k)** Western blots showing indicated proteins levels in iWAT and eWAT from mice in indicated groups (n=6 mice per group for 4T1/Ctrl; n=7 mice for 4T1/miR-204 KO). **(l-m)** Body weight (n=7 mice per group), iWAT, eWAT, BAT (n=8 mice per group), GA and TA skeletal muscle (n=5 mice per group, normalized to body weight) in each group of mice. **(n)** Oxygen consumption (VO<sub>2</sub>) in each group mice. Dot plots represent the 48-hour average values (n=6 mice per group). **(o)** Daily food intake of mice in indicated groups (n=6 mice per group). Data are presented as mean ± s.e.m; \**P*<0.05, \*\* *P*<0.01, \*\*\**P*<0.001, \*\*\*\**P*<0.0001, ns: not significant. One-way ANOVA followed by Dunnett multiple comparison test was used for a, c, i. Unpaired two-tailed t-test was used for f, g, m, n and o. Two-way ANOVA followed by Bonferroni's multiple comparisons test was used for e, l. Source data and exact *P* value are provided as a Source data file.



**Supplementary Figure 3. Cancer-secreted miR-204 induces leptin signaling pathway in white adipose tissue.**

**(a-b)** MCF-10A sEVs, MDA-MB-231 sEVs and 10A/miR-204 sEVs labelled CFSE (Green) treat with 3T3L-1 adipocytes and iWAT stromal vascular fraction (SVF). **(c)** The levels of leptin from cultured medium for differentiated primary adipocytes of iWAT under normoxia or hypoxia were determined by ELISA kit (n= 3 biologically independent samples per group; one-way ANOVA followed by Dunnett multiple comparison test). **(d)** Protein levels in mature 3T3-L1 treated as indicated and transfected with Ctrl shRNA or shRNA targeting HIF1A under 1% O<sub>2</sub>. Repeated three times independently with similar results obtained. **(e)** Western blots showing indicated proteins in mature 3T3-L1 transfected with WT HIF1A or the dPA mutant with or without indicated sEVs treatment. Repeated three times independently with similar results obtained. **(f)** Western blots showing levels of indicated proteins in differentiated primary adipocytes transfected with miR-204 or control mimic under 1% O<sub>2</sub>. Repeated three times independently with similar results obtained. **(g)** Protein levels in differentiated primary adipocytes from iWAT and eWAT treated as indicated under 21% or 1% O<sub>2</sub>. Repeated three times independently with similar results obtained. **(h)** Western blots showing indicated proteins in differentiated primary adipocytes from iWAT and eWAT treated as indicated with or without 100 mM CoCl<sub>2</sub>.

Repeated three times independently with similar results obtained. **(i)** Oil Red O staining of differentiated primary adipocytes from iWAT and eWAT treated by indicated sEVs. Repeated three times independently with similar results obtained. Scale bar, 200  $\mu$ m. Data are presented as mean  $\pm$  s.e.m; \* $P$ <0.05, \*\*  $P$ <0.01, \*\*\*\* $P$ <0.0001, ns: not significant. Source data and exact  $P$  value are provided as a Source data file.

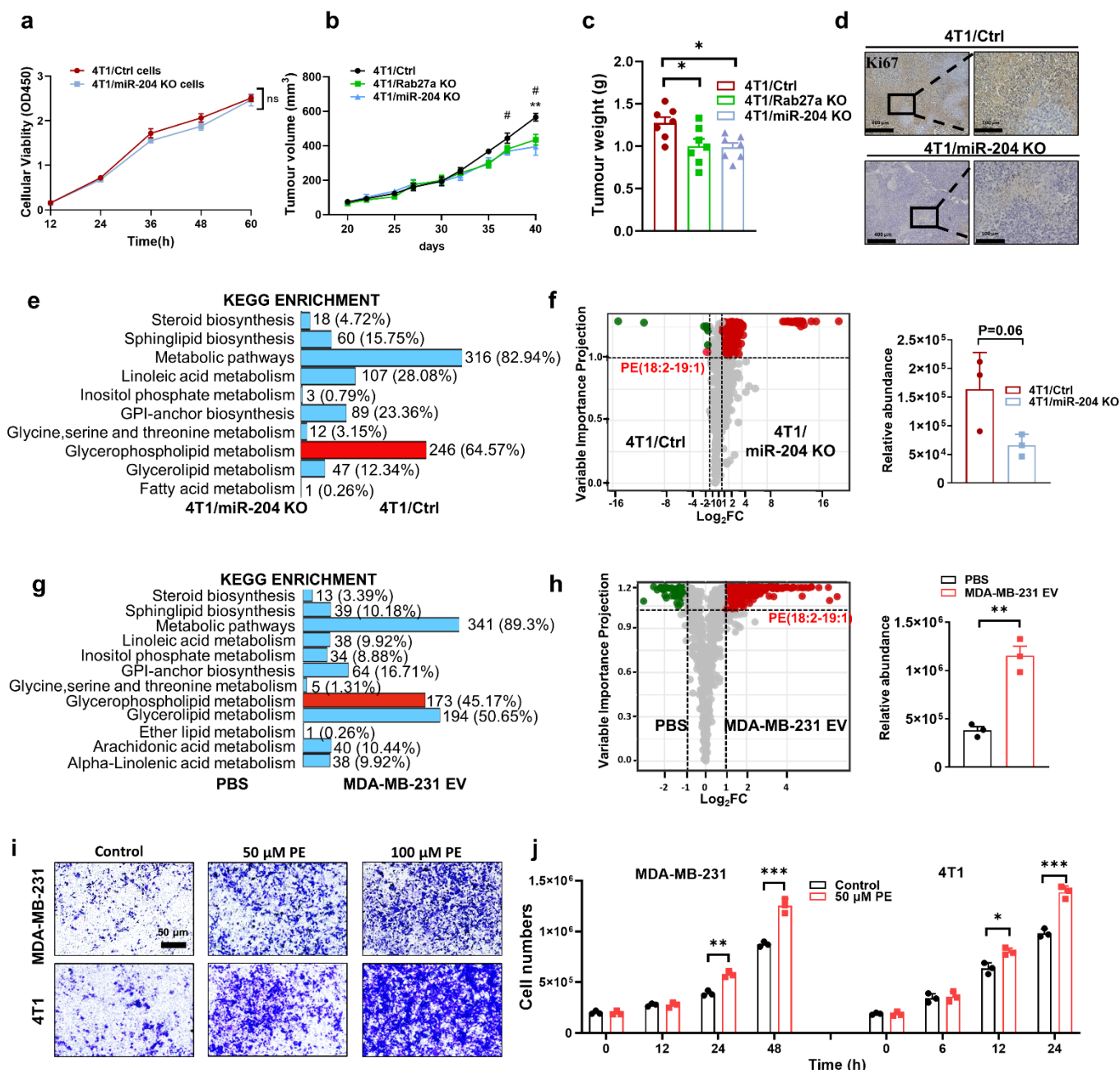


**Supplementary Figure 4. Leptin receptor deficiency blocks miR-204 mediated white adipose tissue browning.**

BKS-Lepr<sup>em2Cd479</sup>/Gpt (BKS-db), C57BL/6J mice were tail vein injected with ~10 μg sEVs per injection per mouse (MCF-10A sEVs, 10A/miR-204 sEVs) twice a week. **(a)** Body weight (n=5 mice for db/db mice; n=6 mice for C57BL/6J mice). **(b)** Relative fat tissues weight (normalized to body weight) (n=5 mice for db/db mice; n=6 mice for C57BL/6J mice). **(c)** Rectal temperatures of the mice at different times after cold exposure (n=3 mice for db/db mice; n=6 mice for C57BL/6J mice). **(d)** Daily food intake (n=5 mice examined by 10 independent experiments for db/db mice; n=6 mice examined by 10 independent experiments for C57BL/6J mice). **(e)** Oxygen consumption (VO<sub>2</sub>) and **(f)** heat production. Dot plots were monitored over a 48-h period in each group mice. n=4 mice for db/db mice; n=6 mice for C57BL/6J mice. **(g-h)** Relative mRNA abundance of the indicated mRNA levels in iWAT and eWAT from each group mice (n=4 mice for db/db mice; n=5 mice for C57BL/6J mice). **(i)** Western blots showing indicated proteins in iWAT and eWAT from indicated groups

of db/db mice (MCF-10A sEVs group, n=4 mice per group; 10A/miR-204 sEVs group, n=5 mice per group). **(j)** Western blots showing indicated proteins in iWAT and eWAT from indicated groups of C57BL/6J mice (MCF-10A sEVs group, n=6 mice per group; 10A/miR-204 sEVs group, n=7 mice per group). **(k)** eWAT (yellow) and iWAT (green) distribution in db/db mice and C57BL/6J mice receiving MCF-10A sEVs or 10A/miR-204 sEVs, as visualized by micro-CT (n=3 mice per group). **(l)** Representative H&E staining with iWAT/eWAT from each group mice. Repeated three times independently with similar results obtained. Scale bar, 200  $\mu$ m. Data are presented as mean  $\pm$  s.e.m; \* $P$ <0.05, \*\* $P$ <0.01, \*\*\* $P$ <0.001, \*\*\*\* $P$ <0.0001, ns: not significant. Unpaired two-tailed t-test was used for a, b, c, d, e, f, g, h. Source data and exact  $P$  value are provided as a Source data file.



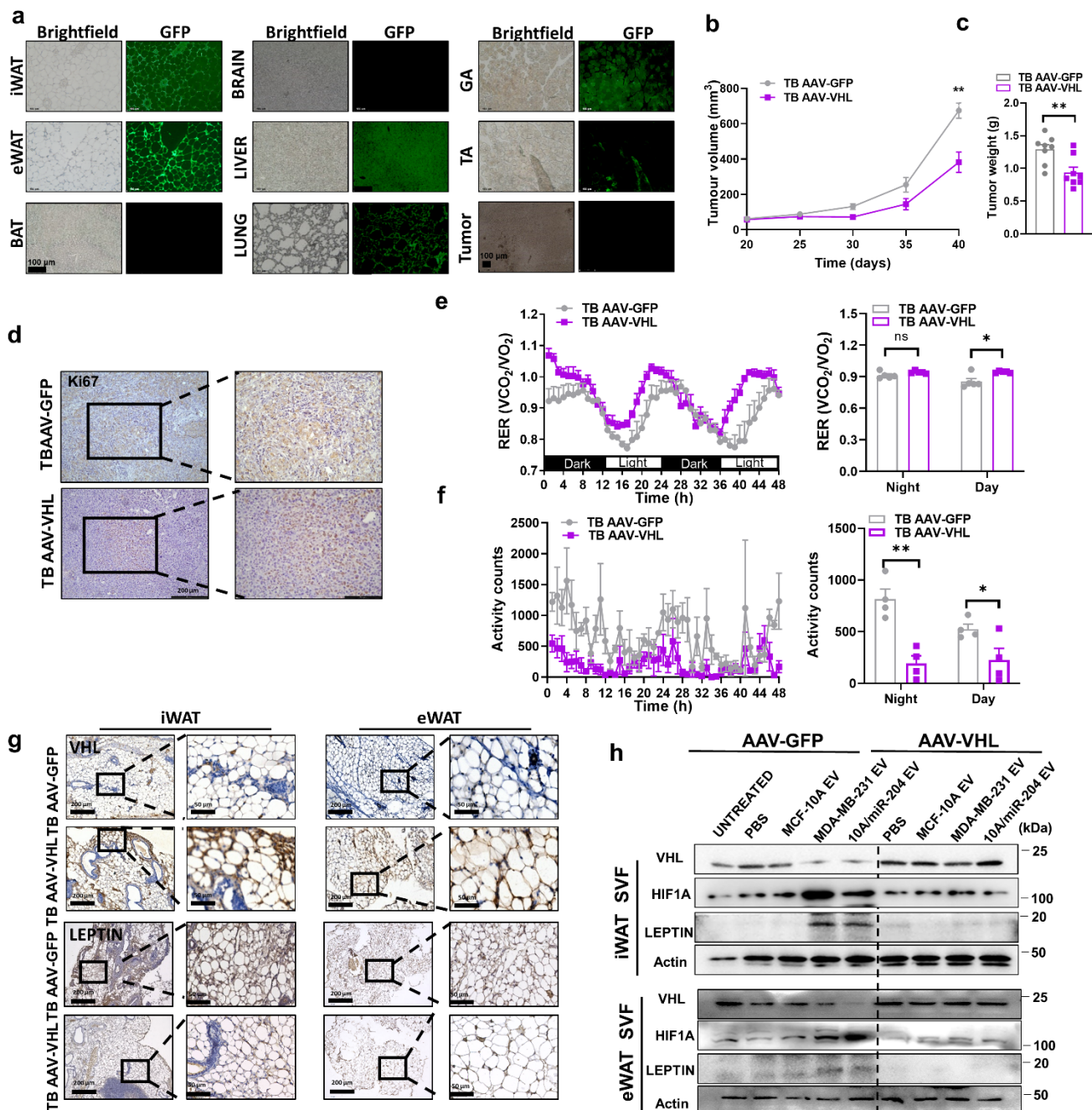


**Supplementary Figure 5. Circulating miR-204 enhances phosphatidylethanolamines production to promote cancer-associated cachexia.**

(a) CCK-8 was used to determine cell viability for 4T1/Ctrl and 4T1/miR-204 KO cells (n=6 biologically independent samples per group). (b) Tumour volumes were measured every 3 days. n=8 mice per group. (c) Tumour weight. n=8 mice per group. (d) Immunohistochemistry staining of Ki67 in 4T1/Ctrl and 4T1/miR-204 KO tumour. Repeated three times independently with similar results obtained. (e) KEGG pathway enrichment of metabolomics showing Glycerophospholipid metabolism and (f) volcano plot depicting levels of phosphatidylethanolamines in serum from 4T1 tumour-bearing mice and 4T1/miR-204 KO mice. n=3 mice per group. (g) KEGG pathway enrichment of metabolomics showing Glycerophospholipid metabolism and (h) volcano plot depicting levels of phosphatidylethanolamines in 3T3-L1 treated by MDA-MB-231 sEVs (n= 3 biologically independent samples per group). (i) Transwell assays for evaluating the migration and invasion capacity of MDA-MB-231 and 4T1 cells cocultured with PE. (j) Cell number counting under different PE concentrations (n=3 biologically independent plates). Data are presented as mean  $\pm$  s.e.m; \* $P$ <0.05, \*\* $P$ <0.01, \*\*\* $P$ <0.001, # $P$ <0.05, ns: not significant. Source data and exact  $P$  value are provided as

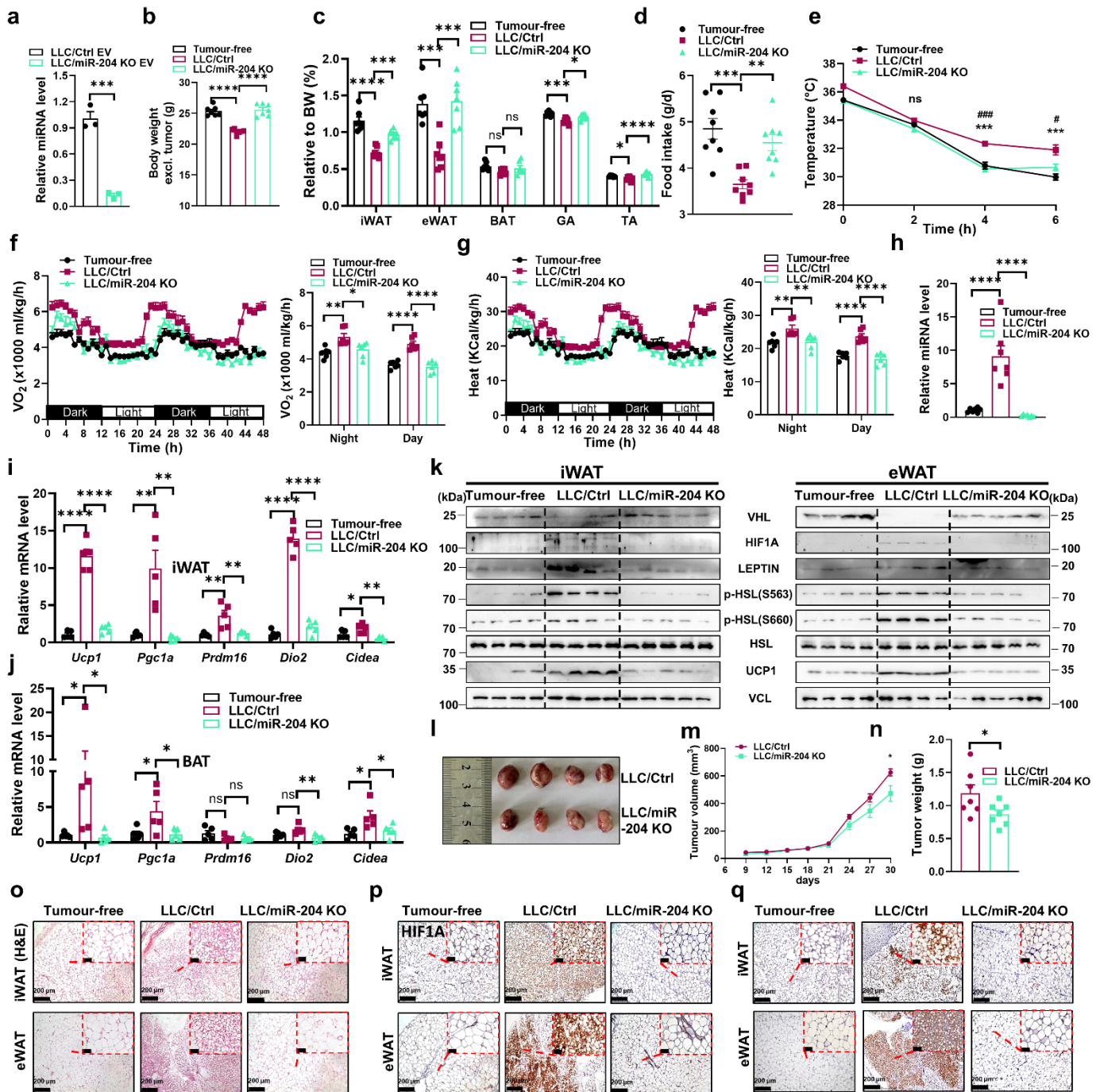


a Source data file. Unpaired two-tailed t-test was used for f, h, j. Two-way ANOVA followed by Bonferroni's multiple comparisons test was used for a. One-way ANOVA was used for b and c.



**Supplementary Figure 6. Exogenous VHL expression suppresses white adipose tissue browning.**

(a) The levels of GFP (green) in tissues of tumour-bearing mice injected with AAV virus, included iWAT, eWAT, BAT, brain, liver, lung, GA, TA and tumour. Repeated three times independently with similar results obtained. Scale bar, 100  $\mu$ m. (b) Tumour volumes were measured every 5 days ( $n=8$  mice per group) and (c) tumour weight ( $n=8$  mice per group). (d) Immunohistochemistry staining of Ki67 in AAV-GFP and AAV-VHL tumour. Scale bar, 200  $\mu$ m. (e-f) Respiratory exchange ratio (RER) and physical activity were monitored over a 48-h period in the two group mice ( $n=5$  mice per group in e and 4 mice per group in f). (g) Representative immunohistochemistry staining of VHL and LEPTIN in iWAT and eWAT. Scale bar, 200  $\mu$ m. (h) Western blots showing indicated proteins in control or transfected with AAV-Vhl in SVF cells from iWAT and eWAT. Repeated three times independently with similar results obtained. Data are presented as mean  $\pm$  s.e.m; \* $P<0.05$ , \*\* $P<0.01$ , ns: not significant. Unpaired two-tailed t-test was used for b, c, e, f. Source data and exact  $P$  value are provided as a Source data file.

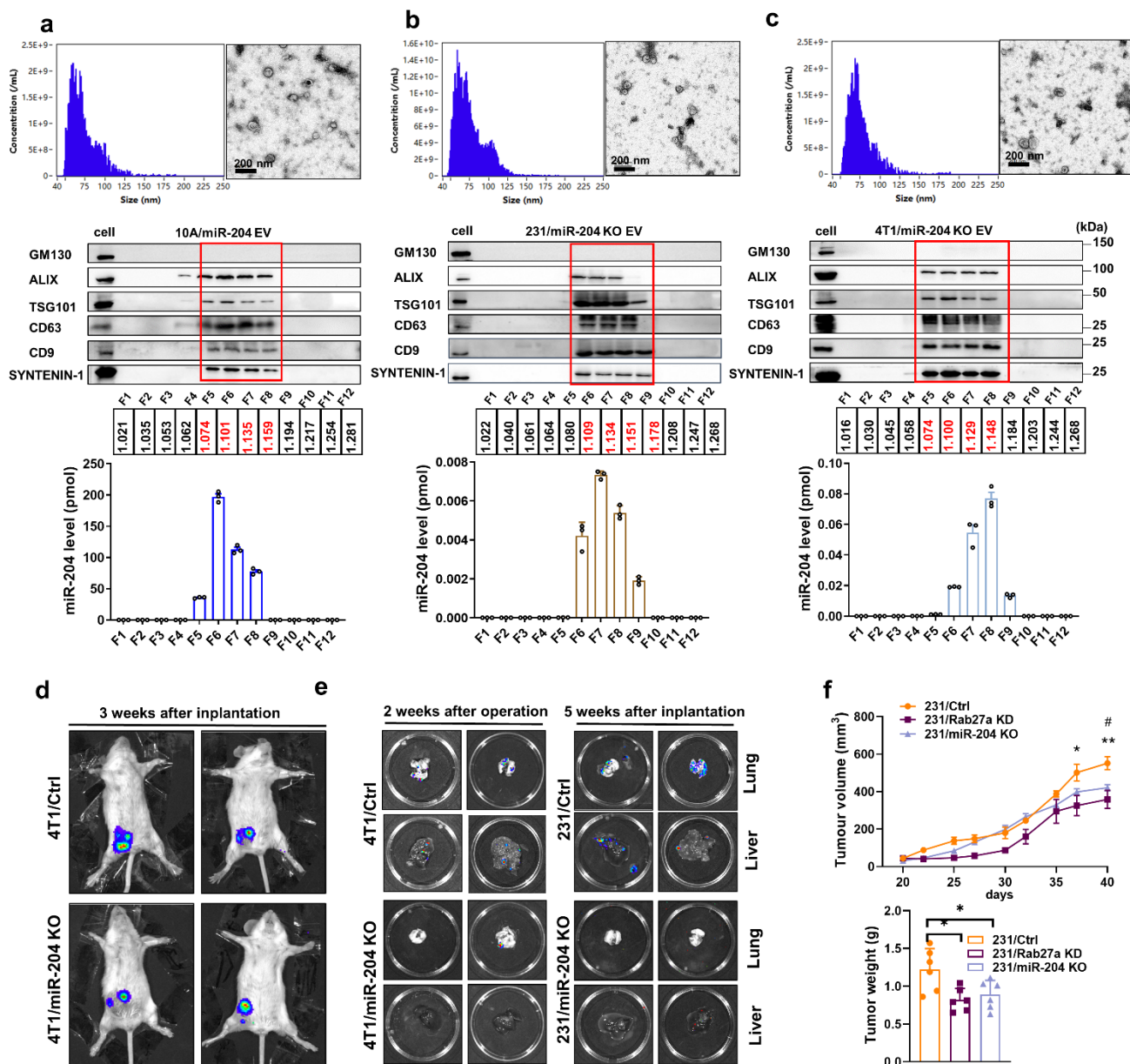


**Supplementary Figure 7. Exosomal miR-204 promotes thermogenesis and lipolysis in LLC-xenografted tumour mice.**

(a) The expression level of miR-204 in LLC and LLC/miR-204 KO derived sEVs. n=3 biologically independent samples per group. (b) Body weight (calculated by subtracting tumour weight from the total weight) of mice bearing LLC tumours (n=7 mice per group). (c) Relative tissues weight (normalized to body weight) (n=7 mice per group). (d) Daily food intake (n=8 mice per group). (e) Rectal temperatures of the mice at different times after cold exposure (n=5 mice per group). (f) Oxygen consumption (VO<sub>2</sub>) and (g) heat production. Dot plots represent the 48-hour average values (n=6 mice per group). (h) The qRT-PCR analysis of miR-204 abundance of serum from each group mice (n=7 mice per group). (i-j) Relative mRNA abundance of the indicated thermogenic markers in iWAT and BAT from each group mice (n=5 mice per group). (k) Western blots showing indicated proteins in iWAT and eWAT from each group mice (tumour-free group, n=4 mice per group; LLC/Ctrl group, n=4 mice per group; LLC/miR-204 KO group, n=5 mice per group). (l) Image of excised

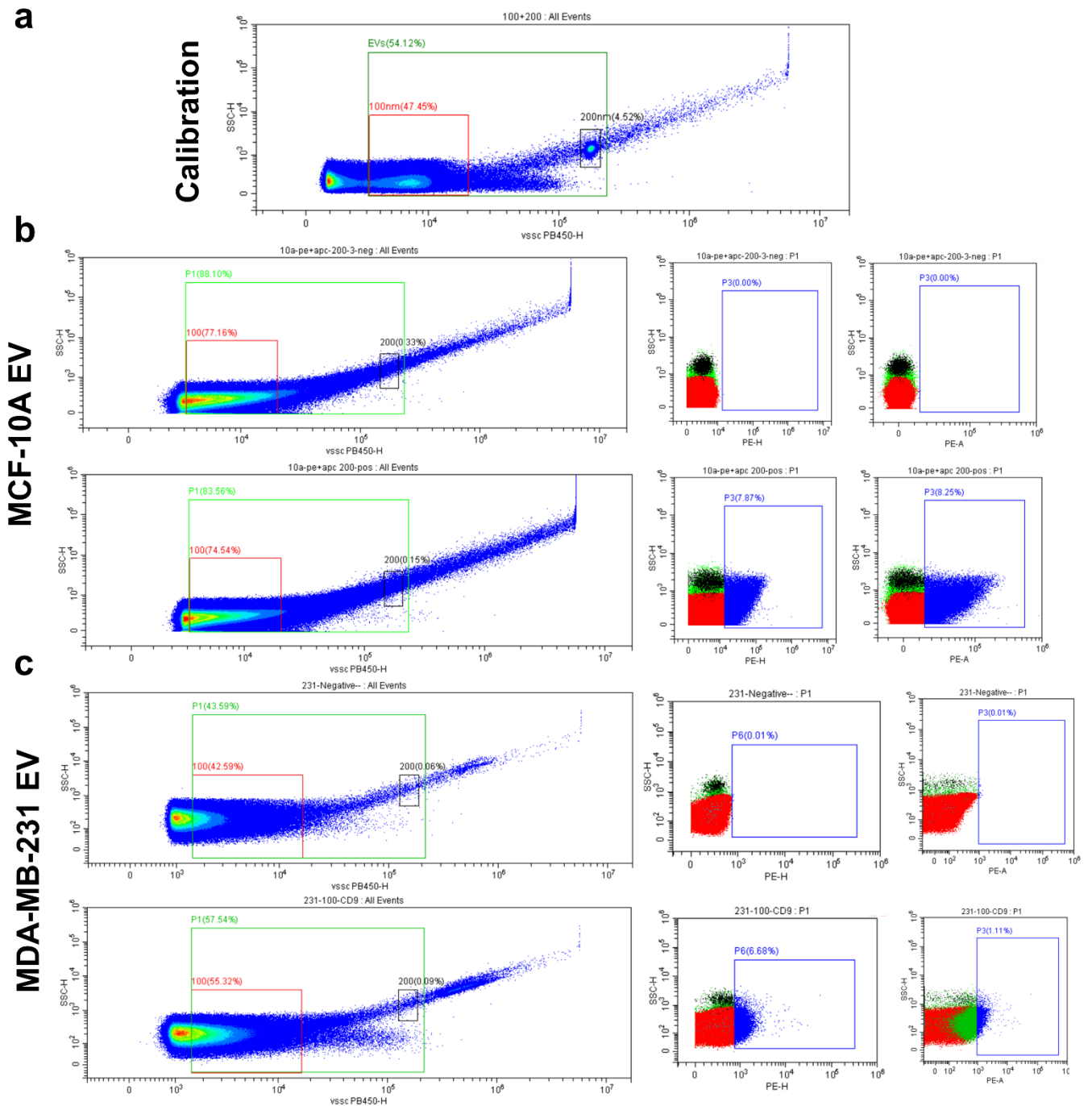
tumours (n=4 mice per group). **(m)** Tumour volume were measured every 3 days (n=7 mice per group) and **(n)** tumour weight (n=7 mice per group). **(o)** Representative H&E staining with iWAT/eWAT from each group mice. Repeated three times independently with similar results obtained. Scale bar, 200  $\mu$ m. **(p)** Representative immunohistochemistry staining of HIF1A in iWAT and eWAT. Repeated three times independently with similar results obtained. Scale bar, 200  $\mu$ m. **(q)** Representative immunohistochemistry staining of UCP1 in iWAT and eWAT. Repeated three times independently with similar results obtained. Scale bar, 200  $\mu$ m. Data are presented as mean  $\pm$  s.e.m; \* $P$ <0.05, \*\* $P$ <0.01, \*\*\* $P$ <0.001, \*\*\*\* $P$ <0.0001, # $P$ <0.05, ### $P$ <0.001, ns: not significant. One-way ANOVA followed by Dunnett multiple comparison test was used for b, c, d, e, f, g, h, I and j. Unpaired two-tailed t-test was used for a, m and n. Source data and exact  $P$  value are provided as a Source data file.





**Supplementary Figure 8. sEV characterization and the effect of miR-204 sEVs on the tumour metastatic.**

Characterization of sEVs derived from 10A/miR-204 (a), 231/miR-204 KO (b) and 4T1/miR-204 KO (c) cells cultured medium: size distribution (top panel), western blotting (middle panel) of indicated whole cell lysates and sEVs fractions showing both positive markers and sEV negative marker (GM130) and relative miR-204 levels (bottom panel) in optiprep gradient fractions (n=3 biological replicates). In the western blot analyses, each well was loaded with 20 µg protein. (d-e) Bioluminescence imaging of 4T1 tumour mice at the 3 weeks after implantation, bioluminescence imaging (green) of 231 tumour mice dissected liver, lung tissues at the end of the experiment. One-way ANOVA followed by Dunnett multiple comparison test. (f) Tumour volume were measured every 3 days (n=6 mice per group) and tumour weight (n=6 mice per group). Data are presented as mean ± s.e.m. \* $P < 0.05$ , \*\* $P < 0.01$ , # $P < 0.05$ . Source data and exact  $P$  value are provided as a Source data file.



**Supplementary Figure 9. Nanoflow cytometry gating strategy for MDA-MB-231 sEVs and MCF-10A sEVs.**

Gating strategies used to analyze CD9 expression on the MDA-MB-231 sEV and MCF-10A sEV membranes.

**(a)** Latex Beads (100 nm and 200 nm mean particle size) (PCS Control Mixed Kit, Beckman Coulter) are characterized and serve as a vesicle size standard. **(b-c)** Representative sEV samples immunostained with the indicated PE-labelled CD9 antibodies.



**Supplementary Table 1: Predicted miRNA regulators of VHL.**

miRNA	Position in the UTR	seed match	context++ score	context++ score percentile	weighted context++ score	conserved branch length	Pct
<i>hsa-miR-223-3p</i>	146-152	7mer-m8	-0.25	91	-0.25	3.383	0.35
<i>hsa-miR-211-5p</i>	631-638	8mer	-0.37	98	-0.37	3.269	0.32
<b><i>hsa-miR-204-5p</i></b>	<b>631-638</b>	<b>8mer</b>	<b>-0.37</b>	<b>98</b>	<b>-0.37</b>	<b>3.269</b>	<b>0.32</b>
<i>hsa-miR-5590-3p</i>	1657-1664	8mer	-0.16	96	-0.16	2.22	< 0.1
<i>hsa-miR-142-5p</i>	1657-1664	8mer	-0.17	96	-0.16	2.22	< 0.1
<i>hsa-miR-340-5p</i>	1658-1665	8mer	-0.08	96	-0.08	2.381	N/A
<i>hsa-miR-21-5p</i>	1689-1695	7mer-m8	-0.2	91	-0.19	3.157	0.2
<i>hsa-miR-590-5p</i>	1689-1695	7mer-m8	-0.15	91	-0.14	3.157	0.2
<i>hsa-miR-101-3p.1</i>	3506-3512	7mer-m8	-0.26	95	-0.25	2.964	0.52
<i>hsa-miR-181b-5p</i>	3585-3592	8mer	-0.24	95	-0.19	3.333	0.22
<i>hsa-miR-181d-5p</i>	3585-3592	8mer	-0.24	95	-0.19	3.333	0.22
<i>hsa-miR-181c-5p</i>	3585-3592	8mer	-0.25	95	-0.19	3.333	0.22
<i>hsa-miR-181a-5p</i>	3585-3592	8mer	-0.24	95	-0.19	3.333	0.22
<i>hsa-miR-4262</i>	3585-3592	8mer	-0.13	91	-0.1	3.333	0.22
<i>hsa-miR-374c-5p</i>	3589-3596	8mer	-0.17	89	-0.13	3.625	N/A
<i>hsa-miR-655-3p</i>	3589-3596	8mer	-0.17	88	-0.13	3.625	N/A
<i>hsa-miR-29a-3p</i>	3684-3690	7mer-m8	-0.43	97	-0.29	4.977	0.86
<i>hsa-miR-29c-3p</i>	3684-3690	7mer-m8	-0.43	97	-0.29	4.977	0.86
<i>hsa-miR-29b-3p</i>	3684-3690	7mer-m8	-0.43	97	-0.29	4.977	0.86
<i>hsa-miR-329-3p</i>	3710-3716	7mer-m8	-0.23	98	0	2.993	N/A
<i>hsa-miR-362-3p</i>	3710-3716	7mer-m8	-0.22	97	0	2.993	N/A

**Supplementary Table 2: Oligonucleotide primers for qRT-PCR analysis.**

Species	Gene	Forward primer (5'-3')	Reverse primer (5'-3')
Mouse	<i>Leptin</i>	GAGACCCCTGTGTCGGTTC	CTGCGTGTGTGAAATGTCATTG
	<i>Npy</i>	TACTCCGCTCTGCGACACTACA	GGCGTTTTCTGTGCTTTCCTTCA
	<i>Agrp</i>	GCCTCAAGAAGACAACCTGCAGAC	AAGCAGGACTCGTGCAGCCTTA
	<i>Pgc1a</i>	GAATCAAGCCACTACAGACACCG	CATCCCTCTTGAGCCTTTCGTG
	<i>Dio2</i>	GGTGGTCAACTTTGGTTCAGCC	AAGTCAGCCACCGAGGAGAACT
	<i>Prdm16</i>	ATCCACAGCACGGTGAAGCCAT	ACATCTGCCCACAGTCCTTGCA
	<i>Ucp1</i>	AGGCTTCCAGTACCATTAGGT	CTGAGTGAGGCAAAGCTGATTT
	<i>Mt-Co1</i>	CACCAGTCAATCCCTGTTGTTACT	GGTAGTTGTCGAGGCCAAAGC
	<i>Ndufv1</i>	GTGGAGGAAGAGATGTCTGTGC	ATGAGCCACCAGGAATCACAGC
	<i>Vhl</i>	GTTTGTGCCATCCCTCAATGTGC	ACCTGACGATGTCCAGTCTCCT
	<i>Hif1a</i>	CCTGCACTGAATCAAGAGGTTGC	CCATCAGAAGGACTTGCTGGCT
	<i>Cidea</i>	GGTGGACACAGAGGAGTTCTTTC	CGAAGGTGACTCTGGCTATTCC
	<i>Actin</i>	CATTGCTGACAGGATGCAGAAGG	TGCTGGAAGGTGGACAGTGAGG
HUMAN	<i>VHL</i>	GCTGTGCCCATCCAAAAAGTCC	CCCAGGAATGAAGTCCAAACCG
	<i>LEPTIN</i>	GACACACGATGGGCTTCTGGTT	ACAACCTGGAGGCATCGCTCTT
	<i>ACTIN</i>	CACCATTGGCAATGAGCGGTTC	AGGTCTTTGCGGATGTCCACGT

**Supplementary Table 3: Antibodies used in this study.**

Antibodies	SOURCE	IDENTIFIER	Dilutions/amounts	Manufacturer's website including validation information
Rab27a (E-8) Mouse monoclonal antibodies	Santa Cruz	Cat# sc-74586	1:100 (WB)	<a href="https://www.scbt.com/zh/p/rab-27a-antibody-e-8?requestFrom=search">https://www.scbt.com/zh/p/rab-27a-antibody-e-8?requestFrom=search</a>
Alix Rabbit polyclonal antibodies	Proteintech	Cat# 12422-1-AP	1:5000 (WB)	<a href="https://www.ptgcn.com/products/PDCD6IP-Antibody-12422-1-AP.htm">https://www.ptgcn.com/products/PDCD6IP-Antibody-12422-1-AP.htm</a>
CD9 Rabbit polyclonal antibodies	Proteintech	Cat# 20597-1-AP	1:1000 (WB)	<a href="https://www.ptgcn.com/products/CD9-Antibody-20597-1-AP.htm">https://www.ptgcn.com/products/CD9-Antibody-20597-1-AP.htm</a>
CD63 Rabbit polyclonal antibody	Proteintech	Cat# 25682-1-AP	1:200 (WB)	<a href="https://www.ptgcn.com/products/CD63-Antibody-25682-1-AP.htm">https://www.ptgcn.com/products/CD63-Antibody-25682-1-AP.htm</a>
CD63 (MX-49.129.5) mouse monoclonal antibodies	Santa Cruz	Cat# sc-5275	1:100 (WB)	<a href="https://www.scbt.com/zh/p/cd63-antibody-mx-49-129-5?requestFrom=search">https://www.scbt.com/zh/p/cd63-antibody-mx-49-129-5?requestFrom=search</a>
TSG101 Rabbit polyclonal antibody	Proteintech	Cat# 28283-1-AP	1:2000 (WB)	<a href="https://www.ptgcn.com/products/TSG101-Antibody-28283-1-AP.htm">https://www.ptgcn.com/products/TSG101-Antibody-28283-1-AP.htm</a>
GOLGA2/GM130 Rabbit polyclonal	Proteintech	Cat# 11308-1-AP	1:2000 (WB)	<a href="https://www.ptgcn.com/products/GOLGA2,GM130-Antibody-11308-1-AP.htm">https://www.ptgcn.com/products/GOLGA2,GM130-Antibody-11308-1-AP.htm</a>
Syntenin-1 Rabbit polyclonal antibody	Proteintech	Cat# 22399-1-AP	1:500 (WB)	<a href="https://www.ptgcn.com/products/SDCBP-Antibody-22399-1-AP.htm">https://www.ptgcn.com/products/SDCBP-Antibody-22399-1-AP.htm</a>
ACTIN (2D4H5) Mouse monoclonal antibodies	Proteintech	Cat# 66009-1-Ig	1:20000 (WB)	<a href="https://www.ptgcn.com/products/Pan-Actin-Antibody-66009-1-Ig.htm">https://www.ptgcn.com/products/Pan-Actin-Antibody-66009-1-Ig.htm</a>
HIF1a Rabbit polyclonal antibody	Proteintech	Cat# 20960-1-AP	1:2000 (WB); 1:200 (IHC)	<a href="https://www.ptgcn.com/products/HIF1A-Antibody-20960-1-AP.htm">https://www.ptgcn.com/products/HIF1A-Antibody-20960-1-AP.htm</a>
HIF1 alpha (AFirm8002(AFB17813)) Mouse monoclonal Antibody	Affinity	Cat# BF8002	1:1000 (WB)	<a href="https://www.affbiotech.cn/goods-16579-BF8002-HIF1_alpha_Mouse_monoclonal_Antibody.html">https://www.affbiotech.cn/goods-16579-BF8002-HIF1_alpha_Mouse_monoclonal_Antibody.html</a>
Anti-UCP1	Abcam	Cat# ab10983	1:1000 (WB); 1:500 (IHC)	<a href="https://www.abcam.cn/ucp1-antibody-ab10983.html">https://www.abcam.cn/ucp1-antibody-ab10983.html</a>
Vinculin Rabbit mAb	Abclonal	Cat# A23468	1:1000 (WB)	<a href="https://abclonal.com.cn/catalog/A23468">https://abclonal.com.cn/catalog/A23468</a>
CREB (48H2) Rabbit mAb	Cell Signaling Technology	Cat# 9197	1:1000 (WB)	<a href="https://www.cellsignal.cn/products/primary-antibodies/creb-48h2-rabbit-mab/9197?site-search-type=Products&amp;N=4294956287&amp;Ntt=9197&amp;fromPage=plp&amp;requestid=3098024">https://www.cellsignal.cn/products/primary-antibodies/creb-48h2-rabbit-mab/9197?site-search-type=Products&amp;N=4294956287&amp;Ntt=9197&amp;fromPage=plp&amp;requestid=3098024</a>
Phospho-CREB (Ser133) (87G3) Rabbit mAb	Cell Signaling Technology	Cat# 9198	1:1000 (WB)	<a href="https://www.cellsignal.cn/products/primary-antibodies/phospho-creb-ser133-87g3-rabbit-mab/9198?site-search-type=Products&amp;N=4294956287&amp;Ntt=9198&amp;fromPage=plp&amp;requestid=3100133">https://www.cellsignal.cn/products/primary-antibodies/phospho-creb-ser133-87g3-rabbit-mab/9198?site-search-type=Products&amp;N=4294956287&amp;Ntt=9198&amp;fromPage=plp&amp;requestid=3100133</a>
ATGL Antibody	Cell Signaling Technology	Cat# 2138	1:1000 (WB)	<a href="https://www.cellsignal.cn/products/primary-antibodies/atgl-antibody/2138?site-search-type=Products&amp;N=4294956287&amp;Ntt=2138&amp;fromPage=plp&amp;requestid=3100495">https://www.cellsignal.cn/products/primary-antibodies/atgl-antibody/2138?site-search-type=Products&amp;N=4294956287&amp;Ntt=2138&amp;fromPage=plp&amp;requestid=3100495</a>
HSL Antibody	Cell Signaling Technology	Cat# 4107	1:1000 (WB)	<a href="https://www.cellsignal.cn/products/primary-antibodies/hsl-antibody/4107?site-search-type=Products&amp;N=4294956287&amp;Ntt=4107&amp;fromPage=plp&amp;requestid=3100652">https://www.cellsignal.cn/products/primary-antibodies/hsl-antibody/4107?site-search-type=Products&amp;N=4294956287&amp;Ntt=4107&amp;fromPage=plp&amp;requestid=3100652</a>
Phospho-HSL (Ser563) Antibody	Cell Signaling Technology	Cat# 4139	1:1000 (WB)	<a href="https://www.cellsignal.cn/products/primary-antibodies/phospho-hsl-ser563-antibody/4139?site-search-type=Products&amp;N=4294956287&amp;Ntt=4139&amp;fromPage=plp&amp;requestid=3102740">https://www.cellsignal.cn/products/primary-antibodies/phospho-hsl-ser563-antibody/4139?site-search-type=Products&amp;N=4294956287&amp;Ntt=4139&amp;fromPage=plp&amp;requestid=3102740</a>
Phospho-HSL (Ser660) Antibody	Cell Signaling Technology	Cat# 45804	1:1000 (WB)	<a href="https://www.cellsignal.cn/products/primary-antibodies/phospho-hsl-ser660-antibody/45804?site-search-type=Products&amp;N=4294956287&amp;Ntt=45804&amp;fromPage=plp&amp;requestid=3103130">https://www.cellsignal.cn/products/primary-antibodies/phospho-hsl-ser660-antibody/45804?site-search-type=Products&amp;N=4294956287&amp;Ntt=45804&amp;fromPage=plp&amp;requestid=3103130</a>
VHL Antibody	Affinity	Cat# AF6292	1:1000 (WB)	<a href="https://www.affbiotech.cn/goods-1890-AF6292-VHL_Antibody.html">https://www.affbiotech.cn/goods-1890-AF6292-VHL_Antibody.html</a>
Leptin Antibody	Affinity	Cat# DF8583	1:1000 (WB); 1:100 (IHC)	<a href="https://www.affbiotech.cn/goods-12056-DF8583-Leptin_Antibody.html">https://www.affbiotech.cn/goods-12056-DF8583-Leptin_Antibody.html</a>
STAT3 Antibody	Affinity	Cat# AF6294	1:1000 (WB)	<a href="https://www.affbiotech.cn/goods-1892-AF6294-STAT3_Antibody.html">https://www.affbiotech.cn/goods-1892-AF6294-STAT3_Antibody.html</a>
Phospho-STAT3 (Tyr705) Antibody	Affinity	Cat# AF3293	1:1000 (WB)	<a href="https://www.affbiotech.cn/goods-1458-AF3293-Phospho_STAT3_Tyr705_Antibody.html">https://www.affbiotech.cn/goods-1458-AF3293-Phospho_STAT3_Tyr705_Antibody.html</a>

Ki67 Antibody	Affinity	Cat# AF0198	1:100 (IHC)	<a href="https://www.affbiotech.cn/goods-897-AF0198-Ki67_Antibody.html">https://www.affbiotech.cn/goods-897-AF0198-Ki67_Antibody.html</a>
Sheep Anti-Digoxigenin Fab fragments Antibody, AP Conjugated	Roche	Cat# 110932749 10	5mU/ml (ISH)	<a href="https://www.sigmaaldrich.cn/CN/zh/product/roche/11093274910">https://www.sigmaaldrich.cn/CN/zh/product/roche/11093274910</a>
Goat anti-Mouse IgG (H+L) Secondary Antibody	Thermo Fisher Scientific	Cat# 31430	1:5000 (WB)	<a href="https://www.thermofisher.cn/cn/zh/antibody/product/Goat-anti-Mouse-IgG-H-L-Secondary-Antibody-Polyclonal/31430">https://www.thermofisher.cn/cn/zh/antibody/product/Goat-anti-Mouse-IgG-H-L-Secondary-Antibody-Polyclonal/31430</a>
Goat anti-Rabbit IgG (H+L) Secondary Antibody	Thermo Fisher Scientific	Cat# 31460	1:10000 (WB)	<a href="https://www.thermofisher.cn/cn/zh/antibody/product/Goat-anti-Rabbit-IgG-H-L-Secondary-Antibody-Polyclonal/31460">https://www.thermofisher.cn/cn/zh/antibody/product/Goat-anti-Rabbit-IgG-H-L-Secondary-Antibody-Polyclonal/31460</a>
PE anti-human CD9 Antibody	BioLegend	Cat# 312106	1:50 (FC)	<a href="https://www.biolegend.com/en-us/products/pe-anti-human-cd9-antibody-2213">https://www.biolegend.com/en-us/products/pe-anti-human-cd9-antibody-2213</a>

**Supplementary Table 4: Clinical information included in this study.**

Patient No.	Medical record No.	Age	Height(cm)	Weight(kg)	BMI	Grade	ER	PR	HER2	Notes
1	10228646	71	160	50	19.53125	Moderately	+	+	+	
2	10223146	80	158	59	23.63403	Moderately	+	+	+	
3	10222574	69	158	52	20.83	Poorly	+	+	++	
4	10220569	71	157	55	22.31328	Moderately	+	+	+	
5	10221730	61	155	56	23.30905	Moderately	-	-	+++	
6	10221708	63	148	48	21.91381	Poorly	-	-	-	
7	10213169	51	156	51	20.95661	Poorly	+	+	+	
8	10210360	53	158	55	22.03173	Moderately	+	+	-	
9	10208290	47	158	54	21.63115	Poorly	+	-	+	
10	10196171	52	158	56	22.4323	Poorly	-	-	++	
11	10170776	74	170	62	21.45329	Moderately	+	+	-	
12	10189802	53	170	71	24.56747	Moderately	+	+	++	
13	10174233	70	156	55	22.60026	Poorly	-	-	-	
14	10173299	42	162	62	23.62445	Poorly	-	-	-	
15	10191000	38	153	42.5	18.15541	Poorly	-	-	-	cachexia
16	10197923	40	155	41	17.06556	Poorly	-	-	-	cachexia
17	10195792	56	158	52	20.83	Poorly	-	-	-	
18	10194890	58	164	59	21.93635	Moderately	+	+	+	
19	10190185	58	168	55	19.48696	Moderately	-	-	+	
20	10251715	52	168	67	23.73866	Poorly	-	-	-	
21	10256435	68	160	58	22.65625	Poorly	-	-	+	
22	10235511	31	167	44	15.77683	Poorly	-	-	-	cachexia
23	10254206	66	160	45	17.57813	Poorly	-	-	-	cachexia
24	10254249	33	162	162	17.90886	Poorly	-	-	-	cachexia
25	10232071	30	160	47	18.35938	Poorly	-	-	-	cachexia
26	10248783	50	159	47	18.59104	Poorly	-	-	+	
27	10232278	32	160	48	18.75					
28	10253886	70	158	78	31.24499	Moderately	+	+	+	
29	10241992	35	163	82.5	31.05123					
30	10228564	67	165	80	29.38476	Moderately	+	+	+	
31	10190333	58	155	68	28.30385					
32	10190975	70	155	68	28.30385	Moderately	+	+	+	
33	10232250	58	158	70	28.04038					
34	10276291	44	158	70	28.04038	Moderately	+	+	++	
35	10274128	70	164	75	27.88519					
36	10210360	53	158	55	22.03173	Moderately	+	+	-	
37	10213169	51	156	51	20.95661	Moderately	+	+	+	
38	10214357	64	155	60	24.97399	Moderately	+	+	-	
39	10216929	36	165	65	23.87511	Moderately	+	+	++	
40	10221062	73	164	60	22.30815	Moderately	+	+	+	
41	10221708	63	148	48	21.91381	Poorly	-	-	-	
42	10180033	37	162	69	26.29172	Moderately	+	+	-	
43	10178763	52	150	55.5	24.66667	Moderately	+	+	+	
44	10180782	65	162	61.5	23.43393	Moderately	+	+	+	
45	10181069	52	164	63	23.42356	Moderately	+	+	-	
46	10181305	38	170	65	22.49135	Moderately	+	+	+++	