

Review

Metabolic Dysfunction-Associated Fatty Liver Disease and Chronic Viral Hepatitis: The Interlink

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Abstract: Metabolic dysfunction-associated fatty liver disease (MAFLD) has now affected nearly one-third of the global population and has become the number one cause of chronic liver disease in the world because of the obesity pandemic. Chronic hepatitis resulting from hepatitis B virus (HBV) and hepatitis C virus (HCV) remain significant challenges to liver health even in the 21st century. The co-existence of MAFLD and chronic viral hepatitis can markedly alter the disease course of individual diseases and can complicate the management of each of these disorders. A thorough understanding of the pathobiological interactions between MAFLD and these two chronic viral infections is crucial for appropriately managing these patients. In this comprehensive clinical review, we discuss the various mechanisms of chronic viral hepatitis-mediated metabolic dysfunction and the impact of MAFLD on the progression of liver disease.

Keywords: metabolic dysfunction-associated fatty liver disease (MAFLD); chronic viral hepatitis; hepatitis B virus (HBV); hepatitis C virus (HCV); hepatic fibrosis; cirrhosis; hepatocellular carcinoma



Citation: Fernandez, C.J.; Alkhalifah, M.; Afsar, H.; Pappachan, J.M. Metabolic Dysfunction-Associated Fatty Liver Disease and Chronic Viral Hepatitis: The Interlink. *Pathogens* **2024**, *13*, 68. <https://doi.org/10.3390/pathogens13010068>

Academic Editors: Ivana Lazarevic and Valentina Svicher

Received: 13 November 2023

Revised: 5 January 2024

Accepted: 7 January 2024

Published: 10 January 2024



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1. Introduction

Metabolic dysfunction-associated fatty liver disease (MAFLD) has become the most common cause of chronic liver disease in recent years, affecting nearly one-third of the global population because of the obesity pandemic [1]. Although a good proportion of MAFLD cases can remain clinically nonprogressive, some cases can develop severe forms of the disease, such as hepatic fibrosis, cirrhosis, and hepatocellular carcinoma (HCC). Several factors, including environmental, epigenetic, genetic, metabolic, and infective causes, can influence the progression of MAFLD to advanced stages of liver disease [2]. The previous terminology, nonalcoholic fatty liver disease (NAFLD), was changed in 2020 to MAFLD by an international consensus panel to reflect these associations of the disease [3]. With the new nomenclature, several uncertainties concerning the pathobiology and consequences of the disease have been resolved [4]. Unlike NAFLD, patients with chronic viral hepatitis, alcohol excess, drug-induced steatosis, or other chronic liver diseases can have a diagnosis of MAFLD.

Chronic viral hepatitis resulting from hepatitis B virus (HBV) and hepatitis C virus (HCV) remains an important cause of advanced liver disease in several regions of the world. Hepatic steatosis is a common feature of both chronic HBV [5] and HCV [6] infections. When patients with MAFLD acquire these chronic viral infections or vice versa,

the pathobiological characteristics of either disease can be markedly altered, and the risk of progression to advanced liver disease, including fibrosis, can be perpetuated. Diagnosing MAFLD in these patients would facilitate the early initiation of lifestyle interventions and multidisciplinary management to improve the prognosis of these patients. Moreover, managing individual disorders can be more complex when they co-exist. Therefore, it is important to understand the pathophysiological interlink between these infections and MAFLD when they co-exist to plan appropriate management, which is the aim of this clinical update review.

2. MAFLD and Chronic HBV Infection

Epidemiology

According to a World Health Organization (WHO) report, in 2015, more than 250 million people globally were suffering from chronic hepatitis B (CHB) infection [7]. Additionally, 887,000 people died from complications related to CHB, including cirrhosis and liver cancer, in the same year. These data underscores the immense burden that CHB places on global public health. There is no direct evidence that CHB is associated with an increased risk of hepatic steatosis. Several meta-analyses have examined this phenomenon. In a meta-analysis of 17 studies, which included 4100 HBV-infected patients and 8 of which also included 945 HCV-infected patients, it was reported that approximately 29.6% of patients with HBV developed fatty liver, like in the general population [8]. The same study observed that 60% of the patients with HCV developed fatty liver. Moreover, the study observed a statistically significant positive association with the male sex (OR 1.74, 95% CI [1.28–2.38], $p < 0.001$) and body mass index (SMD 2.17, 95% CI [1.23, 3.11], $p < 0.001$); and a negative association with HBV-DNA (SMD -74.12 , 95% CI [-82.93 , -65.31], $p < 0.001$). This strong negative association between HBV-DNA and steatosis may indicate a protective effect of HBV infection on steatosis. Another meta-analysis of 54 studies, involving 28,648 CHB patients, found a pooled prevalence of hepatic steatosis of up to 32.8% [9]. A more recent meta-analysis, which included 98 studies and 48,472 patients, demonstrated an even higher global prevalence of hepatic steatosis among CHB patients, reaching 34.93% [10].

3. Effect of MAFLD on CHB Infection and Chronic Liver Disease Progression

MAFLD is associated with increased Th17 cell-related gene expression, increased IL-21 levels, activation of T and B cells, production of inflammatory cytokines, elimination of HBV proliferation with resultant immune clearance of HBV DNA, and HbeAg [11]. The NASH stage of MAFLD is associated with increased expression of toll-like receptors (TLRs) in hepatocytes, Kupffer cells (KCs), hepatic stellate cells (HSCs), sinusoidal endothelial cells, and hepatic dendritic cells (DCs) [12]. Lipopolysaccharide (LPA) induces activation of the TLR4 and myeloid differentiation factor 88 (MyD88)-mediated pathways in obese individuals [13]. Activation of the TLR4/MyD88 pathway contributes to the activation of HSCs and the production of chemokines, which recruits further KCs [14]. TLR4 activation in KCs induces the secretion of pro-inflammatory cytokines (IL-1, IL-6, IL-8, TNF- α , and chemokines) and profibrogenic factors (TGF- β) to activate the inflammation–fibrosis–carcinoma sequence [14]. TLR4/MyD88 signaling also induces the production of IFN- β , IL-6, and TNF- α to inhibit HBV replication [13]. Thus, activation of innate immunity through TLR signaling is associated with the inhibition of HBV replication and the retardation of the progression of MAFLD to NASH, fibrosis, and HCC [15].

MAFLD-associated metabolic stress could reduce peroxisome proliferator-activated receptor- γ coactivator 1 alpha (PGC-1 α), which in turn could inhibit HBV replication and induce Fas-mediated apoptosis of HBV-infected cells, resulting in HBV-clearance and reduction of HBV-related liver disease progression [16]. CHB is associated with a decreased risk of hyperlipidemia [17,18] and raised serum adiponectin levels [19], which could contribute to a lower risk of hepatic steatosis.

On the other hand, the production of saturated fatty acid—palmitic acid as a metabolic component of MAFLD could be associated with impaired function of hepatic DCs and impaired HBsAg processing/presentation, leading to inadequate immune response/HBV-clearance and subsequent development of severe HBV-related liver disease progression [20]. Neutrophil-derived reactive oxygen species (ROS) induced by MAFLD could result in the activation of p38 mitogen-activated protein kinase (MAPK), which in turn could augment HBV replication and result in the progression of MAFLD to NASH [21].

4. Effects of CHB Infection on the MAFLD and Chronic Liver Disease Progression

Some of the transcription factors (including CEBP [22], CREB [23], HNF3 [24], HNF4 [25], FXR [26], RXR [27], and PPAR [28]) involved in the transcription of HBV DNA are involved in hepatic glucose, lipid, bile acid, and xenobiotic metabolism [28] may either inhibit or induce regeneration, inflammation, fibrosis, and malignant transformation of hepatic cells. Differential expressions of IL-13, G-CSF, CCL11, IL-6, and IL-4 are thought to play a role in developing steatosis and fibrosis in patients with CHB infection. IL-13 facilitates hepatic steatosis and fibrosis, the latter through mechanisms including the stimulation of TGF- β 1 gene expression [29] and through activation of the JAK-STAT-6 pathway, in turn results in the production of CCL11, an eosinophil chemotactic protein [30]. CCL11-mediated hepatic eosinophilic infiltration and activation results in hepatic steatosis and fibrosis [31]. G-CSF ameliorates hepatic steatosis by reducing the expression of *SREBP-1c* [32]. IL-4 and IL-6 protect against hepatic fibrosis [33], IL-4 through secretion of matrix metalloproteinase-12 (MMP-12) [34], and IL-6 through the promotion of proliferation/survival of HSCs [35].

In patients with CHB infection, hepatitis B protein X (HBx)—a 17 kDa soluble protein coded by the HBV DNA induces expression of various genes related to lipid accumulation including PPAR [36], *SREBP* [36], *FABP1* [37], *LXR* [38], and *FATP2* [39], thereby promoting lipogenesis. HBx also stimulates various transcription factors, including STAT3, NF- κ B, PI3K/AKT, and Src [40], which promote hepatocyte proliferation [40], inhibit apoptosis [40], and stimulate inflammation [41], thus leading to the development of HCC. Moreover, the pre-S1 domain of the HBV envelope binds to sodium taurocholate cotransporting polypeptide (NCTP), limiting the function of NCTP, thus promoting compensatory bile acid synthesis, cholesterol provision, and hepatic steatosis [42]. Steatosis associated with MAFLD, and the resultant oxidative stress might generate an intra-hepatic pro-fibrotic and pro-cancerous environment [43]. Additionally, CHB-associated deficiency of PML (promyelocytic leukemia protein) results in altered lipid metabolism and steatosis-associated carcinogenesis [44]. Reduced levels of global DNA methylation in patients with concurrent MAFLD and CHB lead to chromosomal abnormality, instability, fragility, and HCC development [45].

Hepatic steatosis was observed in nearly 18% of patients with biopsy-proven CHB infection [46]. Steatosis had an independent association with body mass index and fasting blood glucose levels, and it does not correlate with the degree of hepatic fibrosis [46]. There is a possible genetic susceptibility to develop steatosis in CHB infection, with the rs1010023 polymorphism in the *PNPLA3* gene and rs58542926 polymorphism in the *TM6SF2* gene increasing the tendency to develop MAFLD among patients with CHB infection [43]. HBx could play an important role in increasing the risk of HBV-induced steatosis. On the other hand, the reduced risk of hyperlipidemia and the increased adiponectin levels could reduce the risk of HBV-induced steatosis. Although MAFLD is associated with lower HBV viral load and with an increased rate of HBsAg clearance, both CHB and MAFLD could act synergistically to promote the progression of liver disease, causing hepatocyte injury, inflammation, fibrosis, and HCC. Figure 1 shows the pathobiological interlink between chronic HBV infection and metabolic dysfunction and the impact of MAFLD on HBV replication.

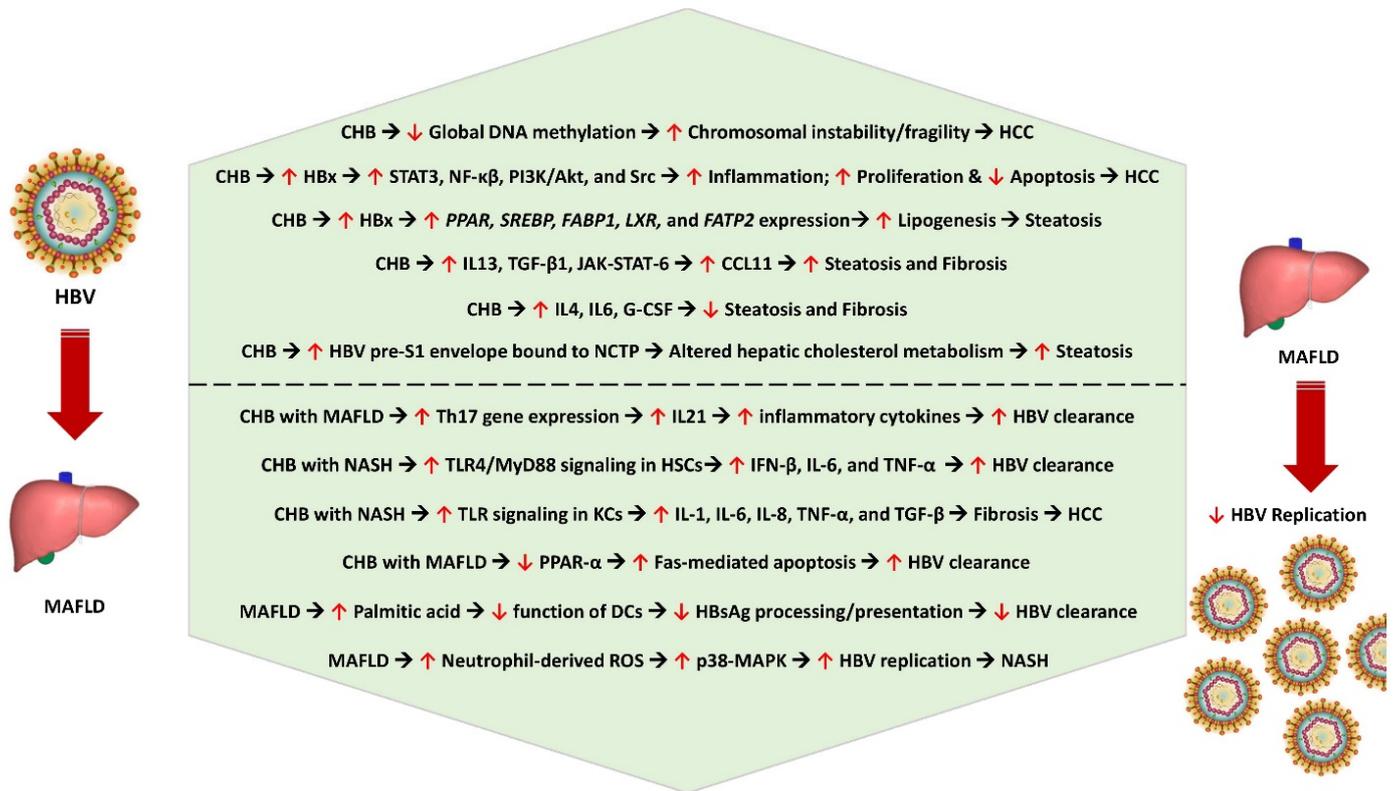


Figure 1. Pathobiological interlink between CHB and metabolic dysfunction and the impact of MAFLD on HBV replication. CHB—chronic hepatitis B, HCC—hepatocellular carcinoma, HBx—hepatitis B protein X, STAT3—signal transducer and activator of transcription 3, NF-κβ—nuclear factor kappa B subunit, PI3K/AKT—phosphoinositide 3-kinase/protein kinase B, PPAR—peroxisome proliferator-activated receptor gene, SREBP—sterol regulatory element-binding protein gene, FABP1—fatty acid-binding protein 1 gene, LXR—liver X receptor gene, FATP2—fatty acid transport protein 2 gene, IL13—interleukin 13, TGF-β1—transforming growth factor beta 1, JAK-STAT-6—Janus kinase-signal transducer and activator of transcription 6, CCL11—C-C motif ligand 11 (eosinophil chemotactic protein or eotaxin-1), IL4—interleukin 4, IL6—interleukin 6, G-CSF—granulocyte colony-stimulating factor, NCTP—sodium taurocholate cotransporting polypeptide, Th17—T helper 17 cell, IL21—interleukin 21, TLR4/Myd88—Toll-like receptor-myeloid differentiation factor 88, IFN-β—interferon beta, KCs—Kupffer cells, HSCs—hepatic stellate cells, IL8—interleukin 8, TNF-α—tumor necrosis factor alpha, Fas or FasR—Fas receptor (apoptosis antigen 1), DCs—dendritic cells, HbsAg—hepatitis B surface antigen, ROS—reactive oxygen species, p38-MAPK—p38-mitogen-activated protein kinase, NASH—nonalcoholic steatohepatitis.

A retrospective study involving 1076 CHB patients with a median follow-up period of 9.8 years evaluated the importance of MAFLD in patients with CHB [47]. The study observed that MAFLD is associated with reduced event-free (aHR 2.00, 95% CI 1.26–3.19), HCC-free (aHR 1.93, 95% CI 1.17–3.21), and transplant-free survival (aHR 1.80, 95% CI 0.98–3.29), implying higher risk for liver-related events and death. A prospective study of 10,546 CHB patients observed that after a median follow-up period of 5.1 years, MAFLD is associated with a 58% reduced risk of HCC (adjusted hazard ratio or aHR 0.42, 95% CI 0.25–0.68, $p < 0.001$) [48]. The steatosis and metabolic dysfunction had distinctive effects on the risk for HCC. While steatosis was protective against HCC (aHR 0.45, 95% CI 0.30–0.67, $p < 0.001$), a greater burden of metabolic dysfunction increased the HCC risk (aHR 1.40 per dysfunction increase, 95% CI 1.19–1.66, $p < 0.001$) [48]. MAFLD can have both metabolic and non-metabolic complications in patients with co-existing CHB as given in Table 1.

Table 1. Metabolic and non-metabolic complications of co-existing MAFLD and CHB [49–54].

Metabolic complications
Insulin Resistance
Dyslipidemia—elevated triglyceride and LDL cholesterol levels
Obesity
Hypertension
Cardiovascular disease
Non-Metabolic Complications
Hepatic fibrosis
Hepatocellular Carcinoma (HCC)
Chronic liver disease-related complications—ascites, encephalopathy, and variceal bleeding
Increased risk of infection
Impaired quality of life—fatigue, discomfort, and the need for ongoing medical care

Management

The management of MAFLD in patients with CHB involves a multifaceted approach. Traditional liver biopsy, considered the gold standard for diagnosis of hepatic steatosis, is associated with a high risk of internal bleeding [55], making non-invasive methods a more appropriate approach. One such method is the controlled attenuation parameter (CAP) via fibro-scan [56,57], which measures attenuation during ultrasonography to estimate the degree of steatosis. CAP has a relatively low cost and is suitable for most first-line clinical settings [58]. In CHB, patients' CAP demonstrated a high degree of accuracy for steatosis assessment compared to other noninvasive methods [59,60]. It has been used in predicting the presence and severity of MAFLD in CHB patients [61].

CHB management requires antiviral treatments such as nucleotide analogs like tenofovir alafenamide or entecavir to suppress viral replication [62], although a cure is often difficult. Patients with concurrent MAFLD may experience variations in viral activity and liver enzymes due to the presence of NASH [63]. Conflicting evidence exists in the response to treatment in patients with co-existent MAFLD and CHB. While some studies indicate lower treatment response in CHB patients with hepatic steatosis, others show comparable responses. Monitoring serum ALT and HBV DNA levels and timely intervention for poor responders are crucial for managing CHB in the presence of MAFLD [64,65].

Acute intervention for concurrent MAFLD is crucial, given its adverse impact on overall health. Lifestyle modifications, including strict diet control aiming at weight loss and adherence to certain dietary practices, such as a hypocaloric diet and avoidance of food high in saturated fats or ultra-processed foods, coupled with regular exercise, form the cornerstones of therapy [66,67]. Several pharmacological treatment options for steatohepatitis are currently being developed, such as semaglutide [68], lanifibranor (pan-peroxisome proliferator-activated receptor agonist) [69], resmetirom (selective thyroid hormone receptor- β agonist) [70,71] and obeticholic acid (selective farnesoid X receptor agonist) [72,73], with some promising results, but their routine use in CHB patients with concurrent MAFLD requires further evaluation.

Improvement of hepatic steatosis may affect HBV replication, necessitating careful monitoring during metabolic correction. Factors like diabetes mellitus, obesity, and dyslipidemia contribute to the progression of both MAFLD and CHB infection [74], making the aggressive management of both conditions essential. These metabolic risk factors are independently associated with liver disease progression, hepatocarcinogenesis, and overall mortality in CHB patients [75,76]. Therefore, addressing metabolic dysfunction is the key to improving co-existent CHB in patients with MAFLD.

5. MAFLD and Chronic HCV Infection

5.1. Epidemiology

According to global estimates, approximately 71.1 million people have chronic hepatitis C virus infection, with a global prevalence of 1% in 2015 [77]. Globally, the most common HCV genotype is genotype 1 (nearly 50% of all adults with HCV infection), followed by genotypes 3, 2, 4, 6, and 5 respectively [78]. HCV infection, especially genotype 3, is well known to be associated with hepatic steatosis. Genotype 3 is highly steatogenic [79], and it exhibits a steatosis prevalence of up to 86% while other phenotypes possess a steatosis prevalence of around 50% [80]. The mean prevalence of steatosis in chronic HCV is around 55% across all HCV genotypes [80]. HCV genotype 3 is reported to exert a direct cytopathic effect on the liver in direct proportion to the viral load, even in the absence of other metabolic risk factors like visceral obesity and/or diabetes mellitus [7]. The term ‘viral steatosis’ is used for this entity [80].

With the change in nomenclature from NAFLD to MAFLD, those patients with HCV infection who also meet the criteria for the diagnosis of MAFLD are classified as hepatitis C with MAFLD. Thus, there are now two types of HCV: hepatitis C with MAFLD and hepatitis C without MAFLD. The term ‘metabolic steatosis’ is used for the entity seen in patients with hepatitis C and MAFLD [80]. Contrary to metabolic steatosis, ‘viral steatosis’ is associated with reduced LDL cholesterol and triglyceride levels [81]. Genotypes 1, 2, and 4 essentially promote insulin resistance associated with host metabolic risk factors, including visceral obesity [79]. MAFLD patients with hepatitis C have a higher risk for advanced hepatic fibrosis but with a similar atherosclerotic CVD risk in comparison to those with MAFLD alone without CHC infection (CHC) [82].

A recent Australian study [83] observed a 43.1% prevalence of MAFLD in patients with CHC infection in contrast to the global prevalence of MAFLD of 25% in the general population [84]. This dual etiology group is associated with an increased risk for hepatic injury, inflammation, and fibrosis (all $p < 0.001$). This study observed that those with CHC and lean MAFLD had a similar rate of advanced fibrosis (31.6%) in comparison to those who had obesity and/or diabetes mellitus (31.8% and 46.2%, respectively, with $p = 0.325$). However, those with dual etiology are at a greater risk of developing advanced fibrosis and HCC even after HCV clearance, implying that managing MAFLD is equally as important as HCV clearance to prevent the progression of hepatic disease and death from HCC or cardiovascular disease [84].

5.2. Disease Characteristics

Table 2 shows the differences between HCV genotype 3 and other genotypes of HCV in their pathobiological characteristics, response to treatment, and disease outcomes on long-term follow-up.

Table 2. Comparison of disease characteristics between various genotypes of hepatitis C with regard to the cause of hepatic steatosis and responsiveness to the antiviral therapy [85].

	Genotype 3 HCV	Non-Genotype 3 HCV
Mechanism of steatosis	Viral steatosis	Metabolic steatosis
Location	Periportal zone (acinar 1)	Centrilobular (acinar 3)
HCV RNA viral load	Corelation with MAFLD severity	No relation to MAFLD severity
Response to antiviral	MAFLD reversible with SVR	Reduced response to therapy
Consequence	High rate of steatosis, more rapid progression to advanced fibrosis, and increased HCC risk	Lower rates of steatosis, slower progression to advanced fibrosis, and lower HCC risk

6. Effect of MAFLD on CHC Infection and Chronic Liver Disease Progression

Lipid droplets are involved in the replication and virion assembly of HCV, and stimulation of de novo lipogenesis (DNL) via MAFLD (and CHC) facilitates the entry of the virus into the hepatocytes [86]. Moreover, upon release from hepatocytes, the mature HCVs in circulation are complexed with lipoproteins [87]. A complex metabolic network exists in the fatty liver to regulate HCV replication. While saturated and monounsaturated fatty acids are required for replication, polyunsaturated fatty acids inhibit HCV RNA replication [88]. Lipid peroxidation, a feature of NASH, inhibits HCV replication [89]. HCV-infected cells have phosphatidylcholines and triglycerides with longer fatty acyl chains [90]. Knocking down fatty acid elongases [90], fatty acid desaturases [90], or phosphatidyl ethanolamine transferase [91] (PEMT) can inhibit HCV RNA replication.

7. Effect of CHC Infection on the MAFLD and Chronic Liver Disease Progression

Development of MAFLD in patients with CHC depends on the host's genetic background, including the rs738409 polymorphism in the *PNPLA3* gene and the rs58542926 polymorphism in *TM6SF2* gene [92]. CHC infection appears to downregulate the intrahepatic expression of *PPAR- α* , and its target known as carnitine palmitoyl acyl-CoA transferase 1A (CPT1A), thereby reducing fatty acid β -oxidation [93]. The presence of HCV core protein results in mitochondrial dysfunction, oxidative stress, and disruption of fatty acid metabolism, leading to steatosis [94]. MAFLD from genotypes 1 and 4 are associated with insulin resistance mediated by reduced expression of insulin receptor substrates (IRS1 and IRS2), thereby reducing signaling through phosphoinositide 3-kinase (PI3K) and Akt [95]. Insulin resistance is also mediated by an increased hepatic expression of fatty acid transporter (CD36), which is involved in increasing fatty acid uptake [96].

MAFLD from genotype 3 is associated with the inhibition of microsomal triglyceride transfer protein (MTTP), resulting in the impaired assembly of ApoB and lipids to form VLDL, thereby impairing triglyceride secretion and thus intracellular triglyceride accumulation in hepatocytes [97]. Another pathophysiological mechanism for MAFLD from genotype 3 is that HCV-3a core protein induces the PI3K-Akt pathway, increases sterol regulatory element-binding protein-1c (*SREBP-1c*) activity, which in turn increases the expression of the fatty acid synthase (FAS) [98]. HCV-3a core protein results in the downregulation of phosphatase and tensin (*PTEN*) homologs inside the hepatocytes triggering the formation of large lipid droplets [99]. HCV-3a core protein acts as an inhibitor of *PPAR- α* activity, resulting in lowered triglyceride breakdown and intrahepatic accumulation of fatty acids [100].

The inhibition of *PPAR- α* activity that accompanies CHC infection increases nuclear factor kappa B (NF- κ B) and activator protein 1 (AP-1) levels, leading to the progression of MAFLD to NASH [101]. Similarly, the increased levels of soluble TNF- α receptors that develop in CHC infection also can cause progression to NASH [102]. Kupffer cells exposed to HCV secrete CCL5, which in turn triggers NF- κ B and ERK signaling in hepatic stellate cells. The resultant pro-inflammatory (NLRP3, IL1B, IL-6, and CCL5) and pro-fibrotic (TGF- β 1, COL4A1, MMP2, and α -SMA) products promote the progression of MAFLD to fibrosis in patients with CHC infection [103].

7.1. Complications

CHC patients could develop insulin resistance, hyperinsulinemia, and diabetes mellitus. This could occur independently from obesity but is associated with a higher HCV replication rate and an enhanced risk of progression to fibrosis. Host factors including obesity, obesity-mediated insulin resistance, and co-existent MAFLD in patients with CHC are associated with a higher degree of hepatic fibrosis, increased risk of HCC, reduced response to interferon alpha-based therapy, and accelerated atherosclerosis in comparison to CHC without MAFLD [104–107]. However, results from the German hepatitis C registry do not show a significant fibrotic progression in patients with co-existent MAFLD and CHC infection [108]. A recent study observed that single nucleotide polymorphism (*rs12979860*) in interferon- λ 4 (*IFNL4*) has an independent strong association with inflammation and

fibrosis, especially in young women with CHC genotype 3 [109]. Figure 2 shows the pathobiological interlink between chronic HCV infection and metabolic dysfunction and the impact of MAFLD on HCV replication.

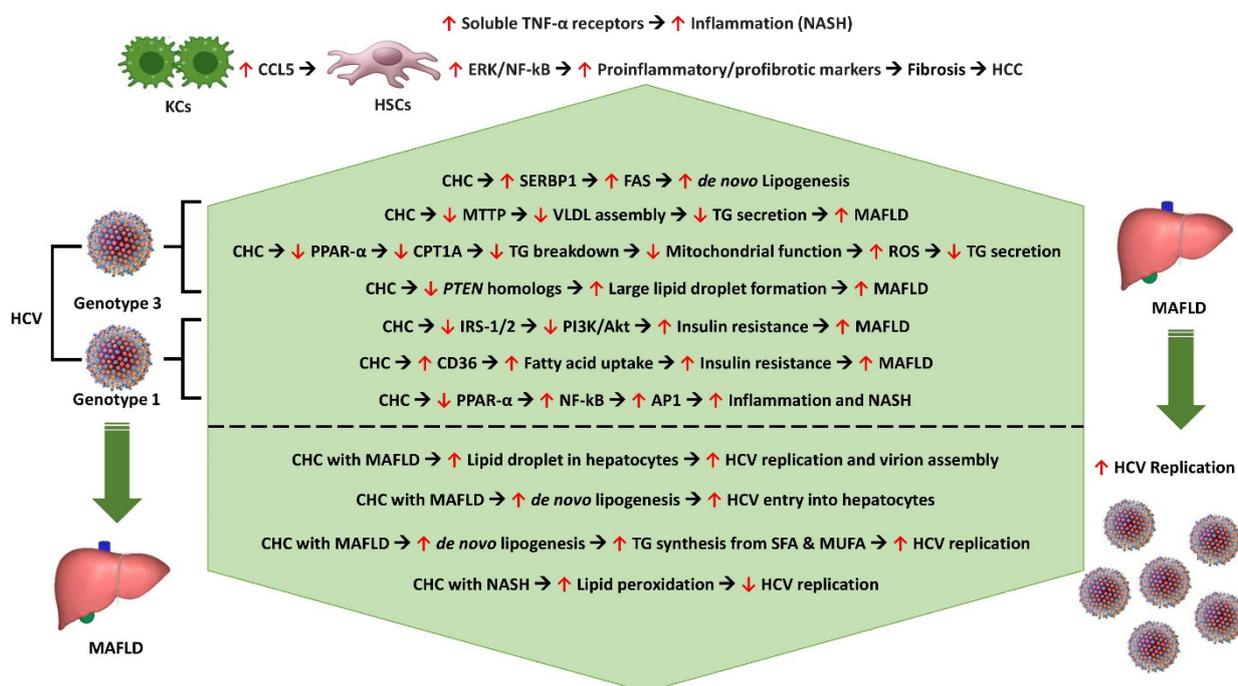


Figure 2. Pathobiological interlink between CHC and metabolic dysfunction and the impact of MAFLD on HCV replication. KCs—Kupffer cells, CCL5—C-C motif ligand 5, HSCs—hepatic stellate cells, TNF- α —tumor necrosis factor alpha, NASH—nonalcoholic steatohepatitis, ERK/NF- κ B—extracellular signal-regulated kinase/nuclear factor kappa B subunit, HCC—hepatocellular carcinoma, CHC—chronic hepatitis C, SERBP1—Serpine1 mRNA-binding protein 1, FAS—fatty acid synthase, MTP—microsomal triglyceride transfer protein, VLDL—very low-density lipoprotein, TG—triglyceride, PPAR—peroxisome proliferator-activated receptor, CPT1A—carnitine palmitoyl acyl-CoA transferase 1A, ROS—reactive oxygen species, PTEN—phosphatase and tensin gene, IRS1/2—insulin receptor substrates 1 and 2, PI3K/AKT—phosphoinositide 3-kinase/protein kinase B, STAT3—signal transducer and activator of transcription 3, CD36—cluster of differentiation 36 (fatty acid translocase), AP1—activator protein 1, SFA—saturated fatty acid, MUFA—monounsaturated fatty acid.

7.2. Management

Obesity is well known to trigger the development of MAFLD and the progression of CHC infection. Though in the interferon era of CHC treatment, obesity was a hindrance to achieving SVR [106], in the era of direct-acting antiviral (DAA) therapy, this is no longer the case [110]. In a prospective study comprising 11,469 patients with CHC infection, up to 78% of patients were either overweight or obese at the treatment initiation [111]. At a follow-up of 2 years, patients who managed to achieve SVR had gained 0.56 ± 12.8 lbs compared to 3.43 ± 14.6 lbs of weight loss in those who failed to achieve SVR ($p < 0.0001$). Moreover, 22% of CHC patients with BMI ≤ 25 at DAA therapy onset became overweight during the follow-up period [111].

In 1991, the FDA approved IFN- α as the first antiviral medication for HCV, and seven years later, ribavirin was introduced [112]. A few years later, three different combinations of DAAs, namely NS3 protease inhibitors, NS5B polymerase inhibitors, and NS5A inhibitors, were approved [113]. The combination of IFN- α and ribavirin decreased SVR in patients with CHC infection [114]. Adding rosuvastatin to this combination could improve the SVR rates along with a reduction in steatosis and fibrosis [115]. Though statins are a viable option, further randomized controlled trials are needed. A combination of IFN- α and vitamin E could achieve a significant reduction in the viral load [116]. In CHC patients who

are refractory to IFN- α therapy, the addition of an antioxidant d- α -tocopherol reduced the rate of progression of fibrosis through inhibition of stellate cell activation [117].

DAA (which are currently the first-line agents, with improved tolerability and superior efficacy for HCV clearance) achieved a median decrease of liver stiffness measurement (LSM) by 0.9 (−0.6–3.2) kPa, $p < 0.001$, but with a median increase of CAP values by 25 (−12.5–61.5) dB/m, $p < 0.001$, indicating that DAAs could increase hepatic steatosis [118]. Though DAAs could achieve HCV clearance, the co-existing MAFLD can persist, particularly in patients with obesity, thereby increasing the risk of progression of hepatic disease. Hence, co-existing MAFLD should be treated with therapeutic lifestyle changes. However, DAAs have added beneficial effects on cardiovascular risk factors—with an increase in the triglyceride-to-cholesterol ratio in the VLDL molecules [119], improvement in glycemic control [120], and significant reduction in the risk of cardiovascular events [121]. Due to potential drug–drug interactions, DAAs should be carefully selected with statins or antihypertensive drugs [122].

8. Areas of Uncertainty/Emerging Concepts

Co-existing CHB infection and MAFLD are becoming increasingly common, and it is important to identify the etiology when hepatitis develops. A novel noninvasive diagnostic model has been developed using various parameters including CAP, LSM, HBV DNA, and AST in predicting HBV-related inflammation in CHB with concurrent MAFLD to identify patients who need anti-HBV therapy [123]. Uncertainty and challenges exist in the management of patients with co-existing CHB and MAFLD in the absence of long-term follow-up data. Though there are inconsistent results on the impact of hepatic steatosis on the efficacy of antiviral therapy for CHB (with some showing reduced and others showing comparable therapy response), and there are insufficient data to confirm a direct link between nucleoside analogues and hepatic steatosis, the onset/progression of MAFLD should be monitored as a potential adverse effect [124]. Antiviral drugs may have effects on the metabolism. For example, tenofovir disoproxil fumarate could significantly reduce the lipoprotein levels in patients with CHB [125]. Statins could retard the decompensation of HBV-associated cirrhosis [126] and HCC [127]. As PPAR- α could promote HBV replication [128], patients should be cautioned when CHB co-exists with MAFLD. Figure 3 summarizes the impact of CHB and CHC on various stages of MAFLD progression.

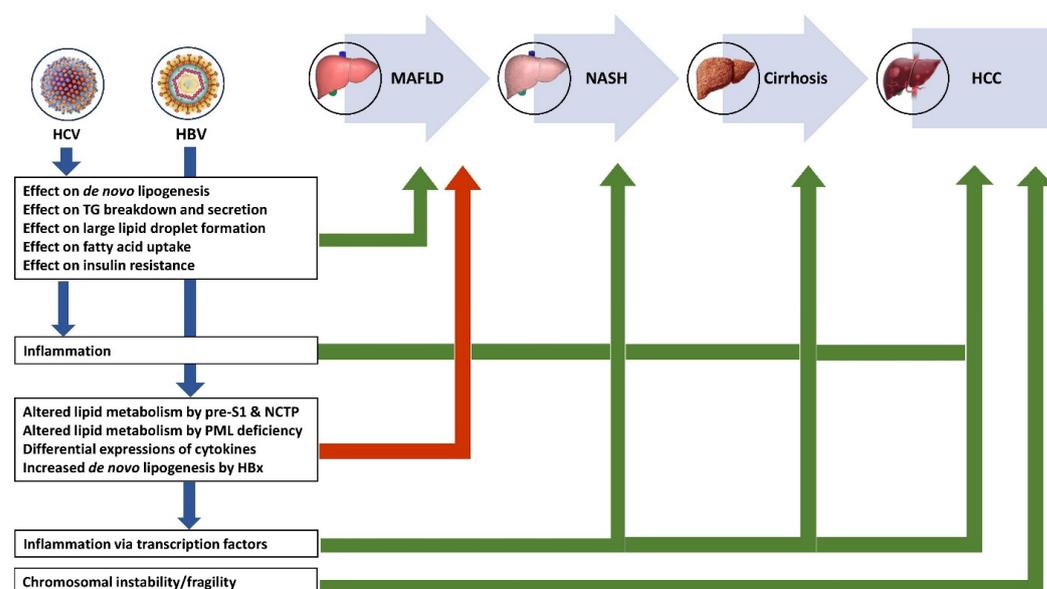


Figure 3. Impact of CHB and CHC on various stages of MAFLD progression. HCV—hepatitis C virus, HBV—hepatitis B virus, MAFLD—metabolic dysfunction-associated fatty liver disease, NASH—nonalcoholic steatohepatitis, HCC—hepatocellular carcinoma, NCTP—sodium taurocholate cotransporting polypeptide.

Let us try to answer a few important questions on this topic.

Does CHB or CHC occur in a patient who already has a confirmed diagnosis of MAFLD?

CHB in patients with MAFLD is associated with reduced HBV replication, whereas CHC in patients with MAFLD is associated with increased HCV replication. In patients with CHB or CHC, the co-existence of MAFLD is associated with progression to CHB/CHC-related fibrosis and HCC.

Does CHB or CHC in their natural evolution determine the development of MAFLD?

Despite several associated steatogenic mechanisms, CHB has a negative association with the risk of developing MAFLD [8,129]. On the other hand, CHC has a positive association with the risk of developing MAFLD [130], with the various genotypes increasing the risk by distinctive mechanisms. However, some believe that MAFLD in patients with chronic viral hepatitis either existed before the infection (or at least the risk factors for MAFLD already existed) and was not diagnosed, or MAFLD developed simultaneously with chronic viral hepatitis due to the development of other conditions that determine MAFLD.

Does the treatment given for CHB or CHC lead to the development of MAFLD?

There are insufficient data to confirm a direct link between CHB/CHC therapy and MAFLD.

Does the treatment given for MAFLD lead to an increase in viral replication in CHB?

Better clinical and mechanistic evidence is needed to reach any definite conclusions.

The following table (Table 3) summarizes some of the answered and unanswered topics related to co-existent chronic viral hepatitis and MAFLD.

Table 3. Summary of the interactions between chronic viral hepatitis and MAFLD [131].

	HBV	HCV
CHB/CHC promoting fatty liver	No	Yes
CHB/CHC predisposing patients to diabetes	Unknown	Yes
CHB/CHC worsening lipid profile	No	No
MAFLD promoting CHB/CHC-related fibrosis	Yes	Yes
MAFLD promoting CHB/CHC-related HCC	Yes	Yes
MAFLD promoting viral replication	No	Yes
MAFLD reducing the antiviral response	Unknown	IFN- α : Yes DAAs: unknown
Drugs for diabetes, hypertension, and dyslipidemia reducing antiviral response	Unknown	IFN- α : unknown Some DAAs: Yes

9. Conclusions

MAFLD and chronic viral hepatitis from HBV and HCV remain significant challenges to liver health across the globe. Disease progression occurs when MAFLD co-exists with HBV or HCV in the same individual, resulting in higher complication rates, and the management of either disease becomes more complex. Timely clinical suspicion and appropriate therapeutic interventions might modify the disease outcomes concerning this dangerous co-existence. More research is needed to improve our understanding regarding the pathobiology and interactions between these diseases when they co-exist and the therapeutic strategies to improve clinical outcomes.

Author Contributions: C.J.F. drafted the initial manuscript with extensive literature search and scientific review of the data. Additional help for drafting and literature search was provided by H.A. and M.A.; J.M.P. conceptualized the idea and provided overall supervision of the drafting and revision process. All authors inputted in the final revision of the paper and agreed in the current form. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data are contained within the article.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. Fouada, S.; Jeeyavudeen, M.S.; Pappachan, J.M.; Jayanthi, V. Pathobiology of Metabolic-Associated Fatty Liver Disease. *Endocrinol. Metab. Clin. N. Am.* **2023**, *52*, 405–416. [[CrossRef](#)] [[PubMed](#)]
2. Tanaka, N.; Kimura, T.; Fujimori, N.; Nagaya, T.; Komatsu, M.; Tanaka, E. Current status, problems, and perspectives of non-alcoholic fatty liver disease research. *World J. Gastroenterol.* **2019**, *25*, 163–177. [[CrossRef](#)]
3. Eslam, M.; Sanyal, A.J.; George, J.; International Consensus Panel. MAFLD: A Consensus-Driven Proposed Nomenclature for Metabolic Associated Fatty Liver Disease. *Gastroenterology* **2020**, *158*, 1999–2014.e1. [[CrossRef](#)]
4. Crane, H.; Gofton, C.; Sharma, A.; George, J. MAFLD: An optimal framework for understanding liver cancer phenotypes. *J. Gastroenterol.* **2023**, *Epub ahead of print*. [[CrossRef](#)] [[PubMed](#)]
5. Baclig, M.O.; Reyes, K.G.; Liles, V.R.; Mapua, C.A.; Dimamay, M.P.S.; Gopez-Cervantes, J. Hepatic steatosis in chronic hepatitis B: A study of metabolic and genetic factors. *Int. J. Mol. Epidemiol. Genet.* **2018**, *9*, 13–19.
6. Asselah, T.; Rubbia-Brandt, L.; Marcellin, P.; Negro, F. Steatosis in chronic hepatitis C: Why does it really matter? *Gut* **2006**, *55*, 123–130. [[CrossRef](#)] [[PubMed](#)]
7. Younossi, Z.; Tacke, F.; Arrese, M.; Chander Sharma, B.; Mostafa, I.; Bugianesi, E.; Wai-Sun Wong, V.; Yilmaz, Y.; George, J.; Fan, J.; et al. Global Perspectives on Nonalcoholic Fatty Liver Disease and Nonalcoholic Steatohepatitis. *Hepatology* **2019**, *69*, 2672–2682. [[CrossRef](#)]
8. Machado, M.V.; Oliveira, A.G.; Cortez-Pinto, H. Hepatic steatosis in hepatitis B virus infected patients: Meta-analysis of risk factors and comparison with hepatitis C infected patients. *J. Gastroenterol. Hepatol.* **2011**, *26*, 1361–1367. [[CrossRef](#)]
9. Zheng, Q.; Zou, B.; Wu, Y.; Yeo, Y.; Wu, H.; Stave, C.D.; Cheung, R.C.; Nguyen, M.H. Systematic review with meta-analysis: Prevalence of hepatic steatosis, fibrosis and associated factors in chronic hepatitis B. *Aliment. Pharmacol. Ther.* **2021**, *54*, 1100–1109. [[CrossRef](#)]
10. Jiang, D.; Chen, C.; Liu, X.; Huang, C.; Yan, D.; Zhang, X.; Zhou, Y.; Lin, Y.; Zhou, Y.; Guan, Z.; et al. Concurrence and impact of hepatic steatosis on chronic hepatitis B patients: A systematic review and meta-analysis. *Ann. Transl. Med.* **2021**, *9*, 1718. [[CrossRef](#)] [[PubMed](#)]
11. Li, H.J.; Kang, F.B.; Li, B.S.; Yang, X.Y.; Zhang, Y.G.; Sun, D.X. Interleukin-21 inhibits HBV replication in vitro. *Antivir. Ther.* **2015**, *20*, 583–590. [[CrossRef](#)]
12. Kiziltsas, S. Toll-like receptors in pathophysiology of liver diseases. *World J. Hepatol.* **2016**, *8*, 1354–1369. [[CrossRef](#)] [[PubMed](#)]
13. Zhang, R.N.; Pan, Q.; Zhang, Z.; Cao, H.X.; Shen, F.; Fan, J.G. Saturated Fatty Acid inhibits viral replication in chronic hepatitis B virus infection with nonalcoholic fatty liver disease by toll-like receptor 4-mediated innate immune response. *Hepat. Mon.* **2015**, *15*, e27909. [[CrossRef](#)]
14. Seki, E.; De Minicis, S.; Osterreicher, C.H.; Kluwe, J.; Osawa, Y.; Brenner, D.A.; Schwabe, R.F. TLR4 enhances TGF-beta signaling and hepatic fibrosis. *Nat. Med.* **2007**, *13*, 1324–1332. [[CrossRef](#)] [[PubMed](#)]
15. Soares, J.B.; Pimentel-Nunes, P.; Afonso, L.; Rolanda, C.; Lopes, P.; Roncon-Albuquerque, R., Jr.; Gonçalves, N.; Boal-Carvalho, I.; Pardo, F.; Lopes, S.; et al. Increased hepatic expression of TLR2 and TLR4 in the hepatic inflammation-fibrosis-carcinoma sequence. *Innate Immun.* **2012**, *18*, 700–708. [[CrossRef](#)] [[PubMed](#)]
16. Piccinin, E.; Villani, G.; Moschetta, A. Metabolic aspects in NAFLD, NASH and hepatocellular carcinoma: The role of PGC1 coactivators. *Nat. Rev. Gastroenterol. Hepatol.* **2019**, *16*, 160–174. [[CrossRef](#)]
17. Huang, C.Y.; Lu, C.W.; Liu, Y.L.; Chiang, C.H.; Lee, L.T.; Huang, K.C. Relationship between chronic hepatitis B and metabolic syndrome: A structural equation modeling approach. *Obesity* **2016**, *24*, 483–489. [[CrossRef](#)]
18. Jarčuška, P.; Janičko, M.; Kružliak, P.; Novák, M.; Veselíny, E.; Fedačko, J.; Senajová, G.; Dražilová, S.; Madarasová-Gecková, A.; Mareková, M.; et al. Hepatitis B virus infection in patients with metabolic syndrome: A complicated relationship. Results of a population based study. *Eur. J. Intern. Med.* **2014**, *25*, 286–291. [[CrossRef](#)]
19. Yoon, S.; Jung, J.; Kim, T.; Park, S.; Chwae, Y.J.; Shin, H.J.; Kim, K. Adiponectin, a downstream target gene of peroxisome proliferator-activated receptor γ , controls hepatitis B virus replication. *Virology* **2011**, *409*, 290–298. [[CrossRef](#)]

20. Martinet, J.; Dufeu-Duchesne, T.; Bruder Costa, J.; Larrat, S.; Marlu, A.; Leroy, V.; Plumas, J.; Aspod, C. Altered functions of plasmacytoid dendritic cells and reduced cytolytic activity of natural killer cells in patients with chronic HBV infection. *Gastroenterology* **2012**, *143*, 1586–1596.e8. [[CrossRef](#)]
21. Chang, W.W.; Su, I.J.; Chang, W.T.; Huang, W.; Lei, H.Y. Suppression of p38 mitogen-activated protein kinase inhibits hepatitis B virus replication in human hepatoma cell: The antiviral role of nitric oxide. *J. Viral Hepat.* **2008**, *15*, 490–497. [[CrossRef](#)]
22. López-Cabrera, M.; Letovsky, J.; Hu, K.Q.; Siddiqui, A. Multiple liver-specific factors bind to the hepatitis B virus core/pregenomic promoter: Trans-activation and repression by CCAAT/enhancer binding protein. *Proc. Natl. Acad. Sci. USA* **1990**, *87*, 5069–5073. [[CrossRef](#)]
23. Kim, B.K.; Lim, S.O.; Park, Y.G. Requirement of the cyclic adenosine monophosphate response element-binding protein for hepatitis B virus replication. *Hepatology* **2008**, *48*, 361–373. [[CrossRef](#)]
24. Raney, A.K.; Zhang, P.; McLachlan, A. Regulation of transcription from the hepatitis B virus large surface antigen promoter by hepatocyte nuclear factor 3. *J. Virol.* **1995**, *69*, 3265–3272. [[CrossRef](#)]
25. Yu, X.; Mertz, J.E. Distinct modes of regulation of transcription of hepatitis B virus by the nuclear receptors HNF4alpha and COUP-TF1. *J. Virol.* **2003**, *77*, 2489–2499. [[CrossRef](#)]
26. Ramière, C.; Scholtès, C.; Diaz, O.; Icard, V.; Perrin-Cocon, L.; Trabaud, M.A.; Lotteau, V.; André, P. Transactivation of the hepatitis B virus core promoter by the nuclear receptor FXRalpha. *J. Virol.* **2008**, *82*, 10832–10840. [[CrossRef](#)]
27. Reese, V.C.; Oropeza, C.E.; McLachlan, A. Independent activation of hepatitis B virus biosynthesis by retinoids, peroxisome proliferators, and bile acids. *J. Virol.* **2013**, *87*, 991–997. [[CrossRef](#)]
28. Tang, H.; McLachlan, A. Transcriptional regulation of hepatitis B virus by nuclear hormone receptors is a critical determinant of viral tropism. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 1841–1846. [[CrossRef](#)]
29. Lee, C.G.; Homer, R.J.; Zhu, Z.; Lanone, S.; Wang, X.; Kotliansky, V.; Shipley, J.M.; Gotwals, P.; Noble, P.; Chen, Q.; et al. Interleukin-13 induces tissue fibrosis by selectively stimulating and activating transforming growth factor beta(1). *J. Exp. Med.* **2001**, *194*, 809–821. [[CrossRef](#)]
30. Tarantino, G.; Cabibi, D.; Cammà, C.; Alessi, N.; Donatelli, M.; Petta, S.; Craxi, A.; Di Marco, V. Liver eosinophilic infiltrate is a significant finding in patients with chronic hepatitis C. *J. Viral Hepat.* **2008**, *15*, 523–530. [[CrossRef](#)]
31. Tacke, F.; Trautwein, C.; Yagmur, E.; Hellerbrand, C.; Wiest, R.; Brenner, D.A.; Schnabl, B. Up-regulated eotaxin plasma levels in chronic liver disease patients indicate hepatic inflammation, advanced fibrosis and adverse clinical course. *J. Gastroenterol. Hepatol.* **2007**, *22*, 1256–1264. [[CrossRef](#)] [[PubMed](#)]
32. Song, Y.S.; Fang, C.H.; So, B.I.; Park, J.Y.; Jun, D.W.; Kim, K.S. Therapeutic effects of granulocyte-colony stimulating factor on non-alcoholic hepatic steatosis in the rat. *Ann. Hepatol.* **2013**, *12*, 115–122. [[CrossRef](#)] [[PubMed](#)]
33. Wong, S.W.; Ting, Y.W.; Yong, Y.K.; Tan, H.Y.; Barathan, M.; Riazalhosseini, B.; Bee, C.J.; Tee, K.K.; Larsson, M.; Velu, V.; et al. Chronic inflammation involves CCL11 and IL-13 to facilitate the development of liver cirrhosis and fibrosis in chronic hepatitis B virus infection. *Scand. J. Clin. Lab. Investig.* **2021**, *81*, 147–159. [[CrossRef](#)] [[PubMed](#)]
34. Weng, S.Y.; Wang, X.; Vijayan, S.; Tang, Y.; Kim, Y.O.; Padberg, K.; Regen, T.; Molokanova, O.; Chen, T.; Bopp, T.; et al. IL-4 Receptor Alpha Signaling through Macrophages Differentially Regulates Liver Fibrosis Progression and Reversal. *EBioMedicine* **2018**, *29*, 92–103. [[CrossRef](#)] [[PubMed](#)]
35. Nieto, N. Oxidative-stress and IL-6 mediate the fibrogenic effects of [corrected] Kupffer cells on stellate cells. *Hepatology* **2006**, *44*, 1487–1501, Erratum in *Hepatology* **2007**, *45*, 546. [[CrossRef](#)] [[PubMed](#)]
36. Kim, K.H.; Shin, H.J.; Kim, K.; Choi, H.M.; Rhee, S.H.; Moon, H.B.; Kim, H.H.; Yang, U.S.; Yu, D.Y.; Cheong, J. Hepatitis B virus X protein induces hepatic steatosis via transcriptional activation of SREBP1 and PPAR-gamma. *Gastroenterology* **2007**, *132*, 1955–1967. [[CrossRef](#)] [[PubMed](#)]
37. Wu, Y.L.; Peng, X.E.; Zhu, Y.B.; Yan, X.L.; Chen, W.N.; Lin, X. Hepatitis B virus X Protein induces hepatic steatosis by enhancing the expression of liver fatty acid binding protein. *J. Virol.* **2015**, *90*, 1729–1740. [[CrossRef](#)] [[PubMed](#)]
38. Na, T.Y.; Shin, Y.K.; Roh, K.J.; Kang, S.A.; Hong, I.; Oh, S.J.; Seong, J.K.; Park, C.K.; Choi, Y.L.; Lee, M.O. Liver X receptor mediates hepatitis B virus X protein-induced lipogenesis in hepatitis B virus-associated hepatocellular carcinoma. *Hepatology* **2009**, *49*, 1122–1131. [[CrossRef](#)]
39. Lu, Y.; Yang, X.; Kuang, Q.; Wu, Y.; Tan, X.; Lan, J.; Qiang, Z.; Feng, T. HBx induced upregulation of FATP2 promotes the development of hepatic lipid accumulation. *Exp. Cell Res.* **2023**, *430*, 113721. [[CrossRef](#)]
40. Sivasudhan, E.; Blake, N.; Lu, Z.; Meng, J.; Rong, R. Hepatitis B Viral Protein HBx and the Molecular Mechanisms Modulating the Hallmarks of Hepatocellular Carcinoma: A Comprehensive Review. *Cells* **2022**, *11*, 741. [[CrossRef](#)]
41. Cho, H.K.; Kim, S.Y.; Yoo, S.K.; Choi, Y.H.; Cheong, J. Fatty acids increase hepatitis B virus X protein stabilization and HBx-induced inflammatory gene expression. *FEBS J.* **2014**, *281*, 2228–2239. [[CrossRef](#)] [[PubMed](#)]
42. Oehler, N.; Volz, T.; Bhadra, O.D.; Kah, J.; Allweiss, L.; Giersch, K.; Bierwolf, J.; Riecken, K.; Pollok, J.M.; Lohse, A.W.; et al. Binding of hepatitis B virus to its cellular receptor alters the expression profile of genes of bile acid metabolism. *Hepatology* **2014**, *60*, 1483–1493. [[CrossRef](#)]
43. Tong, X.; Song, Y.; Yin, S.; Wang, J.; Huang, R.; Wu, C.; Shi, J.; Li, J. Clinical impact and mechanisms of hepatitis B virus infection concurrent with non-alcoholic fatty liver disease. *Chin. Med. J.* **2022**, *135*, 1653–1663. [[CrossRef](#)]
44. Chung, Y.L.; Wu, M.L. The role of promyelocytic leukemia protein in steatosis-associated hepatic tumors related to chronic hepatitis B virus Infection. *Transl. Oncol.* **2018**, *11*, 743–754. [[CrossRef](#)]

45. Li, F.; Ou, Q.; Lai, Z.; Pu, L.; Chen, X.; Wang, L.; Sun, L.; Liang, X.; Wang, Y.; Xu, H.; et al. The co-occurrence of chronic hepatitis B and fibrosis is associated with a decrease in hepatic global DNA methylation levels in patients with non-alcoholic fatty liver disease. *Front. Genet.* **2021**, *12*, 671552. [[CrossRef](#)]
46. Thomopoulos, K.C.; Arvaniti, V.; Tsamantas, A.C.; Dimitropoulou, D.; Gogos, C.A.; Siagris, D.; Theocharis, G.J.; Labropoulou-Karatza, C. Prevalence of liver steatosis in patients with chronic hepatitis B: A study of associated factors and of relationship with fibrosis. *Eur. J. Gastroenterol. Hepatol.* **2006**, *18*, 233–237. [[CrossRef](#)] [[PubMed](#)]
47. van Kleef, L.A.; Choi, H.S.J.; Brouwer, W.P.; Hansen, B.E.; Patel, K.; de Man, R.A.; Janssen, H.L.A.; de Knegt, R.J.; Sonneveld, M.J. Metabolic dysfunction-associated fatty liver disease increases risk of adverse outcomes in patients with chronic hepatitis B. *JHEP Rep.* **2021**, *3*, 100350. [[CrossRef](#)] [[PubMed](#)]
48. Huang, S.C.; Su, T.H.; Tseng, T.C.; Chen, C.L.; Hsu, S.J.; Liao, S.H.; Hong, C.M.; Liu, C.H.; Lan, T.Y.; Yang, H.C.; et al. Distinct effects of hepatic steatosis and metabolic dysfunction on the risk of hepatocellular carcinoma in chronic hepatitis B. *Hepatol. Int.* **2023**, *17*, 1139–1149. [[CrossRef](#)]
49. Kim, K.; Choi, S.; Park, S.M. Association of fasting serum glucose level and type 2 diabetes with hepatocellular carcinoma in men with chronic hepatitis B infection: A large cohort study. *Eur. J. Cancer* **2018**, *102*, 103–113. [[CrossRef](#)] [[PubMed](#)]
50. Kang, S.H.; Cho, Y.; Jeong, S.W.; Kim, S.U.; Lee, J.W.; Korean NAFLD Study Group. From nonalcoholic fatty liver disease to metabolic-associated fatty liver disease: Big wave or ripple? *Clin. Mol. Hepatol.* **2021**, *27*, 257–269. [[CrossRef](#)]
51. Wang, X.; Wei, S.; Wei, Y.; Wang, X.; Xiao, F.; Feng, Y.; Zhu, Q. The impact of concomitant metabolic dysfunction-associated fatty liver disease on adverse outcomes in patients with hepatitis B cirrhosis: A propensity score matching study. *Eur. J. Gastroenterol. Hepatol.* **2023**, *35*, 889–898. [[CrossRef](#)] [[PubMed](#)]
52. Zhang, P.; Liu, Z.; Fan, H.; Shi, T.; Han, X.; Suo, C.; Chen, X.; Zhang, T. Positive hepatitis B core antibody is associated with advanced fibrosis and mortality in nonalcoholic fatty liver disease. *Eur. J. Gastroenterol. Hepatol.* **2023**, *35*, 294–301. [[CrossRef](#)] [[PubMed](#)]
53. Kim, D.; Konyin, P.; Sandhu, K.K.; Dennis, B.B.; Cheung, A.C.; Ahmed, A. Metabolic dysfunction-associated fatty liver disease is associated with increased all-cause mortality in the United States. *J. Hepatol.* **2021**, *75*, 1284–1291. [[CrossRef](#)] [[PubMed](#)]
54. Cai, J.; Zhang, X.J.; Ji, Y.X.; Zhang, P.; She, Z.G.; Li, H. Nonalcoholic Fatty Liver Disease Pandemic Fuels the Upsurge in Cardiovascular Diseases. *Circ. Res.* **2020**, *126*, 679–704. [[CrossRef](#)] [[PubMed](#)]
55. Tapper, E.B.; Lok, A.S. Use of Liver Imaging and Biopsy in Clinical Practice. *N. Engl. J. Med.* **2017**, *377*, 756–768. [[CrossRef](#)] [[PubMed](#)]
56. Middleton, M.S.; Heba, E.R.; Hooker, C.A.; Bashir, M.R.; Fowler, K.J.; Sandrasegaran, K.; Brunt, E.M.; Kleiner, D.E.; Doo, E.; Van Natta, M.L.; et al. Agreement between Magnetic Resonance Imaging Proton Density Fat Fraction Measurements and Pathologist-Assigned Steatosis Grades of Liver Biopsies from Adults with Nonalcoholic Steatohepatitis. *Gastroenterology* **2017**, *153*, 753–761. [[CrossRef](#)] [[PubMed](#)]
57. Gu, J.; Liu, S.; Du, S.; Zhang, Q.; Xiao, J.; Dong, Q.; Xin, Y. Diagnostic value of MRI-PDFF for hepatic steatosis in patients with non-alcoholic fatty liver disease: A meta-analysis. *Eur. Radiol.* **2019**, *29*, 3564–3573. [[CrossRef](#)] [[PubMed](#)]
58. Karlas, T.; Petroff, D.; Sasso, M.; Fan, J.G.; Mi, Y.Q.; de Lédinghen, V.; Kumar, M.; Lupsor-Platon, M.; Han, K.H.; Cardoso, A.C.; et al. Individual patient data meta-analysis of controlled attenuation parameter (CAP) technology for assessing steatosis. *J. Hepatol.* **2017**, *66*, 1022–1030. [[CrossRef](#)]
59. Xu, L.; Lu, W.; Li, P.; Shen, F.; Mi, Y.Q.; Fan, J.G. A comparison of hepatic steatosis index, controlled attenuation parameter and ultrasound as noninvasive diagnostic tools for steatosis in chronic hepatitis B. *Dig. Liver Dis.* **2017**, *49*, 910–917. [[CrossRef](#)]
60. Liang, J.; Liu, F.; Wang, F.; Han, T.; Jing, L.; Ma, Z.; Gao, Y. A Noninvasive Score Model for Prediction of NASH in Patients with Chronic Hepatitis B and Nonalcoholic Fatty Liver Disease. *Biomed. Res. Int.* **2017**, *2017*, 8793278. [[CrossRef](#)]
61. Newsome, P.N.; Sasso, M.; Deeks, J.J.; Paredes, A.; Boursier, J.; Chan, W.K.; Yilmaz, Y.; Czernichow, S.; Zheng, M.H.; Wong, V.W.; et al. FibroScan-AST (FAST) score for the non-invasive identification of patients with non-alcoholic steatohepatitis with significant activity and fibrosis: A prospective derivation and global validation study. *Lancet Gastroenterol. Hepatol.* **2020**, *5*, 362–373, Erratum in *Lancet Gastroenterol. Hepatol.* **2020**, *5*, e3. [[CrossRef](#)] [[PubMed](#)]
62. Charatcharoenwitthaya, P.; Pongpaibul, A.; Kaosombatwattana, U.; Bhanthumkomol, P.; Bandidniyamanon, W.; Pausawasdi, N.; Tanwandee, T. The prevalence of steatohepatitis in chronic hepatitis B patients and its impact on disease severity and treatment response. *Liver Int.* **2017**, *37*, 542–551. [[CrossRef](#)] [[PubMed](#)]
63. Li, J.; Le, A.K.; Chaung, K.T.; Henry, L.; Hoang, J.K.; Cheung, R.; Nguyen, M.H. Fatty liver is not independently associated with the rates of complete response to oral antiviral therapy in chronic hepatitis B patients. *Liver Int.* **2020**, *40*, 1052–1061. [[CrossRef](#)]
64. Chen, J.; Wang, M.L.; Long, Q.; Bai, L.; Tang, H. High value of controlled attenuation parameter predicts a poor antiviral response in patients with chronic hepatitis B. *Hepatobiliary Pancreat. Dis. Int.* **2017**, *16*, 370–374. [[CrossRef](#)] [[PubMed](#)]
65. Jin, X.; Chen, Y.P.; Yang, Y.D.; Li, Y.M.; Zheng, L.; Xu, C.Q. Association between hepatic steatosis and entecavir treatment failure in Chinese patients with chronic hepatitis B. *PLoS ONE* **2012**, *7*, e34198. [[CrossRef](#)] [[PubMed](#)]
66. Younossi, Z.M.; Corey, K.E.; Lim, J.K. AGA clinical practice update on lifestyle modification using diet and exercise to achieve weight loss in the management of nonalcoholic fatty liver disease: Expert review. *Gastroenterology* **2021**, *160*, 912–918. [[CrossRef](#)]
67. Dufour, J.F.; Anstee, Q.M.; Bugianesi, E.; Harrison, S.; Loomba, R.; Paradis, V.; Tilg, H.; Wong, V.W.; Zelber-Sagi, S. Current therapies and new developments in NASH. *Gut* **2022**, *71*, 2123–2134. [[CrossRef](#)] [[PubMed](#)]

68. Newsome, P.N.; Buchholtz, K.; Cusi, K.; Linder, M.; Okanou, T.; Ratziu, V.; Sanyal, A.J.; Sejing, A.S.; Harrison, S.A.; NN9931-4296 Investigators. A placebo-controlled trial of subcutaneous Semaglutide in nonalcoholic steatohepatitis. *N. Engl. J. Med.* **2021**, *384*, 1113–1124. [[CrossRef](#)]
69. Francque, S.M.; Bedossa, P.; Ratziu, V.; Anstee, Q.M.; Bugianesi, E.; Sanyal, A.J.; Loomba, R.; Harrison, S.A.; Balabanska, R.; Mateva, L.; et al. A randomized, controlled trial of the pan-PPAR agonist Lanifibranor in NASH. *N. Engl. J. Med.* **2021**, *385*, 1547–1558. [[CrossRef](#)]
70. Harrison, S.A.; Bashir, M.R.; Guy, C.D.; Zhou, R.; Moylan, C.A.; Frias, J.P.; Alkhoury, N.; Bansal, M.B.; Baum, S.; Neuschwander-Tetri, B.A.; et al. Resmetirom (MGL-3196) for the treatment of non-alcoholic steatohepatitis: A multicentre, randomised, double-blind, placebo-controlled, phase 2 trial. *Lancet* **2019**, *394*, 2012–2024. [[CrossRef](#)]
71. Younossi, Z.M.; Stepanova, M.; Taub, R.A.; Barbone, J.M.; Harrison, S.A. Hepatic Fat Reduction Due to Resmetirom in Patients with Nonalcoholic Steatohepatitis Is Associated with Improvement of Quality of Life. *Clin. Gastroenterol. Hepatol.* **2022**, *20*, 1354–1361.e7. [[CrossRef](#)] [[PubMed](#)]
72. Younossi, Z.M.; Ratziu, V.; Loomba, R.; Rinella, M.; Anstee, Q.M.; Goodman, Z.; Bedossa, P.; Geier, A.; Beckebaum, S.; Newsome, P.N.; et al. Obeticholic acid for the treatment of non-alcoholic steatohepatitis: Interim analysis from a multicentre, randomised, placebo-controlled phase 3 trial. *Lancet* **2019**, *394*, 2184–2196. [[CrossRef](#)] [[PubMed](#)]
73. Younossi, Z.M.; Stepanova, M.; Nader, F.; Loomba, R.; Anstee, Q.M.; Ratziu, V.; Harrison, S.; Sanyal, A.J.; Schattenberg, J.M.; Barritt, A.S.; et al. Obeticholic acid impact on quality of life in patients with nonalcoholic steatohepatitis: REGENERATE 18-month interim analysis. *Clin. Gastroenterol. Hepatol.* **2022**, *20*, 2050–2058.e12. [[CrossRef](#)] [[PubMed](#)]
74. Wong, G.L.; Chan, H.L.; Yu, Z.; Chan, A.W.; Choi, P.C.; Chim, A.M.; Chan, H.Y.; Tse, C.H.; Wong, V.W. Coincidental metabolic syndrome increases the risk of liver fibrosis progression in patients with chronic hepatitis B—A prospective cohort study with paired transient elastography examinations. *Aliment. Pharmacol. Ther.* **2014**, *39*, 883–893. [[CrossRef](#)] [[PubMed](#)]
75. Fan, R.; Niu, J.; Ma, H.; Xie, Q.; Cheng, J.; Rao, H.; Dou, X.; Xie, J.; Zhao, W.; Peng, J.; et al. Association of central obesity with hepatocellular carcinoma in patients with chronic hepatitis B receiving antiviral therapy. *Aliment. Pharmacol. Ther.* **2021**, *54*, 329–338. [[CrossRef](#)] [[PubMed](#)]
76. Chan, K.E.; Ng, C.H.; Fu, C.E.; Quek, J.; Kong, G.; Goh, Y.J.; Zeng, R.W.; Tseng, M.; Aggarwal, M.; Nah, B.; et al. The Spectrum and Impact of Metabolic Dysfunction in MAFLD: A Longitudinal Cohort Analysis of 32,683 Overweight and Obese Individuals. *Clin. Gastroenterol. Hepatol.* **2023**, *21*, 2560–2569.e15. [[CrossRef](#)]
77. Polaris Observatory HCV Collaborators. Global prevalence and genotype distribution of hepatitis C virus infection in 2015: A modeling study. *Lancet Gastroenterol. Hepatol.* **2017**, *2*, 161–176. [[CrossRef](#)]
78. Gower, E.; Estes, C.; Blach, S.; Razavi-Shearer, K.; Razavi, H. Global epidemiology and genotype distribution of the hepatitis C virus infection. *J. Hepatol.* **2014**, *61* (Suppl. 1), S45–S57. [[CrossRef](#)]
79. Moucari, R.; Asselah, T.; Cazals-Hatem, D.; Voitot, H.; Boyer, N.; Ripault, M.P.; Sobesky, R.; Martinot-Peignoux, M.; Maylin, S.; Nicolas-Chanoine, M.H.; et al. Insulin resistance in chronic hepatitis C: Association with genotypes 1 and 4, serum HCV RNA level, and liver fibrosis. *Gastroenterology* **2008**, *134*, 416–423. [[CrossRef](#)]
80. Adinolfi, L.E.; Rinaldi, L.; Guerrero, B.; Restivo, L.; Marrone, A.; Giordano, M.; Zampino, R. NAFLD and NASH in HCV Infection: Prevalence and Significance in Hepatic and Extrahepatic Manifestations. *Int. J. Mol. Sci.* **2016**, *17*, 803. [[CrossRef](#)]
81. Marzouk, D.; Sass, J.; Bakr, I.; El Hosseiny, M.; Abdel-Hamid, M.; Rekaewicz, C.; Chaturvedi, N.; Mohamed, M.K.; Fontanet, A. Metabolic and cardiovascular risk profiles and hepatitis C virus infection in rural Egypt. *Gut* **2007**, *56*, 1105–1110. [[CrossRef](#)] [[PubMed](#)]
82. Tsai, P.S.; Cheng, Y.M.; Wang, C.C.; Kao, J.H. The impact of concomitant hepatitis C virus infection on liver and cardiovascular risks in patients with metabolic-associated fatty liver disease. *Eur. J. Gastroenterol. Hepatol.* **2023**, *35*, 1278–1283. [[CrossRef](#)] [[PubMed](#)]
83. Al-Omary, A.; Byth, K.; Weltman, M.; George, J.; Eslam, M. The importance and impact of recognizing metabolic dysfunction-associated fatty liver disease in patients with chronic hepatitis C. *J. Dig. Dis.* **2022**, *23*, 33–43. [[CrossRef](#)] [[PubMed](#)]
84. Younossi, Z.; Anstee, Q.M.; Marietti, M.; Hardy, T.; Henry, L.; Eslam, M.; George, J.; Bugianesi, E. Global burden of NAFLD and NASH: Trends, predictions, risk factors, and prevention. *Nat. Rev. Gastroenterol. Hepatol.* **2018**, *15*, 11–20. [[CrossRef](#)] [[PubMed](#)]
85. Negro, F. Facts and fictions of HCV and comorbidities: Steatosis, diabetes mellitus, and cardiovascular diseases. *J. Hepatol.* **2014**, *61* (Suppl. 1), S69–S78. [[CrossRef](#)] [[PubMed](#)]
86. Miyanari, Y.; Atsuzawa, K.; Usuda, N.; Watashi, K.; Hishiki, T.; Zayas, M.; Bartenschlager, R.; Wakita, T.; Hijikata, M.; Shimotohno, K. The lipid droplet is an important organelle for hepatitis C virus production. *Nat. Cell Biol.* **2007**, *9*, 1089–1097, Erratum in *Nat. Cell Biol.* **2007**, *9*, 1216. [[CrossRef](#)] [[PubMed](#)]
87. André, P.; Komurian-Pradel, F.; Deforges, S.; Perret, M.; Berland, J.L.; Sodoyer, M.; Pol, S.; Bréchet, C.; Paranhos-Baccalà, G.; Lotteau, V. Characterization of low- and very-low-density hepatitis C virus RNA-containing particles. *J. Virol.* **2002**, *76*, 6919–6928. [[CrossRef](#)]
88. Kapadia, S.B.; Chisari, F.V. Hepatitis C virus RNA replication is regulated by host geranylgeranylation and fatty acids. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 2561–2566. [[CrossRef](#)]
89. Yamane, D.; McGivern, D.R.; Wauthier, E.; Yi, M.; Madden, V.J.; Welsch, C.; Antes, I.; Wen, Y.; Chugh, P.E.; McGee, C.E.; et al. Regulation of the hepatitis C virus RNA replicase by endogenous lipid peroxidation. *Nat. Med.* **2014**, *20*, 927–935. [[CrossRef](#)]

90. Hofmann, S.; Krajewski, M.; Scherer, C.; Scholz, V.; Mordhorst, V.; Truschow, P.; Schöbel, A.; Reimer, R.; Schwudke, D.; Herker, E. Complex lipid metabolic remodeling is required for efficient hepatitis C virus replication. *Biochim. Biophys. Acta Mol. Cell Biol. Lipids.* **2018**, *1863*, 1041–1056. [[CrossRef](#)]
91. Abomughaid, M.; Tay, E.S.E.; Pickford, R.; Malladi, C.; Read, S.A.; Coorsen, J.R.; Gloss, B.S.; George, J.; Douglas, M.W. PEMT Mediates Hepatitis C Virus-Induced Steatosis, Explains Genotype-Specific Phenotypes and Supports Virus Replication. *Int. J. Mol. Sci.* **2023**, *24*, 8781. [[CrossRef](#)] [[PubMed](#)]
92. Coppola, N.; Rosa, Z.; Cirillo, G.; Stanzione, M.; Macera, M.; Boemio, A.; Grandone, A.; Pisaturo, M.; Marrone, A.; Adinolfi, L.E.; et al. TM6SF2 E167K variant is associated with severe steatosis in chronic hepatitis C, regardless of PNPLA3 polymorphism. *Liver Int.* **2015**, *35*, 1959–1963. [[CrossRef](#)] [[PubMed](#)]
93. Cheng, Y.; Dharancy, S.; Malapel, M.; Desreumaux, P. Hepatitis C virus infection down-regulates the expression of peroxisome proliferator-activated receptor alpha and carnitine palmitoyl acyl-CoA transferase 1A. *World J. Gastroenterol.* **2005**, *11*, 7591–7596. [[CrossRef](#)]
94. Okuda, M.; Li, K.; Beard, M.R.; Showalter, L.A.; Scholle, F.; Lemon, S.M.; Weinman, S.A. Mitochondrial injury, oxidative stress, and antioxidant gene expression are induced by hepatitis C virus core protein. *Gastroenterology* **2002**, *122*, 366–375. [[CrossRef](#)]
95. Kawaguchi, T.; Yoshida, T.; Harada, M.; Hisamoto, T.; Nagao, Y.; Ide, T.; Taniguchi, E.; Kumemura, H.; Hanada, S.; Maeyama, M.; et al. Hepatitis C virus down-regulates insulin receptor substrates 1 and 2 through up-regulation of suppressor of cytokine signaling 3. *Am. J. Pathol.* **2004**, *165*, 1499–1508. [[CrossRef](#)] [[PubMed](#)]
96. Miquilena-Colina, M.E.; Lima-Cabello, E.; Sánchez-Campos, S.; García-Mediavilla, M.V.; Fernández-Bermejo, M.; Lozano-Rodríguez, T.; Vargas-Castrillón, J.; Buqué, X.; Ochoa, B.; Aspichueta, P.; et al. Hepatic fatty acid translocase CD36 upregulation is associated with insulin resistance, hyperinsulinaemia and increased steatosis in non-alcoholic steatohepatitis and chronic hepatitis C. *Gut* **2011**, *60*, 1394–1402. [[CrossRef](#)]
97. Serfaty, L.; Andreani, T.; Giral, P.; Carbonell, N.; Chazouillères, O.; Poupon, R. Hepatitis C virus induced hypobetalipoproteinemia: A possible mechanism for steatosis in chronic hepatitis C. *J. Hepatol.* **2001**, *34*, 428–434. [[CrossRef](#)]
98. Jackel-Cram, C.; Qiao, L.; Xiang, Z.; Brownlie, R.; Zhou, Y.; Babiuk, L.; Liu, Q. Hepatitis C virus genotype-3a core protein enhances sterol regulatory element-binding protein-1 activity through the phosphoinositide 3-kinase-Akt-2 pathway. *J. Gen. Virol.* **2010**, *91 Pt 6*, 1388–1395. [[CrossRef](#)]
99. Clément, S.; Peyrou, M.; Sanchez-Pareja, A.; Bourgoin, L.; Ramadori, P.; Suter, D.; Vinciguerra, M.; Guilloux, K.; Pascarella, S.; Rubbia-Brandt, L.; et al. Down-regulation of phosphatase and tensin homolog by hepatitis C virus core 3a in hepatocytes triggers the formation of large lipid droplets. *Hepatology* **2011**, *54*, 38–49. [[CrossRef](#)]
100. de Gottardi, A.; Paziienza, V.; Pugnale, P.; Bruttin, F.; Rubbia-Brandt, L.; Juge-Aubry, C.E.; Meier, C.A.; Hadengue, A.; Negro, F. Peroxisome proliferator-activated receptor-alpha and -gamma mRNA levels are reduced in chronic hepatitis C with steatosis and genotype 3 infection. *Aliment. Pharmacol. Ther.* **2006**, *23*, 107–114. [[CrossRef](#)]
101. Dharancy, S.; Malapel, M.; Perlemuter, G.; Roskams, T.; Cheng, Y.; Dubuquoy, L.; Podevin, P.; Conti, F.; Canva, V.; Philippe, D.; et al. Impaired expression of the peroxisome proliferator-activated receptor alpha during hepatitis C virus infection. *Gastroenterology* **2005**, *128*, 334–342. [[CrossRef](#)] [[PubMed](#)]
102. Zylberberg, H.; Rimaniol, A.C.; Pol, S.; Masson, A.; De Groote, D.; Berthelot, P.; Bach, J.F.; Bréchet, C.; Zavala, F. Soluble tumor necrosis factor receptors in chronic hepatitis C: A correlation with histological fibrosis and activity. *J. Hepatol.* **1999**, *30*, 185–191. [[CrossRef](#)] [[PubMed](#)]
103. Sasaki, R.; Devhare, P.B.; Steele, R.; Ray, R.; Ray, R.B. Hepatitis C virus-induced CCL5 secretion from macrophages activates hepatic stellate cells. *Hepatology* **2017**, *66*, 746–757. [[CrossRef](#)] [[PubMed](#)]
104. Fartoux, L.; Chazouillères, O.; Wendum, D.; Poupon, R.; Serfaty, L. Impact of steatosis on progression of fibrosis in patients with mild hepatitis C. *Hepatology* **2005**, *41*, 82–87. [[CrossRef](#)] [[PubMed](#)]
105. Ohata, K.; Hamasaki, K.; Toriyama, K.; Matsumoto, K.; Saeki, A.; Yanagi, K.; Abiru, S.; Nakagawa, Y.; Shigeno, M.; Miyazoe, S.; et al. Hepatic steatosis is a risk factor for hepatocellular carcinoma in patients with chronic hepatitis C virus infection. *Cancer* **2003**, *97*, 3036–3043. [[CrossRef](#)] [[PubMed](#)]
106. Poynard, T.; Ratziu, V.; McHutchison, J.; Manns, M.; Goodman, Z.; Zeuzem, S.; Younossi, Z.; Albrecht, J. Effect of treatment with peginterferon or interferon alfa-2b and ribavirin on steatosis in patients infected with hepatitis C. *Hepatology* **2003**, *38*, 75–85. [[CrossRef](#)]
107. Adinolfi, L.E.; Restivo, L.; Zampino, R.; Guerrera, B.; Lonardo, A.; Ruggiero, L.; Riello, F.; Loria, P.; Florio, A. Chronic HCV infection is a risk of atherosclerosis. Role of HCV and HCV-related steatosis. *Atherosclerosis* **2012**, *221*, 496–502. [[CrossRef](#)]
108. Rau, M.; Buggisch, P.; Mauss, S.; Boeker, K.H.W.; Klinker, H.; Müller, T.; Stoehr, A.; Schattenberg, J.M.; Geier, A. Prognostic impact of steatosis in the clinical course of chronic HCV infection—Results from the German Hepatitis C-Registry. *PLoS ONE* **2022**, *17*, e0264741. [[CrossRef](#)]
109. Eslam, M.; Hashem, A.M.; Leung, R.; Romero-Gomez, M.; Berg, T.; Dore, G.J.; Chan, H.L.; Irving, W.L.; Sheridan, D.; Abate, M.L.; et al. Interferon-λ rs12979860 genotype and liver fibrosis in viral and non-viral chronic liver disease. *Nat. Commun.* **2015**, *6*, 6422. [[CrossRef](#)]
110. Krassenburg, L.A.P.; Maan, R.; Ramji, A.; Manns, M.P.; Cornberg, M.; Wedemeyer, H.; de Knegt, R.J.; Hansen, B.E.; Janssen, H.L.A.; de Man, R.A.; et al. Clinical outcomes following DAA therapy in patients with HCV-related cirrhosis depend on disease severity. *J. Hepatol.* **2021**, *74*, 1053–1063. [[CrossRef](#)]

111. Do, A.; Esserman, D.A.; Krishnan, S.; Lim, J.K.; Taddei, T.H.; Hauser RG 3rd Tate, J.P.; Re VL 3rd Justice, A.C. Excess weight gain after cure of hepatitis C infection with direct-acting antivirals. *J. Gen. Intern. Med.* **2020**, *35*, 2025–2034, Erratum in *J. Gen. Intern. Med.* **2020**, *35*, 3140. [[CrossRef](#)] [[PubMed](#)]
112. Tomasiewicz, K.; Flisiak, R.; Jaroszewicz, J.; Małkowski, P.; Pawłowska, M.; Piekarska, A.; Simon, K.; Zarebska-Michaluk, D. Recommendations of the Polish group of experts for HCV for the treatment of hepatitis C in 2023. *Clin. Exp. Hepatol.* **2023**, *9*, 1–8. [[CrossRef](#)] [[PubMed](#)]
113. Sun, D.; Dai, M.; Shen, S.; Li, C.; Yan, X. Analysis of naturally occurring resistance-associated variants to NS3/4A protein inhibitors, NS5A protein inhibitors, and NS5B polymerase inhibitors in patients with chronic hepatitis C. *Gene Expr.* **2018**, *18*, 63–69. [[CrossRef](#)] [[PubMed](#)]
114. Harrison, S.A.; Brunt, E.M.; Qazi, R.A.; Oliver, D.A.; Neuschwander-Tetri, B.A.; Di Bisceglie, A.M.; Bacon, B.R. Effect of significant histologic steatosis or steatohepatitis on response to antiviral therapy in patients with chronic hepatitis C. *Clin. Gastroenterol. Hepatol.* **2005**, *3*, 604–609. [[CrossRef](#)] [[PubMed](#)]
115. Malaguarnera, M.; Vacante, M.; Russo, C.; Gargante, M.P.; Giordano, M.; Bertino, G.; Neri, S.; Malaguarnera, M.; Galvano, F.; Li Volti, G. Rosuvastatin reduces nonalcoholic fatty liver disease in patients with chronic hepatitis C treated with α -interferon and ribavirin: Rosuvastatin reduces NAFLD in HCV patients. *Hepat. Mon.* **2011**, *11*, 92–98. [[PubMed](#)]
116. Look, M.P.; Gerard, A.; Rao, G.S.; Sudhop, T.; Fischer, H.P.; Sauerbruch, T.; Spengler, U. Interferon/antioxidant combination therapy for chronic hepatitis C—a controlled pilot trial. *Antiviral Res.* **1999**, *43*, 113–122. [[CrossRef](#)] [[PubMed](#)]
117. Houglum, K.; Venkataramani, A.; Lyche, K.; Chojkier, M. A pilot study of the effects of d-alpha-tocopherol on hepatic stellate cell activation in chronic hepatitis C. *Gastroenterology* **1997**, *113*, 1069–1073. [[CrossRef](#)]
118. Rout, G.; Nayak, B.; Patel, A.H.; Gunjan, D.; Singh, V.; Kedia, S. Shalimar Therapy with oral directly acting agents in hepatitis C infection is associated with reduction in fibrosis and increase in hepatic steatosis on transient elastography. *J. Clin. Exp. Hepatol.* **2019**, *9*, 207–214. [[CrossRef](#)]
119. Sun, H.Y.; Cheng, P.N.; Tseng, C.Y.; Tsai, W.J.; Chiu, Y.C.; Young, K.C. Favouring modulation of circulating lipoproteins and lipid loading capacity by direct antiviral agents grazoprevir/elbasvir or ledipasvir/sofosbuvir treatment against chronic HCV infection. *Gut* **2018**, *67*, 1342–1350. [[CrossRef](#)]
120. Hum, J.; Jou, J.H.; Green, P.K.; Berry, K.; Lundblad, J.; Hettinger, B.D.; Chang, M.; Ioannou, G.N. Improvement in glycemic control of type 2 diabetes after successful treatment of hepatitis C virus. *Diabetes Care* **2017**, *40*, 1173–1180. [[CrossRef](#)]
121. Butt, A.A.; Yan, P.; Shuaib, A.; Abou-Samra, A.B.; Shaikh, O.S.; Freiberg, M.S. Direct-acting antiviral therapy for HCV infection is associated with a reduced risk of cardiovascular disease events. *Gastroenterology* **2019**, *156*, 987–996.e8. [[CrossRef](#)] [[PubMed](#)]
122. Bhattacharya, D.; Aronsohn, A.; Price, J.; Lo Re, V.; AASLD-IDSAs HCV Guidance Panel. Hepatitis C Guidance 2023 Update: AASLD-IDSAs recommendations for testing, managing, and treating hepatitis C virus infection. *Clin. Infect. Dis.* **2023**, ciad319. [[CrossRef](#)] [[PubMed](#)]
123. Tao, X.; Chen, L.; Zhao, Y.; Liu, Y.; Shi, R.; Jiang, B.; Mi, Y.; Xu, L. A novel noninvasive diagnostic model of HBV-related inflammation in chronic hepatitis B virus infection patients with concurrent nonalcoholic fatty liver disease. *Front. Med.* **2022**, *9*, 862879. [[CrossRef](#)] [[PubMed](#)]
124. Zhang, J.; Lin, S.; Jiang, D.; Li, M.; Chen, Y.; Li, J.; Fan, J. Chronic hepatitis B and non-alcoholic fatty liver disease: Conspirators or competitors? *Liver Int.* **2020**, *40*, 496–508. [[CrossRef](#)] [[PubMed](#)]
125. Shaheen, A.A.; AlMattooq, M.; Yazdanfar, S.; Burak, K.W.; Swain, M.G.; Congly, S.E.; Borman, M.A.; Lee, S.S.; Myers, R.P.; Coffin, C.S. Tenofovir disoproxil fumarate significantly decreases serum lipoprotein levels compared with entecavir nucleos(t)ide analogue therapy in chronic hepatitis B carriers. *Aliment. Pharmacol. Ther.* **2017**, *46*, 599–604. [[CrossRef](#)] [[PubMed](#)]
126. Chang, F.M.; Wang, Y.P.; Lang, H.C.; Tsai, C.F.; Hou, M.C.; Lee, F.Y.; Lu, C.L. Statins decrease the risk of decompensation in hepatitis B virus- and hepatitis C virus-related cirrhosis: A population-based study. *Hepatology* **2017**, *66*, 896–907. [[CrossRef](#)] [[PubMed](#)]
127. Simon, T.G.; Duberg, A.S.; Aleman, S.; Hagstrom, H.; Nguyen, L.H.; Khalili, H.; Chung, R.T.; Ludvigsson, J.F. Lipophilic statins and risk for hepatocellular carcinoma and death in patients with chronic viral hepatitis: Results from a nationwide Swedish population. *Ann. Intern. Med.* **2019**, *171*, 318–327. [[CrossRef](#)] [[PubMed](#)]
128. Bar-Yishay, I.; Shaul, Y.; Shlomai, A. Hepatocyte metabolic signaling pathways and regulation of hepatitis B virus expression. *Liver Int.* **2011**, *31*, 282–290. [[CrossRef](#)]
129. Xiong, J.; Zhang, H.; Wang, Y.; Wang, A.; Bian, J.; Huang, H.; Zheng, Y.; Sang, X.; Xu, Y.; Lu, X.; et al. Hepatitis B virus infection and the risk of nonalcoholic fatty liver disease: A meta-analysis. *Oncotarget* **2017**, *8*, 107295–107302. [[CrossRef](#)]
130. Boeckmans, J.; Rombaut, M.; Demuyser, T.; Declerck, B.; Piérard, D.; Rogiers, V.; De Kock, J.; Waumans, L.; Magerman, K.; Cartuyvels, R.; et al. Infections at the nexus of metabolic-associated fatty liver disease. *Arch. Toxicol.* **2021**, *95*, 2235–2253. [[CrossRef](#)]
131. Wang, X.; Xie, Q. Metabolic dysfunction-associated fatty liver disease (MAFLD) and viral hepatitis. *J. Clin. Transl. Hepatol.* **2022**, *10*, 128–133. [[CrossRef](#)] [[PubMed](#)]

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