



Review

PAI-1: A Major Player in the Vascular Dysfunction in Obstructive Sleep Apnea?

Mohammad Badran ¹ and David Gozal ^{1,2,*}

¹ Department of Child Health and Child Health Research Institute, School of Medicine, University of Missouri, 400 N Keene St, Suite 010, Columbia, MO 65201, USA; mbadran@health.missouri.edu

² Department of Medical Pharmacology and Physiology, School of Medicine, University of Missouri, Columbia, MO 65201, USA

* Correspondence: gozald@health.missouri.edu; Tel.: +1-573-882-6882; Fax: +1-573-884-5179

Abstract: Obstructive sleep apnea is a chronic and prevalent condition that is associated with endothelial dysfunction, atherosclerosis, and imposes excess overall cardiovascular risk and mortality. Despite its high prevalence and the susceptibility of CVD patients to OSA-mediated stressors, OSA is still under-recognized and untreated in cardiovascular practice. Moreover, conventional OSA treatments have yielded either controversial or disappointing results in terms of protection against CVD, prompting the need for the identification of additional mechanisms and associated adjuvant therapies. Plasminogen activator inhibitor-1 (PAI-1), the primary inhibitor of tissue-type plasminogen activator (tPA) and urinary-type plasminogen activator (uPA), is a key regulator of fibrinolysis and cell migration. Indeed, elevated PAI-1 expression is associated with major cardiovascular adverse events that have been attributed to its antifibrinolytic activity. However, extensive evidence indicates that PAI-1 can induce endothelial dysfunction and atherosclerosis through complex interactions within the vasculature in an antifibrinolytic-independent manner. Elevated PAI-1 levels have been reported in OSA patients. However, the impact of PAI-1 on OSA-induced CVD has not been addressed to date. Here, we provide a comprehensive review on the mechanisms by which OSA and its most detrimental perturbation, intermittent hypoxia (IH), can enhance the transcription of PAI-1. We also propose causal pathways by which PAI-1 can promote atherosclerosis in OSA, thereby identifying PAI-1 as a potential therapeutic target in OSA-induced CVD.

Keywords: obstructive sleep apnea; intermittent hypoxia; plasminogen activator inhibitor-1; endothelial dysfunction; atherosclerosis



Citation: Badran, M.; Gozal, D. PAI-1: A Major Player in the Vascular Dysfunction in Obstructive Sleep Apnea? *Int. J. Mol. Sci.* **2022**, *23*, 5516. <https://doi.org/10.3390/ijms23105516>

Academic Editor: Shin Takasawa

Received: 13 April 2022

Accepted: 12 May 2022

Published: 15 May 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Obstructive sleep apnea (OSA) is a chronic condition affecting up to one billion people worldwide [1]. OSA is defined as a sleep-breathing disorder that involves a decrease or complete cessation of airflow despite ongoing efforts to breathe due to a collapsed upper airway. This leads to partial reductions (hypopneas) and complete pauses (apneas) in breathing that usually last between 10 and 30 s, but some may persist longer. This can lead to abrupt reductions in blood oxygen saturation, with oxygen levels falling as much as 40% or more in severe cases [2]. As a result, several pathological mechanisms ensue such as intermittent hypoxia (IH), sleep fragmentation, episodic hypercapnia, and increased intrathoracic pressure swings [3–5]. Consequently, these processes can induce major changes in the autonomic nervous system balance with both increased tonic and reactive sympathetic activity along with parasympathetic withdrawal, disruption of the hypothalamic–pituitary–adrenal-axis, systemic and cellular oxidative stress, and inflammation, fibrosis, and accelerated cellular senescence, all of which resulted in neurocognitive deficits, endothelial dysfunction, hypertension, and atherosclerosis [6–13]. Predictably, OSA is considered as an independent risk factor for cardiovascular disease (CVD) including coronary artery disease (CAD), ischemic

stroke, and myocardial infarction (MI) [14]. The majority of strokes and MIs seem to be prompted by atherothrombotic events along with compromised fibrinolytic activity, increasing the propensity for such events [15,16]. The fibrinolytic system is designed to cleave the insoluble polymeric network of fibrin from the vascular system to prevent clot overgrowth and vessel occlusion. Generally, plasminogen is activated by serine proteases plasminogen activators (PAs) including tissue-type PA (tPA) and urokinase-type PA (uPA) into plasmin, which in turn lyses the fibrin and other extracellular matrix components [17,18]. To prevent bleeding, plasminogen activator inhibitor-1 (PAI-1) is normally synthesized in equimolar amounts to PAs, forms a covalent bond with PAs, and stabilizes fibrin [19]. However, many processes including oxidative stress [20], inflammation [21], and fibrosis [22] can lead to elevated levels of PAI-1, which have been implicated in a multitude of diseases and conditions including CVD [23], cancer [24], metabolic disease [25], renal disease [26], behavioral and psychiatric conditions [27], and aging processes [28]. Furthermore, PAI-1 has been shown to induce endothelial dysfunction and atherosclerosis through antifibrinolytic-dependent mechanisms including inflammation [29], endothelial nitric oxide synthase (eNOS) inhibition [30], neointimal hyperplasia [31], and vascular senescence [28]. Despite the fact that PAI-1 levels are consistently elevated in OSA patients [32–42], and that OSA can trigger processes that can upregulate PAI-1 production, little to no attention has been given to PAI-1 as a biomarker or as a promoter of OSA-induced CVD in clinical practice. Here, we will summarize the mechanisms involved in upregulating PAI-1, the pathological role of PAI-1 in CVD, and underline the mechanisms by which OSA could upregulate PAI-1, thus, highlighting PAI-1 as a potential therapeutic target in OSA-induced CVD. Finally, we will discuss some of the therapeutic approaches to reduce PAI-1 levels, which may hold promise as adjuvant therapies in OSA since existing treatments (e.g., continuous positive airway pressure (CPAP)) appear to be ineffective in reversing or mitigating the frequency and severity of cardiovascular events in OSA patients [43,44].

2. PAI-1 Sources, Structure, and Function

PAI-1 can be synthesized by numerous types of cells including platelets, macrophages, adipocytes, hepatocytes, vascular smooth muscle cells, endothelial cells, and others [45–47]. Approximately 10% of the PAI-1 produced circulates in the blood or is deposited in the subendothelial matrix, while the rest is retained in platelets [48,49]. Platelets can de novo synthesize PAI-1 despite lacking nuclei through activated PAI-1 mRNA, with the synthesis rates being increased upon platelet activation [50]. The circulating PAI-1 fraction exists in its active conformation at levels of 5–50 ng/mL with large intra- and inter-personal variability, while platelet PAI-1 concentrations can reach up to 300 ng/mL with 50% shown to be biologically active [48,51,52]. Ultimately, PAI-1 plasma levels are increased under numerous pathological conditions [53]. The structure and function of PAI-1 have been extensively reviewed previously [25,54,55]. Briefly, PAI-1 is a single chain molecule with two interactive domains including a surface-exposed reactive center loop (RCL) that presents as a substrate peptide becoming the primary site for uPA/tPA binding, and a flexible joint region with helices D, E, and F that bind to vitronectin and stabilize PAI-1 in its active form while enhancing its binding affinity to uPA/tPA 200-fold [24,56–61]. PAI-1 exists in three distinct structurally and functionally distinct conformations including active, latent, and cleaved [54,62]. Unless bound to vitronectin, the active form can be readily converted to the more energetically favorable inactive latent form by internalizing the RCL, which may serve as a regulatory mechanism to prevent excessive anti-fibrinolysis [63–65]. However, the latent form can be reactivated. In its cleaved form, PAI-1 is still able to bind to other proteins with its helix, but its ability to inhibit uPA/tPA is abrogated [63]. As alluded to earlier, PAI-1 is a master regulator of the plasminogen system. PAI-1 can rapidly inactivate uPA/tPA with a second-order-rate constant between 10^6 and $10^7 \text{ m}^{-1} \text{ s}^{-1}$, forming a non-covalent Michaelis-like complex and eventually forming an ester bond between the serine residue of the protease and the carboxyl group of the P1 residue of PAI-1 [66,67]. PAI-1 also plays an important role in extracellular matrix (ECM) remodeling by indirectly modulating

the activity of matrix metalloproteinases (MMPs) [68]. Indeed, by inhibiting the plasmin activation required for the cleavage of pro-MMP, PAI-1 can block ECM degradation [55].

3. Mechanisms Involved in PAI-1 Upregulation

The human PAI-1 promoter shows a high degree of homology with mice and rats, suggesting that they are regulated by similar mechanisms. The 5'-flanking region contains a 'TATA' box with several transcription binding sites including hypoxia inducible factor-1 α (HIF-1 α), Smads, activator protein-1 (AP-1), specificity protein-1 (SP-1), and nuclear factor kappa B (NF- κ B). In the next section, we will discuss the major contributors to PAI-1 upregulation (Figure 1).

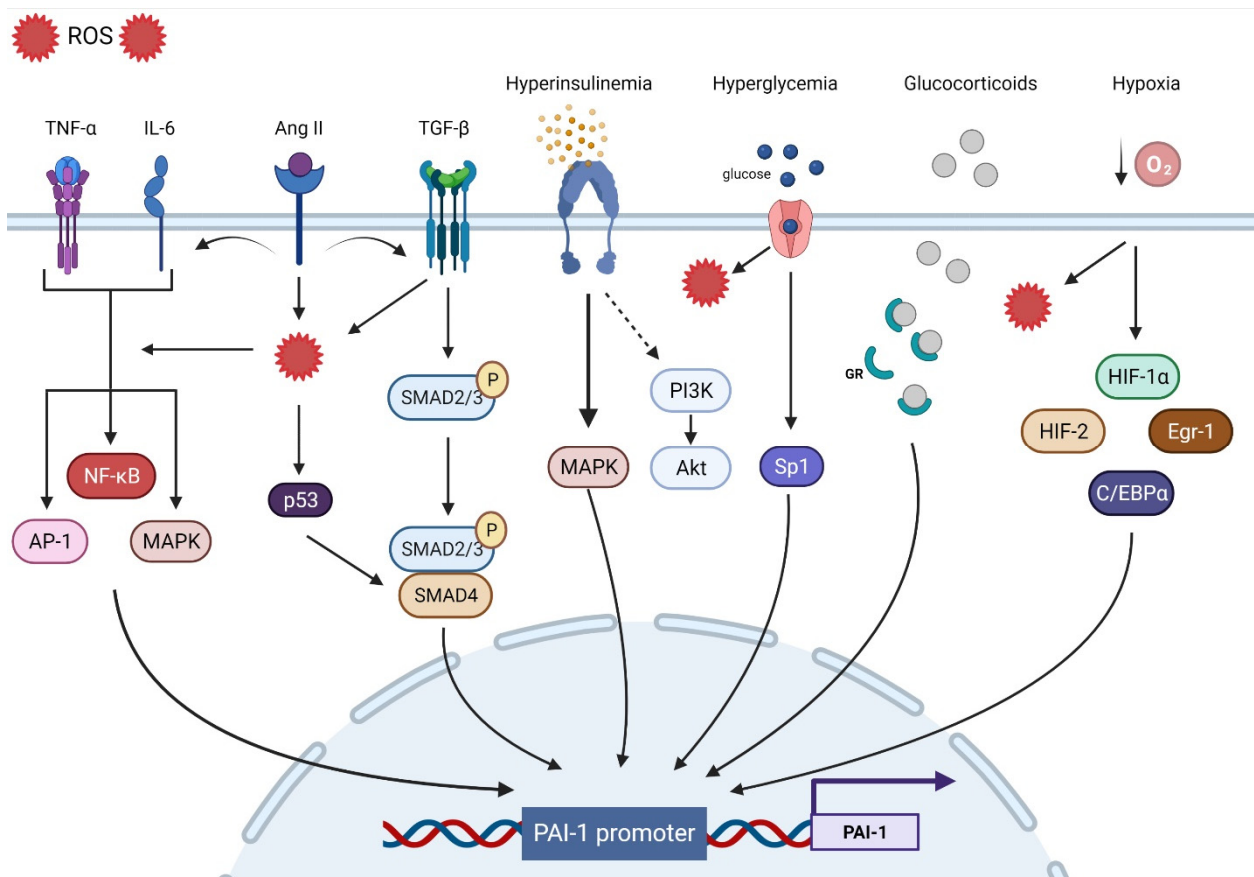


Figure 1. Transcriptional regulatory pathways implicated in PAI-1 synthesis.

3.1. Oxidative Stress

Oxidative stress (OS) is the end result of an imbalance between the production of oxidants and the capacity of the antioxidant system. Although they play an important role in regulating cellular function and signal transduction, free radicals such as reactive oxygen species (ROS) can be detrimental when produced in excess, given their ability to damage lipids, proteins, and DNA [69]. OS is undeniably a major contributor to multiorgan dysfunction in many disease states including CVD [70]. Indeed, ROS overproduction directly decreases nitric oxide (NO) bioavailability, uncouples eNOS, oxidizes low-density lipoprotein (OxLDL), and induces vascular inflammation [71]. OS is a significant upregulator of PAI-1 transcription. Indeed, incubating endothelial cells with H₂O₂ induced marked increases in PAI-1 mRNA and protein expression [72]. Conversely, the PAI-1 promoter was suppressed by up to 75% in the presence of antioxidants [73]. Furthermore, inhibiting NADPH oxidase, a major source of ROS, abolished the PAI-1 release and promoter activity in cultured endothelial cells [74]. Other experimental *in vitro* and *in vivo* studies performed in animal models as well as in humans have shown that the administration of antioxidants

can decrease PAI-1 expression [20,75–83]. Due to their intricate interactions with multiple signaling pathways and transcription factors, ROS are involved in most of the mechanisms regulating PAI-1 expression. For instance, ROS-induced PAI-1 increased transcription and expression is mediated through the activation of mitogen-activated protein kinase (MAPK) and NF- κ B pathways that are tightly involved in pro-fibrotic and pro-inflammatory pathways [74,84]. ROS signaling can also stimulate AP-1, HIF-1 α , and p53, all of which can increase the transcription of PAI-1 [85,86] (Figure 1).

3.2. Inflammation

Inflammation is a complex constellation of reactions between the host normal defense processes to internal and external stressors that have been implicated in many conditions and age-related diseases, especially in promoting atherosclerosis, a hallmark of CVD [87–89]. Low-grade inflammation induces endothelial dysfunction and subintimal cholesterol accumulation, leading to the upregulation of intercellular adhesion molecules and selectins that promote the binding and transmigration of inflammatory cells including monocytes and T-helper cells into the vessel wall. Infiltrating monocytes can transform into resident macrophages that express and activate inflammasomes that are key to the propagation of inflammation through the generation of multiple cytokines that amplify the inflammatory cascade within the vessel wall [89]. Coupled with enhanced ROS production, inflammation enters a vicious cycle in combination with OS, further aggravating atherosclerosis [90]. The link between inflammation and the fibrinolytic system is well-established. Experimental *in vitro* and *in vivo* studies as well as clinical studies have identified tumor necrosis factor- α (TNF- α) as a substantial contributor to increased PAI-1 expression [91–96]. In endothelial cells, TNF- α upregulated PAI-1 levels and was abolished by N-acetyl cysteine, indicating ROS as a mediator [73]. Administration of TNF- α in mice significantly increased the PAI-1 levels in adipose tissue, while obese mice treated with antibodies targeting TNF- α exhibited reduced plasma PAI-1 expression and adipose tissue-PAI-1 levels [97,98]. It is suggested that TNF- α can induce PAI-1 gene expression via redox-sensitive mechanisms triggering NF- κ B translocation and interaction with a regulatory region that is present on the PAI-1 promoter [21,96]. These data showcase the interplay between inflammation and OS and their integral role in upregulating PAI-1. Other pathways have been suggested in TNF- α -mediated PAI-1 induction including MAPK and protein kinase C [93]. Interleukin-6 (IL-6) is another inflammatory cytokine involved in PAI-1 upregulation. IL-6 is an acute phase inflammatory reaction protein that can induce C-reactive protein (CRP) synthesis and cortisol production [99]. Animals injected with IL-6 had significant increases in PAI-1 levels, while using IL-6 receptor antagonist reduced the PAI-1 expression in COVID patients [100,101]. IL-6 can activate NF- κ B and MAPK, leading to increased PAI-1 transcription [55,102] (Figure 1).

3.3. Fibrosis

Progressive vascular fibrosis is a prominent feature of atherosclerosis and CVD [103]. Transforming growth factor- β (TGF- β) is a major regulator of the fibroproliferative response to tissue damage [104]. TGF- β can control cell proliferation and migration, matrix synthesis, calcification, and immunomodulation, all being integral components of atherosclerosis [105]. TGF- β can be produced by all cells composing the vasculature and can also be produced in atherosclerotic lesions. However, TGF- β is mainly released by activated platelets adherent to activated endothelium. As a result, TGF- β induces the transcription of platelet-derived growth factor, collagens, fibronectin, and thrombospondins while suppressing the breakdown of ECM by inducing the transcription of PAI-1 and metalloprotease inhibitors, leading to the accumulation of the fibrotic matrix followed by calcification [103,105]. Overall, TGF- β production in atherosclerotic lesions can result in negative remodeling and progressive narrowing of the arteries, leading to MI and stroke [103]. TGF- β is considered as one of the major drivers of PAI-1 upregulation. *In vitro* studies have shown that PAI-1 expression is induced by TGF- β in various types of cells, while elevated PAI-1 levels are associated

with enhanced TGF- β expression and ECM deposition under many pathological conditions [22,106–111]. TGF- β can induce PAI-1 production through the activation of the Smad pathway via the nuclear translocation of the Smad 2/3 and Smad 4 complex and binding to the PAI-1 promoter [112]. Interestingly, TGF- β can induce ROS production and suppress antioxidant activity in various types of cells and in vivo [113–119]. Thus, PAI-1 expression can also be mediated through TGF- β -induced ROS production. MAPK and NF- κ B signaling are redox sensitive pathways that can be induced by TGF- β [55,114,120,121]. In TGF- β treated cells, inhibition of NADPH oxidase blocked TGF- β induced MAPK activated PAI-1 expression [85]. Furthermore, TGF- β can upregulate PAI-1 through Smad interactions with p53 and the transcription factors AP-1 and SP-1 [22,85,122] (Figure 1).

3.4. Hypoxia

Hypoxia triggers many cellular processes both in physiological and pathological conditions and has been associated with vascular dysfunction and atherosclerosis [123]. Vascular wall cells respond to hypoxia by tuning metabolism, angiogenesis, inflammation, cell survival signaling, and ultimately, may develop endothelial dysfunction [124,125]. The main regulator of such processes is the transcription factor HIF-1 α . Under normoxic conditions, HIF-1 α is constantly degraded, whereas hypoxia promotes its stability and transcriptional activity [126]. However, HIF-1 α is stabilized in atherosclerotic lesions even under normoxic conditions. ROS, OxLDL, NF- κ B, and other factors are promoted by HIF-1 α and in return, enhance HIF-1 α stability [123]. PAI-1 is one of the main transcriptional targets of HIF-1 α . Indeed, cells exposed to hypoxia display increased PAI-1 mRNA expression and stability [127–131]. HIF-1 α knockdown limited irradiation-induced PAI-1 upregulation in endothelial cells [132]. ROS production in endothelial cells induced HIF-1 α and subsequently PAI-1 production [133,134]. Additionally, ROS induced HIF-1 α via a specific NF- κ B binding site in the HIF-1 promoter [135]. Indeed, upregulation of the pulmonary artery smooth muscle PAI-1 was induced by an NF- κ B-dependent HIF-1 α transcription [136]. Although HIF-1 α appears to dominate the PAI-1 transcriptional response to hypoxia, other pathways including HIF-2 α , early growth response protein-1 (Egr-1), and CCAAT-enhancer-binding protein- α (C/EBP α) can augment this response independently of HIF-1 α [137,138] (Figure 1).

3.5. Hormones

Insulin can directly stimulate PAI-1 production in hepatocytes, an effect that is augmented by the presence of insulin-like growth factor [139,140]. The same effect was observed in cocultured endothelial cells and smooth muscle cells (SMCs) [141]. In the context of insulin resistance, compensatory hyperinsulinemia decreases the activity of the PI3-K/Akt pathway and augments the MAPK/ERK pathway, a major driver of PAI-1 production [142,143]. Elevated levels of glucose can also directly increase the expression of PAI-1 in endothelial cells and SMC through an effect on two adjacent Sp1 sites [122]. These data explain the elevated levels of PAI-1 in conditions characterized by hyperinsulinemia and hyperglycemia such as obesity, metabolic syndrome, and type 2 diabetes mellitus [25,144,145]. Under intense stress, very high levels of glucocorticoid hormones can increase the production of PAI-1 protein [146]. Glucocorticoids bind to their cytoplasmic glucocorticoid receptor and the complex is translocated to the nucleus and directly binds to the glucocorticoid response element that enhances PAI-1 transcription [86]. Angiotensin II, a major vasoconstrictor and contributor to hypertension upregulated by the activation of the renin–angiotensin–aldosterone system (RAAS), has been reported to induce PAI-1 expression in cultured endothelial cells in an angiotensin receptor independent manner [147]. Ang II can increase ROS production, fibrotic signaling (TGF- β), and inflammation, all of which can increase the expression of PAI-1 [148–150] (Figure 1).

4. Pathological Role PAI-1 Role in Cardiovascular Disease

In humans, PAI-1 deficiency is a rare disorder that is attributed to mutations in the SERPINE1 gene that leads to either the absence of PAI-1 plasma detectable levels or the production of a non-functional PAI-1 protein [151–153]. The disease is characterized mainly by delayed mild to moderate bleeding following a traumatic event or injury, or during surgeries and in the contest of pregnancy complications [154,155]. Difficulty in establishing an accurate diagnosis stems from the fact that the PAI-1 activity assay detects elevated levels but is much less performant at the lowest detectable ranges [155]. Thus, the true prevalence of this rare condition is not well-established. On the other hand, two frequent PAI-1 gene polymorphisms have been shown to affect the PAI-1 levels [156,157]. The 4G/5G polymorphism that refers to single guanosine insertion/deletion at the transcription site is associated with higher PAI-1 activity, and the G/A polymorphism that refers to the single nucleotide substitution of guanine with adenine upstream of the transcription site leads to increases in the transcription rate [157,158]. Several clinical studies have suggested that PAI-1 polymorphisms (possibly leading to increased PAI-1 levels or activity) are an independent risk factor for major adverse cardiovascular events (MACE) including atherosclerosis, CAD, MI, stroke, and venous thrombosis [159–166]. Even in the absence of polymorphisms, elevated PAI-1 levels have been linked to the aforementioned events [23,167–170]. The Framingham Heart Study showed that PAI-1 levels are predictive of CVD events after accounting for established risk factors, while a serial increase in PAI-1 is associated with a further increase in risk [168]. Additionally, a recent meta-analysis identified 38 articles between 1991 and 2016 that reported PAI-1 levels in 11,557 patients. In studies assessing PAI-1 concentrations and activity levels, 15.1% and 29.6% of the patients included in these studies experienced MACE, respectively. Furthermore, patients with MACE had higher PAI-1 concentrations with a mean difference of 6.11 ng/mL [171]. However, not all studies confirmed a direct link between the elevated PAI-1 levels and CVD, especially after adjusting for the confounding factors [172–175]. It is very likely that the absence of such an association may be explained by the fact that factors such as age, sex, obesity, insulin resistance, and diabetes are positively correlated with plasma PAI-1 levels [25,175–178].

In order to comprehensively evaluate the pathological role of PAI-1, several mouse models have been developed. These murine lines are either completely PAI-1 deficient (PAI-1^{-/-}) or overexpress native or stabilized human or murine PAI-1. PAI-1^{-/-} mice develop normally with no apparent macroscopic or microscopic histological abnormalities [179]. Although the deficiency of PAI-1 has been shown to increase the resistance to thrombosis and is protective against atherosclerosis [180–182], other studies have shown that the absence of PAI-1 can promote atherosclerosis and cardiac fibrosis [183–185]. It is suggested that abrogating the controlling effect of PAI-1 on the plasminogen system can contribute to the atherogenic and fibrotic role of plasmin, since the latter can mediate inflammation, foam cell formation, and ECM remodeling [186–188]. These data highlight the importance of the balance required between all the components of the fibrinolytic system to maintain homeostasis. For mice overexpressing PAI-1, transgenic mice overexpressing a stable active form of human PAI-1 (PAI-1 stab) display phenotypic abnormalities including alopecia and hepatosplenomegaly with age-dependent coronary arterial thrombosis, even in the absence of severe hypercholesterolemia [189,190]. In addition, transgenic mice overexpressing native human PAI-1 develop venous, but not arterial thrombosis [191]. For transgenic mice overexpressing stable murine PAI-1, they appear to suffer from an occasional tail autoamputation with no evidence of thrombosis [191]. The phenotypic differences observed could be attributable to cross-species differences and to the nature of the stable variant [54]. Although the major vascular pathological role of PAI-1 is related to its ability to create a hypofibrinolytic environment, the function of the PAI-1 extends beyond controlling fibrinolysis through the inhibition of plasmin formation as plasmin is involved in other physiological processes including ECM remodeling, angiogenesis, cell growth, and differentiation [192]. PAI-1 can also affect cell migration and signaling through the interaction with vitronectin and LDL receptor related protein 1 (LRP1). Several

studies have noted additional anti-fibrinolytic independent mechanisms by which PAI-1 can induce endothelial dysfunction and atherosclerosis (Figure 2).

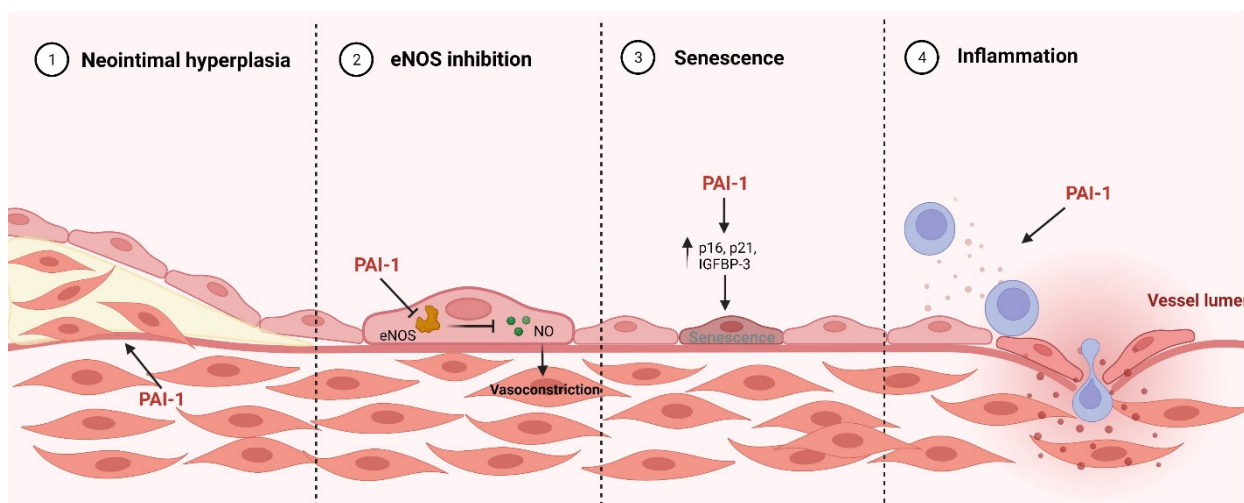


Figure 2. Pathological role of PAI-1 in the vasculature.

4.1. Pro-Inflammatory

As mentioned before, proinflammatory cytokines such as TNF- α and IL-6 can upregulate PAI-1 expression [93,100]. However, PAI-1 possesses the intrinsic ability to modulate inflammation. In alveolar epithelial cells stimulated by cigarette smoke extraction and lipopolysaccharides (LPS), expression of inflammatory factors and monocyte migration were detected. After transfection with siRNA-targeted PAI-1, these inflammatory indicators were attenuated, suggesting a proinflammatory role of PAI-1 at least in chronic obstructive pulmonary disease (COPD) [193]. Moreover, PAI-1 can modulate inflammation and induce macrophage infiltration in murine lungs after LPS-infusion through toll-like Receptor-4 (TLR4) [194]. More recently, it has been shown that PAI-1 promotes neutrophil diapedesis and tissue injury after ischemia-reperfusion (I/R). After I/R, PAI-1 accumulates on the endothelial cell surface and encounters rolling neutrophils expressing LRP1. PAI-1 then facilitates the adhesion of neutrophils through the intracellular adhesion molecule-1 (ICAM-1) triggering endothelial permeability, transmigration of neutrophils to the sub-endothelium, and ultimately inflammation and vascular injury [29] (Figure 2). Although the proinflammatory roles of PAI-1 have not been extensively studied in the setting of endothelial dysfunction, the few studies described earlier support the assumption that such effects may be involved in PAI-1-induced CVD.

4.2. eNOS Inhibition

NO is a gaseous molecule that is synthesized by nitric oxide synthases from L-arginine with a half-life of 2–30 s [195]. In the endothelium, eNOS is the major producer of NO that diffuses to the smooth muscle cells and stimulates soluble guanylate cyclase, thereby relaxing SMCs and initiating vasodilation [195]. NO has also anti-thrombotic, antiproliferative, and anti-inflammatory properties [196]. An imbalance in NO production or in its bioavailability can induce endothelial dysfunction and subsequent CVD [197,198]. Several protein–protein interactions have been shown to modulate eNOS activity such as caveolin-1, heat shock protein 90, and hemoglobin- α [199]. Very recently, it has been uncovered that PAI-1 can be endocytosed by endothelial cells and directly bind to and suppress the ability of eNOS to produce NO [30] (Figure 2). Additionally, chemical inhibition of PAI-1 was shown to impair its interaction with eNOS and to enhance endothelium-dependent vasodilation in blood vessels [30]. Another recent study showed that delivery of recombinant PAI-1 to carotid arteries resulted in reductions in NO signaling and the enhancement of

endothelial-derived hyperpolarization signaling [200]. This evidence incriminates PAI-1 as a direct mediator of endothelial dysfunction.

4.3. Senescence

Senescence is an orchestrated cellular process characterized by the permanent termination of cellular proliferation. Tissue resident cells exhibit hallmarks of the cellular senescent phenotype predominantly during the development of age-related disorders including atherosclerosis [201]. Stress-induced premature cellular senescence is the major contributor to age-dependent vascular pathologies [202]. Quintessential senescent stimuli include ROS-mediated DNA damage, telomere erosion, and the activation of certain transforming genes [203,204]. Still, senescent cells are metabolically active and capable of producing factors called the senescence messaging secretome (SMS). Extensive evidence has identified PAI-1 as a prominent member of the SMS [28,205]. PAI-1 levels increase with age in many different tissues, which are associated with the increased incidence of stress-induced thrombosis in aged mice [206]. In a murine model of thrombosis, plasma PAI-1 levels were elevated in old thrombosed mice when compared to age-matched non-thrombosed mice or younger thrombosed mice [207]. These results indicate that the elevation of PAI-1 with age could predict the onset and progression of atherothrombosis in the elderly population. In endothelial cells, the majority of high passage cells were senescent and had upregulated levels of PAI-1, p21, and monocyte adhesion molecule, while the overexpression of SIRT-1 prevented stress-induced senescence by suppressing the PAI-1 levels and enhancing eNOS expression [208]. Several other in vitro studies showed that TGF- β and p53 pathways elevated PAI-1 levels and inhibited the proliferation of fibroblasts and keratinocytes. However, with the absence of PAI-1, TGF- β and p53 were unable to inhibit proliferation in both cells [209,210]. More importantly, overexpressing PAI-1 was sufficient to promote replicative senescence in fibroblasts [209]. These data strongly indicate that PAI-1 is not only a marker, but also a *bona fide* mediator of senescence. To confirm that PAI-1 induces vascular senescence in vivo, experiments using the inhibition of PAI-1 have been shown to reduce p16 levels and telomere attrition induced by eNOS inhibition in murine aortic tissue [211]. Additionally, in a murine model of accelerated aging (klotho hypomorph), plasma levels of PAI-1 were 45-fold higher than in wild-type mice with increased renal expression of p16 that was reduced after PAI-1 pharmacological inhibition with a noticeable increase in life span [212]. The mechanisms involved in PAI-1-mediated senescence are still unclear. One suggested pathway was the inhibition of insulin-like growth factor binding protein-3 (IGFBP-3) degradation. IGFBP-3 has been shown to directly induce cellular senescence and its depletion was protective against doxorubicin-induced senescence [213]. PAI-1 inhibition also decreased IGFBP-3, p21, p16, and p53 levels in doxorubicin-treated endothelial cells, fibroblasts, and cardiomyocytes [214] (Figure 2). Overall, it is evident that the PAI-1 plays an important role in mediating and controlling cellular senescence.

4.4. Neointimal Hyperplasia

Neointimal hyperplasia is a prominent process involved in CVD such as atherosclerosis and restenosis after balloon angioplasty. Migration of SMCs from the media through the ECM into the intima is a key step in neointimal hyperplasia [215]. PAI-1 levels have been shown to increase in human vascular diseases characterized by neointima formation [216,217]. Through its interactions with vitronectin and LRP1, PAI-1 can mediate SMC adhesion and migration. PAI-1 binding to vitronectin inhibits its interactions with its receptors on SMC, thereby attenuating SMC adhesion and migration [218,219]. On the other hand, PAI-1 binding to LRP1 could promote SMC migration [220]. Thus, the concentrations of PAI-1 and vitronectin can influence neointimal formation. Pharmacological inhibition of PAI-1 in vitro and in vivo can prevent SMC migration and neointimal hyperplasia [31,221]. Indeed, targeting PAI-1 inhibited SMC migration through collagen gels including those supplemented with vitronectin, but did not inhibit the migration in endothelial cells and PAI-1 deficient SMCs [31]. Moreover, PAI-1 inhibition decreased the LRP-mediated signal

transduction in SMCs that was markedly lower in endothelial cells. Importantly, targeting PAI-1 blocked intimal hyperplasia and inflammation in murine models of pathological vascular remodeling, but did not impair reendothelialization after mechanical denudation of the vascular endothelium [31]. These findings suggest an important role of PAI-1 in neointima formation, at least in settings involving atherosclerosis and restenosis (Figure 2).

5. Is PAI-1 a Mediator of OSA-Induced CVD?

OSA is a chronic condition that is highly prevalent globally, especially among obese subjects. Extensive evidence links OSA to increased risk of CVD and overall mortality. The prevalence of OSA among stroke patients is estimated to be 50–70% [222], while up to 65% of patients who seek medical attention for a cardiovascular event are diagnosed with OSA [223]. Despite its high prevalence in patients with CVD and the susceptibility of cardiac patients to OSA-related stressors and adverse cardiovascular outcomes, OSA often remains under-recognized in the field of cardiovascular medicine. During sleep, OSA triggers IH coupled with sleep fragmentation that can induce elevations in blood pressure, OS, and inflammation [3,5,224]. Using rodent models of IH, hemodynamic changes emerge and lead to blood pressure alterations, along with impairments in vascular reactivity, ROS production, activation of proinflammatory cytokines, and altered lipid metabolism, all of which are important factors promoting endothelial dysfunction and atherosclerosis [4,5]. Unfortunately, the beneficial effects of current OSA therapies such as continuous positive airway pressure (CPAP) on CVD outcomes are inconsistent and fraught with scientific controversy. For instance, the largest randomized control study to date (SAVE) failed to demonstrate conclusive evidence of significant reductions in the primary end point (composite CVD) among patients treated with CPAP after a mean of 3.7 years follow-up [44]. A similar randomized clinical trial involving approximately 2500 subjects failed to identify OSA as an independent factor increasing the prevalence of ischemic coronary events, whereas treatment with CPAP did not significantly reduce the CAD prevalence [43]. Moreover, although incident CAD events are significantly enhanced by OSA, this risk is apparent only in those patients without a previous history of CAD [225]. This suggests that once the atherosclerotic vascular pathological processes reach more advanced stages, their reversibility with OSA treatment may not be possible, a finding that was recapitulated in mice exposed to IH for prolonged periods of time [226]. Furthermore, differential sex-specific responses to CPAP for OSA, at least for circulating inflammatory biomarkers even after adjusting for confounding factors, warrant further investigation to inform sex-specific personalized treatment approaches [227]. Ultimately, the need for additional adjuvant therapies aimed at the cardiovascular disturbances induced by OSA are needed.

Circulating PAI-1 levels are elevated in OSA patients [32–42]. Indeed, OSA has been associated with a hypercoagulable state and a decrease in fibrinolytic activity [228], putting OSA patients at high risk of developing thrombosis [229–231]. As described earlier, ROS and proinflammatory cytokines are major drivers of PAI-1 transcription. Extensive evidence from clinical and experimental studies shows that lipid, protein, and DNA oxidative stress markers are all elevated in OSA patients and in animals exposed to IH [232–239]. Additionally, neutrophils and monocytes isolated from OSA patients were shown to be activated and exhibited increased ROS production [240,241]. Evidence from animals and cells exposed to IH also shows that NADPH oxidases, xanthine oxidase, and mitochondria are all major sources of ROS [224]. NF-KB has been shown to be activated in OSA and pro-inflammatory cytokines such as TNF- α , IL-6, and CRP are also all elevated in OSA patients [5,33,242–245]. Indeed, neutrophils from OSA patients showed an 8-fold greater NF-KB binding activity [246]. A recent meta-analysis identified a significant association between OSA and elevated TNF- α levels, while TNF- α levels were consistently correlated with the severity of OSA [247]. Furthermore, the hypoxic stimulus resulting from IH can promote HIF-1 α signaling and contribute to the upregulation of PAI-1 [248]. Although clinical studies show normal or even reduced levels of plasma TGF- β levels (another major

driver of PAI-1 transcription) in OSA [249,250], it has been shown that TGF- β increased with OSA severity in exhaled breath condensate, which can be normalized by CPAP treatment [250]. Furthermore, several animal studies have reported increased TGF- β /Smad signaling in renal, lung, and cardiac tissues when exposed to IH [251–253]. The majority of OSA patients have other or more coexisting co-morbidities including obesity, hypertension, diabetes, and metabolic syndrome [254–257]. Thus, the increased RAAS activation and the enhanced levels of Ang II, along with dyslipidemia, hyperglycemia, and insulin resistance may impose a synergistic effect on PAI-1 levels in OSA patients. Collectively, OSA appears to positively affect the PAI-1 levels as the majority of the mechanisms involved in PAI-1 upregulation can be triggered by OSA (Figure 3).

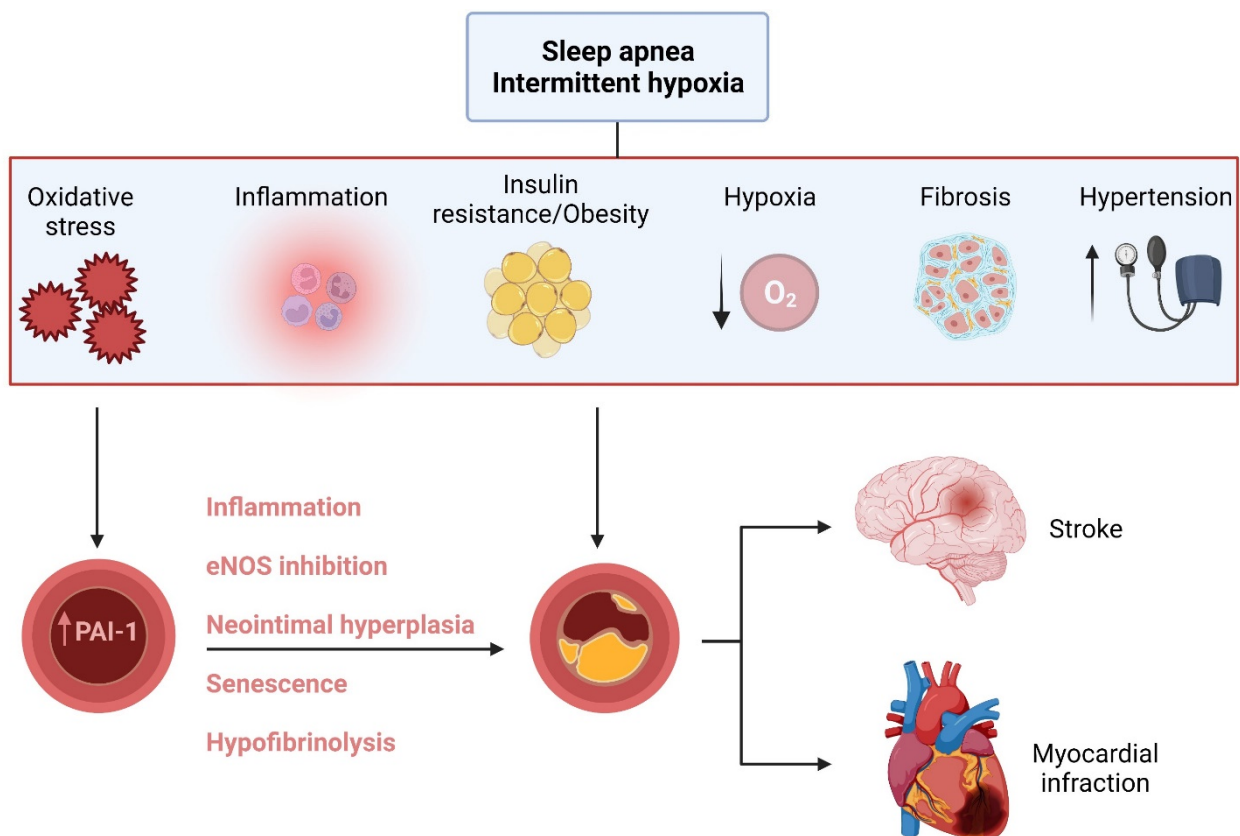


Figure 3. Putative role of PAI-1 in OSA induced CVD.

As indicated in the aforementioned paragraphs, PAI-1 contributes to endothelial dysfunction and atherosclerosis through inflammation, decreased eNOS function, neointimal formation, and vascular senescence, all of which have been reported in OSA and animals exposed to IH (Figure 3). Impaired endothelial function has been reported in both children and adult patients with OSA [258,259]. In animals, a recent meta-analysis analyzed over 125 studies evaluating the impact of IH on vascular function reported that IH altered vasodilation and induced increases in vasoconstrictive responses [260]. Several other studies have reported that IH can uncouple vascular eNOS, reduce eNOS phosphorylation, or directly reduce NO bioavailability [7,8,261–263]. However, no studies have examined the potential inhibitory effects of PAI-1 on eNOS under IH settings. A meta-analysis of 18 studies confirmed that OSA is an independent risk factor for carotid intima media thickness (cIMT), even after adjusting for confounding factors [264]. Another meta-analysis in animals showed that cIMT significantly increases upon IH exposure and that IH increased atherosclerotic plaque size in ApoE^{-/-} mice [260]. OSA is considered as an acceleration trigger of cellular senescence. Indeed, it has been suggested that OSA can cause telomere shortening through enhanced oxidative stress, hypoxia, inflammation, and circadian clock

disturbances [265]. Recently, plasma exosomes isolated from untreated OSA patients were shown to increase the senescence markers of naïve endothelial cells including p16 and x-gal, while similar cells exposed to IH recapitulated the same senescent phenotype [266]. Furthermore, accelerated epigenetic age clock was detected in patients with OSA when compared to the matched controls, and furthermore, adherent treatment with CPAP resulted in the deceleration of epigenetic aging [267]. However, the role of PAI-1 in promoting neointimal formation and mediating vascular senescence has yet to be evaluated in OSA. Thus, it is plausible that OSA-induced vascular dysfunction can be mediated, at least in part, by deregulated PAI-1-related pathways (Figure 3). Future experimental studies assessing the impact of IH in vitro and transgenic mouse lines of PAI-1 will provide valuable insights into the mechanisms by which PAI-1 induces vascular dysfunction in the context of OSA.

Given that PAI-1 is an independent risk factor for MACE, that PAI-1 shows elevated levels in OSA patients, and that there is a failure of conventional treatments to prevent adverse cardiovascular outcomes in OSA patients, it is tempting to propose that targeting PAI-1 may be advantageous in OSA patients with a risk of CVD. Many approaches have been dedicated to the development of PAI-1 inhibitors including small molecules, synthetic peptides, RNA aptamers, and monoclonal antibodies. The mechanisms of action by which these inhibitors are operationally active include: (i) blocking the initial formation of the Michalis complex between PAs and PAI-1; (ii) accelerating the transformation of active PAI-1 to its latent inactive form; or (iii) impeding the formation of the final inhibitory complex, leading to the substrate behavior of PAI-1 [54]. Several experimental studies have shown that PAI-1 inhibitors can inhibit metabolic dysregulation, improve endothelial function, and prevent atherosclerosis in the setting of diet-induced obesity [31,211,268,269]. Despite the extensive characterization of PAI-1 inhibitors and the promising results from the in vitro and in vivo studies, none of the existing PAI-1 inhibitors have yet to be approved for use in humans. This is mainly due to the affinity and specificity issues, structural plasticity of PAI-1, and the counteraction of PAI-1 binding proteins that can modulate its activity (such as vitronectin) [54]. However, evaluating the potential beneficial effects of PAI-1 inhibitors in the setting of IH is essential to assess whether PAI-1 is potentially a recommended approach as a therapeutic target in OSA-mediated CVD.

6. Conclusions

OSA is a chronic and extremely frequent condition that is associated with endothelial dysfunction, atherosclerosis, and overall cardiovascular risk and mortality. PAI-1 is a key regulator of the plasminogen system required for control fibrin stabilization to prevent bleeding. However, elevated levels of PAI-1 may increase the risk of thrombosis and promote atherosclerosis through antifibrinolytic-independent mechanisms. OSA can trigger several signaling pathways involved in enhancing PAI-1 transcription. Thus, being elevated in OSA patients, PAI-1 could play an additive role in OSA-induced CVD. However, PAI-1 influence on CVD in the setting of OSA has yet to be addressed. To this effect, experimental studies evaluating the impact of IH in PAI-1 deficient, overexpressing, and vascular-specific deletion transgenic animals are critically needed to elucidate the role of PAI-1 in OSA-induced CVD. Furthermore, the use of PAI-1 inhibitors under IH conditions may also provide insights into the effectiveness of PAI-1 antagonism in preventing or mitigating OSA-mediated CVD. Therefore, PAI-1 could spark clinical interest as a putative drug target for the treatment of PAI-related CVD in OSA.

Author Contributions: M.B. and D.G. wrote and edited the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: M.B. is the recipient of the American Thoracic Society Unrestricted Research Grant. D.G. is supported by NIH grant AG061824, and by Tier 2 and TRIUMPH grants from the University of Missouri.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: Figures were created with [BioRender.com](https://www.biorender.com).

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

AP1	Activation protein 1
CAD	Coronary artery disease
CPAP	Continuous positive airway pressure
CRP	C-reactive protein
CVD	Cardiovascular disease
ECM	Extracellular matrix
eNOS	Endothelial nitric oxide synthase
GR	Glucocorticoid receptor
HIF-1 α	Hypoxia-inducing factor-1 α
IH	Intermittent hypoxia
IL-6	Interleukin-6
LRP1	Low density lipoprotein receptor-related protein 1
MACE	Major adverse cardiovascular events
MAPK	Mitogen-activated protein kinase
MI	Myocardial infarction
MMP	Matrix metalloproteinase
NF- κ B	Nuclear factor kappa B
OSA	Obstructive sleep apnea
PAI-1	Plasminogen activator inhibitor-1
RCL	Reactive center loop
ROS	Reactive oxygen species
SMC	Smooth muscle cell
Sp1	Specificity protein 1
TGF- β	Transforming growth factor- β
TNF- α	Tumor necrosis factor
tPA	Tissue-type plasminogen activator
uPA	Urokinase-type plasminogen activator
uPAR	Urokinase-type plasminogen activator receptor

References

1. Benjafield, A.V.; Ayas, N.T.; Eastwood, P.R.; Heinzer, R.; Ip, M.S.M.; Morrell, M.J.; Nunez, C.M.; Patel, S.R.; Penzel, T.; Pépin, J.L.D.; et al. Estimation of the Global Prevalence and Burden of Obstructive Sleep Apnoea: A Literature-Based Analysis. *Lancet Respir. Med.* **2019**, *7*, 687–698. [[CrossRef](#)]
2. Kapur, V.K.; Auckley, D.H.; Chowdhuri, S.; Kuhlmann, D.C.; Mehra, R.; Ramar, K.; Harrod, C.G. Clinical Practice Guideline for Diagnostic Testing for Adult Obstructive Sleep Apnea: An American Academy of Sleep Medicine Clinical Practice Guideline. *J. Clin. Sleep Med.* **2017**, *13*, 479–504. [[CrossRef](#)] [[PubMed](#)]
3. Badran, M.; Yassin, B.A.; Fox, N.; Laher, I.; Ayas, N. Epidemiology of Sleep Disturbances and Cardiovascular Consequences. *Can. J. Cardiol.* **2015**, *31*, 873–879. [[CrossRef](#)] [[PubMed](#)]
4. Golbidi, S.; Badran, M.; Ayas, N.; Laher, I. Cardiovascular Consequences of Sleep Apnea. *Lung* **2012**, *190*, 113–132. [[CrossRef](#)] [[PubMed](#)]
5. Badran, M.; Ayas, N.; Laher, I. Insights into Obstructive Sleep Apnea Research. *Sleep Med.* **2014**, *15*, 485–495. [[CrossRef](#)]
6. Badran, M.; Yassin, B.A.; Lin, D.T.S.; Kobor, M.S.; Ayas, N.; Laher, I. Gestational Intermittent Hypoxia Induces Endothelial Dysfunction, Reduces Perivascular Adiponectin and Causes Epigenetic Changes in Adult Male Offspring. *J. Physiol.* **2019**, *597*, 5349–5364. [[CrossRef](#)]
7. Badran, M.; Abuyassin, B.; Golbidi, S.; Ayas, N.; Laher, I. Uncoupling of Vascular Nitric Oxide Synthase Caused by Intermittent Hypoxia. *Oxid. Med. Cell. Longev.* **2016**, 2354870. [[CrossRef](#)]
8. Badran, M.; Golbidi, S.; Devlin, A.; Ayas, N.; Laher, I. Chronic Intermittent Hypoxia Causes Endothelial Dysfunction in a Mouse Model of Diet-Induced Obesity. *Sleep Med.* **2014**, *15*, 596–602. [[CrossRef](#)]

9. Castro-Grattoni, A.L.; Alvarez-Buvé, R.; Torres, M.; Farré, R.; Montserrat, J.M.; Dalmases, M.; Almendros, I.; Barbé, F.; Sánchez-De-La-Torre, M. Intermittent Hypoxia-Induced Cardiovascular Remodeling Is Reversed by Normoxia in a Mouse Model of Sleep Apnea. *Chest* **2016**, *149*, 1400–1408. [[CrossRef](#)]
10. Trzepizur, W.; Cortese, R.; Gozal, D. Murine Models of Sleep Apnea: Functional Implications of Altered Macrophage Polarity and Epigenetic Modifications in Adipose and Vascular Tissues. *Metabolism* **2018**, *84*, 44–55. [[CrossRef](#)]
11. Carreras, A.; Kayali, F.; Zhang, J.; Hirotsu, C.; Wang, Y.; Gozal, D. Metabolic Effects of Intermittent Hypoxia in Mice: Steady versus High-Frequency Applied Hypoxia Daily during the Rest Period. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2012**, *303*, R700–R709. [[CrossRef](#)] [[PubMed](#)]
12. Pollicina, I.; Maniaci, A.; Lechien, J.R.; Iannella, G.; Vicini, C.; Cammaroto, G.; Cannavici, A.; Magliulo, G.; Pace, A.; Cocuzza, S.; et al. Neurocognitive Performance Improvement after Obstructive Sleep Apnea Treatment: State of the Art. *Behav. Sci.* **2021**, *11*, 180. [[CrossRef](#)] [[PubMed](#)]
13. Seda, G.; Han, T.S. Effect of Obstructive Sleep Apnea on Neurocognitive Performance. *Sleep Med. Clin.* **2020**, *15*, 77–85. [[CrossRef](#)] [[PubMed](#)]
14. Tietjens, J.R.; Claman, D.; Kezirian, E.J.; de Marco, T.; Mirzayan, A.; Sadroonri, B.; Goldberg, A.N.; Long, C.; Gerstenfeld, E.P.; Yeghiazarians, Y. Obstructive Sleep Apnea in Cardiovascular Disease: A Review of the Literature and Proposed Multidisciplinary Clinical Management Strategy. *J. Am. Heart Assoc.* **2019**, *8*, e010440. [[CrossRef](#)]
15. Boehme, A.K.; Esenwa, C.; Elkind, M.S.V. Stroke Risk Factors, Genetics, and Prevention. *Circ. Res.* **2017**, *120*, 472. [[CrossRef](#)]
16. Juhan-Vague, I.; Pyke, S.D.M.; Alessi, M.C.; Jespersen, J.; Haverkate, F.; Thompson, S.G. Fibrinolytic Factors and the Risk of Myocardial Infarction or Sudden Death in Patients with Angina Pectoris. *Circulation* **1996**, *94*, 2057–2063. [[CrossRef](#)]
17. Rijken, D.C.; Lijnen, H.R. New Insights into the Molecular Mechanisms of the Fibrinolytic System. *J. Thromb. Haemost.* **2009**, *7*, 4–13. [[CrossRef](#)]
18. Chapin, J.C.; Hajjar, K.A. Fibrinolysis and the Control of Blood Coagulation. *Blood Rev.* **2015**, *29*, 17. [[CrossRef](#)]
19. Binder, B.R.; Christ, G.; Gruber, F.; Grubic, N.; Hufnagl, P.; Krebs, M.; Mihaly, J.; Prager, G.W. Plasminogen Activator Inhibitor 1: Physiological and Pathophysiological Roles. *News Physiol. Sci.* **2002**, *17*, 56–61. [[CrossRef](#)]
20. Eun, A.L.; Ji, Y.S.; Jiang, Z.; Mi, R.Y.; Min, K.K.; Ha, H.; Hi, B.L. Reactive Oxygen Species Mediate High Glucose-Induced Plasminogen Activator Inhibitor-1 up-Regulation in Mesangial Cells and in Diabetic Kidney. *Kidney Int.* **2005**, *67*, 1762–1771. [[CrossRef](#)]
21. Swiatkowska, M.; Szemraj, J.; Cierniewski, C.S. Induction of PAI-1 Expression by Tumor Necrosis Factor Alpha in Endothelial Cells Is Mediated by Its Responsive Element Located in the 4G/5G Site. *FEBS J.* **2005**, *272*, 5821–5831. [[CrossRef](#)] [[PubMed](#)]
22. Guo, B.; Inoki, K.; Isono, M.; Mori, H.; Kanasaki, K.; Sugimoto, T.; Akiba, S.; Sato, T.; Yang, B.; Kikkawa, R.; et al. MAPK/AP-1-Dependent Regulation of PAI-1 Gene Expression by TGF-Beta in Rat Mesangial Cells. *Kidney Int.* **2005**, *68*, 972–984. [[CrossRef](#)] [[PubMed](#)]
23. Song, C.; Burgess, S.; Eicher, J.D.; O'Donnell, C.J.; Johnson, A.D.; Huang, J.; Sabater-Lleal, M.; Asselbergs, F.W.; Tregouet, D.; Shin, S.Y.; et al. Causal Effect of Plasminogen Activator Inhibitor Type 1 on Coronary Heart Disease. *J. Am. Heart Assoc.* **2017**, *6*, e004918. [[CrossRef](#)]
24. Placencio, V.R.; DeClerck, Y.A. Plasminogen Activator Inhibitor-1 in Cancer: Rationale and Insight for Future Therapeutic Testing. *Cancer Res.* **2015**, *75*, 2969–2974. [[CrossRef](#)] [[PubMed](#)]
25. Altalhi, R.; Pechlivani, N.; Ajjan, R.A. PAI-1 in Diabetes: Pathophysiology and Role as a Therapeutic Target. *Int. J. Mol. Sci.* **2021**, *22*, 3170. [[CrossRef](#)] [[PubMed](#)]
26. Małgorzewicz, S.; Skrzypczak-Jankun, E.; Jankun, J. Plasminogen Activator Inhibitor-1 in Kidney Pathology (Review). *Int. J. Mol. Med.* **2013**, *31*, 503–510. [[CrossRef](#)]
27. Jiang, H.; Li, X.; Chen, S.; Lu, N.; Yue, Y.; Liang, J.; Zhang, Z.; Yuan, Y. Plasminogen Activator Inhibitor-1 in Depression: Results from Animal and Clinical Studies. *Sci. Rep.* **2016**, *6*, 30464. [[CrossRef](#)]
28. Vaughan, D.E.; Rai, R.; Khan, S.S.; Eren, M.; Ghosh, A.K. PAI-1 Is a Marker and a Mediator of Senescence. *Arterioscler. Thromb. Vasc. Biol.* **2017**, *37*, 1446. [[CrossRef](#)]
29. Praetner, M.; Zuchtriegel, G.; Holzer, M.; Uhl, B.; Schaubächer, J.; Mittmann, L.; Fabritius, M.; Fürst, R.; Zahler, S.; Funken, D.; et al. Plasminogen Activator Inhibitor-1 Promotes Neutrophil Infiltration and Tissue Injury on Ischemia-Reperfusion. *Arterioscler. Thromb. Vasc. Biol.* **2018**, *38*, 829–842. [[CrossRef](#)]
30. Garcia, V.; Park, E.J.; Siragusa, M.; Frohlich, F.; Haque, M.M.; Pascale, J.V.; Heberlein, K.R.; Isakson, B.E.; Stuehr, D.J.; Sessa, W.C. Unbiased Proteomics Identifies Plasminogen Activator Inhibitor-1 as a Negative Regulator of Endothelial Nitric Oxide Synthase. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 9497–9507. [[CrossRef](#)]
31. Ji, Y.; Weng, Z.; Fish, P.; Goyal, N.; Luo, M.; Myears, S.P.; Strawn, T.L.; Chandrasekar, B.; Wu, J.; Fay, W.P. Pharmacological Targeting of Plasminogen Activator Inhibitor-1 Decreases Vascular Smooth Muscle Cell Migration and Neointima Formation. *Arterioscler. Thromb. Vasc. Biol.* **2016**, *36*, 2167–2175. [[CrossRef](#)] [[PubMed](#)]
32. Von Känel, R.; Natarajan, L.; Ancoli-Israel, S.; Mills, P.J.; Lored, J.S.; Dimsdale, J.E. Day/Night Rhythm of Hemostatic Factors in Obstructive Sleep Apnea. *Sleep* **2010**, *33*, 371–377. [[CrossRef](#)] [[PubMed](#)]
33. Gileles-Hillel, A.; Alonso-Álvarez, M.L.; Kheirandish-Gozal, L.; Peris, E.; Cordero-Guevara, J.A.; Terán-Santos, J.; Martínez, M.G.; Jurado-Luque, M.J.; Corral-Peñañiel, J.; Duran-Cantolla, J.; et al. Inflammatory Markers and Obstructive Sleep Apnea in Obese Children: The NANOS Study. *Mediators Inflamm.* **2014**, e605280. [[CrossRef](#)] [[PubMed](#)]

34. Zakrzewski, M.; Zakrzewska, E.; Kiciński, P.; Przybylska-Kuć, S.; Dybała, A.; Myśliński, W.; Pastryk, J.; Tomaszewski, T.; Mosiewicz, J. Evaluation of Fibrinolytic Inhibitors: Alpha-2-Antiplasmin and Plasminogen Activator Inhibitor 1 in Patients with Obstructive Sleep Apnoea. *PLoS ONE* **2016**, *11*, 166725. [[CrossRef](#)]
35. Nizankowska-Jędrzejczyk, A.; Almeida, F.R.; Lowe, A.A.; Kania, A.; Nastalek, P.; Mejza, F.; Foley, J.H.; Nizankowska-Mogilnicka, E.; Undas, A. Modulation of Inflammatory and Hemostatic Markers in Obstructive Sleep Apnea Patients Treated with Mandibular Advancement Splints: A Parallel, Controlled Trial. *J. Clin. Sleep Med.* **2014**, *10*, 255–262. [[CrossRef](#)]
36. Bagai, K.; Muldowney, J.A.S.; Song, Y.; Wang, L.; Bagai, J.; Artibee, K.J.; Vaughan, D.E.; Malow, B.A. Circadian Variability of Fibrinolytic Markers and Endothelial Function in Patients with Obstructive Sleep Apnea. *Sleep* **2014**, *37*, 359–367. [[CrossRef](#)]
37. von Känel, R.; Loreda, J.S.; Ancoli-Israel, S.; Dimsdale, J.E. Association between Sleep Apnea Severity and Blood Coagulability: Treatment Effects of Nasal Continuous Positive Airway Pressure. *Sleep Breath.* **2006**, *10*, 139–146. [[CrossRef](#)]
38. Phillips, C.L.; McEwen, B.J.; Morel-Kopp, M.C.; Yee, B.J.; Sullivan, D.R.; Ward, C.M.; Tofler, G.H.; Grunstein, R.R. Effects of Continuous Positive Airway Pressure on Coagulability in Obstructive Sleep Apnoea: A Randomised, Placebo-Controlled Crossover Study. *Thorax* **2012**, *67*, 639–644. [[CrossRef](#)]
39. Rangemark, C.; Hedner, J.A.; Carlson, J.T.; Glerup, G.; Winther, K. Platelet Function and Fibrinolytic Activity in Hypertensive and Normotensive Sleep Apnea Patients. *Sleep* **1995**, *18*, 188–194. [[CrossRef](#)]
40. Kheirandish-Gozal, L.; Gileles-Hillel, A.; Alonso-Álvarez, M.L.; Peris, E.; Bhattacharjee, R.; Terán-Santos, J.; Duran-Cantolla, J.; Gozal, D. Effects of Adenotonsillectomy on Plasma Inflammatory Biomarkers in Obese Children with Obstructive Sleep Apnea: A Community-Based Study. *Int. J. Obes.* **2015**, *39*, 1094–1100. [[CrossRef](#)]
41. Martin, R.A.; Strosnider, C.; Giersch, G.; Womack, C.J.; Hargens, T.A. The Effect of Acute Aerobic Exercise on Hemostasis in Obstructive Sleep Apnea. *Sleep Breath.* **2017**, *21*, 623–629. [[CrossRef](#)] [[PubMed](#)]
42. von Känel, R.; Loreda, J.S.; Ancoli-Israel, S.; Mills, P.J.; Dimsdale, J.E. Elevated Plasminogen Activator Inhibitor 1 in Sleep Apnea and Its Relation to the Metabolic Syndrome: An Investigation in 2 Different Study Samples. *Metabolism* **2007**, *56*, 969–976. [[CrossRef](#)] [[PubMed](#)]
43. Sánchez-de-la-Torre, M.; Sánchez-de-la-Torre, A.; Bertran, S.; Abad, J.; Duran-Cantolla, J.; Cabriada, V.; Mediano, O.; Masdeu, M.J.; Alonso, M.L.; Masa, J.F.; et al. Effect of Obstructive Sleep Apnoea and Its Treatment with Continuous Positive Airway Pressure on the Prevalence of Cardiovascular Events in Patients with Acute Coronary Syndrome (ISAACC Study): A Randomised Controlled Trial. *Lancet Respir. Med.* **2020**, *8*, 359–367. [[CrossRef](#)]
44. McEvoy, R.D.; Antic, N.A.; Heeley, E.; Luo, Y.; Ou, Q.; Zhang, X.; Mediano, O.; Chen, R.; Drager, L.F.; Liu, Z.; et al. CPAP for Prevention of Cardiovascular Events in Obstructive Sleep Apnea. *N. Engl. J. Med.* **2016**, *375*, 919–931. [[CrossRef](#)] [[PubMed](#)]
45. Simpson, A.J.; Booth, N.A.; Moore, N.R.; Bennett, B. Distribution of Plasminogen Activator Inhibitor (PAI-1) in Tissues. *J. Clin. Pathol.* **1991**, *44*, 139–143. [[CrossRef](#)] [[PubMed](#)]
46. Crandall, D.L.; Quinet, E.M.; Morgan, G.A.; Busler, D.E.; Mchendry-Rinde, B.; Kral, J.G. Synthesis and Secretion of Plasminogen Activator Inhibitor-1 by Human Preadipocytes. *J. Clin. Endocrinol. Metab.* **1999**, *84*, 3222–3227. [[CrossRef](#)] [[PubMed](#)]
47. Zhang, L.; Seiffert, D.; Fowler, B.J.; Jenkins, G.R.; Thinnes, T.C.; Loskutoff, D.J.; Parmer, R.J.; Miles, L.A. Plasminogen Has a Broad Extrahepatic Distribution. *Thromb. Haemost.* **2002**, *87*, 493–501. [[CrossRef](#)]
48. Booth, N.A.; Simpson, A.J.; Croll, A.; Bennett, B.; MacGregor, I.R. Plasminogen Activator Inhibitor (PAI-1) in Plasma and Platelets. *Br. J. Haematol.* **1988**, *70*, 327–333. [[CrossRef](#)]
49. Charlton, P. The Status of Plasminogen Activator Inhibitor-1 as a Therapeutic Target. *Expert Opin. Investig. Drugs* **1997**, *6*, 539–554. [[CrossRef](#)]
50. Brogren, H.; Wallmark, K.; Deinum, J.; Karlsson, L.; Jern, S. Platelets Retain High Levels of Active Plasminogen Activator Inhibitor 1. *PLoS ONE* **2011**, *6*, e26762. [[CrossRef](#)]
51. Torr-Brown, S.R.; Sobel, B.E. Attenuation of Thrombolysis by Release of Plasminogen Activator Inhibitor Type-1 from Platelets. *Thromb. Res.* **1993**, *72*, 413–421. [[CrossRef](#)]
52. Morrow, G.B.; Whyte, C.S.; Mutch, N.J. Functional Plasminogen Activator Inhibitor 1 Is Retained on the Activated Platelet Membrane Following Platelet Activation. *Haematologica* **2020**, *105*, 2824–2833. [[CrossRef](#)] [[PubMed](#)]
53. Sillen, M.; Declerck, P.J. A Narrative Review on Plasminogen Activator Inhibitor-1 and Its (Patho)Physiological Role: To Target or Not to Target? *Int. J. Mol. Sci.* **2021**, *22*, 2721. [[CrossRef](#)] [[PubMed](#)]
54. Sillen, M.; Declerck, P.J. Targeting PAI-1 in Cardiovascular Disease: Structural Insights Into PAI-1 Functionality and Inhibition. *Front. Cardiovasc. Med.* **2020**, *7*, 364. [[CrossRef](#)] [[PubMed](#)]
55. Rahman, F.A.; Krause, M.P. PAI-1, the Plasminogen System, and Skeletal Muscle. *Int. J. Mol. Sci.* **2020**, *21*, 7066. [[CrossRef](#)]
56. Wind, T.; Hansen, M.; Jensen, J.K.; Andreasen, P.A. The Molecular Basis for Anti-Proteolytic and Non-Proteolytic Functions of Plasminogen Activator Inhibitor Type-1: Roles of the Reactive Centre Loop, the Shutter Region, the Flexible Joint Region and the Small Serpin Fragment. *Biol. Chem.* **2002**, *383*, 21–36. [[CrossRef](#)]
57. Schroeck, F.; Arroyo de Prada, N.; Sperl, S.; Schmitt, M.; Magdolen, V. Interaction of Plasminogen Activator Inhibitor Type-1 (PAI-1) with Vitronectin (Vn): Mapping the Binding Sites on PAI-1 and Vn. *Biol. Chem.* **2002**, *383*, 1143–1149. [[CrossRef](#)]
58. Wilczynska, M.; Fa, M.; Ohlsson, P.I.; Ny, T. The Inhibition Mechanism of Serpins. Evidence That the Mobile Reactive Center Loop Is Cleaved in the Native Protease-Inhibitor Complex. *J. Biol. Chem.* **1995**, *270*, 29652–29655. [[CrossRef](#)]
59. Lawrence, D.A.; Ginsburg, D.; Day, D.E.; Berkenpas, M.B.; Verhamme, I.M.; Kvassman, J.O.; Shore, J.D. Serpin-Protease Complexes Are Trapped as Stable Acyl-Enzyme Intermediates. *J. Biol. Chem.* **1995**, *270*, 25309–25312. [[CrossRef](#)]

60. Boudier, C.; Gils, A.; Declerck, P.J.; Bieth, J.G. The Conversion of Active to Latent Plasminogen Activator Inhibitor-1 Is an Energetically Silent Event. *Biophys. J.* **2005**, *88*, 2848–2854. [[CrossRef](#)]
61. Gettins, P.G.W.; Olson, S.T. Inhibitory Serpins. New Insights into Their Folding, Polymerization, Regulation and Clearance. *Biochem. J.* **2016**, *473*, 2273–2293. [[CrossRef](#)] [[PubMed](#)]
62. Simone, T.M.; Higgins, P.J. Low Molecular Weight Antagonists of Plasminogen Activator Inhibitor-1: Therapeutic Potential in Cardiovascular Disease. *Mol. Med. Ther.* **2012**, *1*, 101. [[CrossRef](#)] [[PubMed](#)]
63. Dupont, D.M.; Madsen, J.B.; Kristensen, T.; Bodker, J.S.; Blouse, G.E.; Wind, T.; Andreassen, P.A. Biochemical Properties of Plasminogen Activator Inhibitor-1. *Front. Biosci.* **2009**, *14*, 1337–1361. [[CrossRef](#)] [[PubMed](#)]
64. Fjellström, O.; Deinum, J.; Sjögren, T.; Johansson, C.; Geschwindner, S.; Nerme, V.; Legnehed, A.; McPheat, J.; Olsson, K.; Bodin, C.; et al. Characterization of a Small Molecule Inhibitor of Plasminogen Activator Inhibitor Type 1 That Accelerates the Transition into the Latent Conformation. *J. Biol. Chem.* **2013**, *288*, 873–885. [[CrossRef](#)]
65. Lawrence, D.A.; Ginsburg, D.; Olson, S.T.; Palaniappan, S. Engineering Plasminogen Activator Inhibitor 1 Mutants with Increased Functional Stability. *Biochemistry* **1994**, *33*, 3643–3648. [[CrossRef](#)]
66. Van De Craen, B.; Declerck, P.J.; Gils, A. The Biochemistry, Physiology and Pathological Roles of PAI-1 and the Requirements for PAI-1 Inhibition in Vivo. *Thromb. Res.* **2012**, *130*, 576–585. [[CrossRef](#)]
67. Thorsen, S.; Philips, M.; Selmer, J.; Lecander, I.; Åstedt, B. Kinetics of Inhibition of Tissue-Type and Urokinase-Type Plasminogen Activator by Plasminogen-Activator Inhibitor Type 1 and Type 2. *Eur. J. Biochem.* **1988**, *175*, 33–39. [[CrossRef](#)]
68. Lee, E.; Vaughan, D.E.; Parikh, S.H.; Grodzinsky, A.J.; Libby, P.; Lark, M.W.; Lee, R.T. Regulation of Matrix Metalloproteinases and Plasminogen Activator Inhibitor-1 Synthesis by Plasminogen in Cultured Human Vascular Smooth Muscle Cells. *Circ. Res.* **1996**, *78*, 44–49. [[CrossRef](#)]
69. Pizzino, G.; Irrera, N.; Cucinotta, M.; Pallio, G.; Mannino, F.; Arcoraci, V.; Squadrito, F.; Altavilla, D.; Bitto, A. Oxidative Stress: Harms and Benefits for Human Health. *Oxid. Med. Cell. Longev.* **2017**, *2017*, 8416763. [[CrossRef](#)]
70. Senoner, T.; Dichtl, W. Oxidative Stress in Cardiovascular Diseases: Still a Therapeutic Target? *Nutrients* **2019**, *11*, 2090. [[CrossRef](#)]
71. D’Oria, R.; Schipani, R.; Leonardini, A.; Natalicchio, A.; Perrini, S.; Cignarelli, A.; Laviola, L.; Giorgino, F. The Role of Oxidative Stress in Cardiac Disease: From Physiological Response to Injury Factor. *Oxid. Med. Cell. Longev.* **2020**, *2020*, 5732956. [[CrossRef](#)] [[PubMed](#)]
72. Oszajca, K.; Bieniasz, M.; Brown, G.; Swiatkowska, M.; Bartkowiak, J.; Szemraj, J. Effect of Oxidative Stress on the Expression of T-PA, u-PA, u-PAR, and PAI-1 in Endothelial Cells. *Biochem. Cell Biol.* **2008**, *86*, 477–486. [[CrossRef](#)] [[PubMed](#)]
73. Swiatkowska, M.; Szemraj, J.; Al-Nedawi, K.; Pawlowska, Z. Reactive Oxygen Species Upregulate Expression of PAI-1 in Endothelial Cells. *Cell. Mol. Biol. Lett.* **2002**, *7*, 1065–1071. [[PubMed](#)]
74. Jaulmes, A.; Sansilvestri-Morel, P.; Rolland-Valognes, G.; Bernhardt, F.; Gaertner, R.; Lockhart, B.P.; Cordi, A.; Wierzbicki, M.; Rupin, A.; Verbeuren, T.J. Nox4 Mediates the Expression of Plasminogen Activator Inhibitor-1 via P38 MAPK Pathway in Cultured Human Endothelial Cells. *Thromb. Res.* **2009**, *124*, 439–446. [[CrossRef](#)]
75. Orbe, J.; Rodriguez, J.A.; Calvo, A.; Grau, A.; Belzunce, M.S.; Martinez-Caro, D.; Páramo, J.A. Vitamins C and E Attenuate Plasminogen Activator Inhibitor-1 (PAI-1) Expression in a Hypercholesterolemic Porcine Model of Angioplasty. *Cardiovasc. Res.* **2001**, *49*, 484–492. [[CrossRef](#)]
76. Gomaa, A.M.S.; Abd El-Mottaleb, N.A.; Amer, H.A. Antioxidant and Anti-Inflammatory Activities of Alpha Lipoic Acid Protect against Indomethacin-Induced Gastric Ulcer in Rats. *Biomed. Pharmacother.* **2018**, *101*, 188–194. [[CrossRef](#)]
77. Martina, V.; Bruno, G.A.; Pannocchia, A.; Zumpano, E.; Tagliabue, M.; Trucco, F.; Giorgianni, A.; Stella, S.; Pescarmona, G.P. PAI-1 Reduction after Treatment with Glutathione in NIDDM. *Fibrinolysis* **1996**, *10*, 63–65. [[CrossRef](#)]
78. Bonfigli, A.R.; Pieri, C.; Manfrini, S.; Testa, I.; Sirolla, C.; Ricciotti, R.; Marra, M.; Compagnucci, P.; Testa, R. Vitamin E Intake Reduces Plasminogen Activator Inhibitor Type 1 in T2DM Patients. *Diabetes Nutr. Metab.* **2001**, *14*, 71–77.
79. Antoniadis, C.; Tousoulis, D.; Tentolouris, C.; Toutouza, M.; Marinou, K.; Goumas, G.; Tsioufis, C.; Toutouzas, P.; Stefanadis, C. Effects of Antioxidant Vitamins C and E on Endothelial Function and Thrombosis/Fibrinolysis System in Smokers. *Thromb. Haemost.* **2003**, *89*, 990–995. [[CrossRef](#)]
80. Zhao, R.; Ma, X.; Xie, X.; Shen, G.X. Involvement of NADPH Oxidase in Oxidized LDL-Induced Upregulation of Heat Shock Factor-1 and Plasminogen Activator Inhibitor-1 in Vascular Endothelial Cells. *Am. J. Physiol. Endocrinol. Metab.* **2009**, *297*, 104–111. [[CrossRef](#)]
81. Hagiwara, H.; Seki, T.; Ariga, T. The Effect of Pre-Germinated Brown Rice Intake on Blood Glucose and PAI-1 Levels in Streptozotocin-Induced Diabetic Rats. *Biosci. Biotechnol. Biochem.* **2004**, *68*, 444–447. [[CrossRef](#)] [[PubMed](#)]
82. Görlach, A.; Diebold, I.; Schini-Kerth, V.B.; Berchner-Pfannschmidt, U.; Roth, U.; Brandes, R.P.; Kietzmann, T.; Busse, R. Thrombin Activates the Hypoxia-Inducible Factor-1 Signaling Pathway in Vascular Smooth Muscle Cells. *Circ. Res.* **2001**, *89*, 47–54. [[CrossRef](#)] [[PubMed](#)]
83. Ren, S.; Shen, G.X. Impact of Antioxidants and HDL on Glycated LDL-Induced Generation of Fibrinolytic Regulators from Vascular Endothelial Cells. *Arterioscler. Thromb. Vasc. Biol.* **2000**, *20*, 1688–1693. [[CrossRef](#)] [[PubMed](#)]
84. Kwon, I.S.; Kim, J.; Rhee, D.K.; Kim, B.O.; Pyo, S. Pneumolysin Induces Cellular Senescence by Increasing ROS Production and Activation of MAPK/NF-KB Signal Pathway in Glial Cells. *Toxicol.* **2017**, *129*, 100–112. [[CrossRef](#)] [[PubMed](#)]

85. Vayalil, P.K.; Iles, K.E.; Choi, J.; Yi, A.K.; Postlethwait, E.M.; Liu, R.M. Glutathione Suppresses TGF-Beta-Induced PAI-1 Expression by Inhibiting P38 and JNK MAPK and the Binding of AP-1, SP-1, and Smad to the PAI-1 Promoter. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2007**, *293*, L1281–L1292. [[CrossRef](#)]
86. Dimova, E.Y.; Kietzmann, T. Metabolic, Hormonal and Environmental Regulation of Plasminogen Activator Inhibitor-1 (PAI-1) Expression: Lessons from the Liver. *Thromb. Haemost.* **2008**, *100*, 992–1006. [[CrossRef](#)]
87. Libby, P. Inflammation and Cardiovascular Disease Mechanisms. *Am. J. Clin. Nutr.* **2006**, *83*, 456S–460S. [[CrossRef](#)]
88. Ruparelina, N.; Chai, J.T.; Fisher, E.A.; Choudhury, R.P. Inflammatory Processes in Cardiovascular Disease: A Route to Targeted Therapies. *Nat. Rev. Cardiol.* **2016**, *14*, 133–144. [[CrossRef](#)]
89. Alfaddagh, A.; Martin, S.S.; Leucker, T.M.; Michos, E.D.; Blaha, M.J.; Lowenstein, C.J.; Jones, S.R.; Toth, P.P. Inflammation and Cardiovascular Disease: From Mechanisms to Therapeutics. *Am. J. Prev. Cardiol.* **2020**, *4*, 100130. [[CrossRef](#)]
90. Steven, S.; Frenis, K.; Oelze, M.; Kalinovic, S.; Kuntic, M.; Jimenez, M.T.B.; Vujacic-Mirski, K.; Helmstädter, J.; Kröller-Schön, S.; Münzel, T.; et al. Vascular Inflammation and Oxidative Stress: Major Triggers for Cardiovascular Disease. *Oxid. Med. Cell. Longev.* **2019**, *2019*, 7092151. [[CrossRef](#)]
91. Cesari, M.; Pahor, M.; Incalzi, R.A. REVIEW: Plasminogen Activator Inhibitor-1 (PAI-1): A Key Factor Linking Fibrinolysis and Age-Related Subclinical and Clinical Conditions. *Cardiovasc. Ther.* **2010**, *28*, e72–e91. [[CrossRef](#)] [[PubMed](#)]
92. Hube, F.; Hauner, H. The Role of TNF-Alpha in Human Adipose Tissue: Prevention of Weight Gain at the Expense of Insulin Resistance? *Horm. Metab. Res.* **1999**, *31*, 626–631. [[CrossRef](#)] [[PubMed](#)]
93. Takeshita, Y.; Takamura, T.; Hamaguchi, E.; Shimizu, A.; Ota, T.; Sakurai, M.; Kaneko, S. Tumor Necrosis Factor-Alpha-Induced Production of Plasminogen Activator Inhibitor 1 and Its Regulation by Pioglitazone and Cerivastatin in a Nonmalignant Human Hepatocyte Cell Line. *Metabolism* **2006**, *55*, 1464–1472. [[CrossRef](#)] [[PubMed](#)]
94. Pandey, M.; Loskutoff, D.J.; Samad, F. Molecular Mechanisms of Tumor Necrosis Factor-Alpha-Mediated Plasminogen Activator Inhibitor-1 Expression in Adipocytes. *FASEB J.* **2005**, *19*, 1317–1319. [[CrossRef](#)]
95. Macfelda, K.; Weiss, T.W.; Kaun, C.; Breuss, J.M.; Kapeller, B.; Zorn, G.; Oberndorfer, U.; Voegelé-Kadletz, M.; Huber-Beckmann, R.; Ullrich, R.; et al. Plasminogen Activator Inhibitor 1 Expression Is Regulated by the Inflammatory Mediators Interleukin-1alpha, Tumor Necrosis Factor-Alpha, Transforming Growth Factor-Beta and Oncostatin M in Human Cardiac Myocytes. *J. Mol. Cell. Cardiol.* **2002**, *34*, 1681–1691. [[CrossRef](#)]
96. Hou, B.; Eren, M.; Painter, C.A.; Covington, J.W.; Dixon, J.D.; Schoenhard, J.A.; Vaughan, D.E. Tumor Necrosis Factor Alpha Activates the Human Plasminogen Activator Inhibitor-1 Gene through a Distal Nuclear Factor KappaB Site. *J. Biol. Chem.* **2004**, *279*, 18127–18136. [[CrossRef](#)]
97. Samad, F.; Yamamoto, K.; Loskutoff, D.J. Distribution and Regulation of Plasminogen Activator Inhibitor-1 in Murine Adipose Tissue In Vivo. Induction by Tumor Necrosis Factor-Alpha and Lipopolysaccharide. *J. Clin. Investig.* **1996**, *97*, 37–46. [[CrossRef](#)]
98. Cigolini, M.; Tonoli, M.; Borgato, L.; Frigotto, L.; Manzato, F.; Zeminian, S.; Cardinale, C.; Camin, M.; Chiaramonte, E.; De Sandre, G.; et al. Expression of Plasminogen Activator Inhibitor-1 in Human Adipose Tissue: A Role for TNF-Alpha? *Atherosclerosis* **1999**, *143*, 81–90. [[CrossRef](#)]
99. Papanicolaou, D.A.; Wilder, R.L.; Manolagas, S.C.; Chrousos, G.P. The Pathophysiologic Roles of Interleukin-6 in Human Disease. *Ann. Intern. Med.* **1998**, *128*, 127–137. [[CrossRef](#)]
100. Kang, S.; Tanaka, T.; Inoue, H.; Ono, C.; Hashimoto, S.; Kioi, Y.; Matsumoto, H.; Matsuura, H.; Matsubara, T.; Shimizu, K.; et al. IL-6 Trans-Signaling Induces Plasminogen Activator Inhibitor-1 from Vascular Endothelial Cells in Cytokine Release Syndrome. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 22351–22356. [[CrossRef](#)]
101. Mestries, J.C.; Kruithof, E.K.O.; Gascon, M.P.; Herodin, F.; Agay, D.; Ythier, A. In Vivo Modulation of Coagulation and Fibrinolysis by Recombinant Glycosylated Human Interleukin-6 in Baboons. *Eur. Cytokine Netw.* **1994**, *5*, 275–281. [[PubMed](#)]
102. Kruithof, E.K.O. Regulation of Plasminogen Activator Inhibitor Type 1 Gene Expression by Inflammatory Mediators and Statins. *Thromb. Haemost.* **2008**, *100*, 969–975. [[CrossRef](#)] [[PubMed](#)]
103. Toma, I.; McCaffrey, T.A. Transforming Growth Factor- β and Atherosclerosis: Interwoven Atherogenic and Atheroprotective Aspects. *Cell Tissue Res.* **2012**, *347*, 155. [[CrossRef](#)]
104. Verrecchia, F.; Mauviel, A. Transforming Growth Factor- β and Fibrosis. *World J. Gastroenterol.* **2007**, *13*, 3056. [[CrossRef](#)] [[PubMed](#)]
105. Chen, P.Y.; Qin, L.; Li, G.; Wang, Z.; Dahlman, J.E.; Malagon-Lopez, J.; Gujja, S.; Cilfone, N.A.; Kauffman, K.J.; Sun, L.; et al. Endothelial TGF- β Signalling Drives Vascular Inflammation and Atherosclerosis. *Nat. Metab.* **2019**, *1*, 912–926. [[CrossRef](#)] [[PubMed](#)]
106. Seeland, U.; Haeuseler, C.; Hinrichs, R.; Rosenkranz, S.; Pfitzner, T.; Scharffetter-Kochanek, K.; Böhm, M. Myocardial Fibrosis in Transforming Growth Factor-Beta(1) (TGF-Beta(1)) Transgenic Mice Is Associated with Inhibition of Interstitial Collagenase. *Eur. J. Clin. Investig.* **2002**, *32*, 295–303. [[CrossRef](#)]
107. Grandaliano, G.; Di Paolo, S.; Monno, R.; Stallone, G.; Ranieri, E.; Pontrelli, P.; Gesualdo, L.; Schena, F.P. Protease-Activated Receptor 1 and Plasminogen Activator Inhibitor 1 Expression in Chronic Allograft Nephropathy: The Role of Coagulation and Fibrinolysis in Renal Graft Fibrosis. *Transplantation* **2001**, *72*, 1437–1443. [[CrossRef](#)]
108. De Gouville, A.C.; Boullay, V.; Krysa, G.; Pilot, J.; Brusq, J.M.; Loriolle, F.; Gauthier, J.M.; Papworth, S.A.; Laroze, A.; Gellibert, F.; et al. Inhibition of TGF-Beta Signaling by an ALK5 Inhibitor Protects Rats from Dimethylnitrosamine-Induced Liver Fibrosis. *Br. J. Pharmacol.* **2005**, *145*, 166–177. [[CrossRef](#)]

109. Kutz, S.M.; Hordines, J.; McKeown-Longo, P.J.; Higgins, P.J. TGF-Beta1-Induced PAI-1 Gene Expression Requires MEK Activity and Cell-to-Substrate Adhesion. *J. Cell Sci.* **2001**, *114*, 3905–3914. [[CrossRef](#)]
110. Hirashima, Y.; Kobayashi, H.; Suzuki, M.; Tanaka, Y.; Kanayama, N.; Terao, T. Transforming Growth Factor-Beta1 Produced by Ovarian Cancer Cell Line HRA Stimulates Attachment and Invasion through an up-Regulation of Plasminogen Activator Inhibitor Type-1 in Human Peritoneal Mesothelial Cells. *J. Biol. Chem.* **2003**, *278*, 26793–26802. [[CrossRef](#)]
111. Datta, P.K.; Blake, M.C.; Moses, H.L. Regulation of Plasminogen Activator Inhibitor-1 Expression by Transforming Growth Factor-Beta -Induced Physical and Functional Interactions between Smads and Sp1. *J. Biol. Chem.* **2000**, *275*, 40014–40019. [[CrossRef](#)] [[PubMed](#)]
112. Lund, L.R.; Riccio, A.; Andreasen, P.A.; Nielsen, L.S.; Kristensen, P.; Laiho, M.; Saksela, O.; Blasi, F.; Danø, K. Transforming Growth Factor-Beta Is a Strong and Fast Acting Positive Regulator of the Level of Type-1 Plasminogen Activator Inhibitor MRNA in WI-38 Human Lung Fibroblasts. *EMBO J.* **1987**, *6*, 1281–1286. [[CrossRef](#)] [[PubMed](#)]
113. Jaffer, O.A.; Carter, A.B.; Sanders, P.N.; Dibbern, M.E.; Winters, C.J.; Murthy, S.; Ryan, A.J.; Rokita, A.G.; Prasad, A.M.; Zabner, J.; et al. Mitochondrial-Targeted Antioxidant Therapy Decreases Transforming Growth Factor- β -Mediated Collagen Production in a Murine Asthma Model. *Am. J. Respir. Cell Mol. Biol.* **2015**, *52*, 106–115. [[CrossRef](#)] [[PubMed](#)]
114. You, W.; Hong, Y.; He, H.; Huang, X.; Tao, W.; Liang, X.; Zhang, Y.; Li, X. TGF- β Mediates Aortic Smooth Muscle Cell Senescence in Marfan Syndrome. *Aging* **2019**, *11*, 3574–3584. [[CrossRef](#)]
115. Jain, M.; Rivera, S.; Monclus, E.A.; Synenki, L.; Zirk, A.; Eisenbart, J.; Feghali-Bostwick, C.; Mutlu, G.M.; Budinger, G.R.S.; Chandel, N.S. Mitochondrial Reactive Oxygen Species Regulate Transforming Growth Factor- β Signaling. *J. Biol. Chem.* **2013**, *288*, 770–777. [[CrossRef](#)]
116. García-Trevijano, E.R.; Iraburu, M.J.; Fontana, L.; Domínguez-Rosales, J.A.; Auster, A.; Covarrubias-Pinedo, A.; Rojkind, M. Transforming Growth Factor Beta1 Induces the Expression of Alpha1(I) Procollagen MRNA by a Hydrogen Peroxide-C/EBPbeta-Dependent Mechanism in Rat Hepatic Stellate Cells. *Hepatology* **1999**, *29*, 960–970. [[CrossRef](#)]
117. Herrera, B.; Murillo, M.M.; Álvarez-Barrientos, A.; Beltrán, J.; Fernández, M.; Fabregat, I. Source of Early Reactive Oxygen Species in the Apoptosis Induced by Transforming Growth Factor-Beta in Fetal Rat Hepatocytes. *Free Radic. Biol. Med.* **2004**, *36*, 16–26. [[CrossRef](#)]
118. Franklin, C.C.; Rosenfeld-Franklin, M.E.; White, C.; Kavanagh, T.J.; Fausto, N. TGFbeta1-Induced Suppression of Glutathione Antioxidant Defenses in Hepatocytes: Caspase-Dependent Post-Translational and Caspase-Independent Transcriptional Regulatory Mechanisms. *FASEB J.* **2003**, *17*, 1535–1537. [[CrossRef](#)]
119. Samarakoon, R.; Chitnis, S.S.; Higgins, S.P.; Higgins, C.E.; Krepinsky, J.C.; Higgins, P.J. Redox-Induced Src Kinase and Caveolin-1 Signaling in TGF-B1-Initiated SMAD2/3 Activation and PAI-1 Expression. *PLoS ONE* **2011**, *6*, e22896. [[CrossRef](#)]
120. Furukawa, F.; Matsuzaki, K.; Mori, S.; Tahashi, Y.; Yoshida, K.; Sugano, Y.; Yamagata, H.; Matsushita, M.; Seki, T.; Inagaki, Y.; et al. P38 MAPK Mediates Fibrogenic Signal through Smad3 Phosphorylation in Rat Myofibroblasts. *Hepatology* **2003**, *38*, 879–889. [[CrossRef](#)]
121. Woodward, R.N.; Finn, A.V.; Dichek, D.A. Identification of Intracellular Pathways through Which TGF-Beta1 Upregulates PAI-1 Expression in Endothelial Cells. *Atherosclerosis* **2006**, *186*, 92–100. [[CrossRef](#)] [[PubMed](#)]
122. Chen, Y.Q.; Su, M.; Walia, R.R.; Hao, Q.; Covington, J.W.; Vaughan, D.E. Sp1 Sites Mediate Activation of the Plasminogen Activator Inhibitor-1 Promoter by Glucose in Vascular Smooth Muscle Cells. *J. Biol. Chem.* **1998**, *273*, 8225–8231. [[CrossRef](#)] [[PubMed](#)]
123. Marsch, E.; Sluimer, J.C.; Daemen, M.J.A.P. Hypoxia in Atherosclerosis and Inflammation. *Curr. Opin. Lipidol.* **2013**, *24*, 393–400. [[CrossRef](#)] [[PubMed](#)]
124. Rey, S.; Semenza, G.L. Hypoxia-Inducible Factor-1-Dependent Mechanisms of Vascularization and Vascular Remodelling. *Cardiovasc. Res.* **2010**, *86*, 236–242. [[CrossRef](#)] [[PubMed](#)]
125. Gao, L.; Chen, Q.; Zhou, X.; Fan, L. The Role of Hypoxia-Inducible Factor 1 in Atherosclerosis. *J. Clin. Pathol.* **2012**, *65*, 872–876. [[CrossRef](#)] [[PubMed](#)]
126. Wang, G.L.; Jiang, B.H.; Rue, E.A.; Semenza, G.L. Hypoxia-Inducible Factor 1 Is a Basic-Helix-Loop-Helix-PAS Heterodimer Regulated by Cellular O₂ Tension. *Proc. Natl. Acad. Sci. USA* **1995**, *92*, 5510–5514. [[CrossRef](#)] [[PubMed](#)]
127. Lin, M.T.; Kuo, I.H.; Chang, C.C.; Chu, C.Y.; Chen, H.Y.; Lin, B.R.; Sureshbabu, M.; Shih, H.J.; Kuo, M.L. Involvement of Hypoxia-Inducing Factor-1 α -Dependent Plasminogen Activator Inhibitor-1 up-Regulation in Cyr61/CCN1-Induced Gastric Cancer Cell Invasion. *J. Biol. Chem.* **2016**, *291*, 27433. [[CrossRef](#)]
128. Sanagawa, A.; Iwaki, S.; Asai, M.; Sakakibara, D.; Norimoto, H.; Sobel, B.E.; Fujii, S. Sphingosine 1-phosphate Induced by Hypoxia Increases the Expression of PAI-1 in HepG2 Cells via HIF-1 α . *Mol. Med. Rep.* **2016**, *14*, 1841–1848. [[CrossRef](#)]
129. Kabei, K.; Tateishi, Y.; Nozaki, M.; Tanaka, M.; Shiota, M.; Osada-Oka, M.; Nishide, S.; Uchida, J.; Nakatani, T.; Tomita, S.; et al. Role of Hypoxia-Inducible Factor-1 in the Development of Renal Fibrosis in Mouse Obstructed Kidney: Special References to HIF-1 Dependent Gene Expression of Profibrogenic Molecules. *J. Pharmacol. Sci.* **2018**, *136*, 31–38. [[CrossRef](#)]
130. Uchiyama, T.; Kurabayashi, M.; Ohyama, Y.; Utsugi, T.; Akuzawa, N.; Sato, M.; Tomono, S.; Kawazu, S.; Nagai, R. Hypoxia Induces Transcription of the Plasminogen Activator Inhibitor-1 Gene through Genistein-Sensitive Tyrosine Kinase Pathways in Vascular Endothelial Cells. *Arterioscler. Thromb. Vasc. Biol.* **2000**, *20*, 1155–1161. [[CrossRef](#)]

131. Kimura, D.; Imaizumi, T.; Tamo, W.; Sakai, T.; Ito, K.; Hatanaka, R.; Yoshida, H.; Tsushima, T.; Satoh, K.; Fukuda, I. Hypoxia Enhances the Expression of Plasminogen Activator Inhibitor-1 in Human Lung Cancer Cells, EBC-1. *Tohoku J. Exp. Med.* **2002**, *196*, 259–267. [[CrossRef](#)] [[PubMed](#)]
132. Toullec, A.; Buard, V.; Rannou, E.; Tarlet, G.; Guipaud, O.; Robine, S.; Iruela-Arispe, M.L.; François, A.; Milliat, F. HIF-1 α Deletion in the Endothelium, but Not in the Epithelium, Protects from Radiation-Induced Enteritis. *Cell. Mol. Gastroenterol. Hepatol.* **2017**, *5*, 15–30. [[CrossRef](#)] [[PubMed](#)]
133. Petry, A.; Belaiba, R.S.; Weitnauer, M.; Görlach, A. Inhibition of Endothelial Nitric Oxide Synthase Increases Capillary Formation via Rac1-Dependent Induction of Hypoxia-Inducible Factor-1 α and Plasminogen Activator Inhibitor-1. *Thromb. Haemost.* **2012**, *108*, 849–862. [[CrossRef](#)] [[PubMed](#)]
134. Görlach, A.; Berchner-Pfannschmidt, U.; Wotzlaw, C.; Cool, R.H.; Fandrey, J.; Acker, H.; Jungermann, K.; Kietzmann, T. Reactive Oxygen Species Modulate HIF-1 Mediated PAI-1 Expression: Involvement of the GTPase Rac1. *Thromb. Haemost.* **2003**, *89*, 926–935. [[CrossRef](#)]
135. Bonello, S.; Zähringer, C.; Belaiba, R.S.; Djordjevic, T.; Hess, J.; Michiels, C.; Kietzmann, T.; Görlach, A. Reactive Oxygen Species Activate the HIF-1 α Promoter via a Functional NF κ B Site. *Arterioscler. Thromb. Vasc. Biol.* **2007**, *27*, 755–761. [[CrossRef](#)]
136. Diebold, I.; Djordjevic, T.; Hess, J.; Görlach, A. Rac-1 Promotes Pulmonary Artery Smooth Muscle Cell Proliferation by Upregulation of Plasminogen Activator Inhibitor-1: Role of NF κ B-Dependent Hypoxia-Inducible Factor-1 α Transcription. *Thromb. Haemost.* **2008**, *100*, 1021–1028. [[CrossRef](#)]
137. An, W.G.; Ahn, Y.-T.; Chua, M.-S.; Whitlock, J.P.; Shin, Y.-C.; Song, W.-H.; Kim, Y.; Eom, C.-Y. Rodent-Specific Hypoxia Response Elements Enhance PAI-1 Expression through HIF-1 or HIF-2 in Mouse Hepatoma Cells. *Int. J. Oncol.* **2010**, *37*, 1627–1638. [[CrossRef](#)]
138. Liao, H.; Hyman, M.C.; Lawrence, D.A.; Pinsky, D.J. Molecular Regulation of the PAI-1 Gene by Hypoxia: Contributions of Egr-1, HIF-1 α , and C/EBP α . *FASEB J.* **2007**, *21*, 935–949. [[CrossRef](#)]
139. Anfosso, F.; Chomiki, N.; Alessi, M.C.; Vague, P.; Juhan-Vague, I. Plasminogen Activator Inhibitor-1 Synthesis in the Human Hepatoma Cell Line Hep G2 Metformin Inhibits the Stimulating Effect of Insulin. *J. Clin. Investig.* **1993**, *91*, 2185–2193. [[CrossRef](#)]
140. Schneider, D.J.; Sobel, B.E. Augmentation of Synthesis of Plasminogen Activator Inhibitor Type 1 by Insulin and Insulin-like Growth Factor Type I: Implications for Vascular Disease in Hyperinsulinemic States. *Proc. Natl. Acad. Sci. USA* **1991**, *88*, 9959–9963. [[CrossRef](#)]
141. Schneider, D.J.; Absher, P.M.; Ricci, M.A. Dependence of Augmentation of Arterial Endothelial Cell Expression of Plasminogen Activator Inhibitor Type 1 by Insulin on Soluble Factors Released from Vascular Smooth Muscle Cells. *Circulation* **1997**, *96*, 2868–2876. [[CrossRef](#)] [[PubMed](#)]
142. Montagnani, M.; Golovchenko, I.; Kim, I.; Koh, G.Y.; Goalstone, M.L.; Mundhekar, A.N.; Johansen, M.; Kucik, D.F.; Quon, M.J.; Draznin, B. Inhibition of Phosphatidylinositol 3-Kinase Enhances Mitogenic Actions of Insulin in Endothelial Cells. *J. Biol. Chem.* **2002**, *277*, 1794–1799. [[CrossRef](#)] [[PubMed](#)]
143. Cusi, K.; Maezono, K.; Osman, A.; Pendergrass, M.; Patti, M.E.; Pratipanawatr, T.; DeFronzo, R.A.; Kahn, C.R.; Mandarino, L.J. Insulin Resistance Differentially Affects the PI 3-Kinase- and MAP Kinase-Mediated Signaling in Human Muscle. *J. Clin. Investig.* **2000**, *105*, 311–320. [[CrossRef](#)] [[PubMed](#)]
144. Alessi, M.C.; Juhan-Vague, I. PAI-1 and the Metabolic Syndrome: Links, Causes, and Consequences. *Arterioscler. Thromb. Vasc. Biol.* **2006**, *26*, 2200–2207. [[CrossRef](#)]
145. Shimomura, I.; Funahashi, T.; Takahashi, M.; Maeda, K.; Kotani, K.; Nakamura, T.; Yamashita, S.; Miura, M.; Fukuda, Y.; Takemura, K.; et al. Enhanced Expression of PAI-1 in Visceral Fat: Possible Contributor to Vascular Disease in Obesity. *Nat. Med.* **1996**, *2*, 800–803. [[CrossRef](#)] [[PubMed](#)]
146. Bouarab, C.; Roullot-Lacarrière, V.; Vallée, M.; Le Roux, A.; Guette, C.; Mennesson, M.; Marighetto, A.; Desmedt, A.; Piazza, P.V.; Revest, J.M. PAI-1 Protein Is a Key Molecular Effector in the Transition from Normal to PTSD-like Fear Memory. *Mol. Psychiatry* **2021**, *26*, 4968–4981. [[CrossRef](#)]
147. Vaughan, D.E.; Lazos, S.A.; Tong, K. Angiotensin II Regulates the Expression of Plasminogen Activator Inhibitor-1 in Cultured Endothelial Cells. A Potential Link between the Renin-Angiotensin System and Thrombosis. *J. Clin. Investig.* **1995**, *95*, 995–1001. [[CrossRef](#)]
148. Rüster, C.; Wolf, G. Angiotensin II as a Morphogenic Cytokine Stimulating Renal Fibrogenesis. *J. Am. Soc. Nephrol.* **2011**, *22*, 1189–1199. [[CrossRef](#)]
149. Fogari, R.; Zoppi, A.; Mugellini, A.; Maffioli, P.; Lazzari, P.; Derosa, G. Role of Angiotensin II in Plasma PAI-1 Changes Induced by Imidapril or Candesartan in Hypertensive Patients with Metabolic Syndrome. *Hypertens. Res.* **2011**, *34*, 1321–1326. [[CrossRef](#)]
150. Skurk, T.; Lee, Y.M.; Hauner, H. Angiotensin II and Its Metabolites Stimulate PAI-1 Protein Release from Human Adipocytes in Primary Culture. *Hypertension* **2001**, *37*, 1336–1340. [[CrossRef](#)]
151. Fay, W.P.; Parker, A.C.; Condrey, L.R.; Shapiro, A.D. Human Plasminogen Activator Inhibitor-1 (PAI-1) Deficiency: Characterization of a Large Kindred with a Null Mutation in the PAI-1 Gene. *Blood* **1997**, *90*, 204–208. [[CrossRef](#)] [[PubMed](#)]
152. Lee, M.H.; Vosburgh, E.; Anderson, K.; McDonagh, J. Deficiency of Plasma Plasminogen Activator Inhibitor 1 Results in Hyperfibrinolytic Bleeding. *Blood* **1993**, *81*, 2357–2362. [[CrossRef](#)] [[PubMed](#)]
153. Schleef, R.; Higgins, D.L.; Pillemer, E.; Levitt, L.J. Bleeding Diathesis Due to Decreased Functional Activity of Type 1 Plasminogen Activator Inhibitor. *J. Clin. Investig.* **1989**, *83*, 1747–1752. [[CrossRef](#)] [[PubMed](#)]

154. Iwaki, T.; Nagahashi, K.; Kobayashi, T.; Umemura, K.; Terao, T.; Kanayama, N. The First Report of Uncontrollable Subchorionic and Retroplacental Haemorrhage Inducing Preterm Labour in Complete PAI-1 Deficiency in a Human. *Thromb. Res.* **2012**, *129*, e161–e163. [[CrossRef](#)]
155. Mehta, R.; Shapiro, A.D. Plasminogen Activator Inhibitor Type 1 Deficiency. *Haemophilia* **2008**, *14*, 1255–1260. [[CrossRef](#)]
156. Kathiresan, S.; Gabriel, S.B.; Yang, Q.; Lochner, A.L.; Larson, M.G.; Levy, D.; Tofler, G.H.; Hirschhorn, J.N.; O'Donnell, C.J. Comprehensive Survey of Common Genetic Variation at the Plasminogen Activator Inhibitor-1 Locus and Relations to Circulating Plasminogen Activator Inhibitor-1 Levels. *Circulation* **2005**, *112*, 1728–1735. [[CrossRef](#)]
157. Dawson, S.; Hamsten, A.; Wiman, B.; Henney, A.; Humphries, S. Genetic Variation at the Plasminogen Activator Inhibitor-1 Locus Is Associated with Altered Levels of Plasma Plasminogen Activator Inhibitor-1 Activity. *Arterioscler. Thromb. A J. Vasc. Biol.* **1991**, *11*, 183–190. [[CrossRef](#)]
158. Grubic, N.; Stegnar, M.; Peternel, P.; Kaider, A.; Binder, B.R. A Novel G/A and the 4G/5G Polymorphism within the Promoter of the Plasminogen Activator Inhibitor-1 Gene in Patients with Deep Vein Thrombosis. *Thromb. Res.* **1996**, *84*, 431–443. [[CrossRef](#)]
159. Liu, Y.; Cheng, J.; Guo, X.; Mo, J.; Gao, B.; Zhou, H.; Wu, Y.; Li, Z. The Roles of PAI-1 Gene Polymorphisms in Atherosclerotic Diseases: A Systematic Review and Meta-Analysis Involving 149,908 Subjects. *Gene* **2018**, *673*, 167–173. [[CrossRef](#)]
160. Huang, G.; Wang, P.; Li, T.; Deng, X. Genetic Association between Plasminogen Activator Inhibitor-1 Rs1799889 Polymorphism and Venous Thromboembolism: Evidence from a Comprehensive Meta-Analysis. *Clin. Cardiol.* **2019**, *42*, 1232–1238. [[CrossRef](#)]
161. Liang, Z.; Jiang, W.; Ouyang, M.; Yang, K. PAI-1 4G/5G Polymorphism and Coronary Artery Disease Risk: A Meta-Analysis. *Int. J. Clin. Exp. Med.* **2015**, *8*, 2097. [[PubMed](#)]
162. Tsantes, A.E.; Nikolopoulos, G.K.; Bagos, P.G.; Tsiara, C.G.; Kapsimali, V.; Travlou, A.; Vaiopoulos, G. Plasminogen Activator Inhibitor-1 4G/5G Polymorphism and Risk of Ischemic Stroke: A Meta-Analysis. *Blood Coagul. Fibrinolysis* **2007**, *18*, 497–504. [[CrossRef](#)] [[PubMed](#)]
163. Garcíá-González, I.J.; Valle, Y.; Sandoval-Pinto, E.; Valdés-Alvarado, E.; Valdez-Haro, A.; Francisco Munõz-Valle, J.; Flores-Salinas, H.E.; Figuera-Villanueva, L.E.; Dávalos-Rodríguez, N.O.; Padilla-Gutiérrez, J.R. The -844 G>A PAI-1 Polymorphism Is Associated with Acute Coronary Syndrome in Mexican Population. *Dis. Markers* **2015**, 460974. [[CrossRef](#)] [[PubMed](#)]
164. Saidi, S.; Slamia, L.B.; Mahjoub, T.; Ammou, S.B.; Almawi, W.Y. Association of PAI-1 4G/5G and -844G/A Gene Polymorphism and Changes in PAI-1/TPA Levels in Stroke: A Case-Control Study. *J. Stroke Cerebrovasc. Dis.* **2007**, *16*, 153–159. [[CrossRef](#)]
165. Abboud, N.; Ghazouani, L.; Saidi, S.; Ben-Hadj-Khalifa, S.; Addad, F.; Almawi, W.Y.; Mahjoub, T. Association of PAI-1 4G/5G and -844G/A Gene Polymorphisms and Changes in PAI-1/Tissue Plasminogen Activator Levels in Myocardial Infarction: A Case-Control Study. *Genet. Test. Mol. Biomarkers* **2010**, *14*, 23–27. [[CrossRef](#)]
166. Kollabathula, A.; Sharma, S.; Kumar, N.; Ahluwalia, J.; Das, R.; Varma, N.; Rana, S.S. Plasminogen Activator Inhibitor-1 4G/5G Promoter Polymorphism in Adults with Splanchnic Vein Thrombosis: A Case-Control Study. *Indian J. Hematol. Blood Transfus.* **2022**, *38*, 169–172. [[CrossRef](#)]
167. Frischmuth, T.; Hindberg, K.; Aukrust, P.; Ueland, T.; Brækkan, S.K.; Hansen, J.; Morelli, V.M. Elevated Plasma Levels of Plasminogen Activator Inhibitor-1 Are Associated with Risk of Future Incident Venous Thromboembolism. *J. Thromb. Haemost.* **2022**. [[CrossRef](#)]
168. Tofler, G.H.; Massaro, J.; O'Donnell, C.J.; Wilson, P.W.F.; Vasan, R.S.; Sutherland, P.A.; Meigs, J.B.; Levy, D.; D'Agostino, R.B. Plasminogen Activator Inhibitor and the Risk of Cardiovascular Disease: The Framingham Heart Study. *Thromb. Res.* **2016**, *140*, 30–35. [[CrossRef](#)]
169. Meltzer, M.E.; Lisman, T.; De Groot, P.G.; Meijers, J.C.M.; Le Cessie, S.; Doggen, C.J.M.; Rosendaal, F.R. Venous Thrombosis Risk Associated with Plasma Hypofibrinolysis Is Explained by Elevated Plasma Levels of TAFI and PAI-1. *Blood* **2010**, *116*, 113–121. [[CrossRef](#)]
170. Schneiderman, J.; Sawdey, M.S.; Keeton, M.R.; Bordin, G.M.; Bernstein, E.F.; Dille, R.B.; Loskutoff, D.J. Increased Type 1 Plasminogen Activator Inhibitor Gene Expression in Atherosclerotic Human Arteries. *Proc. Natl. Acad. Sci. USA* **1992**, *89*, 6998–7002. [[CrossRef](#)]
171. Jung, R.G.; Motazedian, P.; Ramirez, F.D.; Simard, T.; Di Santo, P.; Visintini, S.; Faraz, M.A.; Labinaz, A.; Jung, Y.; Hibbert, B. Association between Plasminogen Activator Inhibitor-1 and Cardiovascular Events: A Systematic Review and Meta-Analysis. *Thromb. J.* **2018**, *16*, 12. [[CrossRef](#)] [[PubMed](#)]
172. Sugano, T.; Tsuji, H.; Masuda, H.; Nakagawa, K.; Nishimura, H.; Kasahara, T.; Yoshizumi, M.; Nakahara, Y.; Kitamura, H.; Yamada, K.; et al. Plasminogen Activator Inhibitor-1 Promoter 4G/5G Genotype Is Not a Risk Factor for Myocardial Infarction in a Japanese Population. *Blood Coagul. Fibrinolysis* **1998**, *9*, 201–204. [[CrossRef](#)] [[PubMed](#)]
173. Crainich, P.; Jenny, N.S.; Tang, Z.; Arnold, A.M.; Kuller, L.H.; Manolio, T.; Sharrett, A.R.; Tracy, R.P. Lack of Association of the Plasminogen Activator Inhibitor-1 4G/5G Promoter Polymorphism with Cardiovascular Disease in the Elderly. *J. Thromb. Haemost.* **2003**, *1*, 1799–1804. [[CrossRef](#)] [[PubMed](#)]
174. Johansson, L.; Jansson, J.H.; Boman, K.; Nilsson, T.K.; Stegmayr, B.; Hallmans, G. Tissue Plasminogen Activator, Plasminogen Activator Inhibitor-1, and Tissue Plasminogen Activator/Plasminogen Activator Inhibitor-1 Complex as Risk Factors for the Development of a First Stroke. *Stroke* **2000**, *31*, 26–32. [[CrossRef](#)]
175. Folsom, A.R.; Cushman, M.; Heckbert, S.R.; Rosamond, W.D.; Aleksic, N. Prospective Study of Fibrinolytic Markers and Venous Thromboembolism. *J. Clin. Epidemiol.* **2003**, *56*, 598–603. [[CrossRef](#)]

176. Chen, R.; Yan, J.; Liu, P.; Wang, Z.; Wang, C. Plasminogen Activator Inhibitor Links Obesity and Thrombotic Cerebrovascular Diseases: The Roles of PAI-1 and Obesity on Stroke. *Metab. Brain Dis.* **2017**, *32*, 667–673. [[CrossRef](#)]
177. van der Weerd, N.; van Os, H.J.A.; Ali, M.; Schoones, J.W.; van den Maagdenberg, A.M.J.M.; Kruyt, N.D.; Siegerink, B.; Wermer, M.J.H. Sex Differences in Hemostatic Factors in Patients with Ischemic Stroke and the Relation with Migraine—A Systematic Review. *Front. Cell. Neurosci.* **2021**, *15*, 7116. [[CrossRef](#)]
178. Yamamoto, K.; Takeshita, K.; Kojima, T.; Takamatsu, J.; Saito, H. Aging and Plasminogen Activator Inhibitor-1 (PAI-1) Regulation: Implication in the Pathogenesis of Thrombotic Disorders in the Elderly. *Cardiovasc. Res.* **2005**, *66*, 276–285. [[CrossRef](#)]
179. Carmeliet, P.; Kieckens, L.; Schoonjans, L.; Ream, B.; Van Nuffelen, A.; Prendergast, G.; Cole, M.; Bronson, R.; Collen, D.; Mulligan, R.C. Plasminogen Activator Inhibitor-1 Gene-Deficient Mice. I. Generation by Homologous Recombination and Characterization. *J. Clin. Investig.* **1993**, *92*, 2746–2755. [[CrossRef](#)]
180. Eitzman, D.T.; Westrick, R.J.; Xu, Z.; Tyson, J.; Ginsburg, D. Plasminogen Activator Inhibitor-1 Deficiency Protects against Atherosclerosis Progression in the Mouse Carotid Artery. *Blood* **2000**, *96*, 4212–4215. [[CrossRef](#)]
181. Zhu, Y.; Farrehi, P.M.; Fay, W.P. Plasminogen Activator Inhibitor Type 1 Enhances Neointima Formation after Oxidative Vascular Injury in Atherosclerosis-Prone Mice. *Circulation* **2001**, *103*, 3105–3110. [[CrossRef](#)] [[PubMed](#)]
182. Carmeliet, P.; Stassen, J.M.; Schoonjans, L.; Ream, B.; Van Den Oord, J.J.; De Mol, M.; Mulligan, R.C.; Collen, D. Plasminogen Activator Inhibitor-1 Gene-Deficient Mice. II. Effects on Hemostasis, Thrombosis, and Thrombolysis. *J. Clin. Investig.* **1993**, *92*, 2756–2760. [[CrossRef](#)] [[PubMed](#)]
183. Gupta, K.K.; Donahue, D.L.; Sandoval-Cooper, M.J.; Castellino, F.J.; Ploplis, V.A. Plasminogen Activator Inhibitor-1 Protects Mice against Cardiac Fibrosis by Inhibiting Urokinase-Type Plasminogen Activator-Mediated Plasminogen Activation. *Sci. Rep.* **2017**, *7*, 365. [[CrossRef](#)] [[PubMed](#)]
184. Luttun, A.; Lupu, F.; Storkebaum, E.; Hoylaerts, M.F.; Moons, L.; Crawley, J.; Bono, F.; Poole, A.R.; Tipping, P.; Herbert, J.M.; et al. Lack of Plasminogen Activator Inhibitor-1 Promotes Growth and Abnormal Matrix Remodeling of Advanced Atherosclerotic Plaques in Apolipoprotein E-Deficient Mice. *Arterioscler. Thromb. Vasc. Biol.* **2002**, *22*, 499–505. [[CrossRef](#)]
185. Xu, Z.; Castellino, F.J.; Ploplis, V.A. Plasminogen Activator Inhibitor-1 (PAI-1) Is Cardioprotective in Mice by Maintaining Microvascular Integrity and Cardiac Architecture. *Blood* **2010**, *115*, 2038–2047. [[CrossRef](#)]
186. Kremen, M.; Krishnan, R.; Emery, I.; Jie, H.H.; Slezicki, K.I.; Wu, A.; Qian, K.; Du, L.; Plawman, A.; Stempien-Otero, A.; et al. Plasminogen Mediates the Atherogenic Effects of Macrophage-Expressed Urokinase and Accelerates Atherosclerosis in ApoE-Knockout Mice. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 17109–17114. [[CrossRef](#)]
187. Stempien-Otero, A.; Plawman, A.; Meznarich, J.; Dyamenahalli, T.; Otsuka, G.; Dichek, D.A. Mechanisms of Cardiac Fibrosis Induced by Urokinase Plasminogen Activator. *J. Biol. Chem.* **2006**, *281*, 15345–15351. [[CrossRef](#)]
188. Haka, A.S.; Grosheva, I.; Singh, R.K.; Maxfield, F.R. Plasmin Promotes Foam Cell Formation by Increasing Macrophage Catabolism of Aggregated Low-Density Lipoprotein. *Arterioscler. Thromb. Vasc. Biol.* **2013**, *33*, 1768–1778. [[CrossRef](#)]
189. Eren, M.; Gleaves, L.A.; Atkinson, J.B.; King, L.E.; Declerck, P.J.; Vaughan, D.E. Reactive Site-Dependent Phenotypic Alterations in Plasminogen Activator Inhibitor-1 Transgenic Mice. *J. Thromb. Haemost.* **2007**, *5*, 1500–1508. [[CrossRef](#)]
190. Eren, M.; Painter, C.A.; Atkinson, J.B.; Declerck, P.J.; Vaughan, D.E. Age-Dependent Spontaneous Coronary Arterial Thrombosis in Transgenic Mice That Express a Stable Form of Human Plasminogen Activator Inhibitor-1. *Circulation* **2002**, *106*, 491–496. [[CrossRef](#)]
191. Erickson, L.A.; Fici, G.J.; Lund, J.E.; Boyle, T.P.; Polites, H.G.; Marotti, K.R. Development of Venous Occlusions in Mice Transgenic for the Plasminogen Activator Inhibitor-1 Gene. *Nature* **1990**, *346*, 74–76. [[CrossRef](#)] [[PubMed](#)]
192. Fay, W.P.; Garg, N.; Sunkar, M. Vascular Functions of the Plasminogen Activation System. *Arterioscler. Thromb. Vasc. Biol.* **2007**, *27*, 1231–1237. [[CrossRef](#)] [[PubMed](#)]
193. Xu, X.; Wang, H.; Wang, Z.; Xiao, W. Plasminogen Activator Inhibitor-1 Promotes Inflammatory Process Induced by Cigarette Smoke Extraction or Lipopolysaccharides in Alveolar Epithelial Cells. *Exp. Lung Res.* **2009**, *35*, 795–805. [[CrossRef](#)] [[PubMed](#)]
194. Gupta, K.K.; Xu, Z.; Castellino, F.J.; Ploplis, V.A. Plasminogen Activator Inhibitor-1 Stimulates Macrophage Activation through Toll-like Receptor-4. *Biochem. Biophys. Res. Commun.* **2016**, *477*, 503–508. [[CrossRef](#)] [[PubMed](#)]
195. Förstermann, U.; Sessa, W.C. Nitric Oxide Synthases: Regulation and Function. *Eur. Heart J.* **2012**, *33*, 829. [[CrossRef](#)] [[PubMed](#)]
196. Gkaliagkousi, E.; Ferro, A. Nitric Oxide Signalling in the Regulation of Cardiovascular and Platelet Function. *Front. Biosci.* **2011**, *16*, 1873–1897. [[CrossRef](#)]
197. Farah, C.; Michel, L.Y.M.; Balligand, J.L. Nitric Oxide Signalling in Cardiovascular Health and Disease. *Nat. Rev. Cardiol.* **2018**, *15*, 292–316. [[CrossRef](#)]
198. Walford, G.; Loscalzo, J. Nitric Oxide in Vascular Biology. *J. Thromb. Haemost.* **2003**, *1*, 2112–2118. [[CrossRef](#)]
199. Garcia, V.; Sessa, W.C. Endothelial NOS: Perspective and Recent Developments. *Br. J. Pharmacol.* **2019**, *176*, 189–196. [[CrossRef](#)]
200. Shu, X.; Ruddiman, C.A.; Keller, T.C.S.; Keller, A.S.; Yang, Y.Y.; Good, M.E.; Best, A.K.; Columbus, L.; Isakson, B.E. Heterocellular Contact Can Dictate Arterial Function. *Circ. Res.* **2019**, *124*, 1473. [[CrossRef](#)]
201. Childs, B.G.; Li, H.; Van Deursen, J.M. Senescent Cells: A Therapeutic Target for Cardiovascular Disease. *J. Clin. Investig.* **2018**, *128*, 1217. [[CrossRef](#)] [[PubMed](#)]
202. Childs, B.G.; Durik, M.; Baker, D.J.; Van Deursen, J.M. Cellular Senescence in Aging and Age-Related Disease: From Mechanisms to Therapy. *Nat. Med.* **2015**, *21*, 1424. [[CrossRef](#)] [[PubMed](#)]

203. Ermolaeva, M.; Neri, F.; Ori, A.; Rudolph, K.L. Cellular and Epigenetic Drivers of Stem Cell Ageing. *Nat. Rev. Mol. Cell Biol.* **2018**, *19*, 594–610. [[CrossRef](#)] [[PubMed](#)]
204. Coppé, J.P.; Desprez, P.Y.; Krtolica, A.; Campisi, J. The Senescence-Associated Secretory Phenotype: The Dark Side of Tumor Suppression. *Annu. Rev. Pathol.* **2010**, *5*, 99–118. [[CrossRef](#)]
205. Samarakoon, R.; Higgins, S.P.; Higgins, C.E.; Higgins, P.J. The TGF- β 1/P53/PAI-1 Signaling Axis in Vascular Senescence: Role of Caveolin-1. *Biomolecules* **2019**, *9*, 341. [[CrossRef](#)]
206. Yamamoto, K.; Takeshita, K.; Shimokawa, T.; Yi, H.; Isobe, K.I.; Loskutoff, D.J.; Saito, H. Plasminogen Activator Inhibitor-1 Is a Major Stress-Regulated Gene: Implications for Stress-Induced Thrombosis in Aged Individuals. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 890. [[CrossRef](#)]
207. McDonald, A.P.; Meier, T.R.; Hawley, A.E.; Thibert, J.N.; Farris, D.M.; Wroblewski, S.K.; Henke, P.K.; Wakefield, T.W.; Myers, D.D. Aging Is Associated with Impaired Thrombus Resolution in a Mouse Model of Stasis Induced Thrombosis. *Thromb. Res.* **2010**, *125*, 72–78. [[CrossRef](#)]
208. Ota, H.; Akishita, M.; Eto, M.; Iijima, K.; Kaneki, M.; Ouchi, Y. Sirt1 Modulates Premature Senescence-like Phenotype in Human Endothelial Cells. *J. Mol. Cell. Cardiol.* **2007**, *43*, 571–579. [[CrossRef](#)]
209. Kortlever, R.M.; Higgins, P.J.; Bernards, R. Plasminogen Activator Inhibitor-1 Is a Critical Downstream Target of P53 in the Induction of Replicative Senescence. *Nat. Cell Biol.* **2006**, *8*, 878–884. [[CrossRef](#)]
210. Kortlever, R.M.; Nijwening, J.H.; Bernards, R. Transforming Growth Factor- β Requires Its Target Plasminogen Activator Inhibitor-1 for Cytostatic Activity. *J. Biol. Chem.* **2008**, *283*, 24308–24313. [[CrossRef](#)]
211. Boe, A.E.; Eren, M.; Murphy, S.B.; Kamide, C.E.; Ichimura, A.; Terry, D.; McAnally, D.; Smith, L.H.; Miyata, T.; Vaughan, D.E. The PAI-1 Antagonist TM5441 Attenuates L-NAME-Induced Hypertension and Vascular Senescence. *Circulation* **2013**, *128*, 2318. [[CrossRef](#)] [[PubMed](#)]
212. Eren, M.; Boe, A.E.; Murphy, S.B.; Place, A.T.; Nagpal, V.; Morales-Nebreda, L.; Ulrich, D.; Quaggin, S.E.; Scott Budinger, G.R.; Mutlu, G.M.; et al. PAI-1-Regulated Extracellular Proteolysis Governs Senescence and Survival in Klotho Mice. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 7090–7095. [[CrossRef](#)] [[PubMed](#)]
213. Elzi, D.J.; Lai, Y.; Song, M.; Hakala, K.; Weintraub, S.T.; Shii, Y. Plasminogen Activator Inhibitor 1–Insulin-like Growth Factor Binding Protein 3 Cascade Regulates Stress-Induced Senescence. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 12052–12057. [[CrossRef](#)] [[PubMed](#)]
214. Ghosh, A.K.; Rai, R.; Park, K.E.; Eren, M.; Miyata, T.; Wilsbacher, L.D.; Vaughan, D.E. A Small Molecule Inhibitor of PAI-1 Protects against Doxorubicin-Induced Cellular Senescence. *Oncotarget* **2016**, *7*, 72443–72457. [[CrossRef](#)]
215. Zhang, L.-N.; Parkinson, J.; Haskell, C.; Wang, Y.-X. Mechanisms of Intimal Hyperplasia Learned from a Murine Carotid Artery Ligation Model. *Curr. Vasc. Pharmacol.* **2008**, *6*, 37–43. [[CrossRef](#)]
216. Pandolfi, A.; Cetrullo, D.; Polishuck, R.; Alberta, M.M.; Calafiore, A.; Pellegrini, G.; Vitacolonna, E.; Capani, F.; Consoli, A. Plasminogen Activator Inhibitor Type 1 Is Increased in the Arterial Wall of Type II Diabetic Subjects. *Arterioscler. Thromb. Vasc. Biol.* **2001**, *21*, 1378–1382. [[CrossRef](#)]
217. Stefansson, S.; Lawrence, D.A. The Serpin PAI-1 Inhibits Cell Migration by Blocking Integrin Alpha V Beta 3 Binding to Vitronectin. *Nature* **1996**, *383*, 441–443. [[CrossRef](#)]
218. Brown, S.L.; Lundgren, C.H.; Nordt, T.; Fujii, S. Stimulation of Migration of Human Aortic Smooth Muscle Cells by Vitronectin: Implications for Atherosclerosis. *Cardiovasc. Res.* **1994**, *28*, 1815–1820. [[CrossRef](#)]
219. Degryse, B.; Neels, J.G.; Czekay, R.P.; Aertgeerts, K.; Kamikubo, Y.I.; Loskutoff, D.J. The Low Density Lipoprotein Receptor-Related Protein Is a Motogenic Receptor for Plasminogen Activator Inhibitor-1. *J. Biol. Chem.* **2004**, *279*, 22595–22604. [[CrossRef](#)]
220. Simone, T.M.; Higgins, S.P.; Archambeault, J.; Higgins, C.E.; Ginnan, R.G.; Singer, H.; Higgins, P.J. A Small Molecule PAI-1 Functional Inhibitor Attenuates Neointimal Hyperplasia and Vascular Smooth Muscle Cell Survival by Promoting PAI-1 Cleavage. *Cell. Signal.* **2015**, *27*, 923. [[CrossRef](#)]
221. Hermann, D.M.; Bassetti, C.L. Sleep-Related Breathing and Sleep-Wake Disturbances in Ischemic Stroke. *Neurology* **2009**, *73*, 1313–1322. [[CrossRef](#)] [[PubMed](#)]
222. Garcia-Rio, F.; Alonso-Fernández, A.; Armada, E.; Mediano, O.; Lores, V.; Rojo, B.; Fernández-Lahera, J.; Fernández-Navarro, I.; Carpio, C.; Ramírez, T. CPAP Effect on Recurrent Episodes in Patients with Sleep Apnea and Myocardial Infarction. *Int. J. Cardiol.* **2013**, *168*, 1328–1335. [[CrossRef](#)] [[PubMed](#)]
223. Badran, M.; Ayas, N.; Laher, I. Cardiovascular Complications of Sleep Apnea: Role of Oxidative Stress. *Oxid. Med. Cell. Longev.* **2014**, *985258*. [[CrossRef](#)] [[PubMed](#)]
224. Zapater, A.; Sánchez-De-La-Torre, M.; Benítez, I.D.; Targa, A.; Bertran, S.; Torres, G.; Aldomà, A.; de Batlle, J.; Abad, J.; Duran-Cantolla, J.; et al. The Effect of Sleep Apnea on Cardiovascular Events in Different Acute Coronary Syndrome Phenotypes. *Am. J. Respir. Crit. Care Med.* **2020**, *202*, 1698–1706. [[CrossRef](#)]
225. Cortese, R.; Gileles-Hillel, A.; Khalyfa, A.; Almendros, I.; Akbarpour, M.; Khalyfa, A.A.; Qiao, Z.; Garcia, T.; Andrade, J.; Gozal, D. Aorta Macrophage Inflammatory and Epigenetic Changes in a Murine Model of Obstructive Sleep Apnea: Potential Role of CD36. *Sci. Rep.* **2017**, *7*, 43648. [[CrossRef](#)] [[PubMed](#)]
226. May, A.M.; Wang, L.; Strohl, K.P.; Walia, H.; Hazen, S.L.; Mehra, R. Sex-Specific Differential Responses of Circulating Biomarkers in Obstructive Sleep Apnea Treatment A Post Hoc Analysis of a Randomized Controlled Trial. *Ann. Am. Thorac. Soc.* **2020**, *17*, 605–613. [[CrossRef](#)] [[PubMed](#)]

227. Bikov, A.; Meszaros, M.; Schwarz, E.I. Coagulation and Fibrinolysis in Obstructive Sleep Apnoea. *Int. J. Mol. Sci.* **2021**, *22*, 2834. [[CrossRef](#)]
228. Seckin, Z.I.; Helmi, H.; Weister, T.J.; Lee, A.; Festic, E. Acute Pulmonary Embolism in Patients with Obstructive Sleep Apnea: Frequency, Hospital Outcomes, and Recurrence. *J. Clin. Sleep Med.* **2020**, *16*, 1029–1036. [[CrossRef](#)]
229. Lippi, G.; Mattiuzzi, C.; Franchini, M. Sleep Apnea and Venous Thromboembolism. A Systematic Review. *Thromb. Haemost.* **2015**, *114*, 958–963. [[CrossRef](#)]
230. García-Ortega, A.; Mañas, E.; López-Reyes, R.; Selma, M.J.; García-Sánchez, A.; Oscullo, G.; Jiménez, D.; Martínez-García, M.Á. Obstructive Sleep Apnoea and Venous Thromboembolism: Pathophysiological Links and Clinical Implications. *Eur. Respir. J.* **2019**, *53*, 1800893. [[CrossRef](#)]
231. Park, A.M.; Suzuki, Y.J. Effects of Intermittent Hypoxia on Oxidative Stress-Induced Myocardial Damage in Mice. *J. Appl. Physiol.* **2007**, *102*, 1806–1814. [[CrossRef](#)] [[PubMed](#)]
232. Rosa, D.P.; Martinez, D.; Picada, J.N.; Semedo, J.G.; Marroni, N.P. Hepatic Oxidative Stress in an Animal Model of Sleep Apnoea: Effects of Different Duration of Exposure. *Comp. Hepatol.* **2011**, *10*, 1. [[CrossRef](#)] [[PubMed](#)]
233. Jun, J.; Reinke, C.; Bedja, D.; Berkowitz, D.; Bevans-Fonti, S.; Li, J.; Barouch, L.A.; Gabrielson, K.; Polotsky, V.Y. Effect of Intermittent Hypoxia on Atherosclerosis in Apolipoprotein E-Deficient Mice. *Atherosclerosis* **2010**, *209*, 381–386. [[CrossRef](#)] [[PubMed](#)]
234. Yamauchi, M.; Nakano, H.; Maekawa, J.; Okamoto, J.; Ohnishi, Y.; Suzuki, T.; Kimura, H. Oxidative Stress in Obstructive Sleep Apnea. *Chest* **2005**, *127*, 1674–1679. [[CrossRef](#)]
235. Vatansever, E.; Surmen-Gur, E.; Ursavas, A.; Karadag, M. Obstructive Sleep Apnea Causes Oxidative Damage to Plasma Lipids and Proteins and Decreases Adiponectin Levels. *Sleep Breath.* **2011**, *15*, 275–282. [[CrossRef](#)]
236. Kizawa, T.; Nakamura, Y.; Takahashi, S.; Sakurai, S.; Yamauchi, K.; Inoue, H. Pathogenic Role of Angiotensin II and Oxidised LDL in Obstructive Sleep Apnoea. *Eur. Respir. J.* **2009**, *34*, 1390–1398. [[CrossRef](#)]
237. Maniaci, A.; Iannella, G.; Cocuzza, S.; Vicini, C.; Magliulo, G.; Ferlito, S.; Cammaroto, G.; Meccariello, G.; De Vito, A.; Nicolai, A.; et al. Oxidative Stress and Inflammation Biomarker Expression in Obstructive Sleep Apnea Patients. *J. Clin. Med.* **2021**, *10*, 277. [[CrossRef](#)]
238. Li, Q.; Zheng, X.; Li, Q.; Zheng, X. Tumor Necrosis Factor Alpha Is a Promising Circulating Biomarker for the Development of Obstructive Sleep Apnea Syndrome: A Meta-Analysis. *Oncotarget* **2017**, *8*, 27616–27626. [[CrossRef](#)]
239. Schulz, R.; Mahmoudi, S.; Hattar, K.; Sibelius, U.L.F.; Olschewski, H.; Mayer, K.; Seeger, W.; Grimminger, F. Enhanced Release of Superoxide from Polymorphonuclear Neutrophils in Obstructive Sleep Apnea. Impact of Continuous Positive Airway Pressure Therapy. *Am. J. Respir. Crit. Care Med.* **2000**, *162*, 566–570. [[CrossRef](#)]
240. Dyugovskaya, L.; Lavie, P.; Lavie, L. Increased Adhesion Molecules Expression and Production of Reactive Oxygen Species in Leukocytes of Sleep Apnea Patients. *Am. J. Respir. Crit. Care Med.* **2002**, *165*, 934–939. [[CrossRef](#)]
241. Imani, M.M.; Sadeghi, M.; Khazaie, H.; Emami, M.; Sadeghi Bahmani, D.; Brand, S. Evaluation of Serum and Plasma Interleukin-6 Levels in Obstructive Sleep Apnea Syndrome: A Meta-Analysis and Meta-Regression. *Front. Immunol.* **2020**, *11*, 1343. [[CrossRef](#)] [[PubMed](#)]
242. Unnikrishnan, D.; Jun, J.; Polotsky, V. Inflammation in Sleep Apnea: An Update. *Rev. Endocr. Metab. Disord.* **2015**, *16*, 25. [[CrossRef](#)] [[PubMed](#)]
243. Gozal, D.; Kheirandish-Gozal, L. Cardiovascular Morbidity in Obstructive Sleep Apnea: Oxidative Stress, Inflammation, and Much More. *Am. J. Respir. Crit. Care Med.* **2008**, *177*, 369–375. [[CrossRef](#)]
244. Kheirandish-Gozal, L.; Gozal, D. Obstructive Sleep Apnea and Inflammation: Proof of Concept Based on Two Illustrative Cytokines. *Int. J. Mol. Sci.* **2019**, *20*, 459. [[CrossRef](#)]
245. Htoo, A.K.; Greenberg, H.; Tongia, S.; Chen, G.; Henderson, T.; Wilson, D.; Liu, S.F. Activation of Nuclear Factor KappaB in Obstructive Sleep Apnea: A Pathway Leading to Systemic Inflammation. *Sleep Breath.* **2006**, *10*, 43–50. [[CrossRef](#)] [[PubMed](#)]
246. Cao, Y.; Song, Y.; Ning, P.; Zhang, L.; Wu, S.; Quan, J.; Li, Q. Association between Tumor Necrosis Factor Alpha and Obstructive Sleep Apnea in Adults: A Meta-Analysis Update. *BMC Pulm. Med.* **2020**, *20*, 215. [[CrossRef](#)] [[PubMed](#)]
247. Prabhakar, N.R.; Peng, Y.J.; Nanduri, J. Hypoxia-Inducible Factors and Obstructive Sleep Apnea. *J. Clin. Investig.* **2020**, *130*, 5042–5051. [[CrossRef](#)]
248. Steffanina, A.; Proietti, L.; Antonaglia, C.; Palange, P.; Angelici, E.; Canipari, R. The Plasminogen System and Transforming Growth Factor- β in Subjects with Obstructive Sleep Apnea Syndrome: Effects of CPAP Treatment. *Respir. Care* **2015**, *60*, 1643–1651. [[CrossRef](#)]
249. Lin, C.C.; Liaw, S.F.; Chiu, C.H.; Lin, M.W. Effects of Continuous Positive Airway Pressure on Exhaled Transforming Growth Factor- β and Vascular Endothelial Growth Factor in Patients with Obstructive Sleep Apnea. *J. Thorac. Dis.* **2020**, *12*, 932–941. [[CrossRef](#)]
250. Zhou, J.P.; Lin, Y.N.; Li, N.; Sun, X.W.; Ding, Y.J.; Yan, Y.R.; Zhang, L.; Li, Q.Y. Angiotensin-(1-7) Rescues Chronic Intermittent Hypoxia-Aggravated TGF- β -Mediated Airway Remodeling in Murine and Cellular Models of Asthma. *J. Pharmacol. Exp. Ther.* **2020**, *375*, 268–275. [[CrossRef](#)]
251. Ding, W.X.; Dong, Y.B.; Ding, N.; Zhang, X.F.; Zhang, S.J.; Zhang, X.L.; Liu, J.N.; Lu, G. Adiponectin Protects Rat Heart from Left Ventricular Remodeling Induced by Chronic Intermittent Hypoxia via Inhibition of TGF- β /Smad2/3 Pathway. *J. Thorac. Dis.* **2014**, *6*, 1278. [[CrossRef](#)] [[PubMed](#)]

252. Abuyassin, B.; Badran, M.; Ayas, N.T.; Laher, I. Intermittent Hypoxia Causes Histological Kidney Damage and Increases Growth Factor Expression in a Mouse Model of Obstructive Sleep Apnea. *PLoS ONE* **2018**, *13*, e0192084. [[CrossRef](#)] [[PubMed](#)]
253. Resta, O.; Foschino-Barbaro, M.P.; Legari, G.; Talamo, S.; Bonfitto, P.; Palumbo, A.; Minenna, A.; Giorgino, R.; De Pergola, G. Sleep-Related Breathing Disorders, Loud Snoring and Excessive Daytime Sleepiness in Obese Subjects. *Int. J. Obes. Relat. Metab. Disord.* **2001**, *25*, 669–675. [[CrossRef](#)] [[PubMed](#)]
254. Castaneda, A.; Jauregui-Maldonado, E.; Ratnani, I.; Varon, J.; Surani, S. Correlation between Metabolic Syndrome and Sleep Apnea. *World J. Diabetes* **2018**, *9*, 66. [[CrossRef](#)]
255. Fallahi, A.; Jamil, D.I.; Karimi, E.B.; Baghi, V.; Gheshlagh, R.G. Prevalence of Obstructive Sleep Apnea in Patients with Type 2 Diabetes: A Systematic Review and Meta-Analysis. *Diabetes Metab. Syndr.* **2019**, *13*, 2463–2468. [[CrossRef](#)]
256. Patel, A.R.; Patel, A.R.; Singh, S.; Singh, S.; Khawaja, I. The Association of Obstructive Sleep Apnea and Hypertension. *Cureus* **2019**, *11*, e4858. [[CrossRef](#)]
257. Kato, M.; Roberts-Thomson, P.; Phillips, B.G.; Haynes, W.G.; Winnicki, M.; Accurso, V.; Somers, V.K. Impairment of Endothelium-Dependent Vasodilation of Resistance Vessels in Patients with Obstructive Sleep Apnea. *Circulation* **2000**, *102*, 2607–2610. [[CrossRef](#)]
258. Gozal, D.; Kheirandish-Gozal, L.; Serpero, L.D.; Capdevila, O.S.; Dayyat, E. Obstructive Sleep Apnea and Endothelial Function in School-Aged Nonobese Children: Effect of Adenotonsillectomy. *Circulation* **2007**, *116*, 2307–2314. [[CrossRef](#)]
259. Harki, O.; Boete, Q.; Pépin, J.-L.; Arnaud, C.; Belaidi, E.; Faury, G.; Khouri, C.; Briançon-Marjollet, A. Intermittent Hypoxia-Related Alterations in Vascular Structure and Function: A Systematic Review and Meta-Analysis of Rodent Data. *Eur. Respir. J.* **2021**, *59*, 2100866. [[CrossRef](#)]
260. Varadharaj, S.; Porter, K.; Pleister, A.; Wannemacher, J.; Sow, A.; Jarjoura, D.; Zweier, J.L.; Khayat, R.N. Endothelial Nitric Oxide Synthase Uncoupling: A Novel Pathway in OSA Induced Vascular Endothelial Dysfunction. *Respir. Physiol. Neurobiol.* **2014**, *207*, 40–47. [[CrossRef](#)]
261. Badran, M.; Abuyassin, B.; Ayas, N.; Laher, I. Intermittent Hypoxia Impairs Uterine Artery Function in Pregnant Mice. *J. Physiol.* **2019**, *597*, 2639–2650. [[CrossRef](#)] [[PubMed](#)]
262. Sharma, P.; Dong, Y.; Somers, V.K.; Peterson, T.E.; Zhang, Y.; Wang, S.; Li, G.; Singh, P. Intermittent Hypoxia Regulates Vasoactive Molecules and Alters Insulin-Signaling in Vascular Endothelial Cells. *Sci. Rep.* **2018**, *8*, 14110. [[CrossRef](#)] [[PubMed](#)]
263. Zhou, M.; Guo, B.; Wang, Y.; Yan, D.; Lin, C.; Shi, Z. The Association between Obstructive Sleep Apnea and Carotid Intima-Media Thickness: A Systematic Review and Meta-Analysis. *Angiology* **2017**, *68*, 575–583. [[CrossRef](#)] [[PubMed](#)]
264. Turkiewicz, S.; Ditmer, M.; Sochal, M.; Białasiewicz, P.; Strzelecki, D.; Gabryelska, A. Obstructive Sleep Apnea as an Acceleration Trigger of Cellular Senescence Processes through Telomere Shortening. *Int. J. Mol. Sci.* **2021**, *22*, 12536. [[CrossRef](#)]
265. Khalyfa, A.; Marin, J.M.; Qiao, Z.; Rubio, D.S.; Kheirandish-Gozal, L.; Gozal, D. Plasma Exosomes in OSA Patients Promote Endothelial Senescence: Effect of Long-Term Adherent Continuous Positive Airway Pressure. *Sleep* **2020**, *43*, 217. [[CrossRef](#)]
266. Cortese, R.; Sanz-Rubio, D.; Kheirandish-Gozal, L.; Marin, J.M.; Gozal, D. Epigenetic Age Acceleration in Obstructive Sleep Apnea Is Reversible with Adherent Treatment. *Eur. Respir. J.* **2022**, *59*, 2103042. [[CrossRef](#)]
267. Wang, L.; Chen, L.; Liu, Z.; Liu, Y.; Luo, M.; Chen, N.; Deng, X.; Luo, Y.; He, J.; Zhang, L.; et al. PAI-1 Exacerbates White Adipose Tissue Dysfunction and Metabolic Dysregulation in High Fat Diet-Induced Obesity. *Front. Pharmacol.* **2018**, *9*, 1087. [[CrossRef](#)]
268. Ichimura, A.; Matsumoto, S.; Suzuki, S.; Dan, T.; Yamaki, S.; Sato, Y.; Kiyomoto, H.; Ishii, N.; Okada, K.; Matsuo, O.; et al. A Small Molecule Inhibitor to Plasminogen Activator Inhibitor 1 Inhibits Macrophage Migration. *Arterioscler. Thromb. Vasc. Biol.* **2013**, *33*, 935–942. [[CrossRef](#)]
269. Khoukaz, H.B.; Ji, Y.; Braet, D.J.; Vadali, M.; Abdelhamid, A.A.; Emal, C.D.; Lawrence, D.A.; Fay, W.P. Drug Targeting of Plasminogen Activator Inhibitor-1 Inhibits Metabolic Dysfunction and Atherosclerosis in a Murine Model of Metabolic Syndrome. *Arterioscler. Thromb. Vasc. Biol.* **2020**, *40*, 1479–1490. [[CrossRef](#)]