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Regulatory T cells (Tregs) and their therapeutic potential against autoimmune disorders – Advances and challenges

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ABSTRACT

Autoimmune diseases are caused when immune cells act against self-protein. This biological self-non-self -discrimination phenomenon is controlled by a distinct group of lymphocytes known as regulatory T cells (Tregs), which are key inflammatory response regulators and play a pivotal role in immune tolerance and homeostasis. Treg-mediated robust immunosuppression provides self-tolerance and protection against autoimmune diseases. However, once this system fails to operate or poorly operate, it leads to an extreme situation where immune system reacts against self-antigens and destroys host organs, thus causing autoimmune diseases. Tregs can target both innate and adaptive immunity via modulating multiple immune cells such as neutrophils, monocytes, antigen-presenting cells, B cells, and T cells. This review highlights the Treg-mediated immunosuppression, role of several markers and their interplay during Treg development and differentiation, and advances in therapeutic aspects of Treg cells to reduce severity of autoimmunity-related conditions along with emphasizing limitations and challenges of their usages.

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Introduction

Autoimmune diseases are no longer neglected and selected practitioners treat them using immune therapy. Autoimmune disorders, which lead to chronic illnesses, are one of the leading causes of death. The primary cause of autoimmune disorders is unknown; however, a lack of proper immunological response is a general cause of all autoimmune conditions.¹ The development of autoimmune diseases is a subject of scientific debate with several tricky questions and incomplete answers to be addressed appropriately. Presently, the numbers of cases of autoimmune diseases have increased. However, while all individuals do not suffer from autoimmune diseases, a considerable population contract some form of this disease. Autoimmune diseases are long-term diseases initiated due to an autoimmune state of affairs that leads to the destruction of organs due to humoral and cell-mediated immune mechanisms. Each autoimmune disorder is unique in nature. In autoimmune disease, the most affected parts involve joints in rheumatoid arthritis (RA), pancreatic cells in type 1 diabetes mellitus (T1D), skin, kidney, serous membranes in systemic lupus erythematosus (SLE), mucosal cells of intestine in inflammatory bowel disease and hepatic cells in autoimmune hepatitis. Autoimmune diseases, such as multiple sclerosis, systemic lupus erythematosus, Guillain-Bare syndrome, and Crohn's disease, which cause

chronic immune response against host cells, are some of the most problematic and challenging disease conditions handled by general practitioners. Though success is uncertain, all efforts are given to cure the disease with targeted treatment.

Human immune system recognizes and reacts against foreign protein, but rarely toward its own (self). The immune system sustains this great responsibility because incidentally originating self-reacting lymphocytes are removed by a process known as programmed cell death, also known as apoptosis, beginning from the embryonic stage. However, we cannot expect all self-reactive lymphocytes to be eliminated as the process to distinguish self from non-self is not 100% full proof. Under physiological selection processes, some selfreacting lymphocytes still evade apoptosis, which may affect in the future. Similarly, it is also difficult to explain why a large number of people never contract autoimmune disease in the presence of autoreactive lymphocytes in circulation, while a few individuals definitely suffer and occasionally respond to clinical treatments.² Adequate justification has never been delivered to address this complex issue. However, regulatory T cells (Tregs) have been reported to significantly control this auto reactivity and reduce the adverse effects, so that humans do not suffer from autoimmunity-mediated pathological ailments.

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Tregs are a specialized T cell subpopulation with specific regulatory mechanisms that inhibit the core components of adaptive and innate immune responses. Human Tregs are characterized with Forkhead box P3 (Foxp3) protein along with low and high expression of CD45RA marker. Functionally and phenotypically three distinct subpopulations of Tregs are (1) CD45RA+FoxP3 low, resting Tregs (2) CD45RA-FoxP3, high activated Tregs, both are suppressive Tregs and the last one (3) is cytokine secreting CD45RA-FoxP3, low nonsuppressive T cells.³

Tregs also play a crucial role in immune homeostasis and self-tolerance.^{4,5} As Treg cells with Foxp3 and CD4⁺ maintain immune homeostasis, several studies have reported that Tregs can help to develop efficient and reliable therapeutic regimens to cure autoimmune disorders and several life-threatening allergic reactions.^{1,6–10} Furthermore, the efficient functioning of Treg requires the expression of not only Foxp3, but also several other Treg signature genes, including CD25 and cytotoxic T lymphocyte antigen 4 (CTLA-4). Moreover, antigen-specific Tregs efficiently control immune response severity for treating autoimmune disorders.

Whether Treg-based cellular therapies may offer a reliable and productive therapeutic regimen for treating autoimmune diseases remains questionable despite the strong evidence supporting this claim. However, many disagreements and major knowledge gaps, particularly related to the mechanism of Treg suppression, necessitate the need of further detailed studies. This review summarizes current knowledge on several Tregrelated biological aspects, focusing on their morphogenesis, modes of action, clinical strategies for isolation and expansion, and therapeutic abilities in treating autoimmune diseases. Furthermore, several challenges have been discussed, which need to be addressed in future studies.

Dispute on T suppressor cells versus Treg cells

There are two schools of thoughts; one supports T suppressor cells, while the other advocates Tregs that control autoimmune disorders. However, proponents of T suppressor cells have now realized that these cells are indeed a type of Tregs. Tregs are a T cell subtype that suppresses inflammatory events. Scientific evidence suggests that Treg deficiency contributes to autoimmunity.^{11,12}

Place and time of Treg cell birth

Differentiation of thymus-derived Tregs (tTregs) relies on TCR signaling, particularly the strength and duration of the signal decide the fate of T cells in circulation.¹³ These self-reacting non-eliminated T cells remain in the circulation for lifetime, but may not aggressively react. Tregs, like other lymphocyte sets, are generated from multipotent hematopoietic stem cells. During embryonic development in the presence of cytokines and under the guidance of several transcription factors, double positive T cell expressing both CD4 and CD8 undergo extensive positive and negative selection to express either CD4 or CD8 markers. However, few cells acquire distinct phenotypic markers and are designated as Tregs. Thymic stromal cells, including cortical medullary thymic epithelial cells and

dendritic cells inside thymus, contribute to Treg differentiation and selection. Additionally, both interleukin (IL)-2 and IL-7 are essential for T cell development in the thymic microenvironment.¹⁴ The process of Treg development is a complex phenomenon and has been discussed in detail previously.¹⁵ A previous report has indicated that a subset of thymus-derived cells expressing CD4 marker along with high expression of IL-2 receptor alpha (IL-2Ra) chain (CD25) protect thymectomized (surgically removal of thymus) mice from autoimmunity.¹⁶ These specialized cells quench adaptive immune response and inhibit the proliferation of other cells, thereby suppressing autoreactive immune responses. Tregs leave the thymus to enter the circulation for patrolling and regulate the autoimmune reaction. Further, several inflammatory cytokines, such as IL-7 and IL-15, are also essential for Treg differentiation, and they appear to partly substitute IL-2, leading to reduced Foxp3 Treg formation.¹⁷

Apart from thymus-derived Tregs, extrathymic origin of Tregs has been confirmed. Tregs can still be divided into natural/thymic and effector Tregs.^{18,19} The anti-inflammatory intensity of naïve Tregs, which have just left the thymus, is inadequate to exert its complete potential. Naive Treg cells propagate as well as develop into the hyper repressive and immunologically differentiated effector Tregs (eTregs) following TCR induction within the lymph node. These eTregs then inhibit antigen-specific maturation of APCs, such as dendritic cells (DCs). Effector Tregs, on the contrary, consume IL-2 via an increased sensitivity to IL-2 receptors and secrete several inhibitory cytokines, such as IL-10, transforming growth factor beta (TGF- β), and IL-35, to inhibit immune system function in an antigen-nonspecific way. In the animal model, antigenspecific Tregs outperform antigen nonspecific Tregs in terms of immune suppression, while antigen nonspecific Treg cells also suppress immune response.²⁰ Therefore, while Tregs suppression is partly antigen nonspecific or non-dependent, antigen-dependent Treg activation affects the immune system effectively.²¹ Furthermore, the control mechanism of this selection has not been fully discovered yet; however, it can be concluded that two population of Treg cells exists, natural Tregs and induced/adaptive Tregs, which can be generated under the influence of specific cytokines, such as TGF- β and IL-10.²²

Tissue-resident Tregs and their possible origin

Tregs develop multiple phenotypes and functions depending on their surroundings and microenvironment due to plasticity. Previous studies have revealed that Tregs reside in several tissues/organs, including the visceral adipose tissue (VAT), colon, skeletal muscle, and skin, in addition to being produced in lymphoid tissues and serve several purposes. Tregs residing in adipose tissue are primarily responsible for maintaining the balance between immunity and metabolism, while Tregs residing in colon tissues sustain intestinal homeostasis and protect the body from harmful microbes. Mostly under inflammatory settings, Tregs present in skeletal muscle cells essentially preserve muscle cell homeostasis and stimulate skeletal muscle repair. Tregs in the skin influence hair regrowth. Even though the phenotypes and activities of tissue-specific Tregs differ, they are all linked to autoimmune disorders.^{7,23,24} Tregs residing in several physiological niches share fundamental phenotypic markers, perform immunosuppression, and may serve as therapeutic targets for treating specific autoimmune diseases in a given circumstance. On the contrary, Tregs adapt to their surroundings and express tissue-specific factors that play different roles in tissue repair.^{8,23}

Intestinal Tregs are mixed origin as indicated by their considerable phenotypic variability, though >70% colonic Tregs do not possess thymic markers like Helios. Compared to spleen and lymph node Tregs, colonic Tregs exhibit reduced Nrp1 (a critical thymic marker) expression. This indicates that the thymus is not necessary for the origin of intestinal Tregs and implies peripherally derived Treg (pTreg) origin.^{25,26}

Skin Tregs are considered to derive from the thymus that proliferates throughout a particular neonatal timeframe (6–13 post-natal days). Blocking T cell migration from the thymus or lymph nodes during this period with FTY720, a sphingosine-1-phosphate receptor antagonist, ten-fold reduces the cutaneous Treg count. It is also linked to an elevation in thymic Tregs, suggesting that skin Tregs are produced from the thymus. However, the CDR3 sequence in skin Tregs, skin Teffs, and their respective analogs in the spleen or lymph nodes have not been studied thoroughly. Therefore, the origin of skin Tregs remains a mystery.²⁷

Identity mark on Treg

The constitutively expressed transcriptional regulator Foxp3 and CD25 (IL-2Ra) and CTLA4 can be considered as distinguishing cell-specific marker for CD4+ Treg cells. Out of these markers, Foxp3 is more specific, while CD25 and CTLA4 are relatively less specific and also expressed on conventional T cells. However, CD25 and CTLA4 are crucial factors in some Treg-mediated suppression mechanisms that maintain immune homeostasis.²⁸ Foxp3 acts as a transcription factor with a 100 amino acid long conserved DNA binding domain that folds like a winged helix structure, termed the forkhead box, and plays an essential role in self-tolerance maintenance and autoimmune disease prevention by regulating the function of CD4⁺ Treg cells. These molecules have distinct roles in T cell survival and regulate important biological activities. Cell surface markers, such as CD25, are essential for autocrine IL-2 absorption on its own surface. CTLA4 is involved in Tregmediated suppression by downregulating CD80 and CD86 on APCs, whereas Foxp3 is essential for high CD25 and CTLA4 expression. Despite the improvement in Treg biology, no exact markers have been designated to characterize human Tregs. However, the identification of a conserved non-coding element within the Foxp3 gene locus using gene sequencing technique has been identified to be most appropriate target for "real" human Tregs.²⁹

Tregs are found in the thymus at 13 weeks of gestation in humans and express forkhead/winged helix family transcription factor Foxp3 protein. These cells are functionally suppressive when co-cultured with Teffs.³⁰ Currently, CD4 and CD25 expression and absence of the IL-7R α -chain (CD127) is considered to be an authentic marker for Treg purification.³¹

Furthermore, while human T lymphocytes temporarily express Foxp3 in response to TCR stimulus, Foxp3⁺ T cells are functionally and phenotypically heterogeneous in humans. CD127 is expressed at small concentrations by CD25⁺ CD4⁺ Tregs, and some Tregs express CD4, CD25, and CD127. However, CD127 is downregulated in naïve T (nT) cells activated by TCR signaling and Foxp3 expression upregulation, implying that the CD4⁺ CD25⁺ CD127⁺ T cell fraction may be contaminated by a few other stimulated non-Treg cells expressing all the markers.^{31,32} Recent reports also suggest the absence of CD127 on Tregs; however, it is necessary to identify the exact marker proteins on Tregs to develop effective therapeutic strategies. Moreover, Foxp3-expressing Tregs should be distinguished from conventional Foxp3-expressing T cells. The classification, phenotypes, functional characteristics, and balance of Treg cells in the immune response are shown in Figure 1.

Experimental evidence of Treg-mediated suppression

Treg-mediated immunosuppression involves several immune-suppressive mechanisms, including CTLA4mediated APC suppression, IL-2 consumption, and immunosuppressive cytokine and metabolite production.²⁸ Furthermore, the Treg-mediated inhibitory pathways impeding various immune cells have been classified as direct pathways, where Tregs induce a direct response on the immune cell, and indirect pathways, under which another cell or molecule is influenced, which in turn inhibit the immune cells. The release of various cytokines, including IL-10, TGF- β , and IL-35, as well as the synthesis of lytic enzymes such as granzyme and perforin, causes target immune cell death, usually by apoptosis. In addition, Tregs directly suppress target cells by expressing CD39/CD73, which in turn reduces extracellular ATP concentration by producing adenosine and AMP.³³ Tregs, on the contrary, impact changes in the microenvironment due to their increased CD25 expression. Tregs with increased IL-2 receptor expression will bind to the IL-2, effectively depriving the neighboring tissues.³⁴ While determining the mechanism of Treg-mediated suppression, it is necessary to mention that different interventions are used selectively by the wide range of Treg-related target cells and microenvironments.35 In this context, Treg-mediated suppression mechanisms to maintain homeostasis in the immune response have been discussed in detail.

Several findings indicate that immunosuppression is caused by the direct interaction of Tregs with effector T and B cells or suppressive cytokine secretion. It has been observed that Tregs suppress CD4⁺ T cell activation and proliferation both contactdependently and -independently. Co-culturing Tregs (CD4⁺, CD25⁺) with APCs and other antigen-specific responder T cells (CD4⁺ and CD8⁺) in presence of specific antigen, suppresses responder cell proliferation and reduced IL-2 production. Tregs expressing increased IL-2 receptor on its surface bind to the IL-2 produced from responding T cells, thereby preventing their proliferation.

Tregs may interfere with the antigen-presenting ability of DCs or promote the secretion of suppressive factors, such as indolamine 2,3 dioxygenase (IDO), a potent

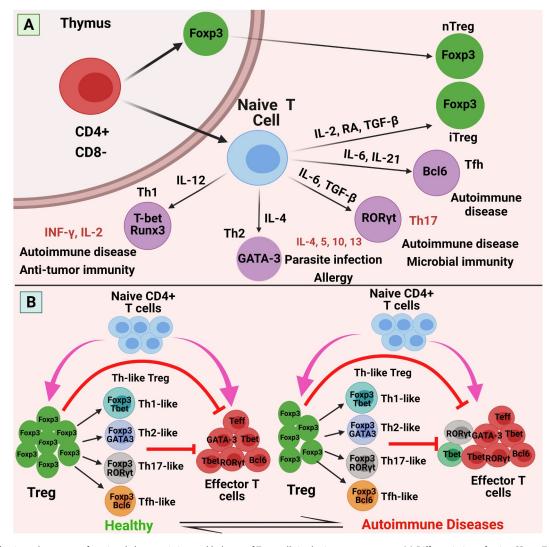


Figure 1. Classification, phenotypes, functional characteristics, and balance of Treg cells in the immune response. (a) Differentiation of naive CD4 + T cells into Tregs or effector T cells. The selection of naïve CD4 + T cells and natural Tregs occurs in the thymus. Naïve CD4 + T cells, subsequently, can differentiate into various different T cell subsets: Th1, Th2, Th17, induced Tregs (iTregs), in the periphery, all heralding distinct immunological functions. These differentiation programs are regulated by different cytokines. The transcription factors T-bet and Runx3, GATA3, or RORγt, are required to differentiate naive T cells into Th1, Th2, or Th17 cells, respectively. For example: T-bet (Th1 cells), GATA3 (Th2 cells), RORγt (Th17 cells), FOXP3 (Tregs). (b) In a healthy individual, the immune system is under regular homeostasis, can suppress autoreactive effector T cells and control a fine balance (left). Aberrant Treg plasticity, quantitative and functional deficiencies of Treg impair immune homeostasis and result in autoimmune diseases (right). Abbreviations: nTreg: natural Treg; iTreg: induced Treg; FOXP3: forkhead Box P3; RA: retinoic acid; CD: cluster of differentiation; IFN: interferon; IL: interleukin; RORγt: retinoid related orphan receptor γ; T-bet: T box transcription factor; TCR: T cell receptor; TGF-β: transforming growth factor-β; Th: T helper cell; GATA3: GATA3-binding factor 3; Tfh: follicular helper T cells; Bcl: B-cell lymphoma 2. Figure was designed by Biorender.com program (https://biorender.com/). Accessed on 13 April 2021.

immunosuppressive enzyme, from DC. IDO induction is dependent upon the high expression of CTLA4, an inhibitory receptor on Treg cell. The mechanism is complex and the conclusive effect of CTLA4 is difficult to prove. Blocking CTLA4 does not affect Tregs either in vitro or in vivo. Tregs isolated from CTLA4-deficient mice have a suppressive effect similar to that of wild mice. On the contrary, in vivo CTLA4 activity blocking in mice can abrogate Treg-mediated suppression of inflammatory disease (IBD), autoimmune gastritis, bowel and transplantation.³⁶ Two Treg subpopulation, naïve (nTreg) and others effective (eTreg), are operational in the body. The nTregs express increased levels of immunosuppressive molecules, particularly IL-10.37 Treg cell depletion acts as an adjuvant effect to enhance DCs and hyperactivity is indicated, speculated for check and balance activity. Treg distribution varies according to the physiological requirements in peripheral location, being abundant in the gut mucosa, where tolerance to residential normal microbiota is essentially required.^{38,39}

Tregs inhibit autoreactive B cell ability to develop detrimental autoantibodies, which also destroys autoreactive B cells. Several markers, such as PD-1 expression on autoreactive B cells and the two PD-1 ligands (PDL-1 and -2) expression on Treg cells, are necessary for autoreactive B cell inhibition.⁴⁰ Furthermore, Tregs also use granzyme B (granular enzyme) and perforin (cause perforation) to destroy effector B cells, which in turn reduces autoantibody production.⁴¹

It is well-established that NK cells, which are innate lymphocytes, destroy tumor cells and remain vigilant throughout human life to reduce tumor burden. Unfortunately, Tregs suppress NK cell activity, resulting in tumor progression. Neutrophil dysfunction results in sustained inflammation and autoimmune disease progression.⁴² Direct interaction of Tregs on polymorphs limits granulocyte infiltration by suppressing the expression of chemokines that act as chemoattractants. Co-culturing neutrophils with Tregs under culture vessel enhances IL-10 and IL-6 production, which are considered to be immunosuppressive.⁴³ In addition, Tregs promote heme oxygenase-1 and IDO expression, and cytokine signaling three molecule (SOCS3) inhibition.⁴³ These findings confirm that human Tregs are effective in preventing graft-versus-host disease (GVHD) and autoimmune diseases, while delaying graft rejection.³⁵

Tregs work specifically on monocytes, preventing their differentiation, cytokine secretion, and antigen expression. Following co-culture with Tregs, monocytes display classic M2 macrophage characteristics such as increased CD206 (mannose scavenger receptor) and CD163 (hemoglobin scavenger receptor) expression, as well as a reduced capacity to respond to pro-inflammatory stimuli as evidenced by decreased IL-6 and TNF-β production and NF-κB activation.⁴⁴ Ex vivo extended Tregs are effective at skewing monocytes toward a tolerogenic phenotype. Compared to monocytes co-cultured freshly isolated Tregs, those cocultured with expanded Tregs have a lower capacity to increase harmful IL-17-emitting T cells.⁴⁵ The suppression mechanism correlates with the reduced CD86 expression by Tregconditioned monocytes.

These immunosuppressive mechanisms should be explored in the future to establish the immunosuppressive mechanisms of Tregs under various other autoimmune disorders. The mechanisms underlying Treg-mediated immunosuppression is shown in Figure 2.

Chemical messengers involved in suppression

Although no consensus has been established to register all the known suppressive agents and their hierarchy of action during immunosuppression, the key molecules governing the suppressive activity of Tregs include IL-10, TGF- β , CTLA4, granzyme, perforin, interferon gamma (IFN- γ), IL-9, cAMP, heme oxygenase-1, CD39, galectins, and IL-35. Treg executes suppressive effect due to IL-10 and TGF- β secretion. However, neutralizing antibodies against these two cytokines may not block Tregs activity and mice lacking these two genes also have similar suppressive activity. No abysmal effect was observed in Treg-deficient mice after the genes for TGF β were knocked out. In vivo models have suggested the requirement of these two cytokines in Tregs, whereas others have advocated that these cytokines are secreted by other cells as well.⁴⁶

Furthermore, since Tregs are more vulnerable to oxidative stress than Teffs, NRF2 has been investigated as a primary transcription factor that protects Tregs against oxidative distress. Tregs undergoing apoptosis due to oxidative damage release a considerable amount of ATP. Hence, Tregs containing high levels of CD37 and CD39 represent an important role in catalysis and adenosine generation due to enhanced extracellular ATP or ADP secretion. Adenosine then binds to the A2A receptors of Teffs, leading to immunosuppression.⁴⁷

Forkhead character of Foxp3 protein

During 1949, a spontaneous scurfy mutation in mice colony was observed at Oak Ridge National Laboratory, Tennessee, USA. Subsequent studies have revealed that the mutation is X-linked, thus affecting only males. Homozygous males carrying the mutant X chromosome exhibit this syndrome, whereas heterozygous female carriers of the mutant X chromosome allele are healthy. Scurfy mice have thick and scaly ears, eyelids, feet, and tail and severe runting.48,49 Internal organs of such mice have characteristic lymphadenopathy, splenomegaly, hepatomegaly, and massive lymphocytic infiltration in the skin and liver. These lesions closely resemble those after graftversus-host reaction. Initial study uncovered that mutation in scurfy mice affects Foxp3-encoding gene and can be rescued by introducing wild-type Foxp3 allele transgene. Conventional T cells do not express Foxp3 protein under normal physiological conditions, whereas some CD25⁺/CD4⁺ population highly expresses Foxp3. Ectopic Foxp3 protein expression in isolated CD4⁺/CD25⁺ T cells acquires the suppressor function in non-Tregs. Similarly, foxp3 introduction in naïve T cell confers suppressive function. Adaptive Treg transfer rescues neonatal Foxp3-deficient mice from the disease. Mice experimentally subjected to germ line foxp3 delineation progressively express autoimmune lesions. Even foxp3 knockout from thymic and dendritic cells outlines the repertoire that developing T cells do not affect dysregulation or alteration in T cell differentiation. Similarly, foxp3 knockout macrophage does not affect tumor growth. These data indicate that foxp3 does not affect any other cells except T cells.¹⁴

Moreover, hyperinflammation in various organs and a severe autoimmune response have been reported in scurfy mice with a mutant foxp3. The hyperinflammatory response has been linked with the lack of Tregs, which in turn leads to enhanced production of inflammatory cytokines.⁵⁰ Besides that, in Homo sapiens, mutated foxp3 tends to cause X chromosome-linked immune dysregulation, polyendocrinopathy, and enteropathy (IPEX) syndrome.⁵¹ The overexpression of *foxp3* in nT cells enhances the immunosuppressive activities of T cells. The nT cells with CD4⁺ and CD25⁻ characteristics transform into Treg-like cells expressing both CD4 and CD25 markers when transfected with foxp3 gene. These cells mimic Tregs by producing antiinflammatory cytokines, exhibiting classic Treg-cell characteristics and their marker proteins, including CTLA-4 and TNF receptorrelated proteins, and showing significant immunosuppressive activities.⁵² Furthermore, it has been concluded that foxp3 is a lineage-specific marker and a key factor in the development, repair, and autoimmune suppressive roles of Tregs. Hence, foxp3 can be presumed to be a key regulatory gene in developing Tregs.

Strategic expansion of Treg cells and its therapeutic applications

Trials are being conducted since last 10 years to verify the safety and feasibility of Treg therapy. The first clinical trials with transferred Tregs were initiated in 2009.¹¹ Medical practitioners are now interested in the clinical application of Tregs following all safety guidelines to reduce autoimmune disorders as an alternative approach to anti-inflammatory drugs.^{53,54}

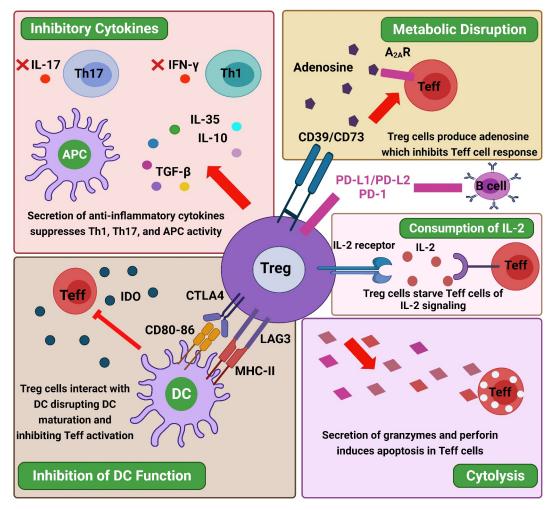


Figure 2. Mechanisms underlying Treg-mediated immunosuppression. Treg cells regulate immune responses by suppressing the functions of effector T cells (Teff) and antigen-presenting cells (APCs) through various mechanisms, including (i) modulation of dendritic cell (DC) function and prevention of DC maturation by the interaction of CTLA4 and LAG3 expressed by Treg cells and the CD80/86 costimulatory molecules and MHC class II expressed by DC, respectively, leading to IDO generation and inhibition of Teff cell activation; (ii) metabolic disruption, Treg cells can disrupt metabolic roles by the expression of the ectoenzymes CD39/73 allowing adenosine generation and binding of adenosine to the adenosine receptor 2A (A2AR) exposed on Teff cells, or by IL-2 deprivation; (iii) generation and secretion of the anti-inflammatory cytokines IL-10, IL-35, and TGF-β that restrain Th1 and Th17 immune responses and the production of IFN-γ and IL-17, respectively; and (iv) direct cytotoxicity, Treg cells can also induce direct killing of effector cells by the release of granzyme A, granzyme B, and perforin, which induce apoptosis in the target cells. Tregs have also been detected to have a direct effect on B-cells via PD-L1/PD-1 interaction. Abbreviations: APC: antigen presenting cell; TGF-β: transforming growth factor-β; A2AR: Adenosine receptor 2A; IL: interleukin; IFN: interferon; Teff: effector T cells; Treg: regulatory T cells; CD: cluster differentiation; IDO: indoleamine 2,3-dioxygenase; DC: dendritic cell; CTLA-4: cytotoxic T lymphocyte antigen-4; TGF-β: transforming growth factor-β; LAG3: lymphocyte-activation gene 3; MHC: major histocompatibility complex; PD-1: programmed death-1; PD-Ls: programmed cell death ligands. Figure was designed by Biorender.com program (https://biorender. com/) Accessed on 13 April 2021.

Adhering to ethical guidelines, Tregs can be isolated from either peripheral blood or umbilical cord blood, while the thymus is also a rich source of Tregs in experimental animals.55-57 Using standard cell isolation procedure employing gradient centrifugation technique, mononuclear cells comprising lymphocytes and monocytes can be isolated. From the isolated mononuclear cell population, further purification and enrichment process is adapted to sort out desired T cell population. The classical approach involves the depletion of CD8 +and/or CD19 + T cells, followed by an enrichment of the CD25 + fraction. This protocol can enrich FOXP3 + expressing T cells to 80% purity due to the presence of cell contaminants. Moreover, flow cytometry-based purification technique is an advanced method of cell isolation with increased purity in the clinics. With this technique, cells are not only selected, but also sorted out for utmost purity (> 99%) and sterility, fulfilling the requirement of in vitro culture. Cells are selected according to

CD4, CD25, and CD127 marker expression.⁵⁸ Treg concentration in cord blood and thymus is high, however it is difficult to isolate. Therefore, post-enrichment Treg expansion in cell culture vessel is essential to achieve optimum concentration for therapeutic applications. Hence, ex vivo polyclonal Treg expansion is now routinely practiced using anti-CD3/CD28coated beads in the presence of IL-2.⁵⁹ Protocol refinement using rapamycin and/or IL-2 cytokine to increase Treg's yield with increased suppressive effect has been followed by several researchers. However, another option is to incorporate TGF β and all trans retinoic acid (ATR) in culture medium for upregulating chemokine receptor expression on Tregs, which may facilitate Treg homing in the gut.

Recently, adoptive cell therapy using genetically engineered Tregs has been attempted to combat transplant rejection and autoimmune diseases.⁶⁰ However, the lack of standardized protocols for expanding and transducing Tregs from mice is a significant drawback of this technique. Purifying, extending, and retrovirally transducing mouse Tregs with a vector encoding a chimeric antigen receptor as a model transgene has been described for obtaining considerable improvement in genetically engineered Tregs. It has been discovered that isolating Tregs based on GFP expression yields appropriately pure cells. Although rapamycin expansion inhibits Treg expansion, it does not influence the suppressive activities of Tregs. This approach enables the study of transduced Tregs in animal models, as well as the investigation of Treg therapy based on genetic engineering for multiple inflammatory diseases.⁶¹ Moreover, in the future, genetically engineered Tregs can solve the issues associated with Treg expansion and purification along with enhancing the suppressive activities of Tregs.

Therapies targeting Treg-mediated suppressive mechanisms have been introduced in several conditions, including organ transplantation and autoimmune disorders. In addition, Tregs have been introduced to cure other clinical manifestations such as cardiovascular disease, obesity, and systemic inflammatory disorders.^{62,63} The therapeutic potentials of Tregs to manage a range of autoimmune diseases have been described in following section and findings of clinical trials are summarized in Table 1.

Inflammatory bowel disease /Crohn`s disease

In vitro-generated, IL-10-producing ovalbumin (OVA)specific Treg cells (Tr1) were reported to prevent colitis induced in SCID mice by CD4+ CD45RBhi T cells. Thus IL-10 secreting CD4+ Treg cells are immunosuppressive and actively downregulate a pathological immune response in vivo.-⁶⁹ Experimental transfer of CD4⁺CD25⁺ Treg cells in immunodeficient mice with clinical signs of colitis was able to ameliorate colitis. Recovery was observed after 2 weeks of postinfusion of Treg cells at a dose of 10⁶ per mice, as compared to transfer of CD4⁻CD25⁻ T cells in control group.⁷⁰ As per published reports, the first clinical trial with single infusion of PBMC-derived freshly isolated Tregs (10^6-10^9) to a patient with active Crohn's disease was safely initiated during 2012 without observation of any severity. Clinical responses were predominantly detected in patients receiving the lowest numbers of Treg cells, suggesting that less suppression may be better.⁶⁴ Crohn's disease has been described in details previously.⁷¹ It has been observed that the inflamed mucosa of IBD cases are modestly populated with Treg cell as compared to other inflammatory conditions like diverticulitis, therefore the inflammation are poorly controlled due to lack of Treg cell number at the site of inflammation.⁷² Knock out mice lacking IL-10 gene is considered to be the suitable model for colitis to establish the efficacy of Treg cell producing IL-10 to reverse the pathological condition once infused, supporting the critical role of IL-10.73 Developing humanized mouse by transplanting fetal small bowel followed by developing colitis by Escherichia coli infection has been used as a model for the study of Treg cell function. After transfusing 10⁶ Treg cells along with rIL-10, it has been observed that Treg cells migrate and preferred to home at the site of inflamed lamina propria and suppress the conventional T cells at the site of inflammation.⁷⁴ Surgically collected specimens from IBD

patients have shown defective CD8+ Treg cells in the lamina propria (LP) whereas CD8⁺ T cells with regulatory activity are present in the LP of healthy individuals. It has been suggested that intestinal epithelial cells play an important role in the generation of such regulatory T cells, because they are able to process and present antigen to T cells. These cells play a central role in maintaining controlled inflammation in the intestinal mucosa.⁷⁵

Type 1 diabetes (T1D)

T1D is a clear example of autoimmune disease as inflammatory cells infiltrate the pancreatic islets and destroy the insulinproducing cells. Recent report has indicated that children with new-onset T1D have an increased proportion of CD45RA(-)CD25(int)FOXP3(low) Treg cells that are not suppressive and secrete significantly more IL-17 than other FOXP3 (+) subsets. Regulatory T cell dysfunctions in T1D have relied on analysis of FOXP3-expressing cells. The number of Tregs reduces during the early phases of the disease.⁷⁶ Proof-ofconcept for the treatment of Type1 diabetes in children by delivering autologous CD3(+)CD4(+)CD25(high)CD127(-) Tregs has been established. Administration of Tregs is safe and tolerable in children with recent-onset of type 1 diabetes.⁵⁸ A randomized trial with patients with T1D in three groups receiving 10×10^6 , 20×10^6 , and 30×10^6 ex vivo expanded autologous Tregs/kg body weight have shown no adverse effect under treatment duration. Safety was proved after one year of follow-up, with eight patients showing signs of clinical remission.⁵⁸ During phase I clinical trials, autologous Tregs derived from patient's blood were administered to evaluate the clinical efficacy. None of the patients showed any unpleasant events up to 31 months of follow-up program, while only three out of 14 patients developed untoward effect without any opportunistic pathogen infection. Further trial with placebo control has shown positive response of Treg therapy to limit the severity of autoimmune disorders.⁶⁶

Hemoglobin concentration and insulin requirement was found to be significantly reduced in some T1D patients after receiving Treg cell therapy. Some patients showed complete independence from the exogenous insulin and revealed significant increase in circulating Tregs. Negligible risks of infections or not having any severe adverse effects were reported following Treg therapy.⁵⁸ Another study has also reported a significant increase in survival of pancreatic β -cells among patients with repeated Treg administration for one year.⁵⁸ Ongoing clinical trials at phase I/II stage (NCT02932826) with Tregs derived from umbilical cord at high and low dose in a single infusion have not shown any adverse effect and final result are yet to be compiled.⁷⁷

Pemphigus vulgaris (PV)

PV is a fatal autoimmune disease caused by IgG autoantibodies targeting the desmosomal adhesion protein desmoglein Dsg1 and Dsg3. The disease is characterized by loss of cell adhesion (also known as acantholysis) in skin and mucous membrane. In pemphigoid diseases, the linear deposition of autoantibodies along the dermal-epidermal junction causes sub-epidermal blistering, resulting in blisters.⁷⁸ Findings of adoptive transfer of splenocytes from Dsg3 -/- mice (mice lacking both the

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Autologous enriched Tregs derived from PBMC, ex vivo expanded NCT01210664 0.5 x 10 ⁸ to 26 × 10 ⁸ Tregs infused as single dose via i/v route per patient NCT01210664 0.5 x 10 ⁸ to 26 × 10 ⁸ Tregs infused as single dose via i/v route per patient 0.5 x 10 ⁸ to 26 × 10 ⁸ Tregs infused as single dose via i/v route per patient CD4 + CD25 + CD127 lowTregs derived from autologous PBMC enriched under in-vitro condition, under Phase-I . 3 x 10 ⁶ - 20 × 10 ⁶ cells per kg, s/c route CD4 + CD25 + CD127 lowTregs derived from autologous PBMC enriched under in-vitro condition, under Phase-I . 3 x 10 ⁶ - 20 × 10 ⁶ cells per kg, s/c route CD4 + CD25 + CD127 lowTregs moder Phase-I . Tregs with insulin and insulin alone MCT0233282.6 Tregs with insulin and insulin alone NCT0233282.6 Tregs with insulin and insulin alone NCT0233382.6 Tregs with insulin and insulin alone NCT0233387.6 Tregs with insulin and insulin alone NCT023339470 10 ¹ 0 ⁸ and NCT02704338 10 -20 × 10 ⁶ cells in a single infusion NCT02704338 Patients aged between 10-70	Type1 diabetes T1D	Autologous ex vivo Tregs CD4+ CD25+ CD127, low ISRCTN06128462	10 x 10 ^{6–} 30 × 0 ⁶ per kg body weight	Absence of serious adverse effect without insulin dependency. Clinical trials completed	58
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Umbilical cord blood derived Tregs with insulin TregsPhase-I and Phase-II trial Tregs with insulin alone NCT02932836 and insulin alone NCT02932836 I x 10 ⁸ deuterium Autologous ex vivo expanded and enriched Tregs, under Phase-I. 1 x 10 ⁸ deuterium Autologous ex vivo expanded and enriched Tregs, under Phase-I. 1 x 10 ⁸ deuterium Ex vivo expanded and onriched Tregs 1 x 10 ⁸ deuterium Ex vivo expanded autologous 2.5 x 10 ⁸ and CD4+ CD127Io/negCD25 2.5 x 10 ⁸ cells per polyclonally expanded autologous Incgs for phase I/II clinical trials 10-20 x 10 ⁶ cells Incgs for phase I/II clinical trials 10-20 x 10 ⁶ cells Incgs for phase I/II clinical trials to patients aged VCT02704338 between 10-70	11D	CD4 + CD25 + CD127 lowTregs derived from autologous PBMC enriched under in-vitro condition, under Phase-I . Clinical trial NCT022772679	3 x 10 ⁶ –20 × 10 ⁶ cells per kg, s/c route	No result posted as on November 2021	35
neous Autologous ex vivo expanded and enriched Tregs, under Phase-I. 1 x 10 ⁸ deuterium labeled Tregs cells Clinical trials NCT02428309 per patients Ex vivo expanded autologous 1.0 x 10 ⁸ and CD4+ CD127lo/negCD25 2.5 x 10 ⁸ cells per polyclonal Tregs ne Polyclonal Tregs ne Polyclonally expanded autologous NCT02204338 10-20 x 10 ⁶ cells ne NCT02704338 nestients age between 10-70 vasts	TID	Umbilical cord blood derived TregsPhase-I and Phase-II trial NCT02932826	Tregs with insulin and insulin alone	40 participants were incorporated in the trial and result not published till Nov 2021	35
Ex vivo expanded autologous 1.0 x 10 ⁸ and Re CD4+ CD127lo/negCD25 2.5 x 10 ⁸ cells per patients of 18 to polyclonal Tregs 75 years age 10-20 x 10 ⁶ cells ne Polyclonally expanded autologous 10-20 x 10 ⁶ cells Ini ne Polyclonally expanded autologous 10-20 x 10 ⁶ cells Ini NCT02704338 tinical trials in a single infusion Normally events	Active cutaneous lupus	Autologous ex vivo expanded and enriched Tregs, under Phase-I. Clinical trials NCT02428309	1 x 10 ⁸ deuterium labeled Tregs cells per patients	Increase population of Tregs and IL-17 production by both CD4 and CD8 cells recorded in skin biopsies. Terminated due to participant recruitment feasibility	67
Polyclonally expanded autologous 10–20 × 10 ⁶ cells Tregs for phase I/II clinical trials in a single infusion NCT 02704338 between 10–70 between 10–70	Pemphigus Vulgaris	Ex vivo expanded autologous CD4+ CD127lo/negCD25 polyclonal Tregs NCT03239470	1.0 x 10 ⁸ and 2.5 x 10 ⁸ cells per patients of 18 to 75 years age	Recovery criteria was to detect CD4+, CD25++, CD127 low Treg in patients using cell shorting system. Results submitted during October 2021 but detail is under quality control process for further recruiting,	8
	Auto immune hepatitis	Polyclonally expanded autologous Tregs for phase I/II clinical trials NCT 02704338	$10-20 \times 10^{\circ}$ cells in a single infusion to patients aged between $10-70$ years	Initiated at Nanjing Medical University China. Further information not yet published	35

Table 1. Clinical trials of Tregs against autoimmune diseases.

parental gene for Dsg3) immunized with recombinant Dsg3 protein to Rag2-/-Dsg3+/+ mice suggested that adoptively transferred Tregs suppressed the antibody production in a dose-dependent manner and the depletion of endogenous Tregs augmented the antibody production.⁷⁹ Similarly, using HLA-DRB104:02 transgenic PV mouse model immunized with human Dsg3 established that *in vivo*, although not statistically significant, Treg cells exert a clear down-regulatory effect on the Dsg3-driven T-cell response and, accordingly, the formation of Dsg3-specific IgG antibodies were detected.⁸⁰

To evaluate the effects of Tregs on the manifestation of PV, a nonrandomized, open-label, phase 1 ongoing clinical trial (NCT03239470) is operational since 2017. In this trial, 12 pemphigus and or pemphigoid patients received single infusion of autologous expanded Tregs (CD4+ CD127lo/negCD25 +) at one of the following doses: either 2.5×10^8 poly Tregs or 10×10^8 poly Tregs. As no adverse effect has been observed, the project is continuing and the final results from this study are awaited.⁸¹

Autoimmune hepatitis (AIH)

Tregs have the potential to serve as an effective therapeutic modality in treating autoimmune hepatitis (AIH), a chronic autoimmune inflammatory disease that usually requires lifelong immunosuppression for management of this condition. Relapses following therapy discontinuation are common, indicating that current treatments do not regain intrahepatic immune regulation. Immune response homeostasis can be reestablished by increasing the functionally active intrahepatic Tregs. However, steroid therapy preferentially depletes intrahepatic Tregs. In recent clinical trials, Tregs have been used to improve the autoimmune disorders with a low dose of cytokines such as IL-2. Mice with complexed IL-2/anti-IL-2 have been used to improve the Treg selectivity and a significant improvement in intrahepatic and systemic Treg levels along with a decrease in activated, intrahepatic Teffs have been reported. Reduced levels of liver-specific ALT (alanine transaminase) and a molecular pattern close to that observed in healthy people have also been reported. Finally, complexed IL-2/anti-IL-2 improves the equilibrium between Tregs and Teffs within the liver. Treg-specific IL-2 augmentation aids in reestablishing immune tolerance and homeostasis in the immune response.⁸² It is noteworthy that incorporating several changes in the therapeutic regimens involving Tregs can be an efficient approach to treat autoimmune disorders like AIH.⁸³ In absence of hepatic infection, detection of elevated level of liver enzyme is considered to be presumptive diagnosis of autoimmune hepatitis.⁸⁴ Divided opinion has been given by researchers to support the role of Tregs in autoimmune hepatitis. A decrease in population of Treg cells in the peripheral blood in few cases of autoimmune hepatitis has been reported,⁸⁵ on the contrary no such difference in Tregs cells in several other cases has raised controversy in this regard.⁸⁶

The stability of Treg cells at the site of hepatic inflammation is crucial, however research data suggested that due to low concentration of IL-2 in hepatic cell, human liver tissues may not be the ideal site for Tregs survival and expansion.⁸⁷ Based on this data, low dose IL-2 was administered in two patients with refractory AIH and an increase in the frequency of Treg cells in the peripheral circulation was observed, which indirectly support the ameliorating role of Tregs for AIH.⁸⁸

Enrichment of effector T cells (Teffs) within the liver tissues of untreated AIH-1 patients with a constant Treg/Teff ratio but without any change of Tregs in the circulation, is not an uncommon finding. The disproportional decrease of intrahepatic Tregs during therapy might explain high rates of relapse after discontinuation of immunosuppression.⁸⁹ Under the guidance of National Institute of Health, a phase-1 clinical trials ((NCT02704338) has been registered to evaluate the efficacy of autologous Tregs from peripheral blood of autoimmune hepatitis patients using ex vivo expansion procedure so that each patient should receive 5×10^6 cells /kg body weight. Results of this study have not been posted yet as the project is continuing.⁹⁰

Systemic lupus erythematosus (SLE)

SLE is an autoimmune, multisystemic disease that may affect virtually any organ due to induction of autoantibodies and the deposition of immune-complexes in tissues that leads to the activation of complement, accumulation of neutrophils and monocytes, and self-reactive lymphocyte.⁹¹ In mice model, regulatory T cell numbers have been reported to decrease in lupus-prone mice as the age and the disease progresses.⁹² Clinical application of Tregs in human patients suffering with SLE have shown variable efficacy and even conflicting reports.⁹³ Such discrepancies in findings may be due to difference in isolation and purification strategies applied for enrichment of Foxp3⁺ CD25⁺ Tregs before being infused to the patients.⁹⁴

Most of the earlier studies on SLE have shown decreased proportion of either CD4 + CD25hi T cells or CD4 + CD25 + FoxP3 + Treg cells once the disease flares.⁹⁵ Deuterium tracking of infused Treg cells in SLE patient with active skin disease revealed transient presence of cells in peripheral blood, accompanied by increased percentages of highly activated Treg cells in diseased skin. Flow cytometric analysis and whole transcriptome RNA sequencing revealed that Treg cell accumulation in skin was associated with a marked attenuation of the interferon-y pathway and a reciprocal augmentation of the interleukin-17 (IL-17) pathway. This phenomenon was more pronounced in skin relative to peripheral blood.⁶⁷ The homeostasis of CD4⁺ T cell subsets is regulated by IL2, and reduced production of IL-2 by T cells is observed in individuals with SLE. It has been reported that the treatment with low-dose of rIL-2 selectively modulated the abundance of T Regs, follicular helper T (T_{FH}) cells and IL-17-producing helper T (T_H17) cells, but not T_H1 or T_H2 cells.⁹⁶ The contributing role of Tregs in SLE has been reported from 15 patients in which the proportion of CD45RA-FoxP3hi activated Treg cells was decreased whereas the proportion of CD45RA+FoxP3lo resting Treg cells was increased.⁹⁷ In a unique case study, single cutaneous lupus adult patient was included in phase I non-randomized clinical trial for evaluating the effects of a single dose of 1×10^8 ex vivo expanded, FACS-sorted, and deuterium-labeled autologous Tregs. The number of circulating labeled cells dropped 4 weeks post-injection, but a marked increase in Tregs and IL-17 production was recorded in skin biopsy.⁶⁷

Rheumatoid arthritis (RA)

RA is a systemic and heterogeneous autoimmune disease characterized with clinical manifestation of symmetrical polyarthritis. It is a chronic inflammatory joint disease, which can cause cartilage and bone damage as well as disability condition in patients.98 The pathogenesis of RA is complex, and the dysfunction of Tregs is one of the proposed mechanisms underlying the breakdown of selftolerance leading to the progression of rheumatoid arthritis. For patients with a poor response to traditional diseasemodifying anti-rheumatic drugs, the clinicians adapted Treg therapy with variable success.⁹⁹ Clinical findings indicated an increase in number of Treg cells in the peripheral blood of several individuals suffering with RA.¹⁰⁰ On the contrary, either an unchanged Treg population¹⁰¹ and even low count of Tregs in several other patients manifesting frank clinical cases has been reported by different research groups, which is a subject of clinical concern.¹⁰² Variability in the interpretation of data of the above discussed cases mostly depends upon the identification of Tregs based upon the expression of FoxP3 and CD25 marker, which are likely to be mediated by inflammation and T cell activation. Invariably, Tregs of RA patients don't secrete copious amount of IL-2 and IFNy in their peripheral circulation as compared to healthy individuals, suggestive of FoxP3 expression to be a hallmark of Tregs.¹⁰³ Recovery in RA patients due to anti-TNF antibody therapy was attributed to the generation of novel Treg subsets expressing FoxP3 marker but not CD-62 ligand.¹⁰⁴ Whether TNF-a plays a direct role to modulate Treg function is still under debate.¹⁰⁵ This indicates that the use of a more stringent method to define Treg cells will reveal decreased number of Treg cells in peripheral blood and increased number in synovial fluid, but whether Treg cells in the synovial fluid function normally is yet to be confirmed.¹⁰⁶ Transfer of Treg cells can slow down the disease process, confirming that these cells have the potential to treat autoimmune diseases.¹⁰⁷ Adaptive transfer of Tregs in mice model using collagen-induced arthritis (CIA) showed inhibitory effects on arthritis, which significantly prevents the development of CIA.¹⁰⁸ Importantly, when arthritis is inhibited in these models, not only T and B cells are inhibited by Treg cells but osteoclast-mediated bone destruction is also directly diminished, preventing joint injury.¹⁰⁹ Adaptive Treg cell therapy supplements the functional activity of existing residential Tregs impaired by the highly inflamed synovial milieu.¹¹⁰

Systemic sclerosis (SSc)

SSc is an autoimmune inflammatory condition that results in potentially widespread fibrosis and vascular abnormalities, which can affect the skin, lungs, gastrointestinal tract, heart, and kidneys. The skin becomes thickened and hard (sclerotic). It is characterized by clinical features that resemble GVH disease. The classification of systemic sclerosis is based on skin involvement. Limited cutaneous systemic sclerosis (LcSSc), formerly known as the CREST syndrome, is associated with skin thickening distal to the elbows, distal to the knees, and/or face without trunk involvement.¹¹¹

The majority of the studies reported decreased frequencies and/or impaired function of circulating Tregs in SSc patients compared to controls.¹¹² Reduced frequency of Tregs, together with that of total TGFβ1 and IL-10, may be responsible for the loss of tolerance observed in SSc.¹¹³ Contrary to this, other studies have reported decreased frequency of circulating Tregs in peripheral blood but not reaching statistical significance.^{114,115} Success of adaptive therapy of Tregs in SSc is not so promising, however uncontrolled studies in SSc have demonstrated that hematopoietic stem cell therapy (HSCT) can halt disease progression and reverse skin fibrosis, and might even reduce mortality.¹¹⁶ In a clinical trial, 26 SSc patients were selected for CD34 ⁺ autologous hematopoietic stem cell transplantation to compare its effectiveness and safety to intravenous cyclophosphamide, the only treatment option for progressive SSc. Sustained improvement of skin thickening and stabilization of organ function for up to 7 years after transplantation was recorded in this clinical trial.¹¹⁷

Primary Sjogren's syndrome (pSS)

pSS is a systemic autoimmune disease characterized by lymphocytic infiltration of the salivary and lacrimal glands. In addition to the exocrine glands, many other organs can be affected by the disease as well. Hyperactivity of B cells is thought to play a central role in the pathogenesis of pSS. Available evidence strongly indicates that this B cell hyperactivity is mediated by T cells. Loss of self-tolerance and secretion of many pro-inflammatory cytokines including IFN-y, IL-17 and IL-21 associated with local inflammation in pSS, are mediated by T-cell activity.¹¹⁸ However, circulating Treg cells in pSS is controversial since either diminished, normal or increased proportions are reported by different workers.¹⁰⁵ Report indicating localization of Treg cells in target organ, i.e. salivary gland is also conflicting because of either the absence of Treg cells¹¹⁹ or presence of infiltrating FoxP3 +CD4+T cells positively correlated to the biopsy scores in lymphocytic sialadenitis but without any change in circulating Tregs in periphery.¹²⁰

Psoriasis

Tregs have also been employed as crucial pla yers in psoriasis, which is a systemic inflammatory disorder with significant involvement of genetic factors induced by environmental influence. Disease pathogenicity is mainly driven by type 1 and Th17 cytokine-producing cells, which are tightly controlled by Tregs in normal people. Tregs play critical roles in maintaining immune homeostasis via blocking inflammatory cells, which in turn prevent autoimmune disorders like psoriasis and other skin-related autoimmune diseases. It has been postulated that Treg-mediated suppression is disrupted in psoriasis, changing the Th17/Treg ratio. While Treg dysfunction is linked to disease worsening in patients with psoriasis, it is unclear how they have been biologically controlled.¹²¹ Recent research suggests that Tregs are essential in psoriasis treatment. While the significance of Treg concentration is unknown, attenuated Treg activity and a disrupted Th17/Treg equilibrium have been linked to psoriasis pathogenesis and exacerbation.¹²¹⁻¹²⁵

Other auto immune diseases

Recently, Huang et al. have reported the critical roles of Tregs in the initiation, treatment, and recovery of uveitis, which is a recurrent autoimmune disease characterized by relapsing and remitting ocular attacks that necessitates corticosteroids and systemic immunosuppression to avoid serious vision loss. Both Th17 (pro-inflammatory cells) and Tregs mediate uveitis, and it has been suggested that modulating Tregs could be an effective potential therapeutic strategy against uveitis.¹²⁶

Further, the protective role of Tregs has been described in Myasthenia gravis (MG), which is a T cell-dependent, B-lymphocyte-mediated autoimmune disease caused by the production of antibodies against the nicotinic acetylcholine receptors and other post-synaptic components at the neuromuscular junction. Adoptive transfer of Tregs has been found to suppress the autoimmune response in the rodent model and the protective role of these cells has been suggested in MG. However, any disturbances in the ratio of T-helper (Th) cells and Tregs might be linked with the poor prognosis of the autoimmune diseases, such as MG.¹²⁷

Herrnstadt et al. postulated the potential roles of Tregs in providing protection against any renal tissue injury, which has been linked with pathogenic driving force of Th cells such as Th1 and Th17, also known as T effector cells, in glomerulonephritis (GN).¹²⁸ Tregs overcome the pathogenic behavior of vital innate immunity components in inflamed kidney cells or tissues. Hence, these findings can be utilized to further elucidate the immunosuppressive function of Treg cell population in renal injuries and kidney inflammation.¹²⁹

Several clinical trials for addressing autoimmune disorders have used polyclonal Tregs and antigen-specific Tregs designed with TCR or chimeric antigen receptor (CAR) constructs that could dramatically boost their performance in the host. Furthermore, it has been proposed that antigen-specific Tregs may be injected into patients with autoimmune skin diseases to restore Treg content and provide resistance against the exacerbated immune response.¹²

Briefly, the present therapies adapted by the clinicians to control the autoimmune disease in population are primarily aimed to restore immune tolerance in the patients. The drugs used to tame down autoimmune diseases are targeted to encourage in vivo Treg induction and expansion. Drugs or biologicals such as mTOR inhibitor rapamycin and cytokines IL-10, tiny dose of IL-2, TNF receptor 2 agonist, or tyrosine kinase 3 ligand are used universally to manage the autoimmune disease under control. Besides these, further explorative researches into Treg persistence and the optimal requirements for Treg survival in peripheral blood are expected in the coming years to properly promote the clinical use of Tregs in immunotherapeutics.

Challenges in clinical applications

Application of Tregs in clinics must follow several stringent guidelines imposed by Food and Drug Administration (FDA). It is mandatory that the cell therapy products should pass the sterility, identity, purity, and potency standards before administration. Demonstration of purity and identity is most grim part of the test system. Foxp3 marker expression is a universally accepted marker for Treg identity. However, ex vivo Treg expansion may not yield 100% Foxp3+ cells, but it should adhere the acceptable limit of contaminating cells, which must satisfy the qualitative as well as quantitative bench marks. Contaminating cells cause allogenic reaction in the recipient; hence, it should be at extremely low concentration. Currently, investigators are looking forward to develop disease-specific models to establish the potency of expanded enriched Treg products, as one product is not suitable for all diseases.¹³⁰ However, some other limitations associated with the use of Treg cells in clinical conditions include the stability of the Treg population in peripheral blood, and, therefore, also limit their effects. Furthermore, Treg cryopreservation is required for its robust use among the patients in clinical settings, which may damage Tregs by reducing its durability, triggering unusual cytokine secretion, and interfering with the expressions of several surface proteins necessary for Treg functioning and processing. Hence, optimum Treg culture techniques, cryopreservation extension, and storage for medicinal use must also be explored and established.131

Conclusion and future perspectives

Recent advancements in technology have enabled scientists to identify and classify Tregs and their immunosuppressive mechanisms, allowing a better comprehension of the relationship among Treg plasticity, which is needed for optimal Treg immune suppression and Treg destabilization or vulnerability from various factors that leads to autoimmune diseases. The plasticity and heterogeneity in the mice- and human-derived Treg population have given new information in Treg biology. Due to the conflicting findings in different studies, the contribution of Tregs to different autoimmune disease models is yet unclear. However, it has been confirmed that Tregs express specific transcription factors, such as Foxp3 protein, which is considered to be a master regulator. Treg isolation is not easy due to the lack of a specific marker for experimental purposes. Human Tregs expressing a low, intermediate, or high CD25 molecule label 1 are difficult to isolate as these boundaries are thin. Using human Tregs as therapeutic agents is a new area of research in clinical application. Clinical findings have provided some hope to use Tregs as an adjunct with approved drugs to support the treatment regimen for reducing the reoccurrence of autoimmune diseases and can add few years of quality life in an individual.

Further, the results of studies on the patients with inflammation and autoimmune diseases are still conflicting and misleading due to a lack of reliable Treg-specific markers, which in turn play a pivotal role in providing immunosuppressive functionality to the Tregs. It is the ultimate need of the time to elucidate the specific markers on the Tregs for designing efficient therapeutic strategies. A vivid elucidation of the Treg-mediated immune suppression and the identification of changes in the immune response of patients with functional Treg deficiency is necessary for efficient therapeutic regimes to control autoimmune diseases. Furthermore, in-depth and specific studies are required to establish the roles of Tregs under specific inflammatory responses, which will pave a path toward a personalized therapeutic strategy e2035117-12 👄 T. K. GOSWAMI ET AL.

to control specific clinical manifestations. Various advanced techniques, such as single-cell RNA sequencing, can discern Treg subsets and provide knowledge on TCR repertoires. Specific Tregs against specific antigens, as well as their hetero-geneity in stable and diseased states, will open up several possibilities, including the identification of TCR-transduced cells for medicinal purposes. In addition, using genetically engineered Tregs and chimeric antigen receptor-Tregs is promising in preclinical autoimmune disorder models, and may have medical applications, particularly when extracellular receptors are important. Finally, it can be well stated that identifying and deciphering immunosuppressive pathways and their interlinked markers will lead to the development of efficient and reliable therapeutic regimes to treat autoimmune diseases.

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Author contributions

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References

- 1. Arellano B, Graber DJ, Sentman CL. Regulatory T cell-based therapies for autoimmunity. Discov Med. 2016;22:73–80.
- 2. Matzinger P. The danger model: a renewed sense of self. Science (80-). 2002;296:301–05. doi:10.1126/science.1071059.
- Miyara M, Yoshioka Y, Kitoh A, Shima T, Wing K, Niwa A, Parizot C, Taflin C, Heike T, Valeyre D, et al. Functional delineation and differentiation dynamics of human CD4+ T cells expressing the FoxP3 transcription factor. Immunity. 2009;30:899–911. doi:10.1016/j.immuni.2009.03.019.

- Gliwiński M, Iwaszkiewicz-Grześ D, Trzonkowski P. Cell-based therapies with T regulatory cells. BioDrugs. 2017;31:335–47. doi:10.1007/s40259-017-0228-3.
- Mikami N, Kawakami R, Sakaguchi S. New Treg cell-based therapies of autoimmune diseases: towards antigen-specific immune suppression. Curr Opin Immunol. 2020;67:36–41. doi:10.1016/j. coi.2020.07.004.
- Palomares O, Yaman G, Azkur AK, Akkoc T, Akdis M, Akdis CA. Role of Treg in immune regulation of allergic diseases. Eur J Immunol. 2010;40:1232–40. doi:10.1002/eji.200940045.
- Wang S, Zou X, Zhang Y, Wang X, Yang W, Li Y. The generation and regulation of tissue-resident Tregs and their role in autoimmune diseases. J Immunol Res. 2020;2020:1–13. doi:10.1155/2020/8815280.
- Rocamora-Reverte L, Melzer FL, Würzner R, and Weinberger B. The complex role of regulatory T cells in immunity and aging. Front Immunol 11. 2021;616949. doi:10.3389/fimmu.2020.616949
- Sampani E, Stangou M, Daikidou DV, Nikolaidou V, Asouchidou D, Dimitriadis C, Lioulios G, Xochelli A, Fylaktou A, Papagianni A. Influence of end stage renal disease on CD28 expression and T-cell immunity. Nephrology. 2021;26:185–96. doi:10.1111/nep.13784.
- Sampani E, Daikidou DV, Lioulios G, Xochelli A, Mitsoglou Z, Nikolaidou V, Dimitriadis C, Fylaktou A, Papagianni A, Stangou M. CD28null and regulatory T cells are substantially disrupted in patients with end-stage renal disease due to diabetes mellitus. Int J Mol Sci. 2021;22:1–13. doi:10.3390/ijms22062975.
- Trzonkowski P, Bieniaszewska M, Juścińska J, Dobyszuk A, Krzystyniak A, Marek N, Myśliwska J, Hellmann A. First-in-man clinical results of the treatment of patients with graft versus host disease with human ex vivo expanded CD4+CD25+CD127-T regulatory cells. Clin Immunol. 2009;133:22–26. doi:10.1016/j. clim.2009.06.001.
- Mukhatayev Z, Ostapchuk YO, Fang D, and Le Poole IC. Engineered antigen-specific regulatory T cells for autoimmune skin conditions. Autoimmun Rev 20 3 . 2021;102761. doi:10.1016/j.autrev.2021.102761
- Singer A, Adoro S, Park JH. Lineage fate and intense debate: myths, models and mechanisms of CD4- versus CD8-lineage choice. Nat Rev Immunol. 2008;8:788–801. doi:10.1038/nri2416.
- Josefowicz SZ, Lu LF, Rudensky AY. Regulatory T cells: mechanisms of differentiation and function. Annu Rev Immunol. 2012;30:531–64. doi:10.1146/annurev.immunol.25.022106.141623.
- Nazzal D, Gradolatto A, Truffault F, Bismuth J, and Berrih-Aknin S. Human thymus medullary epithelial cells promote regulatory T-cell generation by stimulating interleukin-2 production via ICOS ligand. Cell Death Dis 5 9 . 2014;e1420. doi:10.1038/cddis.2014.377
- Sakaguchi S, Sakaguchi N, Asano M, Itoh M, and Toda M. Immunologic self-tolerance maintained by activated T cells expressing IL-2 receptor alpha-chains (CD25). Breakdown of a single mechanism of self-tolerance causes various autoimmune diseases. J Immunol. 1995;155 3 :1151–64. Accessed date 1 August 1995. http://www.ncbi.nlm.nih.gov/pubmed/7636184
- Burchill MA, Yang J, Vogtenhuber C, Blazar BR, Farrar MA. IL-2 receptor β-Dependent STAT5 activation is required for the development of Foxp3 + regulatory T cells. J Immunol. 2007;178:280–90. doi:10.4049/jimmunol.178.1.280.
- Shevach EM, Thornton AM. tTregs, pTregs, and iTregs: similarities and differences. Immunol Rev. 2014;259:88–102. doi:10.1111/ imr.12160.
- Adeegbe DO, Nishikawa H. Natural and induced T regulatory cells in cancer. Front Immunol. 2013;4:4. doi:10.3389/fimmu.2013.00004.
- Tang Q, Adams JY, Tooley AJ, Bi M, Fife BT, Serra P, Santamaria P, Locksley RM, Krummel MF, Bluestone JA. Visualizing regulatory T cell control of autoimmune responses in nonobese diabetic mice. Nat Immunol. 2006;7:83–92. doi:10.1038/ni1289.
- Allos H, Al Dulaijan BS, Choi J, Azzi J. Regulatory T cells for more targeted immunosuppressive therapies. Clin Lab Med. 2019;39:1–13. doi:10.1016/j.cll.2018.11.001.

- 22. Bluestone JA, Abbas AK. Natural versus adaptive regulatory T cells. Nat Rev Immunol. 2003;3:253–57. doi:10.1038/nri1032.
- 23. Lui PPW, Cho I, Ali N. Tissue regulatory T cells. Immunology. 2020;161:4–17. doi:10.1111/imm.13208.
- Panduro M, Benoist C, Mathis D. Tissue Tregs. Annu Rev Immunol. 2016;34(1):609–33. doi:10.1146/annurev-immunol -032712-095948.
- 25. Weiss JM, Bilate AM, Gobert M, Ding Y, de Lafaille MAC, Parkhurst CN, Xiong H, Dolpady J, Frey AB, Ruocco MG, et al. Neuropilin 1 is expressed on thymus-derived natural regulatory T cells, but not mucosagenerated induced Foxp3+ T reg cells. J Exp Med. 2012;209:1723–42. doi:10.1084/jem.20120914.
- Atarashi K, Tanoue T, Shima T, Imaoka A, Kuwahara T, Momose Y, Cheng G, Yamasaki S, Saito T, Ohba Y, et al. Induction of colonic regulatory T cells by indigenous Clostridium species. Science (80-). 2011;331:337–41. doi:10.1126/ science.1198469.
- Scharschmidt TC, Vasquez KS, Truong HA, Gearty SV, Pauli ML, Nosbaum A, Gratz IK, Otto M, Moon JJ, Liese J, et al. A wave of regulatory T cells into neonatal skin mediates tolerance to commensal microbes. Immunity. 2015;43:1011–21. doi:10.1016/j. immuni.2015.10.016.
- Ohue Y, Nishikawa H. Regulatory T (Treg) cells in cancer: can Treg cells be a new therapeutic target? Cancer Sci. 2019;110:2080–89. doi:10.1111/cas.14069.
- Toker A, Engelbert D, Garg G, Polansky JK, Floess S, Miyao T, Baron U, Düber S, Geffers R, Giehr P, et al. Active demethylation of the Foxp3 locus leads to the generation of stable regulatory T cells within the thymus. J Immunol. 2013;190:3180–88. doi:10.4049/ jimmunol.1203473.
- Darrasse J, Mardon G, Salomon BL, Catala MKD, Klatzmann D. Ontogeny of CD4+CD25+ regulatory/suppressor T cells in human fetuses. Blood. 2005;105:4715–21. doi:10.1182/blood-2004-10-4051.
- Liu W, Putnam AL, Xu-yu Z, Szot GL, Lee MR, Zhu S, Gottlieb PA, Kapranov P, Gingeras TR, Barbara BF, et al. CD127 expression inversely correlates with FoxP3 and suppressive function of human CD4+ T reg cells. J Exp Med. 2006;203:1701–11. doi:10.1084/ jem.20060772.
- Onizuka S, Tawara I, Shimizu J, Sakaguchi S, Fujita T, Nakayama E. Tumor rejection by in vivo administration of anti-CD25 (interleukin-2 receptor α) monoclonal antibody. Cancer Res. 1999;59:3128–33.
- Vignali DAA, Collison LW, Workman CJ. How regulatory T cells work. Nat Rev Immunol. 2008;8:523–32. doi:10.1038/nri2343.
- Sojka DK, Huang YH, Fowell DJ. Mechanisms of regulatory T-cell suppression - A diverse arsenal for a moving target. Immunology. 2008;124:13–22. doi:10.1111/j.1365-2567.2008.02813.x.
- 35. Romano M, Fanelli G, Albany CJ, Giganti G, Lombardi G. Past, present, and future of regulatory T cell therapy in transplantation and autoimmunity. Front Immunol. 2019;10:10. doi:10.3389/fimmu.2019.00010.
- Wing K, Sakaguchi S. Regulatory T cells exert checks and balances on self tolerance and autoimmunity. Nat Immunol. 2010;11:7–13. doi:10.1038/ni.1818.
- 37. Feuerer M, Hill JA, Kretschmer K, Von Boehmer H, Mathis D, Benoist C. Genomic definition of multiple ex vivo regulatory T cell subphenotypes. Proc Natl Acad Sci U S A. 2010;107:5919–24. doi:10.1073/pnas.1002006107.
- Omenetti S, Pizarro TT, Bancu I, Lauzurica-Valdemoros R, Borràs FE. The Treg/Th17 axis: a dynamic balance regulated by the gut microbiome. Front Immunol. 2015;6:6. doi:10.3389/ fimmu.2015.00006.
- Liu Y, Tran DQ, Lindsey JW, Rhoads JM. The association of gut microbiota and treg dysfunction in autoimmune diseases. Adv Exp Med Biol. 2021;1278:191–203.
- Gotot J, Gottschalk C, Leopold S, Knolle PA, Yagita H, Kurts C, Ludwig-Portugall I. Regulatory T cells use programmed death 1 ligands to directly suppress autoreactive B cells in vivo. Proc Natl Acad Sci U S A. 2012;109:10468–73. doi:10.1073/pnas.1201131109.

- Iikuni N, Lourenço EV, Hahn BH, La Cava A. Cutting edge: regulatory T cells directly suppress B cells in systemic lupus erythematosus. J Immunol. 2009;183:1518–22. doi:10.4049/ jimmunol.0901163.
- Richards H, Williams A, Jones E, Hindley J, Godkin A, Simon AK, Gallimore A. Novel role of regulatory T cells in limiting early neutrophil responses in skin. Immunology. 2010;131:583–92. doi:10.1111/j.1365-2567.2010.03333.x.
- Lewkowicz N, Klink M, Mycko MP, Lewkowicz P. Neutrophil -CD4+CD25+ T regulatory cell interactions: a possible new mechanism of infectious tolerance. Immunobiology. 2013;218:455–64. doi:10.1016/j.imbio.2012.05.029.
- 44. Tiemessen MM, Jagger AL, Evans HG, Van Herwijnen MJC, John S, Taams LS. CD4+CD25+Foxp3+ regulatory T cells induce alternative activation of human monocytes/macrophages. Proc Natl Acad Sci U S A. 2007;104:19446–51. doi:10.1073/pnas.0706832104.
- 45. Romano M, Fanelli G, Tan N, Nova-Lamperti E, McGregor R, Lechler RI, Lombardi G, Scottà C. Expanded regulatory T cells induce alternatively activated monocytes with a reduced capacity to expand T helper-17 cells. Front Immunol. 2018;9. doi:10.3389/ fimmu.2018.01625.
- 46. Kullberg MC, Hay V, Cheever AW, Mamura M, Sher A, Letterio JJ, Shevach EM, Piccirillo CA. TGF-β1 production by CD4+CD25+ regulatory T cells is not essential for suppression of intestinal inflammation. Eur J Immunol. 2005;35:2886–95. doi:10.1002/ eji.200526106.
- 47. Maj T, Wang W, Crespo J, Zhang H, Wang W, Wei S, Zhao L, Vatan L, Shao I, Szeliga W, et al. Oxidative stress controls regulatory T cell apoptosis and suppressor activity and PD-L1-blockade resistance in tumor. Nat Immunol. 2017;18:1332–41. doi:10.1038/ni.3868.
- Tang Q, Bluestone JA. The Foxp3+ regulatory T cell: a jack of all trades, master of regulation. Nat Immunol. 2008;9:239–44. doi:10.1038/ni1572.
- Ziegler SF. FOXP3: of mice and men. Annu Rev Immunol. 2006;24:209–26. doi:10.1146/annurev.immunol.24.021605.090547.
- 50. Brunkow ME, Jeffery EW, Hjerrild KA, Paeper B, Clark LB, Yasayko SA, Wilkinson JE, Galas D, Ziegler SF, Ramsdell F. Disruption of a new forkhead/winged-helix protein, scurfin, results in the fatal lymphoproliferative disorder of the scurfy mouse. Nat Genet. 2001;27:68–73. doi:10.1038/83784.
- 51. Wildin RS, Ramsdell F, Peake J, Faravelli F, Casanova J-L-R, Buist N, Levy-Lahad E, Mazzella M, Goulet O, Perroni L, et al. The immune dysregulation, polyendocrinopathy, enteropathy, X-linked syndrome (IPEX) is caused by mutations of FOXP3. Nat Genet. 2001;27:20–21. doi:10.1038/83713.
- Shimizu J, Yamazaki S, Takahashi T, and Ishida YSS. Stimulation of CD25+CD4+ regulatory T cells through GITR breaks immunological self-tolerance. Nat Immunol. 2002;3 2 135–142. doi:10.1038/ni759
- Longhi MS, Ma Y, Mieli-Vergani G, Vergani D. Regulatory T cells in autoimmune hepatitis. J Hepatol. 2012;57:932–33. doi:10.1016/j. jhep.2012.05.022.
- Aghabi YO, Yasin A, Kennedy JI, Davies SP, Butler AE, and Stamataki Z. Targeting enclysis in liver autoimmunity, transplantation, viral infection and cancer. Front Immunol. 2021;12 662134. doi:10.3389/fimmu.2021.662134
- 55. Brunstein CG, Miller JS, Cao Q, McKenna DH, Hippen KL, Curtsinger J, DeFor T, Levine BL, June CH, Rubinstein P, et al. Infusion of ex vivo expanded T regulatory cells in adults transplanted with umbilical cord blood: safety profile and detection kinetics. Blood. 2011;117:1061–70. doi:10.1182/blood-2010-07-293795.
- 56. Dijke IE, Hoeppli RE, Ellis T, Pearcey J, Huang Q, McMurchy AN, Boer K, Peeters AMA, Aubert G, Larsen I, et al. Discarded human thymus is a novel source of stable and long-lived therapeutic regulatory T cells. Am J Transplant. 2016;16:58–71. doi:10.1111/ajt.13456.
- 57. Safinia N, Vaikunthanathan T, Fraser H, Thirkell S, Lowe K, Blackmore L, Whitehouse G, Martinez-Llordella M, Jassem W, Sanchez-Fueyo A, et al. Successful expansion of functional and stable regulatory T cells for immunotherapy in liver transplantation. Oncotarget. 2016;7:7563–77. doi:10.18632/oncotarget.6927.

- Marek-Trzonkowska N, Myśliwiec M, Dobyszuk A, Grabowska M, Derkowska I, Juścińska J, Owczuk R, Szadkowska A, Witkowski P, Młynarski W, et al. Therapy of type 1 diabetes with CD4 +CD25highCD127-regulatory T cells prolongs survival of pancreatic islets - Results of one year follow-up. Clin Immunol. 2014;153:23–30. doi:10.1016/j.clim.2014.03.016.
- 59. Fraser H, Safinia N, Grageda N, Thirkell S, Lowe K, Fry LJ, Scottá C, Hope A, Fisher C, Hilton R, et al. A rapamycin-based GMP-compatible process for the isolation and expansion of regulatory T cells for clinical trials. Mol Ther - Methods Clin Dev. 2018;8:198–209. doi:10.1016/j.omtm.2018.01.006.
- Janssens I, and Cools N. Regulating the regulators: is introduction of an antigen-specific approach in regulatory T cells the next step to treat autoimmunity? Cell Immunol. 2020;358 104236. doi:10.1016/j.cellimm.2020.104236
- Wu D, Wong MQ, Vent-Schmidt J, Boardman DA, Steiner TS, and Levings MK. A method for expansion and retroviral transduction of mouse regulatory T cells. J Immunol Methods. 2021;488 112931. doi:10.1016/j.jim.2020.112931
- 62. Roth-Walter F, Adcock IM, Benito-Villalvilla C, Bianchini R, Bjermer L, Boyman O, Caramori G, Cari L, Fan Chung K, Diamant Z, et al. Immune modulation via T regulatory cell enhancement: disease-modifying therapies for autoimmunity and their potential for chronic allergic and inflammatory diseases—An EAACI position paper of the Task Force on Immunopharmacology (TIPCO). Allergy Eur J Allergy Clin Immunol. 2021;76:90–113. doi:10.1111/all.14478.
- Eggenhuizen PJ, Ng BH, Ooi JD. Treg enhancing therapies to treat autoimmune diseases. Int J Mol Sci. 2020;21:1–18. doi:10.3390/ ijms21197015.
- 64. Desreumaux P, Foussat A, Allez M, Beaugerie L, Hébuterne X, Bouhnik Y, Nachury M, Brun V, Bastian H, Belmonte N, et al. Safety and efficacy of antigen-specific regulatory T-cell therapy for patients with refractory Crohn's disease. Gastroenterology. 2012:143. doi:10.1186/1471-230X-12-143
- 65. Goldberg R, Scotta C, Cooper D, Nissim-Eliraz E, Nir E, Tasker S, Irving PM, Sanderson J, Lavender P, Ibrahim F, et al. Correction of defective T-Regulatory cells from patients with crohn's disease by ex vivo ligation of retinoic acid receptor-α. Gastroenterology. 2019;156:1775–87. doi:10.1053/j.gastro.2019.01.025.
- 66. Bluestone JA, Buckner JH, Fitch M, Gitelman SE, Gupta S, Hellerstein MK, Herold KC, Lares A, Lee MR, Li K. Type 1 diabetes immunotherapy using polyclonal Tregs. Sci Transl Med. 2015 Nov 25;7:315ra189. doi:10.1126/scitranslmed.aad4134.
- 67. Dall'Era M, Pauli ML, Remedios K, Taravati K, Sandova PM, Putnam AL, Lares A, Haemel A, Tang Q, Hellerstein M, et al. Adoptive Treg cell therapy in a patient with systemic lupus erythematosus. Arthritis Rheumatol. 2019;71:431–40. doi:10.1002/ art.40737.
- Terry LV, Oo YH. The next frontier of regulatory T cells: promising immunotherapy for autoimmune diseases and organ transplantations. Front Immunol. 2020;11:11. doi:10.3389/fimmu.2020.00011.
- Groux H, O'Garra A, Bigler M, Rouleau M, Antonenko S, De Vries JE, Roncarolo MG. A CD4+ T-cell subset inhibits antigen-specific T-cell responses and prevents colitis. Nature. 1997;389:737-42. doi:10.1038/39614.
- 70. Mottet C, Uhlig HH, Powrie F. Cutting edge: cure of colitis by CD4
 + CD25 + Regulatory T Cells. J Immunol. 2003;170:3939–43. doi:10.4049/jimmunol.170.8.3939.
- Goswami TK, Joardar SN, Ram GC, Banerjee R, Singh DK. Association of mycobacterium paratuberculosis in Crohn's disease and Johne's disease: a possible zoonotic threat. Curr Sci. 2000;79:1076–81.
- Maul J, Loddenkemper C, Mundt P, Berg E, Giese T, Stallmach A, Zeitz M, Duchmann R. Peripheral and intestinal regulatory CD4 +CD25high T cells in inflammatory bowel disease. Gastroenterology. 2005;128:1868–78. doi:10.1053/j.gastro.2005. 03.043.

- Foussat A, Cottrez F, Brun V, Fournier N, Breittmayer J-P, Groux H. A comparative study between T regulatory Type 1 and CD4 + CD25 + T cells in the control of inflammation. J Immunol. 2003;171:5018–26. doi:10.4049/jimmunol.171.10.5018.
- 74. Canavan JB, Scottà C, Vossenkämper A, Goldberg R, Elder MJ, Shoval I, Marks E, Stolarczyk E, Lo JW, Powell N, et al. Developing in vitro expanded CD45RA+ regulatory T cells as an adoptive cell therapy for Crohn's disease. Gut. 2016;65:584–94. doi:10.1136/ gutjnl-2014-306919.
- Brimnes J, Allez M, Dotan I, Shao L, Nakazawa A, Mayer L. Defects in CD8 + regulatory T cells in the lamina propria of patients with inflammatory bowel disease. J Immunol. 2005;174:5814–22. doi:10.4049/jimmunol.174.9.5814.
- Marwaha AK, Crome SQ, Panagiotopoulos C, Berg KB, Qin H, Ouyang Q, Xu L, Priatel JJ, Levings MK, Tan R. Cutting edge: increased IL-17-secreting T cells in children with new-onset type 1 diabetes. J Immunol. 2010;185:3814–18. doi:10.4049/jimmunol.1001860.
- Baeten P, Van Zeebroeck L, Kleinewietfeld M, Hellings N, Broux B. Improving the efficacy of regulatory T cell therapy. Clin Rev Allergy Immunol. 2021. doi:10.1007/s12016-021-08866-1.
- Sugiyama H, Matsue H, Nagasaka A, Nakamura Y, Tsukamoto K, Shibagaki N, Kawamura T, Kitamura R, Ando N, Shimada S. CD4 +CD25high regulatory T cells are markedly decreased in blood of patients with pemphigus vulgaris. Dermatology. 2007;214:210–20. doi:10.1159/000099585.
- 79. Yokoyama T, Matsuda S, Takae Y, Wada N, Nishikawa T, Amagai M, Koyasu S. Antigen-independent development of Foxp31 regulatory T cells suppressing autoantibody production in experimental pemphigus vulgaris. Int Immunol. 2011;23:365-73. doi:10.1093/intimm/dxr020.
- Schmidt T, Willenborg S, Hünig T, Deeg CA, Sonderstrup G, Hertl M, Eming R. Induction of T regulatory cells by the superagonistic anti-CD28 antibody D665 leads to decreased pathogenic IgG autoantibodies against desmoglein 3 in a HLA-transgenic mouse model of pemphigus vulgaris. Exp Dermatol. 2016;25:293–98. doi:10.1111/exd.12919.
- Izumi K, Bieber K, Ludwig RJ. Current clinical trials in pemphigus and pemphigoid. Front Immunol. 2019;10:10.
- Buitrago-Molina LE, Pietrek J, Noyan F, Schlue J, Manns MP, Wedemeyer H, Hardtke-Wolenski M, and Jaeckel E. Treg-specific IL-2 therapy can reestablish intrahepatic immune regulation in autoimmune hepatitis. J Autoimmun. 2021;117 102591. doi:10.1016/j.jaut.2020.102591
- Longhi MS, Mieli-Vergani G, and Vergani D. Regulatory T cells in autoimmune hepatitis: un updated overview. J Autoimmun. 2021;119 102619. doi:10.1016/j.jaut.2021.102619
- Sebode M, Hartl J, Vergani D, Lohse AW. Autoimmune hepatitis: from current knowledge and clinical practice to future research agenda. Liver Int. 2018;38:15–22. doi:10.1111/liv.13458.
- Longhi MS, Ma Y, Bogdanos DP, Cheeseman P, Mieli-Vergani G, Vergani D. Impairment of CD4+CD25+ regulatory T-cells in autoimmune liver disease. J Hepatol. 2004;41:31–37. doi:10.1016/j. jhep.2004.03.008.
- Peiseler M, Sebode M, Franke B, Wortmann F, Schwinge D, Quaas A, Baron U, Olek S, Wiegard C, Lohse AW, et al. FOXP3+ regulatory T cells in autoimmune hepatitis are fully functional and not reduced in frequency. J Hepatol. 2012;57:125–32. doi:10.1016/j. jhep.2012.02.029.
- Jeffery HC, Jeffery LE, Lutz P, Corrigan M, Webb GJ, Hirschfield GM, Adams DH, Oo YH. Low-dose interleukin-2 promotes STAT-5 phosphorylation, Treg survival and CTLA-4-dependent function in autoimmune liver diseases. Clin Exp Immunol. 2017;188:394–411. doi:10.1111/cei.12940.
- Lim TY, Martinez-Llordella M, Kodela E, Gray E, Heneghan MA, Sanchez-Fueyo A. Low-dose interleukin-2 for refractory autoimmune hepatitis. Hepatology. 2018;68:1649–52. doi:10.1002/ hep.30059.

- Taubert R, Hardtke-Wolenski M, Noyan F, Wilms A, Baumann AK, Schlue J, Olek S, Falk CS, Manns MP, Jaeckel E. Intrahepatic regulatory T cells in autoimmune hepatitis are associated with treatment response and depleted with current therapies. J Hepatol. 2014;61:1106–14. doi:10.1016/j.jhep.2014.05.034.
- Esensten JH, Muller YD, Bluestone JA, Tang Q. Regulatory T-cell therapy for autoimmune and autoinflammatory diseases: the next frontier. J Allergy Clin Immunol. 2018;142:1710–18. doi:10.1016/j. jaci.2018.10.015.
- Tsokos GC, Lo MS, Reis PC, Sullivan KE. New insights into the immunopathogenesis of systemic lupus erythematosus. Nat Rev Rheumatol. 2016;12:716–30. doi:10.1038/nrrheum.2016.186.
- 92. Humrich JY, Morbach H, Undeutsch R, Enghard P, Rosenberger S, Weigert O, Kloke L, Heimann J, Gaber T, Brandenburg S, et al. Homeostatic imbalance of regulatory and effector T cells due to IL-2 deprivation amplifies murine lupus. Proc Natl Acad Sci U S A. 2010;107:204–09. doi:10.1073/pnas.0903158107.
- Ohl K, Tenbrock K. Regulatory T cells in systemic lupus erythematosus. Eur J Immunol. 2015;45(2):344–55. doi:10.1002/ eji.201344280.
- 94. Schmidt A, Rieger CC, Venigalla RK, Éliás S, Max R, Lorenz HM, Gröne HJ, Krammer PH, Kuhn A. Analysis of FOXP3+ regulatory T cell subpopulations in peripheral blood and tissue of patients with systemic lupus erythematosus. Immunol Res. 2017;65:551–63. doi:10.1007/s12026-017-8904-4.
- Lee HY, Hong YK, Yun HJ, Kim YM, Kim JR, Yoo WH. Altered frequency and migration capacity of CD4+ CD25+ regulatory T cells in systemic lupus erythematosus. Rheumatology. 2008;47:789–94. doi:10.1093/rheumatology/ken108.
- 96. He J, Zhang X, Wei Y, Sun X, Chen Y, Deng J, Jin Y, Gan Y, Hu X, Jia R, et al. Low-dose interleukin-2 treatment selectively modulates CD4+ T cell subsets in patients with systemic lupus erythematosus. Nat Med. 2016;22:991–93. doi:10.1038/nm.4148.
- Mizui M, Tsokos GC. Targeting regulatory T cells to treat patients with systemic lupus erythematosus. Front Immunol. 2018;9:786. doi:10.3389/fimmu.2018.00786.
- Smolen JS, Aletaha D, McInnes IB. Rheumatoid arthritis. Lancet. 2016;388(10055):2023–38. doi:10.1016/S0140-6736(16)30173-8.
- 99. Cao D, van Vollenhoven R, Klareskog L, Trollmo C, Malmström V. CD25brightCD4+ regulatory T cells are enriched in inflamed joints of patients with chronic rheumatic disease. Arthritis Res Ther. 2004;6:6. doi:10.1186/ar1025.
- 100. Han GM, O'Neil-Andersen NJ, Zurier RB, Lawrence DA. CD4 +CD25high T cell numbers are enriched in the peripheral blood of patients with rheumatoid arthritis. Cell Immunol. 2008;253(1– 2):92–101. doi:10.1016/j.cellimm.2008.05.007.
- 101. Liu MF, Wang CR, Fung LL, Lin LH, Tsai CN. The presence of cytokine-suppressive CD4+CD25+ T cells in the peripheral blood and synovial fluid of patients with rheumatoid arthritis. Scand J Immunol. 2005;62:312–17. doi:10.1111/j.1365-3083.2005.01656.x.
- 102. Lawson CA, Brown AK, Bejarano V, Douglas SH, Burgoyne CH, Greenstein AS, Boylston AW, Emery P, Ponchel F, Isaacs JD. Early rheumatoid arthritis is associated with a deficit in the CD4 +CD25high regulatory T cell population in peripheral blood. Rheumatology. 2006;45:1210–17. doi:10.1093/rheumatology/kel089.
- 103. Flores-Borja F, Jury EC, Mauri C, Ehrenstein MR. Defects in CTLA-4 are associated with abnormal regulatory T cell function in rheumatoid arthritis. Proc Natl Acad Sci U S A. 2008;105:19396–401. doi:10.1073/pnas.0806855105.
- 104. Nadkarni S, Mauri C, Ehrenstein MR. Anti-TNF-α therapy induces a distinct regulatory T cell population in patients with rheumatoid arthritis via TGF-β. J Exp Med. 2007;204:33–39. doi:10.1084/ jem.20061531.
- 105. Miyara M, Gorochov G, Ehrenstein M, Musset L, Sakaguchi S, Amoura Z. Human FoxP3 + regulatory T cells in systemic autoimmune diseases. Autoimmun Rev. 2011;10:744–55. doi:10.1016/j. autrev.2011.05.004.

- 106. Morita T, Shima Y, Wing JB, Sakaguchi S, Ogata A, and Kumanogoh A. The proportion of regulatory T cells in patients with rheumatoid arthritis: a meta-Analysis. PLoS One. 2016;11 9 e0162306 doi:10.1371/journal.pone.0162306.
- 107. Kelchtermans H, De Klerck B, Mitera T, Van Balen M, Bullens D, Billiau A, Leclercq G, Matthys P. Defective CD4+CD25+ regulatory T cell functioning in collagen-induced arthritis: an important factor in pathogenesis, counter-regulated by endogenous IFN-gamma. Arthritis Res Ther. 2005;7:402–15. doi:10.1186/ar1500.
- 108. Morgan ME, Flierman R, Van Duivenvoorde LM, Witteveen HJ, Van Ewijk W, Van Laar JM, De Vries RRP, Toes REM. Effective treatment of collagen-induced arthritis by adoptive transfer of CD25+ regulatory T cells. Arthritis Rheum. 2005;52:2212–21. doi:10.1002/art.21195.
- 109. Zaiss MM, Axmann R, Zwerina J, Polzer K, Gückel E, Skapenko A, Schulze-Koops H, Horwood N, Cope A, Schett G. Treg cells suppress osteoclast formation: a new link between the immune system and bone. Arthritis Rheum. 2007;56:4104–12. doi:10.1002/art.23138.
- 110. van Amelsfort JM, van Roon JA, Noordegraaf M, Jacobs KM, Bijlsma JW, Lafeber FP, Taams LS. Proinflammatory mediator-induced reversal of CD4+,CD25+ regulatory T cell-mediated suppression in rheumatoid arthritis. Arthritis Rheum. 2007;56:732–42. doi:10.1002/art.22414.
- 111. LeRoy EC, Medsger TJ. Criteria for the classification of early systemic sclerosis. J Rheumatol. 2001;28:1573-76.
- Frantz C, Auffray C, Avouac J, Allanore Y. Regulatory T cells in systemic sclerosis. Front Immunol. 2018;9. doi:10.3389/ fimmu.2018.02356.
- 113. Antiga E, Quaglino P, Bellandi S, Volpi W, Del Bianco E, Comessatti A, Osella-Abate S, De Simone C, Marzano A, Bernengo MG, et al. Regulatory T cells in the skin lesions and blood of patients with systemic sclerosis and morphoea. Br J Dermatol. 2010;162:1056–63. doi:10.1111/j.1365-2133.2010.09633.x.
- 114. Kataoka H, Yasuda S, Fukaya S, Oku K, Horita T, Atsumi T, Koike T. Decreased expression of Runx1 and lowered proportion of Foxp3+ CD25+ CD4+ regulatory T cells in systemic sclerosis. Mod Rheumatol. 2015;25:90–95. doi:10.3109/14397595.2014.899736.
- 115. Klein S, Kretz CC, Ruland V, Stumpf C, Haust M, Hartschuh W, Hartmann M, Enk A, Suri-Payer E, Oberle N, et al. Reduction of regulatory T cells in skin lesions but not in peripheral blood of patients with systemic scleroderma. Ann Rheum Dis. 2011;70:1475–81. doi:10.1136/ard.2009.116525.
- 116. Tsukamoto H, Nagafuji K, Horiuchi T, Mitoma H, Niiro H, Arinobu Y, Inoue Y, To K, Miyamoto T, Iwasaki H, et al. Analysis of immune reconstitution after autologous CD34 + stem/progenitor cell transplantation for systemic sclerosis: predominant reconstitution of Th1 CD4 + t cells. Rheumatology. 2011;50:944–52. doi:10.1093/rheumatology/keq414.
- 117. Vonk MC, Marjanovic Z, Van Den Hoogen FHJ, Zohar S, Schattenberg AVMB, Fibbe WE, Larghero J, Gluckman E, Van Laar JM, Farge D, et al. Long-term follow-up results after autologous haematopoietic stem cell transplantation for severe systemic sclerosis. Ann Rheum Dis. 2008;67:98–104. doi:10.1136/ard.2007.071464.
- Verstappen GM, Kroese FGM, Bootsma H. T cells in primary Sjögren's syndrome: targets for early intervention. Rheumatol (United Kingdom). 2021;60:3088–98.
- 119. Liu MF, Lin LH, Weng CT, Weng M. Decreased CD4+CD25+bright T cells in peripheral blood of patients with primary Sjogren's syndrome. Lupus. 2008;17:34–39. doi:10.1177/0961203307085248.
- 120. Christodoulou MI, Kapsogeorgou EK, Moutsopoulos NM, Moutsopoulos HM. Foxp3+ T-regulatory cells in Sjögren's syndrome: correlation with the grade of the autoimmune lesion and certain adverse prognostic factors. Am J Pathol. 2008;173:1389–96. doi:10.2353/ajpath.2008.080246.
- 121. Nussbaum L, Chen YL, Ogg GS. Role of regulatory T cells in psoriasis pathogenesis and treatment. Br J Dermatol. 2021;184:14-24. doi:10.1111/bjd.19380.

- 122. Dong F, Li XH, Zhang KM, Yin GH. Functional characterization of CD4+ CD25+ regulatory T cells differentiated in vitro from bone marrow-derived hematopoietic cells of psoriatic patients. J Clin Dermatology. 2008;37:207–09.
- 123. Zhang L, Yang XQ, Cheng J, Hui RS, Gao TW. Increased Th17 cells are accompanied by FoxP3+ Treg cell accumulation and correlated with psoriasis disease severity. Clin Immunol. 2010;135:108–17. doi:10.1016/j.clim.2009.11.008.
- 124. Sugiyama H, Gyulai R, Toichi E, Garaczi E, Shimada S, Stevens SR, McCormick TS, Cooper KD. Dysfunctional blood and target tissue CD4 + CD25 high regulatory T cells in psoriasis: mechanism underlying unrestrained pathogenic effector T cell proliferation. J Immunol. 2005;174:164–73. doi:10.4049/jimmunol.174.1.164.
- 125. Yun WJ, Lee DW, Chang SE, Yoon GS, Huh JR, Won CH, Lee MW, Kim SE, Kim BJ, Moon KC, et al. Role of CD4 +CD25high+FOXP3+ regulatory T cells in psoriasis. Ann Dermatol. 2010;22:397–403. doi:10.5021/ad.2010.22.4.397.

- 126. Huang Z, Li W, Su W. Tregs in autoimmune uveitis. Adv Exp Med Biol. 2021;1278:205–27. doi:10.1007/978-981-15-6407-9_11.
- 127. Wu Y, Luo J, Garden OA, Christensen CB, Lildal T, Pedersen M, Magnusson M, Borghammer P, Ovesen T. Immunoregulatory cells in myasthenia gravis. Front Neurol. 2020;11:11. doi:10.3389/ fneur.2020.00011.
- 128. Herrnstadt GR, Steinmetz OM. The role of Treg subtypes in glomerulonephritis. Cell Tissue Res. 2020;385(2):293-304. doi:10.1007/s00441-020-03359-7.
- 129. Neumann K, Tiegs G. Immune regulation in renal inflammation. Cell Tissue Res. 2021;385(2):305–22. doi:10.1007/s00441-020-03351-1.
- Riley JL, June CH, Blazar BR. Human T regulatory cells as therapeutic agents: take a billion or so of these and call me in the morning. Immunity. 2010;30:656–65. doi:10.1016/j.immuni.2009.04.006.
- 131. Golab K, Leveson-Gower D, Wang XJ, Grzanka J, Marek-Trzonkowska N, Krzystyniak A, Millis JM, Trzonkowski P, Witkowski P. Challenges in cryopreservation of regulatory T cells (Tregs) for clinical therapeutic applications. Int Immunopharmacol. 2013;16:371–75. doi:10.1016/j.intimp.2013.02.001.