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Hydrogen peroxide induced by the fungicide prothioconazole triggers deoxynivalenol (DON) production by *Fusarium graminearum*

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Abstract

Background: *Fusarium* head blight is a very important disease of small grain cereals with *F. graminearum* as one of the most important causal agents. It not only causes reduction in yield and quality but from a human and animal healthcare point of view, it produces mycotoxins such as deoxynivalenol (DON) which can accumulate to toxic levels. Little is known about external triggers influencing DON production.

Results: In the present work, a combined *in vivo/in vitro* approach was used to test the effect of sub lethal fungicide treatments on DON production. Using a dilution series of prothioconazole, azoxystrobin and prothioconazole + fluoxastrobin, we demonstrated that sub lethal doses of prothioconazole coincide with an increase in DON production 48 h after fungicide treatment. In an artificial infection trial using wheat plants, the *in vitro* results of increased DON levels upon sub lethal prothioconazole application were confirmed illustrating the significance of these results from a practical point of view. In addition, further *in vitro* experiments revealed a timely hyperinduction of H₂O₂ production as fast as 4 h after amending cultures with prothioconazole. When applying H₂O₂ directly to germinating conidia, a similar induction of DON-production by *F. graminearum* was observed. The effect of sub lethal prothioconazole concentrations on DON production completely disappeared when applying catalase together with the fungicide.

Conclusions: These cumulative results suggest that H₂O₂ induced by sub lethal doses of the triazole fungicide prothioconazole acts as a trigger of DON biosynthesis. In a broader framework, this work clearly shows that DON production by the plant pathogen *F. graminearum* is the result of the interaction of fungal genomics and external environmental triggers.

Background

Fusarium graminearum is one of the main causal agents of *Fusarium* head blight (FHB) in small grain cereals [1]. Although FHB symptoms have a classical impact on yield, the major concern referred to FHB is the presence of mycotoxins. *Fusarium* spp. are able to produce a plethora of mycotoxins with diverse chemical and biological features [2]. This toxin fingerprint, inherent to the genetics of each individual strain, determines the chemotype of each particular *Fusarium* isolate. *F. graminearum* chemotypes are mainly characterized by type B trichothecenes among which deoxynivalenol (DON),

acetyldeoxynivalenol (3-ADON and 15-ADON) and nivalenol (NIV) are the most prevalent [3].

Although the genetic background of type B trichothecene production has been studied elaborately, a coherent view on the production profile of these mycotoxins during infection and colonization of a host is lacking and identifying or understanding mechanisms that regulate the production of these secondary metabolites remains a challenge [4-6]. To date, the role of the type B trichothecene DON during infection and colonization of plants remains a controversial issue. Using DON non-producing *Fusarium* strains, the importance of DON production during spread of the fungus throughout the grain host was demonstrated [4]. In concordance, DON production elicits defence responses in wheat [5]. This role for DON as a virulence factor, actively produced

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during the infection process, has been confirmed in many other studies [6-8]. Notwithstanding these compelling lines of evidence, other authors uncouple DON production from colonization and aggressiveness [9-11]. The aforementioned controversy illustrates nicely that besides the genotypical derived DON-chemotype, many environmental triggers are crucial to unequivocally delineate the DON-production by a strain of *Fusarium*. The involvement of external influences triggering DON production is further corroborated by research illustrating modulation of DON production by either abiotic factors such as a_w , temperature, available carbon and/or nitrogen source, and biotic factors such as presence of other fungi [12-16].

The importance of these external triggers in DON production is consolidated by the observation that the production level of mycotoxins in axenic *in vitro* cultures is often orders of magnitude lower than observed during infection and colonization of a host, suggesting that specific host signals are involved in eliciting mycotoxins production. The secondary plant signalling compound hydrogen peroxide (H_2O_2), which is involved in plant-fungi interactions, is highlighted as an possible trigger interfering with type B trichothecene production. In previous work with *F. graminearum*, it was demonstrated that exogenously applied H_2O_2 at time of spore germination resulted in higher DON and A-DON levels 30 days later [17]. In addition, this DON accumulation was accompanied by an up-regulation of the *tri* gene machinery, responsible for DON biosynthesis [18,19]. Moreover, liquid cultures of *F. graminearum* supplied with H_2O_2 started to produce H_2O_2 themselves and the kinetics of this paralleled with DON accumulation [19] indicating a link between DON production and oxidative stress. Notwithstanding this clear observation, underlying mechanisms remain elusive. Recently, evidence is brought forward that the response of *Fusarium* to H_2O_2 is chemotype dependent. Ponts et al. (2009) observed a reduced NIV production in these chemotypes upon exogenous H_2O_2 application while the opposite was observed in DON chemotypes. Furthermore these data suggest that NIV isolates combine this adaptation to oxidative stress with a proliferated virulence [20].

The application of fungicides as possible external triggers for trichothecene biosynthesis remains a controversial issue. Several authors have described that sublethal concentrations of fungicides trigger trichothecene biosynthesis [21-23]. Others report opposite results [24,25].

The objective of the present work, was to investigate the influence of three fungicides i.e. prothioconazole (a triazole fungicide), azoxystrobin (a strobilurin fungicide) and prothioconazole + fluoxastrobin, applied at sub lethal concentrations on DON production by *F. graminearum*.

Triazoles are known inhibitors of the ergosterol biosynthesis in fungi while strobilurin fungicides inhibit mitochondrial electron transport by binding the Qo site of cytochrome bc1 complex. Where the effectiveness of triazole fungicides against *Fusarium* spp. is a certainty, the activity of strobilurins against *Fusarium* spp. is doubtful.

The hypothesis of a fungicide-induced oxidative stress response as a trigger for DON biosynthesis was evaluated by a combined approach of H_2O_2 measurements and application of the H_2O_2 scavenger enzyme catalase. Finally, the work was validated on a laboratory scale in an *in vivo* assay using wheat plants. The present work clearly demonstrates the risks of reduced fungicide doses with respect to DON accumulation.

Results

Effectiveness of fungicides to inhibit conidial germination and to reduce fungal biomass

Strobilurins and triazoles are among the most frequently used fungicides to respectively control *M. nivale* and *F. graminearum*. Nevertheless, application of these chemicals is often suboptimal due to the short vulnerable period of the pathogen in the field (during anthesis of the host), and environmental factors such as rain and wind. To determine if suboptimal fungicide treatments influence germination of *F. graminearum* conidia and DON production, an *in vitro* assay was set up using a dilution series of azoxystrobin, prothioconazole and fluoxastrobin + prothioconazole. Azoxystrobin did not influence the *F. graminearum* conidial germination at any of the given time points in a concentration-dependent way (Figure 1C). In contrast, prothioconazole effectively inhibited conidial germination at field dose and in dilutions 1/10 and 1/100 but did not have a significant effect at lower doses at time point 48 h (Figure 1B). At time intervals 4 h and 24 h, intermediate concentrations caused a temporary delay in germination. Finally the combination of prothioconazole and fluoxastrobin exhibited fungicidal activity at field concentration and inhibited germination in dilutions 1/100 and 1/100 and displayed no or very little effect in dilution 1/1000 (Figure 1A). Similar results were observed at the level of mycelial radial outgrowth (data not shown).

The effect of the different fungicides on conidial germination was also reflected in the amount of fungal biomass as measured by Q-PCR analysis (Table 1). These Q-PCR data clearly highlighted an effect of prothioconazole and prothioconazole + fluoxastrobin on *Fusarium* growth.

Effect of fungicides on DON production

To check whether the effect of the strobilurin fungicides and the triazole fungicide prothioconazole on fungal

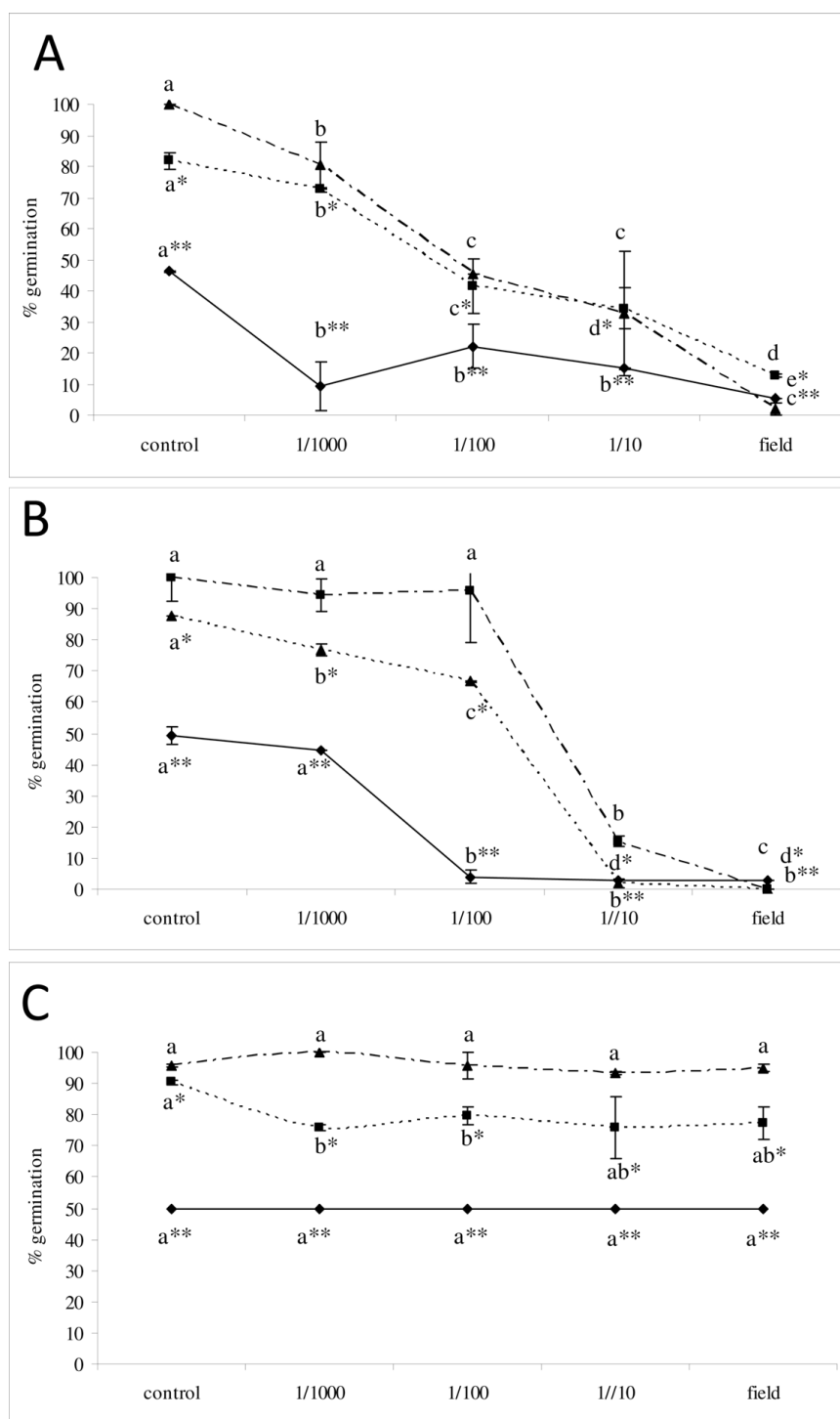


Figure 1 Effect of prothioconazole + fluoxastrobin (a), prothioconazole (b) and azoxystrobin (c) on conidial germination of *F. graminearum*. Conidia at a concentration of 10^6 conidia/ml were challenged with a tenfold dilution series of fluoxastrobin + prothioconazole, azoxystrobin and prothioconazole starting from 0.5 g/l + 0.5 g/l, 0.83 g/l and 0.67 g/l. For each treatment and repetition 50 conidia were scored for their germination and percentage of conidial germination was calculated at 4 h (solid line), 24 h (dashed line) and 48 h (point dashed line) after staining with 0.02% of cotton blue in lactic acid. Experiment consisted of two repetitions for each treatment and the experiment was repeated three times. Graphs represent the average of all three experiments. Different letters at each data point indicate differences from the control treatment at 4 h (**), 24 h (*) and 48 h after analysis with a Kruskal-Wallis and Mann-Whitney test with a sequential Bonferroni correction for multiple comparisons.

Table 1 Effect of a tenfold dilution series of prothioconazole, prothioconazole + fluoxastrobin and azoxystrobin on fungal biomass measured by Q-PCR analysis.

	prothio	prothio+catalase*	prothio+fluoxa	prothio+fluoxa+catalase*	azoxy	azoxy+catalase*
control	235.68 ^a	194.60 ^a	255.68 ^a	245.89 ^a	251.57 ^a	232.45 ^a
1/1000	9.42 ^b	63.03 ^b	76.23 ^b	48.17 ^b	267.16 ^a	230.12 ^a
1/100	2.35 ^c	31.13 ^c	16.58 ^c	44.90 ^b	250.01 ^a	234.93 ^a
1/10	2.51 ^c	50.02 ^{bc}	LD	LD	254.22 ^a	216.00 ^a
field	LD	33.47 ^c	LD	LD	236.54 ^a	170.72 ^a

F. graminearum biomass expressed as ng/μl. In each run, a no-template control was included. The amount of fungal material was measured based on a standard series of *F. graminearum* DNA ranging from 100 ng/μl down to 3.125 ng/μl which was carried out in triplicate.

Different letters indicate significant differences after analysis with a Kruskal-Wallis Mann-Whitney analysis with P = 0.05

Prothio: prothioconazole; azoxy: azoxystrobin; fluoxa:fluoxastrobin

*: Effect of catalase (1000 U/ml) added at the start of the experiment on the *F. graminearum* biomass.

LD: Lower than detection limit.

biomass and germination was paralleled by a reduced production of the type B trichothecene DON, levels of this mycotoxin were measured using a competitive ELISA-approach (Figure 2A, B, C). As expected, application of azoxystrobin did not influence DON production by *F. graminearum* strain 8/1. Remarkably, the combined application of prothioconazole and fluoxastrobin triggered a huge production of DON at the sub lethal doses of dilution 1/10 and 1/100, as early at time point 48 h but not at earlier time points (4 h and 24 h, data not shown). For the sole application of prothioconazole no major effects on DON production were observed since none of the tested concentrations were sub lethal. In an additional experiment using an extra intermediate concentration of 1/50 of the field concentration of prothioconazole, a reduced spore germination of about 50% was observed (data not shown). Concomitant with this observation, this sub lethal dilution resulted in an increased DON production (32 μg/μg of fungal DNA). Hence, application of sub lethal concentrations of respectively prothioconazole + fluoxastrobin and prothioconazole seems to result in the activation of the trichothecene biosynthesis machinery leading to an accumulation of DON as fast as 48 h after the start of the experiment.

Timely production of H₂O₂ precedes DON accumulation in combined strobilurin and triazole fungicide application

As several lines of evidence in literature corroborate an important role for reactive oxygen species (ROS) and more specifically H₂O₂ in stress responses of fungi, the accumulation of H₂O₂ upon fungicide application was monitored in the established *in vitro* germination assay. In these experiments, we unequivocally demonstrated that sole application of respectively azoxystrobin and prothioconazole at the given concentrations did not result in elevated H₂O₂ concentrations at any of the time points (Figure 3). In addition, prothioconazole at field dose resulted in lower H₂O₂ concentrations than

those observed in control samples possibly reflecting the reduction in microbial metabolic activity due to the application of the fungicide. Sub lethal dilutions of the combined application of fluoxastrobin + prothioconazole (i.e. 1/10 and 1/100) resulted in an increased H₂O₂ content in the medium compared to the control and the other treatments as fast as 4 h after the start of the germination assay. Although the increase at concentration 1/100 was less proliferate than the increase at concentration 1/10 of the field dose of fluoxastrobin + prothioconazole, it was consistent in all performed experiments. Moreover, this peak in H₂O₂ disappeared or was less proliferated at later time points 24 h and 48 h. These findings strongly suggest that timely production of H₂O₂ triggers the trichothecene biosynthesis machinery to produce DON in sub lethal fungicide treatments.

To further examine the role of H₂O₂ in fungicide-induced stress, exogenous catalase was added together with the fungicidal treatment. At 4 h after application, catalase resulted in a reduced germination rate (Figure 4A, B) compared to all non-catalase treatments. In addition, at later time points, the application of catalase partially abolished the fungicidal effect of prothioconazole + fluoxastrobin (Figure 4C) and of prothioconazole (Figure 4D) at both the level of conidial germination and fungal biomass (Table 1). No effect was observed in the treatment with azoxystrobin (data not shown). In addition, this partial loss of fungicidal effect due to the application of catalase was accompanied by the disappearance of the H₂O₂ peak previously observed in the prothioconazole + fluoxastrobin treated samples at 4 h after application of prothioconazole (Figure 5A). No peak was observed in the treatment with sole application of prothioconazole (Figure 5B). At later time points, no H₂O₂ accumulation was observed in none of the treatments (data not shown). Finally, completely in line with these observations, the disappearance of the H₂O₂ trigger at 4 h due to the application of catalase resulted in DON production comparable to control treatments (Figure 2D, E, F).

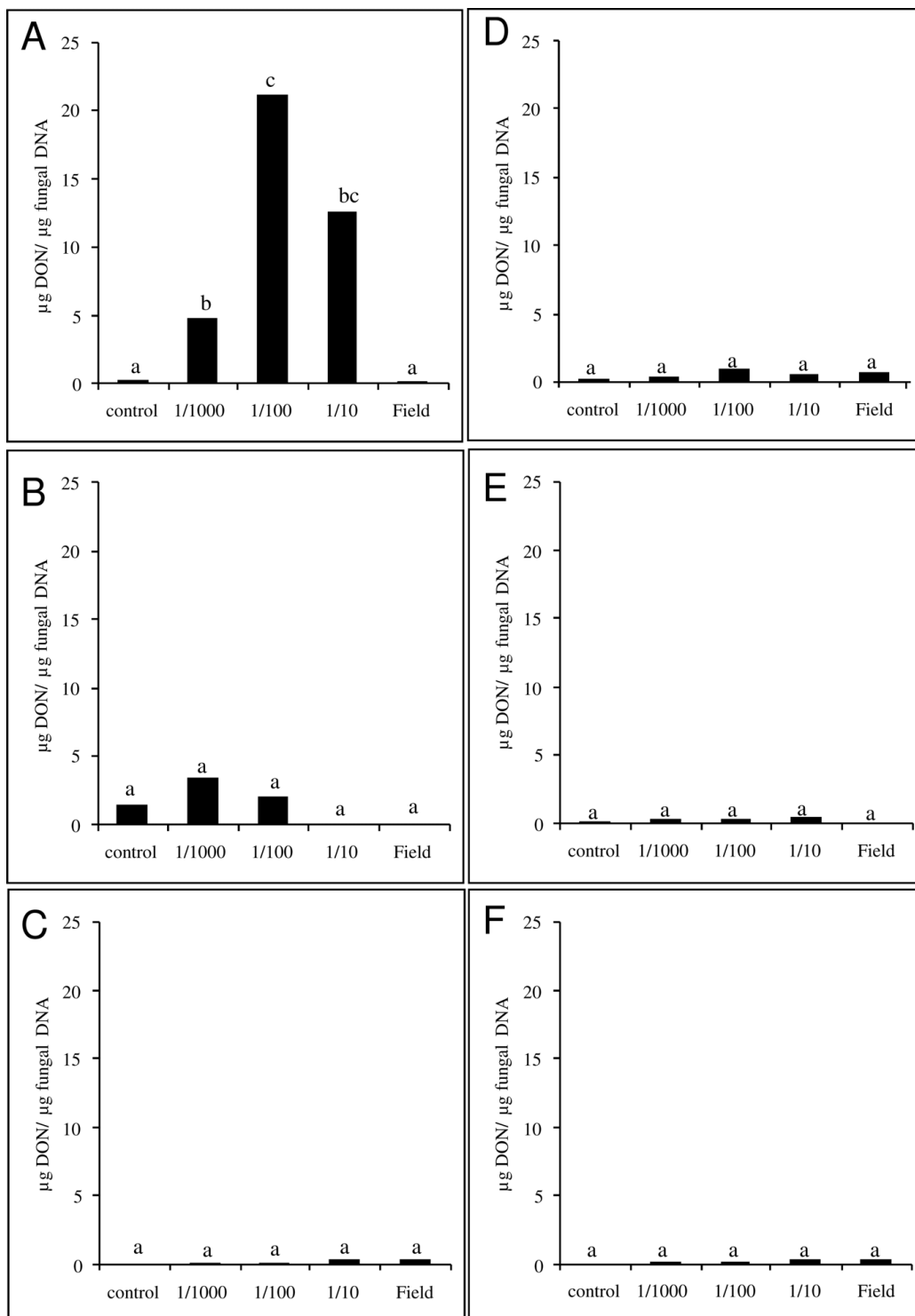


Figure 2 Effect of prothioconazole + fluoxastrobin (a), prothioconazole (b) and azoxystrobin (c) alone or in combination with catalase (d,e,f) on production of deoxynivalenol (DON) by *F. graminearum*. Conidia at a concentration of 10^6 conidia/ml were challenged with a tenfold dilution series of fluoxastrobin + prothioconazole, azoxystrobin and prothioconazole starting from 0.5 g/l + 0.5 g/l, 0.83 g/l and 0.67 g/l in absence (a,b,c) or presence (e,f,g) of 1000 U/ml catalase. DON content in the medium was determined using a competitive ELISA approach 48 h after start of the experiments. Each bar is the result of two pooled samples to reduce variance. The experiment was repeated twice in time of which one representative experiment is shown in the figure. Different letters above bars indicate significant differences after analysis with a Kruskal-Wallis and Mann-Whitney test with a sequential Bonferroni correction for multiple comparisons.

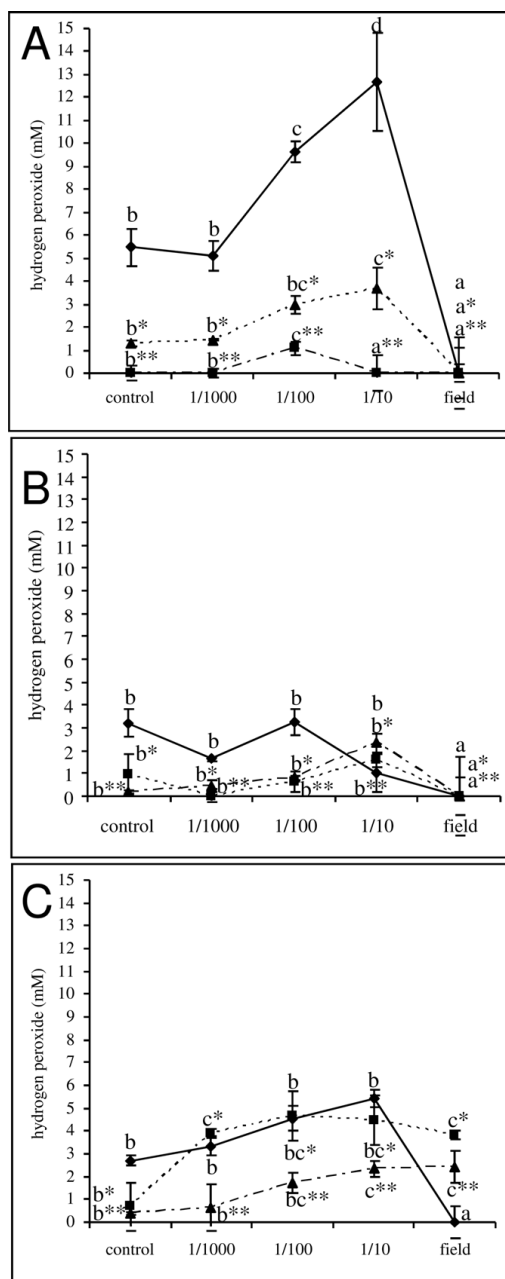


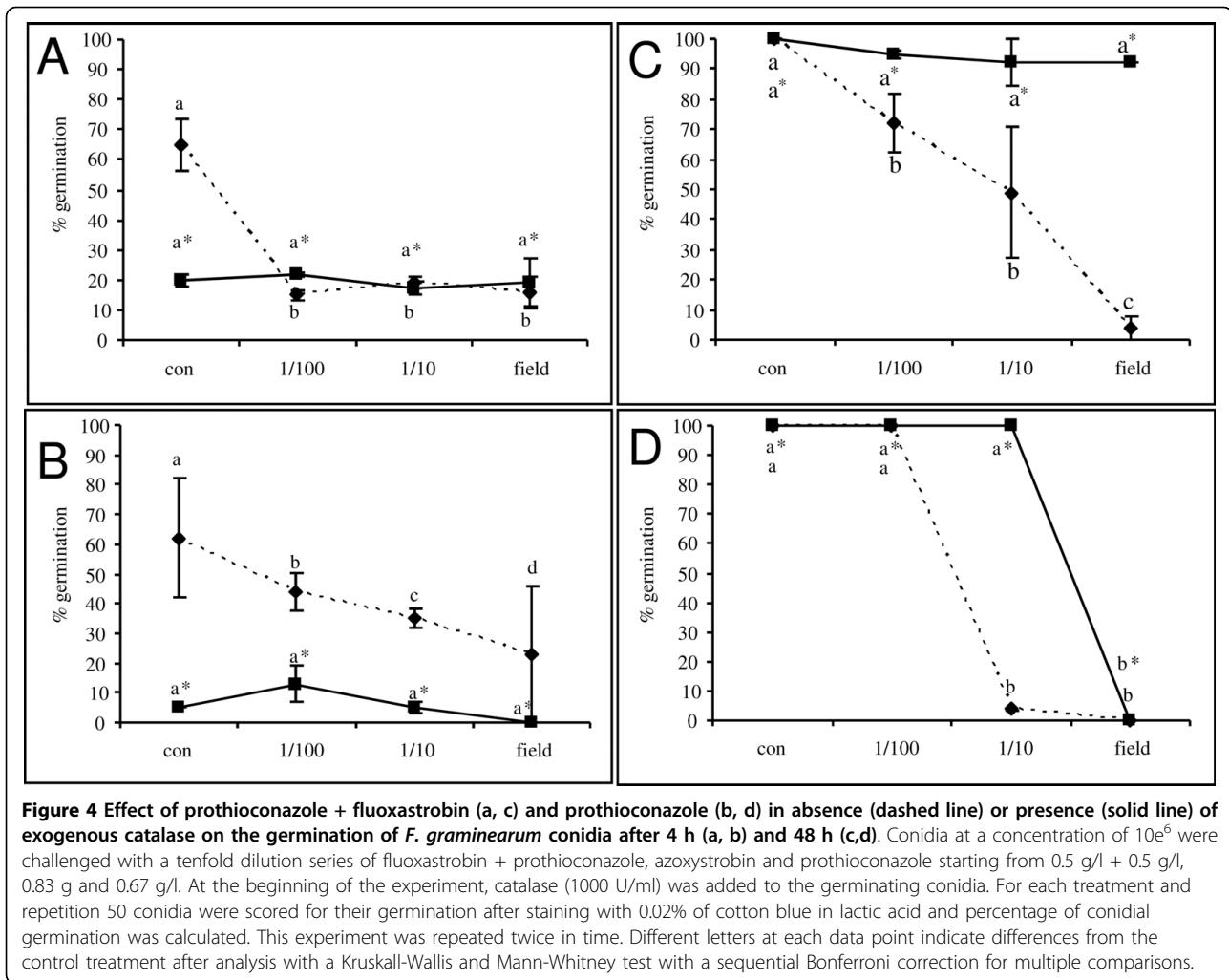
Figure 3 Effect of prothioconazole + fluoxastrobin (a), prothioconazole (b) and azoxystrobin (c) on extracellular H₂O₂ concentrations. Conidia at a concentration of 10⁶ conidia/ml were challenged with a tenfold dilution series of fluoxastrobin + prothioconazole, azoxystrobin and prothioconazole starting from 0.5 g/l + 0.5 g/l, 0.83 g/l and 0.67 g/l. H₂O₂ was measured at 4 h (solid line), 24 h (dashed line) and 48 h (point dashed line) using TMB (trimethylbenzidine) as a substrate in the presence of an overdose of peroxidase. The H₂O₂ concentrations were calculated based on a standard curve included in each experiment. Each data point is the result of three repetitions and the experiments were repeated twice in time. Different letters at each data point indicate differences from the control treatment at 4 h (**), 24 h (*) and 48 h after analysis with a Kruskal-Wallis and Mann-Whitney test with a sequential Bonferroni correction for multiple comparisons.

Stress-induced H₂O₂ accumulation upon fungicide application is necessary and sufficient as a trigger to induce DON

To further decipher a direct link between H₂O₂ at one hand and the production of the mycotoxin DON at the other hand, the accumulation of DON was monitored upon exogenously single pulse application of H₂O₂ ranging from 0.01 mM up to 100 mM. H₂O₂ influenced germination of *F. graminearum* conidia in a concentration-dependent manner (Figure 6). As early as 4 h after the start of the assay, exogenous application of H₂O₂ at concentrations from 1 mM up to 100 mM retarded or stopped conidial germination. The sub lethal concentration of 10 mM H₂O₂ induced DON production as fast as 4 h after application of H₂O₂ in one of the experiments. In the other experiment, 4 h was probably just too early to observe the increased DON production and in this experiment, the increment in DON was observed at 24 h. The ability of 10 mM H₂O₂ to initiate DON production is in concordance with H₂O₂ concentrations induced by sub lethal prothioconazole concentrations (Figure 3A). At later time points, DON did not further accumulate and concentration remained the same for the subsequent 24 and 48 h time points. This effect of H₂O₂ on DON production was confirmed by an experiment in which H₂O₂ was eliminated from the well plates by exogenously applied catalase. This resulted in a fall-back of the DON production in the 10 mM H₂O₂ treatment to levels comparable to control wells (data not shown). Finally, surprisingly, low concentrations of H₂O₂ facilitated conidial germination compared to control samples. Indicating the necessity of low levels of H₂O₂ in optimal germination of conidia and proliferation of fungal cells.

Sublethal prothioconazole + fluoxastrobin application triggers DON production *in vivo*

In an *in vivo* case study with azoxystrobin and prothioconazole + fluoxastrobin, the effect of sub lethal fungicide concentrations on growth and DON production was verified on wheat plants (variety Cadenza) during anthesis. A point inoculation with *F. graminearum* clearly led to typical *Fusarium* symptoms 14 days after inoculation (Figure 7). In the treatment with azoxystrobin, no reduction of symptoms was observed (data not shown) which is in concordance with the previously described *in vitro* data. Application of prothioconazole + fluoxastrobin resulted in a complete control of *Fusarium* at field dose or dilution 1/10 (Figure 7A). At concentration 1/100 symptoms were apparent although they were less proliferate than in the inoculated control plants pointing to a sub lethal concentration. Parallel with the symptom evaluation, DON content was determined in the wheat ears. No DON was apparent in treatments



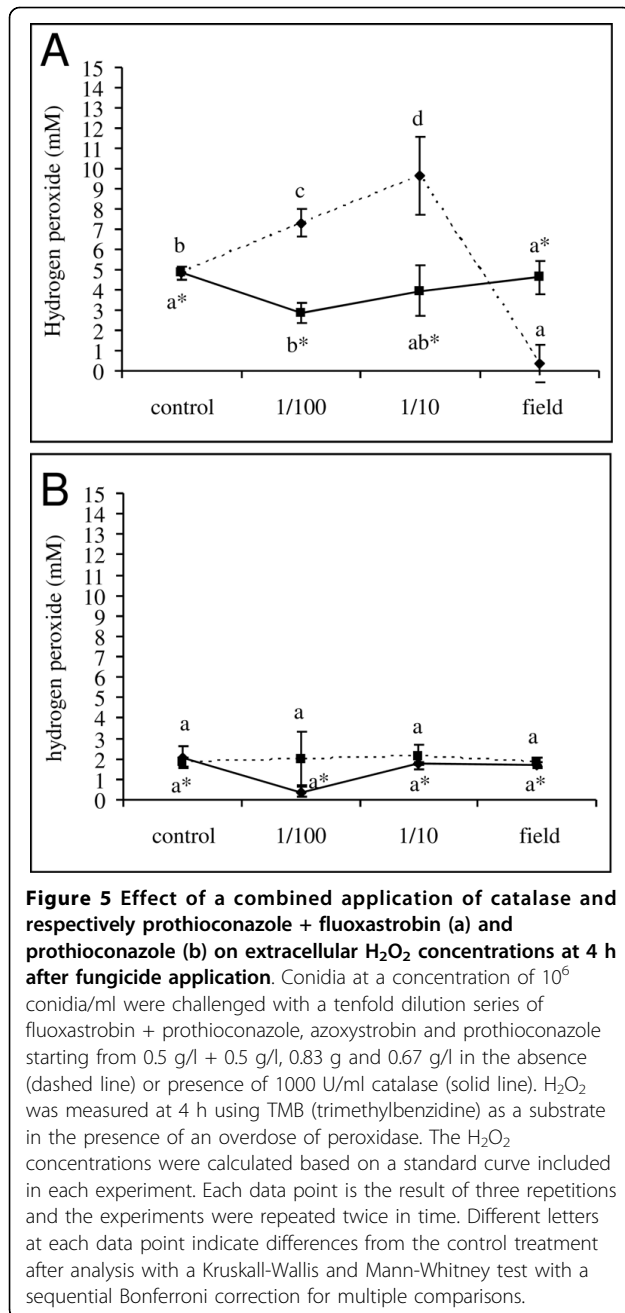
with field dose or dilution 1/10. However, a significant increase in DON content was observed in ears originating from the 1/100 treatment compared to the control treatment (Figure 7B) which is in concordance with the *in vitro* observations.

Discussion

In an effort to broaden our understanding of external triggers influencing the DON production machinery of *F. graminearum*, the effect of strobilurin and triazole fungicides on DON production was investigated. Our results demonstrate that prothioconazole, a triazole fungicide, has good control capacities culminating in reduced vegetative radial outgrowth, a reduced conidial germination and a reduction of *F. graminearum* biomass. Triazoles are known inhibitors of the ergosterol biosynthesis in fungi and have been described for their good control capacities against *Fusarium* spp [21].

On the contrary, the strobilurin fungicide azoxystrobin was not able to induce a reduction in radial outgrowth,

spore germination and fungal biomass. Strobilurin fungicides inhibit mitochondrial electron transport by binding the Qo site of cytochrome bc1 complex. Although the effectiveness of strobilurins against *Fusarium* spp. is doubtful, they have been reported to be effective against *F. culmorum* [24]. Apparently, *F. graminearum* is very resistant to this type of fungicides. Resistance to strobilurin fungicides has been reported in many species to be associated with a single amino acid replacement at position 143 of the cytochrome b gene [26-28]. Although this mechanism was recently described in *Microdochium nivale* it has not yet been described in *F. graminearum*. We assume that the observed resistance is therefore possibly a consequence of the activation of a respiratory chain using an alternative oxidase (AOX) bypassing complexes III and IV in the cytochrome mediated pathway. Activity of this AOX mediates electron transfer directly from ubiquinol to oxygen. Kaneko and Ishii (2009) demonstrated that *F. graminearum* acts very rapidly upon strobilurin application by

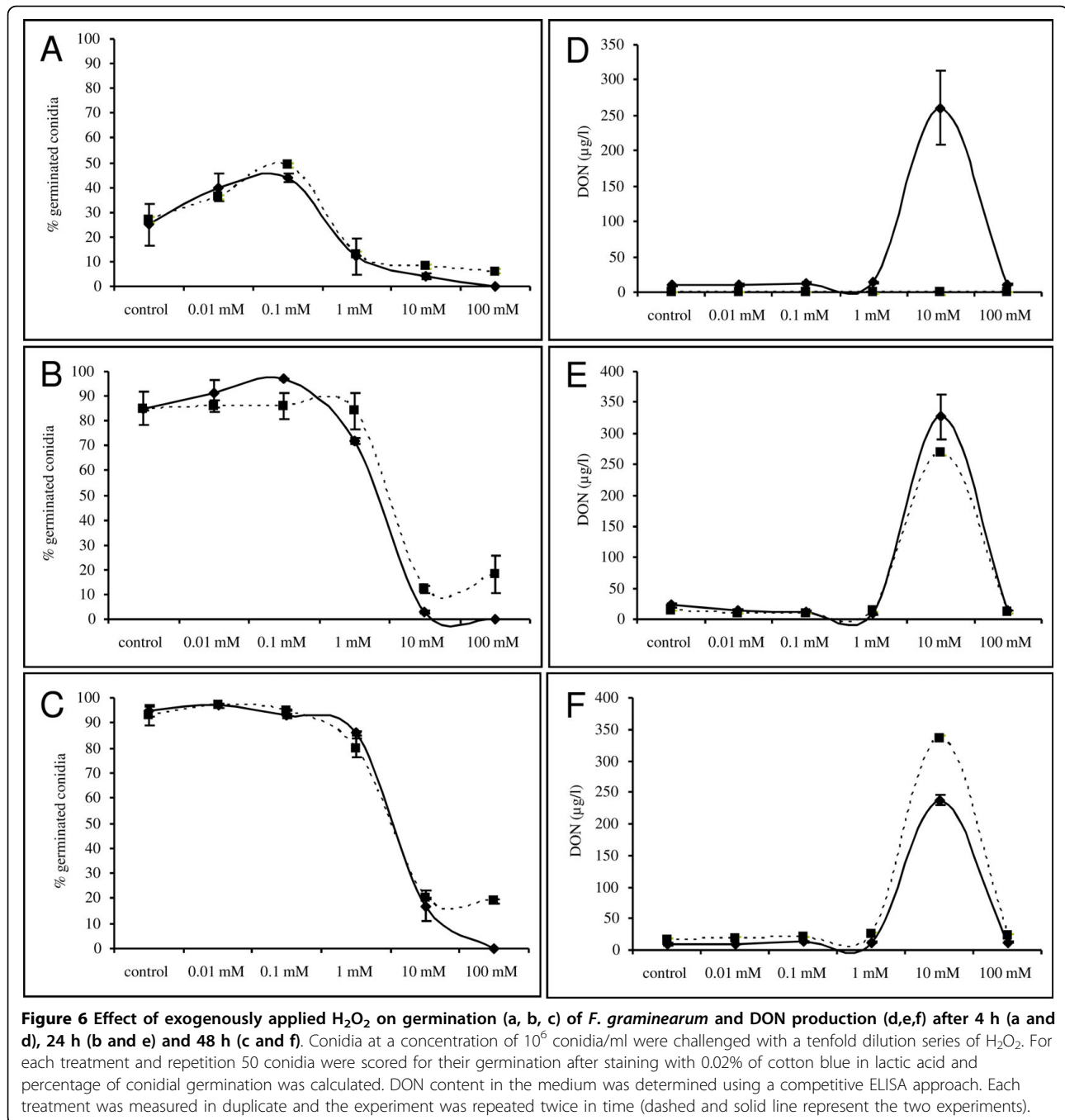


the activation of AOX whereas *M. nivale*, a fungal species susceptible to strobilurins, reacted slowly with a retarded moderate activation of this enzyme [29].

Since the generation of reactive oxygen species such as H₂O₂ is a hallmark of an oxidative stress response, extracellular H₂O₂ was measured upon fungicide application in an *in vitro* assay. Unexpectedly, application of strobilurin fungicides did not result in an increased extracellular H₂O₂ formation, which is at first sight, contradictory to previous findings by Kaneko and Ishii (2009) who found an increased production of H₂O₂

upon strobilurin application. However it is important to notice that in the present work the H₂O₂ released in the medium was measured whereas Kaneko and Ishii (2009) focused on intracellular H₂O₂. Remarkably, the application of sub lethal doses of prothioconazole or the combination of prothioconazole amended with fluoxastrobin resulted in a boosted H₂O₂ production as fast as 4 h after application. This prompt production disappeared at later time points. In addition, a clear induction of DON production was observed 48 h after application of sub lethal prothioconazole + fluoxastrobin concentrations. This induction of DON was confirmed in an *in vivo* experiment in which flowering wheat plants were infected with *F. graminearum* and subjected to a sub lethal dose of prothioconazole + fluoxastrobin. Previous work on *F. culmorum* demonstrated no or a negative effect of several strobilurins and triazoles on DON production [24] so the observed phenomenon of an increased DON production by *F. graminearum* induced by sub lethal concentrations of triazole fungicides might be a strain- or species-specific phenomenon.

It is tempting to speculate whether this accumulation of DON is the consequence of the preceding accumulation of H₂O₂ as such being the first link in a signalling cascade activated upon sub lethal triazole treatment. Although this key role of H₂O₂ is not unambiguously demonstrated in the present study, the amount of evidence is compelling: H₂O₂ precedes accumulation of DON, combined application of catalase (eliminating H₂O₂ from the medium) inhibited DON accumulation. In addition, the application led to a reduced activity of the triazole fungicide. Application of H₂O₂ to *F. graminearum* cultures led to a reduced germination and prompt induction of DON biosynthesis 4 h after H₂O₂ application. This additional experiment proves that H₂O₂ accumulation is necessary and sufficient to initiate DON production. The activation of the DON biosynthesis machinery by H₂O₂ is in concordance with previous observations by the group of Barreau [17,19,20] who demonstrated that exogenously applied H₂O₂ by repeated single or pulse-feeding resulted in accumulation of DON. However, these authors only monitored increases in DON at late time points such as 10 to 30 days after H₂O₂ application whereas we observe a clear prompt activation of DON production within hours. From a physiological point of view the effect of H₂O₂ during the initial germination events is logic and in line with the physiology of an in field *F. graminearum* infection: H₂O₂ is one of the key regulators in the plant defense system upon pathogen attack [30]. Therefore, this molecule is encountered frequently and at early time points by the pathogen in the interaction with its host. Previous work by the group of John Manners demonstrated beautifully that DON itself can induce



hypersensitive cell death and H₂O₂ during infection [5] and as such underpinning the interaction between both molecules.

Astonishingly, very low concentrations of H₂O₂ promoted conidia germination rate where a reduction was expected. We hypothesize that during germination events, very small amounts of H₂O₂ are beneficial and necessary in the primordial germination- and hyphal extension events. It is known that H₂O₂ is necessary in *de novo* synthesis of cell wall and membrane

components during germination and hyphal extension. Indirect evidence for this was provided by Seong et al (2008) who observed high activities of the peroxisomes at the onset of spore germination [31] The need for basal H₂O₂ is subscribed by the observation that catalase treatment results in a reduced spore germination at very early time points in germination. In several independent studies, it was demonstrated that reactive oxygen species such as H₂O₂ are key players and crucial in the regulation of cell differentiation in microbial eukaryotes

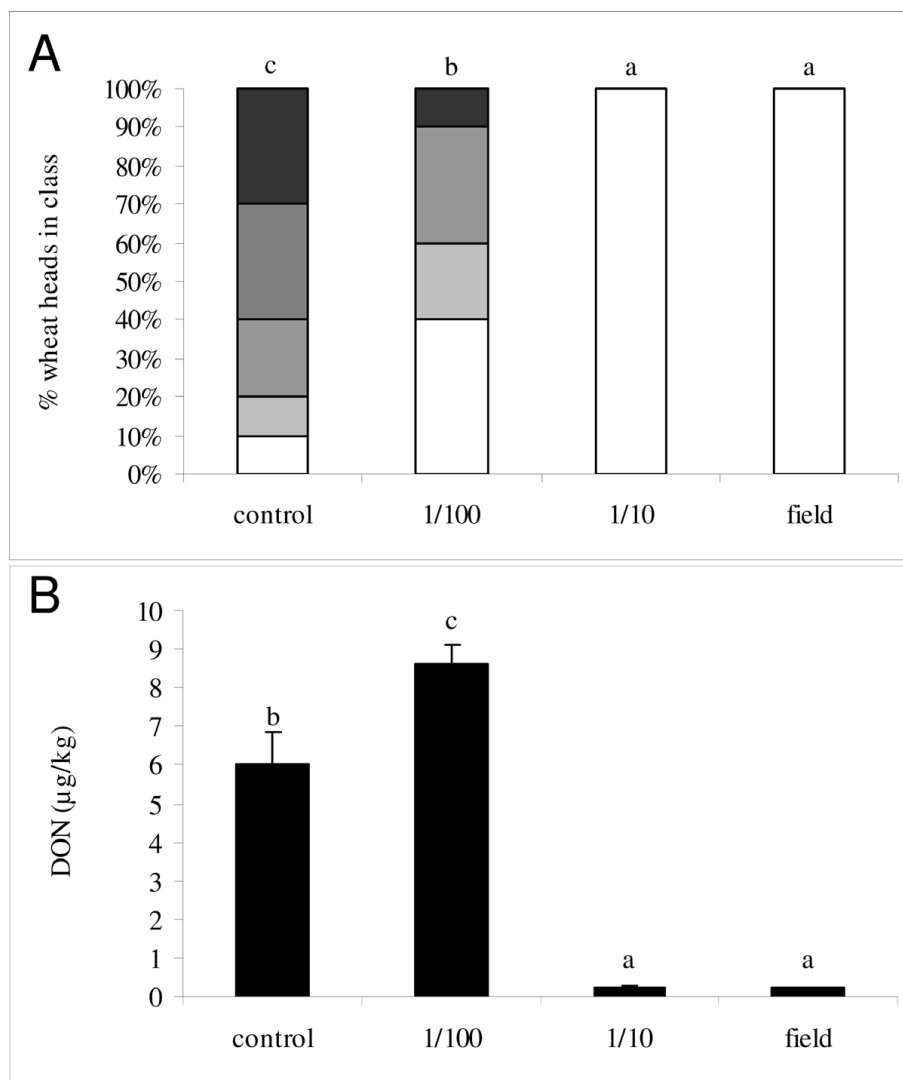


Figure 7 *In vivo* effect of prothioconazole + fluoxastrobin on symptoms of *F. graminearum* (a) and DON content (b) after point inoculation of wheat ears 14 days after infection. Wheat ears (variety Cadenza) were inoculated with two droplets of 20 μl of conidia at a concentration of 10^9 conidia/ml. Infection spots were indicated with a marker. Ears were subsequently treated with a tenfold dilution series of fluoxastrobin + prothioconazole starting from 0.5 g/l + 0.5 g/l. For each treatment, 10 plants were assessed for *Fusarium* symptoms. This experiment was repeated twice in time with analogous results. The figure represents one representative experiment. Different letters at each data point indicate differences from the control treatment after analysis with a Kruskal-Wallis and Mann-Whitney test with a sequential Bonferroni correction for multiple comparisons.

[32,33]. In accordance with this, it was demonstrated that NADPH oxidases which generate reactive oxygen are decisive in fungal cell differentiation and growth in a model system using *Neurospora crassa* [34].

Taken together, these results not only reinforce the hypothesis that H_2O_2 can induce DON biosynthesis but also suggest that DON accumulation induced by sub lethal triazole application is mediated through an increased production or release of H_2O_2 into the medium rendering a physiological interface of H_2O_2 influencing DON production. It is tempting to speculate on the

mechanistics behind these observations. We hypothesize that due to the inhibition of ergosterol biosynthesis by the application of triazole fungicides, an increased cell permeability results in the increased release of H_2O_2 in the medium which in turns activates the trichothecene biosynthesis machinery. Indeed, although H_2O_2 is a very reactive molecule which can diffuse freely across bio membranes, it has been shown in a *Sacharomyces* model system that organisms prevent H_2O_2 diffusion [35,36]. This hypothesis is subscribed by accumulating indirect evidence in many other fungi. As such in

Candida ergosterol depletion increases vulnerability to phagocytic oxidative damage [37]. In *Sacharomyces* it was demonstrated using ergosterol knock out mutants that ergosterol depletion results in a changed biophysical property of the plasma membrane leading to an increased permeability towards H₂O₂[38].

Although beyond the scope of the present paper it is important to notice that triazole fungicides on their own can generate H₂O₂ *in planta* as an intermediate metabolite in plants through activation of antioxidant systems [39] generating as such a greening effect which results in a retardation of the senescence [40]. The effect of this physiological induced H₂O₂ *in planta* on DON production by an invading *F. graminearum* is till now not studied and certainly needs more attention in the future.

Conclusions

In the present work it was shown that sub lethal prothioconazole concentrations resulted in a significant increase in DON production by *F. graminearum* in a combined approach of an *in vitro* assay and an artificial infection trial. In the *in vitro* assay, the stimulated DON production was preceded by a prompt induction of H₂O₂ suggesting that the proliferated DON production was induced by an oxidative stress response in the fungus. This hypothesis was confirmed by exogenous application of catalase which abrogated the elevated DON production observed at the sub lethal doses of prothioconazole. In a broader framework, this work clearly shows that DON production by the plant pathogen *F. graminearum* is the result of the interaction of fungal genomics and external triggers. Further work is needed to characterise the effect of these external triggers influencing DON biosynthesis. This work will certainly lead to a better insight into factors that influence DON production under field conditions.

Methods

Fungal Material, induction of conidia, conidia suspension and conidia counting

A GFP transformant of *Fusarium graminearum* strain 8/1 [41] was grown on potato dextrose agar (PDA) for 7 days at 20°C and kept at 4°C upon use in the germination assays. Conidia of *F. graminearum* were obtained by incubating a mycelium plug on a PDA plate for 7 days under a light regime of UV/darkness (12 h 365 nm 10 W/12 h). Macroconidia were harvested by adding distilled water amended with 0.01% of Tween20 to the fully grown PDA plates and by rubbing the conidia-bearing mycelium with a spatula. Conidia were counted and diluted to a final concentration of 10⁶ conidia/ml. In the germination assays, fungal conidia were visualised using a 0.02% cotton blue solution prepared in lactic acid.

In vitro growth and germination assay, exogenous application of fungicides and H₂O₂

In the present study, 3 fungicides were used i.e. fluoxastrobin+prothioconazole, azoxystrobin and prothioconazole. Field doses of each fungicide were the point of departure for the *in vitro* assay. The field dose of each fungicide differed according to the manufacturers instructions and mounted to 0.5 g/l + 0.5 g/l, 0.83 g and 0.67 g for respectively fluoxastrobin+prothioconazole, azoxystrobin and prothioconazole.

In experiments aiming to measure fungal biomass and conidia germination, a ten-fold dilution series of these three fungicides was prepared to obtain a final concentration of 1/1000, 1/100, 1/10 and field dose of each fungicide in the 24-well plates in which the assay was executed. In these wells, 250 µl of conidial suspension was added and amended with 250 µl of the fungicide dilution. These wells were incubated at 20°C. Each treatment consisted out of 2 repetitions and the experiment was repeated three times independently in time. Control treatments consisted of 250 µl of spore suspension and 250 µl of distilled water.

H₂O₂ was applied once at the beginning of the germination trials in a final concentration ranging from 0.01 mM, 0.1 mM, 1 mM up to 10 mM. 250 µl of H₂O₂ solution was added to 250 µl of spore suspension. Each treatment consisted out of 2 repetitions and the experiment was repeated three times. Control treatments consisted of 250 µl of spore suspension and 250 µl of distilled water.

Infection of wheat plants and application of fungicides *in vivo*

F. graminearum macroconidia were obtained and harvested as previously described. A conidia suspension of 10⁶ conidia/ml was prepared. A dilution series of fluoxastrobin and azoxystrobin + prothioconazole was prepared starting from the field dose as mentioned in the *in vitro* assays. Ten ears of wheat plants at flowering stage (Zadok's stage 60) were infected with 2 droplets of 20 µl of conidia suspension. Subsequently, the infected wheat plants were sprayed with fungicide dilutions till run off and placed in a growth chamber at 22°C under a relative humidity of 100% for 2 days to guarantee the conidial germination and penetration. After 2 days, the plants were incubated for 12 days in a growth chamber at 22°C under a light regime of 16 h light/8 h dark. Fourteen days after inoculation, the infection was assessed based on the surface of the ear covered with *Fusarium* symptoms: 1 = healthy; 2 = up to 25%; 3 = 25 to 50%; 4 = 50 to 75%; 5 = 75 to 100% of the ear covered with symptoms. The experiment was repeated twice in time.

DNA extraction and fungal quantification using a Q-PCR approach

To quantify the amount of *Fusarium* biomass in the *in vitro* assays, fungal biomass retrieved from each individual well was centrifuged and supernatant was eliminated. The pellet freeze-dried for 6 h at -10°C and 4 h at -50°C (Christ Alpha 1-2 LD Plus, Osterode, Deutschland). Samples were stored at -20°C upon extraction.

DNA extraction was performed as previously described by Audenaert et al. (2009) [42] based on the method established by Shaghai and Mahroof et al. (1989) [43]. For PCR, amplification of the EF1 α gene, the forward primer FgramB379 (5'-CCATTCCCTGG-GCGCT-3') and the reverse primer FgramB411 (5'-CCTATTGACAGGTGGTTAGTACTGG-3') were used [44]. The real-time PCR mix consisted of 12.5 μl 2 \times SYBR Green PCR Master Mix (Stratagene), 250 nM of each primer, 0.5 $\mu\text{g}/\mu\text{l}$ bovine serum albumin (BSA) and 2 μl of template DNA. PCR was performed on a 7000 series Detection System (Applied Biosystems) using the following PCR protocol: 2 min at 50°C , 10 min at 95°C , 40 cycles of 95°C for 15 s and 62°C for 1 min followed by a dissociation analysis at 55°C to 95°C .

A standard curve was established in threefold using a twofold dilution series of pure fungal DNA from 100 ng up to 3.125 ng. The amount of fungal DNA was calculated from the cycle threshold (Ct) and the amount of fungal material in control samples.

Measurement of H₂O₂ and DON, application of catalase

H₂O₂ formation in the fungicide experiments was measured 4 h, 24 h and 48 h post inoculation using a TMB (trimethylbenzidine) assay. This assay is based on the conversion of TMB to a blue stain upon reaction with H₂O₂ in the presence of peroxidases. 250 μl of the conidia suspension was removed from a well and amended with an excess of 100 μl horse radish peroxidase (500 U/ml) and 150 μl of TMB (1 mg/ml). TMB was dissolved in 100% ethanol and the stock solution of 1 mg/ml was prepared in 50 mM of Tris-acetate buffer (pH 5.0). H₂O₂ formation was determined by measuring the absorbance at 620 nm in duplicate in each time point and in two independent experiments. In each experiment, a standard curve of pure H₂O₂ was added in a concentration range of 0.01 mM up to 100 mM. The H₂O₂ formed in the *in vitro* assay was calculated based on this standard curve.

DON concentration was measured by ELISA using the Veratox DON 5/5 kit (Biognost, Neogen, Leest, Belgium). The lower limit of detection was 0.1 ppm. A standard curve was established using 0, 0.25, 0.4, 1 and 2 ppm DON. The ELISA kit provides 100% specificity for DON. 200 μl of the conidia suspension was removed

from each well. Two repetitions per treatment were pooled and subsequently centrifuged to eliminate the fungal pellet. 100 μl of this supernatant was used for further analysis in the ELISA assay. Experiments in which DON content was measured were repeated twice in time with two repetitions per experiment and treatment. In the *in vivo* experiments, 1 g of grains was ground and extracted in 10 ml of distilled water. Subsequently, the extract was analyzed by ELISA as described above. The DON content was measured in five fold.

In the *in vitro* experiments using catalase, 125 μl of Catalase from bovine liver (Sigma, Bornem, Belgium) was added to the wells to a final concentration of 1000 U/ml. In the experiments where catalase was applied, 250 μl of conidia were amended with 125 μl of fungicides. Care was taken that the final concentration of the fungicides was the same as aforementioned in the other studies.

Data analysis

Differences in DON levels, H₂O₂ content, disease assessment, germination and fungal diameter were detected using a non-parametric Kruskal-Wallis and Mann-Whitney test with a sequential Bonferroni correction for multiple comparisons. Differences between DON levels and disease severity were considered at $P = 0.05/(n-1)$ with n the number of cases in the study. All data were analyzed using SPSS-software (Originally: Statistical Package for Social Sciences) version 15.0 for WindowsXP.

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Authors' contributions

KA conceived of the study, carried out most of the *in vitro* assays and drafted the manuscript. EC carried out the immunoassays and helped with the *in vitro* assays partim conidial germination. GH, MH and SDS coordinated and helped to draft the manuscript. All authors read and approved the final manuscript.

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References

- Goswami RS, Kistler HC: **Heading for disaster: *Fusarium graminearum* on cereal crops.** *Molecular Plant Pathology* 2004, **5**(6):515-525.
- Bottalico A, Perrone G: **Toxigenic *Fusarium* species and mycotoxins associated with head blight in small-grain cereals in Europe.** *European Journal of Plant Pathology* 2002, **108**(7):611-624.
- Desjardins AE: ***Gibberella* from A (venaceae) to Z (eae).** *Annual Review of Phytopathology* 2003, **41**:177-198.
- Bai GH, Desjardins AE, Plattner RD: **Deoxynivalenol-nonproducing *Fusarium graminearum* causes initial infection, but does not cause disease spread in wheat spikes.** *Mycopathologia* 2002, **153**(2):91-98.
- Desmond OJ, Manners JM, Stephens AE, MacClean DJ, Schenk PM, Gardiner DM, Munn AL, Kazan K: **The *Fusarium* mycotoxin deoxynivalenol elicits hydrogen peroxide production, programmed cell death and defence responses in wheat.** *Molecular Plant Pathology* 2008, **9**(4):435-445.
- Mudge AM, Dill-Macky R, Dong YH, Gardiner DM, White RG, Manners JM: **A role for the mycotoxin deoxynivalenol in stem colonisation during crown rot disease of wheat caused by *Fusarium graminearum* and *Fusarium pseudograminearum*.** *Physiological and Molecular Plant Pathology* 2006, **69**(1-3):73-85.
- Hestbjerg H, Felding G, Elmholt S: ***Fusarium culmorum* infection of barley seedlings: Correlation between aggressiveness and deoxynivalenol content.** *Journal of Phytopathology-Phytopathologische Zeitschrift* 2002, **150**(6):308-312.
- Goswami RS, Kistler HC: **Pathogenicity and *in planta* mycotoxin accumulation among members of the *Fusarium graminearum* species complex on wheat and rice.** *Phytopathology* 2005, **95**(12):1397-1404.
- Liu WZ, Langseth W, Skinnis H, Elen ON, Sundheim L: **Comparison of visual head blight ratings, seed infection levels, and deoxynivalenol production for assessment of resistance in cereals inoculated with *Fusarium culmorum*.** *European Journal of Plant Pathology* 1997, **103**(7):589-595.
- Adams GC, Hart LP: **The role of deoxynivalenol and 15-acetyldeoxynivalenol in pathogenesis by *Gibberella zeae* as elucidated through protoplast fusions between toxigenic and non-toxigenic strains.** *Phytopathology* 1989, **79**(4):404-408.
- Walker SL, Leath S, Hagler WM, Murphy JP: **Variation among isolates of *Fusarium graminearum* associated with *Fusarium* head blight in North Carolina.** *Plant Disease* 2001, **85**(4):404-410.
- Simpson DR, Thomsett MA, Nicholson P: **Competitive interactions between *Microdochium nivale* var. majus, *M-nivale* var. *nivale* and *Fusarium culmorum* in *planta* and *in vitro*.** *Environmental Microbiology* 2004, **6**(1):79-87.
- Schmidt-Heydt M, Magan N, Geisen R: **Stress induction of mycotoxin biosynthesis genes by abiotic factors.** *Fems Microbiology Letters* 2008, **284**(2):142-149.
- Gardiner DM, Kazan K, Manners JM: **Nutrient profiling reveals potent inducers of trichothecene biosynthesis in *Fusarium graminearum*.** *Fungal Genetics and Biology* 2009, **46**(8):604-613.
- Gardiner DM, Osborne S, Kazan K, Manners JM: **Low pH regulates the production of deoxynivalenol by *Fusarium graminearum*.** *Microbiology-SGM* 155(9):3149-3156.
- Magan N, Hope R, Collete A, Baxter ES: **Relationship between growth and mycotoxin production by *Fusarium* species, biocides and environment.** *European Journal of Plant Pathology* 108(7):685-690.
- Ponts N, Pinson-Gadais L, Verdal-Bonnin MN, Barreau C, Richard-Forget F: **Accumulation of deoxynivalenol and its 15-acetylated form is significantly modulated by oxidative stress in liquid cultures of *Fusarium graminearum*.** *FEMS Microbiology Letters* 2006, **258**(1):102-107.
- Ochiai N, Tokai T, Takahashi-Ando N, Fujimura M, Kimura M: **Genetically engineered *Fusarium* as a tool to evaluate the effects of environmental factors on initiation of trichothecene biosynthesis.** *FEMS Microbiology Letters* 2007, **275**(1):53-61.
- Ponts N, Pinson-Gadais L, Barreau C, Richard-Forget F, Ouellet T: **Exogenous H₂O₂ and catalase treatments interfere with Tri genes expression in liquid cultures of *Fusarium graminearum*.** *FEBS Letters* 2007, **581**(3):443-447.
- Ponts N, Couedelo L, Pinson-Gadais L, Verdal-Bonnin MN, Barreau C, Richard-Forget F: ***Fusarium* response to oxidative stress by H₂O₂ is trichothecene chemotype-dependent.** *FEMS Microbiology Letters* 2009, **293**(2):255-262.
- Mullenborn C, Steiner U, Ludwig M, Oerke EC: **Effect of fungicides on the complex of *Fusarium* species and saprophytic fungi colonizing wheat kernels.** *European Journal of Plant Pathology* 2008, **120**(2):157-166.
- Ochiai N, Tokai T, Takahashi-Ando N, Fujimura M, Kimura M: **Genetically engineered *Fusarium* as a tool to evaluate the effects of environmental factors on initiation of trichothecene biosynthesis.** *FEMS Microbiology Letters* 275(1):53-61.
- D'Mello JPF, Macdonald AMC, Postel D, Dijkstra WTP, Dujardin A, Placinta CM: **Pesticide use and mycotoxin production in *Fusarium* and *Aspergillus* phytopathogens.** *European Journal of Plant Pathology* 104(8):741-751.
- Covarelli L, Turner AS, Nicholson P: **Repression of deoxynivalenol accumulation and expression of Tri genes in *Fusarium culmorum* by fungicides *in vitro*.** *Plant Pathology* 2004, **53**(1):22-28.
- Matthies A, Buchenauer H: **Effect of tebuconazole (Folicur (R)) and prochloraz (Sportak (R)) treatments on *Fusarium head scab* development, yield and deoxynivalenol (DON) content in grains of wheat following artificial inoculation with *Fusarium culmorum*.** *Zeitschrift für Pflanzenkrankheiten und Pflanzenschutz/Journal of Plant diseases and Protection* 107(1):33-52.
- Kim YS, Dixon EW, Vincelli P, Farman ML: **Field resistance to strobilurin (Q (o)) fungicides in *Pyricularia grisea* caused by mutations in the mitochondrial cytochrome b gene.** *Phytopathology* 2003, **93**(7):891-900.
- Fisher N, Brown AC, Sexton G, Cook A, Windass J, Meunier B: **Modeling the Q(o) site of crop pathogens in *Saccharomyces cerevisiae* cytochrome b.** *European Journal of Biochemistry* 2004, **271**(11):2264-2271.
- Fraaije BA, Butters JA, Coelho JM, Jones DR, Hollomon DW: **Following the dynamics of strobilurin resistance in *Blumeria graminis* f.sp. *tritici* using quantitative allele-specific real-time PCR measurements with the fluorescent dye SYBR Green I.** *Plant Pathology* 2002, **51**(1):45-54.
- Kaneko I, Ishii H: **Effect of azoxystrobin on activities of antioxidant enzymes and alternative oxidase in wheat head blight pathogens *Fusarium graminearum* and *Microdochium nivale*.** *Journal of General Plant Pathology* 2009, **75**(5):388-398.
- Levine A, Tenhaken R, Dixon R, Lamb C: **H₂O₂ from the oxidative burst orchestrates the plant hypersensitive disease resistance response.** *Cell* 1994, **79**(4):583-593.
- Seong KY, Zhao X, Xu JR, Guldener U, Kistler HC: **Conidial germination in the filamentous fungus *Fusarium graminearum*.** *Fungal Genetics and Biology* 2008, **45**(4):389-399.
- Aguirre J, Rios-Momberg M, Hewitt D, Hansberg W: **Reactive oxygen species and development in microbial eukaryotes.** *Trends in Microbiology* 2005, **13**(3):111-118.
- Hansberg W, Aguirre J: **Hyperoxidant states cause microbial cell-differentiation by cell isolation from dioxygen.** *Journal of Theoretical Biology* 1990, **142**(2):201-221.
- Cano-Dominguez N, Alvarez-Delfin K, Hansberg W, Aguirre J: **NADPH oxidases NOX-1 and NOX-2 require the regulatory subunit NOR-1 to control cell differentiation and growth in *Neurospora crassa*.** *Eukaryotic Cell* 2008, **7**(8):1352-1361.
- Branco MR, Marinho HS, Cyrne L, Antunes F: **Decrease of H₂O₂ plasma membrane permeability during adaptation to H₂O₂ in *Saccharomyces cerevisiae*.** *Journal of Biological Chemistry* 2004, **279**(8):6501-6506.
- Sousa-Lopes A, Antunes F, Cyrne L, Marinho HS: **Decreased cellular permeability to H₂O₂ protects *Saccharomyces cerevisiae* cells in stationary phase against oxidative stress.** *FEBS Letters* 2004, **578**(1-2):152-156.
- Shimokawa O, Nakayama H: **Increased sensitivity of *Candida albicans* cells accumulating 14-alpha-methylated sterols to active oxygen: Possible relevance to *in vivo* efficacies of azole antifungal agents.** *Antimicrobial Agents and Chemotherapy* 1992, **36**(8):1626-1629.
- Folmer V, Pedrosa N, Matias AC, Lopes S, Antunes F, Cyrne L, Marinho HS: **H₂O₂ induces rapid biophysical and permeability changes in the plasma membrane of *Saccharomyces cerevisiae*.** *Biochimica Biophysica Acta-Biomembr* 2008, **1778**(4):1141-1147.
- Wu YX, von Tiedemann A: **Impact of fungicides on active oxygen species and antioxidant enzymes in spring barley (*Hordeum vulgare* L.) exposed to ozone.** *Environmental Pollution* 2002, **116**(1):37-47.
- Wu YX, von Tiedemann A: **Physiological effects of azoxystrobin and epoxiconazole on senescence and the oxidative status of wheat.** *Pesticide Biochemistry and Physiology* 2001, **71**(1):1-10.

41. Jansen C, von Wettstein D, Schafer W, Kogel KH, Felk A, Maier FJ: **Infection patterns in barley and wheat spikes inoculated with wild-type and trichodiene synthase gene disrupted *Fusarium graminearum*.** *Proceedings of the National Academy of Sciences of the United States of America* 2005, **102(46)**:16892-16897.
42. Audenaert K, Van Broeck R, Bekaert B, De Witte F, Heremans B, Messens K, Hofte M, Haesaert G: ***Fusarium* head blight (FHB) in Flanders: population diversity, inter-species associations and DON contamination in commercial winter wheat varieties.** *European Journal of Plant Pathology* 2009, **125(3)**:445-458.
43. Saghaimarouf MA, Soliman KM, Jorgensen RA, Allard RW: **Ribosomal DNA spacer-length polymorphisms in barley: Mendelian inheritance, chromosomal location and population dynamics.** *Proceedings of the National Academy of Sciences of the United States of America-Biological Sciences* 1984, **81(24)**:8014-8018.
44. Nicolaisen M, Supronien S, Nielsen LK, Lazzaro I, Spliid NH, Justesen AF: **Real-time PCR for quantification of eleven individual *Fusarium* species in cereals.** *Journal of Microbiological Methods* 2009, **76(3)**:234-240.

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