



## Maternal and Adult Interleukin-17A Exposure and Autism Spectrum Disorder

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Epidemiological evidence in humans has suggested that maternal infections and maternal autoimmune diseases are involved in the pathogenesis of autism spectrum disorder. Animal studies supporting human results have shown that maternal immune activation causes brain and behavioral alterations in offspring. Several underlying mechanisms, including interleukin-17A imbalance, have been identified. Apart from the pro-inflammatory effects of interleukin-17A, there is also evidence to support the idea that it activates neuronal function and defines cognitive behavior. In this review, we examined the signaling pathways in both immunological and neurological contexts that may contribute to the improvement of autism spectrum disorder symptoms associated with maternal blocking of interleukin-17A and adult exposure to interleukin-17A. We first describe the epidemiology of maternal immune activation then focus on molecular signaling of the interleukin-17 family regarding its physiological and pathological roles in the embryonic and adult brain. In the future, it may be possible to use interleukin-17 antibodies to prevent autism spectrum disorder.

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## INTRODUCTION

Prenatal exposure to maternal immune activation (MIA) has been implicated as an environmental risk factor for autism spectrum disorder (ASD). The relationship between MIA and the pathogenesis of neurodevelopmental disorders including ASD has been discussed at length (1–7).

In the first part of this review, we describe the epidemiology of MIA, including maternal infection and maternal autoimmune diseases, as risk factors for ASD. Subsequently, among immunological factors, we focus on molecular signaling of the interleukin (IL)-17 family regarding its physiological and pathological roles in the embryonic and adult brain, based essentially on animal experiments.

## **MIA AND ASD**

Abnormalities in the immune system have been widely observed in the brain and periphery of patients with ASD. Studies have shown that ASD is associated with chronic neuroinflammation, with increased activation of microglia and astrocytes and the production of cytokines and chemokines in the brain (8, 9).

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Infections during pregnancy can cause prematurity or stillbirth, and pathogens can be vertically transmitted to the fetus, causing congenital infections and severe diseases, known as TORCH syndrome (*Toxoplasma gondii*, other, rubella virus, cytomegalovirus, herpes simplex virus) (10, 11). In addition to the threat from these pathogens, other clinical evidence suggests that ASD is increased in the offspring of pregnancies during seasonal outbreaks and epidemics of influenza, measles, epidemic parotitis, and polio (7). Moreover, animal studies have shown that MIA, including viral infection and mimicry, results in neurodevelopmental abnormalities in rodents and non-human primates similar to human ASD phenotypes (3, 12, 13). However, this relationship has not been elucidated, because a meta-analysis showed that the odds ratio (OR) of offspring with ASD is only 1.13 (95% confidence interval [CI] 1.03–1.23) (14).

Furthermore, chronic inflammatory and allergic conditions in pregnancy, such as autoimmune diseases (15) (OR 1.34, 95% CI 1.23–1.46) or asthma (OR 1.43, 95% CI 1.38–1.49) (16), are prominent risk factors for ASD (6). The correlation between asthma and ASD has been well-demonstrated (17, 18). Among autoimmune diseases, maternal psoriasis is also a significant risk factor for ASD (OR 1.39, 95% CI 1.00–1.95) (18). Maternal psoriasis has recently received attention because IL-17A is one of the most important cytokines in the pathogenesis of psoriasis (19, 20).

## **IL-17 SIGNALING**

IL-17A (commonly known as IL-17) is a signature cytokine of a distinct CD4<sup>+</sup> T helper 17 (Th17) cell that is characterized by the expression of retinoic acid receptor-related orphan receptor gamma t (ROR $\gamma$ t) and is activated by IL-23. IL-17A is most strongly implicated in human disease among the six IL-17 family members (IL-17A, IL-17B, IL-17C, IL-17D, IL-25(also known as IL-17E), and IL-17F). As shown in **Figure 1**, all the family members except IL-17D basically function as homodimers; however, IL-17A and IL-17F form a heterodimer (21). The IL-23/IL-17A signaling axis has been found to play a critical role in autoimmune diseases (21, 23, 24).

It has been revealed that IL-17A is produced by other cell populations, such as IL-17-producing CD8<sup>+</sup> T (Tc17) cells,  $\gamma\delta T$  cells, natural killer T cells, natural Th17 cells, natural killer cells, group 3 innate lymphoid cells, neutrophils, and mast cells (21, 23, 24) (**Figure 1**).

The IL-17 receptor family is composed of five members (IL-17RA to IL-17RE), which are distinct subclasses of receptors characterized by an intracellular motif called SEFIR (SEF [similar expression to FGF receptor]/IL-17 receptor) (**Figure 1**) (21, 24). The initial event in IL-17R signaling is the recruitment of Act1, a multifunctional protein containing the SEFIR domain required for IL-17R-Act1 interaction. Act1 has E3 ubiquitin ligase activity and rapidly recruits and ubiquitinates tumor necrosis factor receptor-associated factor 6 (TRAF6), another E3 ubiquitin ligase (**Figure 1**). Like other receptors that recruit TRAF6, IL-17 triggers the activation of the canonical nuclear factor  $\kappa$ B (NF- $\kappa$ B) cascade and pro-inflammatory and anti-microbial genes (21, 24). TRAF6 also promotes the activation of mitogenactivated protein kinase and activator protein 1 (AP1) pathways, and CCAAT/enhancer-binding protein (C/EBP) transcription factors (21, 24).

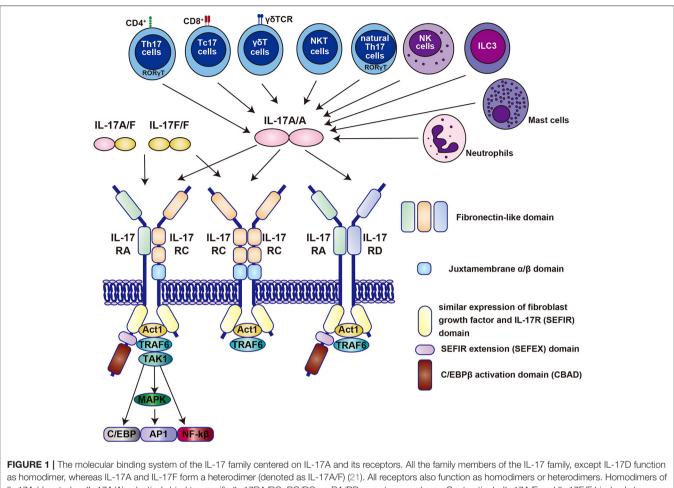
### RELATIONSHIP BETWEEN IL-17 EXPOSURE AND ASD IN THE RODENT EMBRYONIC BRAIN

Little is known about the role of IL-17A in brain development under non-inflammatory conditions. Therefore, we obtained the RNA sequencing results from Mouse Genome Informatics and found that *Il-17a* and *Il-17f* were not or hardly expressed in the mouse embryonic brain (http://www.informatics.jax.org/). In contrast, IL-17 family receptors, *Il-17ra, rc, and rd*, are all expressed in the embryonic and adult mouse brain. These results were confirmed by Choi et al. in 2016, where they demonstrated that IL-17RA is expressed mainly in the cortical plate of the mouse embryonic brain (25).

Accumulating evidence supports a role for Th17 cells and their effector cytokine IL-17A in ASD (26) (Figure 2A). Polyinosinic:polycytidylic acid [poly (I:C)] is structurally similar to double-stranded RNA and is used to model the actions of extracellular double-stranded RNA, such as viral mimicry. Poly (I:C)-induced MIA in the gestating dam is relayed to the embryo via the placenta. Choi et al. (25) showed that increased Th17 cells in the placenta secrete IL-17A, which enters the fetal circulation. Since mouse blood brain barrier begins to form between E11 and E17 (27), circulated IL-17A can enter the brain and regulate development without local production of IL-17A (25). Another group also confirmed the upregulation of IL-17A in maternal blood and the postnatal offspring brain (28). In addition to poly (I:C), Lipopolysaccharide, major component of the outer membrane of Gram-negative bacteria, is also used to induce MIA as a bacterial septic shock model (29).

IL-17A acts directly on the mouse fetal brain on embryonic day (E) 14.5, resulting in an ASD-like phenotype, including abnormal behaviors in ultrasonic vocalization tests, social interaction tests, and marble burying tests (25). Direct injection of IL-17A into the fetal lateral ventricles on E14.5 resulted in phenocopied ASD-like behaviors and cortical disorganization in the offspring induced by poly (I:C)-evoked MIA (25). *Il-17ra* mRNA is detectable in the fetal brain on E14.5 and is upregulated by poly (I:C)-MIA in an IL-17A-dependent manner (25). Direct injection of IL-17A into the fetal brain on E14.5 resulted in thinning of the cortical plate on E18.5, which was not observed in MIA induction on E14.5 (25) (**Figure 2A**). Interestingly, Choi et al. found that poly (I:C)-induced MIA and IL-17A administration to the embryonic brain on E14.5 resulted in patch-like cortical dysplasia on E18.5 (25), which is similar to

Abbreviations: ASD, autism spectrum disorder; CI, confidence interval; CNS, central nervous system; E, embryonic day; EAE, experimental allergic encephalomyelitis; Ig, immunoglobulin; IL, interleukin; MIA, maternal immune activation; OR, odds ratio; S1DZ, primary somatosensory cortex dysgranular zone; SEFIR, similar expression to FGF receptor /IL-17 receptor; SFB, segmented filamentous bacteria; Th17, T helper 17; TRAF6, tumor necrosis factor receptorassociated factor 6.



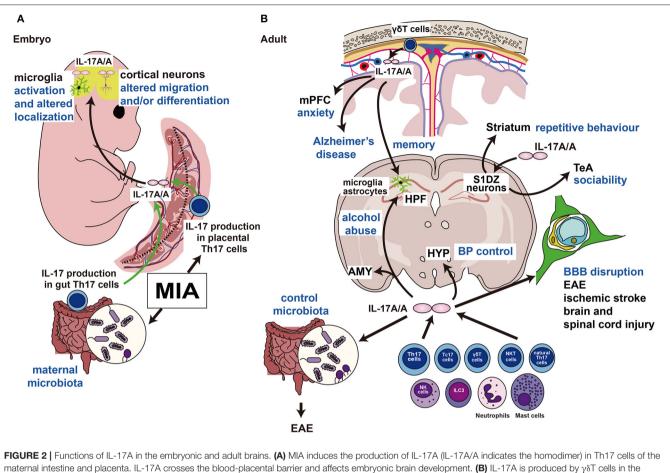
as homodimer, whereas IL-17A and IL-17F form a heterodimer (denoted as IL-17A/F) (21). All receptors also function as homodimers or heterodimers. Homodimers of IL-17A (denoted as IL-17A/A) selectively bind to specific IL-17RA/RC, RC/RC, or RA/RD receptor complexes. Contrastingly, IL-17A/F and IL-17F/F bind only to IL-17RA/RC and RC/RC receptor complexes. Each IL-17 receptor has an extracellular fibronectin-like domain that binds the ligand and an intracellular SEFIR (similar expression of fibroblast growth factor and IL-17R) domain that recruits molecules such as Act1 and TRAF6 (21). The IL-17 receptor family has been shown by Goepfert et al., to be structurally bent between the first and second fibronectin domains (22). IL, interleukin; Th17, T helper 17; Tc17, IL-17-producing CD8<sup>+</sup> T cells; NKT, natural killer T cells; NK; natural killer cells; ILC3, type 3 innate lymphoid cells; NF, nuclear factor; TRAF6, tumor necrosis factor receptor-associated factor 6; MAPK, mitogen-activated protein kinase; TAK1; transforming growth factor-β-activated kinase 1.

some human patients with ASD (30). Their group reproduced the results by another study (31); however, another group mentioned that they could not find any patches after MIA; therefore, the occurrence of cortical patches remains controversial (32).

Kim et al. showed that maternal microbiota, including segmented filamentous bacteria (SFB), promote IL-17A production in maternal gut Th17 cells (33). They treated MIAevoked dams with vancomycin to kill SFB, and this treatment inhibited the ASD-phenotype in offspring, such as abnormal ultrasonic vocalization, repetitive behavior, or sociability, with decreased IL-17A production (**Figure 2A**). More recently, another group showed that the administration of IL-17A during the entire maternal period causes early and persistent cortical abnormalities and ASD-like phenotypes in male offspring (34). The offspring showed abnormal expression of synaptic and cell cycle genes, disrupted adult glia, inhibitory synapses, and abnormal behaviors (34). Moreover, IL-17A injection into the fetal brain on E14.5 resulted in microglial activation and altered localization (35) (**Figure 2A**). In addition, maternal overexpression of IL-17A induced abnormal behavior in offspring, and in parallel, elevated kynurenine levels in maternal serum and fetal plasma were observed. Moreover, maternal kynurenine-injected mice exhibited behavioral abnormalities similar to those observed in the offspring of *Il-17a*-overexpressed dams (36) (**Figure 2A**).

# IL-17A EXPOSURE IN THE ADULT BRAIN OF RODENTS

Contrary to the analysis in the embryonic brain, the expression of IL-17A and its receptors in the adult central nervous system (CNS) has been intensively studied. Das Sarma et al. showed that IL-17RA is expressed in some cultured astrocytes (16.8%) and slightly in microglia (0.80%) (37), and Liu et al. (2014) showed that in the adult dentate gyrus, astrocytes mainly



**PROVE 2** (Protocols of IL-17A in the embryonic and adult brans. (A) MiA indices the production on L-17A (IL-17AA indicates the homodiniter) in Thir 7 cells of the maternal intestine and placenta. IL-17A crosses the blood-placental barrier and affects embryonic brain development. (B) IL-17A is produced by  $\gamma$ 8T cells in the cranial meninges and regulates the mPFC, which controls emotion, and HPF, which regulates memory formation. This pathway may be activated in the Alzheimer's disease model. Direct injection of IL-17A into the S1DZ restores the ASD phenotypes of repetitive behavior or sociability. IL-17A also controls the gut microbiota, and its disruption causes autoimmunity. IL17A is involved in the destruction of the BBB in several neurological disorders. Intravenous injection of IL-17A affects the hypothalamus and alters systemic blood pressure. IL, interleukin; MIA, maternal immune activation; Th17, T helper 17; mPFC; medial prefrontal cortex; HPF, hippocampal formation; S1DZ, primary somatosensory cortex dysgranular zone; ASD, autism spectrum disorder; BBB, blood-brain barrier; HYP, hypothalamus; BP, blood pressure; EAE, experimental allergic encephalomyelitis; To17, IL-17-producing CD8<sup>+</sup> T cells; NKT, natural killer T cells; NK; natural killer cells; ILC3, type 3 innate lymphoid cells; AMY, amygdala; TeA, temporal association area.

express *Il17a* under physiological conditions (38). Their study revealed that IL-17A is a negative regulator of neurogenesis in the adult hippocampus, and *Il17a* knockout enhances synaptic function (38).

In addition to these published results, we obtained the RNA sequencing results from the Human Brain Atlas (https://www.proteinatlas.org/) and Brain RNA-Seq (https://www.brainrnaseq.org/), based on published papers (39, 40). According to these databases, *Il-17a* and *Il-17f* mRNA are rarely expressed in any cell type in the mouse brain; *Il-17ra* mRNA is mainly expressed in macrophages/microglia in small amounts in oligodendrocytes, neurons, and oligodendrocyte precursor cells and is almost absent in astrocytes and endothelial cells. In terms of tissue distribution, a small amount of *Il-17ra* mRNA was observed in the cerebral cortex. Since *Il-17ra* and *rd* mRNA are much more abundant in the pituitary gland, it is necessary to analyze the expression of each isoform of the IL-17 receptor.

Chen et al. used forward genetic methods to show that the Caenorhabditis elegans homolog of Il-17a functions as a neuromodulator in somatosensory neurons (41). Subsequently, Ribeiro et al. showed that IL-17A controls synaptic plasticity and short-term memory (42) (Figure 2B). Intriguingly, IL-17A is secreted by fetal-derived meningeal resident γδT cells and plays an important role in memory formation via glial cell production of brain-derived neurotrophic factor under physiological conditions (42). Furthermore, even under physiological conditions, IL-17A secreted from yoT cells and IL-17RA signaling in neurons of the medial prefrontal cortex controls anxiety-like behaviors, not sociability or memory (43). Alves De Lima et al. also found that the number of meningeal  $\gamma\delta T$  cells increases after birth; therefore, depletion of IL-17A or  $\gamma\delta T$  cells in the postnatal period may affect behavior (43) (Figure 2B).

Reed et al. showed the beneficial effects of IL-17A on social behavior disorders (44) (Figure 2B). They first detected

abnormalities in the neural circuits responsible for repetitive behavior and sociability examined using the marble burying test and social interaction test, respectively (31). The main focus of abnormal circuits in MIA offspring is the primary somatosensory cortex dysgranular zone (S1DZ). Interestingly, using optogenetics, it was reported that S1DZ neurons projecting to the temporal association cortex control sociability, and S1DZ neurons projecting to the striatum regulate repetitive behavior in MIA offspring (31). On the other hand, lipopolysaccharide administration can restore social behavioral deficits in MIAexposed offspring. More interestingly, direct IL-17A delivery into the S1DZ can also restore disturbed social behavior even in monogenic ASD mouse models such as Cntnap2 or Fmr1 mutant mice (44). The authors concluded that the production of IL-17A during inflammation can ameliorate the expression of social behavior deficits by directly affecting neural activity in the brain (44).

In addition to ASD, IL-17A signaling has received strong attention for its pathophysiological functions in various neurological disorders (45–48) (**Figure 2B**). In particular, the importance of IL-17A has been strongly demonstrated in experimental allergic encephalomyelitis (EAE), a model of multiple sclerosis (45, 47, 48). In a recent study, *Il-17a/f*-deficient mice lost sensitivity to EAE, which correlates with changes in the gut microbiota (49). Another important aspect of IL-17A is the regulation of blood-brain barrier functions (50–52) (**Figure 2B**). Furthermore, the involvement of IL-17A signaling has been revealed in various experimental models of ischemic brain injury (53), traumatic brain injury (54), and spinal cord injury (55, 56) (**Figure 2B**).

In addition to the immunological disorders described above, emerging evidence suggests that IL-17A secreted by meningeal  $\gamma\delta T$  cells regulates the pathogenesis of Alzheimer's disease (57, 58), IL-17A secreted by Th17 cells is involved in alcohol abuse (59), and IL-17A regulates blood pressure via the activation of paraventricular nucleus neurons (60) (**Figure 2B**).

### RELATIONSHIP BETWEEN IL-17A EXPOSURE IN THE HUMAN BRAIN AND ASD

It has been reported that neurons, glia, and endothelial cells in the human cortex express receptors for IL-17 (61). However, no information on the specific expression of the IL-17 receptor isoform in the human brain has been reported. Therefore, we obtained the RNA sequencing results from the Human Protein Atlas (https://www.proteinatlas.org/) and Brain RNA-Seq (https://www.brainrnaseq.org/), based on a published paper (62). We found that *IL17A* is rarely expressed in the human brain as revealed by both databases. On the other hand, receptors for the IL-17 family, *IL-17RA* and *RC*, are both expressed in embryonic and adult brains as examined by both databases.

In some patients with ASD, IL-17A has been found at high levels in the blood and correlates with the severity of behavioral symptoms (63, 64). A genome-wide association study showed that copy number variation of the *IL17A* gene is a risk factor

for ASD (65). However, the evidence indicated an indirect correlation. Therefore, to show a direct causal relationship between maternal IL-17A exposure and ASD, we propose the following clinical investigation.

First, psoriasis has received much attention in recent years, since maternal psoriasis is also a significant risk factor for ASD (18), and IL-17A is one of the most important cytokines in the pathogenesis of psoriasis (19, 20). Therefore, maternal psoriasis is a candidate disease to be investigated (**Figure 3**).

To modulate IL-17A signaling, three commercially available antibodies are currently available to treat humans: secukinumab (human monoclonal antibody to IL-17A, immunoglobulin [Ig]G1), ixekizumab (humanized monoclonal antibody to IL-17A, IgG4), and brodalumab (human monoclonal antibody to the IL-17 receptor, IgG2). Both secukinumab and ixekizumab are approved for psoriasis, psoriatic arthritis, and ankylosing spondylitis; brodalumab is only approved for the treatment of psoriasis (66). All subclasses of IgG (IgG1–IgG4) cross the human placenta (67), therefore, all candidate antibodies can block the abnormal upregulation of IL-17 signaling in the fetus. Since the human blood brain barrier also begins to form during pregnancy as well as in mice (68), candidate antibodies may enter the fetal brain after angiogenesis. Clinically, no complication with prenatal usage of secukinumab was reported (69).

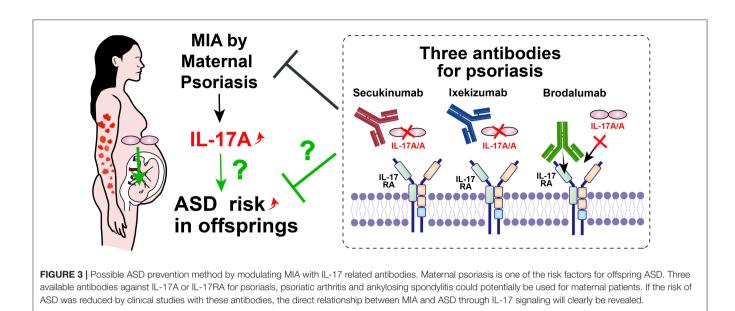
If the incidence of ASD is reduced in the offspring of pregnant patients with psoriasis treated with IL-17-related antibodies, this would indicate a direct causal relationship between MIA and ASD via IL-17 signaling (**Figure 3**).

Lastly, considering the current pandemic situation, I can't avoid mentioning the topic of COVID-19. To the best our knowledge, no evidence that COVID-19 in pregnant mother could be the risk of ASD in offspring has been reported. However, some reports showed that IL-17A is involved in pathophysiology of COVID-19 infection (70), therefore long term observation will elucidate whether maternal COVID-19 infection may impact fetal brain development (71).

### DISCUSSION

The pathophysiological mechanism of ASD or brain development caused by MIA or IL-17A exposure remains to be addressed. As mentioned in section Relationship Between IL-17A Exposure in the Human Brain and ASD, in humans, there is a lack of direct causal relationship between IL-17A and ASD.

Even in experimental animals, the following questions remain elusive. First, MIA has been shown to cause abnormalities in fetal brain development, including unexplained cortical dysgenesis. Wong et al. found that adult offspring exposed to MIA on E14.5 had significantly reduced numbers of either TBR1<sup>+</sup> or SATB2<sup>+</sup> cells in the cortex with cortical patches (26) [see also Shin Yim et al. (31)]. During cortical development, exposure to MIA and IL-17A transiently delays the production of SATB2<sup>+</sup> cells on E14.5 and alters cortical neurogenesis or radial migration only at the medial area with cortical patch formation without changing cortical thickness. Furthermore, IL-17RA is only expressed in cortical plates, and the cell type is unknown (26). These phenotypes cannot



be explained by abnormalities in neurogenesis or radial migration of the entire radial glia. Another study suggested that microglia may alter the neurogenesis of radial glia or neural migration, as IL-17A injection induces microglia to migrate closer to the lateral ventricles (35). In support of this idea, microglia, but not neurons or other glial types, express the highest amount of Il-17ra in the adult stages as examined by databases. Second, it has been suggested that MIA can affect brain development into adulthood with altered systemic immunological responses (30, 72). It has long been unclear why these effects persist, but recent evidence might answer this question. Lim et al. infected pregnant mice with Yersinia pseudotuberculosis. Although the infection was restricted to the dam, the offspring surprisingly harbored more intestinal Th17 cells into adulthood via IL-6 signaling (73). Maternal IL-6 induced immediate and long-term effects based on changes in the epigenetic memory of fetal intestinal epithelial stem cells. Therefore, an enhanced response to the microbiota is trained during pregnancy, and the immune response system is already altered at birth (73).

In this review, we summarize how IL-17A affects brain development and adult brain function mostly based on the animal experiments. In the near future, it may be possible to use IL-17A related antibodies to prevent ASD. However, the

### REFERENCES

- Knuesel I, Chicha L, Britschgi M, Schobel SA, Bodmer M, Hellings JA, et al. Maternal immune activation and abnormal brain development across CNS disorders. *Nat Rev Neurol.* (2014) 10:643–60. doi: 10.1038/nrneurol.2014.187
- Estes ML, Mcallister AK. Maternal immune activation: Implications for neuropsychiatric disorders. *Science*. (2016) 353:772–7. doi: 10.1126/science.aag3194
- Careaga M, Murai T, Bauman MD. Maternal immune activation and autism spectrum disorder: from rodents to nonhuman and human primates. *Biol Psychiatry*. (2017) 81:391–401. doi: 10.1016/j.biopsych.2016.10.020

involvement of IL-17A signaling has not been elucidated yet. Future clinical studies will help to answer this question.

## **AUTHOR CONTRIBUTIONS**

MF: conceptualization and writing—original draft preparation. HM, YO, and XL: writing—review, editing and visualization. All authors contributed to the article and approved the submitted version.

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- Meltzer A, Van De Water J. The role of the immune system in autism spectrum disorder. *Neuropsychopharmacology*. (2017) 42:284–98. doi: 10.1038/npp.2016.158
- Bilbo SD, Block CL, Bolton JL, Hanamsagar R, Tran PK. Beyond infection—maternal immune activation by environmental factors, microglial development, and relevance for autism spectrum disorders. *Exp Neurol.* (2018) 299:241–51. doi: 10.1016/j.expneurol.2017. 07.002
- Han VX, Patel S, Jones HF, Dale RC. Maternal immune activation and neuroinflammation in human neurodevelopmental disorders. *Nat Rev Neurol.* (2021) 17:564–79. doi: 10.1038/s41582-021-00530-8

- Han VX, Patel S, Jones HF, Nielsen TC, Mohammad SS, Hofer MJ, et al. Maternal acute and chronic inflammation in pregnancy is associated with common neurodevelopmental disorders: a systematic review. *Transl Psychiatry*. (2021) 11:71. doi: 10.1038/s41398-021-01198-w
- Matta SM, Hill-Yardin EL, Crack PJ. The influence of neuroinflammation in autism spectrum disorder. *Brain Behav Immun.* (2019) 79:75–90. doi: 10.1016/j.bbi.2019.04.037
- Liao X, Liu Y, Fu X, Li Y. Postmortem studies of neuroinflammation in autism spectrum disorder: A systematic review. *Mol Neurobiol.* (2020) 57:3424– 38. doi: 10.1007/s12035-020-01976-5
- Coyne CB, Lazear HM. Zika virus— reigniting the TORCH. Nat Rev Microbiol. (2016) 14:707–15. doi: 10.1038/nrmicro.2016.125
- Megli CJ, Coyne CB. Infections at the maternal-fetal interface: an overview of pathogenesis and defence. *Nat Rev Microbiol.* (2021) 20:67– 82. doi: 10.1038/s41579-021-00610-y
- Bergdolt L, Dunaevsky A. Brain changes in a maternal immune activation model of neurodevelopmental brain disorders. *Prog Neurobiol.* (2019) 175:1– 19. doi: 10.1016/j.pneurobio.2018.12.002
- Zawadzka A, Cieślik M, Adamczyk A. The role of maternal immune activation in the pathogenesis of autism: a review of the evidence, proposed mechanisms and implications for treatment. *Int J Mol Sci.* (2021) 22:11516. doi: 10.3390/ijms222111516
- 14. Jiang HY, Xu LL, Shao L, Xia RM Yu ZH, Ling ZX, Yang F, et al. Maternal infection during pregnancy and risk of autism spectrum disorders: a systematic review and meta-analysis. *Brain Behav Immun.* (2016) 58:165– 72. doi: 10.1016/j.bbi.2016.06.005
- Chen SW, Zhong XS, Jiang LN, Zheng XY, Xiong YQ, Ma SJ, et al. Maternal autoimmune diseases and the risk of autism spectrum disorders in offspring: a systematic review and meta-analysis. *Behav Brain Res.* (2016) 296:61– 9. doi: 10.1016/j.bbr.2015.08.035
- Gong T, Lundholm C, Rejnö G, Bölte S, Larsson H, D'Onofrio BM, et al. Parental asthma and risk of autism spectrum disorder in offspring: a population and family-based case-control study. *Clin Exp Allergy.* (2019) 49:883–91. doi: 10.1111/cea.13353
- Croen LA, Grether JK, Yoshida CK, Odouli R, Van De Water J. Maternal autoimmune diseases, asthma and allergies, and childhood autism spectrum disorders: a case-control study. *Arch Pediatr Adolesc Med.* (2005) 159:151– 7. doi: 10.1001/archpedi.159.2.151
- Croen LA, Qian Y, Ashwood P, Daniels JL, Fallin D, Schendel D, et al. Family history of immune conditions and autism spectrum and developmental disorders: findings from the study to explore early development. *Autism Res.* (2019) 12:123–35. doi: 10.1002/aur.1979
- Brembilla NC, Senra L, Boehncke WH. The IL-17 family of cytokines in psoriasis: Il-17A and beyond. *Front Immunol.* (2018) 9:1682. doi: 10.3389/fimmu.2018.01682
- Rendon A, Schäkel K. Psoriasis pathogenesis and treatment. Int J Mol Sci. (2019) 20:1475. doi: 10.3390/ijms20061475
- Chung SH, Ye XQ, Iwakura Y. Interleukin-17 family members in health and disease. Int Immunol. (2021) 33:723–9. doi: 10.1093/intimm/dxab075
- 22. Goepfert A, Lehmann S, Blank J, Kolbinger F, Rondeau JM. Structural analysis reveals that the cytokine il-17f forms a homodimeric complex with receptor il-17rc to drive il-17ra-independent signaling. *Immunity.* (2020) 52:499–512.e5. doi: 10.1016/j.immuni.2020.02.004
- Cua DJ, Tato CM. Innate IL-17-producing cells: the sentinels of the immune system. Nat Rev Immunol. (2010) 10:479–89. doi: 10.1038/nri2800
- McGeachy MJ, Cua DJ, Gaffen SL. The IL-17 family of cytokines in health and disease. *Immunity*. (2019) 50:892–906. doi: 10.1016/j.immuni.2019.03.021
- 25. Choi GB, Yim YS, Wong H, Kim S, Kim H, Kim SV, et al. The maternal interleukin-17a pathway in mice promotes autism-like phenotypes in offspring. *Science*. (2016) 351:933–9. doi: 10.1126/science.aad0314
- Wong H, Hoeffer C. Maternal IL-17A in autism. *Exp Neurol.* (2018) 299:228–40. doi: 10.1016/j.expneurol.2017.04.010
- 27. Abbott NJ, Patabendige AAK, Dolman DEM, Yusof SR, Begley DJ. Structure and function of the blood-brain barrier. Neurobiol Dis. (2010) 37:13-25. doi: 10.1016/j.nbd.2009. 07.030
- 28. Garay PA, Hsiao EY, Patterson PH, McAllister AK. Maternal immune activation causes age- and region-specific changes in brain cytokines in

offspring throughout development. Brain Behav Immun. (2013) 31:54-68. doi: 10.1016/j.bbi.2012.07.008

- Yasumatsu K, Nagao JI, Arita-Morioka KI, Narita Y, Tasaki S, Toyoda K, et al. Bacterial-induced maternal interleukin-17A pathway promotes autistic-like behaviors in mouse offspring. *Exp Anim.* (2020) 69:250–60. doi: 10.1538/expanim.19-0156
- Stoner R, Chow ML, Boyle MP, Sunkin SM, Mouton PR, Roy S, et al. Patches of disorganization in the neocortex of children with autism. N Engl J Med. (2014) 370:1209–19. doi: 10.1056/NEJMoa1307491
- Shin Yim Y, Park A, Berrios J, Lafourcade M, Pascual LM, Soares N, et al. Reversing behavioural abnormalities in mice exposed to maternal inflammation. *Nature*. (2017) 549:482–7. doi: 10.1038/nature23909
- Canales CP, Estes ML, Cichewicz K, Angara K, Aboubechara JP, Cameron S, et al. Sequential perturbations to mouse corticogenesis following in utero maternal immune activation. *Elife*. (2021) 10:e60100. doi: 10.7554/eLife.60100
- Kim S, Kim H, Yim YS, Ha S, Atarashi K, Tan TG, et al. Maternal gut bacteria promote neurodevelopmental abnormalities in mouse offspring. *Nature*. (2017) 549:528–32. doi: 10.1038/nature23910
- Gumusoglu SB, Hing BWQ, Chilukuri ASS, Dewitt JJ, Scroggins SM, Stevens HE. Chronic maternal interleukin-17 and autism-related cortical gene expression, neurobiology, and behavior. *Neuropsychopharmacology.* (2020) 45:1008–17. doi: 10.1038/s41386-020-0640-0
- Sasaki T, Tome S, Takei Y. Intraventricular IL-17A administration activates microglia and alters their localization in the mouse embryo cerebral cortex. *Mol Brain.* (2020) 13:93. doi: 10.1186/s13041-020-00635-z
- 36. Murakami Y, Imamura Y, Kasahara Y, Yoshida C, Momono Y, Fang K, et al. The effects of maternal interleukin-17A on social behavior, cognitive function, and depression-like behavior in mice with altered kynurenine metabolites. *Int J Tryptophan Res.* (2021) 14:11786469211026639. doi: 10.1177/11786469211026639
- 37. Das Sarma J, Ciric B, Marek R, Sadhukhan S, Caruso ML, Shafagh J, et al. Functional interleukin-17 receptor A is expressed in central nervous system glia and upregulated in experimental autoimmune encephalomyelitis. *J Neuroinflammation*. (2009) 6:14. doi: 10.1186/1742-2094-6-14
- Liu Q, Xin W, He P, Turner D, Yin J, Gan Y, et al. Interleukin-17 inhibits adult hippocampal neurogenesis. Sci Rep. (2014) 4:7554. doi: 10.1038/srep07554
- Sjöstedt E, Zhong W, Fagerberg L, Karlsson M, Mitsios N, Adori C, et al. An atlas of the protein-coding genes in the human, pig, and mouse brain. *Science*. (2020) 367:eaay5947. doi: 10.1126/science.aay5947
- Zhang Y, Chen K, Sloan SA, Bennett ML, Scholze AR, O'Keeffe S, et al. An RNA-sequencing transcriptome and splicing database of glia, neurons, and vascular cells of the cerebral cortex. J Neurosci. (2014) 34:11929– 47. doi: 10.1523/JNEUROSCI.1860-14.2014
- Chen C, Itakura E, Nelson GM, Sheng M, Laurent P, Fenk LA, et al. IL-17 is a neuromodulator of Caenorhabditis elegans sensory responses. *Nature*. (2017) 542:43–8. doi: 10.1038/nature20818
- Ribeiro M, Brigas HC, Temido-Ferreira M, Pousinha PA, Regen T, Santa C, et al. Meningeal γδ T cell-derived IL-17 controls synaptic plasticity and short-term memory. *Sci Immunol.* (2019) 4:eaay5199. doi: 10.1126/sciimmunol.aay5199
- Alves De Lima K, Rustenhoven J, Da Mesquita S, Wall M, Salvador AF, Smirnov I, et al. Meningeal γδT cells regulate anxiety-like behavior via IL-17a signaling in neurons. *Nat Immunol.* (2020) 21:1421–9. doi: 10.1038/s41590-020-0776-4
- Reed MD, Yim YS, Wimmer RD, Kim H, Ryu C, Welch GM, et al. IL-17a promotes sociability in mouse models of neurodevelopmental disorders. *Nature*. (2020) 577:249–53. doi: 10.1038/s41586-019-1843-6
- Waisman A, Hauptmann J, Regen T. The role of IL-17. Acta Neuropathol. (2015) 129:625–37. doi: 10.1007/s00401-015-1402-7
- Storelli E, Cassina N, Rasini E, Marino F, Cosentino M. Do Th17 lymphocytes and IL-17 contribute to Parkinson's disease? A systematic review of available evidence. *Front Neurol.* (2019) 10:13. doi: 10.3389/fneur.2019.00013
- Milovanovic J, Arsenijevic A, Stojanovic B, Kanjevac T, Arsenijevic D, Radosavljevic G, et al. Interleukin-17 in chronic inflammatory neurological diseases. *Front Immunol.* (2020) 11:947. doi: 10.3389/fimmu.2020.00947
- Wo J, Zhang F, Li Z, Sun C, Zhang W, Sun G. The role of gamma-delta T cells in diseases of the central nervous system. *Front Immunol.* (2020) 11:580304. doi: 10.3389/fimmu.2020.580304

- Regen T, Isaac S, Amorim A, Núñez NG, Hauptmann J, Shanmugavadivu A, et al. IL-17 controls central nervous system autoimmunity through the intestinal microbiome. *Sci Immunol.* (2021) 6:eaaz6563. doi: 10.1126/sciimmunol.aaz6563
- Huppert J, Closhen D, Croxford A, White R, Kulig P, Pietrowski E, et al. Cellular mechanisms of IL-17-induced blood-brain barrier disruption. *FASEB J.* (2010) 24:1023–34. doi: 10.1096/fj.09-141978
- Ni P, Dong H, Wang Y, Zhou Q, Xu M, Qian Y, et al. IL-17A contributes to perioperative neurocognitive disorders through bloodbrain barrier disruption in aged mice. *J Neuroinflammation.* (2018) 15:332. doi: 10.1186/s12974-018-1374-3
- 52. Setiadi AF, Abbas AR, Jeet S, Wong K, Bischof A, Peng I, et al. IL-17A is associated with the breakdown of the blood-brain barrier in relapsing-remitting multiple sclerosis. *J Neuroimmunol.* (2019) 332:147– 54. doi: 10.1016/j.jneuroim.2019.04.011
- Shichita T, Sugiyama Y, Ooboshi H, Sugimori H, Nakagawa R, Takada I, et al. Pivotal role of cerebral interleukin-17-producing gammadeltaT cells in the delayed phase of ischemic brain injury. *Nat Med.* (2009) 15:946– 50. doi: 10.1038/nm.1999
- Tobin RP, Mukherjee S, Kain JM, Rogers SK, Henderson SK, Motal HL, et al. Traumatic brain injury causes selective, CD74-dependent peripheral lymphocyte activation that exacerbates neurodegeneration. *Acta Neuropathol Commun.* (2014) 2:143. doi: 10.1186/s40478-014-0143-5
- Sun G, Yang S, Cao G, Wang Q, Hao J, Wen Q, et al. γδ T cells provide the early source of IFN-γ to aggravate lesions in spinal cord injury. J Exp Med. (2018) 215:521–35. doi: 10.1084/jem.20170686
- Miyajima H, Itokazu T, Tanabe S, Yamashita T. Interleukin-17A regulates ependymal cell proliferation and functional recovery after spinal cord injury in mice. *Cell Death Dis.* (2021) 12:766. doi: 10.1038/s41419-021-04064-1
- Cristiano C, Volpicelli F, Lippiello P, Buono B, Raucci F, Piccolo M, et al. Neutralization of IL-17 rescues amyloid-beta-induced neuroinflammation and memory impairment. *Br J Pharmacol.* (2019) 176:3544–57. doi: 10.1111/bph.14586
- Brigas HC, Ribeiro M, Coelho JE, Gomes R, Gomez-Murcia V, Carvalho K, et al. IL-17 triggers the onset of cognitive and synaptic deficits in early stages of Alzheimer's disease. *Cell Rep.* (2021) 36:109574. doi: 10.1016/j.celrep.2021.109574
- Xu J, Ma HY, Liu X, Rosenthal S, Baglieri J, Mccubbin R, et al. Blockade of IL-17 signaling reverses alcohol-induced liver injury and excessive alcohol drinking in mice. JCI Insight. (2020) 5:e131277. doi: 10.1172/jci.insight.131277
- Cao Y, Yu Y, Xue B, Wang Y, Chen X, Beltz TG, et al. IL (interleukin)-17A acts in the brain to drive neuroinflammation, sympathetic activation, and hypertension. *Hypertension*. (2021) 78:1450–62. doi: 10.1161/HYPERTENSIONAHA.121.18219
- He JJ Li S, Shu HF Yu SX, Liu SY, Yin Q, et al. The interleukin 17 system in cortical lesions in focal cortical dysplasia. J Neuropathol Exp Neurol. (2013) 72:152–63. doi: 10.1097/NEN.0b013e31828 1262e
- 62. Zhang Y, Sloan SA, Clarke LE, Caneda C, Plaza CA, Blumenthal PD, et al. Purification and characterization of progenitor and mature human astrocytes reveals transcriptional and functional differences with mouse. *Neuron.* (2016) 89:37–53. doi: 10.1016/j.neuron.2015. 11.013

- Al-Ayadhi LY, Mostafa GA. Elevated serum levels of interleukin-17A in children with autism. J Neuroinflammation. (2012) 9:158. doi: 10.1186/1742-2094-9-158
- 64. Akintunde ME, Rose M, Krakowiak P, Heuer L, Ashwood P, Hansen R, et al. Increased production of IL-17 in children with autism spectrum disorders and co-morbid asthma. J Neuroimmunol. (2015) 286:33–41. doi: 10.1016/j.jneuroim.2015.07.003
- 65. Van Der Zwaag B, Franke L, Poot M, Hochstenbach R, Spierenburg HA, Vorstman JA, et al. Gene-network analysis identifies susceptibility genes related to glycobiology in autism. *PLoS ONE.* (2009) 4:e5324. doi: 10.1371/journal.pone.0005324
- Pacha O, Sallman MA, Evans SE. COVID-19: a case for IL-17 inhibition. Nat Rev Immunol. (2020) 20:345–6. doi: 10.1038/s41577-020-0328-z
- Simister NE. Placental transport of immunoglobulin G. Vaccine. (2003) 21:3365–9. doi: 10.1016/S0264-410X(03)00334-7
- Saili KS, Zurlinden TJ, Schwab AJ, Silvin A, Baker NC, Hunter ES, et al. Bloodbrain barrier development: systems modeling and predictive toxicology. *Birth Defects Res.* (2017) 109:1680–710. doi: 10.1002/bdr2.1180
- Warren RB, Reich K, Langley RG, Strober B, Gladman D, Deodhar A, et al. Secukinumab in pregnancy: outcomes in psoriasis, psoriatic arthritis and ankylosing spondylitis from the global safety database. *Br J Dermatol.* (2018) 179:1205–7. doi: 10.1111/bjd.16901
- Maione F, Casillo GM, Raucci F, Salvatore C, Ambrosini G, Costa L, et al. Interleukin-17A (IL-17A): a silent amplifier of COVID-19. *Biomed Pharmacother*. (2021) 142:111980. doi: 10.1016/j.biopha.2021.111980
- Joma M, Fovet CM, Seddiki N, Gressens P, Laforge M. COVID-19 and pregnancy: Vertical transmission and inflammation impact on newborns. *Vaccines (Basel)*. (2021) 9:391. doi: 10.3390/vaccines9040391
- Hsiao EY, Mcbride SW, Chow J, Mazmanian SK, Patterson PH. Modeling an autism risk factor in mice leads to permanent immune dysregulation. *Proc Natl Acad Sci U S A.* (2012) 109:12776–81. doi: 10.1073/pnas.1202556109
- Lim AI, Mcfadden T, Link VM, Han SJ, Karlsson RM, Stacy A, et al. Prenatal maternal infection promotes tissue-specific immunity and inflammation in offspring. *Science*. (2021) 373:eabf3002. doi: 10.1126/science.abf3002

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