

METTL14 Gene Polymorphisms Confer Neuroblastoma Susceptibility: An Eight-Center Case-Control Study

Zhenjian Zhuo,^{1,11} Hongting Lu,^{2,11} Jinhong Zhu,^{1,3,11} Rui-Xi Hua,¹ Yong Li,⁴ Zhonghua Yang,⁵ Jiao Zhang,⁶ Jiwen Cheng,⁷ Haixia Zhou,⁸ Suhong Li,⁹ Li Li,¹⁰ Huimin Xia,¹ and Jing He¹

¹Department of Pediatric Surgery, Guangzhou Institute of Pediatrics, Guangdong Provincial Key Laboratory of Research in Structural Birth Defect Disease, Guangzhou Women and Children's Medical Center, Guangzhou Medical University, Guangzhou 510623, Guangdong, China; ²Department of Pediatric Surgery, The Affiliated Hospital of Qingdao University, Qingdao 266000, Shandong, China; ³Department of Clinical Laboratory, Biobank, Harbin Medical University Cancer Hospital, Harbin 150040, Heilongjiang, China; ⁴Department of Pediatric Surgery, Hunan Children's Hospital, Changsha 410004, Hunan, China; ⁵Department of Pediatric Surgery, Shengjing Hospital of China Medical University, Shenyang 110004, Liaoning, China; ⁶Department of Pediatric Surgery, The First Affiliated Hospital of Zhengzhou University, Zhengzhou 450052, Henan, China; ⁷Department of Pediatric Surgery, The Second Affiliated Hospital of Xi'an Jiaotong University, Xi'an 710004, Shaanxi, China; ⁸Department of Hematology, The Second Affiliated Hospital and Yuying Children's Hospital of Wenzhou Medical University, Wenzhou 325027, Zhejiang, China; ⁹Department of Pathology, Children Hospital and Women Health Center of Shanxi, Taiyuan 030013, Shannxi, China; ¹⁰Kunming Key Laboratory of Children's Major Disease Research, Yunnan Institute of Pediatrics Research, Yunnan Medical Center for Pediatric Diseases, Kunming Children's Hospital, Kunming 650228, Yunnan, China

Neuroblastoma is the primary cause of cancer death in childhood. METTL14 is tightly linked to cancer. However, whether single-nucleotide polymorphisms (SNPs) in the METTL14 gene could predispose to neuroblastoma susceptibility lacks evidence. With an epidemiology case-control study, associations between METTL14 gene SNPs and overall risk for neuroblastoma were estimated in 898 cases and 1,734 controls. Following that, stratified analysis was performed. Among the five analyzed SNPs, rs298982 G>A and rs62328061 A>G exhibited a significant association with decreased susceptibility to neuroblastoma, whereas the associations with increased neuroblastoma susceptibility were observed for rs9884978 G>A and rs4834698 T>C. Moreover, subjects carrying two to five risk genotypes were more inclined to develop neuroblastoma than those with zero to one risk genotypes. The stratified analysis further demonstrated the protective effect of rs298982 G>A and rs62328061 A>G, as well as the predisposing effect of rs4834698 T>C and two to five risk genotypes, in certain subgroups. Haplotype analysis was performed. Moreover, falsepositive report probability analysis validated the reliability of the significant results. The expression quantitative trait locus analysis revealed that rs298982 is correlated with the expression levels of its surrounding genes. Our results suggest that some SNPs in the METTL14 gene are associated with predisposition to neuroblastoma.

INTRODUCTION

million among children less than 15 years old in developed countries.² In China, neuroblastoma represents approximately 10% of childhood tumors, with an incidence of approximately 7.7 cases per 1 million children.³ Neuroblastoma is the primary cause of cancer death in childhood, accounting for approximately 15% of all pediatric cancer mortality.^{4,5} Normally, neural crest cell precursors migrate from the dorsal neural tube and then start to differentiate upon reaching their appropriate locations in the sympathetic nervous system. However, neuroblastoma may occur sometimes because of the defects in migration, maturation, or differentiation of neural crest cells.^{6,7} So far, the long-term survival of high-risk neuroblastoma is less than 40%. What is worse, this type of neuroblastoma represents about 50% of all of the newly diagnosed patients.^{8,9}

Remarkable advancement has been achieved in the ever-greater understanding of the genetic etiology of neuroblastoma.^{10–12} Mutations in the *PHOX2B*¹³ and *ALK*^{14,15} genes frequently predispose children to familial neuroblastoma. The introduction of single-nucleotide

E-mail: xia-huimin@foxmail.com

Neuroblastoma is predominantly a cancer of infants and young children, because most of the affected children were diagnosed at under five years of age.¹ The incidence of neuroblastoma is about 11–13 in 1

Received 16 May 2020; accepted 7 August 2020; https://doi.org/10.1016/j.omtn.2020.08.009.

¹¹These authors contributed equally to this work.

Correspondence: Jing He, Department of Pediatric Surgery, Guangzhou Institute of Pediatrics, Guangdong Provincial Key Laboratory of Research in Structural Birth Defect Disease, Guangzhou Women and Children's Medical Center, Guangzhou Medical University, 9 Jinsui Road, Guangzhou 510623, Guangdong, China. **E-mail:** hejing198374@gmail.com

Correspondence: Huimin Xia, Department of Pediatric Surgery, Guangzhou Institute of Pediatrics, Guangdong Provincial Key Laboratory of Research in Structural Birth Defect Disease, Guangzhou Women and Children's Medical Center, Guangzhou Medical University, 9 Jinsui Road, Guangzhou 510623, Guangdong, China.

polymorphism (SNP) array technology in the 1990s¹⁶ enabled genome-wide association studies (GWASs) in the late 2000s.¹⁷ In 2008, the first GWAS regarding neuroblastoma was performed, with 1,752 European American neuroblastoma cases and 4,171 cancer-free controls.¹⁸ This study identified that a chromosome 6p22 locus was strongly associated with neuroblastoma predisposition, specifically aggressive subtype. To date, plenty of neuroblastoma susceptibility SNPs have been identified by GWASs.^{19–23} However, so far all of the genetic variations have not been sufficient to fully understand the genetic landscape of neuroblastoma. Therefore, a better understanding of the genetic alterations that shape neuroblastoma risk becomes necessary to clarify its etiology. Ongoing research identifying novel genetic variations may lead to new approaches to diagnosis, prevention, and treatment of neuroblastoma.

N6-methyladenosine (m⁶A) refers to methylation at the sixth position of N on adenosine, which is the most prevalent RNA posttranscriptional modification in eukaryotic organisms, especially in messenger RNA (mRNA).²⁴ Key regulators of m⁶A modification mainly fall into three categories: RNA methyltransferases (METTL3, METTL14, and WTAP, known as "writers"), demethylases (FTO and ALKBH5, known as "erasers"), and m⁶A-binding proteins (YTHDF1/2/3 and IGF2BP1, known as "readers").^{25,26} Strong evidence suggests that aberrant expression of m⁶A key regulators could cause dysregulated RNA m⁶A modification, resulting in a variety of diseases, including cancers.²⁷⁻²⁹ Some SNPs located in specific regions of m⁶A key regulator genes may alter m⁶A methylation and subsequently affect related biological processes.³⁰ A large number of m⁶A gene SNPs have been identified to associate with major depressive disorder,³¹ osteoporosis,³² and obesity.33 However, evidence regarding m6A modification key regulator genes SNPs and the risk of cancer is very scarce.

To identify novel susceptibility m^6A modification key regulator gene *METTL14* SNPs to neuroblastoma, we performed this multi-center epidemiology study. We addressed the associations between the *METTL14* gene SNPs and neuroblastoma risk in a Chinese population of children.

RESULTS

Effect of METTL14 Gene SNPs on Neuroblastoma Risk

We successfully genotyped the five *METTL14* gene SNPs (rs1064034 T>A, rs298982 G>A, rs62328061 A>G, rs9884978 G>A, and *rs4834698* T>C) in 896 cases and 1,733 controls. The associations of these SNPs with neuroblastoma risk are shown in Table 1. The actual SNP genotype frequencies were in accordance with Hardy-Weinberg equilibrium (HWE) in controls (HWE p > 0.05), except for *rs9884978* G>A. In the single-locus analysis, carriers of the rs298982 (AA versus GG: adjusted odds ratio [OR] = 0.40, 95% confidence interval [CI] = 0.20–0.83, p = 0.014; AA versus GG/GA: adjusted OR = 0.40, 95% CI = 0.20–0.83, p = 0.014) or rs62328061 (AG versus AA: adjusted OR = 0.77, 95% CI = 0.64–0.93, p = 0.007; AG/GG versus AA: adjusted OR = 0.82, 95% CI = 0.68–0.97, p = 0.024) variant alleles showed decreased susceptibility to neuroblastoma. On the contrary, the rs9884978 (AA versus GG/GA:

adjusted OR = 1.39, 95% CI = 1.00–1.94, p = 0.048) and rs4834698 (CC versus TT: adjusted OR = 1.28, 95% CI = 1.02–1.61, p = 0.034) variant alleles contribute to increased susceptibility to neuroblastoma. We then defined rs1064034 AA, rs298982 GG/GA, rs62328061 GG, rs9884978 AA, and rs4834698 TC/CC as risk genotypes. We observed that participants with two to five risk genotypes experienced a 1.47-fold increase in the risk of developing neuroblastoma in comparison with zero to one risk genotypes (adjusted OR = 1.47, 95% CI = 1.15–1.88, p = 0.002).

Stratification Analysis of Identified SNPs

We further explored the association between METTL14 gene polymorphisms and susceptibility to neuroblastoma in certain subgroups classified by age, gender, sites of origins, and clinical stages (Table 2). The protective effect of rs298982 remains prominent in children aged 18 months and over (adjusted OR = 0.29, 95% CI = 0.11-0.75, p = 0.011), male (adjusted OR = 0.38, 95% CI = 0.15-0.99, p = 0.048), and patients in clinical stages I+II+4S (adjusted OR = 0.17, 95% CI = 0.04-0.70, p = 0.014), while the association with rs62328061 was observed for subgroups over 18 months of age (adjusted OR = 0.78, 95% CI = 0.62-0.98, p = 0.033), with tumors originating from the mediastinum (adjusted OR = 0.70, 95% CI = 0.50-0.97, p = 0.032) and with clinical stages III+IV neuroblastoma (adjusted OR = 0.77, 95% CI = 0.60-0.98, p = 0.037). As for the rs4834698 polymorphism, a stronger risk effect of TC/CC genotypes was found among children older than 18 months (adjusted OR = 1.31, 95% CI = 1.03-1.67, p = 0.026) and patients in clinical stages III+IV (adjusted OR = 1.31, 95% CI = 1.01-1.70, p = 0.043). The combined analysis stated that the presence of two to five risk genotypes was associated with an enhanced neuroblastoma risk in all age (\leq 18 months: adjusted OR = 1.49, 95% CI = 1.002-2.22, p = 0.049; >18 months: adjusted OR = 1.43, 95% CI = 1.04–1.95, p = 0.026) and gender groups (female: adjusted OR = 1.54, 95% CI = 1.06-2.25, p = 0.024; male: adjusted OR = 1.41, 95% CI = 1.02-1.95, p = 0.039), children with tumors originating from the adrenal gland (adjusted OR = 1.59, 95% CI = 1.04-2.44, p = 0.034), tumors originating from the mediastinum (adjusted OR = 1.66, 95% CI = 1.04-2.66, p = 0.034), and patients in clinical stages I+II+4S (adjusted OR = 1.53, 95% CI = 1.11-2.11, p = 0.009).

METTL14 Haplotype Analysis

We next determined whether the haplotypes of the five *METTL14* gene SNPs are linked to neuroblastoma risk. As shown in Table 3, the wild-type allele AAAGT was defined as the reference group. When compared with the reference haplotype AAAGT, the following haplo-types were significantly associated with enhanced neuroblastoma risk: AAAGC, AAAAC, AAGGT, AGAGT, AGAGC, AGAAT, AGAAC, AGGGT, TGAGT, TGAAC, and TGGAC.

False-Positive Report Probability (FPRP) Analysis

FPRP analysis was carried out to interrogate whether the significant findings are deserving attention (Table 4). At the prior probability level of 0.1, the significant association for rs62328061 A>G (AG versus AA) remained noteworthy. It was also the case for the association with rs4834698 T>C (CC/CT versus TT) in children over 18 months of

Table 1. Asso	ociation of METTL14 (Gene Polymorphisms with	Neuroblasto	ma Susceptibility			
Genotype	Cases (N = 896)	Controls (N = 1,733)	Controls (N = 1,733) p ^a		р	Adjusted OR (95% CI) ^b	p ^b
rs1064034 T>A	A (HWE = 0.619)						
TT	446 (49.78)	812 (46.86)		1.00		1.00	
ТА	359 (40.07)	755 (43.57)		0.87 (0.73–1.03) 0.098		0.86 (0.73-1.02)	0.084
AA	91 (10.16)	166 (9.58)		1.00 (0.75–1.32) 0.989		1.00 (0.75-1.32)	0.992
Additive			0.386	0.95 (0.84–1.07) 0.386		0.95 (0.84-1.07)	0.367
Dominant	450 (50.22)	921 (53.14)	0.155	0.89 (0.76–1.05) 0.155		0.89 (0.75-1.04)	0.138
Recessive	805 (89.84)	5 (89.84) 1,567 (90.42)		1.07 (0.82-1.40)	0.634	1.07 (0.82–1.40)	0.619
rs298982 G>A	(HWE = 0.092)						
GG	672 (75.00)	1,287 (74.26)		1.00		1.00	
GA	215 (24.00)	403 (23.25)		1.02 (0.85–1.24)	0.824	1.02 (0.84–1.24)	0.833
AA	9 (1.00)	43 (2.48)		0.40 (0.19-0.83)	0.013	0.40 (0.20-0.83)	0.014
Additive			0.271	0.91 (0.77-1.08)	0.271	0.91 (0.77-1.08)	0.273
Dominant	224 (25.00)	446 (25.74)	0.682	0.96 (0.80-1.16)	0.683	0.96 (0.80-1.16)	0.678
Recessive	887 (99.00)	1,690 (97.52)	0.010	0.40 (0.19-0.82)	0.013	0.40 (0.20-0.83)	0.014
rs62328061 A>	>G (HWE = 0.600)						
AA	645 (71.99)	1,176 (67.86)		1.00		1.00	
AG	217 (24.22)	507 (29.26)		0.78 (0.65-0.94) 0.009		0.77 (0.64-0.93)	0.007
GG	34 (3.79)	50 (2.89)		1.24 (0.79–1.94)	0.345	1.24 (0.79–1.94)	0.346
Additive			0.145	0.89 (0.77-1.04)	0.145	0.89 (0.76-1.04)	0.128
Dominant	251 (28.01)	557 (32.14)	0.030	0.82 (0.69-0.98)	0.030	0.82 (0.68-0.97)	0.024
Recessive	862 (96.21) 1,683 (97.11)		0.209	1.33 (0.85–2.07) 0.210		1.33 (0.85-2.07)	0.207
rs9884978 G>.	A (HWE = 0.028)						
GG	576 (64.29)	1,092 (63.01)		1.00		1.00	
GA	255 (28.46)	548 (31.62)		0.88 (0.74-1.06)	0.171	0.88 (0.74-1.05)	0.166
AA	65 (7.25)	93 (5.37)		1.33 (0.95–1.85)	0.097	1.34 (0.96–1.87)	0.087
Additive			0.805	1.02 (0.89–1.16)	0.804	1.02 (0.89–1.16)	0.787
Dominant	320 (35.71)	641 (36.99)	0.520	0.95 (0.80-1.12)	0.521	0.95 (0.80-1.12)	0.521
Recessive	831 (92.75)	1,640 (94.63)	0.054	1.38 (0.99–1.91)	0.054	1.39 (1.00-1.94)	0.048
rs4834698 T>0	C (HWE = 0.587)						
TT	222 (24.78)	484 (27.93)		1.00		1.00	
TC	442 (49.33)	853 (49.22)		1.13 (0.93–1.37) 0.223		1.13 (0.93–1.37)	0.235
CC	232 (25.89)	396 (22.85)		1.28 (1.02–1.60) 0.035		1.28 (1.02-1.61)	0.034
Additive			0.035	1.13 (1.01–1.27) 0.035		1.13 (1.01-1.27)	0.034
Dominant	674 (75.22)	1,249 (72.07)	0.084	1.18 (0.98–1.42)	0.084	1.18 (0.98–1.41)	0.088
Recessive	664 (74.11)	1,337 (77.15)	0.083	1.18 (0.98–1.42)	0.083	1.18 (0.98–1.43)	0.077
Combined Effe	ect of Risk Genotypes ^c						
0-1	100 (11.16)	269 (15.52)		1.00		1.00	
2-5	796 (88.84)	1,464 (84.48)	0.002	1.46 (1.14-1.87)	0.002	1.47 (1.15-1.88)	0.002

Values were in bold if the 95% CIs excluded 1 or P values less than 0.05.

 $^a\chi^2$ test for genotype distributions between neuroblastoma patients and controls.

^bAdjusted for age and gender.

^cRisk genotypes were rs1064034 AA, rs298982 GG/GA, rs62328061 GG, rs9884978 AA, and rs4834698 TC/CC.

age. Regarding the combined analysis, findings for the presence of two to five genotypes in the overall analysis and patients in stages I+II+4S in the stratified analysis could be called noteworthy. Significant findings

remained noteworthy for the haplotypes AAAGC, AGAGT, AGGGT, TGAGT, TGAGC, TGAAT, and TGAAC when compared with reference haplotype AAAGT.

rs298982 (Case/ Control)			rs62328061 (Case/Control)		rs4834698 (Case/Contro!					Risk Genotypes (Case/ Control)						
Variables	GG/ GA	AA	AOR (95% CI) ^a	p ^a	AA	AG/ GG	AOR (95% CI) ^a	p ^a	TT	TC/CC	AOR (95% CI) ^a	p ^a	0-1	2–5	AOR (95% CI) ^a	p ^a
Age, Months																
≤18	340/ 702	4/ 11	0.77 (0.24– 2.44)	0.656	244/ 487	100/ 226	0.88 (0.67– 1.17)	0.380	93/ 190	251/ 523	0.98 (0.73– 1.31)	0.880	37/ 109	307/ 604	1.49 (1.002- 2.22)	0.049
>18	547/ 988	5/ 32	0.29 (0.11- 0.75)	0.011	401/ 689	151/ 331	0.78 (0.62- 0.98)	0.033	129/ 294	423/ 726	1.31 (1.03– 1.67)	0.026	63/ 160	489/ 860	1.43 (1.04– 1.95)	0.026
Gender														_		
Female	402/ 727	4/ 17	0.44 (0.15– 1.33)	0.146	293/ 501	113/ 243	0.79 (0.60– 1.03)	0.077	101/ 197	305/ 547	1.07 (0.81– 1.42)	0.620	42/ 113	364/ 631	1.54 (1.06– 2.25)	0.024
Male	485/ 963	5/ 26	0.38 (0.15- 0.99)	0.048	352/ 675	138/ 314	0.84 (0.66– 1.07)	0.150	121/ 287	369/ 702	1.25 (0.98– 1.60)	0.074	58/ 156	432/ 833	1.41 (1.02- 1.95)	0.039
Sites of Origin																
Adrenal gland	246/ 1,690	2/ 43	0.32 (0.08– 1.34)	0.120	181/ 1,176	67/ 557	0.77 (0.57– 1.04)	0.085	55/ 484	193/ 1249	1.37 (1.00– 1.88)	0.054	26/ 269	222/ 1,464	1.59 (1.04– 2.44)	0.034
Retroperitoneal	312/ 1,690	6/ 43	0.76 (0.32– 1.81)	0.540	222/ 1,176	96/ 557	0.90 (0.69– 1.17)	0.416	90/ 484	228/ 1,249	0.99 (0.76– 1.29)	0.942	40/ 269	278/ 1,464	1.29 (0.90– 1.84)	0.163
Mediastinum	212/ 1,690	1/ 43	0.19 (0.03– 1.37)	0.099	160/ 1,176	53/ 557	0.70 (0.50- 0.97)	0.032	48/ 484	165/ 1,249	1.31 (0.94– 1.84)	0.115	21/ 269	192/ 1,464	1.66 (1.04– 2.66)	0.034
Others	105/ 1,690	0/ 43	/	/	71/ 1,176	34/ 557	1.02 (0.67– 1.55)	0.943	27/ 484	78/ 1,249	1.11 (0.71– 1.74)	0.649	13/ 269	92/ 1,464	1.29 (0.71– 2.34)	0.400
Clinical Stages																
I+II+4S	467/ 1,690	2/ 43	0.17 (0.04- 0.70)	0.014	333/ 1,176	136/ 557	0.86 (0.69– 1.08)	0.194	124/ 484	346/ 1,249	1.09 (0.86– 1.37)	0.479	50/ 269	419/ 1,464	1.53 (1.11- 2.11)	0.009
III+IV	387/ 1,690	7/ 43	0.74 (0.33– 1.66)	0.464	286/ 1,176	108/ 557	0.77 (0.60- 0.98)	0.037	91/ 484	303/ 1,249	1.31 (1.01– 1.70)	0.043	48/ 269	346/ 1,464	1.36 (0.97– 1.89)	0.073

AOR, adjusted odds ratio.

^aAdjusted for age and gender, omitting the corresponding stratify factor.

Effect of rs298982 G>A on the Expression of Surrounding Genes To further assess whether the functional relevance of rs298982 G>A affects mRNA expression, we explored Cis-expression quantitative trait loci (eQTLs) target genes of the rs298982 G>A using released data from GTEx. It showed that the rs298982 G allele was significantly associated with increased SNHG8 mRNA levels in the cultured fibroblasts (Figure 1A). However, the rs298982 G allele was significantly associated with lower expression levels of RP11-384K6.6 in the whole blood (Figure 1B) and cultured fibroblasts (Figure 1C). The overall workflow is shown in Figure 2.

DISCUSSION

More and more novel neuroblastoma susceptibility genetic variants have been identified. Yet it remains a challenge to unearth the full range of neuroblastoma susceptibility variations. In this study, we provided evidence that common variations in the METTL14 gene were significantly associated with the risk of neuroblastoma. Our data also shed light on the biological mechanisms by which METTL14 gene SNP rs298982 G>A enhances hepatoblastoma risk. This is the most extensive study to date

studying the association of METTL14 gene SNPs and neuroblastoma risk.

Recent research has uncovered the importance of METTL14 in cancer development. Chen et al.³⁴ detected a lower expression level of METTL14 in colorectal cancer tissues and cell lines. The low METTL14 was significantly associated with poor overall survival. Furthermore, functional experiments demonstrated that METTL14 suppressed colorectal cancer via the miR-375/SP1 and miR-375/ YAP1 pathways. Ma et al.³⁵ reported that METTL14 was significantly downregulated in hepatocellular carcinoma. Reduced METTL14 expression had worse recurrence-free survival and overall survival. Functionally, METTL14 facilitates the maturation of primary miR-126 in an m⁶A-dependent manner by binding to microprocessor protein DGCR8. In contrast, Weng et al.²⁹ found that METTL14 was highly expressed in acute myeloid leukemia cells and played an oncogenic role. Lang et al.³⁶ indicated that METTL14 was an important driver in Epstein-Barr virus (EBV)-induced oncogenesis. They showed that knockdown of METTL14 reduced tumorigenic activity of EBV-transformed cells in the xenograft animal model systems.

Haplotypes ^a	Cases (N = 1,792)	Controls ($N = 3,466$)	Crude OR (95% CI)	р	Adjusted OR ^b (95% CI)	p ^b	
AAAGT	23 (1.28)	85 (2.45)	1.00		1.00		
AAAGC	117 (6.53)	222 (6.41)	1.95 (1.17-3.25)	0.011	1.95 (1.17-3.25)	0.011	
AAAAT	35 (1.95)	100 (2.89)	1.29 (0.71-2.35)	0.401	1.29 (0.71-2.36)	0.403	
AAAAC	4 (0.22)	2 (0.06)	7.39 (1.27-42.91)	0.026	7.30 (1.25-42.51)	0.027	
AAGGT	42 (2.34)	76 (2.19)	2.04 (1.13-3.70)	0.019	2.01 (1.11-3.64)	0.022	
AAGGC	3 (0.17)	3 (0.09)	3.70 (0.70–19.54)	0.124	3.70 (0.70–19.57)	0.124	
AAGAT	0 (0.00)	1 (0.03)	1	/	/	/	
AGAGT	50 (2.79)	74 (2.14)	2.50 (1.39-4.48)	0.002	2.47 (1.38-4.44)	0.002	
AGAGC	19 (1.06)	1 (0.03)	70.22 (8.92-552.56)	<0.0001	70.21 (8.91-553.13)	<0.0001	
AGAAT	9 (0.50)	1 (0.03)	33.26 (4.01-276.19)	0.001	32.69 (3.93-271.78)	0.001	
AGAAC	10 (0.56)	1 (0.03)	36.96 (4.50-303.80)	0.0008	35.34 (4.29-290.73)	0.0009	
AGGGT	62 (3.46)	99 (2.86)	2.31 (1.32-4.05)	0.003	2.34 (1.34-4.10)	0.003	
AGGGC	123 (6.86)	300 (8.66)	1.52 (0.91-2.51)	0.108	1.50 (0.90-2.49)	0.118	
AGGAT	43 (2.40)	122 (3.52)	1.30 (0.73-2.32)	0.369	1.29 (0.73-2.32)	0.390	
AGGAC	1 (0.06)	0 (0.00)	1	/	/	/	
TAAGT	3 (0.17)	0 (0.00)	1	/	/	/	
TAAGC	2 (0.11)	0 (0.00)	1	/	/	/	
TAAAT	4 (0.22)	0 (0.00)	1	/	/	/	
TGAGT	509 (28.40)	1,077 (31.07)	1.75 (1.09-2.80)	0.021	1.74 (1.08-2.79)	0.022	
GAGC	448 (25.00)	790 (22.79)	2.10 (1.30-3.37)	0.002	2.09 (1.30-3.37)	0.002	
GAAT	102 (5.69)	185 (5.34)	2.04 (1.21-3.43)	0.007	2.05 (1.22-3.45)	0.007	
ſGAAC	172 (9.60)	321 (9.26)	1.98 (1.21-3.25)	0.007	1.97 (1.20-3.24)		
TGGGT	2 (0.11)	1 (0.03)	7.39 (0.64-85.16)	0.109	5.89 (0.51-68.68)	0.157	
ſGGGC	4 (0.22)	4 (0.12)	3.70 (0.86-14.92)	0.079	3.65 (0.85-15.74)	0.082	
ſGGAT	2 (0.11)	0 (0.00)	/	/	/	/	
ГGGAC	3 (0.17)	1 (0.03)	11.09 (1.10-111.65)	0.041	10.84 (1.07-109.36)	0.043	

^bObtained in logistic regression models with adjustment for age and gender.

There is only one publication regarding the association of METTL14 gene SNPs with cancer risk so far. In brief, Meng et al.³⁷ explored the association between m⁶A gene SNPs and colorectal cancer risk in a two-stage case-control study with 1,150 cases and 1,342 controls in the discovery stage, and with 932 cases and 966 controls in the validation stage. Out of 240 SNPs in 20 m⁶A modification-related genes, only 1 SNP, rs118049207, located in the SND1 gene, was identified to predispose to colorectal cancer in the Chinese population. None of the five studied METTL14 gene SNPs (rs115267066, rs298981, rs2029399, rs167246, and rs441216) was associated with colorectal cancer risk. Clearly, these results can provide genetic insights into the origins of colorectal cancer risk. However, to date, the roles of the METTL14 gene SNPs in neuroblastoma risk are unknown. To fill this gap, we designed the current case-control investigation to determine the correlation of METTL14 gene polymorphisms and neuroblastoma risk in the Chinese population. In the present study, significant relationships were detected among neuroblastoma risk and the four METTL14 gene polymorphisms (rs298982 G>A,

rs62328061 A>G, rs9884978 G>A, and rs4834698 T>C). Compared with the genotypes of single SNPs, association studies based on haplotypes of multiple markers significantly improve the power of mapping and characterizing disease-causing genes.^{38,39} Therefore, we explored whether various haplotypes consisting of the five METTL14 gene polymorphisms are associated with neuroblastoma risk. Expectedly, METTL14 gene haplotypes significantly confer a higher risk of neuroblastoma. These results suggest that these variants may interact with each other to modify the risk of neuroblastoma. We further attempted to interpret the possible mechanism of METTL14 gene SNP-mediated neuroblastoma risk. eQTLs evidence suggested that the G allele in rs298982 is significantly associated with increased long noncoding RNA SNHG8 (lncRNA SNHG8) level in the cultured fibroblasts. lncRNA SNHG8 was documented to play oncogenic roles in several kinds of cancers.⁴⁰⁻⁴⁴ We propose the higher expression of lncRNA SNHG8 caused by G allele in rs298982 may facilitate the development of neuroblastoma. On the contrary, with the increase of G genotype, the average expression of RP11-

				Prior Probability					
Genotype	Crude OR (95% CI)	p ^a	Statistical Power ^b	0.25	0.1	0.01	0.001	0.0001	
rs298982 G>A									
AA versus GG	0.40 (0.19-0.83)	0.013	0.113	0.263	0.517	0.922	0.992	0.999	
AA versus GG/GA	0.40 (0.19-0.82)	0.013	0.109	0.259	0.512	0.920	0.991	0.999	
>18	0.28 (0.11-0.73)	0.009	0.063	0.299	0.561	0.934	0.993	0.999	
Stages I+II+4S	0.17 (0.04-0.70)	0.014	0.056	0.430	0.694	0.961	0.996	1.000	
rs62328061 A>G									
AG versus AA	0.78 (0.65-0.94)	0.009	0.944	0.027	0.078	0.483	0.904	0.989	
AG/GG versus AA	0.82 (0.69-0.98)	0.030	0.986	0.083	0.214	0.750	0.968	0.997	
>18	0.78 (0.62-0.99)	0.037	0.904	0.109	0.268	0.801	0.976	0.998	
Mediastinum	0.70 (0.51-0.97)	0.032	0.605	0.137	0.323	0.840	0.981	0.998	
rs4834698 T>C									
CC versus TT	1.28 (1.02-1.60)	0.035	0.987	0.095	0.239	0.776	0.972	0.997	
CC/CT versus TT	·								
>18	1.33 (1.05–1.69)	0.020	0.833	0.068	0.178	0.705	0.960	0.996	
Risk Genotypes									
2-5 versus 0-1	1.46 (1.14–1.87)	0.002	0.582	0.012	0.036	0.290	0.805	0.976	
≤18	1.50 (1.01-2.23)	0.047	0.507	0.216	0.452	0.901	0.989	0.999	
>18	1.44 (1.06–1.97)	0.021	0.593	0.096	0.242	0.778	0.973	0.997	
Female	1.55 (1.06-2.26)	0.022	0.439	0.132	0.314	0.834	0.981	0.998	
Male	1.40 (1.01-1.93)	0.044	0.667	0.164	0.370	0.866	0.985	0.998	
Adrenal gland	1.57 (1.02-2.40)	0.039	0.427	0.213	0.448	0.899	0.989	0.999	
Mediastinum	1.68 (1.05-2.69)	0.030	0.330	0.215	0.451	0.900	0.989	0.999	
Stages I+II+4S	1.54 (1.12–2.12)	0.008	0.446	0.053	0.145	0.651	0.949	0.995	
Risk Haplotypes									
AAAGC versus AAAGT	1.95 (1.17-3.25)	0.011	0.884	0.035	0.098	0.545	0.924	0.992	
AAAAC versus AAAGT	7.39 (1.27-42.91)	0.026	0.041	0.651	0.848	0.984	0.998	1.000	
AAGGT versus AAAGT	2.04 (1.13-3.70)	0.019	0.485	0.104	0.258	0.793	0.975	0.997	
AGAGT versus AAAGT	2.50 (1.39-4.48)	0.002	0.227	0.027	0.077	0.478	0.902	0.989	
AGAGC versus AAAGT	70.22 (8.92-552.56)	< 0.0001	0.000	0.443	0.705	0.963	0.996	1.000	
AGAAT versus AAAGT	33.26 (4.01-276.19)	0.001	0.003	0.574	0.802	0.978	0.998	1.000	
AGAAC versus AAAGT	36.96 (4.50-303.80)	0.0008	0.002	0.557	0.790	0.976	0.998	1.000	
AGGGT versus AAAGT	2.31 (1.32-4.05)	0.003	0.383	0.025	0.072	0.460	0.896	0.989	
TGAGT versus AAAGT	1.75 (1.09–2.80)	0.021	1.000	0.058	0.157	0.672	0.954	0.995	
TGAGC versus AAAGT	2.10 (1.30-3.37)	0.002	0.999	0.007	0.020	0.186	0.697	0.958	
TGAAT versus AAAGT	2.04 (1.21-3.43)	0.007	0.784	0.027	0.077	0.480	0.903	0.989	
TGAAC versus AAAGT	1.98 (1.21-3.25)	0.007	0.955	0.022	0.062	0.420	0.880	0.987	
TGGAC versus AAAGT	11.09 (1.10-111.65)	0.041	0.048	0.719	0.885	0.988	0.999	1.000	

- _ D. -----, Val . . . -Del -1 N -- 1-... ~

 $^{a}\chi^{2}$ test was used to calculate the genotype frequency distributions.

^bStatistical power was calculated using the number of observations in the subgroup and the OR and p values in this table.

384K6.6 is gradually decreased. This conclusion requires further interpretation as the role of RP11-384K6.6 remains to be revealed. In all, more functional experiments are needed to support this possible mechanism.

The major strength of this study includes novelty, the relatively large sample size in neuroblastoma cases, and multiple-center participants in a single Chinese population. However, our results should be interpreted in light of three limitations. First, the study population



Figure 1. Functional Relevance of rs298982 G>A on Genes Expression in GTEx Database

(A–C) The rs298982 G genotype had a significantly (A) higher *SNHG8* mRNA level in the cell-cultured fibroblasts (p = 1.8×10^{-6}), but a significantly lower *RP11-384K6.6* level in the (B) whole blood (p = 3.9×10^{-9}) and (C) cell-cultured fibroblasts (p = 9.4×10^{-9}).

involved only Chinese subjects and was limited to volunteers. Therefore, the generalizability of these findings to the general population was reduced. On the other hand, a single population background here may strengthen the reliability of the conclusion in the study. It is worth pointing out that information from diverse ethnic backgrounds is very useful for elucidating pathogenetic mechanisms in greater detail. Further comparative studies are needed to clarify whether the SNPs are also associated with neuroblastoma in other ethnicities with different genetic backgrounds. Second, only genetic factors, but not environmental factors, were taken into account. Third, the number of SNPs included was relatively small.

In this study, we, for the first time, identified a significant association of *METTL14* gene SNPs with neuroblastoma risk. These SNPs in the *METTL14* gene are intriguing loci for further studies, and the underlying biological mechanisms should be revealed.

MATERIALS AND METHODS

Sample Selection

The current study was approved by the institutional review board of Guangzhou Women and Children's Medical Center. We recruited 898 neuroblastoma patients registered in eight hospitals located in eight cities (Guangzhou, Zhengzhou, Wenzhou, Xi'an, Taiyuan, Kunming, Changsha, and Shenyang). We also included 1,734 age- and gendermatched healthy controls having visited the participating hospitals. Written consent was obtained from all participants at enrolment into the study. More details could refer to our previous work.^{45,46}

Population Characteristics

The clinical characteristics of the participants are depicted in Table S1. The mean age for cases was 33.11 ± 28.07 months, and the mean age for controls was 30.41 ± 24.90 months. Overall, 898 cases and 1,734 controls were well matched in terms of age (p = 0.155) and gender (p = 0.236). According to the International Neuroblastoma Staging System (INSS),⁴⁷ 310 neuroblastoma cases (34.52%) were diagnosed with clinical stage I, 160 (17.82%) with clinical stage II, 163 (18.15%) with clinical stage III, 231 (25.72%) with clinical stage IV, 18 (2.00%) with clinical stage 4S disease, and 16 (1.78%) without adequate information. Overall, 248 (27.62%) neuroblastomas occurred in the adrenal gland, 319 (35.52%) in the retroperitoneal region, 214 (23.83%) in the mediastinum, 105 (11.69%) in other regions, and 12 (1.34%) to be determined.

Polymorphism Selection and Genotyping

METTL14 gene SNPs with potential functions were retrieved from the dbSNP database and SNPinfo software, with details reported elsewhere.⁴⁸ In brief, selection criteria were as follows: (1) located at the two ends of the METTL14 gene (i.e., the 5' near gene, 5' untranslated region [UTR], 3' UTR, and 3' near gene); (2) the minor allele frequency \geq 5% for Chinese Han subjects reported in 1000 Genomes (https://www.ncbi.nlm.nih.gov/variation/tools/1000genomes/) and (3) affecting transcription factor binding site (TFBS) activity or the miRNA binding sites activity; and (4) SNPs in low linkage disequilibrium (LD) with each other ($R^2 < 0.8$). Five SNPs (rs1064034 T>A, rs298982 G>A, rs62328061 A>G, rs9884978 G>A, and rs4834698 T>C) fell into the scope of criteria. Among them, rs298982 G>A and rs9884978 G>A are located in the 5' near gene, rs1064034 T>A and rs4834698 T>C are located in the 3' UTR, and the rs62328061 A>G is located in the coding region. Moreover, rs1064034 T>A and rs4834698 T>C affect miRNA binding sites activity, whereas rs298982 G>A and rs9884978 G>A affect TFBS activity. In the last, rs62328061 A>G modulates splicing activity. There was no significant LD ($R^2 < 0.8$) among these five *METTL14* SNPs ($R^2 = 0.033$ between rs298982 G>A and rs9884978 G>A; R² = 0.023 between rs298982 G>A and rs62328061 A>G; R² = 0.135 between rs298982 G>A and rs4834698 T>C; R² = 0.322 between rs298982 G>A and rs1064034 T>A; $R^2 = 0.054$ between rs9884978 G>A and rs62328061 A>G; R^2 = 0.252 between rs9884978 G>A and rs4834698 T>C; R^2 = 0.103 between rs9884978 G>A and rs1064034 T>A; $R^2 = 0.218$ between rs62328061 A>G and rs4834698 T>C; R² = 0.521 between rs62328061 A>G and rs1064034 T>A; $R^2 = 0.419$ between rs4834698 T>C and rs1064034 T>A) (Figure S1). Genomic DNA was extracted from peripheral blood using the standard procedure. The genotyping of the SNPs was conducted using the TaqMan SNP genotyping assay.⁴⁹⁻⁵¹ Laboratory technicians were blind to the sample information, including the identities of the replicate aliquots. A repeated genotyping was analyzed by an arbitrarily chosen 10% of the samples from both cases and controls. A concordance rate of 100% was obtained.

Statistical Analysis

We checked HWE for each SNP in controls using a goodness-of-fit χ^2 test. A two-sided χ^2 test was used to analyze the difference in demographic and clinical variables between the cases and controls. The homozygotes of the common allele served as the reference group. The



Figure 2. The Workflow of the Current Study

remaining genotypes were classified as variants. ORs and 95% CIs were calculated for the variant compared with the reference category using logistic regression analyses. Data were further stratified by age, gender, sites of origins, and clinical stages. The estimation of haplotype frequency and the analysis of their effect on neuroblastoma risk were performed using logistic regression analysis.^{46,52} We also applied the FPRP analysis to evaluate noteworthy associations by the means as described elsewhere.^{53,54} In brief, three parameters were employed to determine FPRP values, statistical power, p value, and prior probability π representing a real association between the SNP and a disease. We set 0.2 as an FPRP threshold and assigned a prior probability of 0.1 to detect an OR of 1.50 (for risk effects) or 0.67 (for protective effects) for an association with genotypes under investigation. eQTLs analysis in the GTEx portal (https://www. gtexportal.org/home/) was adopted to determine the correlation between the SNPs and levels of nearby genes expression.⁵⁵ p values below 0.05 were considered significant. All statistical calculations were carried out using SAS 9.1 (SAS Institute, Cary, NC, USA).

Novelty

We genotyped five *METTL14* polymorphisms in 898 neuroblastoma cases and 1,734 controls enrolled from eight hospitals. We found that rs298982, rs62328061, rs9884978, and rs4834698 were associated with neuroblastoma susceptibility. The significant associations were further validated by stratified analyses, haplotype analyses, and FPRP analyses. eQTL analysis suggested a potential functional role of rs298982 in neuroblastoma. Our results first highlight the critical roles of *METTL14* polymorphisms in the etiology of neuroblastoma.

SUPPLEMENTAL INFORMATION

Supplemental Information can be found online at https://doi.org/10. 1016/j.omtn.2020.08.009.

AUTHOR CONTRIBUTIONS

Z.Z., H.L., J. Zhu, J.H., and H.X. designed and performed the study and wrote the manuscript; Y.L., Z.Y., J. Zhang, J.C., H.Z., S.L., L.L., and J.H. collected the samples and information; R.-X.H. and J.H. participated in analyzing data; Z.Z., J.H., and H.X. coordinated the study over the entire time. All authors reviewed the final manuscript.

CONFLICTS OF INTEREST

The authors declare no competing interests.

ACKNOWLEDGMENTS

This study was supported by grants from the Natural Science Foundation of Guangdong Province (grant 2019A1515010360), Pearl River S&T Nova Program of Guangzhou (grant 201710010086), and Guangdong Provincial Key Laboratory of Research in Structural Birth Defect Disease (grant 2019B030301004).

REFERENCES

- Maris, J.M. (2010). Recent advances in neuroblastoma. N. Engl. J. Med. 362, 2202– 2211.
- Tas, M.L., Reedijk, A.M.J., Karim-Kos, H.E., Kremer, L.C.M., van de Ven, C.P., Dierselhuis, M.P., van Eijkelenburg, N.K.A., van Grotel, M., Kraal, K.C.J.M., Peek, A.M.L., et al. (2020). Neuroblastoma between 1990 and 2014 in the Netherlands: Increased incidence and improved survival of high-risk neuroblastoma. Eur. J. Cancer 124, 47–55.
- Bao, P.P., Li, K., Wu, C.X., Huang, Z.Z., Wang, C.F., Xiang, Y.M., Peng, P., Gong, Y.M., Xiao, X.M., and Zheng, Y. (2013). [Recent incidences and trends of childhood malignant solid tumors in Shanghai, 2002-2010]. Zhonghua Er Ke Za Zhi 51, 288–294.
- Matthay, K.K., Maris, J.M., Schleiermacher, G., Nakagawara, A., Mackall, C.L., Diller, L., and Weiss, W.A. (2016). Neuroblastoma. Nat. Rev. Dis. Primers 2, 16078.
- Maris, J.M., Hogarty, M.D., Bagatell, R., and Cohn, S.L. (2007). Neuroblastoma. Lancet 369, 2106–2120.
- Newman, E.A., Abdessalam, S., Aldrink, J.H., Austin, M., Heaton, T.E., Bruny, J., Ehrlich, P., Dasgupta, R., Baertschiger, R.M., Lautz, T.B., et al.; APSA Cancer committee (2019). Update on neuroblastoma. J. Pediatr. Surg. 54, 383–389.
- 7. Cheung, N.K., and Dyer, M.A. (2013). Neuroblastoma: developmental biology, cancer genomics and immunotherapy. Nat. Rev. Cancer 13, 397–411.
- Almstedt, E., Elgendy, R., Hekmati, N., Rosén, E., Wärn, C., Olsen, T.K., Dyberg, C., Doroszko, M., Larsson, I., Sundström, A., et al. (2020). Integrative discovery of treatments for high-risk neuroblastoma. Nat. Commun. 11, 71.

- 9. Esposito, M.R., Aveic, S., Seydel, A., and Tonini, G.P. (2017). Neuroblastoma treatment in the post-genomic era. J. Biomed. Sci. 24, 14.
- Barr, E.K., and Applebaum, M.A. (2018). Genetic Predisposition to Neuroblastoma. Children (Basel) 5, 119.
- Zhong, X., Liu, Y., Liu, H., Zhang, Y., Wang, L., and Zhang, H. (2018). Identification of Potential Prognostic Genes for Neuroblastoma. Front. Genet. 9, 589.
- Tonini, G.P., and Capasso, M. (2020). Genetic predisposition and chromosome instability in neuroblastoma. Cancer Metastasis Rev. 39, 275–285.
- 13. Trochet, D., Bourdeaut, F., Janoueix-Lerosey, I., Deville, A., de Pontual, L., Schleiermacher, G., Coze, C., Philip, N., Frébourg, T., Munnich, A., et al. (2004). Germline mutations of the paired-like homeobox 2B (PHOX2B) gene in neuroblastoma. Am. J. Hum. Genet. 74, 761–764.
- Ogawa, S., Takita, J., Sanada, M., and Hayashi, Y. (2011). Oncogenic mutations of ALK in neuroblastoma. Cancer Sci. 102, 302–308.
- Aygün, Z., Batur, Ş., Emre, Ş., Celkan, T., Özman, O., and Comunoglu, N. (2019). Frequency of ALK and GD2 Expression in Neuroblastoma. Fetal Pediatr. Pathol. 38, 326–334.
- 16. Wang, D.G., Fan, J.B., Siao, C.J., Berno, A., Young, P., Sapolsky, R., Ghandour, G., Perkins, N., Winchester, E., Spencer, J., et al. (1998). Large-scale identification, mapping, and genotyping of single-nucleotide polymorphisms in the human genome. Science 280, 1077–1082.
- Hirschhorn, J.N., and Daly, M.J. (2005). Genome-wide association studies for common diseases and complex traits. Nat. Rev. Genet. 6, 95–108.
- Maris, J.M., Mosse, Y.P., Bradfield, J.P., Hou, C., Monni, S., Scott, R.H., Asgharzadeh, S., Attiyeh, E.F., Diskin, S.J., Laudenslager, M., et al. (2008). Chromosome 6p22 locus associated with clinically aggressive neuroblastoma. N. Engl. J. Med. 358, 2585–2593.
- Capasso, M., Diskin, S., Cimmino, F., Acierno, G., Totaro, F., Petrosino, G., Pezone, L., Diamond, M., McDaniel, L., Hakonarson, H., et al. (2014). Common genetic variants in NEFL influence gene expression and neuroblastoma risk. Cancer Res. 74, 6913–6924.
- 20. Capasso, M., McDaniel, L.D., Cimmino, F., Cirino, A., Formicola, D., Russell, M.R., Raman, P., Cole, K.A., and Diskin, S.J. (2017). The functional variant rs34330 of CDKN1B is associated with risk of neuroblastoma. J. Cell. Mol. Med. 21, 3224–3230.
- 21. Diskin, S.J., Capasso, M., Schnepp, R.W., Cole, K.A., Attiyeh, E.F., Hou, C., Diamond, M., Carpenter, E.L., Winter, C., Lee, H., et al. (2012). Common variation at 6q16 within HACE1 and LIN28B influences susceptibility to neuroblastoma. Nat. Genet. 44, 1126–1130.
- 22. McDaniel, L.D., Conkrite, K.L., Chang, X., Capasso, M., Vaksman, Z., Oldridge, D.A., Zachariou, A., Horn, M., Diamond, M., Hou, C., et al. (2017). Common variants upstream of MLF1 at 3q25 and within CPZ at 4p16 associated with neuroblastoma. PLoS Genet. 13, e1006787.
- 23. Oldridge, D.A., Wood, A.C., Weichert-Leahey, N., Crimmins, I., Sussman, R., Winter, C., McDaniel, L.D., Diamond, M., Hart, L.S., Zhu, S., et al. (2015). Genetic predisposition to neuroblastoma mediated by a LMO1 super-enhancer polymorphism. Nature 528, 418–421.
- Cao, G., Li, H.B., Yin, Z., and Flavell, R.A. (2016). Recent advances in dynamic m6A RNA modification. Open Biol. 6, 160003.
- 25. Liang, Y., Zhan, G., Chang, K.J., Yang, Y.P., Wang, L., Lin, J., and Hsu, C.H. (2020). The roles of m6A RNA modifiers in human cancer. J. Chin. Med. Assoc. 83, 221–226.
- Meyer, K.D., and Jaffrey, S.R. (2017). Rethinking m⁶A Readers, Writers, and Erasers. Annu. Rev. Cell Dev. Biol. 33, 319–342.
- 27. Li, T., Hu, P.S., Zuo, Z., Lin, J.F., Li, X., Wu, Q.N., Chen, Z.H., Zeng, Z.L., Wang, F., Zheng, J., et al. (2019). METTL3 facilitates tumor progression via an m⁶A-IGF2BP2dependent mechanism in colorectal carcinoma. Mol. Cancer 18, 112.
- Orouji, E., Peitsch, W.K., Orouji, A., Houben, R., and Utikal, J. (2020). Oncogenic Role of an Epigenetic Reader of m(6)A RNA Modification: YTHDF1 in Merkel Cell Carcinoma. Cancers (Basel) 12, 202.
- 29. Weng, H., Huang, H., Wu, H., Qin, X., Zhao, B.S., Dong, L., Shi, H., Skibbe, J., Shen, C., Hu, C., et al. (2018). METTL14 Inhibits Hematopoietic Stem/Progenitor Differentiation and Promotes Leukemogenesis via mRNA m⁶A Modification. Cell Stem Cell 22, 191–205.e9.

- 30. Zheng, Y., Nie, P., Peng, D., He, Z., Liu, M., Xie, Y., Miao, Y., Zuo, Z., and Ren, J. (2018). m6AVar: a database of functional variants involved in m6A modification. Nucleic Acids Res. 46 (D1), D139–D145.
- 31. Du, T., Rao, S., Wu, L., Ye, N., Liu, Z., Hu, H., Xiu, J., Shen, Y., and Xu, Q. (2015). An association study of the m6A genes with major depressive disorder in Chinese Han population. J. Affect. Disord. 183, 279–286.
- Mo, X.B., Zhang, Y.H., and Lei, S.F. (2018). Genome-wide identification of m⁶A-associated SNPs as potential functional variants for bone mineral density. Osteoporos. Int. 29, 2029–2039.
- 33. Kalantari, N., Keshavarz Mohammadi, N., Izadi, P., Doaei, S., Gholamalizadeh, M., Eini-Zinab, H., Salonurmi, T., Rafieifar, S., Janipoor, R., and Azizi Tabesh, G. (2018). A haplotype of three SNPs in FTO had a strong association with body composition and BMI in Iranian male adolescents. PLoS ONE 13, e0195589.
- 34. Chen, X., Xu, M., Xu, X., Zeng, K., Liu, X., Sun, L., Pan, B., He, B., Pan, Y., Sun, H., et al. (2020). METTL14 Suppresses CRC Progression via Regulating N6-Methyladenosine-Dependent Primary miR-375 Processing. Mol. Ther. 28, 599–612.
- 35. Ma, J.Z., Yang, F., Zhou, C.C., Liu, F., Yuan, J.H., Wang, F., Wang, T.T., Xu, Q.G., Zhou, W.P., and Sun, S.H. (2017). METTL14 suppresses the metastatic potential of hepatocellular carcinoma by modulating N⁶-methyladenosine-dependent primary MicroRNA processing. Hepatology 65, 529–543.
- 36. Lang, F., Singh, R.K., Pei, Y., Zhang, S., Sun, K., and Robertson, E.S. (2019). EBV epitranscriptome reprogramming by METTL14 is critical for viral-associated tumorigenesis. PLoS Pathog. 15, e1007796.
- 37. Meng, Y., Li, S., Gu, D., Xu, K., Du, M., Zhu, L., Chu, H., Zhang, Z., Wu, Y., Fu, Z., and Wang, M. (2020). Genetic variants in m6A modification genes are associated with colorectal cancer risk. Carcinogenesis 41, 8–17.
- Akey, J., Jin, L., and Xiong, M. (2001). Haplotypes vs single marker linkage disequilibrium tests: what do we gain? Eur. J. Hum. Genet. 9, 291–300.
- Manolio, T.A., Brooks, L.D., and Collins, F.S. (2008). A HapMap harvest of insights into the genetics of common disease. J. Clin. Invest. 118, 1590–1605.
- 40. Dong, J., Teng, F., Guo, W., Yang, J., Ding, G., and Fu, Z. (2018). IncRNA SNHG8 Promotes the Tumorigenesis and Metastasis by Sponging miR-149-5p and Predicts Tumor Recurrence in Hepatocellular Carcinoma. Cell. Physiol. Biochem. 51, 2262– 2274.
- Qu, X., Li, Y., Wang, L., Yuan, N., Ma, M., and Chen, Y. (2020). LncRNA SNHG8 accelerates proliferation and inhibits apoptosis in HPV-induced cervical cancer through recruiting EZH2 to epigenetically silence RECK expression. J. Cell. Biochem. Published online January 21, 2020. https://doi.org/10.1002/jcb.29646.
- 42. Song, H., Song, J., Lu, L., and Li, S. (2019). SNHG8 is upregulated in esophageal squamous cell carcinoma and directly sponges microRNA-411 to increase oncogenicity by upregulating KPNA2. OncoTargets Ther. 12, 6991–7004.
- 43. Song, Y., Zou, L., Li, J., Shen, Z.P., Cai, Y.L., and Wu, X.D. (2018). LncRNA SNHG8 promotes the development and chemo-resistance of pancreatic adenocarcinoma. Eur. Rev. Med. Pharmacol. Sci. 22, 8161–8168.
- 44. Zhen, Y., Ye, Y., Wang, H., Xia, Z., Wang, B., Yi, W., and Deng, X. (2019). Knockdown of SNHG8 repressed the growth, migration, and invasion of colorectal cancer cells by directly sponging with miR-663. Biomed. Pharmacother. 116, 109000.
- 45. Zhuo, Z.J., Liu, W., Zhang, J., Zhu, J., Zhang, R., Tang, J., Yang, T., Zou, Y., He, J., and Xia, H. (2018). Functional Polymorphisms at ERCC1/XPF Genes Confer Neuroblastoma Risk in Chinese Children. EBioMedicine 30, 113–119.
- 46. Hua, R.X., Zhuo, Z., Ge, L., Zhu, J., Yuan, L., Chen, C., Liu, J., Cheng, J., Zhou, H., Zhang, J., et al. (2020). LIN28A gene polymorphisms modify neuroblastoma susceptibility: A four-centre case-control study. J. Cell. Mol. Med. 24, 1059–1066.
- 47. Brodeur, G.M., Pritchard, J., Berthold, F., Carlsen, N.L., Castel, V., Castelberry, R.P., De Bernardi, B., Evans, A.E., Favrot, M., Hedborg, F., et al. (1994). Revisions of the international criteria for neuroblastoma diagnosis, staging and response to treatment. Prog. Clin. Biol. Res. 385, 363–369.
- 48. He, J., Wang, F., Zhu, J., Zhang, R., Yang, T., Zou, Y., and Xia, H. (2016). Association of potentially functional variants in the XPG gene with neuroblastoma risk in a Chinese population. J. Cell. Mol. Med. 20, 1481–1490.
- 49. Chen, X., Wang, Y., Chen, X., Cheng, K., Li, J., Lou, J., Ke, J., Yang, Y., Gong, Y., Zhu, Y., et al. (2016). Genetic variants in the regulatory region of SLC10A1 are not

associated with the risk of hepatitis B virus infection and clearance. Infect. Genet. Evol. 44, 495–500.

- 50. Li, J., Chang, J., Tian, J., Ke, J., Zhu, Y., Yang, Y., Gong, Y., Zou, D., Peng, X., Yang, N., et al. (2018). A Rare Variant P507L in TPP1 Interrupts TPP1-TIN2 Interaction, Influences Telomere Length, and Confers Colorectal Cancer Risk in Chinese Population. Cancer Epidemiol. Biomarkers Prev. 27, 1029– 1035.
- 51. Zou, D., Lou, J., Ke, J., Mei, S., Li, J., Gong, Y., Yang, Y., Zhu, Y., Tian, J., Chang, J., et al. (2018). Integrative expression quantitative trait locus-based analysis of colorectal cancer identified a functional polymorphism regulating SLC22A5 expression. Eur. J. Cancer 93, 1–9.
- Lin, D.Y., Zeng, D., and Millikan, R. (2005). Maximum likelihood estimation of haplotype effects and haplotype-environment interactions in association studies. Genet. Epidemiol. 29, 299–312.
- 53. Wacholder, S., Chanock, S., Garcia-Closas, M., El Ghormli, L., and Rothman, N. (2004). Assessing the probability that a positive report is false: an approach for molecular epidemiology studies. J. Natl. Cancer Inst. 96, 434–442.
- 54. He, J., Zou, Y., Liu, X., Zhu, J., Zhang, J., Zhang, R., Yang, T., and Xia, H. (2018). Association of Common Genetic Variants in Pre-microRNAs and Neuroblastoma Susceptibility: A Two-Center Study in Chinese Children. Mol. Ther. Nucleic Acids 11, 1–8.
- Carithers, L.J., and Moore, H.M. (2015). The Genotype-Tissue Expression (GTEx) Project. Biopreserv. Biobank. 13, 307–308.