

Comparative analysis of palmitoylation sites of serotonin (5-HT) receptors in vertebrates

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Funding information

Ministry of Education, Culture, Sports,
Science and Technology of Japan (MEXT)
Grant/Award Number: 16K07078; Japan
Agency for Medical Research and
Development (AMED) Grant/Award
Number: JP17gm5910009; Takeda Science
Foundation; Mitsubishi Foundation; Brain
Science Foundation; Suzuken Memorial
Foundation; Astellas Foundation for
Research on Metabolic Disorders

Abstract

Background: In the vertebrate central nervous system as well as in the periphery, serotonin, also known as 5-hydroxytryptamine (5-HT), function as a neurotransmitter, a hormone or a mitogen. 5-HT receptors are composed of 7 family 5-HT₁₋₇ receptors, comprising of 14 structurally and pharmacologically distinct 5-HT receptor subtypes. Previous experimental studies showed that mouse 5-HT_{1A}, 5-HT₄ and 5-HT₇ receptors are regulated by post-translational protein palmitoylation, the reversible attachment of the lipid palmitate to intracellular cysteine residues. Here, we further focused on conservation of these putative palmitoylation sites found in vertebrate 5-HT receptor orthologs.

Methods and Results: Analysis of sequence databases provides evidence to suggest that palmitoylation sites of these 5-HT receptors have been extremely conserved in the vertebrate lineages from jawless fishes to human, in spite of the divergence of 5-HT_{1A}, 5-HT₄ or 5-HT₇ receptors full-length amino acid sequences during molecular evolution.

Conclusion: Our findings mean that dynamic regulation of 5-HT receptors made possible by reversible post-translational protein palmitoylation may be critical for refined functions of the vertebrate serotonergic systems.

KEYWORDS

5-hydroxytryptamine, orthologs, post-translational protein palmitoylation, receptor, serotonin, vertebrate

1 | INTRODUCTION

Serotonin, also chemically known as 5-hydroxytryptamine (5-HT), is an important monoamine neurotransmitter and a local hormone, which diversely acts in the vertebrate central nervous system as well as in the various peripheral organs.¹ Serotonergic dysfunction induces multiple psychiatric disorders, including mood disorders, such as major depressive disorder and bipolar disorder.²⁻⁵ Evolutionally, serotonin is widely used in invertebrates, plants, and even in unicellular organism.^{1,6} In mammals, serotonin effects are mediated via 7 family 5-HT₁₋₇ receptors, comprising of 14 structurally and pharmacologically distinct subtypes: 5-HT_{1A}, 5-HT_{1B}, 5-HT_{1D},

5-HT_{1E}, 5-HT_{1F}, 5-HT_{2A}, 5-HT_{2B}, 5-HT_{2C}, 5-HT₃, 5-HT₄, 5-HT_{5A}, 5-HT_{5B}, 5-HT₆, and 5-HT₇.⁷ These 5-HT receptors are classified into G protein-coupled receptors (GPCRs) with the exception of the 5-HT₃ receptor, which is a ligand-gated sodium/potassium cation channel.^{7,8} All GPCR-type 5-HT receptor isoforms have evolutionarily conserved 7 transmembrane regions with the extracellular N-terminal domain and the C-terminal cytoplasmic domain.

One key modification of mammalian 5-HT receptors is the reversible attachment of the lipid palmitate to intracellular cysteine residues. Previous biochemical studies showed that 4 5-HT receptor subtypes, mouse 5-HT_{1A}, 5-HT_{1B}, 5-HT₄, and 5-HT₇ receptors are palmitoylated within their C-terminal intracellular region like many other GPCRs.^{9,10}

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This process, post-translational protein palmitoylation, acts as a sticky "tag" that can direct channels and receptors to specific regions of the plasma membrane, or to specific intracellular membranes or vesicles.^{11–14} Genetic evidence strongly links impaired palmitoylation to abnormal mammalian brain development and/or function, including human neuropsychiatric disorders.^{15–20} Biochemical experiments have already determined the palmitoylation sites of mouse 5-HT_{1A} receptor,²¹ 5-HT₄ receptor,^{22,23} and 5-HT₇ receptor.²⁴

In this report, we further focused on conservation of 5-HT_{1A}, 5-HT₄, and 5-HT₇ receptor palmitoylation sites found in their vertebrate orthologs. Analysis of sequence databases provides evidence for the acquisition and conservation of palmitoylation mechanism in these 5-HT receptor regulations during vertebrate evolution.

2 | METHODS

For analysis of the 5-HT receptor orthologs, currently available protein sequences, cDNA sequences, expressed sequence tags (ESTs), and genomic sequences were obtained by searching the National Center for Biotechnology Information (NCBI) databases, GenBank, EST banks, elephant shark genome project (<http://esharkgenome.imcb.a-star.edu.sg/>), Joint Genome Institute (<http://genome.jgi-psf.org/>), and the Ensembl database (<http://www.ensembl.org/>) by sequence homologies. Amino acid sequence alignments among 5-HT receptor orthologs were performed using a Basic Local Alignment Search Tool (BLASTP) search of the NCBI (<https://blast.ncbi.nlm.nih.gov/> with BLOSUM62). For the evaluation of sequence homology between humans and animal species in each class, full-length data of 5-HT_{1A}, 5-HT_{4(a)}, and 5-HT_{7(a)} receptor orthologs were used for calculation (Table 1). Both splice variants, 5-HT_{4(a)} and 5-HT_{7(a)} receptors, uniquely possess variant-specific palmitoylation sites in addition to the common palmitoylation sites of 5-HT₄ and 5-HT₇ receptor variants, respectively (for detail, see Figure 3A,B). To analyze the conservation and exchange of all palmitoylation sites, we select 5-HT_{4(a)} and 5-HT_{7(a)} receptors, rather than other 5-HT₄ and 5-HT₇ variants, for the homology comparison. Average of all identity percentages across 5-HT receptor orthologs is shown in the text.

A full phylogenetic tree of 5-HT_{1A} orthologs can be found at: http://www.ensembl.org/Homo_sapiens/Gene/Compara_Tree?db=c&ore;g=ENSG00000178394;r=5:63960356-63962507.

A full phylogenetic tree of 5-HT₄ orthologs can be found at: http://www.ensembl.org/Homo_sapiens/Gene/Compara_Tree?db=c&ore;g=ENSG00000164270;r=5:148451032-148677235.

A full phylogenetic tree of 5-HT₇ orthologs can be found at: http://www.ensembl.org/Homo_sapiens/Gene/Compara_Tree?db=c&ore;g=ENSG00000148680;r=10:90740823-90857698.

3 | RESULTS

Recent expansive progress in genome analyses revealed that many vertebrate species possess 5-HT_{1A}, 5-HT₄, and 5-HT₇ receptor

orthologs (Figure 1). Generally speaking, structurally or functionally important amino acid residues are conserved during molecular evolution against mutation pressure. The homology comparison of full-length amino acid sequence of 5-HT_{1A} receptor orthologs showed ~93% identity among mammalian species, ~80% identity between humans and birds, ~76% identity between humans and reptiles, ~74% identity between humans and amphibians, ~71% identity between humans and ray-finned fishes (the class Actinopterygii) (Figure 2A). The homology comparison of full-length amino acid sequence of 5-HT_{4(a)} (or 5-HT_{4 isoform X2}) receptor orthologs showed ~96% identity among mammalian species, ~87% identity between humans and birds, ~88% identity between humans and reptiles, ~85% identity between humans and amphibians, ~76% identity between humans and ray-finned fishes (Figure 2B). The homology comparison of full-length amino acid sequence of 5-HT_{7(a)} receptor orthologs showed ~95% identity among mammalian species, ~83% identity between humans and birds, ~86% identity between humans and reptiles, ~77% identity between humans and amphibians. 5-HT_{7(a)}-type splice variants have not been identified for fishes (Figure 2C). Random mutations are observed all over vertebrate 5-HT receptor sequences during vertebrate evolution. Sequence alignment among 5-HT_{1A}, 5-HT₄, or 5-HT₇ receptor orthologs revealed that cysteine residues at the putative palmitoylation sites are extremely conserved across vertebrates. (All currently available sequence data are shown in Table 1.) Details for each 5-HT receptor subtype are described below. While there is no strict consensus rule in amino acid sequence around known palmitoylated cysteines, hydrophobic residues (Ile, Leu, Val, and Phe) and positively charged basic residues (Arg and Lys) often locate around the palmitoylation sites, which might contribute to membrane binding.^{11,25} Actually, hydrophobic residues exist around the putative palmitoylation sites in vertebrate 5-HT_{1A}, 5-HT₄, and 5-HT₇ receptor orthologs (Table 1). Arginine and lysine residues are notably detected around the putative palmitoylation sites in most vertebrate 5-HT_{1A} and 5-HT₇ receptor orthologs. Concerning 5-HT₄ receptors, arginine, lysine, or histidine residues locate around the putative palmitoylation sites in almost all cartilaginous and ray-finned fishes, but not in coelacanth and tetrapods.

3.1 | 5-HT_{1A} receptor palmitoylation sites in various vertebrate species

A previous work demonstrated that mouse 5-HT_{1A} receptor is doubly palmitoylated at its C-terminal cysteine residues Cys417 and Cys420.²¹ Stable palmitoylation is required for the receptor coupling to the G_i protein that regulates the inhibition of downstream cAMP formation. Sequence comparison of total 216 vertebrate species from jawless fish (sea lamprey) to humans showed that the first palmitoylation site corresponding to mouse/human 5-HT_{1A} receptor Cys417 is completely conserved in all vertebrate species without exception (Table 1). Concerning another palmitoylation site Cys420, 5-HT_{1A} receptor orthologs of all tetrapods, non-tetrapod lobe-finned fish (the class Sarcopterygii), and cartilaginous fish, namely mammals, birds, reptiles, amphibians, coelacanth, and whale shark possess a

TABLE 1 The BLAST alignments of palmitoylation sites in vertebrate 5-HT_{1A}, 5-HT₄, and 5-HT₇ receptor orthologs

Species (common name)		Species (Latin name)	identity 5-HT _{1A} (%) C417C420	identity 5-HT ₄ C328C329 (%)	5-HT ₄ (a) C386	identity 5-HT ₇ (%) C401	5-HT ₇ (a) A435C438	Superorder	Subclass
Phylum: Chordata		Subphylum: Vertebrata							
Superclass: Gnathostomata		Class: Mammalia							
Order: Primates	Human	<i>Homo sapiens</i>	100	100	IILCCDDE	LESFCF	100	LLQCQYR	QNADYCRKK
	Bonobo	<i>Pan paniscus</i>	99	99	IILCCDDE	LESFCF	100	LLQCQYR	QNADYCRKK
	Chimpanzee	<i>Pan troglodytes</i>	99	99	IILCCDDE	LESFCF	100	LLQCQYR	QNADYCRKK
	Western lowland gorilla	<i>Gorilla gorilla gorilla</i>	99	99	IILCCDDE	N. D.	100	LLQCQYR	QNADYCRKK
	Sumatran orangutan	<i>Pongo abelii</i>	99	99	IILCCDDE	N. D.	99	LLQCQYR	QNADYCRKK
	White-cheeked crested gibbon	<i>Nomascus leucogenys</i>	99	99	IILCCDDE	LESFCF	99	LLQCQYR	N. D.
	Rhesus monkey	<i>Macaca mulatta</i>	98	98	IILCCDDE	LESFCF	99	LLQCQYR	QNADYCRKK
	Crab-eating macaque	<i>Macaca fascicularis</i>	98	98	IILCCDDE	LESFCF	99	LLQCQYR	QNADYCRKK
	Pig-tailed macaque	<i>Macaca nemestrina</i>	98	98	IILCCDDE	LESFCF	99	LLQCQYR	QNADYCRKK
	Japanese macaque	<i>Macaca fuscata</i>	98	98	IILCCDDE	LESFCF	N. D.	N. D.	N. D.
	Celebes crested macaque	<i>Macaca nigra</i>	98	98	IILCCDDE	N. D.	N. D.	N. D.	N. D.
	Golden snub-nosed monkey	<i>Rhinopithecus roxellana</i>	98	98	IILCCDDE	LESFCF	99	LLQCQYR	QNADYCRKK
	Black snub-nosed monkey	<i>Rhinopithecus bieti</i>	98	98	IILCCDDE	LESFCF	98	LLQCQYR	QNADYCRKK
	Angola colobus	<i>Colobus angolensis palliatus</i>	98	98	IILCCDDE	N. D.	98	LLQCQYR	N. D.
	Green monkey	<i>Chlorocebus sabaeus</i>	98	98	IILCCDDE	LESFCF	99	LLQCQYR	QNADYCRKK
	African green monkey	<i>Chlorocebus aethiops</i>	98	98	IILCCDDE	N. D.	N. D.	N. D.	N. D.
	Drill	<i>Mandrillus leucophaeus</i>	98	98	IILCCDDE	LESFCF	99	LLQCQYR	QNADYCRKK
	Sooty mangabey	<i>Cercocebus atys</i>	98	98	IILCCDDE	N. D.	99	LLQCQYR	QNADYCRKK
	Olive baboon	<i>Papio anubis</i>	98	98	IILCCDDE	N. D.	98	LLQCQYR	QNADYCRKK
	Ma's night monkey	<i>Aotus nancymariae</i>	97	97	IILCCDDE	LESFCF	99	LLQCQYR	QTTTYCRKK
	Common marmoset	<i>Callithrix jacchus</i>	97	97	IFKCKFCRQ	98	99	LLQCQYR	QTTTYCRKK
	White-headed capuchin	<i>Cebus capucinus imitator</i>	96	96	IILCCDDE	LESFCF	98	LLQCQYR	QTTTYCRKK
	Bolivian squirrel monkey	<i>Saimiri boliviensis boliviensis</i>	96	96	IILCCDDE	LESFCF	98	LLQCQYR	QTTTYCRKK
	Philippine tarsier	<i>Carlito syrichta</i>	96	96	IILCCDDE	LESFCF	97	LLQCQYR	QNSDYCRKK
	Coquerel's sifaka	<i>Propithecus coquereli</i>	93	93	IILCCDDE	LESFCF	97	LLQCQYR	QNSDYCRKK
	Northern greater galago	<i>Otolemur garnettii</i>	94	94	IILCCDDE	LESFCF	97	LLQCQYR	QNSDYCRKK
Order: Dermoptera	Sunda flying lemur	<i>Galeopterus variegatus</i>	94	94	IILCCDDE	N. D.	95	LLQCQYR	QNSDYCRKK
	Gray mouse lemur	<i>Microcebus murinus</i>	93	93	IILCCDDE	LESFCF	97	LLQCQYR	QNSDYCRKK
Order: Scandentia	Chinese tree shrew	<i>Tupaia chinensis</i>	93	93	IILCCDDE	LESFCF	98	LLQCQYR	QNSDYCRKK
	Desu	<i>Octodon degus</i>	91	91	IILCCDDE	LESFCF	94	LLQCQYR	QNSDYCRKK
Order: Rodentia	Long-tailed chinchilla	<i>Chinchilla lanigera</i>	92	92	IILCCDDE	N. D.	95	LLQCQYR	QNSDYCRKK
	Guinea pig	<i>Cavia porcellus</i>	92	92	IILCCDDE	N. D.	92	LLQCQYR	QNSDYCRKK
	Naked mole rat	<i>Heterocephalus glaber</i>	90	90	IILCCDDE	LESFCF	95	LLQCQYR	QNSDYCRKK
	Damara mole-rat	<i>Fukomys damarensis</i>	90	97	IILCCDDE	LESFCF	95	LLQCQYR	QNSDYCRKK
	North American beaver	<i>Castor canadensis</i>	93	93	IILCCDDE	N. D.	97	LLQCQYR	QNSDYCRKK
	Ord's kangaroo rat	<i>Dipodomys ordii</i>	90	90	IILCCDDE	N. D.	95	LLQCQYR	QNSDYCRKK
	Upper Galilee mountains blind mole rat	<i>Nannospalax gallii</i>	92	92	IILCCDDE	N. D.	96	LLQCQYR	QSSDYCRKK
	House mouse	<i>Mus musculus</i>	88	88	IILCCDDE	LESFCF	95	LLQCQYR	QNSDHCGRK
	Ryukyu mouse	<i>Mus caroli</i>	88	88	IILCCDDE	LESFCF	95	LLQCQYR	QNSDHCGRK
	Gairdner's shrewmouse	<i>Mus pahari</i>	87	87	IILCCDDE	LESFCF	95	LLQCQYR	QNSDHCGRK
	Mongolian jird	<i>Meriones unguiculatus</i>	91	91	IILCCDDE	N. D.	95	LLQCQYR	QNSDHCGRK
	Norway rat	<i>Rattus norvegicus</i>	90	94	IILCCDDE	LESFCF	95	LLQCQYR	QNSDHCGRK
	Prairie deer mouse	<i>Peromyscus maniculatus bairdii</i>	91	91	VILCCDDE	LESFCF	95	LLQCQYR	QNSDHCGRK
	Prairie vole	<i>Microtus ochrogaster</i>	90	90	IILCCDDE	N. D.	94	LLQCQYR	QNSDHCGRK
	Chinese hamster	<i>Cricetus griseus</i>	90	90	IILCCDDE	N. D.	94	LLQCQYR	QNSDHCGRK
	Golden hamster	<i>Mesocricetus auratus</i>	90	90	IILCCDDE	N. D.	94	LLQCQYR	QNSDHCGRK
	Lesser Egyptian jerboa	<i>Jaculus jaculus</i>	89	89	IILCCDDE	LESFCF	95	LLQCQYR	QNSDYCRKK
	Thirteen-lined ground squirrel	<i>Ictidomys tridecemlineatus</i>	92	92	IILCCDDE	LESFCF	96	LLQCQYR	N. D.
	Alpine marmot	<i>Marmota marmota marmota</i>	93	93	IILCCDDE	LESFCF	96	LLQCQYR	N. D.
Order: Lagomorpha	Rabbit	<i>Oryctolagus cuniculus</i>	94	94	IILCCDDE	LESFCF	94	LLQCQYR	QNSDYCRKK
	American pika	<i>Ochotona princeps</i>	91	91	IILCCDDE	N. D.	94	LLQCQYR	QNSDYCRKK
Order: Carnivora	Cat	<i>Felis catus</i>	94	94	IILCCDDE	N. D.	95	LLQCQYR	N. D.
	Amur tiger	<i>Panthera tigris altaica</i>	94	94	IILCCDDE	N. D.	95	LLQCQYR	N. D.
	Leopard	<i>Panthera pardus</i>	94	94	IILCCDDE	LESFCF	96	LLQCQYR	QNSDYCRKK
	Cheetah	<i>Acinonyx jubatus</i>	93	93	IILCCDDE	N. D.	98	LLQCQYR	N. D.
	Dog	<i>Canis lupus familiaris</i>	92	92	IILCCDDE	LESFCF	98	LLQCQYR	QNSDYCRKK
	Polar bear	<i>Ursus maritimus</i>	95	95	IILCCDDE	N. D.	97	LLQCQYR	QNSDYCRKK
	Giant panda	<i>Ailuropoda melanoleuca</i>	95	95	IILCCDDE	LESFCF	95	LLQCQYR	QNSDYCRKK
	Ferret	<i>Mustela putorius furo</i>	94	94	IILCCDDE	LESFCF	96	LLQCQYR	QNSDYCRKK
	American mink	<i>Neovison vison</i>	N. D.	N. D.	N. D.	N. D.	96	LLQCQYR	QNSDYCRKK
	Pacific walrus	<i>Odobenus rosmarus divergens</i>	95	95	IILCCDDE	LESFCF	97	LLQCQYR	N. D.
	Weddell seal	<i>Leptonychotes weddellii</i>	95	95	IILCCDDE	N. D.	97	LLQCQYR	N. D.
	Hawaiian monk seal	<i>Neomonachus schauinslandi</i>	94	94	IILCCDDE	N. D.	96	LLQCQYR	QNSDYCRKK
Order: Pholidota	Malayan pangolin	<i>Manis javanica</i>	91	91	IILCCDDE	N. D.	94	LLQCQYR	QNSDYCRKK
Order: Perissodactyla	Western white rhinoceros	<i>Ceratotherium simum simum</i>	93	93	IILCCDDE	LESFCF	96	LLQCQYR	QNSDYCRKK
	Przewalski's horse	<i>Equus przewalskii</i>	93	93	IILCCDDE	N. D.	96	LLQCQYR	QNSDYCRKK
	Horse	<i>Equus caballus</i>	94	94	IILCCDDE	N. D.	93	LLQCQYR	QNSDYCRKK
	Donkey	<i>Equus asinus</i>	N. D.	N. D.	N. D.	N. D.	96	LLQCQYR	QNSDYCRKK
Order: Chiroptera	Brandt's bat	<i>Myotis brandtii</i>	90	90	IILCCDDE	N. D.	97	LLQCQYR	QKSDCRKK
	David's myotis	<i>Myotis davidii</i>	89	89	IILCCDDE	LESFCF	95	LLQCQYR	QKSDCRKK
	Little brown bat	<i>Myotis lucifugus</i>	90	90	IILCCDDE	LESFCF	83	LLQCQYR	QKSDCRKK
	Great roundleaf bat	<i>Hipposideros armiger</i>	92	92	IILCCDDE	N. D.	96	LLQCQYR	QNSDYCRKK
	Chinese rufous horseshoe bat	<i>Rhinolophus sinicus</i>	90	90	IILCCDDE	LESFCF	96	LLQCQYR	QNSDYCRKK
	Big brown bat	<i>Eptesicus fuscus</i>	92	92	IILCCDDE	LESFCF	95	LLQCQYR	QKSDCRKK
	Natal long-fingered bat	<i>Miniopterus natalensis</i>	88	88	IILCCDDE	LESFCF	96	LLQCQYR	QKSDCRKK
	Large flying fox	<i>Pteropus vampyrus</i>	94	94	IILCCDDE	LESFCF	95	LLQCQYR	QNSDYCRKK
	Black flying fox	<i>Pteropus alecto</i>	94	94	IILCCDDE	LESFCF	95	LLQCQYR	QNSDYCRKK
	Egyptian fruit bat	<i>Rousettus aegyptiacus</i>	93	93	IILCCDDE	N. D.	96	LLQCQYR	QNSDYCRKK
Order: Cetartiodactyla	Common minke whale	<i>Balaenoptera acutorostrata scammoni</i>	92	92	IILCCDDE	N. D.	95	LLQCQYR	QNSDYRRKK
	Bottlenose dolphin	<i>Tursiops truncatus</i>	93	93	IILCCDDE	LESFCF	96	LLQCQYR	QnsdyCrkK
	Killer whale	<i>Orcinus orca</i>	93	93	IILCCDDE	LESFCF	96	LLQCQYR	QnsdyCrkK
	Yangtze River dolphin	<i>Lipotes vexillifer</i>	92	92	IILCCDDE	LESFCF	95	LLQCQYR	QNSDYCRKK
	Sperm whale	<i>Physeter catodon</i>	92	92	IILCCDDE	N. D.	96	LLQCQYR	QnsdyCrkK
	Arabian camel	<i>Camelus dromedarius</i>	92	92	IILCCDDE	LESFCF	94	LLQCQYR	QNSDYCRKK
	wild Bactrian camel	<i>Camelus ferus</i>	93	93	IILCCDDE	LESFCF	94	LLQCQYR	QNSDYCRKK
	Bactrian camel	<i>Camelus bactrianus</i>	93	93	IILCCDDE	N. D.	94	LLQCQYR	QNSDYCRKK
	Cattle	<i>Bos taurus</i>	92	92	IILCCDDE	LESFCF	94	LLQCQYR	QnsdyCrkK
	Wild yak	<i>Bos mutus</i>	92	92	IILCCDDE	LESFCF	96	LLQCQYR	QnsdyCrkK
	Zebu	<i>Bos indicus</i>	92	92	IILCCDDE	N. D.	95	LLQCQYR	N. D.
	Water buffalo	<i>Bubalus bubalis</i>	91	91	IILCCDDE	LESFCF	94	LLQCQYR	QnsdyCrkK
	American bison	<i>Bison bison</i>	92	92	IILCCDDE	LESFCF	94	LLQCQYR	QNSDYCRKK
	White-tailed deer	<i>Odocoileus virginianus texanus</i>	92	92	IILCCDDE	N. D.	94	LLQCQYR	QNSDYCRKK
	pig (wild boar)	<i>Sus scrofa</i>	91	91	IILCCDDE	N. D.	96	LLQCQYR	QKSDYCRKK
	Goat	<i>Capra hircus</i>	91	91	IILCCDDE	LESFCF	95	LLQCQYR	N. D.
	Sheep	<i>Ovis aries</i>	92	92	IILCCDDE	LESFCF	95	LLQCQYR	N. D.
	Mouflon	<i>Ovis aries musimon</i>	92	92	IILCCDDE	LESFCF	95	LLQCQYR	N. D.
	Chiru	<i>Pantholops hodgsonii</i>	92	92	IILCCDDE	N. D.	96	LLQCQYR	QnsdyCrkK
	Alpaca	<i>Vicugna pacos</i>	93	93	IILCCDDE	N. D.	95	LLQCQYR	QnsdyCrkK
Order: Erinaceomorpha	Western European hedgehog	<i>Eriacus europaeus</i>	92	92	IILCCDDE	N. D.	96	LLQCQYR	QNSDYCRKK
Order: Soricomorpha	Star-nosed mole	<i>Condylura cristata</i>	92	92	IILCCDDE	LESFCF	96	LLQCQYR	QNSDYCRKK
	European shrew	<i>Sorex araneus</i>	90	90	IILCCDDE	N. D.	96	LLQCQYR	N. D.
Order: Afrosoricida	Cape golden mole	<i>Chrysochloris asiatica</i>	93	93	IILCCDDE	LESFCF	96	LLQCQYR	N. D.
	Lesser hedgehog tenrec	<i>Echinops telfairi</i>	89	89	IILCCDDE	N. D.	96	LLQCQYR	N. D.
Order: Macroscelidea	Cape elephant shrew	<i>Elephantulus edwardii</i>	90	90	IILCCDDE	N. D.	96	LLQCQYR	N. D.
Order: Tubulidentata	Aardvark	<i>Orycteropus afer afer</i>	92	92	IILCCDDE	LESFCF	96	LLQCQYR	N. D.
Order: Proboscidea	African savanna elephant	<i>Loxodonta africana</i>	90	90	IILCCDDE	LESFCF	96	LLQCQYR	N. D.
Order: Sirenia	Florida manatee	<i>Trichechus manatus latirostris</i>	92	92	IILCCDDE	LESFCF	96	LLQCQYR	N. D.
Order: Cingulata	Nine-banded armadillo	<i>Dasyops novemcinctus</i>	90	90	IILCCDDE	LESFCF	96	LLQCQYR	QNSDYSRKK
Order: Didelphimorphia	Gray short-tailed opossum	<i>Monodelphis domestica</i>	84	84	IILCCDDE	N. D.	84	LLQCQYR	QNSDYCRKK
Order: Dasyuromorphia	Tasmanian devil	<i>Sarcophilus harrisii</i>	80	80	IILCCDDE	N. D.	85	LLQCQYR	QNSDYSRKK
Order: Diprotodontia	Koala	<i>Phascolarctos cinereus</i>	83	83	IILCCDDE	N. D.	84	LLQCQYR	QNSDYSRKK
Order: Monotremata	Platypus	<i>Ornithorhynchus anatinus</i>	57	57	IILCCDDE	N. D.	80	LLQCQYR	RNSDSRKK

(Continues)

TABLE 1 (Continued)

Superclass: Gnathostomata class: Aves							
Order: Psittaciformes	Kea	<i>Nestor notabilis</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
	Budgerigar	<i>Melopsittacus undulatus</i>	80	IILCCGDE	QESCF	LLQCRYR	N. D.
	Turquoise-fronted amazon	<i>Amazona aestiva</i>	79	IILCCGDE	N. D.	86	LLQCRYR RSDFSREK
Order: Passeriformes	American crow	<i>Corvus brachyrhynchos</i>	83	N. D.	80	IILCCGDE	QESCF 86
	Hooded crow	<i>Corvus corax corax</i>	80	IILCCGDE	QESCF	87	LLQCRYR RSDFSREK
	Canary	<i>Canis canaria</i>	81	IILCCGDE	N. D.	77	LLQCRYR RSDFSREK
	Collared flycatcher	<i>Ficedula albicollis</i>	79	IILCCGDE	QESCF	80	LLQCRYR RSDFSREK
	Ground tit	<i>Pseudopodoces humilis</i>	81	IILCCGDE	QESCF	80	LLQCRYR N. D.
	Great tit	<i>Parus major</i>	81	IILCCGDE	QESCF	81	LLQCRYR RSDFSREK
	Medium ground-finch	<i>Geospiza fortis</i>	73	IILCCGDE	QESCF	81	LLQCRYR N. D.
	Zebra finch	<i>Taeniopygia guttata</i>	79	IILCCGDE	N. D.	81	LLQCRYR N. D.
	Society finch	<i>Lonchura striata domestica</i>	79	IILCCGDE	QESCF	81	LLQCRYR RSDFSREK
	Rifleman	<i>Acanthisitta chloris</i>	N. D.	IILCCGDE	N. D.	83	LLQCRYR N. D.
	White-throated sparrow	<i>Zonotrichia albicollis</i>	80	IILCCGDE	QESCF	83	LLQCRYR RSDFSREK
	Golden-collared manakin	<i>Mniotilta varia</i>	79	IILCCGDE	QESCF	83	LLQCRYR N. D.
	Blue-crowned manakin	<i>Lepidothrix coronata</i>	79	IILCCGDE	QESCF	83	LLQCRYR N. D.
	Common starling	<i>Sturnus vulgaris</i>	80	IILCCGDE	N. D.	81	LLQCRYR RSDFSREK
	Blackbird	<i>Turdus merula</i>	80	IILCCGDE	N. D.	N. D.	N. D.
Order: Falconiformes	Saker falcon	<i>Falco cherrug</i>	81	IILCCGDE	QESCF	LLQCRYR	N. D.
	Peregrine falcon	<i>Falco peregrinus</i>	80	IILCCGDE	QESCF	LLQCRYR	N. D.
Order: Bucconiformes	Rhinoceros hornbill	<i>Buceros rhinoceros silvestris</i>	80	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Piciformes	Downy woodpecker	<i>Picoides pubescens</i>	80	IILCCGDE	QESCF	LLQCRYR	N. D.
Order: Coraciiformes	Carmine bee-eater	<i>Merops rubicus</i>	N. D.	IILCCGDE	N. D.	85	LLQCRYR RSDFSREK
Order: Cariamiformes	Red-legged seriema	<i>Cariama cristata</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Accipitriformes	White-tailed eagle	<i>Haliaeetus albicilla</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
	Bald eagle	<i>Haliaeetus leucocephalus</i>	81	IILCCGDE	N. D.	LLQCRYR	N. D.
	Golden eagle	<i>Aquila chrysaetos canadensis</i>	81	IILCCGDE	QESCF	81	LLQCRYR RSDFSREK
	Turkey vulture	<i>Cathartes aura</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Strigiformes	Barn owl	<i>Tyto alba</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Coliiformes	Speckled mousebird	<i>Colius striatus</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Leptosomatiformes	Cuckoo roller	<i>Leptosomus discolor</i>	80	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Trogoniformes	Bar-tailed trogon	<i>Apaloderma vittatum</i>	64	IILCCGDE	N. D.	88	LLQCRYR RSDFSREK
Order: Gaviformes	Red-throated loon	<i>Gavia stellata</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Sphenisciformes	Emperor penguin	<i>Aptenodytes forsteri</i>	80	IILCCGDE	HESCF	LLQCRYR	N. D.
	Adelie penguin	<i>Pygoscelis adeliae</i>	77	N. D.	IILCCGDE	N. D.	LLQCRYR N. D.
Order: Procellariiformes	Northern fulmar	<i>Fulmarus glacialis</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Pelecaniformes	Dalmatian pelican	<i>Pelecanus crispus</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
	Crested ibis	<i>Nipponia nippon</i>	79	IILCCGDE	N. D.	LLQCRYR	N. D.
	White-throated tinamou	<i>Tinamus guttatus</i>	85	N. D.	83	IILCCGDE	QESCF LLQCRYR N. D.
Order: Ciconiiformes	Little egret	<i>Egretta garzetta</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Suliformes	Great cormorant	<i>Phalacrocorax carbo</i>	N. D.	IILCCGDE	N. D.	87	LLQCRYR RSDFSREK
Order: Caprimulgiformes	Chuk-wills-widow	<i>Caprimulgus carolinensis</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Apodiformes	Chimney swift	<i>Chaetura pelagica</i>	N. D.	IILCCGDE	N. D.	87	LLQCRYR N. D.
	Anna's hummingbird	<i>Calypte anna</i>	83	IILCCGDE	QESCF	89	LLQCRYR N. D.
Order: Podicipediformes	Great crested grebe	<i>Podiceps cristatus</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Phoenicopteriformes	American flamingo	<i>Phoenicopterus ruber ruber</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Pteroclidiformes	Yellow-throated sandgrouse	<i>Pterocles gutturalis</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Columbiformes	Rock pigeon	<i>Columba livia</i>	79	IILCCGDE	QESCF	LLQCRYR	N. D.
	Band-tailed pigeon	<i>Patagioenas fasciata monilis</i>	80	IILCCGDE	QESCF	LLQCRYR	N. D.
Order: Gruiformes	Grey crowned crane	<i>Balearica regulorum gibbericeps</i>	70	IILCCGDE	N. D.	LLQCRYR	N. D.
	Eurypyga helias	<i>Eurypyga helias</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
	Brown mesite	<i>Mesitornis unicolor</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Otidiformes	Macqueen's bustard	<i>Chlamydotis macquenei</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Opisthocomiformes	Hoatzin	<i>Opisthocomus hoazin</i>	85	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Charadriiformes	Ruff	<i>Calidris pugnax</i>	81	IILCCGDE	QESCF	81	LLQCRYR RSDFSREK
	Killdeer	<i>Charadrius vociferus</i>	68	IILCCGDE	QESCF	89	LLQCRYR N. D.
Order: Cuculiformes	Common cuckoo	<i>Cuculus canorus</i>	80	IILCCGDE	QESCF	88	LLQCRYR N. D.
Order: Musophagiformes	Red-crested turaco	<i>Tauraco erythrolophus</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
Order: Galliformes	Scaled quail	<i>Callipepla squamata</i>	81	IILCCGDE	N. D.	N. D.	LLQCRYR N. D.
	Japanese quail	<i>Coturnix japonica</i>	80	IILCCGDE	QESCF	80	LLQCRYR RSDFSREK
	Northern bobwhite	<i>Coturnix coturnix</i>	81	IILCCGDE	N. D.	LLQCRYR	N. D.
	Chicken	<i>Gallus gallus</i>	80	IILCCGDE	QESCF	80	LLQCRYR RSDFSREK
	Turkey	<i>Meleagris gallopavo</i>	81	IILCCGDE	QESCF	87	LLQCRYR RSDFSREK
	Helmeted guineafowl	<i>Numida meleagris</i>	80	IILCCGDE	QESCF	89	LLQCRYR N. D.
Order: Anseriformes	Mallard	<i>Anas platyrhynchos</i>	80	IILCCGDE	QESCF	88	LLQCRYR RSDFSREK
	Swan goose	<i>Anser cygnoides</i>	81	IILCCGDE	QESCF	89	LLQCRYR RSDFSREK
	Chinese geese	<i>Anser cygnoides domesticus</i>	89	IILCCGDE	QESCF	87	LLQCRYR RSDFSREK
Order: Struthioniformes	Southern ostrich	<i>Struthio camelus australis</i>	N. D.	IILCCGDE	QESCF	89	LLQCRYR N. D.
Order: Tinamiformes	White-throated tinamou	<i>Tinamus guttatus</i>	85	N. D.	89	IILCCGDE	LLQCRYR N. D.
Order: Apterygiformes	Kiwi	<i>Apteryx australis mantelli</i>	82	N. D.	89	IILCCGDE	QESCF LLQCRYR N. D.
Superclass: Gnathostomata class: Reptilia							
Order: Crocodylia	Saltwater crocodile	<i>Crocodylus porosus</i>	76	IILCCGDE	QESCF	84	LLQCRYR RSDYSRKK
	American alligator	<i>Alligator mississippiensis</i>	76	IILCCGDE	QESCF	87	LLQCRYR RSDYSRKK
	Chinese alligator	<i>Alligator sinensis</i>	81	IILCCGDE	QESCF	87	LLQCRYR N. D.
	Gharial	<i>Gavialis gangeticus</i>	76	IILCCGDE	QESCF	84	LLQCRYR RSDYSRKK
Order: Testudines	Green sea turtle	<i>Chelonia mydas</i>	81	IILCCGDE	QESCF	88	LLQCRYR N. D.
	Chinese soft-shelled turtle	<i>Pelodiscus sinensis</i>	84	IILCCGDE	QESCF	89	LLQCRYR RSDYSRKK
	Painted turtle	<i>Chrysemys picta bellii</i>	81	IILCCGDE	QESCF	89	LLQCRYR N. D.
Order: Squamata	Green anole	<i>Anolis carolinensis</i>	71	IILCCGDE	QESCF	87	LLQCRYR N. D.
	Schlegel's Japanese gecko	<i>Gekko japonicus</i>	73	IILCCGDE	N. D.	81	LLQCRYR RSDYSRKK
	Central bearded dragon	<i>Pogona vitticeps</i>	73	IILCCGDE	QESCF	88	LLQCRYR RSDYSRKK
	Burmese python	<i>Python bivittatus</i>	74	IILCCGDE	N. D.	89	LLQCRYR RSDYSRKK
	King cobra	<i>Ophiophagus hannah</i>	N. D.	IILCCGDE	N. D.	LLQCRYR	N. D.
	Pit vipers	<i>Protobothrops mucrosquamatus</i>	72	IILCCGDE	QESCF	87	LLQCRYR N. D.
	Common garter snake	<i>Thamnophis sirtalis</i>	73	IILCCGDE	IILCCGDE	N. D.	LLQCRYR N. D.
Superclass: Gnathostomata class: Amphibia							
Order: Anura	Western clawed frog	<i>Xenopus (Silurana) tropicalis</i>	76	IILCCGDE	QESCF	84	LLQCRYR -TRRHSREH
	African clawed frog	<i>Xenopus laevis</i>	72	IILCCGDE	QESCF	85	LLQCRYR RTRRQSRN
	High Himalaya frog	<i>Nanorana parkeri</i>	75	IILCCGDE	QESCF	86	LLQCRYR K--HYSRD
Superclass: Gnathostomata class: Sarcopterygii							
Order: Coelacanthiformes	Coelacanth	<i>Latimeria chalumnae</i>	79	IILCCGDE	QESCF	84	LLQCRYR N. D.
Superclass: Gnathostomata class: Actinopterygii							
Order: Cypriniformes	Common carp	<i>Cyprinus carpio</i>	74	IILCCGDE	N. D.	N. D.	LLQCRYR N. D.
	Common carp	<i>Cyprinus carpio</i>	69	IILCCGDE	N. D.	N. D.	LLQCRYR N. D.
		<i>Sinocyclocheilus anshuiensis</i>	75	IILCCGDE	QESCF	77	LLQCRYR N. D.
		<i>Sinocyclocheilus anshuiensis</i>	70	IILCCGDE	QESCF	78	LLQCRYR N. D.
	Golden-line barbel	<i>Sinocyclocheilus grahami</i>	75	IILCCGDE	QESCF	78	LLQCRYR N. D.
	Golden-line barbel	<i>Sinocyclocheilus grahami</i>	74	IILCCGDE	QESCF	78	LLQCRYR N. D.
	Golden-line barbel	<i>Sinocyclocheilus grahami</i>	70	IILCCGDE	QESCF	78	LLQCRYR N. D.
		<i>Sinocyclocheilus rhinoceros</i>	75	IILCCGDE	QESCF	76	LLQCRYR N. D.
		<i>Sinocyclocheilus rhinoceros</i>	69	IILCCGDE	QESCF	76	LLQCRYR N. D.
	Zebrafish	<i>Danio rerio</i>	75	IILCCGDE	QESCF	76	LLQCRYR N. D.
	Zebrafish	<i>Danio rerio</i>	69	IILCCGDE	QESCF	76	LLQCRYR N. D.
Order: Perciformes	Burton's mouthbrooder	<i>Haplochromis burtoni</i>	75	IILCCGDE	HESCL	77	LLQCRYR N. D.
	Burton's mouthbrooder	<i>Haplochromis burtoni</i>	67	IILCCGDE	HESCL	76	LLQCRYR N. D.
	Nile tilapia	<i>Oreochromis niloticus</i>	76	IILCCGDE	HESCL	76	LLQCRYR N. D.
	Nile tilapia	<i>Oreochromis niloticus</i>	74	IILCCGDE	HESCL	76	LLQCRYR N. D.
	Mozambique tilapia	<i>Oreochromis mossambicus</i>	67	IILCCGDE	N. D.	N. D.	LLQCRYR N. D.
	Lyretail cichlid	<i>Neolamprologus brichardi</i>	76	IILCCGDE	HESCL	76	LLQCRYR N. D.
	Nyerere's Victoria cichlid	<i>Pundamilia nyererei</i>	75	IILCCGDE	N. D.	N. D.	LLQCRYR N. D.
	Nyerere's Victoria cichlid	<i>Pundamilia nyererei</i>	67	IILCCGDE	N. D.	N. D.	LLQCRYR N. D.
	Asian sea bass	<i>Lates calcarifer</i>	71	IILCCGDE	HESCL	76	LLQCRYR N. D.
	Asian sea bass	<i>Lates calcarifer</i>	68	IILCCGDE	HESCL	76	LLQCRYR N. D.
	Spiny chromis	<i>Acanthochromis polyacanthus</i>	75	IILCCGDE	IILCCGDE	N. D.	LLQCRYR N. D.
	Spiny chromis	<i>Acanthochromis polyacanthus</i>	68	IILCCGDE	IILCCGDE	N. D.	LLQCRYR N. D.

(Continues)



TABLE 1 (Continued)

	Zebra mbuna	<i>Maylandia zebra</i>	75	IIKCKFHRP		76	IILCCGRQ	HESCL		LLRCRYR	N.D.
	Zebra mbuna	<i>Maylandia zebra</i>	67	IIKCHFRCR						LLRCRYR	N.D.
	Bicolor damselfish	<i>Stegastes partitus</i>	75	IIKCKFHRP		76	IILCCGRK	HESCL		LLRCRYR	N.D.
	Bicolor damselfish	<i>Stegastes partitus</i>	67	IIKCHFRCR						LLRCRYR	N.D.
	Mudskipper	<i>Boleophthalmus pectinirostris</i>	73	IIKCKFHRP			IILCCGRK	N.D.		LLRCRYR	N.D.
	Croceine croaker	<i>Larimichthys crocea</i>	74	IIKCKFHRP		79	IILCCGRK	HESCL		LLRCRYR	N.D.
	Croceine croaker	<i>Larimichthys crocea</i>	67	IIKCHFRCR						LLRCRYR	N.D.
	Ballan wrasse	<i>Labrus bergylla</i>	72	IIKCKFHRA			N.D.	N.D.		LLRCRYR	N.D.
	Ballan wrasse	<i>Labrus bergylla</i>	68	IIKCHFRCR						LLRCRYR	N.D.
Order: Cyprinodontiformes		<i>Austrofundulus limnaeus</i>	71	IIKCKFHRA		76	IILCCGRK	RESCL		LLRCRYR	N.D.
	Mummichog	<i>Fundulus heteroclitus</i>	72	IIKCKFHRP		73	IILCCGRK	HESCL		LLRCRYR	N.D.
	Mummichog	<i>Fundulus heteroclitus</i>	66	IIKCHFPCN						LLRCRYR	N.D.
	Sheepshead minnow	<i>Cyprinodon variegatus</i>	72	IIKCKFHRP			N.D.	N.D.		LLRCRYR	N.D.
	Sheepshead minnow	<i>Cyprinodon variegatus</i>	66	IIKCHFPCP						LLRCRYR	N.D.
	Turquoise killifish	<i>Nothobranchius furzeri</i>	73	IIKCKFHRA		74	IILCCGRK	HESCL		LLRCRYR	N.D.
	Turquoise killifish	<i>Nothobranchius furzeri</i>	66	IIKCYFCKP						LLRCRYR	N.D.
	Mangrove rivulus	<i>Kryptolebias marmoratus</i>	72	IIKCKFHRP		75	IILCCGRK	RESCL		LLRCRYR	N.D.
	Mangrove rivulus	<i>Kryptolebias marmoratus</i>	66	IIKCHFRCR						LLRCRYR	N.D.
	Guppy	<i>Poecilia reticulata</i>	72	IIKCKFHRP		74	IILCCGRK	RESCL		LLRCRYR	N.D.
	Guppy	<i>Poecilia reticulata</i>	66	IIKCHFPCP						LLRCRYR	N.D.
	Amazon molly	<i>Poecilia formosa</i>	72	IIKCKFHRP		74	IILCCGRK	RESCL		LLRCRYR	N.D.
	Amazon molly	<i>Poecilia formosa</i>	66	IIKCHFPCP						LLRCRYR	N.D.
	Sailfin molly	<i>Poecilia latipinna</i>	72	IIKCKFHRP		74	IILCCGRK	RESCL		LLRCRYR	N.D.
	Atlantic molly	<i>Poecilia mexicana</i>		N.D.		74	IILCCGRK	RESCL		LLRCRYR	N.D.
	Platyfish	<i>Xiphophorus maculatus</i>	71	IIKCKFHRP			IILCCGRK	N.D.		LLRCRYR	N.D.
Order: Belontiiformes		<i>Oryzias latipes</i>	72	IIKCKFHRP		76	IILCCGRK	HESCL		LLRCRYR	N.D.
	Japanese medaka	<i>Oryzias latipes</i>	67	IIKCHFRCR						LLRCRYR	N.D.
Order: Batrachoidiformes		<i>Opsanus beta</i>	71	IIKCKFHRA			N.D.	N.D.		N.D.	N.D.
Order: Scorpaeniformes		<i>Notothenia coriiceps</i>	70	IIKCKFHRA			VILCCGDE	N.D.		LLRCRYR	N.D.
	Black rockcod	<i>Notothenia coriiceps</i>	67	IIKCHFRCR						LLRCRYR	N.D.
	False kelpfish	<i>Sebastes marmoratus</i>		N.D.			N.D.	N.D.		N.D.	N.D.
Order: Characiformes		<i>Pygocentrus nattereri</i>	69	IIKCHFRCR		77	IILCCGHR	QESCF		LLRCRYR	N.D.
	Hed piranha	<i>Pygocentrus nattereri</i>								IICCRYP	N.D.
	Red piranha	<i>Pygocentrus nattereri</i>								IICCRYP	N.D.
	Mexican tetra	<i>Astyanax mexicanus</i>								IICCRYP	N.D.
	Mexican tetra	<i>Astyanax mexicanus</i>	68	IIKCHFRCR		77	IILCCGHK	QESCF		IICCRYP	N.D.
Order: Esociformes		<i>Esox lucius</i>	70	IIKCKFHRP		77	IILCCGRK	PESCF		LLRCRYR	N.D.
	Northern pike	<i>Esox lucius</i>								IICCRYP	N.D.
Order: Clupeiformes		<i>Clupea harengus</i>	70	IVKCKNFRP IIC			IILCCGRK	N.D.		LLRCRYR	N.D.
	Atlantic herring	<i>Clupea harengus</i>								LLRCRYR	N.D.
Order: Salmoniformes		<i>Oncorhynchus mykiss</i>	71	IIKCKFHRQ		75	IILCCGRK	QESCF		LLRCRYR	N.D.
	Rainbow trout	<i>Oncorhynchus mykiss</i>								IICCRYP	N.D.
	Rainbow trout	<i>Oncorhynchus mykiss</i>	72	IIKCKFHRP			N.D.	N.D.		LLRCRYR	N.D.
	Coho salmon	<i>Oncorhynchus kisutch</i>	71	IIKCKFHRQ						IICCRYP	N.D.
	Coho salmon	<i>Oncorhynchus kisutch</i>	71	IIKCKFHRP		76	IILCCGRK	QESCF		LLRCRYR	N.D.
	Atlantic Salmon	<i>Salmo salar</i>	71	IIKCKFHRQ						IICCRYP	N.D.
	Atlantic Salmon	<i>Salmo salar</i>	73	IIKCKFHRH			N.D.	N.D.		LLRCRYR	N.D.
Order: Tetraodontiformes		<i>Takifugu rubripes</i>	66	IIKCHFRCR						LLRCRYR	N.D.
	Japanese pufferfish	<i>Takifugu rubripes</i>	70	IIKCKFHRH			N.D.	N.D.		LLRCRYR	N.D.
	Spotted green pufferfish	<i>Tetraodon nigroviridis</i>	70	IIKCKFHRH			N.D.	N.D.		LLRCRYR	N.D.
Order: Pleuronectiformes		<i>Cynoglossus semilaevis</i>	73	IIKCKFHRP			IILCCGRK	N.D.		LLRCRYR	N.D.
	Tongue sole	<i>Cynoglossus semilaevis</i>	72	IIKCKFHRP		79	IILCCGRK	SESCL		LLRCRYR	N.D.
	Olive flounder	<i>Paralichthys olivaceus</i>	66	IIKCHFRCR						LLRCRYR	N.D.
	Olive flounder	<i>Paralichthys olivaceus</i>	77	IIKCKFHRP			N.D.	N.D.		LLRCRYR	N.D.
	European flounder	<i>Platichthys flesus</i>	76	IVKCKFHRP		75	SIILCCGRK	AESCL		LLRCRYR	N.D.
Order: Gasterosteiformes		<i>Hippocampus comes</i>	68	IIKCHFRCR						LLRCRFR	N.D.
	Tiger tail seahorse	<i>Hippocampus comes</i>	66	IIKCHFRCR		78	IILCCGHQ	QESCF		LLACRYR	N.D.
Order: Siluriformes		<i>Ictalurus punctatus</i>								LI---YR	N.D.
	Channel catfish	<i>Ictalurus punctatus</i>	75	IIKCKFHRP		77	IILCCGRK	HESCL		LLRCRYR	N.D.
Order: Anguilliformes		<i>Monopterus albus</i>	68	IIKCHFRCR						LLRCRYR	N.D.
	Swamp eel	<i>Monopterus albus</i>	75	IVKCKFYR		78	IILCCGHK	QESCL		LLRCRYR	N.D.
Order: Osteoglossiformes		<i>Scleropages formosus</i>	79	IIKCKFCR		78	IILCCGQE	QESCF		LLRCRYR	N.D.
Order: Lepisosteiformes		<i>Lepisosteus oculatus</i>								LLCCRYR	N.D.
	Spotted gar	<i>Lepisosteus oculatus</i>								LLCCRYR	N.D.
	Spotted gar	<i>Lepisosteus oculatus</i>								LLCCRYR	N.D.
superclass: Gnathostomata	Class: Chondrichthyes										
Order: Orectolobiformes		<i>Rhinodon typus</i>	75	IIKCKFRCRQ			RILCCGDR	N.D.		LLRCRYR	N.D.
	Whale shark	<i>Rhinodon typus</i>								LLRCRYR	N.D.
Order: Chimaeriformes		<i>Callorhynchus milii</i>	76	IIKCKF			TILCCGNR	N.D.		LLRCRYR	N.D.
	Australian ghostshark	<i>Callorhynchus milii</i>								LLRCQFR	N.D.
	Australian ghostshark	<i>Callorhynchus milii</i>								LLRCQFR	N.D.
superclass: Agnatha	Class: Petromyzontida										
Order: Petromyzontiformes		<i>Petromyzon marinus</i>	62	IIKCKFCRQ			N.D.	N.D.		N.D.	N.D.
	Sea lamprey	<i>Petromyzon marinus</i>									
phylum: Chordata	subphylum: Cephalochordata										
Order: Amphioxiformes		<i>Branchiostoma belcheri</i>	49	ILFGR		47	RILSCWSCCDDVDN	N.D.		N.D.	N.D.
	Belcher's lancelet	<i>Branchiostoma belcheri</i>									
phylum: Chordata	subphylum: Urochordata										
Order: Enterogona		<i>Ciona intestinalis</i>	32	LFRWNR			N.D.	N.D.		N.D.	N.D.
	Sea squirt	<i>Ciona intestinalis</i>									

Amino acid sequences around putative palmitoylation sites corresponding to human 5-HT_{1A} receptor Cys417 and Cys420; those corresponding to human 5-HT₄ receptor Cys328, Cys329, and human 5-HT_{4(a)} receptor Cys386; those corresponding to human 5-HT₇ receptor Cys401 and human 5-HT_{7(a)} receptor Ala435 and Cys438 in vertebrate 5-HT receptor orthologs are shown. Percent identities between orthologs across 2 species were obtained by performing BLAST search (with BLOSUM62) with full-length amino acid sequence of human 5-HT receptor orthologs. N. D., sequence not determined.

cysteine residue there (Table 1). In these animals, the corresponding site is exceptionally substituted or lost only in 2 bat species (great roundleaf bat and Chinese rufous horseshoe bat) and ghost shark. Similar to many other teleost fish genes, most teleost fishes have more than 2 different types of 5-HT_{1A} receptor. Redundant 5-HT_{1A} receptor orthologs in teleost fishes are consistent with their additional whole-genome duplication that occurred in the ancestor of teleosts.^{26,27} Basically, there exist 2 cysteine residues at least in one ortholog of most ray-finned fishes. Characteristic sequences “-IIKCHFRCR-stop” or “-IIKCKFHRP-stop” and their several variations are detected in the C-termini of fish 5-HT_{1A} receptor orthologs. Even sea lamprey 5-HT_{1A} receptor holds both putative palmitoylation sites “-IIKCKFCRQ-stop.”

3.2 | 5-HT₄ receptor palmitoylation sites in various vertebrate species

5-HT₄ receptors are coupled to G_s protein that stimulate adenylyl cyclases to produce cAMP. Some splice variants of the 5-HT₄ receptor have been identified for many vertebrate species. For instance, there exist at least 11 5-HT₄ receptor splice variants in humans.²⁸ These variants show different tissue distribution and exhibit different functional activities.^{28,29} Mouse 5-HT_{4(a)} (also called 5-HT_{4 isoform X2}) receptor contains 387 amino acids that is palmitoylated at distinct sites, Cys328/Cys329 and Cys386.^{22,23} While Cys328/Cys329 are commonly conserved among all mouse splice variants, Cys386 specifically locates close to the C-terminus

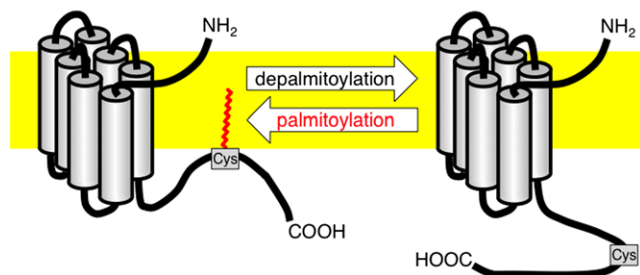
vertebrate 5-HT_{1A}, 4, 7 receptors

FIGURE 1 Palmitoylation of vertebrate 5-HT receptors. Schematic structure of 5-HT_{1A}, 5-HT₄, and 5-HT₇ receptors and their C-terminal palmitoylation sites. 5-HT receptors contain 7 transmembrane domains and intracellular C-terminal domain. Squared “Cys” represents multiple palmitoylation sites of 5-HT_{1A}, 5-HT₄, and 5-HT₇ receptors

of 5-HT_{4(a)}-type variant (Figure 3A). The 5-HT_{4(a)} receptor C-terminus around the palmitoylation site (-Ser-Cys386-Phe-stop) may also form a class I PDZ ligand (-Ser/Thr-Xaa-φ-stop, where φ represents a hydrophobic residue), which controls protein-protein interactions between receptors and PDZ domain-containing binding proteins.³⁰ Previous reports showed that agonist stimulation-dependent palmitoylation on both sites is involved in regulation of receptor constitutive activity.^{22,23} Palmitoylation sites corresponding to mouse 5-HT₄ receptor Cys328/Cys329 are completely conserved in all vertebrate 5-HT₄ receptor orthologs including other splice variants than 5-HT_{4(a)} receptor (227 vertebrate species examined, Table 1). In addition to these common palmitoylation sites among 5-HT₄ receptor splice variants, 5-HT_{4(a)} receptor-specific C-terminal palmitoylation site Cys386 and PDZ ligands are almost completely conserved in known 5-HT_{4(a)}-type splice variants (Table 1). This palmitoylation site is exceptionally absent only in sifaka 5-HT_{4(a)} receptor in total 142 vertebrate species.

3.3 | 5-HT₇ receptor palmitoylation sites in various vertebrate species

Several isoforms exist in mammalian 5-HT₇ receptors.²⁸ Both humans and mouse possess 3 5-HT₇ receptor C-terminal splice variants. Dynamic palmitoylation of mouse 5-HT_{7(a)} receptor regulates cAMP formation by activating adenylyl cyclase via a G_s-mediated signaling pathway.²⁴ Mouse 5-HT_{7(a)} (or 5-HT_{7 isoform 1}) receptor holds 3 palmitoylation sites at Cys404, Cys438, and Cys441 (Figure 3B). Mouse 5-HT₇ receptor Cys404 corresponds to human 5-HT₇ receptor Cys401, which is commonly conserved in all splice variants of vertebrate 5-HT₇ receptor orthologs (240 vertebrate species examined, Table 1). In contrast to this common palmitoylation site, cysteine residue corresponding to mouse 5-HT_{7(a)} receptor Cys438 is specifically found only in 2 species, mouse and saltwater crocodile (Table 1). In other 114 vertebrate species, Ser dominantly exists at the corresponding sites, which is substituted to Ala, Thr, or Pro in some species.

(A)

5-HT _{1A} orthologs	Human	Mouse	Chicken	Anole	Frog	Zebrafish
Human	100%	88%	80%	71%	72%	75%
Mouse		100%	79%	68%	69%	73%
Chicken			100%	69%	72%	77%
Anole				100%	65%	67%
Frog					100%	70%
Zebrafish						100%

(B)

5-HT _{4(a)} orthologs	Human	Mouse	Chicken	Anole	Frog	Zebrafish
Human	100%	94%	89%	88%	85%	76%
Mouse		100%	86%	86%	83%	73%
Chicken			100%	92%	90%	77%
Anole				100%	89%	76%
Frog					100%	78%
Zebrafish						100%

(C)

5-HT _{7(a)} orthologs	Human	Mouse	Chicken	Anole	Frog	Zebrafish (5-HT ₇)
Human	100%	95%	80%	70%	74%	78%
Mouse		100%	79%	70%	74%	78%
Chicken			100%	74%	73%	77%
Anole				100%	77%	70%
Frog					100%	70%
Zebrafish (5-HT ₇)						100%

FIGURE 2 The BLAST alignments of vertebrate 5-HT receptors. A, 5-HT_{1A} receptor orthologs; B, 5-HT_{4(a)} receptor orthologs; C, 5-HT_{7(a)} receptor orthologs. Percent identity among orthologs across any 2 species was obtained by performing BLAST search (with BLOSUM62) with full-length amino acid sequences of vertebrate 5-HT receptor orthologs. *Homo sapiens* (human being), *Mus musculus* (mouse), *Gallus gallus* (chicken), *Anolis carolinensis* (green anole), *Xenopus laevis* (African clawed frog), *Danio rerio* (zebrafish) are compared as representative of each vertebrate class

The class Mammalia recognizes 3 subclasses: the Prototheria (platypus and several species of echidna), the Metatheria (extant Marsupialia, eg, koala), and the Eutheria (extant Placentalia, eg, mouse) (Figure 4A). Eutherian (placental) mammals further comprise 4 superorders; Afrotheria, Xenarthra, Laurasiatheria, and Euarchontoglires. All of the eutherian species diverged from the same root around 100 million years ago in the early Cretaceous period.^{31,32} Mouse 5-HT_{7(a)} receptor Cys441 corresponds to human 5-HT_{7(a)} receptor Cys438, which is extremely conserved in almost all eutherian 5-HT_{7(a)}-type splice variants (83 eutherian species examined, Table 1). The site is exceptionally substituted only in 2 species, minke whale and armadillo. Moreover, many molecular studies based on DNA analysis have supported an integrated classification as the magorder Boreoeutheria.^{31–33} The Boreoeutheria is a clade that is composed of 2 superordinal sister taxa Laurasiatheria (cat, etc.) and Euarchontoglires (humans, mouse, etc.). Our analysis showed that 81

**(A)**
mouse 5-HT₄ receptor splice variants

isoform X1/(b): 388 aa (C328, C329)

LIILCCDDERYKRPPILGOTVPCSTTTINGSTHVLRDTVECGGWESRCHLTATSPLVAAQPSDT

isoform X2/(a): 387 aa (C328, C329, C386)

LIILCCDDERYKRPPILGOTVPCSTTTINGSTHVLRYTVLHSGHHQELEKLP IHNDPESLE**SCF**

isoform X3: 377 aa (C328, C329)

LIILCCDDERYKRPPILGOTVPCSTTTINGSTHVLRTSYDYDTYLQSLGSDKSVI

isoform X4/(e): 371 aa (C328, C329)

LIILCCDDERYKRPPILGOTVPCSTTTINGSTHVLSFPLLFRNRVPV

isoform X5: 368 aa (C328, C329)

LIILCCDDERYKRPPILGOTVPCSTTTINGSTHVLRAGGDQLVTP**(B)**
mouse 5-HT₇ receptor splice variants

isoform 1/(a): 448 aa (C404, C438, C441)

LLQCOYRNINRKLSAAGMHEALKLAERPERSEFVLQNC**CDHCGKKGHDT**

isoform 2: 470 aa (C404)

LLQCOYRNINRKLSAAGMHEALKLAERPERSEFVLMTGASGVQKALENLPWNGVNTGIKAVNSVALTKL

isoform 3: 435 aa (C404)

LLQCOYRNINRKLSAAGMHEALKLAERPERSEFVLhuman 5-HT₇ receptor splice variants

isoform (a): 445 aa (C401, A435, C438)

LLQCOYRNINRKLSAAGMHEALKLAERPERPEFVLQADY**CRKKGHDS**

isoform (b): 432 aa (C401)

LLQCOYRNINRKLSAAGMHEALKLAERPERPEFVL

isoform (d): 479 aa (C401, C435(?))

LLQCOYRNINRKLSAAGMHEALKLAERPERPEFVLR**ACTRRVLLRPEKRPPVSVVWLQSPDHHNLADKM**
LTTVEKKVMIHD**FIGURE 3** Palmitoylation sites in the 5-HT receptor splice variants. A, C-terminal sequence alignment of the mouse 5-HT₄ receptor splice variants; B, C-terminal sequence alignments of the mouse and the human 5-HT₇ receptor splice variants. Consensus amino acid sequence among splice variants is underlined. Palmitoylation sites are marked in red. The box shows the canonical class I PDZ ligand specifically located on the C-terminus of 5-HT_{4(a)} receptor

5-HT_{7(a)} receptor orthologs contain the corresponding cysteine residue in total 82 boreoeutherian species (Figure 4B). In metatherians, opossum holds cysteine residue at the corresponding site, whereas Tasmanian devil and koala lack the palmitoylation site. The serine residue is shared at the corresponding site in Tasmanian devil, koala, platypus, sauropsids (all birds and all reptiles), and all amphibians. 5-HT_{7(a)}-type splice variants have not been detected in fishes.

4 | DISCUSSION

Sequence comparison of 5-HT receptor orthologs made it possible to clarify the process of acquisition, conservation, substitution, or loss of these protein modification sites in vertebrate evolution. As described above, sea lamprey 5-HT_{1A} receptor ortholog has completely same sequence with humans in its C-terminus “-IICKFCRQ-

stop.” There exists no palmitoylation site in nonvertebrate chordates such as lancelet and sea squirt. Thus, acquisition event of these palmitoylation sites in the 5-HT_{1A} receptor may occur in the common vertebrate ancestor around 500 million years ago in the late Cambrian to the early Ordovician periods. These sites are evolutionarily conserved against mutation pressure throughout vertebrate species. They have been partly lost later in limited species. Conserved expression pattern of 5-HT_{1A} receptor orthologs was observed in most of the brain regions between sea lamprey and other vertebrates.³⁴ In conclusion, C-terminal palmitoylation sites of 5-HT_{1A} receptor, which is likely to play crucial roles in the vertebrate central nervous system, are completely conserved in the vertebrate lineage from the superclass Agnatha (jawless fishes) to the superclass Gnathostomata (jawed vertebrate).

Similarly, all 5-HT₄ receptor splice variants have the common palmitoylation sites. Even the lancelet 5-HT₄ receptor ortholog

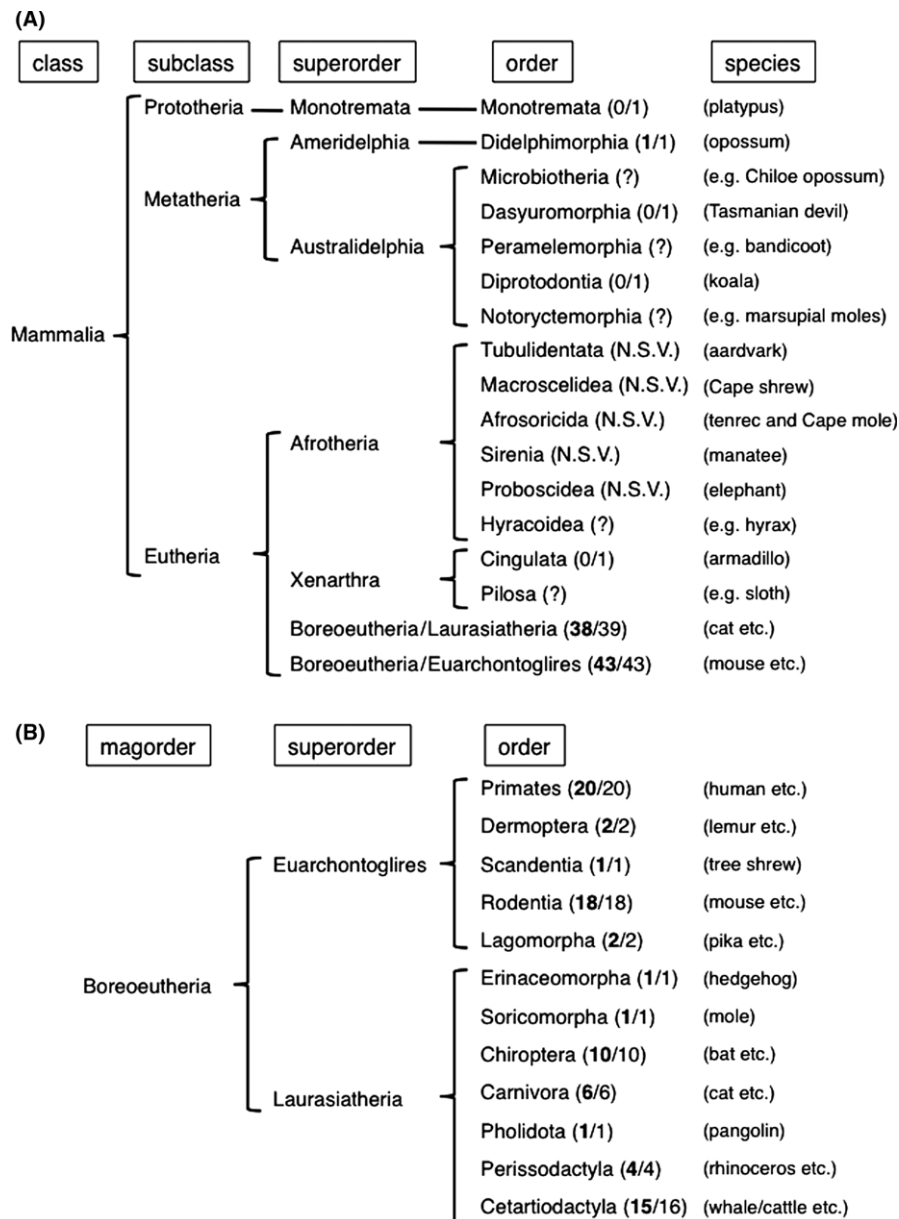


FIGURE 4 Conserved palmitoylation motifs in the mammalian 5-HT_{7(a)} receptors. A, Classification of mammalian species and existence of C-terminal putative palmitoylation site corresponding to mouse 5-HT_{7(a)} receptor Cys441/human 5-HT_{7(a)} receptor Cys438. Numbers of putative palmitoylation site-containing species (shown in bold) per those of examined species in 17 reported mammalian orders are indicated. "?" means that sequence information is currently unavailable for indicated orders. N. S. V.: no information has been reported on 5-HT_{7(a)}-type splice variant sequence yet; B, Classification of boreoeutherian species and existence of C-terminal putative palmitoylation site corresponding to mouse 5-HT_{7(a)} receptor Cys441/human 5-HT_{7(a)} receptor Cys438. Numbers of putative palmitoylation site-containing species (shown in bold) per those of examined species in 12 mammalian orders are indicated

possesses a primitive palmitoylation sequence "-RILSCWSCCDVDN-." Repetitive deletions and substitutions have presumably occurred to generate the common palmitoylation motif for vertebrate 5-HT₄ receptors "typically, -IILCCGDE-," followed by Gly to Asp mutation in mammals "typically, -IILCCDDE-." (Figure 5A). 5-HT_{4(a)} receptor-specific C-terminal palmitoylation site corresponding to mouse/human Cys386 had appeared in the ancestor of bony fishes. 5-HT_{4(a)}-type splice variants have not been reported for cartilaginous fishes, and no information about

any 5-HT₄ receptor is currently available for cyclostomes (hagfishes and lampreys). Future analysis about 5-HT₄ receptor orthologs of hagfishes, lampreys, more sharks, and rays will reveal the detailed history of acquisition and establishment of palmitoylation sites among 5-HT₄ receptor variants. Reversible palmitoylation and depalmitoylation cycle of this 5-HT_{4(a)} receptor-specific site may be critical for dynamic regulation of 5-HT_{4(a)} receptor localization and membrane trafficking through its binding to PDZ domain-containing scaffold proteins.

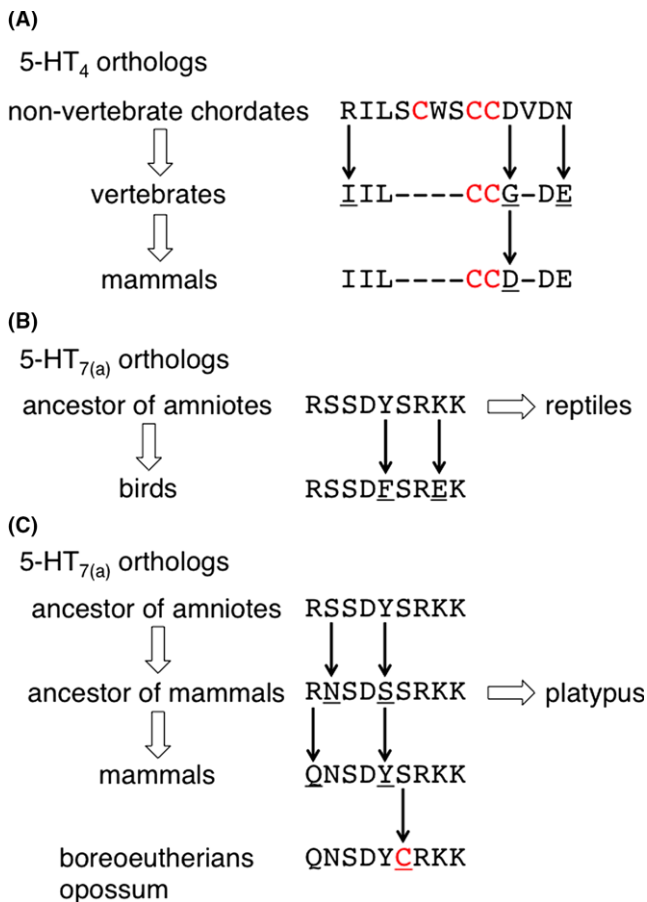


FIGURE 5 Evolutionary models of stepwise mutations around palmitoylation sites in vertebrate 5-HT receptor orthologs. A, A model of mutations in 5-HT₄ receptor orthologs developed from a putative ancestor of chordates to mammals. Sequence of the ancestral chordate is predicted from nonvertebrate chordate, lancelet; B, A model of mutations in 5-HT_{7(a)} receptor orthologs developed from a putative ancestor of amniotes to birds. Sequence of the ancestral amniote is predicted from reptiles; C, A model of mutations in 5-HT_{7(a)} receptor orthologs developed from a putative ancestor of amniotes to mammals. Sequence of the ancestral mammal is predicted from platypus. (-): amino acid deletions; arrows: amino acid changes. Putative palmitoylation sites are marked in red

Likewise, all tetrapod 5-HT₇ receptor orthologs have the common palmitoylation site corresponding to mouse Cys404/human Cys401. The origin and development of the palmitoylation motifs in 5-HT₇ receptors still remains unclear because there is no available information on lancelet, sea squirt, and cyclostomes. By contrast, 5-HT_{7(a)}-type variant appeared in the ancestor of tetrapods, in which serine residue locates at the corresponding site to mouse Cys441/human Cys438. Both reptiles and birds hold serine residue at the corresponding site (Figure 5B). Cysteine residue at the palmitoylation site on 5-HT_{7(a)} receptor C-terminus is almost completely conserved in all orders belonging to boreoeutherians (Figure 5C). Highly conserved 5-HT_{7(a)} receptor C-terminal motif around the palmitoylation site in boreoeutherian lineages strongly suggests that this C-terminal motif was shared at least in the common ancestor of

boreoeutherians (Figure 4B). This feature was acquired at a certain point during the mammalian evolution. It is still difficult to speculate when acquisition events of this 5-HT_{7(a)} receptor-specific modification site initially took place, because 5-HT_{7(a)}-type variants have not been identified for afrotherian mammals (Figure 4A). On the other hand, Gray short-tailed opossum possesses cysteine residue at the corresponding site in its 5-HT_{7(a)} receptor. This sequence enables us to predict that a replacement of Ser to Cys specifically happened in this marsupial species and in the common ancestor of boreoeutherians. In contrast, Tasmanian devil and koala hold serine residue there. The difference may reflect the marsupial divergence history in Australasia and the Americas. Another possibility is that 5-HT_{7(a)} receptor-specific palmitoylation site was initially established in the common ancestor between the metatherian and the eutherian lineages and has been lost later in afrotherians, xenarthrans, and most metatherian species.

Further accumulation of sequence data will fill in the blanks of the sequence list concerning vertebrate 5-HT_{4(a)}-type variants and tetrapod 5-HT_{7(a)}-type variants and will reveal the timeline of establishment and divergence of these palmitoylation sites-containing motifs in more detail. Especially, sequence information on urochordates (ascidians or sea squirts), cephalochordates (lancelets), and cyclostomes will clarify the initial acquisition of the mechanism of 5-HT receptor palmitoylation.

Dynamic regulation of 5-HT receptors made possible by reversible post-translational protein palmitoylation may be critical for more effective membrane localization and trafficking in refined functions of the vertebrate tissues. Previous researches have revealed that many other GPCRs are functionally regulated by their direct palmitoylation.^{35,36} So far, we have shown that palmitoylation sites of ionotropic glutamate receptors (iGluRs), the major excitatory neurotransmitter receptors in vertebrate central nervous system, and those of iGluRs-binding proteins are extremely conserved in various species of whole vertebrate.³⁷⁻³⁹ Furthermore, palmitoylation sites of hyperpolarization-activated cyclic nucleotide-gated (HCN)-2 channel⁴⁰ and water channel aquaporin (AQP)-4⁴¹ are conserved across vertebrates. By contrast, palmitoylation sites of dopamine D₁-like, D₁ and D₅, receptors, are broadly found in vertebrates and invertebrates.⁴² Future genome analysis would permit us to understand detailed history of acquisition and refinement of the post-translational protein palmitoylation in vertebrates.

ACKNOWLEDGMENTS

This work was supported in part by the Grants-in-Aid from the Ministry of Education, Culture, Sports, Science and Technology of Japan (MEXT) Grant Number 16K07078, AMED-RRIME from the Japan Agency for Medical Research and Development (AMED) Grant Number JP17gm5910009, the Takeda Science Foundation, the Mitsubishi Foundation, the Brain Science Foundation, the Suzuken Memorial Foundation, and the Astellas Foundation for Research on Metabolic Disorders.



CONFLICT OF INTEREST

The authors declare no conflict of interest for this article.

DATA REPOSITORY

The authors do not deposit any data because this manuscript does not use data.

APPROVAL OF THE RESEARCH PROTOCOL BY AN INSTITUTIONAL REVIEWER BOARD

n/a.

INFORMED CONSENT

n/a.

REGISTRY AND THE REGISTRATION NO. OF THE STUDY/TRIAL

n/a.

ANIMAL STUDIES

n/a.

AUTHOR CONTRIBUTION

TK executed data collection and analyzed the results. TH conceived this project, analyzed the results, and wrote the manuscript.

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REFERENCES

- Mohammad-Zadeh LF, Moses L, Gwaltney-Brant SM. Serotonin: a review. *J Vet Pharmacol Ther.* 2008;31:187–99.
- Andrews PW, Bharwani A, Lee KR, Fox M, Thomson JA Jr. Is serotonin an upper or a downer? The evolution of the serotonergic system and its role in depression and the antidepressant response. *Neurosci Biobehav Rev.* 2015;51:164–88.
- Kaufman J, DeLorenzo C, Choudhury S, Parsey RV. The 5-HT_{1A} receptor in major depressive disorder. *Eur Neuropsychopharmacol.* 2016;26:397–410.
- Vadodaria KC, Stern S, Marchetto MC, Gage FH. Serotonin in psychiatry: in vitro disease modeling using patient-derived neurons. *Cell Tissue Res.* 2017;371:161–70.
- Yohn CN, Gergues MM, Samuels BA. The role of 5-HT receptors in depression. *Mol Brain.* 2017;10:28.
- Gillette R. Evolution and function in serotonergic systems. *Integr Comp Biol.* 2006;46:838–46.
- Barnes NM, Sharp T. A review of central 5-HT receptors and their function. *Neuropharmacology.* 1999;38:1083–152.
- Koyama Y, Kondo M, Shimada S. The significance of complete distributional analysis of the serotonin receptor. *Neurotransmitter.* 2017;4:e1564.
- Gorinski N, Ponimaskin E. Palmitoylation of serotonin receptors. *Biochem Soc Trans.* 2013;41:89–94.
- Ng GY, George SR, Zastawny RL, et al. Human serotonin_{1B} receptor expression in Sf9 cells: phosphorylation, palmitoylation, and adenylyl cyclase inhibition. *Biochemistry.* 1993;32:11727–33.
- Fukata Y, Fukata M. Protein palmitoylation in neuronal development and synaptic plasticity. *Nat Rev Neurosci.* 2010;11:161–75.
- Linder ME, Deschenes RJ. Palmitoylation: policing protein stability and traffic. *Nat Rev Mol Cell Biol.* 2007;8:74–84.
- Nadolski MJ, Linder ME. Protein lipidation. *FEBS J.* 2007;274:5202–10.
- Resh MD. Fatty acylation of proteins: new insights into membrane targeting of myristoylated and palmitoylated proteins. *Biochim Biophys Acta.* 1999;1451:1–16.
- Fallin MD, Lasseter VK, Wolyniec PS, et al. Genomewide linkage scan for bipolar-disorder susceptibility loci among Ashkenazi Jewish families. *Am J Hum Genet.* 2004;75:204–19.
- Mansouri MR, Marklund L, Gustavsson P, et al. Loss of ZDHC15 expression in a woman with a balanced translocation t(X;15)(q13.3;cen) and severe mental retardation. *Eur J Hum Genet.* 2005;13:970–7.
- Mukai J, Dhilla A, Drew LJ, et al. Palmitoylation-dependent neurodevelopmental deficits in a mouse model of 22q11 microdeletion. *Nat Neurosci.* 2008;11:1302–10.
- Otani K, Ujike H, Tanaka Y, et al. The ZDHC8 gene did not associate with bipolar disorder or schizophrenia. *Neurosci Lett.* 2005;390:166–70.
- Raymond FL, Tarpey PS, Edkins S, et al. Mutations in ZDHC9, which encodes a palmitoyltransferase of NRAS and HRAS, cause X-linked mental retardation associated with a Marfanoid habitus. *Am J Hum Genet.* 2007;80:982–7.
- Young FB, Butland SL, Sanders SS, Sutton LM, Hayden MR. Putting proteins in their place: palmitoylation in Huntington disease and other neuropsychiatric diseases. *Prog Neurobiol.* 2012;97:220–38.
- Papoucheva E, Dumuis A, Sebben M, Richter DW, Ponimaskin EG. The 5-hydroxytryptamine(1A) receptor is stably palmitoylated, and acylation is critical for communication of receptor with Gi protein. *J Biol Chem.* 2004;279:3280–91.
- Ponimaskin EG, Heine M, Joubert L, et al. The 5-hydroxytryptamine (4a) receptor is palmitoylated at two different sites, and acylation is critically involved in regulation of receptor constitutive activity. *J Biol Chem.* 2002;277:2534–46.
- Ponimaskin EG, Schmidt MF, Heine M, Bickmeyer U, Richter DW. 5-Hydroxytryptamine 4(a) receptor expressed in Sf9 cells is palmitoylated in an agonist-dependent manner. *Biochem J.* 2001;353(Pt 3):627–34.
- Kvachnina E, Dumuis A, Wlodarczyk J, et al. Constitutive Gs-mediated, but not G12-mediated, activity of the 5-hydroxytryptamine 5-HT₇(a) receptor is modulated by the palmitoylation of its C-terminal domain. *Biochim Biophys Acta.* 2009;1793:1646–55.
- El-Husseini Ael D, Brecht DS. Protein palmitoylation: a regulator of neuronal development and function. *Nat Rev Neurosci.* 2002;3:791–802.
- Glasauer SM, Neuhauss SC. Whole-genome duplication in teleost fishes and its evolutionary consequences. *Mol Genet Genomics.* 2014;289:1045–60.
- Hermansen RA, Hvidsten TR, Sandve SR, Liberles DA. Extracting functional trends from whole genome duplication events using comparative genomics. *Biol Proced Online.* 2016;18:11.
- Coupar IM, Desmond PV, Irving HR. Human 5-HT(4) and 5-HT(7) receptor splice variants: are they important? *Curr Neuropharmacol.* 2007;5:224–31.



29. Claeysen S, Sebben M, Becamel C, Bockaert J, Dumuis A. Novel brain-specific 5-HT₄ receptor splice variants show marked constitutive activity: role of the C-terminal intracellular domain. *Mol Pharmacol*. 1999;55:910–20.
30. Joubert L, Hanson B, Barthet G, et al. New sorting nexin (SNX27) and NHERF specifically interact with the 5-HT₄ receptor splice variant: roles in receptor targeting. *J Cell Sci*. 2004;117(Pt 22):5367–79.
31. Springer MS, Murphy WJ. Mammalian evolution and biomedicine: new views from phylogeny. *Biol Rev Camb Philos Soc*. 2007;82:375–92.
32. Springer MS, Meredith RW, Janecka JE, Murphy WJ. The historical biogeography of Mammalia. *Philos Trans R Soc Lond B Biol Sci*. 2011;366:2478–502.
33. Ruiz-Herrera A, Farre M, Robinson TJ. Molecular cytogenetic and genomic insights into chromosomal evolution. *Heredity*. 2012;108:28–36.
34. Cornide-Petronio ME, Anadon R, Barreiro-Iglesias A, Rodicio MC. Serotonin 1A receptor (5-HT_{1A}) of the sea lamprey: cDNA cloning and expression in the central nervous system. *Brain Struct Funct*. 2013;218:1317–35.
35. Escriba PV, Wedegaertner PB, Goni FM, Vogler O. Lipid-protein interactions in GPCR-associated signaling. *Biochim Biophys Acta*. 2007;1768:836–52.
36. Norskov-Lauritsen L, Brauner-Osborne H. Role of post-translational modifications on structure, function and pharmacology of class C G protein-coupled receptors. *Eur J Pharmacol*. 2015;763(Pt B):233–40.
37. Thomas GM, Hayashi T. Smarter neuronal signaling complexes from existing components: how regulatory modifications were acquired during animal evolution: evolution of palmitoylation-dependent regulation of AMPA-type ionotropic glutamate receptors. *BioEssays*. 2013;35:929–39.
38. Hayashi T. Evolutionarily conserved palmitoylation-dependent regulation of ionotropic glutamate receptors in vertebrates. *Neurotransmitter*. 2014;1:e388.
39. Hayashi T. The origin and diversity of PICK1 palmitoylation in the Eutheria. *Neurotransmitter*. 2015;2:e802.
40. Itoh M, Kaizuka T, Hayashi T. Evolutionary acquisition and divergence of vertebrate HCN2 palmitoylation. *Neurotransmitter*. 2017;4:e1603.
41. Hayashi T. Conservation and phylogenetic stepwise changes of aquaporin (AQP) 4 palmitoylation in vertebrate evolution. *Neurotransmitter*. 2017;4:e1608.
42. Adachi T, Hayashi T. Evolutionarily conserved phosphorylation and palmitoylation-dependent regulation of dopamine D1-like receptors in vertebrates. *Neurotransmitter*. 2016;3:e1434.

How to cite this article: Kaizuka T, Hayashi T. Comparative analysis of palmitoylation sites of serotonin (5-HT) receptors in vertebrates. *Neuropsychopharmacol Rep*. 2018;38:75–85. <https://doi.org/10.1002/npr2.12011>