

ILLUSTRATED REVIEW

Illustrated State-of-the-Art Capsules of the ISTH 2022 Congress

Robert A. Ariëns PhD¹ | Beverley J. Hunt MD OBE² |
 Ejaife O. Agbani BPharm, MSc, PhD³ | Josefin Ahnström PhD⁴ |
 Robert Ahrends PhD⁵ | Raza Alikhan MD, FRCP, FRCPath⁶ | Alice Assinger PhD⁷ |
 Zsuzsa Bagoly MD, PhD⁸ | Alessandra Balduini MD⁹ |
 Elena Barbon PhD¹⁰ | Christopher D. Barrett MD^{11,12,13} | Paul Batty MBBS, PhD¹⁴ |
 Jorge David Aivazoglou Carneiro MD, PhD¹⁵ | Wee Shian Chan MD, MSc, FRCP¹⁶ |
 Moniek de Maat PhD¹⁷ | Kerstin de Wit MBChB, MD, MSc¹⁸ | Cécile Denis PhD¹⁹ |
 Martin H. Ellis MD²⁰ | Renee Eslick BMed, FRACP, FRCPA²¹ | Hongxia Fu PhD²² |
 Catherine P. M. Hayward MD, PhD²³ | Benoit Ho-Tin-Noé PhD²⁴ |
 Frederikus A. Klok MD, PhD²⁵ | Riten Kumar MD, MSc²⁶ | Karin Leiderman PhD²⁷ |
 Rustem I. Litvinov MD, PhD²⁸ | Nigel Mackman PhD²⁹ | Zoe McQuilten MBBS, PhD³⁰ |
 Matthew D. Neal MD³¹ | William A. E. Parker MB, PhD, MRCP³² |
 Roger J. S. Preston PhD³³ | Julie Rayes PhD³⁴ | Alireza R. Rezaie PhD³⁵ |
 Lara N. Roberts MBBS, MD Res³⁶ | Bianca Rocca MD, PhD³⁷ |
 Susan Shapiro MD, PhD^{38,39} | Deborah M. Siegal MD, MSc, FRCPC⁴⁰ |
 Lirlândia P. Sousa PhD⁴¹ | Katsue Suzuki-Inoue MD, PhD⁴² |
 Tahira Zafar MB, DCP, FRCPath^{43,44} | Jiaxi Zhou PhD⁴⁵

¹University of Leeds, Leeds, UK²King's Healthcare Partners, London, UK³Department of Physiology and Pharmacology, Cumming School of Medicine, University of Calgary, Calgary, Alberta, Canada⁴Centre for Haematology, Imperial College London, London, UK⁵Institute of Analytical Chemistry, University of Vienna, Vienna, Austria⁶Haemostasis & Thrombosis, University Hospital of Wales, Cardiff, UK⁷Medical University of Vienna, Vienna, Austria⁸Faculty of Medicine, Department of Laboratory Medicine, Division of Clinical Laboratory Sciences and ELKH-DE Neurodegenerative and Cerebrovascular Research Group, University of Debrecen, Debrecen, Hungary⁹University of Pavia, Pavia, Italy¹⁰San Raffaele Telethon Institute for Gene Therapy, IRCCS San Raffaele Scientific Institute, Milan, Italy¹¹Division of Acute Care Surgery and Surgical Critical Care, Department of Surgery, University of Nebraska Medical Center, Omaha, Nebraska, USA¹²Koch Institute, Center for Precision Cancer Medicine, Massachusetts Institute of Technology, Cambridge, Massachusetts, USA¹³Division of Surgical Critical Care, Department of Surgery, Boston University Medical Center, Boston University School of Medicine, Boston, Massachusetts, USA¹⁴Royal Free Hospital, London, UK

This is an open access article under the terms of the [Creative Commons Attribution-NonCommercial-NoDerivs](https://creativecommons.org/licenses/by-nc-nd/4.0/) License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

© 2022 The Authors. *Research and Practice in Thrombosis and Haemostasis* published by Wiley Periodicals LLC on behalf of International Society on Thrombosis and Haemostasis (ISTH).

- ¹⁵Instituto da Criança do Hospital das Clínicas da Faculdade de Medicina da Universidade de São Paulo, São Paulo, Brazil
- ¹⁶University of British Columbia, Vancouver, British Columbia, Canada
- ¹⁷Department of Hematology, Erasmus MC, Rotterdam, The Netherlands
- ¹⁸Queen's University and McMaster University, Kingston, ON, Canada
- ¹⁹INSERM UMRS 1176, Le Kremlin-Bicetre Cedex, France
- ²⁰Hematology Institute and Blood Bank, Meir Medical Center and Sackler School of Medicine, Tel Aviv University, Tel Aviv, Israel
- ²¹Haematology Department, Canberra Hospital, Garran, Australian Capital Territory, Australia
- ²²Division of Hematology, Department of Medicine, University of Washington, Seattle, Washington, USA
- ²³Hamilton Health Sciences, McMaster University Medical Center, Hamilton, Ontario, Canada
- ²⁴INSERM Unit 1148, Paris, France
- ²⁵Department of Medicine – Thrombosis and Hemostasis, Leiden University Medical Center, Leiden, The Netherlands
- ²⁶Dana Farber/Boston Children's Cancer and Blood Disorders Center, Boston, Massachusetts, USA
- ²⁷Colorado School of Mines, Golden, Colorado, USA
- ²⁸Department of Cell and Developmental Biology, University of Pennsylvania School of Medicine, Philadelphia, Pennsylvania, USA
- ²⁹UNC Blood Research Center, Division of Hematology, Department of Medicine, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina, USA
- ³⁰Monash University, Melbourne, Victoria, Australia
- ³¹Trauma and Transfusion Medicine Research Center, Department of Surgery, University of Pittsburgh, Pittsburgh, Pennsylvania, USA
- ³²Cardiovascular Research Unit, Northern General Hospital, University of Sheffield, Sheffield, UK
- ³³Irish Centre for Vascular Biology, Department of Pharmacy & Biomolecular Sciences, Royal College of Surgeons in Ireland, Dublin 2, Ireland
- ³⁴University of Birmingham, Birmingham, UK
- ³⁵Cardiovascular Biology Research Program, Oklahoma Medical Research Foundation, Oklahoma City, Oklahoma, USA
- ³⁶King's Thrombosis Centre, Department of Haematological Medicine, King's College Hospital NHS Foundation Trust, London, UK
- ³⁷Department of Safety and Bioethics, Section of Pharmacology, Catholic University School of Medicine, Rome, Italy
- ³⁸Oxford University Hospitals NHS Foundation Trust, Oxford, UK
- ³⁹Radcliffe Department of Medicine, Oxford University, Oxford, UK
- ⁴⁰Ottawa Hospital Research Institute and University of Ottawa, Ottawa, Ontario, Canada
- ⁴¹Departamento de Análises Clínicas e Toxicológicas, Faculdade de Farmácia, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil
- ⁴²Department of Clinical and Laboratory Medicine, University of Yamanashi, Yamanashi, Japan
- ⁴³Frontier Medical College, Abbotabad, Pakistan
- ⁴⁴Hemophilia Treatment Center, Rawalpindi, Pakistan
- ⁴⁵Institute of Hematology & Blood Diseases Hospital, Chinese Academy of Medical Sciences & Peking Union Medical College, Tianjin, China

Handling Editor: Dr Michelle Sholzberg

Abstract

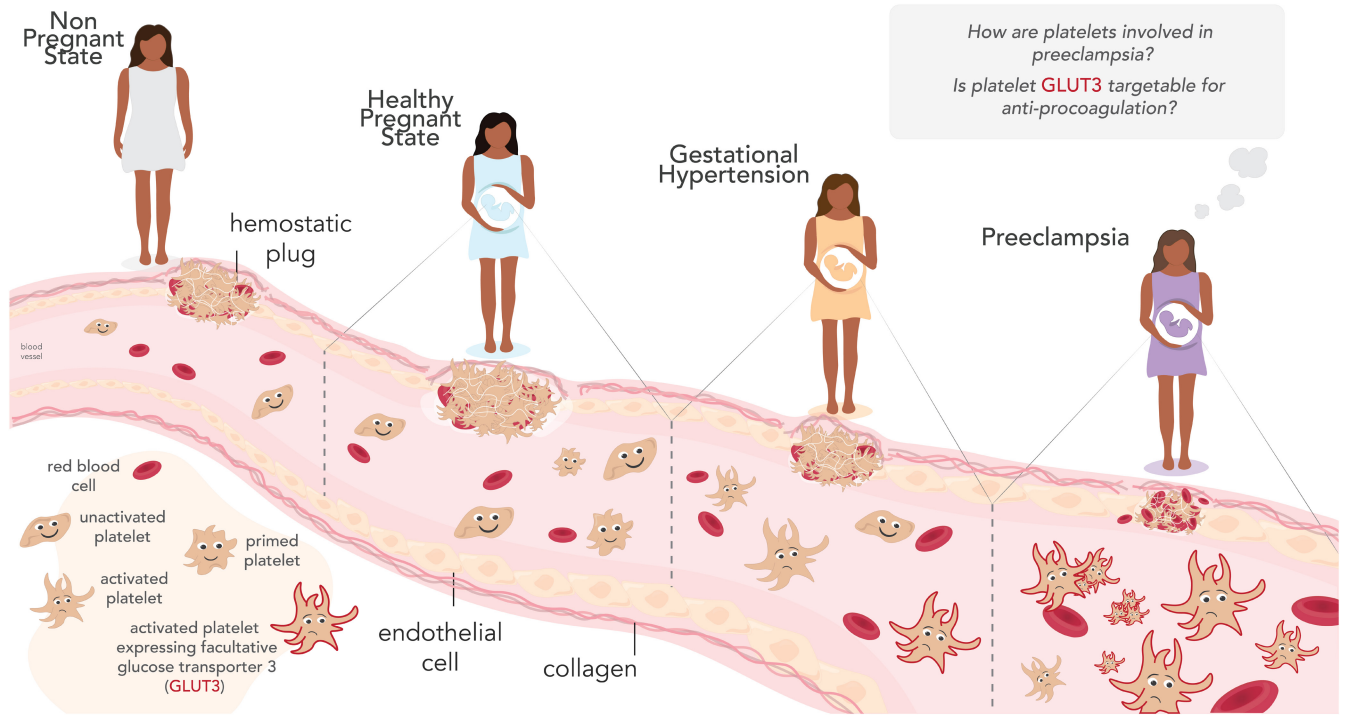
The ISTH London 2022 Congress is the first held (mostly) face-to-face again since the COVID-19 pandemic took the world by surprise in 2020. For 2 years we met virtually, but this year's in-person format will allow the ever-so-important and quintessential creativity and networking to flow again. What a pleasure and joy to be able to see everyone! Importantly, all conference proceedings are also streamed (and available recorded) online for those unable to travel on this occasion. This ensures no one misses out. The 2022 scientific program highlights new developments in hemophilia and its treatment, acquired and other inherited bleeding disorders, thromboinflammation, platelets and coagulation, clot structure and composition, fibrinolysis, vascular biology, venous thromboembolism, women's health, arterial thrombosis, pediatrics, COVID-related thrombosis, vaccine-induced thrombocytopenia with thrombosis, and omics and diagnostics. These areas are elegantly reviewed in this Illustrated Review article. The Illustrated Review is a highlight of the ISTH Congress. The format lends itself very well to explaining the science, and the collection of beautiful graphical summaries of recent developments in the field are stunning and self-explanatory. This clever and effective way to communicate research is revolutionary and different from traditional formats. We hope you enjoy this article and will be inspired by its content to generate new research ideas.

CONTENTS

| | |
|------------------------------------|--|
| Ejaife O. Agbani | Preeclampsia and platelet procoagulant membrane dynamics |
| Josefin Ahnstrom | Protein S, tissue factor pathway inhibitor, and factor V |
| Robert Ahrends | Probing the membrane landscape and identifying key lipids critical for platelet activation by lipidomics |
| Raza Alikhan | Cancer-associated thrombosis: What the guidelines say and what they do not say |
| Alice Assinger | Platelet-leukocyte interaction: Detection and functional relevance in infection and sterile inflammation |
| Zsuzsa Bagoly | Monitoring efficacy of fibrinolysis in stroke |
| Alessandra Balduini | Megakaryocytes and different thrombopoietic environments |
| Elena Barbon | Engineering of adeno-associated viruses to enhance cell-specific transduction |
| Chris Barrett | Fibrinolysis and thrombolytic therapy in COVID-19 respiratory failure |
| Paul Batty | Adeno-associated virus integration |
| Jorge David Aivazoglou Carneiro | Update on the pathogenesis of COVID-19 and multisystem inflammatory syndrome in children |
| Wee Shian Chan | Can pregnancy-adapted algorithms rule out venous thromboembolism in pregnancy? |
| Moniek P. M. de Maat | Hemostatic biomarkers for stroke prediction |
| Kerstin de Wit | Risk stratification for acute venous thromboembolism |
| Cecile Denis | New animal models for bleeding disorders and treatments |
| Martin H. Ellis | Myeloproliferative neoplasms in pregnancy: Implications for mother and child |
| Renee Eslick | Management of thrombocytopenia in pregnancy |
| HongXia Fu | Binding of flow-activated von Willebrand factor to glycoprotein Ib |
| Caterine P. M. Hayward | Hemostatic defects in fibrinolytic disorders |
| Benoit Ho-Tin-Noe | Thrombus composition and thrombolysis resistance in stroke |
| Erik Klok | Long-term outcomes of venous thromboembolism |
| Riten Kumar | Multisystem inflammatory syndrome in children and thrombotic complications of COVID-19 in children |
| Karin Leiderman | Mathematical modeling to study variability in bleeding |
| Rustem I. Litvinov | Blood clot contraction: Mechanisms, pathophysiology, and disease |
| Nigel Mackman | Mechanisms of cancer-associated thrombosis |
| Zoe McQuilten | When to use tranexamic acid in bleeding |
| Matthew D. Neal | Platelet and endothelial biology in bleeding |
| William A. E. Parker | Drug therapies for stroke prevention |
| Roger Preston | Coagulation factor signaling |
| Julie Rayes | Novel mechanism for thromboinflammation in COVID-19 |
| Alireza R. Rezaie | Thrombomodulin regulates endothelial quiescence |
| Lara Roberts | Systematic approach to venous thromboembolism prevention |
| Bianca Rocca | Personalizing antiplatelet therapy based on platelet turnover and metabolic phenotype |
| Susie Shapiro | Venous thromboembolism prevention—What have we learned from COVID-19? |
| Deborah Michelle Siegal | Restarting anticoagulants after major bleeding |
| Lirlandia Piers Sousa | The plasminogen/plasmin system beyond fibrinolysis: Emerging players in resolution of inflammation |
| Katsue Suzuki-Inoue | Crosstalk between hemostasis and lymphangiogenesis |
| Tahira Zafar | Raising awareness of women's bleeding/health issues in culturally challenging settings |
| Jiaxi Zhou | Megakaryocyte single-cell transcriptomics |

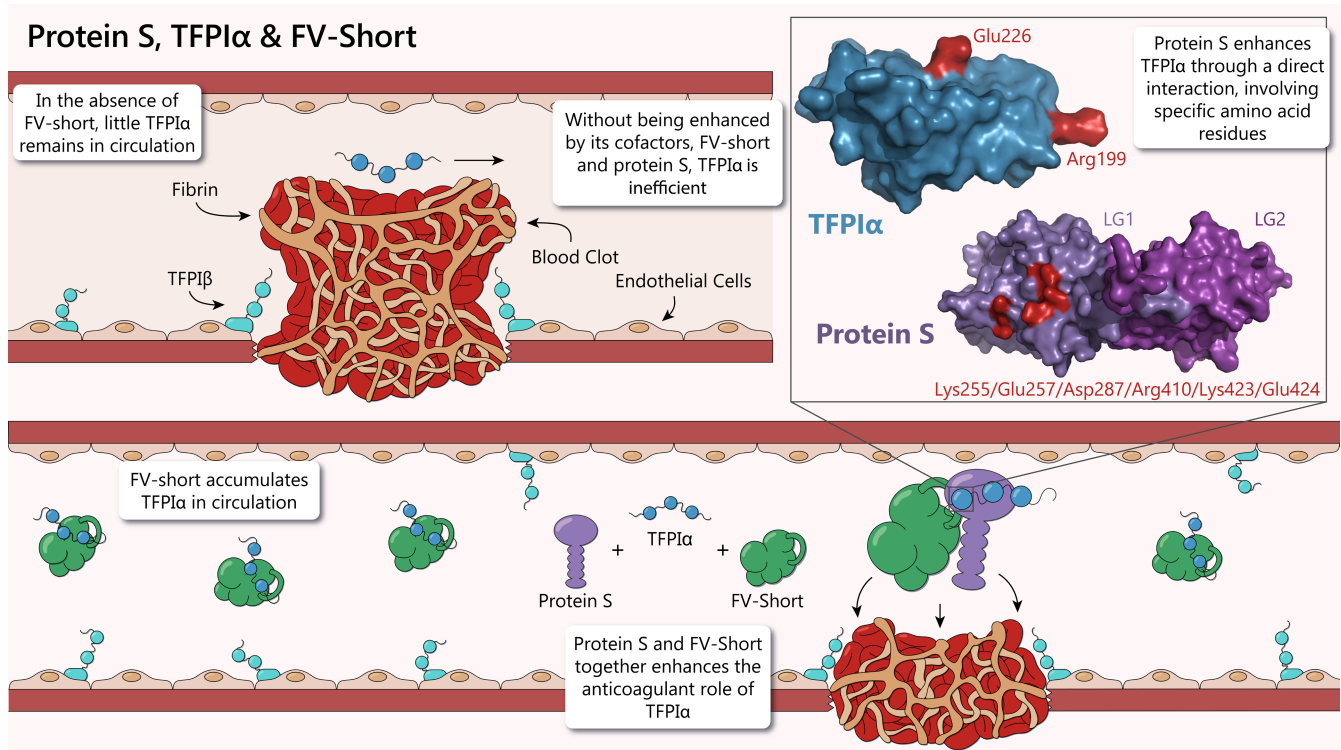
Preeclampsia and platelet procoagulant membrane dynamics

Ejaife O. Agbani BPharm, MSc, PhD



Protein S, tissue factor pathway inhibitor, & factor V

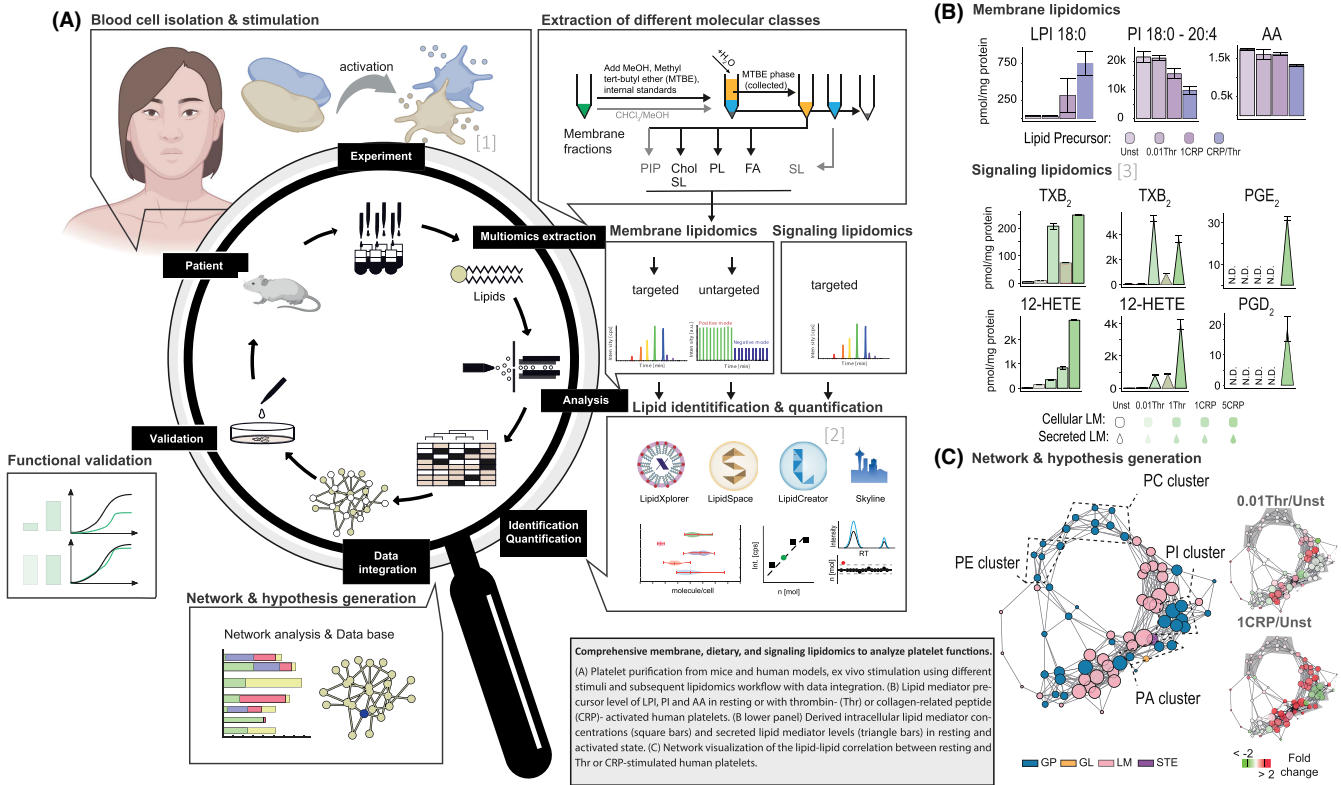
Josefin Ahnström PhD



The initiation phase of coagulation is regulated by tissue factor pathway inhibitor (TFPI), which efficiently reduces/delays thrombin generation. In contrast to the endothelium-bound TFPI β , which in itself is an effective regulator of coagulation, the soluble form, TFPI α , is completely dependent on cofactors (compare the top and bottom figures for the absence and presence of cofactors, respectively).¹ Protein S enhances the anticoagulant properties of TFPI α through a direct protein-protein interaction, involving specific amino acid residues in TFPI α Kunitz 3 and protein S laminin G-type 1.^{1,2} This interaction enables TFPI α to interact with and inhibit membrane-bound factor Xa more efficiently. A splice variant of factor V (FV), FV-short, regulates TFPI α levels through a high-affinity interaction, likely resulting in an increased half-life, as well as functioning as a synergistic TFPI α cofactor, together with protein S.^{1,3}

Probing the membrane landscape and identifying key lipids critical for platelet activation by lipidomics

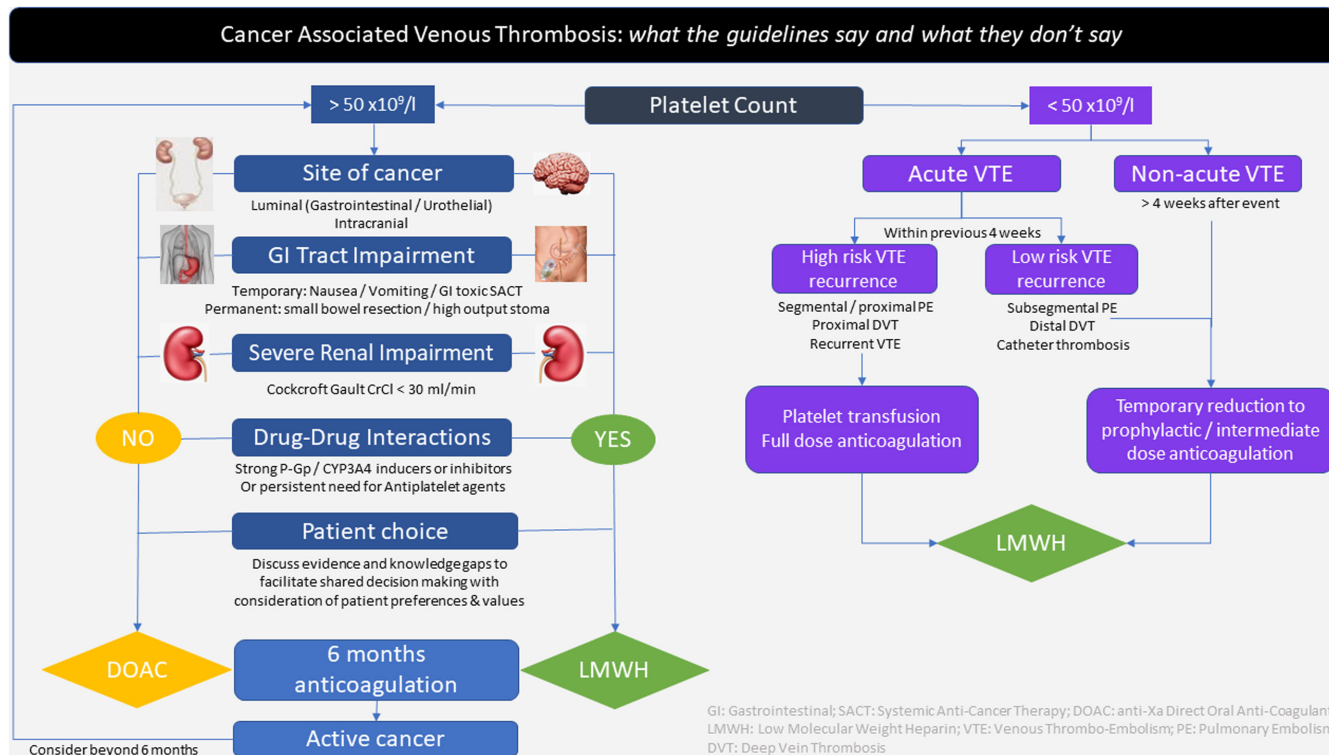
Robert Ahrends PhD



For references, see Peng et al.^{4,5}; Holinstat⁶

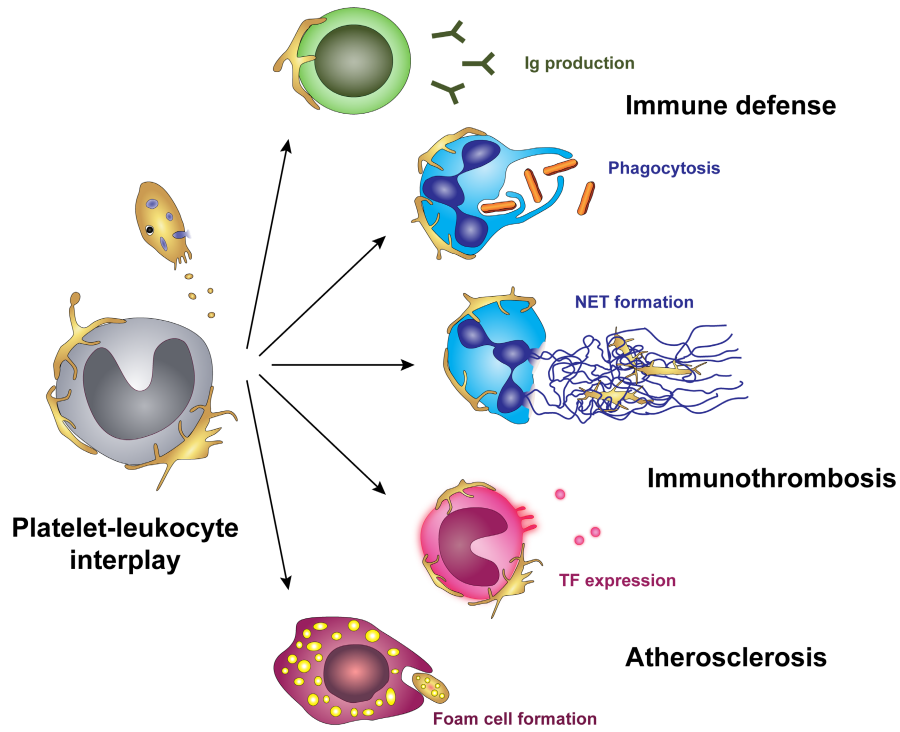
Cancer associated thrombosis: What the guidelines say and what they do not say

Raza Alikhan MD, FRCP, FRCPath



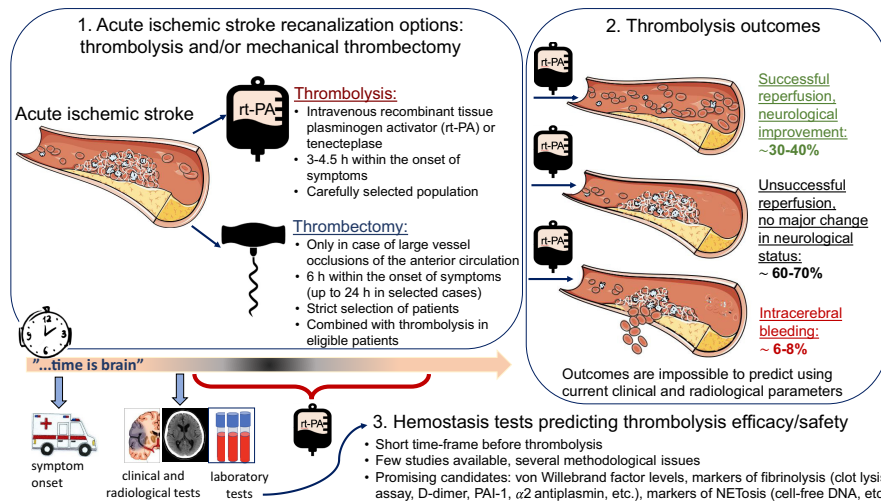
Platelet-leukocyte interaction: Detection and functional relevance in infection and sterile inflammation

Alice Assinger PhD



Monitoring efficacy of fibrinolysis in stroke

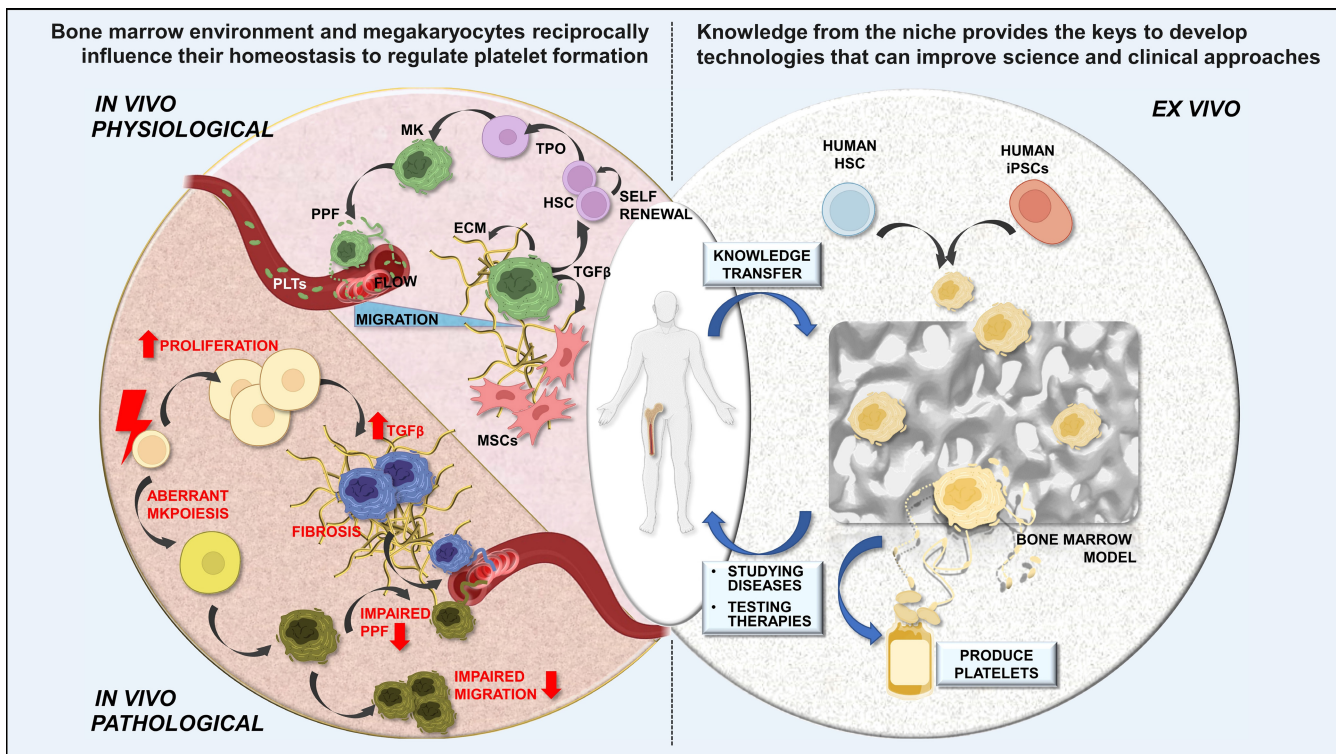
Zsuzsa Bagoly MD, PhD



Currently, there are two proven reperfusion strategies for the opening of the occluded vessel in patients with acute ischemic stroke (AIS): intravenous thrombolysis using recombinant tissue-type plasminogen activator (rt-PA; alteplase) or tenecteplase, and mechanical thrombectomy.⁷ Both therapies must be delivered within a rapid time frame in selected patients; moreover, mechanical thrombectomy is eligible in only a fraction of patients with large-artery occlusion. Despite the unquestionable effectiveness of rt-PA as first-line treatment of AIS, successful reperfusion is achieved in only \approx 30% to 40% of patients, while \approx 6% to 8% of patients develop intracranial hemorrhage as a side effect. As of today, outcomes cannot be foreseen at the initiation of therapy, and this remains one of the greatest challenges of AIS treatment.⁸ Due to the short time frame before treatment, few studies are available on hemostasis tests, and several methodological issues are raised.^{8,9} Nevertheless, some assays show promising results and need to be further investigated and validated in large populations.

Megakaryocytes and different thrombopoietic environments

Alessandra Balduini MD

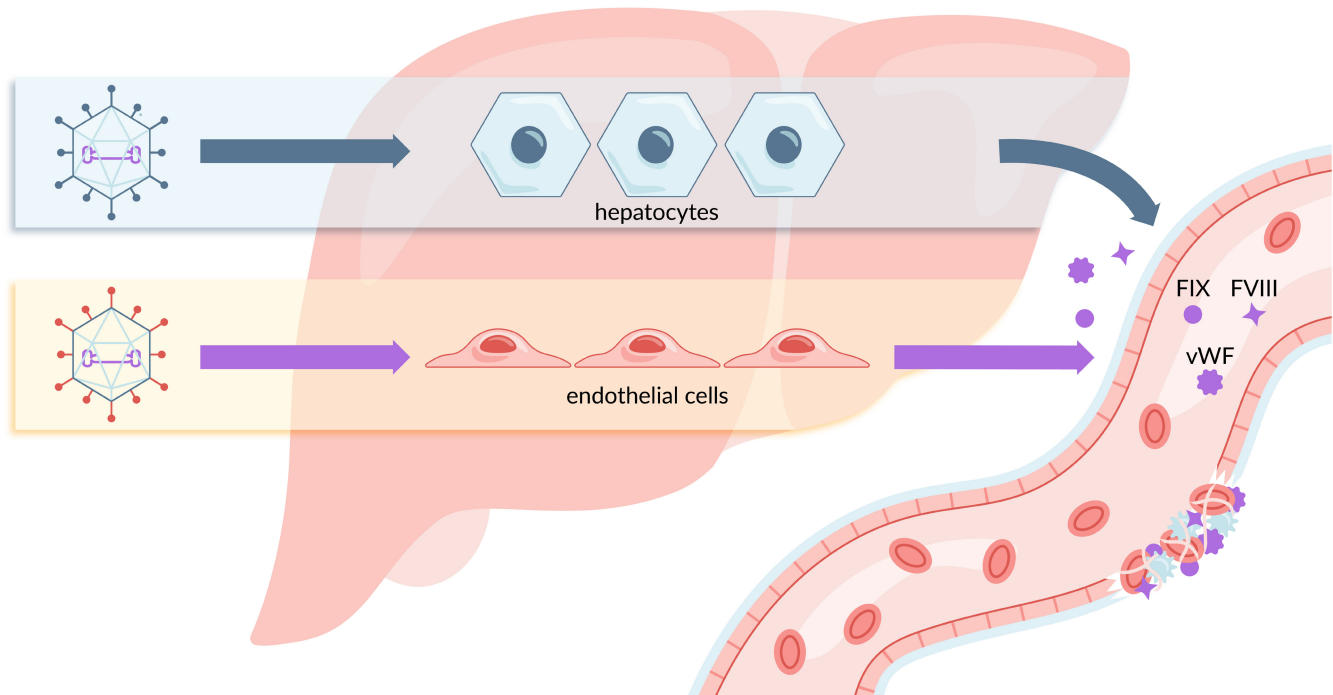


Circulating platelets are specialized cells produced by megakaryocytes through the formation of proplatelets. Leading studies point to the bone marrow niche as the core of hematopoietic stem cell (HSC) differentiation, revealing interesting and complex environmental factors for consideration. Megakaryocytes take cues from the physicochemical bone marrow microenvironment, which includes contact with extracellular matrix components, interaction with endothelium, and contact with the turbulent flow generated by the blood circulation into the sinusoid lumen.¹⁰ Germinal and acquired mutation in HSCs may manifest in altered megakaryocyte maturation, proliferation, and platelet production. Alterations of the whole hematopoietic niche may also occur, highlighting the central role of megakaryocytes in the control of the physiological bone marrow homeostasis. Tissue-engineering approaches have been developed to create a functional mimic of the native tissue.^{11,12} Reproducing the thrombopoietic environment is instrumental to gaining new insight into its activity and answering the growing demand for human platelets for fundamental studies and clinical applications in transfusion medicine. (ECM, extracellular matrix; MK, megakaryocyte; iPSC, induced pluripotent stem cell; MSC, mesenchymal stem cell; PPF, proplatelet formation; PLT, platelet; TPO, thrombopoietin; TGF β , transforming growth factor- β).

The figure was created with [BioRender.com](https://www.biorender.com)

Engineering of adeno-associated viruses to enhance cell-specific transduction

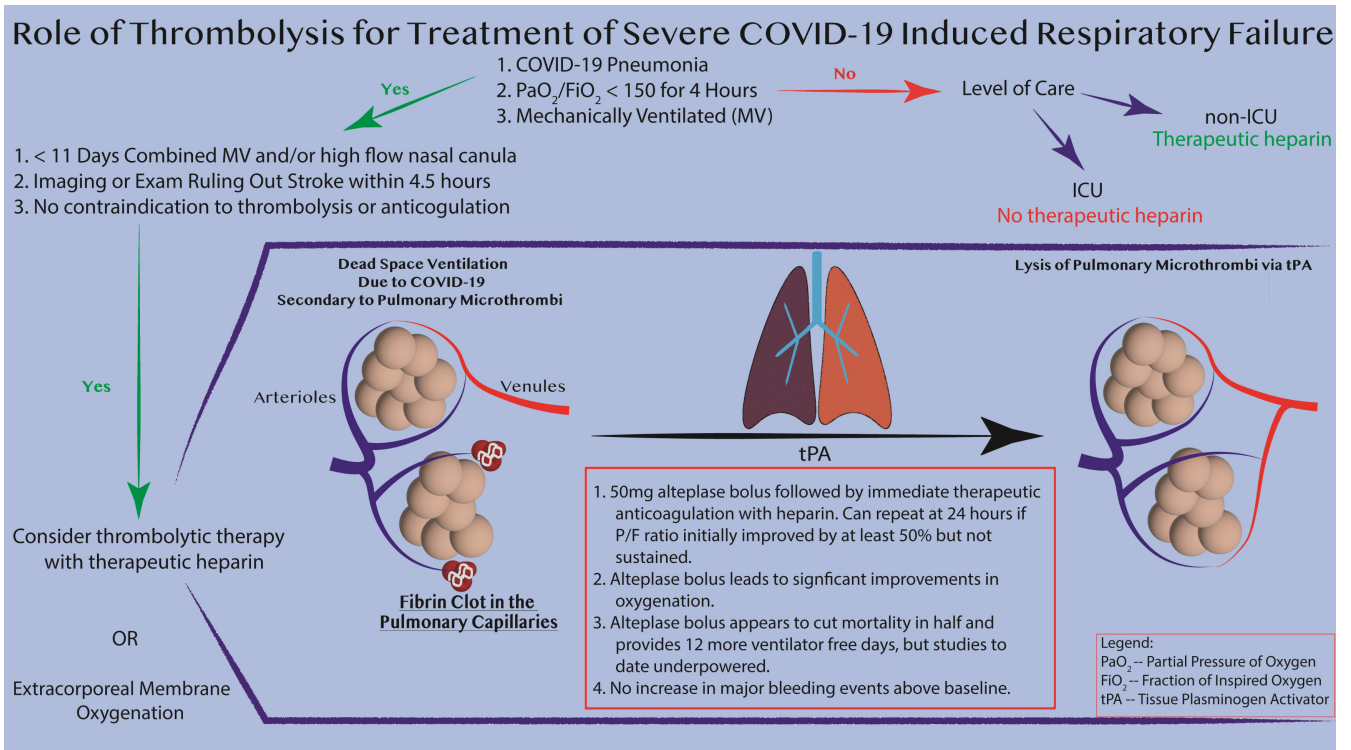
Elena Barbon PhD



Adeno-associated viral vectors have emerged as one of the most promising gene delivery platforms for *in vivo* liver-directed gene therapy for the treatment of blood coagulation disorders, particularly hemophilia A (HA) and B (HB). The ongoing clinical trials are based on adeno-associated virus (AAV)-mediated hepatocyte-targeted expression of coagulation factor VIII or IX for HA and HB, respectively.¹³ Nevertheless, there is increasing interest in the possibility of improving the targeting of different cell types. To this aim, AAV capsid engineering represents an attractive strategy to enhance cell-specific transduction.^{14,15} This may represent a promising approach in coagulation diseases such as HA or von Willebrand disease, where expressing the therapeutic protein from its natural biosynthetic site, that is, endothelial cells, could be beneficial to guarantee an optimized protein biosynthesis, secretion, and activity.

Fibrinolysis and thrombolytic therapy in COVID-19 respiratory failure

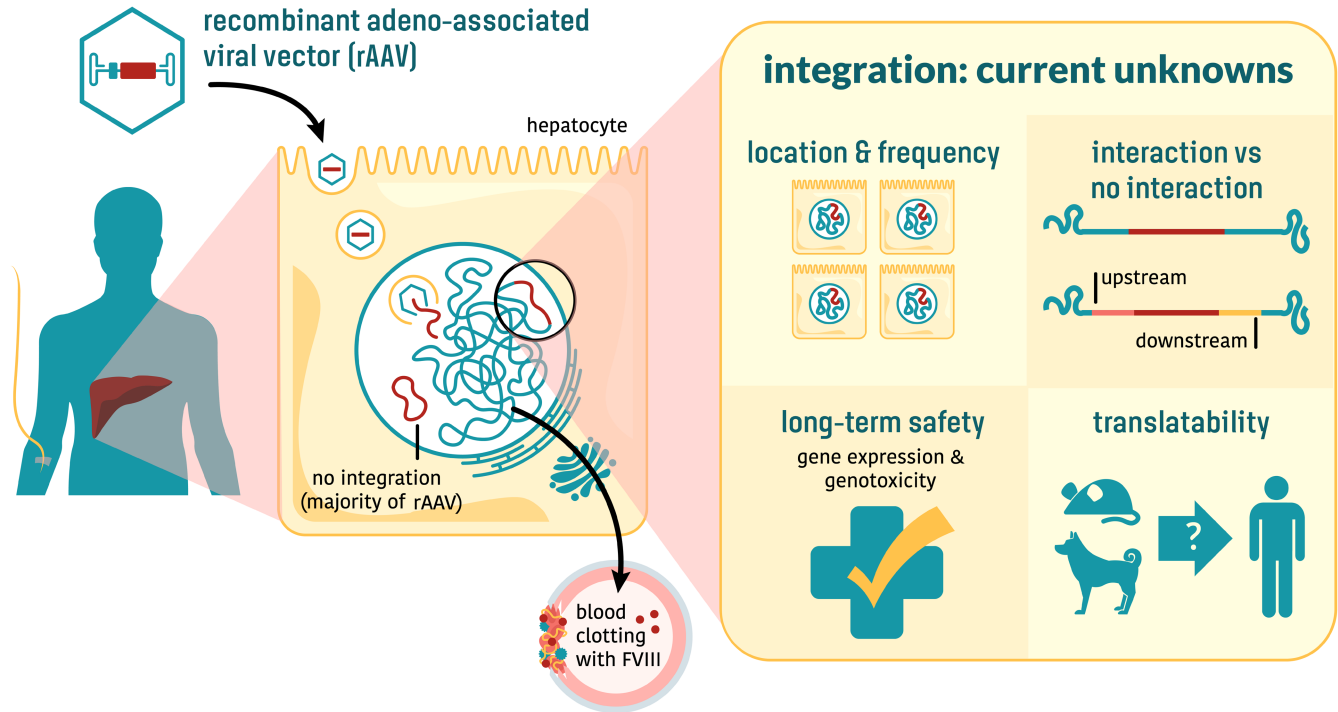
Christopher D. Barrett MD



For references, see Barrett et al.^{16,17}; ATTACC Investigators et al.¹⁸

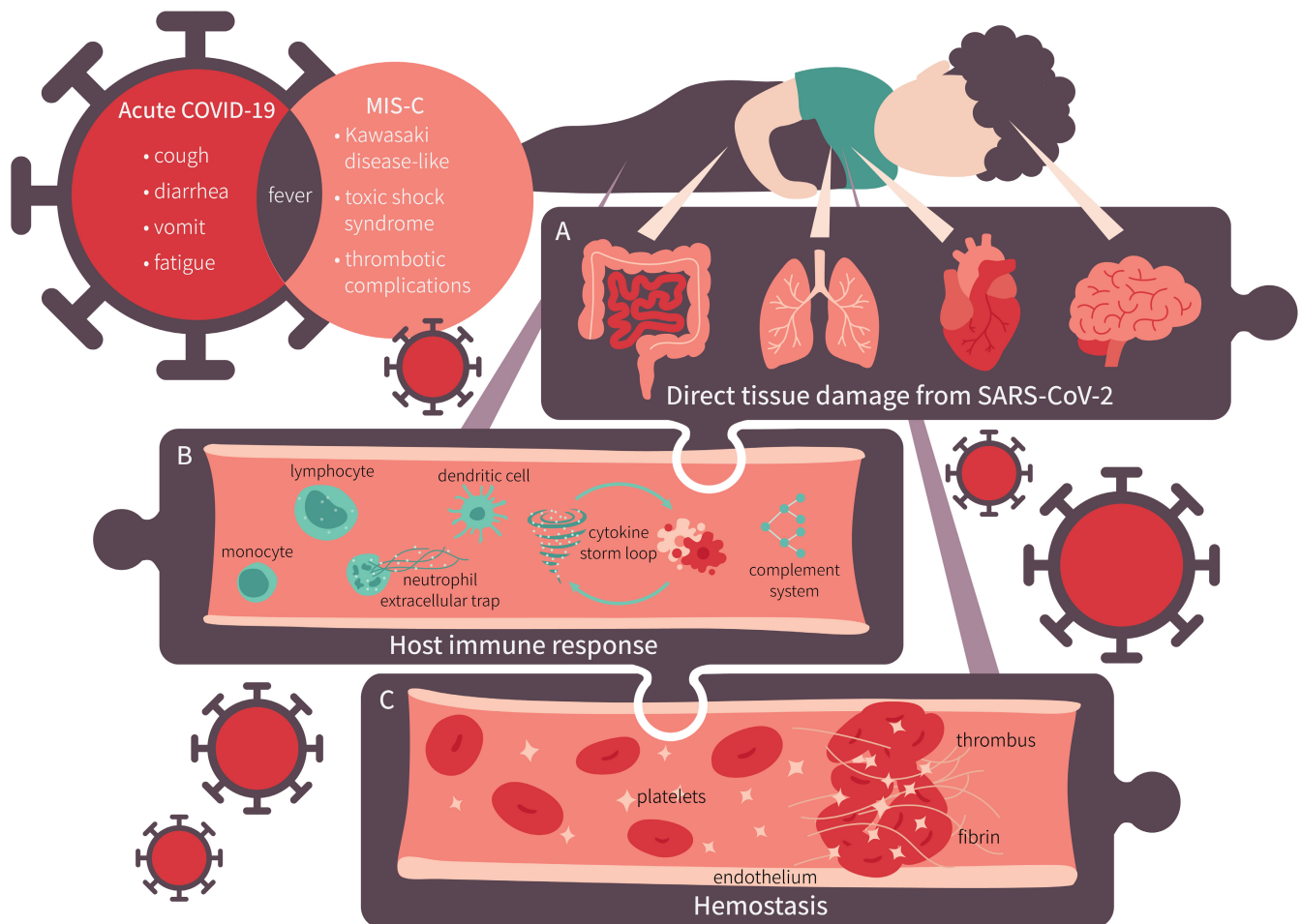
Adeno-associated virus integration

Paul Batty MBBS, PhD



Update on the pathogenesis of COVID-19 and multisystem inflammatory syndrome in children

Jorge David Aivazoglou Carneiro MD, PhD



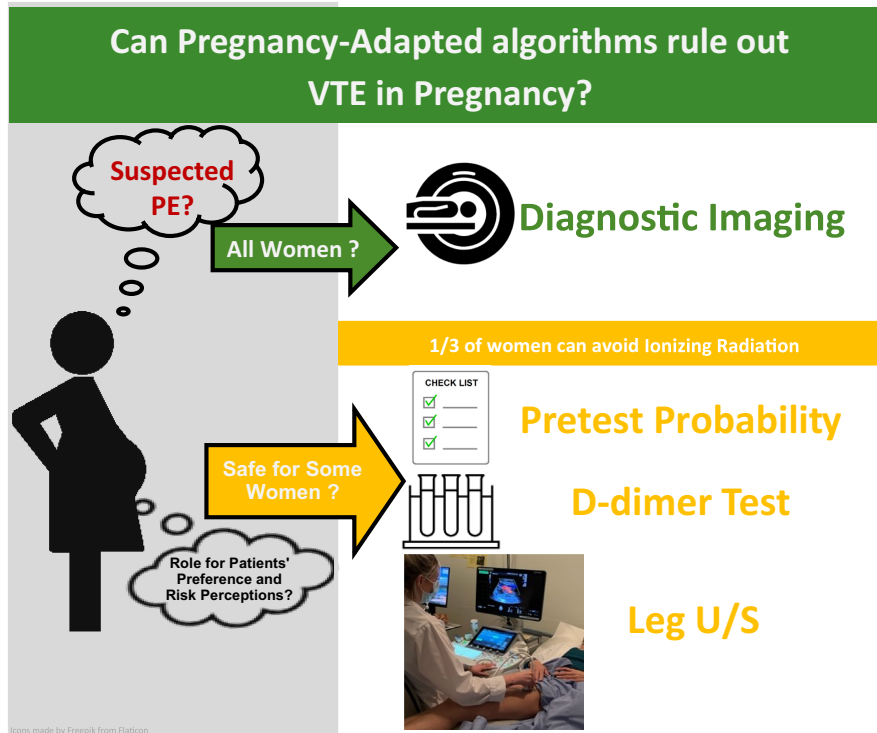
Pathogenesis of COVID-19 and multisystem inflammatory syndrome in children (MIS-C)

1. Severe acute respiratory syndrome coronavirus 2 infection affects several body organs in special respiratory, cardiovascular, gastrointestinal, and nervous systems, causing direct tissue damage.
2. Innate immune system activation is the first step of host immune response initiating inflammatory pathways that provide viral clearance. However, an innate immune response out of control may result in severe COVID-19 or MIS-C.
3. The endothelium plays an important role in the balance of hemostasis. Therefore, endothelial damage and inflammation promote a hypercoagulable state and thrombosis.

For references, see Yuki et al.¹⁹; Gustine and Jones²⁰; Diamond and Kanneganti²¹

Can pregnancy-adapted algorithms rule out venous thromboembolism in pregnancy?

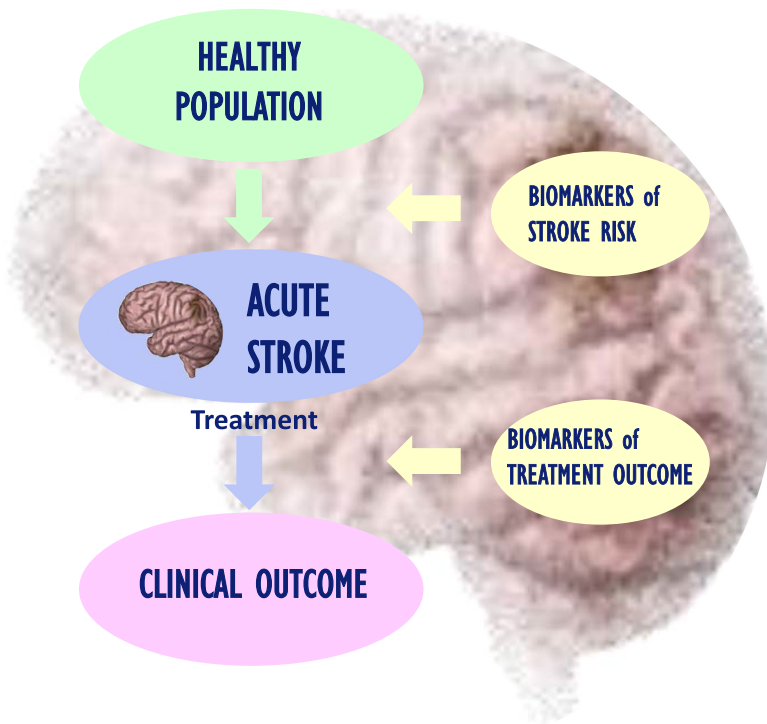
Wee Shian Chan MD, MSc, FRCP



For references, see Konstantinides et al.²²; van der Pol et al.²³; Righini et al.²⁴

Biomarkers for stroke prediction

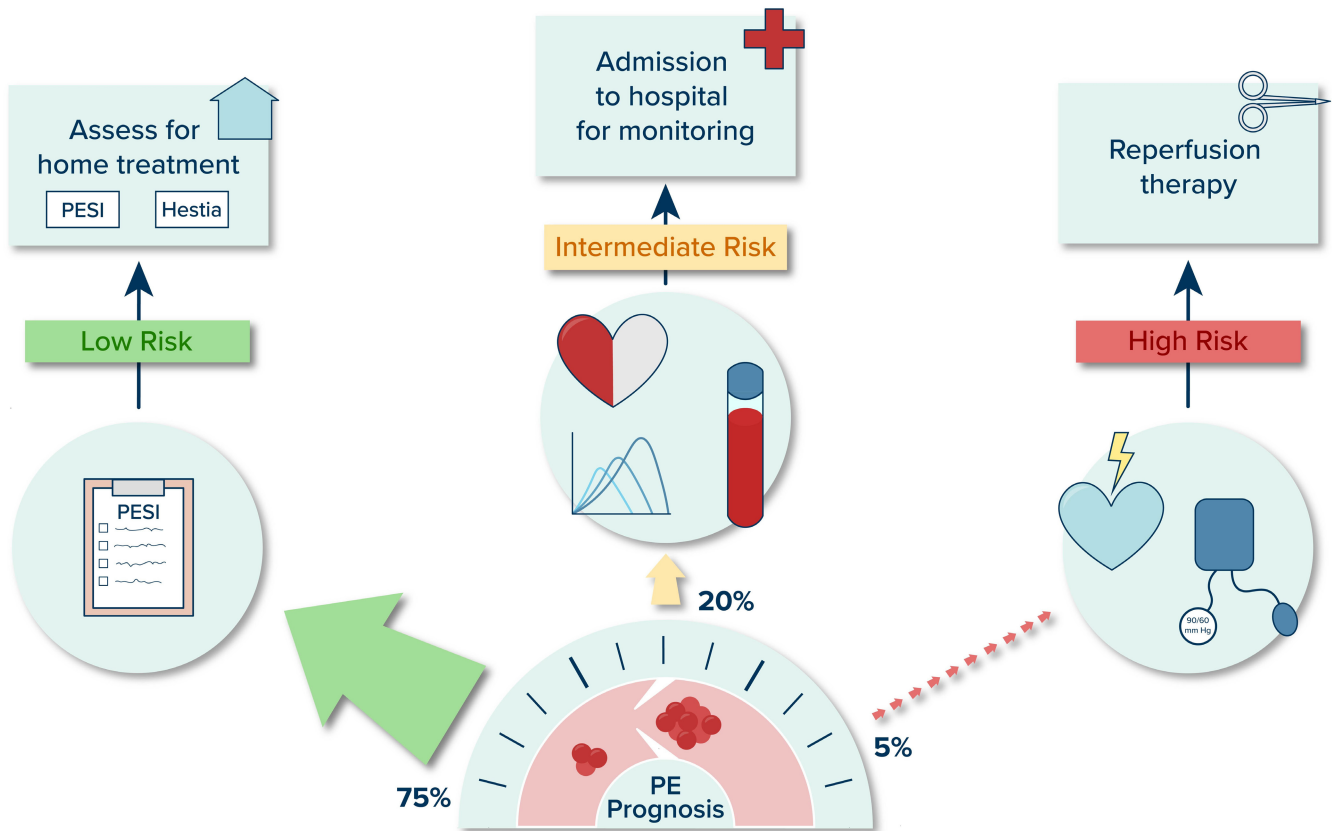
Moniek de Maat PhD



| | ischemic stroke risk in healthy population | outcome after stroke | outcome after tPA treatment |
|---|--|-----------------------|-----------------------------|
| Primary hemostasis von Willebrand factor ADAMTS13 platelet activation markers | ↑ ↓ -/↑ | -/↑ -/↓ -/↑ | - ↓ no data |
| Secondary hemostasis fibrinogen FVIII thrombin generation | ↑ ↑ ↓↑ | -/↑ -/↑ no data | - - no data |
| Fibrinolysis PAI-1 clot lysis time D-dimer | - no data ↑ | -/↑ no data ↑ | -/↑ ↓ no data |
| Thrombus characteristics mechanical, fibrin network | no data | no data | no data |
| Inflammation C-reactive protein, cytokines | ↑ | no data | no data |
| Immunothrombosis NETs, eDNA | no data | no data | ↓ |

Risk stratification for acute venous thromboembolism

Kerstin de Wit MBChB, MD, MSc

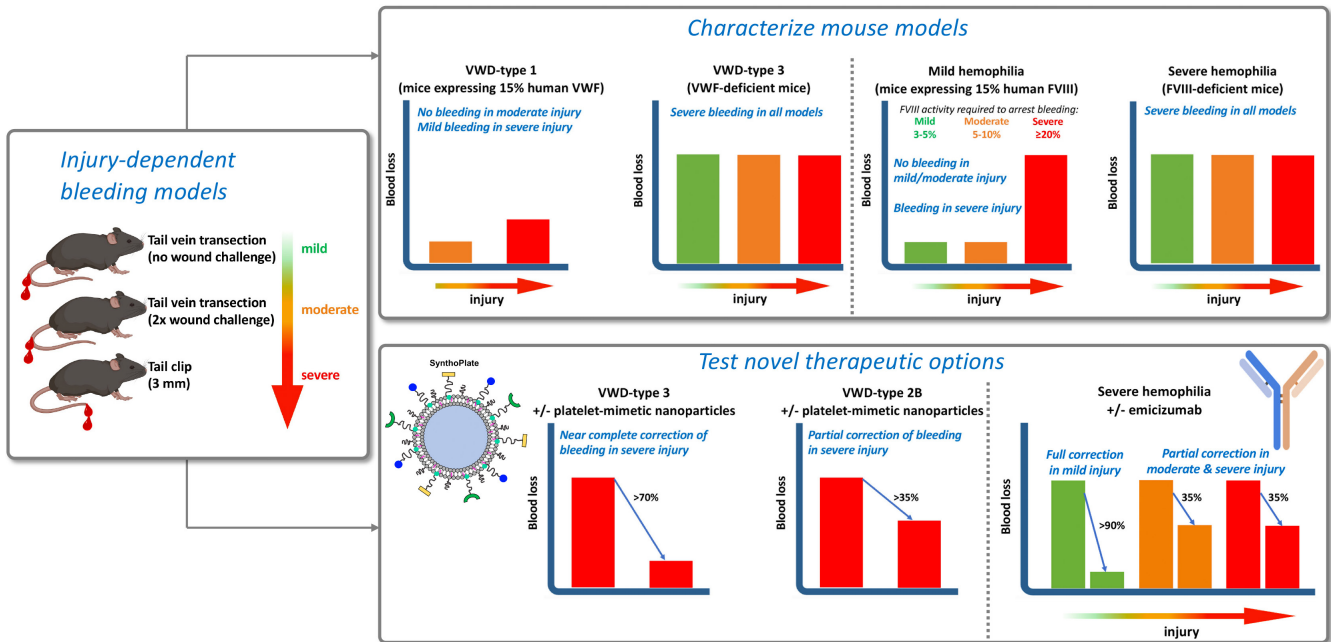


Patients with acute pulmonary embolism (PE) can be classified as having low-risk PE (75% of cases), intermediate-risk PE (20% of cases), or high-risk PE (5% of cases). Low-risk patients can be assessed for outpatient treatment with rapid follow-up. Intermediate-risk patients are admitted for monitoring, and high-risk patients require immediate reperfusion therapy.²²

PESI, pulmonary embolism severity index; VTE, venous thromboembolism.

New animal models for bleeding disorders and treatments

Cécile Denis PhD



FVIII, factor VIII; VWD, von Willebrand disease; VWF, von Willebrand factor.

Myeloproliferative neoplasms in pregnancy: Implications for mother and child

Martin H. Ellis MD

MPN in pregnancy: Incidence



3.2 per 100 00 pregnancies per year

Pregnancy-related complications are common in MPN

Pregnancy loss=28.7%

Adverse outcomes* =9.8%

*VTE, ATE, preeclampsia, abruption, IUGR



MPN in pregnancy: Treatment recommendations

First pregnancy

Maternal VTE /ATE prophylaxis

- LMWH (VTE)
(Only for co-existent VTE risk factors: previous VTE, C/S, advanced age, obesity)
- Aspirin (ATE)

Placenta-related prophylaxis

- Observation or aspirin

Subsequent pregnancies

(in case of previous placenta-related complications)

Maternal VTE/ATE prophylaxis

- As for first pregnancy

Placenta-related prophylaxis

- Interferon
- Aspirin-low dose

MPN=myeloproliferative neoplasm IUGR=Intrauterine growth restriction

VTE=venous thromboembolism ATE=arterial thromboembolism C/S=Cesarean section LMWH=low molecular weight heparin

Polycythemia vera, essential thrombocythemia, and primary myelofibrosis (termed *myeloproliferative neoplasms* [MPNs]) are clonal diseases that may result in fatal end-stage bone marrow fibrosis or acute leukemia. During the long natural history of these diseases, thrombosis is an important complication.

The median age at diagnosis of the MPNs is >60years; however, 20% of patients are <40years old when diagnosed, making MPNs in pregnancy a relevant clinical issue. Indeed, the estimated incidence of MPNs among pregnant women in the United Kingdom is 3.2 in 100 000 pregnancies per year.²⁵

The risk of maternal (venous or arterial thrombosis or hemorrhage), or placenta-related (fetal loss or preeclampsia/eclampsia) complications is higher in pregnancies in patients with MPNs than in the general population. Meta-analysis data demonstrate that 28.7% pregnancies are lost among these patients and that an additional 9.6% of women experience an adverse outcome such as thrombosis, preeclampsia, eclampsia, placental abruption, or intrauterine growth retardation.²⁶

Treatment is based on observational data and expert opinion and includes aspirin, low-molecular-weight heparin, and interferon- α . Prospective studies are needed to better define appropriate treatment for these patients.

Management of thrombocytopenia in pregnancy

Renee Eslick BMed, FRACP, FRCPA

Management of Thrombocytopenia in Pregnancy

Eslick R



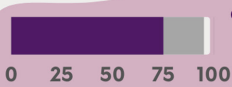
Frequency

1 in 10 women develop thrombocytopenia in pregnancy.



Treatment

- Optimise antenatal iron and haemoglobin
- Delivery for PET
- Corticosteroids and/or IVIg for ITP



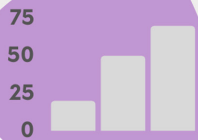
Aetiology

- 75% gestational
- 20% preeclampsia (PET)
- 3% immune (ITP)



Mode of Delivery

- Per obstetrical indications
- Active management of third stage of labour - prophylactic uterotonics and assisted delivery of the placenta



Platelet Thresholds

- Limited evidence and individualised decision making recommended
- In the absence of bleeding, consider aiming for the following thresholds:
- >20x10⁹/L during pregnancy
- >50x10⁹/L for Caesarean or vaginal birth
- >70x10⁹/L for neuraxial anaesthesia



Neonatal Issues

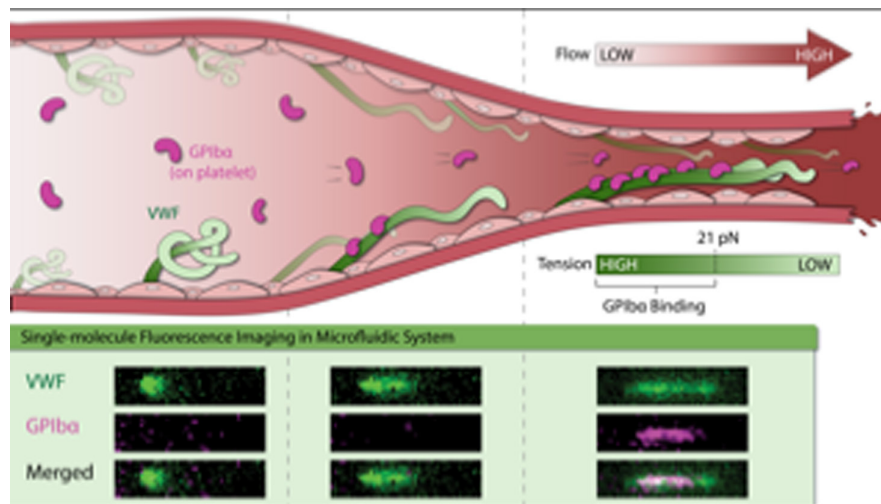
- Avoid instrumental delivery if possible
- For maternal ITP / hereditary thrombocytopenia:
 - Check cord platelet count
 - Evaluate infant for signs/symptoms of bleeding

IVIg, intravenous immunoglobulin; ITP, immune thrombocytopenia; PET, preeclampsia.

For reference, see Eslick et al.²⁷

Binding of flow-activated von Willebrand factor to glycoprotein Ib

Hongxia Fu PhD



Blood protein von Willebrand factor (VWF; green) is essential in thrombosis and hemostasis. Force induced by blood flow is an important regulator to mediate VWF hemostatic function. Dynamic single-molecule imaging in a microfluidic system reveals that a two-step conformational transition induces VWF activation mechanism.^{28,29} First, VWF elongates from compact to linear form. Second, a tension above 21 pN (dark green shading) induces VWF transition to a state with high affinity for platelet glycoprotein Ib α (GPIb α ; magenta). For clarity, platelets are not shown. GPIb α dissociates rapidly when tension is under 21 pN. This mechanism allows VWF to be activated by hydrodynamic force at sites of hemorrhage but avoid thrombus formation downstream. (Single-molecule fluorescence images were reproduced from Fu et al.,²⁸ *Nature Communications*, 2017 with modifications and shared via a Creative Commons 4.0 license.)

Hemostatic defects in fibrinolytic disorders

Catherine P. M. Hayward MD, PhD

Inherited Fibrinolytic Diseases Phenotype

| | |
|--|--|
| <p>SHARED</p> <ul style="list-style-type: none"> • Increased bleeding, delayed onset • Responsive to fibrinolytic inhibitors • Frequently: delayed diagnosis | <p>PAI-1 DEFICIENCY ONLY</p> <ul style="list-style-type: none"> • Antenatal bleeds • Cardiac fibrosis • Longevity <p>QPD ONLY</p> <ul style="list-style-type: none"> • Reduced platelet counts |
|--|--|

PLASMINOGEN

PAI-1 (1) → tPA → Plasminogen → Plasmin

PAI-1 (2) → uPA → Plasminogen → Plasmin

(F11a, F12a, kallikrein) → Plasminogen

Plasmin → α₂ antiplasmin (α₂macroglobulin) (3)

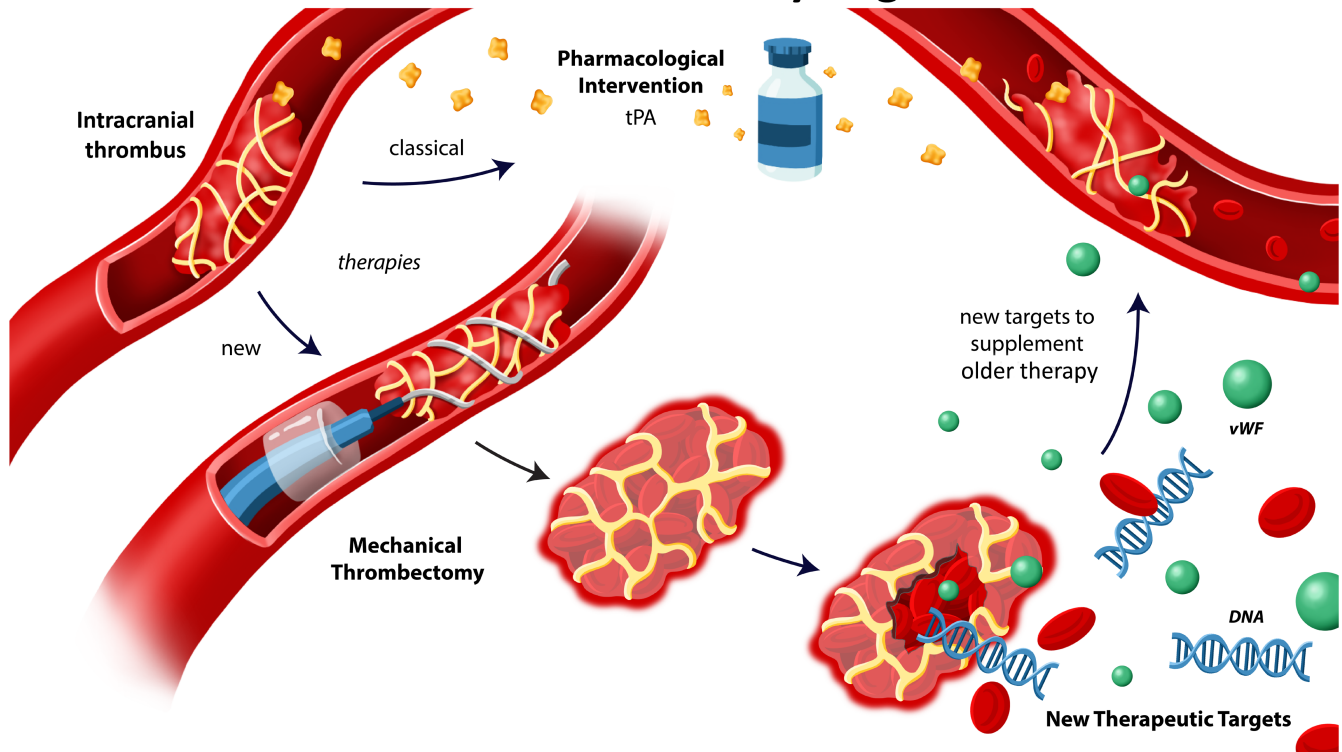
| Inherited fibrinolytic disorder | Pathophysiology | Diagnosis |
|---------------------------------|--|---|
| 1. PAI-1 deficiency | <ul style="list-style-type: none"> • ↑↑↑ fibrinolysis (loss-of-inhibitor defect) • Cause: ↓↓↓ uPA & tPA inhibition resulting in ↑↑↑ plasmin generation | <ul style="list-style-type: none"> • ↓↓↓ ELT • ↓↓↓ PAI-1 antigen • Can ↓α2 antiplasmin • Genetic investigations |
| 2. Quebec platelet disorder | <ul style="list-style-type: none"> • ↑↑↑ fibrinolysis (gain-of-function defect) • Cause: PLAU "rewired" in megakaryocytes → ↑↑↑ platelet uPA • ↑↑↑ plasmin generation in wounds (& in α-granules) | <ul style="list-style-type: none"> • Genetic test for QPD duplication mutation |
| 3. α2 antiplasmin deficiency | <ul style="list-style-type: none"> • ↑↑↑ fibrinolysis (loss-of-inhibitor defect) • Cause: ↓↓↓ plasmin inhibition | <ul style="list-style-type: none"> • ↓↓↓ α2-antiplasmin • Genetic investigations |

Accelerated Lysis of Platelet Fibrin Clot

Thrombus composition and thrombolysis resistance in stroke

Benoit Ho-Tin-Noé PhD

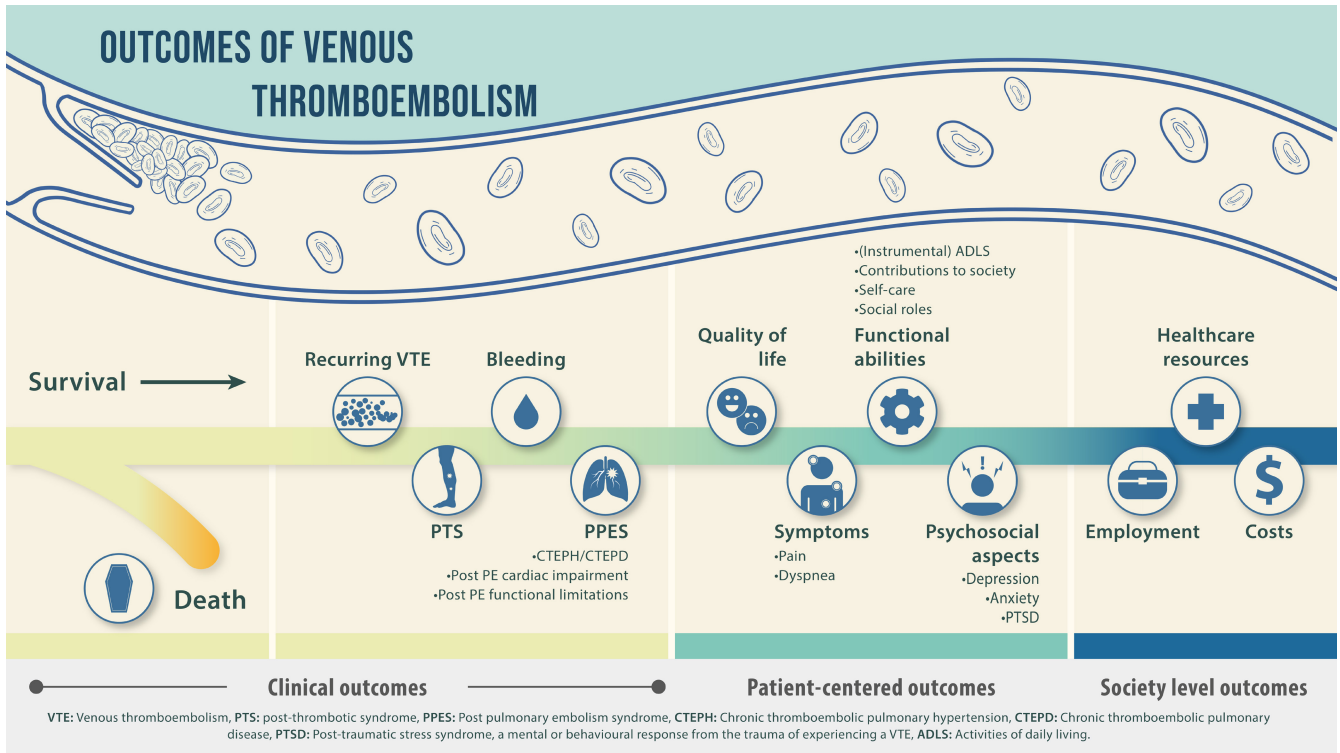
Past, present and future recanalization therapies for acute ischemic stroke caused by large vessel occlusion



The introduction of mechanical thrombectomy has considerably increased the rate of successful recanalization in large-vessel occlusion stroke. From a basic research standpoint, it has enabled the analysis of stroke thrombi and a better understanding of the mechanisms underlying resistance to intravenous thrombolysis. Therefore, although the efficacy of mechanical thrombectomy could have caused the end of thrombolysis, it has instead opened the way for its rethinking and improvement, through the development of innovative thrombolytic strategies.

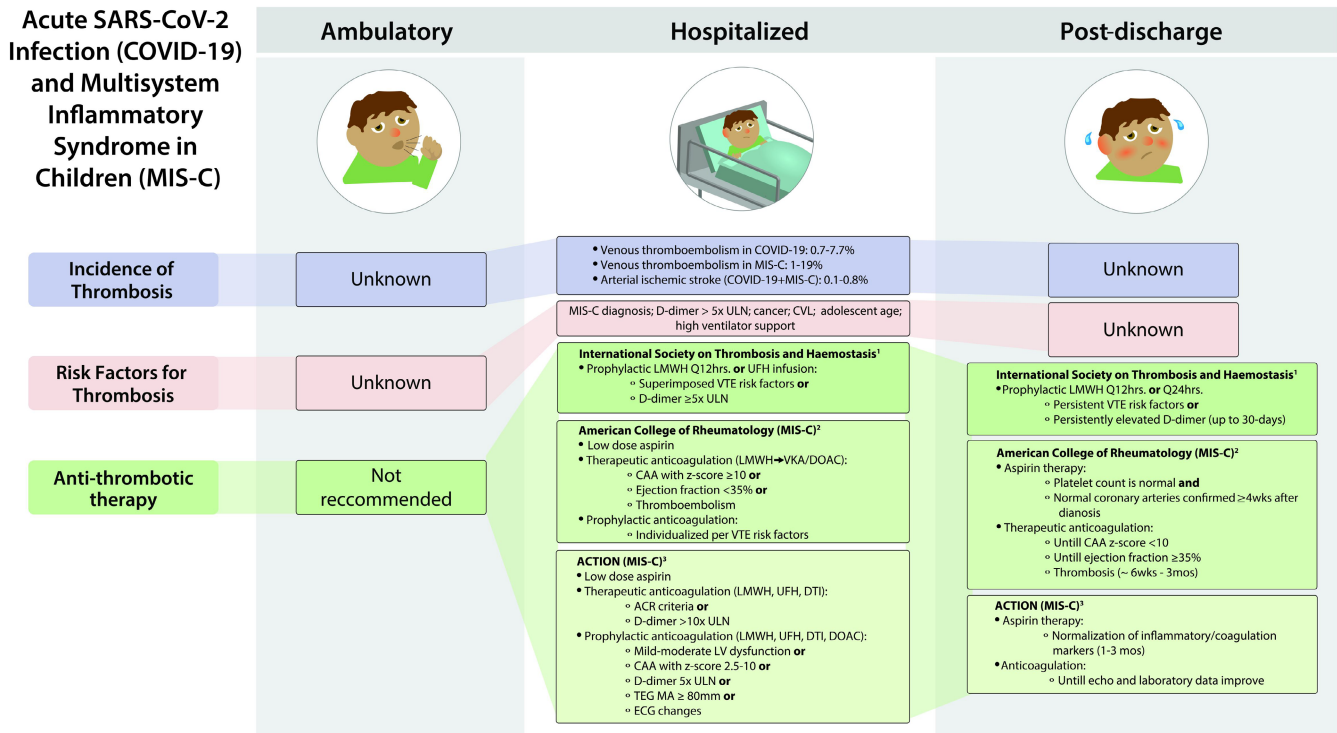
Outcomes of venous thromboembolism

Frederikus A. Klok MD, PhD



Multisystem inflammatory syndrome in children and thrombotic complications of COVID-19 in children

Riten Kumar MD, MSc



ACR, American College of Rheumatology; ACTION, Advanced Cardiac Therapies Improving Outcomes Network; anti-FXa, anti-factor Xa; CAA, coronary artery aneurysm; CVL, central venous line; DOAC, direct oral anticoagulant; DTI, direct thrombin inhibitor; ECG, electrocardiogram; LMWH, low-molecular-weight heparin; LV, left ventricular; MA, maximum amplitude; TEG, thromboelastogram; UFH, unfractionated heparin; ULN, upper limit of normal; VKA, vitamin K antagonist; VTE, venous thromboembolism

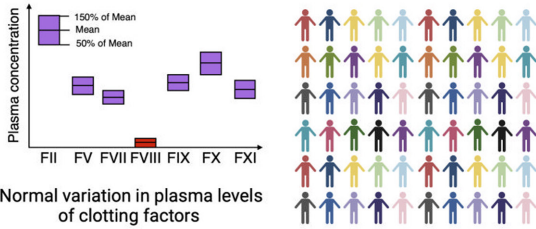
For references, see Goldenberg et al.³⁰; Henderson et al.³¹; Bansal et al.³²

Mathematical modeling to study variability in bleeding

Karin Leiderman PhD

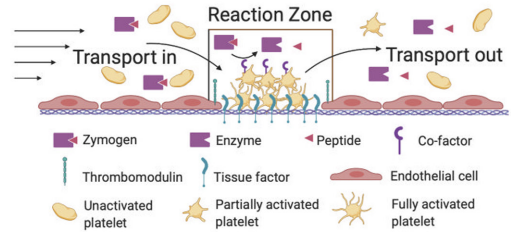
Mathematical Modeling to Study Variability in Bleeding

Create synthetic data set representing the plasma levels of thousands of hemophilia patients



Normal variation in plasma levels of clotting factors

Mathematical Model of Flow-Mediated Coagulation

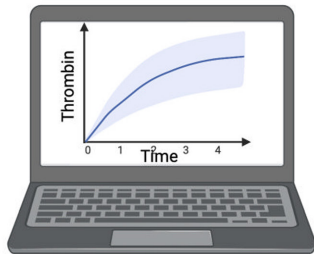


$$\left(\text{Rate of change of concentrations in reaction zone} \right) = \left(\text{Rate of supply} \right) + \left(\text{Rate of removal} \right) + / - \left(\text{Rate of generation, inhibition, etc.} \right)$$

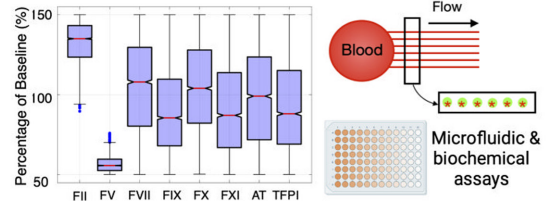
Perform thousands of simulations using synthetic data set as input to the mathematical model



Each simulation takes only seconds of real time



Statistically analyze the model output, identify modifiers of bleeding, experimentally validate

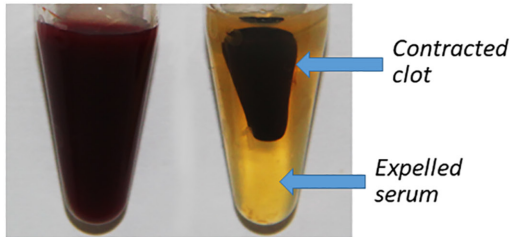


Blood clot contraction: Mechanisms, pathophysiology, and disease

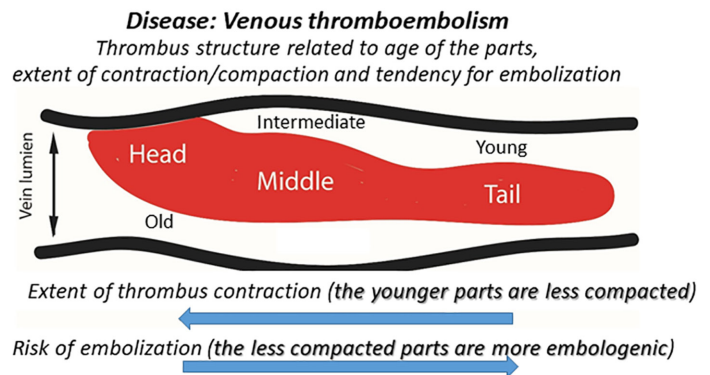
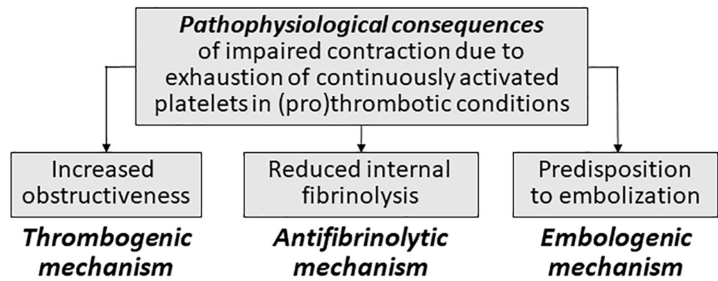
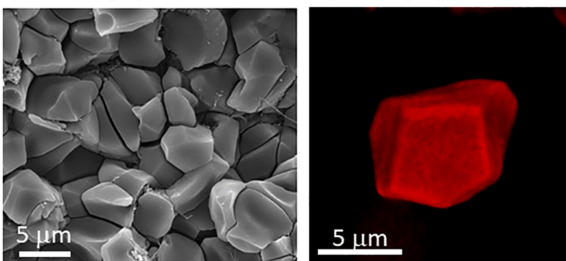
Rustem I. Litvinov MD, PhD

Blood Clot Contraction: Mechanisms, Pathophysiology, and Disease

Contraction (retraction, shrinkage) of *in vitro* clots and *in vivo* thrombi is driven by activated platelets



A new pathogenic mechanism: Contraction of clots and thrombi results in formation of **polyhedrocytes** - tightly packed, highly impermeable, compressed polyhedral RBCs

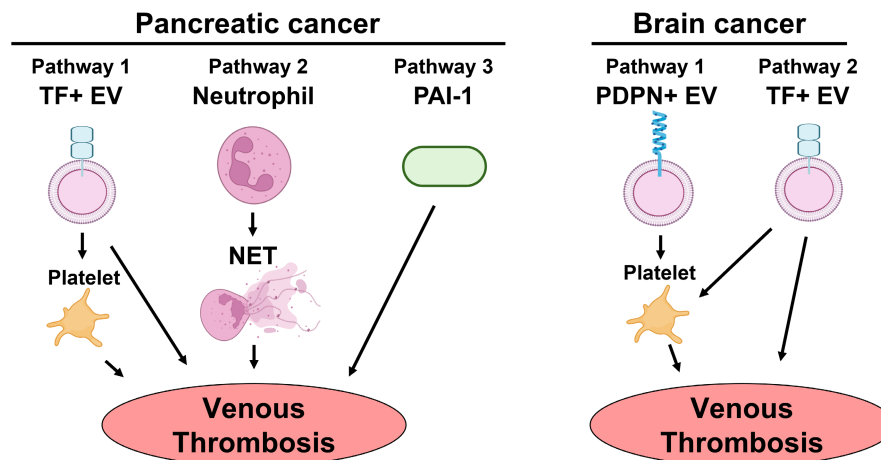


Contraction (retraction) of blood clots and thrombi is clinically important since it can affect their obstructiveness, permeability, susceptibility to fibrinolysis, and the propensity to rupture (embologenicity). The rate and extent of clot contraction *in vitro* and *in vivo* can be enhanced or inhibited by altered platelet functionality as well as pathological cellular and molecular composition of the blood.³³ Clot contraction is reduced in the blood of patients with various (pro)thrombotic conditions, due to continuous platelet activation followed by their exhaustion and reduced contractility.³⁴ Impaired clot contraction is a significant but understudied and underappreciated pathogenic factor that can influence the course and outcomes of thrombotic disorders.³⁵

Mechanisms of cancer-associated thrombosis

Nigel Mackman PhD

Pathways of Cancer-Associated Thrombosis








The majority of studies on the mechanisms of cancer-associated thrombosis (CAT) have focused on pancreatic cancer and brain cancer because they have the highest rates of venous thromboembolism (VTE). There are several pathways that contribute to venous thrombosis in pancreatic and brain cancer. Human studies have identified circulating biomarkers that are associated with VTE. Mouse cancer models are used to directly examine the role of a pathway in venous thrombosis. Taken together, these studies suggest that tissue factor-positive extracellular vesicles (EVs) directly and indirectly (via activation of platelets) enhance venous thrombosis in both pancreatic and brain cancer.^{36,37} Neutrophils form neutrophil extracellular traps that can enhance venous thrombosis in pancreatic cancer.³⁶ Plasminogen activator inhibitor-1 contributes to venous thrombosis in pancreatic cancer by inhibiting fibrinolysis.³⁸ In brain cancer, podoplanin-positive EVs activate platelets and enhance venous thrombosis.³⁷ Targeting these pathways may reduce CAT.

Image created with [BioRender.com](https://www.biorender.com)

When to use tranexamic acid in bleeding

Zoe McQuilten MBBS, PhD

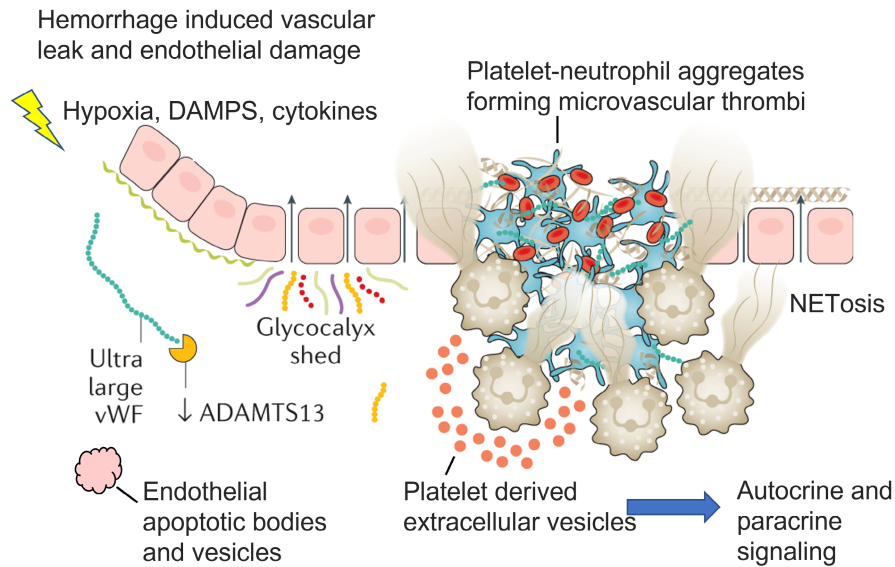
SUMMARY OF RANDOMIZED CONTROLLED TRIALS OF TRANEXAMIC ACID FOR BLEEDING

| CLINICAL SITUATION | | TRANEXAMIC ACID | | | EVIDENCE TO SUPPORT USE? | | |
|--------------------------------------|--|---|--|--|--|---------------|---|
| | RCT# | DOSE (IV)* | MORTALITY | OTHER | THROMBO-EMBOLISM | SEIZURES | |
| TRAUMATIC HEMORRHAGE |  5 | Loading: 1g/10 min Infusion: 1g/8 hr vs. placebo | EVIDENCE OF BENEFIT | | NO DIFFERENCE | NO DIFFERENCE | ✓ YES, best if given within 3 hrs |
| TRAUMATIC BRAIN INJURY |  9 | Loading: 1g/10 min Infusion: 1g/8 hr vs. placebo | EVIDENCE OF BENEFIT if given <3hrs of mild-moderate injury | Possibly reduced hematoma expansion | NO DIFFERENCE | NO DIFFERENCE | ✓ MAYBE, if given to mild-moderate TBI within 3 hrs |
| SPONTANEOUS INTRACEREBRAL HEMORRHAGE |  4 | Loading : 1g Infusion: 1g/8 hr vs. placebo | NO EVIDENCE OF BENEFIT | Reduced hematoma expansion | NO DIFFERENCE | NO DIFFERENCE | ● NO, further research needed |
| POSTPARTUM HEMORRHAGE |  2 | 1g + 1g if bleeding after 30 min or stopped and restarted within 24hrs, vs. placebo | EVIDENCE OF REDUCTION IN DEATH due to bleeding | No change in hysterectomy or transfusion rates | NO DIFFERENCE | NO DIFFERENCE | ✓ YES, best if given within 3 hrs of birth |
| GASTRO-INTESTINAL HEMORRHAGE |  5 | Loading Dose: 1g Infusion: 3g/24 hrs vs. placebo | NO EVIDENCE OF BENEFIT | | HIGHER RISK OF VENOUS THROMBO-EMBOLISM | HIGHER RISK | ● NO |

*STUDIED IN LARGEST TRIAL

Platelet and endothelial biology in bleeding

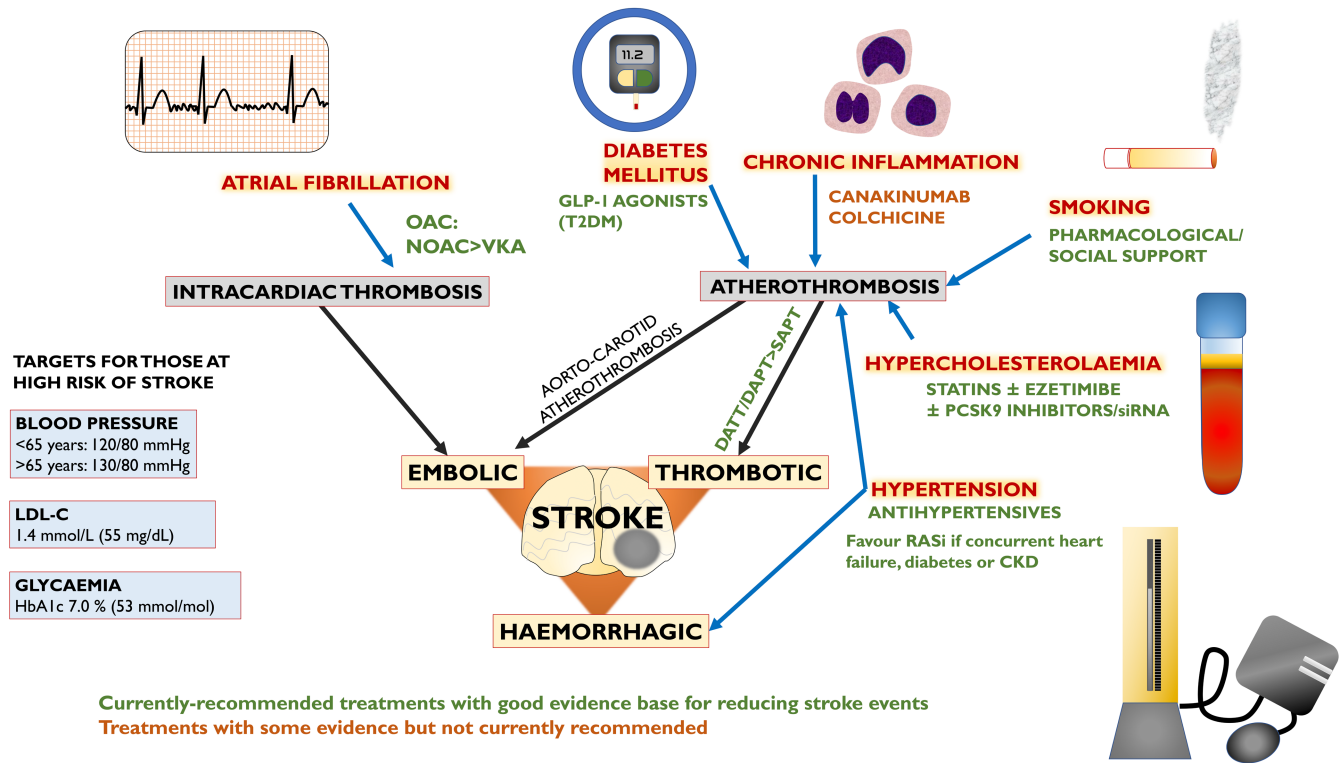
Matthew D. Neal MD



Following trauma and hemorrhage, the systemic signals of hypoxia, danger-associated molecular patterns and cytokines produce endothelial injury in addition to direct endothelial disruption.³⁹ The platelet is the sentinel responder to injury, orchestrating the complex processes of hemostasis and coagulation through interaction with damaged endothelium.⁴⁰ This State of the Art will review the adaptive and maladaptive responses of platelets and endothelium to bleeding with a focus on key mediators including von Willebrand factor, the metalloprotease ADAMTS-13,⁴¹ neutrophil extracellular trap release, and the role of endothelial and platelet extracellular vesicles in cell signaling.

Drug therapy for stroke prevention

William A. E. Parker MB, PhD, MRCP

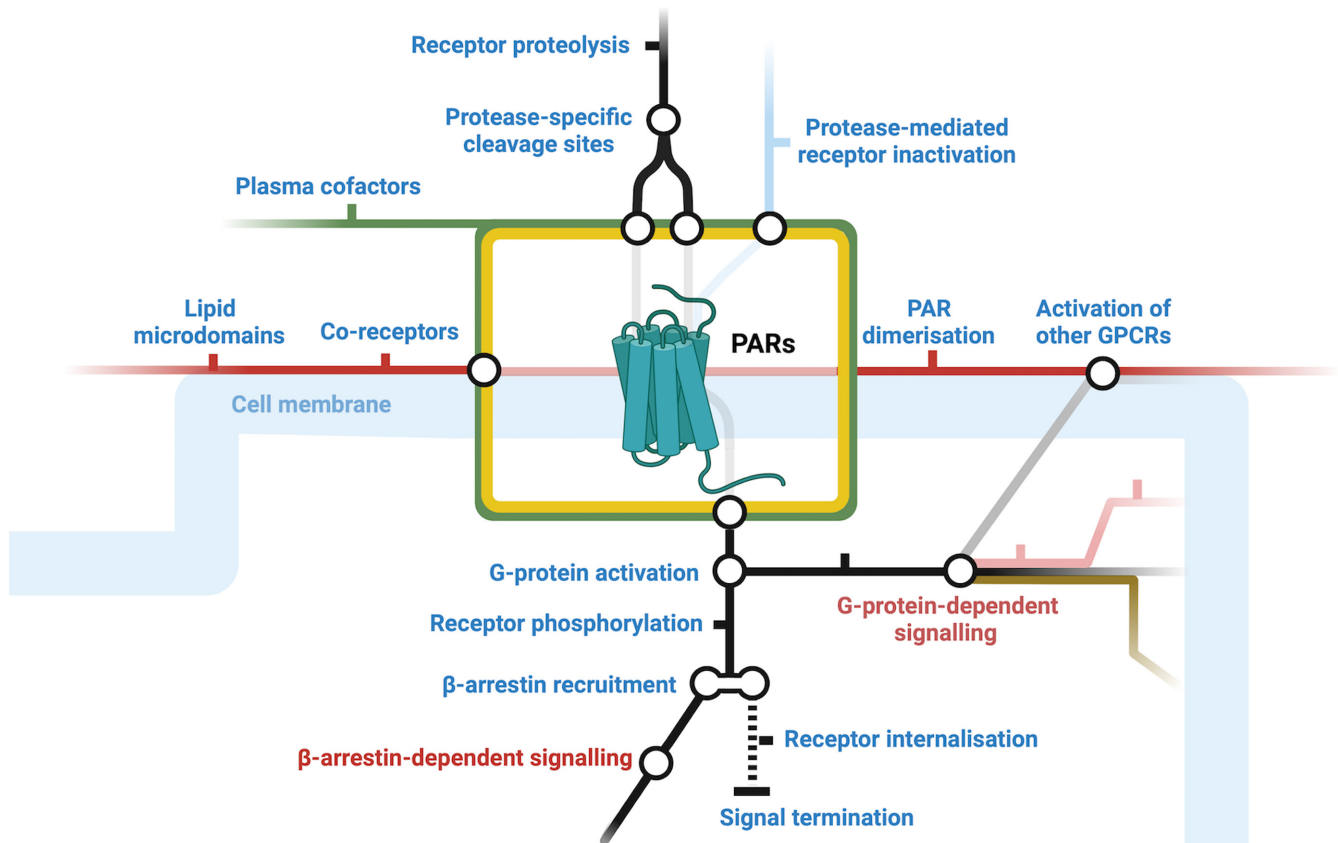


Drugs for control of modifiable risk factors in the prevention of stroke. Modified with permission from Parker et al. 2020 under Creative Commons CC-BY-NC license.⁴² CKD, chronic kidney disease; DAPT, dual antiplatelet therapy; DATT, dual antithrombotic therapy; GLP-1, glucagonlike peptide 1; HbA_{1c}, glycated hemoglobin; LDL-C, low-density lipoprotein cholesterol; LEAD, lower-extremity arterial disease; NOAC, non-vitamin-K antagonist oral anticoagulant; OAC, oral anticoagulant; PCSK9, proprotein convertase subtilisin/kexin type 9; RASi, renin-angiotensin system inhibitor; SAPT, single antiplatelet therapy; T2DM, type 2 diabetes mellitus; VKA, vitamin K antagonist.

Coagulation factor signaling

Roger J. S. Preston PhD

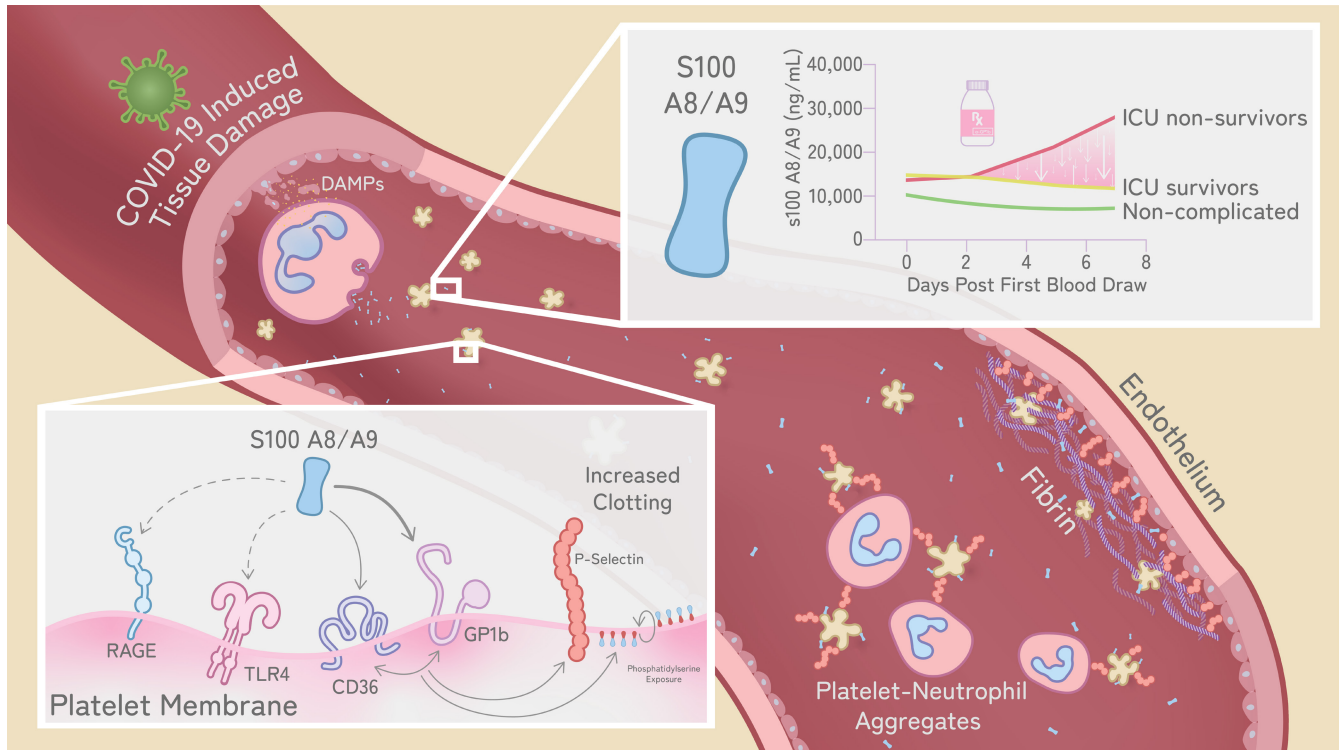
Determinants of coagulation protease signalling



Most coagulation factor cell signaling is mediated by protease-activated receptors (PARs), a unique family of G-protein-coupled receptors whose activation is triggered by receptor proteolysis. Cell signaling output arising from PAR activation is shaped by both the activating protease and the specific cellular contexts in which receptor activation has occurred. Although not yet fully understood, intra- and extracellular regulatory determinants of PAR signaling diversity include protease-specific cleavage sites, association with protease coreceptors, receptor oligomerization, and recruitment of distinct signaling intermediates. These mechanisms combine to enable coagulation proteases to initiate a spectrum of downstream signaling pathways in multiple cell types.

Novel mechanisms of thromboinflammation during infection and hemolysis

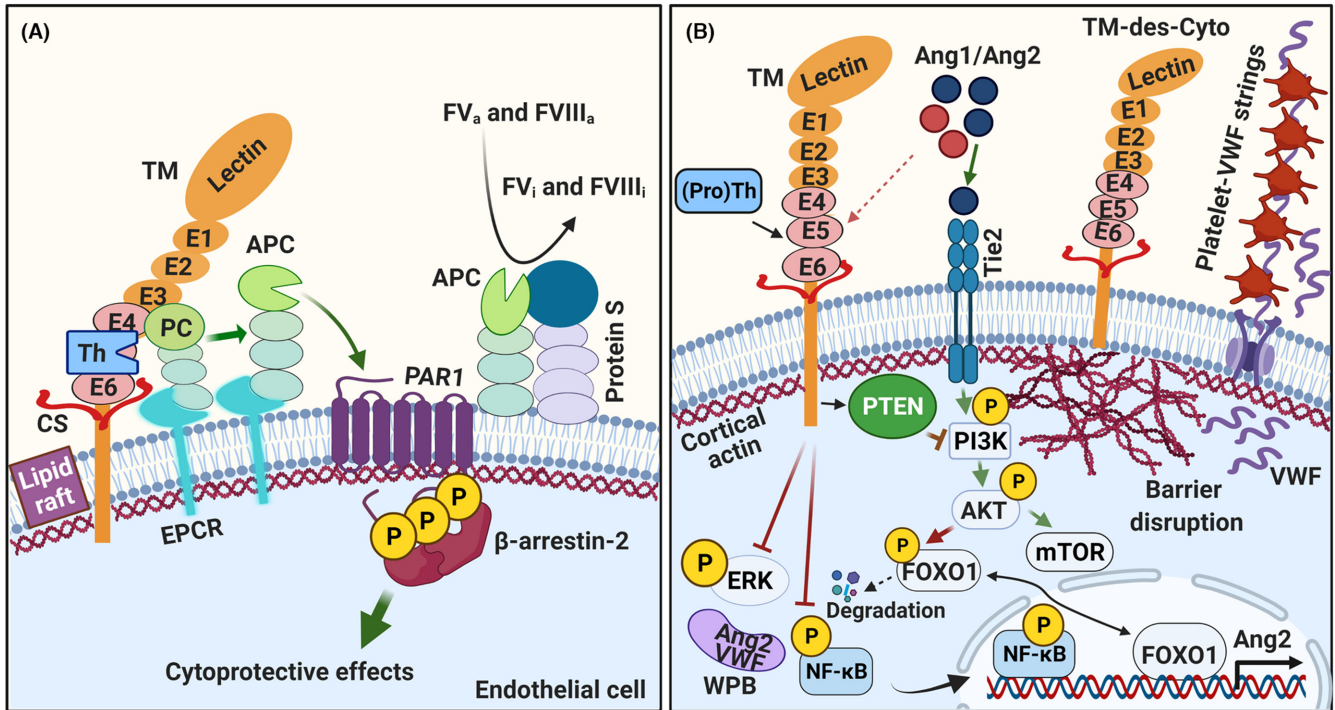
Julie Rayes PhD



- S100A8/A9 is a damage-associated molecular pattern with prothrombotic and proinflammatory properties.
- S100A8/A9 levels are increased in the plasma of patients with COVID-19.
- Deposition of S100A8/A9 on the vessel wall in patients with COVID-19 correlates with thrombotic complications.
- S100A8/A9 induces the formation of procoagulant platelets and amplifies fibrin generation.
- Glycoprotein Iba is the main receptor for S100A8/A9 on platelets, with a supporting role for CD36.

Thrombomodulin regulates endothelial quiescence

Alireza R. Rezaie PhD



(A)

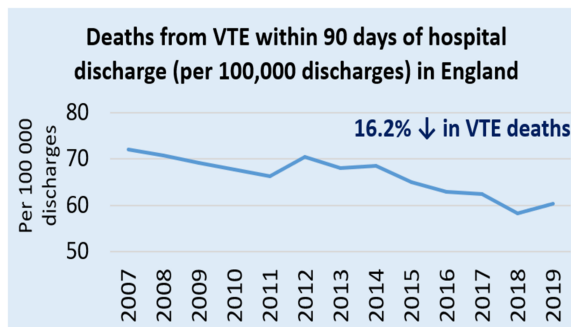
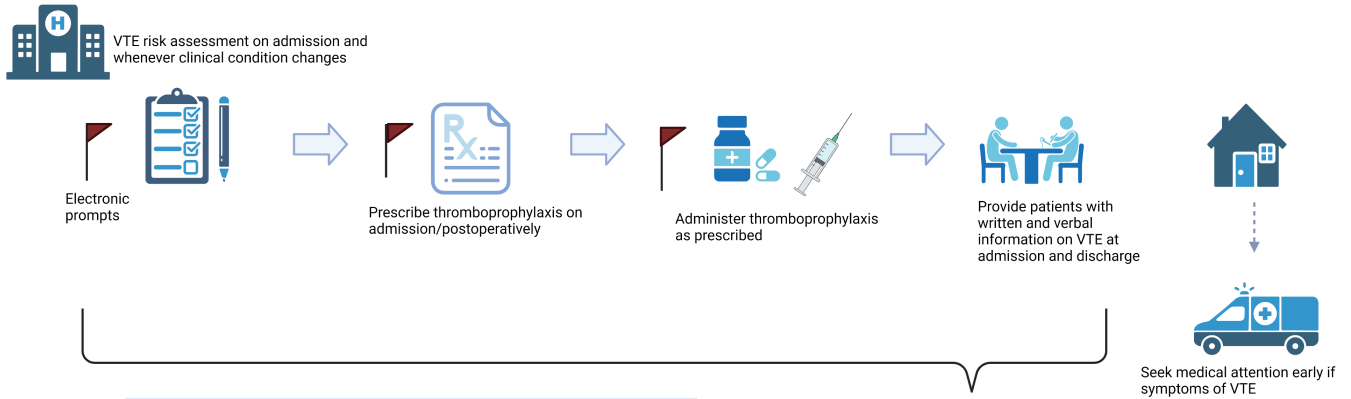
1. Thrombomodulin (TM), endothelial protein C receptor (EPCR), and protease-activated receptor-1 (PAR1), are colocalized in lipid rafts of endothelial cells.
2. Thrombin binds TM and activates EPCR-bound protein C.
3. Activated protein C (APC) in association with EPCR functions in the anti-inflammatory pathway by activating PAR1 and inducing cytoprotective β -arrestin-2 biased signaling.
4. APC functions in the anticoagulant pathway by binding protein S and inactivating factors Va and VIIIa ($FV_i/FVIII_i$).

(B)

1. TM has anti-inflammatory and barrier-stabilizing functions and extracellular signal-regulated and nuclear factor kappa-light-chain-enhancer of activated B cells signaling.
2. (Pro)exosite-1 of (pro)thrombin binds epidermal growth factor (EGF)-like domains of TM and induces cytoprotective signaling.
3. Cytoplasmic domain of TM is involved in recruiting phosphatase and tensin homolog to plasma membrane, thereby regulating endothelial cell proliferation, angiogenesis, and metabolism through PI3K/AKT/FOXO1/mTOR signaling.
4. TM knockdown or deletion of its cytoplasmic-domain leads to barrier destabilization, constitutive expression/secretion of von Willebrand factor (VWF), cell surface platelet-VWF strings formation, reduced angiopoietin 2 expression and deregulated endothelial cell proliferation, angiogenesis and Tie2 receptor signaling. (Pro)Th, (pro)thrombin; WPB, Weibel Palade body.

Systematic approach to venous thromboembolism prevention

Lara N. Roberts MBBS, MD Res



- Local audit/learning from hospital-associated VTE
- Central monitoring of process and clinical outcomes

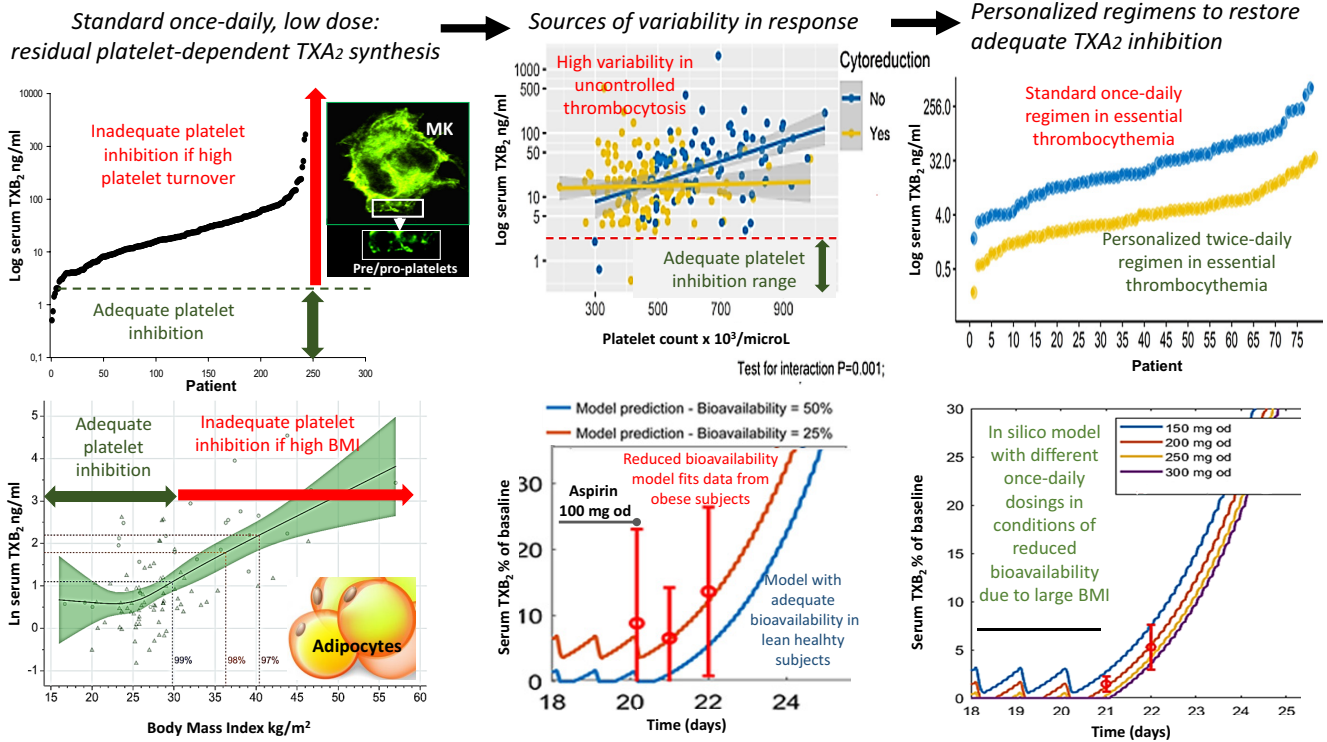
Created with BioRender.com

VTE, venous thromboembolism.

For references, see Roberts et al.⁴³; NHS Digital⁴⁴

Personalizing antiplatelet therapy based on platelet turnover and metabolic phenotype

Bianca Rocca MD, PhD



Variability in the response to fixed-dose antiplatelet drugs can be increased by primary (essential thrombocythemia) or secondary (reactive inflammatory states) pathological conditions associated with increased platelet turnover and count, that is, pharmacodynamic (PD) variability, as well as by high-degree obesity, that is, pharmacokinetic (PK) variability.

Understanding PK- and/or PD-related mechanisms of variability is central to design personalized regimens and to optimize the risk/benefit balance of common antiplatelet drugs by increasing the daily frequency or the once-daily dosing.

Moreover, in silico PK/PD modeling and in silico trials can be instrumental in designing clinical trials, developing antithrombotic therapy, and generating new personalized drug regimens.

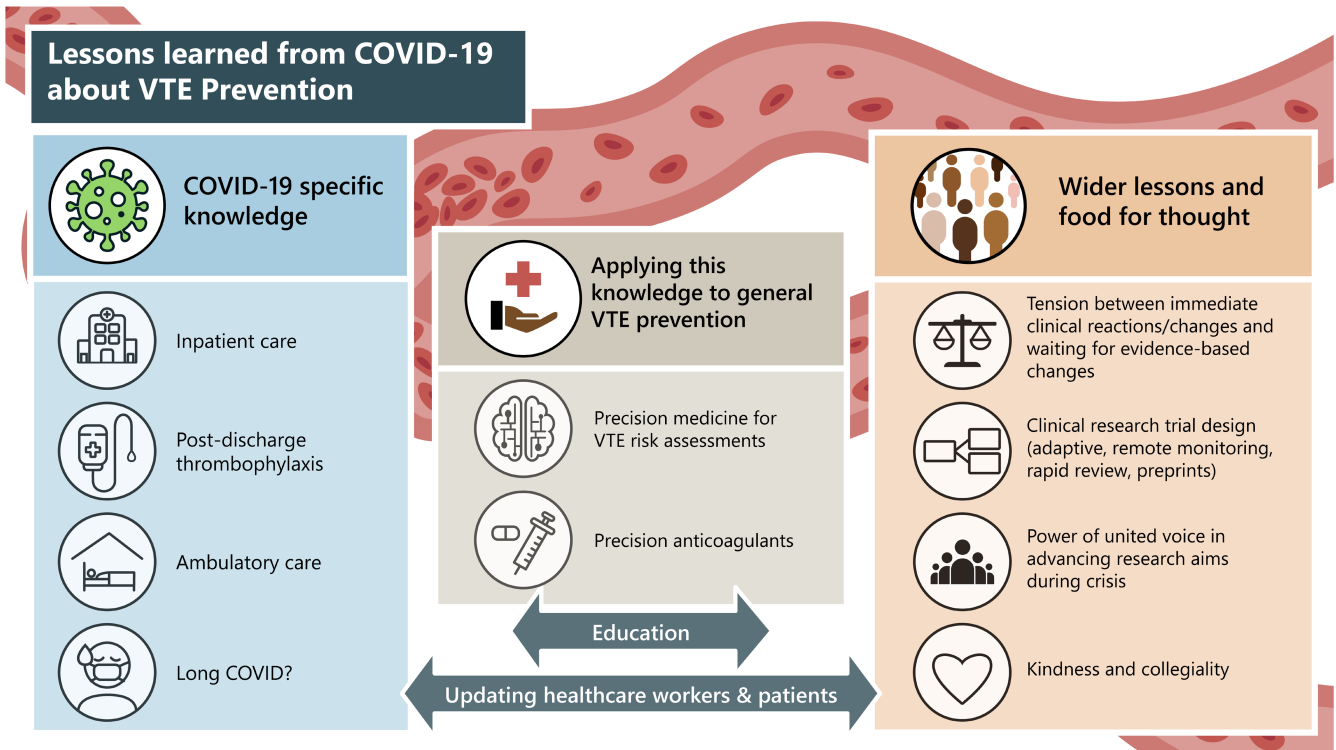
Finally, in silico PK/PD modelling can be essential in rare diseases and “special” populations such as extreme ages (elderly and children), extreme body sizes (underweight and high-degree obesity), severe comorbidities (severe kidney and liver dysfunction), which are either underrepresented or excluded from cardiovascular randomized trials.

A, acetylsalicylic acid; MK, megakaryocyte; od, once daily; P, placebo; TXB₂, thromboxane B₂.

For references, see Tosetto et al.⁴⁵; Rocca et al.⁴⁶; Petrucci et al.⁴⁷

VTE prevention—What have we learned from COVID-19?

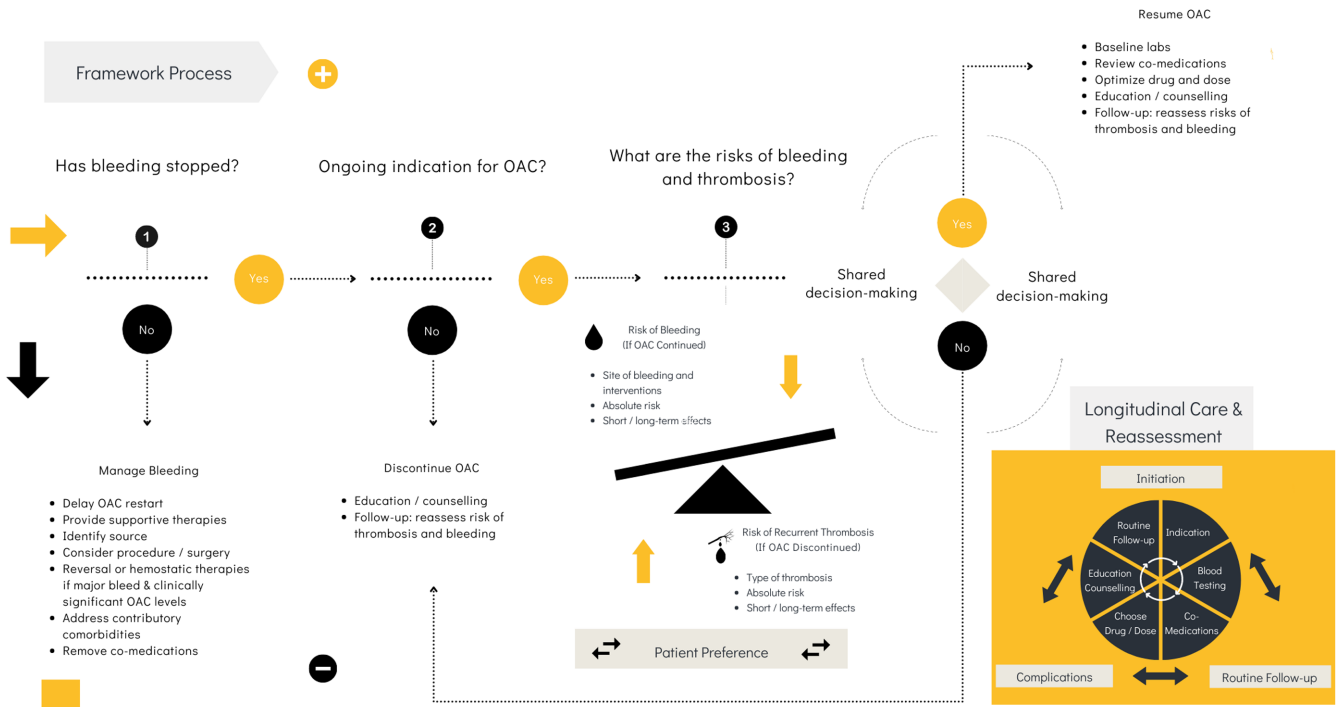
Susan Shapiro MD, PhD



Restarting anticoagulants after major bleeding

Deborah M. Siegal MD, MSc, FRCPC

Restarting Antithrombotics after Major Bleeding

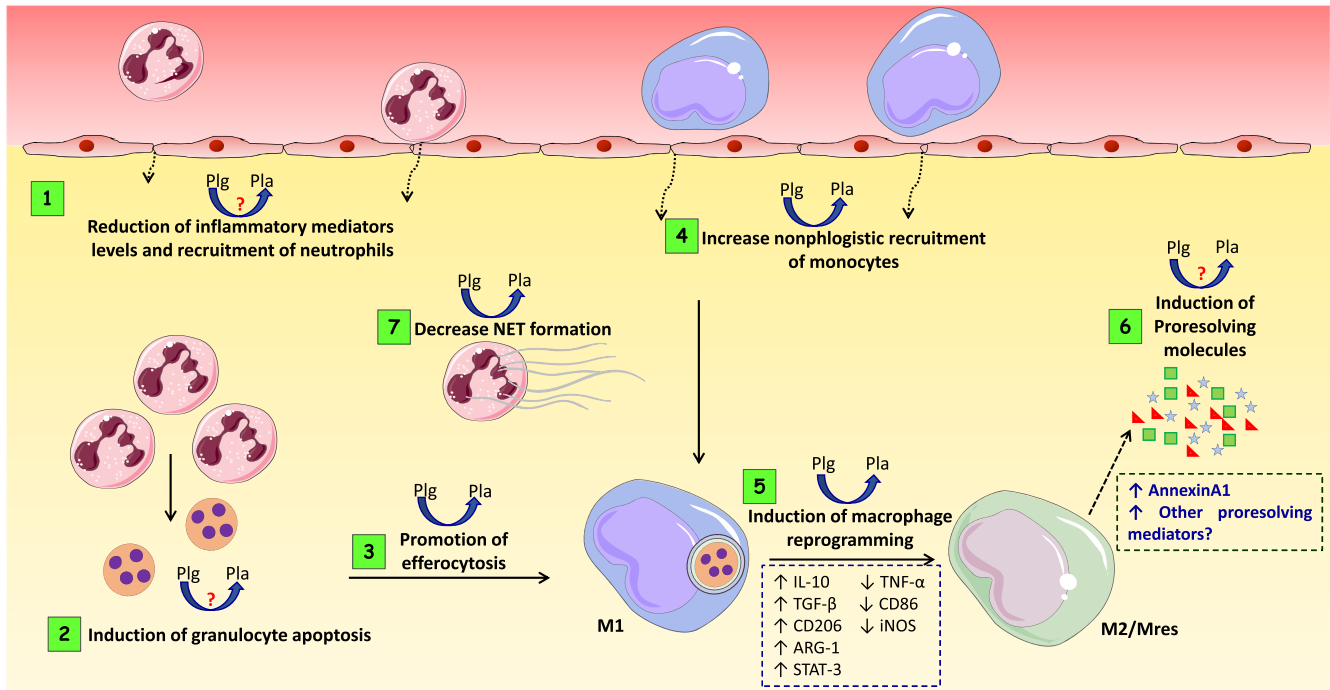


For reference, see Xu and Siegal⁴⁸

The plasminogen/plasmin system beyond fibrinolysis: Emerging players in resolution of inflammation

Lirlândia P. Sousa PhD

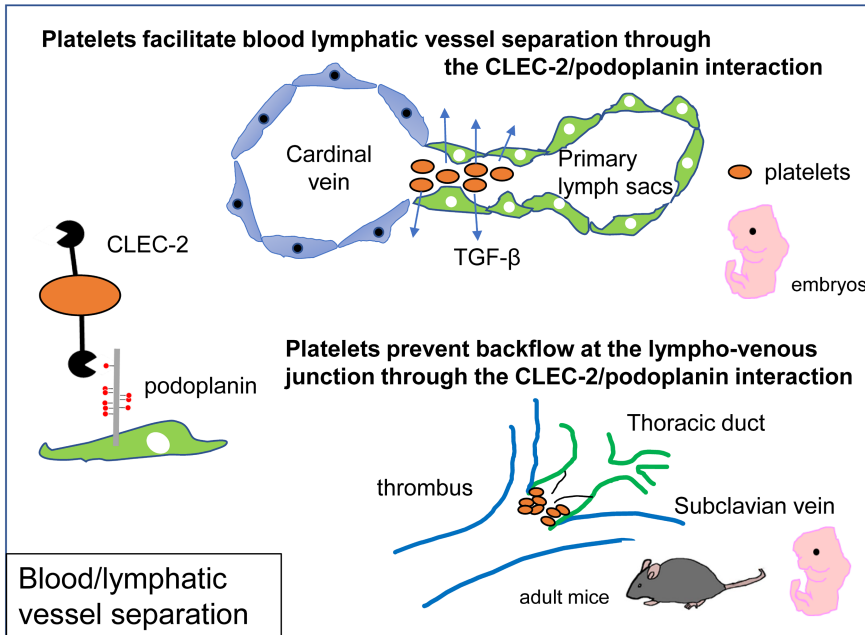
The Plasminogen/Plasmin system beyond fibrinolysis: Emerging players in resolution of inflammation



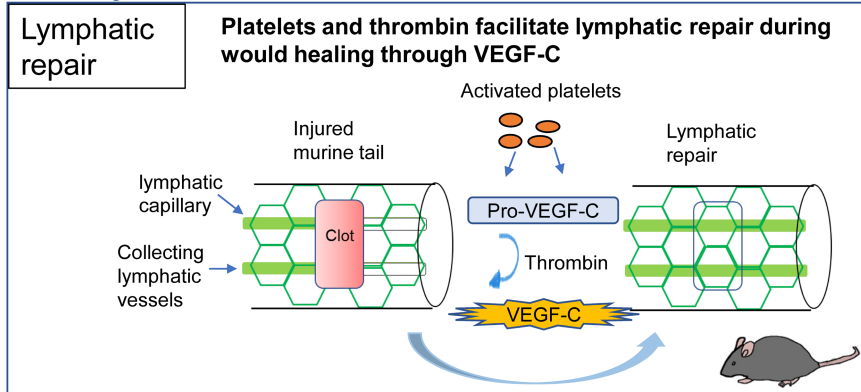
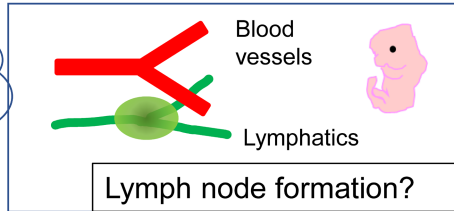
The plasminogen/plasmin (Plg/Pla) system is associated with a variety of biological activities beyond the classical dissolution of fibrin clots, including cell migration, tissue repair, and inflammation. Inflammation is an evolutionarily conserved response that guarantees the maintenance of tissue homeostasis through resolution—an active process mediated by molecules named proresolving mediators.⁴⁹ Despite the classical view of the Plg/Pla system on inflammation, emerging studies from our group^{50,51} and others have revealed its anti-inflammatory and proresolving actions that includes reduction of proinflammatory mediators (1) enhanced neutrophil apoptosis (2), and efferocytosis (3); promotion of nonphlogistic recruitment of mononuclear cells (4); induction of macrophage reprogramming toward resolving phenotypes (5); and induction of expression of the proresolving mediator annexin A1 (6). We have now identified a novel proresolving feature of Plg/Pla during sepsis that is the regulation of neutrophil extracellular trap release (7). Red question marks indicate specific processes that Pla-protease activity need to be covered.

Crosstalk between hemostasis and lymphangiogenesis

Katsue Suzuki-Inoue MD, PhD



**Crosstalk
between hemostasis
and lymphangiogenesis**



Raising awareness of women's bleeding/health issues in culturally challenging settings

Tahira Zafar MB, DCP, FRCPath

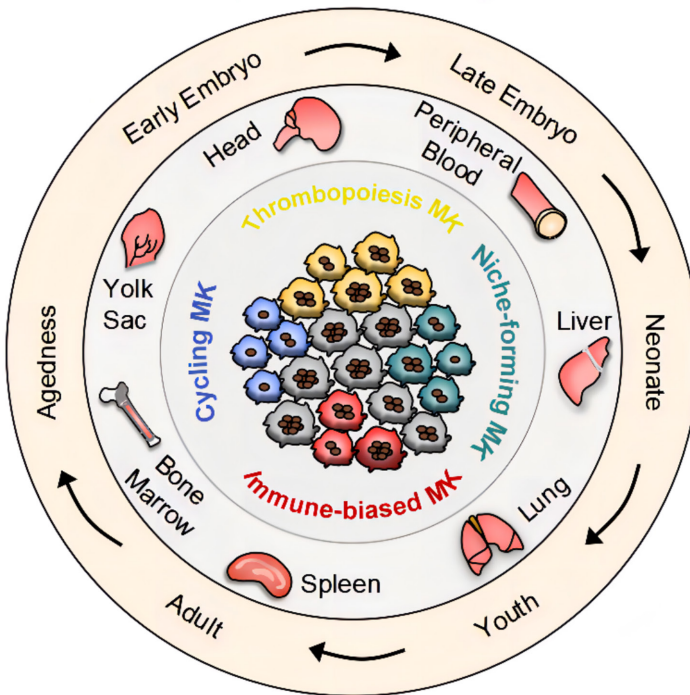


For references, see Ali⁵²; Butt⁵³; UN.⁵⁴

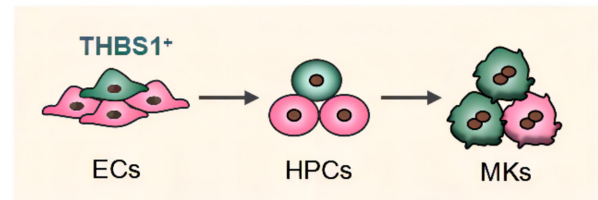
Megakaryocyte single-cell transcriptomics

Jiaxi Zhou PhD

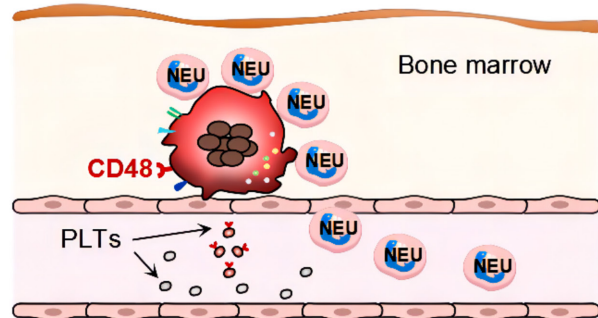
MKs are spatio-temporal heterogeneous.



THBS1⁺ ECs are biased toward megakaryopoiesis^[1], probably generating niche-forming MKs that prompt malignant HSC expansion.



CD48⁺ MKs function as immune-surveillance cells by neutrophil recruitment and platelet release^[2-3].



EC, endothelial cell; HPC, hematopoietic progenitor cell; HSC, hematopoietic stem cell; MK, megakaryocyte; NEU, neutrophil; PLT, platelet. For references, see Wang et al.⁵⁵; Liu et al.^{56,57}

RELATIONSHIP DISCLOSURE

JA has received grant funding from the British Heart Foundation and is a consultant for Silence Therapeutics; RAA is a consultant for Bristol Myers Squibb, Bayer, and Pfizer; AB has received support from Horizon 2020 FETOpen SilkFusion, NIH, and Novartis; CDB has received grant funding from NHLBI and NIH and is a consultant for Camurus AB; PB has received funding from BioMarin and is a consultant for BioMarin, Pfizer, and the Institute for Medical & Nursing Education; KdW has received grant funding from Pfizer; CD has received grant funding from Haima Therapeutics; MHE is a consultant for Novartis and Gad Medical; CPMH has received grant funding from the Canadian Institutes for Health Research and is a consultant for Werfen and Diagnostica Stago Inc; FAK has received research support from Bayer, Actelion, the Dutch Thrombosis Association, Actelion, BSCI, The Netherlands Organisation for Health Research and Development, and the Dutch Heart foundation; ZM has received grant funding from CSL Behring; MDN has received grant funding from Haemonetics, Instrumentation Laboratories, and Janssen Pharmaceuticals, is consultant for Haemonetics and Janssen Pharmaceuticals, and serves on the advisory board for Haima Therapeutics; BR has received grant funding from Bayer AG and is a consultant for SOBI and Bayer AG; SS is a consultant for Pfizer, Takeda, and Roche and received support for attending meetings from CSL Behring; DMS is a consultant for BMS-Pfizer, Leo Pharma, Portola Pharmaceuticals, and Servier; KS is a consultant for Novo Nordisk, Chugai Pharmaceutical Co, and Mitsubishi Tanabe Pharma Co; all other authors declare no conflicts of interest.

TWITTER

Robert A. Ariëns  @RobertAriens
 Beverley J. Hunt  @bhwords
 Ejaife O. Agbani  @DrEjaifeAgbani
 Josefin Ahnström  @AhnstromJosefin
 Robert Ahrends  @Ahrends15
 Raza Alikhan  @dvtfdc
 Alice Assinger  @PlateletVienna
 Zsuzsa Bagoly  @bagoly18
 Alessandra Balduini  @ABalduiniLab
 Christopher D. Barrett  @ChrisBarrettMD
 Paul Batty  @paul_batty1
 Wee Shian Chan  @WeeShianChan
 Kerstin Wit  @kerstindewit
 Cécile Denis  @cecile_denis
 Renee Eslick  @renee_eslick
 Hongxia Fu  @hongxiaUW
 Benoit Ho-Tin-Noé  @Team6LVTSINSERM
 Frederikus A. Klok  @Erik_Klok_MD
 Riten Kumar  @kumar_riten
 Karin Leiderman  @karin_leiderman
 Matthew D. Neal  @mackey_neal
 William A. E. Parker  @TheClotDoc
 Roger J. S. Preston  @rogerjpreston

Julie Rayes  @julierayes

Lara N. Roberts  @LaraNRoberts1

Susan Shapiro  @shapiro_susie

Deborah M. Siegal  @DebSiegal

REFERENCES

- Gierula M, Ahnström J. Anticoagulant protein S-New insights on interactions and functions. *J Thromb Haemost.* 2020;18:2801-2811.
- Teraz-Orosz A, Gierula M, Petri A, et al. Laminin G1 residues of protein S mediate its TFPI cofactor function and are competitively regulated by C4BP. *Blood Adv.* 2022;6(2):704-715.
- Dahlbäck B, Tran S. The preAR2 region (1458-1492) in factor V-Short is crucial for the synergistic TFPI α -cofactor activity with protein S and the assembly of a trimolecular factor Xa-inhibitory complex comprising FV-Short, protein S, and TFPI α . *J Thromb Haemost.* 2022;20(1):58-68.
- Peng B, Geue S, Coman C, et al. Identification of key lipids critical for platelet activation by comprehensive analysis of the platelet lipidome. *Blood.* 2018;132:e1-e12. doi:10.1182/blood-2017-12-822890
- Peng B, Kopczyński D, Pratt BS, et al. LipidCreator workbench to probe the lipidomic landscape. *Nat Commun.* 2020;11:2057. doi:10.1038/s41467-020-15960-z
- Holinstat M. Controlling the Clot: ANXA7 regulates collagen activation of platelet through 12-LOX. *Circ Res.* 2021;121:508-510. doi:10.1161/CIRCRESAHA.121.319736
- Gauberti M, Martinez de Lizarrondo S, Vivien D. Thrombolytic strategies for ischemic stroke in the thrombectomy era. *J Thromb Haemost.* 2021;19(7):1618-1628.
- Bagoly Z, Szegei I, Kalmadi R, Toth NK, Csiba L. Markers of coagulation and fibrinolysis predicting the outcome of acute ischemic stroke thrombolysis treatment: a review of the literature. *Front Neurol.* 2019;10:513.
- Donkel SJ, Benaddi B, Dippel DWJ, Ten Cate H, de Maat MPM. Prognostic hemostasis biomarkers in acute ischemic stroke. *Arterioscler Thromb Vasc Biol.* 2019;39(3):360-372.
- Di Buduo CA, Aguilar A, Soprano PM, et al. Latest culture techniques: cracking the secrets of bone marrow to mass-produce erythrocytes and platelets ex vivo. *Haematologica.* 2021;106(4):947-957. doi:10.3324/haematol.2020.262485
- Di Buduo CA, Wray LS, Tozzi L, et al. Programmable 3D silk bone marrow niche for platelet generation ex vivo and modeling of megakaryopoiesis pathologies. *Blood.* 2015;125(14):2254-2264. doi:10.1182/blood-2014-08-595561
- Di Buduo CA, Laurent PA, Zaninetti C, et al. Miniaturized 3D bone marrow tissue model to assess response to Thrombopoietin-receptor agonists in patients. *elife.* 2021;10:e58775. doi:10.7554/eLife.58775
- Arruda VR, Weber J, Samelson-Jones BJ. Gene therapy for inherited bleeding disorders. *Semin Thromb Hemost.* 2021;47(2):161-173. doi:10.1055/s-0041-1722862
- Rodríguez-Márquez E, Meumann N, Büning H. Adeno-associated virus (AAV) capsid engineering in liver-directed gene therapy. *Expert Opin Biol Ther.* 2021;21(6):749-766. doi:10.1080/14712598.2021.1865303
- Barbon E, Kaweck C, Marmier S, et al. Development of a dual hybrid AAV vector for endothelial-targeted expression of von Willebrand factor. *Gene Ther.* 2021. doi:10.1038/s41434-020-00218-6
- Barrett CD, Moore HB, Moore EE, Wang J, Hajizadeh N, Biffi WL, Lottenberg L, Patel PR, Truitt MS, McIntyre R, Talmor DS, Sauer A, Yaffe MB. Study of Alteplase for Respiratory failure in SARS-Cov2 COVID-19 (STARS): a vanguard multicenter, rapidly adaptive, pragmatic, randomized, controlled trial. *Chest.* 2022;161(3): 710-727. doi:10.1016/j.chest.2021.09.024

17. Barrett CD, Moore HB, Moore EE, et al. MULTicenter STudy of tissue plasminogen activator use in COVID-19 severe respiratory failure (MUST COVID): a retrospective cohort study. *Res Pract Thromb Haemost.* 2022;6(2): e12669. doi:10.1002/rth2.12669
18. ATTACC Investigators, ACTIV-4a Investigators, REMAP-CAP Investigators, et al. Therapeutic anticoagulation with heparin in noncritically ill patients with Covid-19. *N Engl J Med* 2021;385(9): 790-802.
19. Yuki K, Fujiori M, Koutsogiannaki S. COVID-19 pathophysiology: a review. *Clin Immunol.* 2020;215:108427.
20. Gustine JN, Jones D. Immunopathology of hyperinflammation in COVID-19. *Am J Pathol.* 2021;191:4-17.
21. Diamond MS, Kanneganti TD. Innate immunity: the first line of defense against SARS-Cov-2. *Nat Immunol.* 2022;23:165-176. doi:10.1038/s41590-021-01091-0
22. Konstantinides SV, Meyer G, Becattini C, et al. 2019 ESC Guidelines for the diagnosis and management of acute pulmonary embolism developed in collaboration with the European Respiratory Society (ERS): the task force for the diagnosis and management of acute pulmonary embolism of the European Society of Cardiology (ESC). *Eur Heart J.* 2020;41:543-603.
23. van der Pol LM, Tromeur C, Bistervels IM, et al. Pregnancy-adapted YEARS algorithm for diagnosis of suspected pulmonary embolism. *N Engl J Med.* 2019;380(12):1139-1149.
24. Righini M, Robert-Ebadi H, Elias A, et al. Diagnosis of pulmonary embolism during pregnancy: a multicenter prospective management outcome study. *Ann Intern Med.* 2018;169(11):766-773.
25. Alimam, S., Bewley, S., Chappell, L.C, Knight, M., Seed, P., Gray, G., Harrison, CN, Robinson, S.E. (2016) Pregnancy outcomes in myeloproliferative neoplasms: UK prospective cohort study. *Br J Haematol.* 175, 31-36.
26. Maze D, Kazi S, Gupta V, et al. Association of treatments for myeloproliferative neoplasms during pregnancy with birth rates and maternal outcomes: a systematic review and meta-analysis. *JAMA Netw Open.* 2019;2(10):e1912666.
27. Eslick R, Cutts B, Merriman E, et al. HOW Collaborative position paper on the management of thrombocytopenia in pregnancy. *Aust N Z J Obstet Gynaecol.* 2021;61:195-204.
28. Fu H, Jiang Y, Yang D, Scheiflinger F, Wong WP, Springer TA. Flow-induced elongation of von Willebrand factor precedes tension-dependent activation. *Nat Commun.* 2017;8:324. doi:10.1038/s41467-017-00230-2
29. Jiang Y, Fu H, Springer TA, Wong WP. Electrostatic steering enables flow-activated Von Willebrand factor to bind platelet glycoprotein, revealed by single-molecule stretching and imaging. *J Mol Biol.* 2019;431:1380-1396. doi:10.1016/j.jmb.2019.02.014
30. Goldenberg NA, Sochet A, Albisetti M, et al. Consensus-based clinical recommendations and research priorities for anticoagulant thromboprophylaxis in children hospitalized for COVID-19-related illness. *J Thromb Haemost.* 2020;18:3099-3105.
31. Henderson LA, Canna SW, Friedman KG, et al. American College of Rheumatology clinical guidance for multisystem inflammatory syndrome in children associated with SARS-CoV-2 and hyperinflammation in pediatric COVID-19: version 3. *Arthritis Rheumatol.* 2022;74:e1-e20.
32. Bansal N, Azeka E, Neunert C, et al. Multisystem inflammatory syndrome associated with COVID-19 anti-thrombosis guideline of care for children by action. *Pediatr Cardiol.* 2021;42(7):16235-11639.
33. Tutwiler V, Litvinov RI, Lozhkin AP, et al. Kinetics and mechanics of clot contraction are governed by molecular and cellular blood composition. *Blood.* 2016;127(1):149-159.
34. Peshkova AD, Malyasyov DV, Bredikhin RA, et al. Reduced contraction of blood clots in patients with venous thromboembolism is a possible thrombogenic and embologenic mechanism. *TH Open.* 2018;2(1):e104-e115.
35. Weisel JW, Litvinov RI. Visualizing thrombosis to improve thrombus resolution. *Res Pract Thromb Haemost.* 2021;5(1):38-50.
36. Hisada Y, Mackman N. Update from the laboratory: mechanistic studies of pathways of cancer-associated venous thrombosis using mouse models. *Hematology Am Soc Hematol Educ Program.* 2019;2019(1):182-186.
37. Tawil N, Bassawon R, Meehan B, et al. Glioblastoma cell populations with distinct oncogenic programs release podoplanin as procoagulant extracellular vesicles. *Blood Adv.* 2021;5(6):1682-1694.
38. Hisada Y, Garratt KB, Maqsood A, et al. Plasminogen activator inhibitor 1 and venous thrombosis in pancreatic cancer. *Blood Adv.* 2021;5(2):487-495.
39. Moore EE, Moore HB, Kornblith LZ, et al. Trauma-induced coagulopathy. *Nat Rev Dis Primers.* 2021;7(1):30. doi:10.1038/s41572-021-00264-3
40. Vulliamy P, Kornblith LZ, Kutcher ME, Cohen MJ, Brohi K, Neal MD. Alterations in platelet behavior after major trauma: adaptive or maladaptive? *Platelets.* 2021;32(3):295-304. doi:10.1080/09537104.2020.1718633
41. Plautz WE, Haldeman SH, Dyer MR, et al. Reduced cleavage of von willebrand factor by ADAMTS13 is associated with microangiopathic acute kidney injury following trauma. *Blood Coagul Fibrinolysis.* 2022;33(1):14-24. doi:10.1097/MBC.0000000000001089
42. Parker WAE, Gorog DA, Geisler T, et al. Prevention of stroke in patients with chronic coronary syndromes or peripheral arterial disease. *Eur Heart J Suppl.* 2020;22(Supplement_M):M26-M34.
43. Roberts LN, Durkin M, Arya R. Annotation: developing a national programme for VTE prevention. *Br J Haematol.* 2017;178(1):162-170.
44. NHS Digital. NHS Outcomes Framework Indicators dataset. 5.1 Deaths from venous thromboembolism (VTE) related events within 90days post hospital discharge. 2021. <https://digital.nhs.uk/data-and-information/publications/statistical/nhs-outcomes-framework/february-2021/domain-5-treating-and-caring-for-people-in-a-safe-environment-and-protecting-them-from-avoidable-harm/nof/5.1-deaths-from-venous-thromboembolism-vte-related-event-s-within-90-days-post-discharge-from-hospital>
45. Tosetto A, Rocca B, Petrucci G, et al. Association of platelet thromboxane inhibition by low-dose aspirin with platelet count and cytoreductive therapy in essential thrombocythemia. *Clin Pharm Ther.* 2022;111:939-949.
46. Rocca B, Tosetto A, Betti S, et al. A randomized double-blind trial of 3 aspirin regimens to optimize antiplatelet therapy in essential thrombocythemia. *Blood.* 2020;136(2):171-182.
47. Petrucci G, Zaccardi F, Giaretta A, et al. Obesity is associated with impaired responsiveness to once-daily low-dose aspirin and in vivo platelet activation. *J Thromb Haemost.* 2019;17(6):885-895.
48. Xu Y, Siegal DM. Anticoagulant-associated gastrointestinal bleeding: framework for decisions about whether, when and how to resume anticoagulants. *J Thromb Haemost.* 2021;19:2383-2393.
49. Sousa LP, Alessandri AL, Pinho V, Teixeira MM. Pharmacological strategies to resolve acute inflammation. *Curr Opin Pharmacol.* 2013;13(4):625-631.
50. Sugimoto MA, Ribeiro ALC, Costa BRC, et al. Plasmin and plasminogen induce macrophage reprogramming and regulate key steps of inflammation resolution via annexin A1. *Blood.* 2017;129(21):2896-2907.
51. Vago JP, Sugimoto MA, Lima KM, et al. Plasminogen and the plasminogen receptor, Plg-R_{KT} regulate macrophage phenotypic, and functional changes. *Front Immunol.* 2019;10:1458.
52. Original art by Rahimeen Qasim Ali, Grade 9 student, Presentation Convent School, Lal Kurti, Rawalpindi, Pakistan.
53. Butt MA. Women's health problems in Pakistan. *Middle East J Fam Med.* 2004;2(2).
54. UN report on Pakistani women health matters. <https://www.dawn.com/news/1389532>

55. Wang H, He J, Xu C, et al. Decoding human megakaryocyte development. *Cell Stem Cell*. 2021;28(3):535-549.
56. Liu C, Wu D, Xia M, et al. Characterization of cellular heterogeneity and an immune subpopulation of human megakaryocytes. *Adv Sci (Weinh)*. 2021;8(15):e2100921.
57. Liu C, Huang B, Wang H, Zhou J. The heterogeneity of megakaryocytes and platelets and implications for ex vivo platelet generation. *Stem Cells Transl Med*. 2021;10(12):1614-1620.

How to cite this article: Ariëns RA, Hunt BJ, Agbani EO, et al. Illustrated State-of-the-Art Capsules of the ISTH 2022 Congress. *Res Pract Thromb Haemost*. 2022;6:e12747. doi: [10.1002/rth2.12747](https://doi.org/10.1002/rth2.12747)