


Draft Genome Sequence of *Actinomyces odontolyticus* subsp. *actinosynbacter* Strain XH001, the Basibiont of an Oral TM7 Epibiont

Jeffrey S. McLean,^a Quanhui Liu,^a Batbileg Bor,^b Joseph K. Bedree,^b Lujia Cen,^b Michael Watling,^c  Thao T. To,^a Roger E. Bumgarner,^c Xuesong He,^b Wenyuan Shi^b

Department of Periodontics, University of Washington, Seattle, Washington, USA^a; School of Dentistry, University of California Los Angeles, Los Angeles, California, USA^b; Department of Microbiology, University of Washington, Seattle, Washington, USA^c

Here, we present the draft genome sequence of *Actinomyces odontolyticus* subsp. *actinosynbacter* strain XH001, isolated from the human oral cavity. Uniquely, it was discovered as a host bacterium to the ultrasmall epibiont TM7x, which is the first cultivated member of “*Candidatus* Saccharibacteria” (formerly candidate phylum TM7).

Received 7 December 2015 Accepted 9 December 2015 Published 4 February 2016

Citation McLean JS, Liu Q, Bor B, Bedree JK, Cen L, Watling M, To TT, Bumgarner RE, He X, Shi W. 2016. Draft genome sequence of *Actinomyces odontolyticus* subsp. *actinosynbacter* strain XH001, the basibiont of an oral TM7 epibiont. *Genome Announc* 4(1):e01685-15. doi:10.1128/genomeA.01685-15.

Copyright © 2016 McLean et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Jeffrey S. McLean, jsmclean@uw.edu.

Although *Actinomyces odontolyticus* is a commensal oral species, it is an opportunistic pathogen that has been linked to many diseases, most notably its association with actinomycosis, the formation of painful abscesses in the mouth, lungs, or gastrointestinal tract (1). Furthermore, oral *Actinomyces* spp. have been linked to childhood caries, periodontitis, and human oral carcinomas (2–4). A newly reported interaction between an obligate ultrasmall epibiont, TM7x (a recently cultivated member of “*Candidatus* Saccharibacteria,” formerly candidate phylum TM7), and its basibiont *A. odontolyticus* subsp. *actinosynbacter* strain XH001 (5) provides a great system to study parasitic epibiont symbiosis in the bacterial kingdom. The genome presented here from the isolated strain derived from human saliva (6, 7) will enable further research into this unique interaction.

A. odontolyticus subsp. *actinosynbacter* strain XH001 was cultured in brain heart infusion (BHI) medium and incubated at 37°C under microaerobic conditions until exponential phase was reached. Genomic DNA was extracted using the Epicentre MasterPure DNA purification kit. The complete genome sequence was determined via Illumina sequencing using paired-end 300-bp reads. All quality-trimmed reads were *de novo* assembled using SPAdes version 3.61 (8, 9).

The draft genome is 2,336,127 bp and assembled into 5 contigs, with an overall G+C content of 65.9%. Gene annotation using the Prokaryotic Genome Automatic Annotation Pipeline (PGAAP) provided by National Center for Biotechnology Information (NCBI) identified a total of 1,998 genes, consisting of 1,936 coding sequences, 49 tRNAs, and 6 rRNAs. A set of 49 single-copy genes was extracted and aligned to 100 sequenced genomes using the Department of Energy Systems Biology Knowledgebase (KBase; <http://kbase.us>). The resulting likelihood-based tree was built from the 49 concatenated genes and indicated relatedness to the sequenced species *A. odontolyticus* and *Actinomyces* sp. ICM39; however, the closest sequenced genome, *A. odontolyticus* F0309, produced an average nucleotide identity of only 93.7% (10). Using an additional method described for the delineation of species us-

ing 40 universal marker genes (11), XH001 was just above the cutoff for the *A. odontolyticus* species. These results support that this strain conservatively represents a subspecies of *A. odontolyticus*, and the name “*A. odontolyticus* subsp. *actinosynbacter* strain XH001” is proposed. The report by Bor et al. (5) presents further physiological characteristics of this strain.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. LLVT00000000. The version described in this paper is version LLVT00000000.1.

ACKNOWLEDGMENTS

We declare no conflicts of interest.

FUNDING INFORMATION

HHS | NIH | National Institute of General Medical Sciences (NIGMS) provided funding to Jeffrey S. McLean under grant number 1R01GM095373. HHS | NIH | National Institute of Dental and Craniofacial Research (NIDCR) provided funding to Jeffrey S. McLean under grant number 1R01DE023810.

REFERENCES

1. Aas JA, Paster BJ, Stokes LN, Olsen I, Dewhirst FE. 2005. Defining the normal bacterial flora of the oral cavity. *J Clin Microbiol* 43:5721–5732. <http://dx.doi.org/10.1128/JCM.43.11.5721-5732.2005>.
2. Colombo AV, Silva CM, Haffajee A, Colombo AP. 2006. Identification of oral bacteria associated with crevicular epithelial cells from chronic periodontitis lesions. *J Med Microbiol* 55:609–615. <http://dx.doi.org/10.1099/jmm.0.46417-0>.
3. Becker MR, Paster BJ, Leys EJ, Moeschberger ML, Kenyon SG, Galvin JL, Boches SK, Dewhirst FE, Griffen AL. 2002. Molecular analysis of bacterial species associated with childhood caries. *J Clin Microbiol* 40:1001–1009. <http://dx.doi.org/10.1128/JCM.40.3.1001-1009.2002>.
4. Nagy KN, Sonkodi I, Szöke I, Nagy E, Newman HN. 1998. The microflora associated with human oral carcinomas. *Oral Oncol* 34:304–308. [http://dx.doi.org/10.1016/S1368-8375\(98\)80012-2](http://dx.doi.org/10.1016/S1368-8375(98)80012-2).
5. Bor B, Poweleit N, Bois J, Cen L, Bedree J, Zhou ZH, Gunsalus R, Lux R, McLean J, He X, Shi W. 2015. Phenotypic and physiological charac-

- terization of the epibiotic interaction between TM7x and its basibiont *Actinomyces*. *Microb. Ecol.*
6. Edlund A, Yang Y, Hall AP, Guo L, Lux R, He X, Nelson KE, Neelson KH, Yooseph S, Shi W, McLean JS. 2013. An *in vitro* biofilm model system maintaining a highly reproducible species and metabolic diversity approaching that of the human oral microbiome. *Microbiome* 1:25. <http://dx.doi.org/10.1186/2049-2618-1-25>.
 7. He X, McLean JS, Edlund A, Yooseph S, Hall AP, Liu SY, Dorrestein PC, Esquenazi E, Hunter RC, Cheng G, Nelson KE, Lux R, Shi W. 2015. Cultivation of a human-associated TM7 phylotype reveals a reduced genome and epibiotic parasitic lifestyle. *Proc Natl Acad Sci USA* 112: 244–249. <http://dx.doi.org/10.1073/pnas.1419038112>.
 8. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <http://dx.doi.org/10.1089/cmb.2012.0021>.
 9. Nurk S, Bankevich A, Antipov D, Gurevich AA, Korobeynikov A, Lapidus A, Prjibelski AD, Pyshkin A, Sirotkin A, Sirotkin Y, Stepanauskas R, Clingenpeel SR, Woyke T, McLean JS, Lasken R, Tesler G, Alekseyev MA, Pevzner PA. 2013. Assembling single-cell genomes and mini-metagenomes from chimeric MDA products. *J Comput Biol* 20: 714–737. <http://dx.doi.org/10.1089/cmb.2013.0084>.
 10. Varghese NJ, Mukherjee S, Ivanova N, Konstantinidis KT, Mavrommatis K, Kyrpides NC, Pati A. 2015. Microbial species delineation using whole genome sequences. *Nucleic Acids Res* 43:6761–6771. <http://dx.doi.org/10.1093/nar/gkv657>.
 11. Mende DR, Sunagawa S, Zeller G, Bork P. 2013. Accurate and universal delineation of prokaryotic species. *Nat Methods* 10:881–884. <http://dx.doi.org/10.1038/nmeth.2575>.