



OPEN

## Paternal exposure to a common pharmaceutical (Ritalin) has transgenerational effects on the behaviour of Trinidadian guppies

Alex R. De Serrano<sup>1</sup>✉, Kimberly A. Hughes<sup>2</sup> & F. Helen Rodd<sup>1</sup>

Evidence is emerging that paternal effects, the nongenetic influence of fathers on their offspring, can be transgenerational, spanning several generations. Methylphenidate hydrochloride (MPH; e.g. Ritalin) is a dopaminergic drug that is highly prescribed to adolescent males for the treatment of Attention-deficit/hyperactivity disorder. It has been suggested that MPH could cause transgenerational effects because MPH can affect the male germline in rodents and because paternal effects have been observed in individuals taking similar drugs (e.g. cocaine). Despite these concerns, the transgenerational effects of paternal MPH exposure are unknown. Therefore, we exposed male and female Trinidadian guppies (*Poecilia reticulata*) to a low, chronic dose of MPH and observed that MPH affected the anxiety/exploratory behaviour of males, but not females. Because of this male-specific effect, we investigated the transgenerational effects of MPH through the paternal line. We observed behavioural effects of paternal MPH exposure on offspring and great-grandoffspring that were not directly administered the drug, making this the first study to demonstrate that paternal MPH exposure can affect descendants. These effects were not due to differential mortality or fecundity between control and MPH lines. These results highlight the transgenerational potential of MPH.

The parental environment can have a strong influence on offspring development and subsequent phenotype<sup>1</sup>. Evidence is now emerging that these parental effects can be transgenerational, i.e. spanning several generations; for example, stressful situations (e.g. early life stress)<sup>2,3</sup>, and exposure to some drugs and pollutants can have transgenerational effects on multiple traits including physiology and behaviour, e.g.<sup>4–13</sup>. If these effects are common, they would have important implications for understanding how natural populations respond to changing environments, including anthropogenic effects (e.g. climate change, exposure to pollutants) and how the experiences of our ancestors can affect our health and susceptibility to disease<sup>14</sup>. However, it is not clear if these results are generalizable beyond the few taxa where these effects have been investigated. In this study, we examine the transgenerational effects of a common pharmaceutical on natural behaviours of Trinidadian guppies, a freshwater, tropical fish.

Parental effects can be transmitted by the mother and/or father. It is well established that the maternal environment can affect offspring phenotype in both adaptive<sup>1</sup> and nonadaptive ways<sup>15</sup>. In contrast, paternal effects, the influence of fathers on their offspring by nongenetic means, have received much less attention because, unlike maternal effects, there was no clear mechanism for their effects, except in species where males provide parental care or resources to females<sup>16</sup>. Recent evidence suggests that nongenetic factors, such as compounds in ejaculate and epigenetic/cytoplasmic modifications to sperm, can affect offspring phenotype<sup>16,17</sup>. Fathers can have additional effects on their offspring through their interactions with females, for example, when females differentially allocate resources to offspring based on the quality of their mate<sup>16–18</sup>.

Evidence is accumulating that parental effects can extend beyond offspring, influencing subsequent generations. Maternal effects can span multiple generations, affecting progeny that were not directly exposed to the original stimulus: ‘grandmaternal effects’ in externally fertilizing species<sup>19–21</sup>, and ‘great-grandmaternal effects’

<sup>1</sup>Department of Ecology and Evolutionary Biology, University of Toronto, 25 Willcocks St, Toronto, ON M5S 3B2, Canada. <sup>2</sup>Department of Biological Science, Florida State University, 319 Stadium Dr, Tallahassee, FL 32304, USA. ✉email: a.deserrano@utoronto.ca

in internally fertilizing species<sup>22–27</sup>. There is some evidence that paternal effects can extend beyond the offspring generation. These studies have demonstrated that stressors, including fear conditioning<sup>28</sup>, maternal separation<sup>3</sup>, restraint<sup>29</sup>, obesity<sup>30–32</sup>, drug exposure<sup>4</sup>, and exposure to pollutants<sup>5,6</sup> are associated with altered phenotypes that span multiple generations through the paternal line. However, the conclusions that can be drawn from these studies may be limited for several reasons. First, many of these studies do not report potentially confounding effects of differences in survival or reproductive success between control and treatment lines. Second, some studies only investigate the effects in progeny of one sex, despite evidence that parental effects are sometimes sex-specific<sup>33–37</sup>. Third, most examples come from rodents (but see<sup>5,14,27,37</sup>), which makes it unclear if these results can be generalized to other taxa.

For our study of transgenerational effects, we manipulated the dopaminergic pathway with a pharmaceutical for several reasons. Dopamine is a neurotransmitter that is conserved across vertebrates<sup>38</sup> and is involved in a variety of behaviours that are ecologically relevant in natural populations, such as sexual motivation<sup>39,40</sup> and novelty-seeking behaviour<sup>41,42</sup>. Novelty-seeking behaviour can have fitness consequences for individuals in natural populations, including increased access to novel food sources, mating opportunities, and suitable habitats<sup>41,43</sup>, but at the cost of increased risk of predation<sup>43</sup>. We also focused on dopamine because it has been postulated that some drugs that affect the dopaminergic system will cause transgenerational effects<sup>44</sup> and some of those drugs are widely used in humans.

In this study, we manipulated Methylphenidate hydrochloride (MPH), commercially known as Ritalin (Novartis) or Concerta (Janssen Pharmaceuticals Inc.). It is a stimulant that affects the dopaminergic pathway and was predicted to have transgenerational effects<sup>44</sup>. MPH provides therapeutic benefits by binding dopamine transporters on the plasma membrane of presynaptic neurons; this blocks the reuptake of this neurotransmitter (as well as norepinephrine), resulting in an increase in synaptic dopamine<sup>45,46</sup>. MPH is widely prescribed to individuals with Attention-deficit/hyperactivity disorder (ADHD<sup>47–50</sup>); this disorder is characterized by imbalances in the dopaminergic and noradrenergic systems of the brain, which results in reduced focus and increased impulsivity/over-activity. Further, ADHD is often comorbid with other psychological disorders and/or drug abuse and, therefore, can have detrimental effects on quality of life if left untreated<sup>51</sup>.

Despite its widespread use, the transgenerational effects of MPH are relatively unexplored. Studies in lab rodents and zebrafish have shown that maternal exposure to MPH can affect the neurochemistry and behaviour of offspring<sup>52–55</sup>. Further, it has been suggested that MPH can affect the male germline, as male rodents exposed to MPH through adolescence and into early adulthood (postnatal days 21 to 60) exhibited altered sperm morphology and germ cell epithelium structure<sup>56</sup>. However, the effects of paternal MPH on offspring and later generations in any species are currently unknown, and we are not aware of any transgenerational epidemiological studies on the effects of MPH in humans. This is surprising, in light of the high prescription rates to adolescent males (> 7%<sup>49</sup>), and the fact that similar drugs (e.g. cocaine) can cause paternal effects<sup>57–61</sup>. Specifically, MPH shares some neurochemical properties with cocaine and, in rodents, repeated exposure to either MPH or cocaine can lead to an increased prevalence of stereotypical behaviours, drug-seeking, and affective disorders, although some of these effects are only observed at high doses of MPH<sup>62</sup>.

In this study, we investigated if MPH causes transgenerational effects in Trinidadian guppies, *Poecilia reticulata*. Guppies and other fish, in general, are increasingly being used for neurological research because of their similarity to mammals in patterns of neurodevelopment, functional brain organization, and neurocircuitry, including the dopaminergic system<sup>63,64</sup>. In addition, female guppies carry eggs internally for approximately 25 days<sup>65</sup> with small amounts of maternal transfer<sup>66</sup>. Further, novelty-seeking behaviour is ecologically relevant for wild guppies: guppies are highly attracted to novel stimuli<sup>67–70</sup>, and exploration and subsequent dispersal can increase mating success in both sexes<sup>71,72</sup>. Finally, previous studies have demonstrated that responses by guppies to MPH are similar to those observed in mammals<sup>73</sup>.

For this study, we administered MPH to the first-generation (G<sub>1</sub>) male and female guppies and asked if there were effects on their behaviour, and on the behaviour and whole brain dopamine levels of three subsequent generations that were not administered the drug directly. Because chronic treatment with therapeutic levels of MPH reduced rodent exploration<sup>74</sup> and increased anxiety<sup>74,75</sup>, we predicted that guppies chronically treated with MPH would reduce their exploration of a novel environment. Indeed, we did observe a significant effect of chronic MPH on the exploratory behaviour of first generation (G<sub>1</sub>) males, but not females, so we subsequently focused on the transgenerational paternal effects of MPH exposure. Because paternal exposure to cocaine increased anxiety in rodent offspring<sup>60</sup>, we also predicted that offspring of MPH treated male guppies would exhibit increased anxiety, and therefore reduced exploration, as adults. However, we were unable to predict the direction of the behavioural responses of later generations (i.e. grandoffspring, great-grandoffspring) as there is not a consistent pattern in the literature about the direction or strength of induced phenotypes transmitted across multiple generations<sup>76</sup>. Finally, for transgenerational effects on brain dopamine levels, Lepetiller et al.<sup>55</sup> observed that semi-chronic, prenatal MPH exposure increased dopamine levels in adult male rodents, so we predicted that progeny would also have increased dopamine levels relative to controls.

## Results

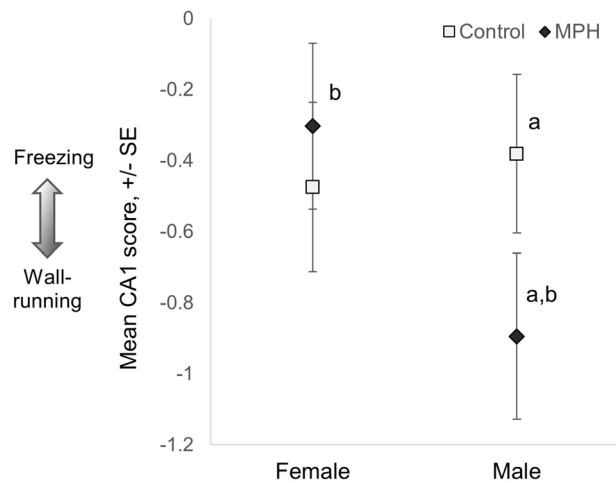
**Effects of chronic MPH administration.** For the first step of this project, first generation (G<sub>1</sub>) juvenile guppies assigned to the MPH treatment began receiving a low, chronic dose of MPH ( $2.5 \times 10^{-8}$  g/mL) from one month of age until testing, while Control individuals were treated in the same manner, except that they were only administered the vehicle. To mimic the therapeutic use of this drug in humans<sup>47</sup>, we treated guppies chronically by administering MPH three days/week to ensure that levels remained relatively stable in the aquarium water. Adult guppies were tested in the open field test, a standardized test that is widely used to measure locomotion, anxiety, and exploratory behaviour<sup>77</sup>. On average, we tested seven G<sub>1</sub> individuals per brood (mean: 6.8, min: 1,

| Generation                                  | Original model  | Response                      | Final model  | Estimate   | DF <sub>num</sub> | DF <sub>den</sub> | F value     | P value      |
|---|---|-------------------------------|--|--|-------------------|-------------------|-------------|--------------|
| G <sub>1</sub>                              | Treatment * sex * age, date tested, handling, time, drug administration | CA1                           | Intercept  | -0.892   |                   | 74.1              |             | 0.0003       |
|   |   |                               | Treatment  | 0.5134   | 1                 | 166               | 1.79        | 0.18         |
|   |   |                               | Sex  | 0.5911   | 1                 | 170               | 3.68        | 0.057        |
|   |   |                               | <b>Treatment * sex</b>   | <b>-0.6847</b>   | <b>1</b>          | <b>175</b>        | <b>6.62</b> | <b>0.011</b> |
|   |   |                               | Date   | 0.00219  | 1                 | 178               | 2.9         | 0.09         |
|   |   | CA2                           | Intercept  | 0.08642  |                   | 39.5              |             | 0.57         |
|   |   |                               | Treatment  | -0.00959   | 1                 | 168               | 0.01        | 0.94         |
|   |   |                               | Sex  | 0.252  | 1                 | 172               | 3.63        | 0.059        |
|   |   | G <sub>2</sub>                | G <sub>1</sub> male treatment * sex * age, brood size, date tested, days until isolation, handling, time | CA1  | Intercept         | -0.4604           |             | 45.3         |
| G <sub>1</sub> male treatment               | -0.0057   |                               |  |  | 1                 | 35.1              | 0.001       | 0.97         |
| Age   | -0.00346  |                               |  |  | 1                 | 272               | 5.37        | 0.02         |
| <b>G<sub>1</sub> male treatment * age</b>   | <b>-0.00307</b>   |                               |  |  | <b>1</b>          | <b>230</b>        | <b>5.11</b> | <b>0.025</b> |
| Sex   | 0.3318  |                               |  |  | 1                 | 277               | 6.99        | 0.009        |
| Sex * age                                   | 0.00599   |                               |  |  | 1                 | 278               | 14.57       | 0.0002       |
| Time  | 0.00141   |                               |  |  | 1                 | 274               | 2.96        | 0.087        |
| CA2   | Intercept   |                               |  | -0.06745   |                   | 70.7              |             | 0.6          |
|   | <b>G<sub>1</sub> male treatment</b>                                     |                               |  | <b>0.266</b>   | <b>1</b>          | <b>61</b>         | <b>3.96</b> | <b>0.051</b> |
|   | Age   |                               |  | 0.00003  | 1                 | 162               | 1.92        | 0.17         |
|   | Sex   |                               |  | 0.07042  | 1                 | 278               | 0.28        | 0.59         |
|   | Sex * age   |                               |  | -0.00431   | 1                 | 276               | 6.83        | 0.01         |
|   | Date  |                               |  | 0.00421  | 1                 | 151               | 9.26        | 0.003        |
|   | G <sub>3</sub>  |                               |  | G <sub>1</sub> male treatment * sex * age, brood size, date tested, days until isolation, handling, time | CA1               | Intercept         | -0.3052     |              |
| G <sub>1</sub> male treatment               |   | 0.07873                       | 1  |  |                   | 42.3              | 0.28        | 0.6          |
| Sex   |   | -0.08446                      | 1  |  |                   | 208               | 0.33        | 0.57         |
| Days until isolation                        |   | 0.00135                       | 1  |  |                   | 188               | 3.69        | 0.056        |
| Handling                                    |   | 0.08132                       | 1  |  |                   | 205               | 11.23       | 0.001        |
| CA2   |   | Intercept                     | -0.6538  |  |                   | 73.4              |             | 0.0003       |
|   |   | G <sub>1</sub> male treatment | 0.1478   |  | 1                 | 75.4              | 0.73        | 0.4          |
|   |   | Sex                           | 0.6919   |  | 1                 | 201               | 17.96       | <0.0001      |
|   |   | Age                           | -0.00552   |  | 1                 | 203               | 10.27       | 0.002        |
|   |   | Sex * age                     | 0.00415  |  | 1                 | 203               | 4.3         | 0.04         |
|   |   | G <sub>4</sub>                | G <sub>1</sub> male treatment * sex * age, brood size, date tested, days until isolation, handling, time |  | CA1               | Intercept         | -0.442      |              |
| G <sub>1</sub> male treatment               | 0.1786  |                               |  | 1  |                   | 27.3              | 0.37        | 0.55         |
| Sex   | 0.4158  |                               |  | 1  |                   | 155               | 4.17        | 0.043        |
| G <sub>1</sub> male treatment * sex         | -0.1401   |                               |  | 1  |                   | 141               | 0.2         | 0.65         |
| Age   | -0.00514  |                               |  | 1  |                   | 58.8              | 0.04        | 0.85         |
| G <sub>1</sub> male treatment * age         | 0.01121   |                               |  | 1  |                   | 92.9              | 0.39        | 0.53         |
| Sex * age                                   | 0.00859   |                               |  | 1  |                   | 157               | 0.001       | 0.99         |
| <b>G<sub>1</sub> male treatment*sex*age</b> | <b>-0.01723</b>   |                               |  | <b>1</b>   |                   | <b>158</b>        | <b>4.74</b> | <b>0.031</b> |
| Date  | -0.00603  |                               |  | 1  | 83.1              | 4.33              | 0.04        |              |
| CA2   | Intercept   |                               |  | -0.2083  |                   | 30.8              |             | 0.02         |
|   | <b>G<sub>1</sub> male treatment</b>                                     |                               |  | <b>0.3839</b>  | <b>1</b>          | <b>49.7</b>       | <b>5.55</b> | <b>0.022</b> |
|   | Sex   |                               |  | 0.09821  | 1                 | 163               | 0.43        | 0.51         |

**Table 1.** Final results of mixed model analyses of G<sub>1</sub>–G<sub>4</sub> open field trials for CA1 and CA2. \*Bolted text indicates that the model term ‘G<sub>1</sub> male treatment’ or interactions involving this term are statistically significant at  $P < 0.05$ .

max: 12) from one to three broods per pair (mean: 1.6), with half belonging to the MPH treatment group and the other half belonging to the Control group. We summarized recorded behaviours (freezing, ‘cautious swimming’ (i.e. slow swimming without movement of caudal fin), ‘wall-running’ (i.e. thigmotaxis), swimming, and duration located in the central area (Table S1)) with a Correspondence Analysis (CA): an ordination method conceptually similar to PCA<sup>78</sup>. We analyzed the first two components of CA (CA1 and CA2) using linear mixed effects models.

For CA1, cautious behaviours (freezing and cautious swimming) loaded in the opposite direction of ‘wall-running’ (CA1 loadings: freezing: 1.57, cautious swimming: 1.26, inner squares: 0.75, swimming: -0.19, wall-running: -1.68; biplot in Fig. S1). As predicted, G<sub>1</sub> male guppies chronically treated with MPH exhibited increased ‘wall-running’ (Table 1; Fig. 1;  $n = 188$ ; Table S2), which we interpret as anxiety/avoidance in response to a novel



**Figure 1.** First generation ( $G_1$ ) CA1 scores for the open field test. MPH treated males performed significantly more wall-running than Control males and MPH treated females, and there was a trend for them to perform more wall-running than Control females. Positive CA1 values indicate relatively more freezing and negative values indicate relatively more wall-running. Symbols represent least-square means for each response variable, +/- one standard error. Letters indicate points that are significantly different from one another at  $P < 0.05$ .

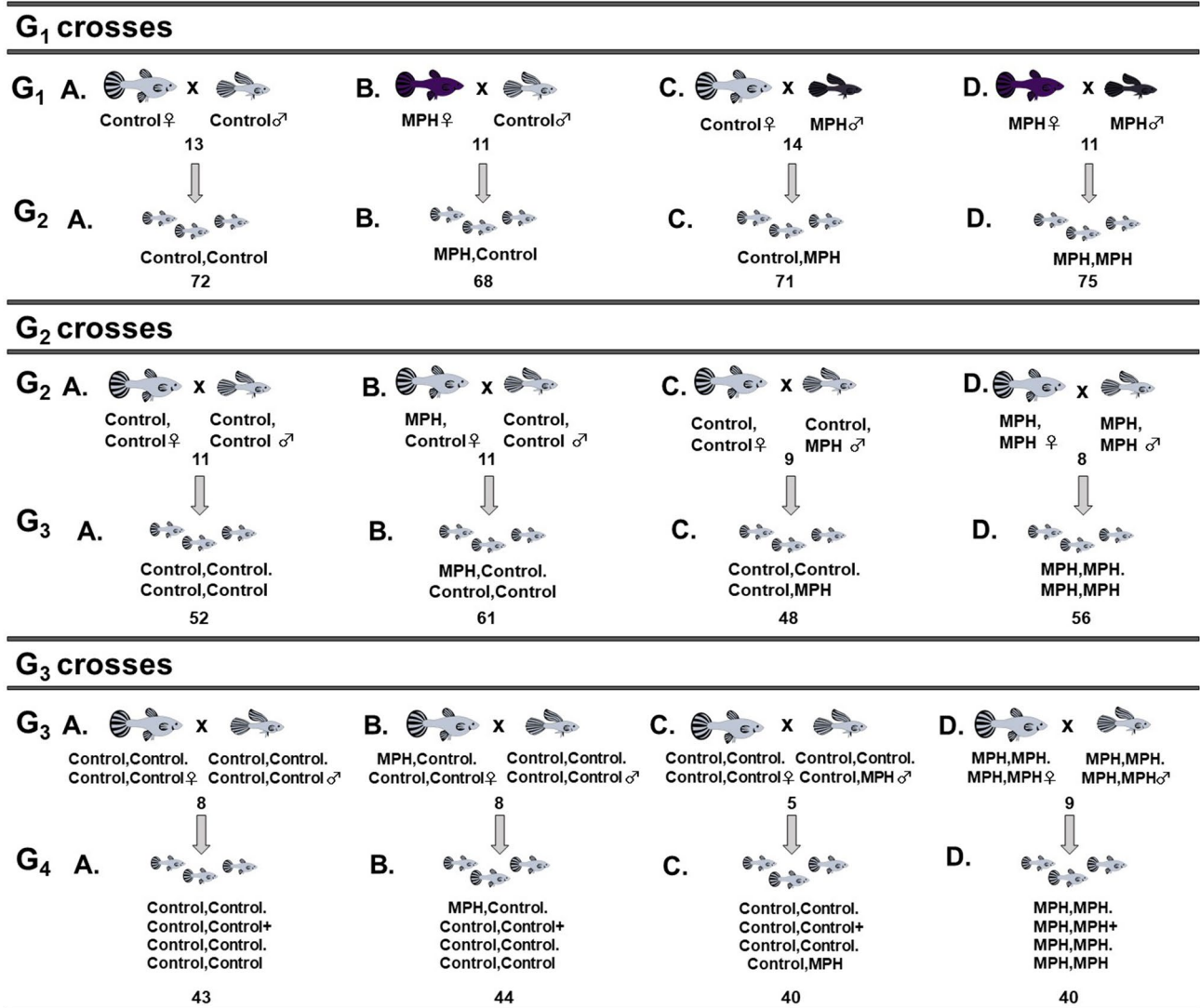
environment<sup>73</sup>. In contrast, Control males froze more; freezing is involved in predator evasion<sup>79</sup> (Table 1; Fig. 1). This suggests that MPH treatment disrupted typical anti-predator responses to a novel environment<sup>53,80,81</sup>. However, we did not observe a significant effect of MPH treatment on female behaviour (post hoc tests in Table S3). There was not a significant effect of MPH treatment on the behavioural variation summarized by CA2 (Table 1).

**Paternal effects of MPH.** To investigate the transgenerational effects of MPH treatment, first-generation fish were mated using a factorial design to produce four offspring ( $G_2$ ) treatment groups, which were maintained to produce the third ( $G_3$ ) and fourth ( $G_4$ ) generations (Fig. 2). The  $G_2$ – $G_4$  cohorts were not administered MPH or vehicle. For  $G_2$ – $G_4$  cohorts, we tested an average of three offspring per brood ( $G_2$  mean: 3.3, min: 1, max: 11;  $G_3$  mean: 2.7, min: 1, max: 8;  $G_4$  mean: 2.7, min: 1, max: 8) from one to five broods per pair ( $G_2$  mean: 1.8, min: 1, max: 4;  $G_3$  mean: 2, min: 1, max: 5;  $G_4$  mean: 2.1, min: 1, max: 5).

There was not a significant effect of  $G_1$  female treatment on the behaviour of any progeny generation ( $G_2$ – $G_4$ ), so we focus on the effects of  $G_1$  male treatment. Note that analyses that retained the distinction between  $G_1$  female and  $G_1$  male treatment groups produced very similar results to the results presented here (see Table S4). Offspring of MPH treated fathers and Control fathers (hereafter called ‘MPH treated’ and ‘Control’ offspring for brevity) differed in the behaviours summarized by both CA1 and CA2. Similar to their parents, for CA1, there were sex-specific effects of MPH but, this time, female offspring differed based on their fathers’ treatment status (Table 1; Fig. 3A;  $n = 286$ ). Female behaviour depended on their fathers’ treatment and the age at which they were tested: the behaviour of ‘Control’ females did not change significantly with age, but ‘MPH treated’ female offspring that were tested at a relatively old age froze more than younger individuals (slopes in Table S5). In contrast, ‘MPH treated’ and ‘Control’ males did not differ in how their behaviour changed with age.

For CA2, cautious behaviours loaded in the opposite direction of swimming and duration in the center of the arena (representing exploration or boldness<sup>82</sup>), so this axis represents a continuum between shyness/avoidance and boldness/exploration in response to a novel environment (CA2 loadings: freezing: 2.07, wall-running: 1.14, cautious swimming: 0.71, inner squares: -0.57, swimming: -0.77). As predicted, ‘MPH treated’ offspring again differed from ‘Control’ offspring (Table 1; Fig. 4A;  $n = 286$ ), with ‘Control’ offspring freezing more than ‘MPH treated’ offspring. This suggests that ‘MPH treated’ offspring were less cautious when investigating a novel environment. There was not a significant difference between male and female offspring. Taken together, results for CA1 and CA2 suggest that  $G_1$  male MPH treatment affected offspring response to a novel environment, with a stronger effect on the anxiety/avoidance behaviours of their daughters than their sons.

**Transgenerational effects of MPH.** To determine if these effects were transgenerational, we also tested the behaviour of ‘Control’ and ‘MPH treated’ grandoffspring ( $G_3$ ) and great-grandoffspring ( $G_4$ ). There was not a significant effect of  $G_1$  male treatment on grandoffspring ( $G_3$ ) behaviour (Table 1;  $n = 213$ ). Due to logistical constraints, the median ages for  $G_3$  males and females were higher than for the other generations and this may have influenced the results for this generation (Female median age (days):  $G_1 = 132$ ,  $G_2 = 267$ ,  $G_3 = 394$ ,  $G_4 = 269$ ; Male median age:  $G_1 = 117$ ,  $G_2 = 231$ ,  $G_3 = 300$ ,  $G_4 = 248$ ; Table S6 Figs. S2–S3). However, for great-grandoffspring ( $G_4$ ) behaviour, similar to the pattern we observed for the  $G_2$  (offspring), there were significant effects of  $G_1$  male treatment on both CA1 and CA2 (Table 1;  $n = 167$ ; CA1: Fig. 3B; CA2: Fig. 4B). All great-grandoffspring treatment groups exhibited behavioural patterns similar to their  $G_2$  ancestors with one exception: the association between age and CA1 score appeared to differ for  $G_2$  and  $G_4$  ‘Control’ males (lower left panels, Fig. 3A,B). To

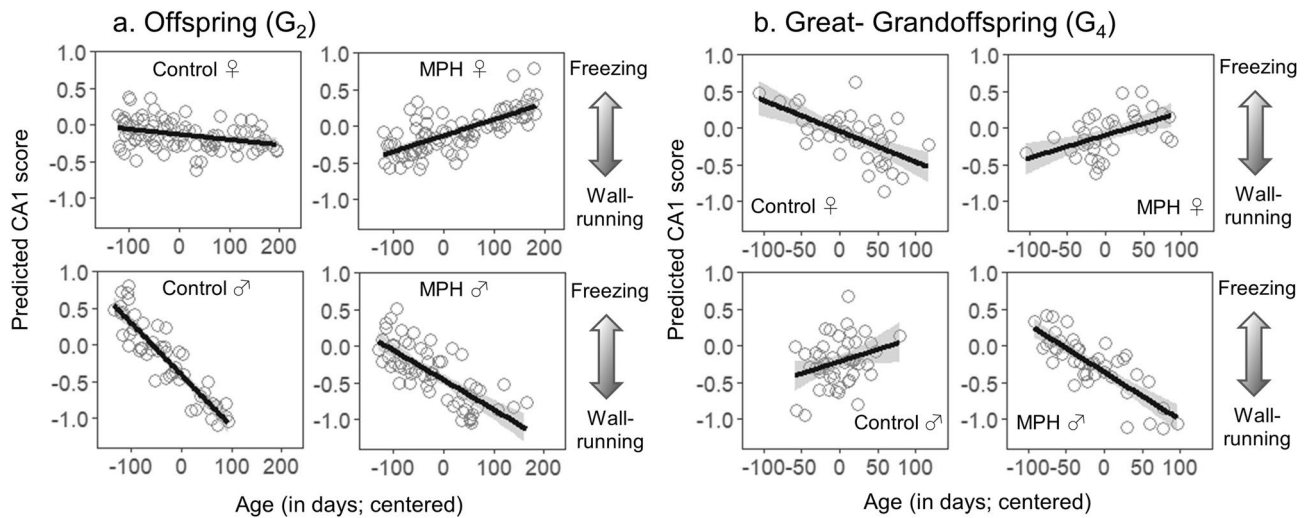


**Figure 2.** Crossing design for the G<sub>2</sub>-G<sub>4</sub> generations. First-generation fish were mated using a factorial design to produce four offspring (G<sub>2</sub>) treatment groups: (A) Control female × Control male; (B) MPH treated female × Control male; (C) Control female × MPH treated male; and (D) MPH treated female × MPH treated male. These treatment lineages were maintained to produce the third (G<sub>3</sub>) and fourth (G<sub>4</sub>) generations. Because we saw an effect of MPH treatment on G<sub>1</sub> males, but not G<sub>1</sub> females, for statistical analyses of G<sub>2</sub>-G<sub>4</sub> cohorts, we pooled the original treatment groups into two categories: progeny that were descendants of (i) G<sub>1</sub> Control males (A. + B.) or (ii) G<sub>1</sub> MPH treated males (C. + D.). Numbers underneath pairs of fish represent the number of unique lines that contributed to the following generation. Numbers underneath the corresponding progeny groups (small fish) represent the sample size for that group in the open field test. In total, we measured the behaviour of 858 fish. Dark shading in G<sub>1</sub> fish represents MPH treatment.

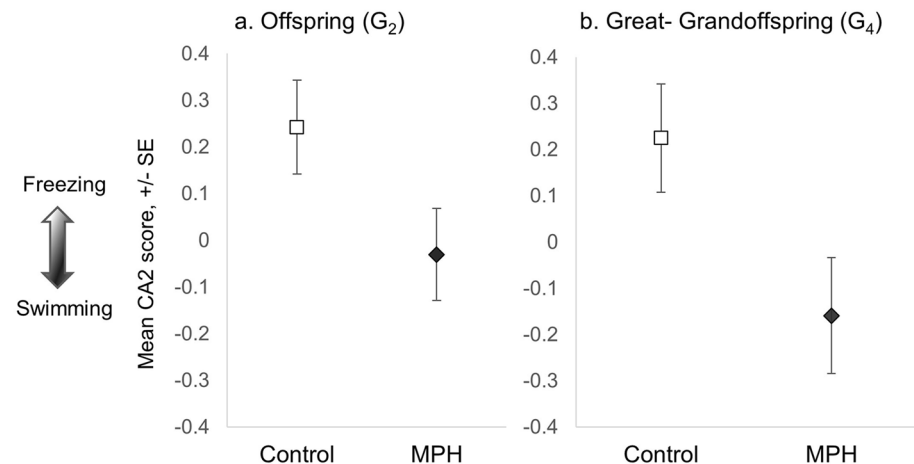
determine if this apparent difference was statistically significant, we performed an additional analysis comparing only G<sub>2</sub> and G<sub>4</sub> ‘Control’ males, including the interaction between ‘generation’ and the covariate age. This analysis revealed that the CA1 scores for G<sub>2</sub> and G<sub>4</sub> ‘Control’ males did not significantly differ ( $F_{1,6,85} = 0.02$ ;  $P = 0.9$ ;  $n = 97$ ; Table S7, suggesting that behavioural patterns were consistent for both generations.

While there is no evidence to suggest that MPH has mutagenic or clastogenic properties<sup>83</sup>, it is possible that our behavioural results were due to differential survival or reproduction in ‘Control’ and ‘MPH treated’ lineages during the course of the experiment. To examine this possibility, we performed several analyses that we describe briefly here (details in SI Methods). To determine if survival differed between ‘Control’ and ‘MPH treatment’ individuals for each of the four cohorts (G<sub>1</sub>-G<sub>4</sub> cohorts), we performed separate survival analyses, by sex, in SAS using *Proc Lifetest*, which uses the Kaplan–Meier estimator<sup>84</sup>. There was not a significant difference between ‘Control’ and ‘MPH treatment’ group survival curves for either sex for any of the four generations (Table S8; Figs. S4–S5). Further, there was not a significant difference between ‘Control’ and ‘MPH treated’ groups in their fertility (whether or not they produced offspring; Table S9), fecundity (number of offspring produced; Table S10), or number of broods produced (Table S11). Finally, to determine if MPH treatment affected offspring sex ratio, we asked if the sex ratio of ‘Control’ and ‘MPH’ groups deviated from expected (1:1), and if the proportion of





**Figure 3.** Associations between age and CA1 scores in the open field test for offspring ( $G_2$ ; panel **a**) and great-grandoffspring ( $G_4$ ; panel **b**). Panel (**a**): Male offspring of both Control ('Control ♂') and MPH treated ('MPH ♂') sires that were tested at a younger age froze relatively more than individuals tested at older ages. For female offspring of Control sires ('Control ♀'), there was not an association between age and CA1 score but, in contrast, female offspring of MPH treated sires ('MPH ♀') froze more if tested at a relatively older age. Panel (**b**): All groups for the great-grandoffspring ( $G_4$ ) cohort exhibited qualitatively similar patterns to the same groups from the  $G_2$  cohort, except for male great-grandoffspring descended from  $G_1$  Control males ('Control ♂'); however, this difference was not statistically significant. Positive CA1 values indicate relatively more freezing and negative values indicate relatively more wall-running. Lines were fit to the predicted values (which incorporate model estimates and random effects) using 'lm' in R and shading corresponds to the 95% confidence intervals.



**Figure 4.** CA2 scores for offspring ( $G_2$ ; panel **a**) and great-grandoffspring ( $G_4$ ; panel **b**) in the open field test. Panel (**a**): Offspring of MPH treated sires spent significantly more time swimming in the open field tub than offspring of Control sires. Panel (**b**): Great-grandoffspring ( $G_4$ ) descended from  $G_1$  MPH treated males spent significantly more time swimming throughout the open field tub than great-grandoffspring descended from  $G_1$  Control males. Positive CA2 values indicate relatively more freezing and negative values indicate relatively more swimming. Symbols represent least-square means for each response variable, +/- one standard error.

offspring that were male per pair (number male offspring/total offspring produced by a given pair) differed for 'Control' and 'MPH' groups for the  $G_2$ – $G_4$  cohorts. There was not a significant effect of MPH treatment on sex ratio (Table S12) nor on the inter-pair variation in proportion of offspring that were male (Table S13). Taken together, these results suggest that the effects of these factors on the behavioural differences between 'Control' and 'MPH treated' lines were minimal.

Because MPH functions by regulating the dopaminergic pathway<sup>45,46</sup>, we measured whole brain dopamine concentration for a subset of individuals from the  $G_2$  to  $G_4$  cohorts immediately following behavioural testing (SI Methods for detail). Contrary to our prediction, there was not a significant effect of  $G_1$  male treatment on whole brain dopamine concentration for any generation tested ( $G_2$  to  $G_4$ , all  $P > 0.5$ ; Table S15).

## Discussion

The main aim of this study was to determine if parental exposure to MPH, a drug commonly prescribed for ADHD treatment, leads to transgenerational effects in generations that were not administered the drug directly. We exposed  $G_1$  male and female guppies to a chronic, low dose of MPH, which affected male behaviour in the open field test. The offspring and great-grandoffspring of MPH treated males also exhibited altered behaviour relative to controls, demonstrating that MPH can cause transgenerational effects through the paternal line. Below, we discuss possible explanations for the lack of a significant effect of MPH on  $G_1$  females and  $G_3$  fish. Further, MPH treatment did not have a significant effect on whole brain dopamine levels, suggesting that the behavioural differences in progeny were not directly modulated by altered dopamine levels in the brain. Because fish and mammals share functional similarities in their brain neurochemistry and behaviour<sup>63,64</sup>, we suggest that our results may be relevant for mammals, including male adolescents and adults who are prescribed MPH.

The behavioural results for  $G_1$  males are similar to those observed in rodents that were chronically administered MPH<sup>74,75</sup>. One way to interpret our results is to consider behavioural patterns that would be adaptive in the wild: freezing in response to an unfamiliar environment allows individuals to assess potential threats and reduce detection by predators<sup>80,81</sup>.  $G_1$  Control males froze in response to the novel environment, which suggests that the reduced freezing and increased ‘wall-running’ of  $G_1$  MPH treated males (and their offspring and great-grandoffspring) would not be adaptive in the wild. Zebrafish exposed to MPH during development also showed reduced freezing in a novel environment<sup>53</sup>. Further, our results for female progeny were age-dependent with the female descendants of  $G_1$  MPH treated males freezing more when they were older (and therefore, larger) when tested. Because predation risk declines with size in the source population used for this experiment, selection pressure on larger females to freeze should be relaxed<sup>85–87</sup>, suggesting that ancestral exposure to MPH interfered with this adaptive response. An exciting next step will be to test if environmentally relevant levels of MPH<sup>88,89</sup> can also lead to transgenerational effects.

While MPH treatment caused a significant effect in  $G_1$  males but not  $G_1$  females, this sex-specific effect was reversed in their progeny for one behavioural metric (CA1), with only female offspring and great-grandoffspring differing in behaviour based on their father’s/great-grandfather’s treatment. Other studies have observed transgenerational effects expressed in the sex opposite to the one originally affected by an environmental stimulus<sup>33–37</sup>. This result further emphasizes the importance of studying transgenerational effects in both sexes.

Determining how MPH caused paternal effects was not part of this study, but it is possible that the effects on offspring resulted from nongenetic modifications to sperm/ejaculate and/or female-mediated paternal effects, both of which would be relevant in natural populations<sup>16,17</sup>. In rodents, MPH alters sperm morphology and germ cell epithelium structure<sup>56</sup>, and drugs that are neurochemically similar to MPH (e.g. cocaine) can cause epigenetic modifications to sperm with subsequent effects on offspring behaviour<sup>59</sup>. To determine the relative contributions of nongenetic epigenetic effects and female-mediated paternal effects in studies of paternal effects, future studies could use artificial insemination to tease apart the effects of male behaviour on female preference/physiology and paternal nongenetic effects.

An important finding of our study is the evidence for transgenerational effects in great-grandoffspring, a generation that was not directly exposed to MPH. Epigenetic modifications have been implicated as a mechanism for transgenerational effects in rodents (although this is contentious<sup>90,91</sup>). Mammalian embryos and germlines undergo epigenetic reprogramming (e.g. erasure of methylation marks), but it has been proposed that epigenetic marks can resist this reprogramming if the environmental stimulus occurs during a critical period<sup>92</sup>. The  $G_1$  male guppies in our study began receiving MPH at one month of age, so their germ cells were exposed to MPH during two critical periods: (i) prior to ‘puberty’ (when germ cells differentiate into spermatogonia), and (ii) during spermatogenesis<sup>92</sup>. In medaka (*Oryzias latipes*), a fish relatively closely related to guppies<sup>93</sup>, epigenetic reprogramming mechanisms are similar to rodents<sup>94,95</sup>. Therefore, if guppies have reprogramming mechanisms like medaka’s, it is possible that epigenetic mechanisms may be responsible for our transgenerational results; future studies could use in vitro fertilization to disentangle the effects of epigenetic modifications to sperm and other potential factors including the effects of male behaviour on female physiology or provisioning to offspring<sup>16,17</sup>. Further, because guppies have internally developing embryos, we suggest that guppies could be a ‘natural’ comparator for investigating transgenerational mechanisms in live bearing species.

We observed significant effects of  $G_1$  male MPH treatment on offspring ( $G_2$ ) and great-grandoffspring ( $G_4$ ) behaviour, but not grandoffspring ( $G_3$ ) behaviour. There are at least two explanations for this pattern: (i) we may have missed the effect in the  $G_3$  because of logistical issues with running this large experiment, and (ii) the expression of behaviour skipped a generation. We may have missed the ‘window of effect’ for the  $G_3$  because many individuals of this generation were tested at an older age than other generations (significantly higher median age). Age had a significant impact on the behaviour of  $G_2$  and  $G_4$  guppies, and rodent behaviour in the open field test also changes with increasing adult age<sup>77,96–98</sup>. Alternatively, it may be that the effect of MPH does disappear for a generation to return in the next. While other studies have observed the disappearance of a phenotype in one generation with a subsequent return in the following generation<sup>99,100</sup>, the mechanism for this is unclear.

Surprisingly, we did not see significant effects of MPH administration on  $G_1$  females nor on their offspring exposed in utero, i.e. there was not a significant effect of  $G_1$  female treatment on the behaviour of their offspring (or on any other generation tested). One possible explanation for this is that mature female guppies become significantly larger than mature males and, thus, females may have taken up a relatively lower amount of MPH per mg body mass from the water in their aquarium (doses were not adjusted for body size). However, given the significant effects of an acute dose of the same concentration on female guppies in the open field test<sup>73</sup>, this explanation seems unlikely. Alternatively, given the sex-specific differences in the dopaminergic pathway<sup>101,102</sup> and other aspects of their biology, it is possible that chronic MPH administration had different effects on males and females.

In this study, we used individuals from an outbred population to investigate transgenerational effects. Therefore, one caveat is that we cannot rule out the possibility that genetic differences between MPH and control lines contributed to our results, however, this issue is not unique to this study<sup>103–106</sup>. We did attempt to standardize initial variation between Control and MPH lines by assigning full siblings to both treatments, and we determined that MPH treatment and Control lines did not significantly differ in their mortality, fecundity, fertility, offspring sex ratio or number of broods produced.

We measured dopamine levels in progeny generations ( $G_2$  to  $G_4$  cohorts) because MPH functions by increasing available dopamine<sup>45,46</sup>, and because rodents prenatally exposed to MPH had increased dopamine levels as adults<sup>55</sup>. However, we did not observe a significant association between paternal MPH treatment and progeny dopamine concentration. Although this is speculative, one possible explanation for why we observed behavioural effects of MPH across generations, but did not see the accompanying effect on dopamine levels, is that chronic  $G_1$  male MPH treatment affected the expression of genes related to other neurotransmitter pathways (e.g. serotonin, glutamate), and this altered expression was transmitted across generations. Other studies have shown that dopamine levels returned to baseline levels after chronic MPH use has ceased but, despite this, altered behavioural patterns persisted; those studies suggested that other neuroadaptations were responsible for these persistent behavioural changes<sup>53,107</sup>. In rats, chronic MPH use during adolescence is associated with persistent changes in the expression of genes involved with glutamate and serotonin receptors, and these have been linked to reward dysfunction, reduced impulsivity, and perseverative behaviour<sup>108</sup>. An important next step would be to determine if chronic exposure to MPH affects the expression of genes involved with glutamate and serotonin receptors.

As far as we are aware, this is the first study of any species to demonstrate that paternal exposure to low, clinically relevant levels of MPH during adolescence and early adulthood can affect offspring phenotype. Because of the similarities between fish and mammals in the dopaminergic system<sup>63</sup> and in behavioural responses to MPH<sup>53,73</sup>, our results are likely relevant to humans. This could have widespread implications given the relatively high prescription rates of MPH to males<sup>49,50</sup>, and because there are currently no precautions for MPH use for men planning to have children<sup>109,110</sup>. Because drug-seeking behaviour is associated with novelty-seeking<sup>41</sup>, offspring of men taking MPH could be prone to an increased propensity for drug abuse<sup>41</sup> and other affective disorders<sup>51</sup>. The next step will be to verify our results in mammalian models.

This study contributes to the accumulating evidence that paternal effects have the potential to span multiple generations. Further, because the behavioural responses of guppies to acute<sup>73</sup> and chronic doses of MPH resemble behavioural responses in mammals, we suggest that the Trinidadian guppy is an excellent comparator for studying the effects of drugs on behaviour and neurochemistry. Future studies should continue to explore transgenerational effects in other natural systems to determine the generality of these effects.

## Methods

To produce the first generation ( $G_1$ ), virgin females were each housed with one male. Thus, all offspring of a given pair were full siblings and were part of the same 'lineage' (Fig. 2). For up to two broods per female, offspring were moved into sibling tanks in the same experimental chamber within 24 h of birth. At one month of age, all focal individuals were moved to their own tank for treatment. All focal fish were housed with an unrelated, non-focal juvenile to avoid the stress of social isolation (details in SI Methods).

**MPH administration.** Treatment with MPH (or vehicle) began at one month of age (30 days old) because this roughly corresponds to human adolescence (~12–18 years old), a period when neurotransmitter systems, including the dopaminergic system, undergo maturation and rearrangement<sup>111</sup>. Based on gonopodium development<sup>112</sup>, the male guppies in our study became sexually mature at approximately 60 days after birth (median: 60 days; range: 42–74 days) and began exhibiting sexual behaviour one to two weeks before maturity<sup>112</sup> (note: there was not a significant effect of MPH treatment on age at sexual maturity ( $F_{1,51.4} = 0.45$ ;  $P = 0.5$ )). Female guppies become sexually mature at approximately the same time as males<sup>112</sup>.

An acute, low dose of MPH ( $2.5 \times 10^{-8}$  g/mL) affected female guppies' behaviour in a novel environment<sup>73</sup> so we used the same dose for chronic treatment in this study. We administered MPH to treatment individuals every other day, as high-performance liquid chromatography showed that this drug delivery schedule maintained aquarium MPH levels at approximately  $2.5 \times 10^{-8}$  g/mL. We dissolved MPH in dechlorinated water before administering doses to treatment tanks with a syringe. Control individuals received water, without MPH, with a syringe. Because we wanted to ensure that MPH concentration in treatment tanks remained relatively consistent over the course of our study and because the densities of guppies were low, we did not perform partial water changes on any experimental tanks. All tanks were covered with fitted lids to reduce water evaporation and, once per week, water was topped up to the original volume. To ensure tanks remained clean and aerated, each tank was equipped with an airstone and ~ three snails. When tanks needed to be cleaned (~ once every two months), the fish and tank water were gently added to another container, the tank was thoroughly cleaned, and then the original water was poured back into the tank. The same cleaning procedures were used for MPH treatment and Control tanks. The  $G_2$ – $G_4$  cohorts were not administered MPH or vehicle, but tank cleaning and maintenance was performed in the same way as for the  $G_1$  cohort.

**Behavioural assays.** We assessed novelty-seeking behaviour of guppies using the widely used open field test, which assesses the behaviour of an individual placed in a novel, open environment<sup>73,77,82,113</sup>. The metrics used for this test can represent an interplay between curiosity/motivation to explore and fearfulness, and it has been validated for these behaviours in guppies<sup>82</sup>. Fish and the water from their 'home tanks' were moved to a new tank in the testing room at least four days before testing to allow them to acclimate to room conditions. On the day of testing, fish were fed at least 30 min prior to testing. For the  $G_1$  cohort only, if it was a 'drug



administration' day, fish were given their respective treatment (MPH for MPH treatment fish or the vehicle for Controls) at least 30 min prior to testing by a research assistant who knew the treatment status of the fish. As in previous studies<sup>73,82</sup>, the open field test was conducted in a 33 × 28 × 12 cm green plastic tub, with black lines delimiting 5.5 × 7 cm rectangles on the bottom, containing 5L fresh conditioned water. Behavioural observations commenced 10–15 s after the experimental fish was gently netted into the tub to ensure that we captured the initial responses to the novel environment. Behaviours were recorded for seven minutes using JWatcher (version 1.0<sup>114</sup>). The observer (AD) was blind to treatment status when scoring behaviour. The water in the tub was replaced with 5L fresh conditioned water after each test. G<sub>1</sub> tests were conducted between 9:00am to 1:00 pm, and this timeframe was further reduced to 9:00 am–11:30 am for the G<sub>2</sub>–G<sub>4</sub> cohorts, because an assistant was no longer required to administer treatments. After testing, fish were either returned to their 'home' tank in the environmental room or were immediately prepared for brain dissection for dopamine quantification (SI Methods).

We summarized all behaviours in the open field test with a CA, except for the proportion of inner/total squares traversed (a metric of exploration<sup>82,115,116</sup>), as this behaviour was measured on a different scale than other behaviours. This behaviour was moderately correlated with CA1 and results were similar to those for CA1 (see Table S16 and Figs. S6–S7). We measured activity level (total squares traversed) because stimulant drugs, including MPH, can affect activity level independent of exploratory behaviour<sup>117,118</sup>; however, MPH treatment did not have a significant effect on activity (Table S17).

**Statistical analyses.** We performed CA in R (version 3.4.4<sup>119</sup>) using the “corresp” function of the MASS package (version 7.3–49<sup>120</sup>). CA works well with data that are zero-inflated and that add to a fixed total<sup>78</sup>. Collectively, the first two components of the CA explained 77% of behavioural variation in the open field test (CA1: 49%; CA2: 28%; Fig. S1). We analyzed CA1, CA2 and other response variables with linear mixed effects models (*Proc Mixed*) in SAS (version 9.4<sup>84</sup>); all models met normality assumptions (based on residual plots). For the G<sub>1</sub> cohort, we included treatment (MPH treated or control) and sex as main effects. We included age as a covariate, as behaviour in the open field test has been observed to change with increasing adult age<sup>77,96–98</sup>. Because MPH functions by affecting dopamine<sup>45,46</sup> and because dopamine function can be sex and age dependent<sup>121–123</sup>, we anticipated that the behavioural effects of MPH could also be sex and age dependent. Therefore, we considered all interactions between MPH treatment, sex, and age. To account for variation in testing conditions, the following were included as additional covariates/cofactor: drug administration day ('yes' or 'no'), time of day tested, date tested, and handling time (time to net the fish from their tank to the open field tub; see SI Methods). Guppy exploratory behaviour can be associated with body size<sup>116</sup>, and MPH can affect body size<sup>124</sup>, however, in this study, we did not observe that variation in body size (standard length) significantly contributed to the behavioural results (Tables S18–S19). We included parental lineage and brood identifier (because some females contributed multiple broods (we note that this happened at similar frequencies for females from each treatment)) as random effects. For analyses of the G<sub>2</sub>–G<sub>4</sub> cohorts, we included G<sub>1</sub> male treatment and sex as main effects and age as a covariate. We considered two additional covariates in these analyses: 'days until isolation from siblings' (because the number of days focal fish remained with siblings varied) and number of broodmates (see SI Methods for details). Because the large number of random and fixed effects created a risk of over parameterization, we only included biologically relevant interactions for which there was a priori justification; specifically, we considered all interactions among G<sub>1</sub> male treatment, sex, and age, and removed non-significant interactions among those terms and other non-significant covariates in a stepwise fashion ( $P > 0.1$ ).

Because individuals in the G<sub>2</sub>–G<sub>4</sub> generations varied in how related they were to one another, we input the pedigree into *Proc Inbreed* to create a genetic covariance matrix. We included this covariance matrix in the model as a random effect<sup>125</sup>. In one case, a model with the full pedigree did not converge (G<sub>4</sub> male dopamine analyses), so we removed this factor and instead included great-grandparental lineages and all relevant interactions as random effects (Table S15).

For all continuous covariates, we centered each variable about its mean<sup>126</sup>. We used the Kenward-Roger method to determine approximate degrees of freedom because sample sizes were not equal for each level of fixed effects<sup>127</sup>. When interactions between main effects were significant, post hoc comparisons between least-square means were performed using “simulate” in *Proc Mixed*.

In separate analyses of the G<sub>1</sub>, G<sub>2</sub>, and G<sub>4</sub> generations, we observed significant effects of G<sub>1</sub> male MPH treatment on behaviour. In total, 5 tests involving G<sub>1</sub> male MPH treatment were significant at  $P < 0.05$ , whereas only 1.6 would be expected by chance (for each generation, 4 terms involved treatment for each of the 2 behaviours, for a total of 32 tests).

All procedures outlined in this study were approved by the Animal Care Committee at the University of Toronto (protocol numbers: 20008920, 20008921, 20009555, 20010160, 20010588, 20009045, 20010020, 20010527). Authorization to import and administer methylphenidate hydrochloride was obtained from the National Compliance and Exemption Division, Office of Controlled Substances, Health Canada (Authorization number: 26982.01.12). All experiments were performed in accordance with ARRIVE guidelines.

## Data availability

All data generated and analyzed for this study are included in this published article (and its Supplementary Information files).

Received: 29 June 2020; Accepted: 2 February 2021

Published online: 17 February 2021

## References

- Mousseau, T. A. & Fox, C. W. The adaptive significance of maternal effects. *Trends Ecol. Evol.* **13**, 403–407 (1998).
- Franklin, T. B., Linder, N., Russig, H., Thöny, B. & Mansuy, I. M. Influence of early stress on social abilities and serotonergic functions across generations in mice. *PLoS ONE* **6**, e21842. <https://doi.org/10.1371/journal.pone.0021842> (2011).
- Gapp, K. *et al.* Implication of sperm RNAs in transgenerational inheritance of the effects of early trauma in mice. *Nat. Neurosci.* **17**, 667–669 (2014).
- McCarthy, D. M. *et al.* Nicotine exposure of male mice produces behavioral impairment in multiple generations of descendants. *PLoS Biol.* **16**, e2006497. <https://doi.org/10.1371/journal.pbio.2006497> (2018).
- Alfonso, S. *et al.* Examining multi- and transgenerational behavioral and molecular alterations resulting from parental exposure to an environmental PCB and PBDE mixture. *Aquat. Toxicol.* **208**, 29–38 (2019).
- Anway, M. D., Memon, M. A., Uzumcu, M. & Skinner, M. K. Transgenerational effect of the endocrine disruptor vinclozolin on male spermatogenesis. *J. Androl.* **27**, 868–879 (2006).
- Crews, D. *et al.* Transgenerational epigenetic imprints on mate preference. *PNAS* **104**, 5942–5946 (2007).
- Crews, D. *et al.* Epigenetic transgenerational inheritance of altered stress responses. *PNAS* **109**, 9143–9148 (2012).
- Gillette, R. *et al.* Sexually dimorphic effects of ancestral exposure to vinclozolin on stress reactivity in rats. *Endocrinology* **155**, 3853–3866 (2014).
- Gillette, R., Son, M. J., Ton, L., Gore, A. C. & Crews, D. Passing experiences on to future generations: endocrine disruptors and transgenerational inheritance of epimutations in brain and sperm. *Epigenetics* **13**, 1106–1126 (2018).
- Bhandari, R., Saal, F. & vom Tillitt, D. Transgenerational effects from early developmental exposures to bisphenol A or 17 $\alpha$ -ethinylestradiol in medaka *Oryzias latipes*. *Sci. Rep.* **5**, 9303. <https://doi.org/10.1038/srep09303> (2015).
- Kidd, K. A. *et al.* Collapse of a fish population after exposure to a synthetic estrogen. *PNAS* **104**, 8897–8901 (2007).
- Skinner, M. K. *et al.* Gene bionetworks involved in the epigenetic transgenerational inheritance of altered mate preference: environmental epigenetics and evolutionary biology. *BMC Genom.* **15**, 377. <https://doi.org/10.1186/1471-2164-15-377> (2014).
- Pembrey, M. E. *et al.* Sex-specific, male-line transgenerational responses in humans. *Eur. J. Hum. Genet.* **14**, 159–166 (2006).
- Moisiadis, V. G. & Matthews, S. G. Glucocorticoids and fetal programming part 1: outcomes. *Nature* **10**, 391–402 (2014).
- Crean, A. J. & Bonduriansky, R. What is a paternal effect?. *Trends Ecol. Evol.* **29**, 554–559 (2014).
- Champagne, F. A. Interplay between paternal germline and maternal effects in shaping development: the overlooked importance of behavioural ecology. *Funct. Ecol.* **34**, 401–413 (2019).
- Sheldon, B. C. Differential allocation: tests, mechanisms and implications. *Trends Ecol. Evol.* **15**, 397–402 (2000).
- Reznik, S. Y., Vaghina, N. P. & Voinovich, N. D. Multigenerational maternal effect on diapause induction in *Trichogramma* species (Hymenoptera: Trichogrammatidae). *Biocontrol Sci. Technol.* **22**, 429–445 (2012).
- Rechavi, O. *et al.* Starvation-induced transgenerational inheritance of small RNAs in *C. elegans*. *Cell* **158**, 277–287 (2014).
- Shama, L. N. S. *et al.* Transgenerational effects persist down the maternal line in marine sticklebacks: gene expression matches physiology in a warming ocean. *Evol. Appl.* **9**, 1096–1111 (2016).
- Dunn, G. A. & Bale, T. L. Maternal high-fat diet effects on third-generation female body size via the paternal lineage. *Endocrinology* **152**, 2228–2236 (2011).
- Skinner, M. K. *et al.* Ancestral dichlorodiphenyltrichloroethane (DDT) exposure promotes epigenetic transgenerational inheritance of obesity. *BMC Med.* **11**, 228. <https://doi.org/10.1186/1741-7015-11-228> (2013).
- Zhu, J., Lee, K. P., Spencer, T. J., Biederman, J. & Bhide, P. G. Transgenerational transmission of hyperactivity in a mouse model of ADHD. *J. Neurosci.* **34**, 2768–2773 (2014).
- Leroux, S. *et al.* Embryonic environment and transgenerational effects in quail. *Genet. Sel. Evol.* **49**, 14. <https://doi.org/10.1186/s12711-017-0292-7> (2017).
- Vera-Chang, M. N. *et al.* Transgenerational hypocortisolism and behavioral disruption are induced by the antidepressant fluoxetine in male zebrafish *Danio rerio*. *PNAS* **115**, E12435–E12442 (2018).
- Sheriff, M. J., McMahon, E. K., Krebs, C. J. & Boonstra, R. Risk severity predicts generational impact. *J. Zool.* **296**, 305–310 (2015).
- Dias, B. G. & Ressler, K. J. Parental olfactory experience influences behavior and neural structure in subsequent generations. *Nat. Neurosci.* **17**, 89–96 (2014).
- He, N. *et al.* Parental life events cause behavioral difference among offspring: adult pre-gestational restraint stress reduces anxiety across generations. *Sci. Rep.* **6**, 39497. <https://doi.org/10.1038/srep39497> (2016).
- Pentinat, T., Ramon-Krauel, M., Cebria, J., Diaz, R. & Jimenez-Chillaron, J. C. Transgenerational inheritance of glucose intolerance in a mouse model of neonatal overnutrition. *Endocrinology* **151**, 5617–5623 (2010).
- Wei, Y. *et al.* Paternally induced transgenerational inheritance of susceptibility to diabetes in mammals. *PNAS* **111**, 1873–1878 (2014).
- Cropley, J. E. *et al.* Male-lineage transmission of an acquired metabolic phenotype induced by grand-paternal obesity. *Mol. Metab.* **5**, 699–708 (2016).
- Dunn, G. A., Morgan, C. P. & Bale, T. L. Sex-specificity in transgenerational epigenetic programming. *Horm. Behav.* **59**, 290–295 (2011).
- Glover, V. & Hill, J. Sex differences in the programming effects of prenatal stress on psychopathology and stress responses: an evolutionary perspective. *Physiol. Behav.* **106**, 736–740 (2012).
- Saavedra-Rodríguez, L. & Feig, L. A. Chronic social instability induces anxiety and defective social interactions across generations. *Biol. Psychiatry* **73**, 44–53 (2013).
- Moisiadis, V. G., Constantinof, A., Kostaki, A., Szyf, M. & Matthews, S. G. Prenatal glucocorticoid exposure modifies endocrine function and behaviour for 3 generations following maternal and paternal transmission. *Sci. Rep.* **7**, 11814. <https://doi.org/10.1038/s41598-017-11635-w> (2017).
- Hellmann, J. K., Carlson, E. R. & Bell, A. M. Sex-specific plasticity across generations II: grandpaternal effects are lineage specific and sex specific. *J. Anim. Ecol.* **89**, 2800–2812 (2020).
- gene duplications and functional diversification in Craniates. Le Crom, S., Kapsimali, M., Barome, P.-O. & Vernier, P. Dopamine receptors for every species. *J. Struct. Funct. Genomics* **3**, 161–176 (2003).
- Melis, M. R. & Argiolas, A. Dopamine and sexual behavior. *Neurosci. Biobehav. R.* **19**, 19–38 (1995).
- Pfaus, J. G., Ismail, N. & Coria-Avila, G. A. Sexual motivation. In *Encyclopedia of Behavioral Neuroscience* (eds. Koob, G. F., Le Moal, M. & Thompson, R. F.) 201–209 (Oxford, Oxford Academic Press, 2010).
- Bardo, M. T., Donohew, R. L. & Harrington, N. G. Psychobiology of novelty seeking and drug seeking behavior. *Behav. Brain Res.* **77**, 23–43 (1996).
- Mällo, T. *et al.* Rats with persistently low or high exploratory activity: behaviour in tests of anxiety and depression and extracellular levels of dopamine. *Behav. Brain Res.* **177**, 269–281 (2006).
- Smith, B. R. & Blumstein, D. T. Fitness consequences of personality: a meta-analysis. *Behav. Ecol.* **19**, 448–455 (2007).
- Csoka, A. B. & Szyf, M. Epigenetic side-effects of common pharmaceuticals: a potential new field in medicine and pharmacology. *Med. Hypotheses* **73**, 770–780 (2009).

45. Kuczenski, R. & Segal, D. S. Effects of methylphenidate on extracellular dopamine serotonin, and norepinephrine: comparison with amphetamine. *J. Neurochem.* **68**, 2032–2037 (1997).
46. Gamo, N. J., Wang, M. & Arnsten, A. F. T. Methylphenidate and atomoxetine enhance prefrontal function through  $\alpha_2$ -adrenergic and dopamine D<sub>1</sub> receptors. *J. Am. Acad. Child Adolesc. Psychiatry* **49**, 1011–1023 (2010).
47. Greenhill, L. L. *et al.* Guidelines and algorithms for the use of methylphenidate in children with attention-deficit/hyperactivity disorder. *J. Atten. Disord.* **6**, S89–S100 (2002).
48. Kessler, R. C. *et al.* The prevalence and correlates of adult ADHD in the United States: results from the national comorbidity survey replication. *Am. J. Psychiatry* **163**, 716–723 (2006).
49. Visser, S. N. *et al.* Trends in the parent-report of health care provider-diagnosed and medicated attention-deficit/hyperactivity disorder: United States, 2003–2011. *J. Am. Acad. Child. Psychiatry* **53**, 34–46 (2014).
50. Karlstad, Ø. *et al.* Use of drugs for ADHD among adults—a multinational study among 15.8 million adults in the Nordic countries. *Eur. J. Clin. Pharmacol.* **72**, 1507–1514 (2016).
51. Biederman, J. Attention-deficit/hyperactivity disorder: a selective overview. *Biol. Psychiatry* **57**, 1215–1220 (2005).
52. McFadyen-Leussis, M. P., Lewis, S. P., Bond, T. L. Y., Carrey, N. & Brown, R. E. Prenatal exposure to methylphenidate hydrochloride decreases anxiety and increases exploration in mice. *Pharmacol. Biochem. Behav.* **77**, 491–500 (2004).
53. Levin, E. D. *et al.* 2011. Persistent behavioral impairment caused by embryonic methylphenidate exposure in zebrafish. *Neurotoxicol. Teratol.* **33**, 668–673 (2011).
54. Lloyd, S. A. *et al.* Prenatal exposure to psychostimulants increases impulsivity, compulsivity, and motivation for rewards in adult mice. *Physiol. Behav.* **119**, 43–51 (2013).
55. Lepelletier, F. X. *et al.* Prenatal exposure to methylphenidate affects the dopamine system and the reactivity to natural reward in adulthood in rats. *Int. J. Neuropsychoph.* <https://doi.org/10.1093/ijnp/pyu044> (2015).
56. Montagnini, B. G. *et al.* Effects of repeated administration of methylphenidate on reproductive parameters in male rats. *Physiol. Behav.* **133**, 122–129 (2014).
57. He, F., Lidow, I. A. & Lidow, M. S. Consequences of paternal cocaine exposure in mice. *Neurotoxicol. Teratol.* **28**, 198–209 (2006).
58. Killinger, C. E., Robinson, S. & Stanwood, G. D. Subtle biobehavioral effects produced by paternal cocaine exposure. *Synapse* **66**, 902–908 (2012).
59. Vassoler, F. M., White, S. L., Schmidt, H. D., Sadri-Vakili, G. & Pierce, R. C. Epigenetic inheritance of a cocaine-resistance phenotype. *Nat. Neurosci.* **16**, 42–67 (2013).
60. Fischer, D. K., Rice, R. C., Rivera, A. M., Donohoe, M. & Rajadhyaksha, A. M. Altered reward sensitivity in female offspring of cocaine-exposed fathers. *Behav. Brain Res.* **332**, 23–31 (2017).
61. Wimmer, M. E. *et al.* Paternal cocaine taking elicits epigenetic remodeling and memory deficits in male progeny. *Mol. Psychiatry* **22**, 1641–1650 (2017).
62. Yano, M. & Steiner, H. Methylphenidate and cocaine: the same effects on gene regulation?. *Trends Pharmacol. Sci.* **28**, 588–596 (2007).
63. Hall, Z. J., De Serrano, A. R., Rodd, F. H. & Tropepe, V. Casting a wider fish net on animal models in neuropsychiatric research. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **55**, 7–15 (2014).
64. Fontana, B. D., Mezzomo, N. J., Kalueff, A. V. & Rosemberg, D. B. The developing utility of zebrafish models of neurological and neuropsychiatric disorders: a critical review. *Exp. Neurol.* **299**, 157–171 (2018).
65. Reznick, D. N. The impact of predation on life history evolution in Trinidadian guppies: genetic basis of observed life history patterns. *Evolution* **36**, 1236–1250 (1982).
66. DeMarais, A. & Oldis, D. Matrotrophic transfer of fluorescent microspheres in Poeciliid fishes. *Copeia* **3**, 632–636 (2005).
67. Hughes, K. A., Du, L., Rodd, F. H. & Reznick, D. N. Familiarity leads to female mate preference for novel males in the guppy *Poecilia reticulata*. *Anim. Behav.* **58**(907), 916 (1999).
68. Rodd, F. H., Hughes, K. A., Grether, G. F. & Baril, C. T. A possible non-sexual origin of mate preference: are male guppies mimicking fruit?. *Proc. R. Soc. B Biol. Sci.* **269**, 475–481 (2002).
69. Valvo, J., Rodd, F. H. & Hughes, K. A. Consistent female preference for rare and unfamiliar male color patterns in wild guppy populations. *Behav. Ecol.* **30**, 1672–1681 (2019).
70. Daniel, M. J., Koffinas, L. & Hughes, K. A. Mating preference for novel phenotypes can be explained by general neophilia in female guppies. *Am. Nat.* **196**, 414–428 (2020).
71. Deacon, A. E., Ramnarine, I. W. & Magurran, A. E. How reproductive ecology contributes to the spread of a globally invasive fish. *PLoS ONE* **6**, e24416. <https://doi.org/10.1371/journal.pone.0024416> (2011).
72. Hughes, K. A., Houde, A. E., Price, A. C. & Rodd, F. H. Mating advantage for rare males in wild guppy populations. *Nature* **503**, 108–110 (2013).
73. De Serrano, A. R., Fong, C. & Rodd, F. H. Effects of methylphenidate on responses to novelty in a teleost fish (*Poecilia reticulata*). *Behav. Brain Res.* **302**, 53–59 (2016).
74. Schmitz, F. *et al.* Methylphenidate causes behavioral impairments and neuron and astrocyte loss in the hippocampus of juvenile rats. *Mol. Neurobiol.* **54**, 4201–4216 (2016).
75. Bolaños, C. A., Barrot, M., Berton, O., Wallace-Black, D. & Nestler, E. J. Methylphenidate treatment during pre- and periadolescence alters behavioral responses to emotional stimuli at adulthood. *Biol. Psychiatry* **54**, 1317–1329 (2003).
76. Bell, A. M. & Hellman, J. K. An integrative framework for understanding the mechanisms and multigenerational consequences of transgenerational plasticity. *Annu. Rev. Ecol. Evol. S.* **50**, 97–118 (2019).
77. Walsh, R. N. & Cummins, R. A. Open-field test—critical review. *Psychol. Bull.* **83**, 482–504 (1976).
78. Hill, M. O. Correspondence analysis: a neglected multivariate method. *J. R. Stat. Soc. C Appl.* **23**, 340–354 (1974).
79. Godin, J. G. J. Evading predators. In *Behavioural Ecology of Teleost Fishes* (ed. Godin, J. G. J.) 191–236 (Oxford, Oxford University Press, 1997).
80. Sih, A. Foraging strategies and the avoidance of predation by an aquatic insect *Notonecta Hoffmanni*. *Ecology* **63**(786), 796 (1982).
81. McPeck, M. A., Grace, M. & Richardson, J. M. L. Physiological and behavioral responses to predators shape the growth/predation risk trade-off in damselflies. *Ecology* **82**, 1535–1545 (2001).
82. Burns, J. G. The validity of three tests of temperament in guppies (*Poecilia reticulata*). *J. Comp. Psychol.* **122**, 344–356 (2008).
83. Morris, S. M. *et al.* The genetic toxicity of methylphenidate: a review of the current literature. *J. Appl. Toxicol.* **32**, 756–764 (2012).
84. SAS Institute. *SAS/STAT 9.4 User's Guide* (SAS Institute, Cary, 2013).
85. Seghers, B. H. Feeding behavior and terrestrial locomotion in the cyprinodontid fish, *Rivulus hartii* (Boulenger). *Verh. Internat. Verein. Limnol.* **20**, 2055–2059 (1978).
86. Mattingly, H. T. & Butler, M. J. Laboratory predation on the Trinidadian guppy: implications for the size-selective predation hypothesis and guppy life history evolution. *OIKOS* **69**, 54–64 (1994).
87. Reznick, D. N., Butler, M. J., Rodd, F. H. & Ross, P. N. Life history evolution in guppies (*Poecilia reticulata*): 6—differential mortality as a mechanism for natural selection. *Evolution* **50**, 1651–1660 (1996).
88. Bijlsma, L., Emke, E., Hernandez, F. & de Voogt, P. Investigation of drugs of abuse and relevant metabolites in Dutch sewage water by liquid chromatography coupled to high resolution mass spectrometry. *Chemosphere* **89**, 1399–1406 (2012).

89. Racamonde, I., Rodil, R., Quintana, J. B., Villaverde-de-Saa, E. & Cela, R. Determination of benzodiazepines, related pharmaceuticals and metabolites in water by solid-phase extraction and liquid-chromatography-tandem mass spectrometry. *J. Chromatogr. A* **1352**, 69–79 (2014).
90. Laland, K. *et al.* Does evolutionary theory need a rethink?. *Nature* **514**, 161–164 (2014).
91. Horsthemke, B. A critical view on transgenerational epigenetic inheritance in humans. *Nat. Commun.* **9**, 2973. <https://doi.org/10.1038/s41467-018-05445-5> (2018).
92. Soubry, A., Hoyo, C., Jirtle, R. L. & Murphy, S. K. A paternal environmental legacy: evidence for epigenetic inheritance through the male germ line. *BioEssays* **36**, 359–371 (2014).
93. Hughes, L. C. *et al.* Comprehensive phylogeny of ray-finned fishes (Actinopterygii) based on transcriptomic and genomic data. *PNAS* **115**, 6249–6254 (2018).
94. Wang, X. & Bhandari, R. K. DNA methylation dynamics during epigenetic reprogramming of medaka embryo. *Epigenetics* **14**, 611–622 (2019).
95. Wang, X. & Bhandari, R. K. The dynamics of DNA methylation during epigenetic reprogramming of primordial germ cells in medaka (*Oryzias latipes*). *Epigenetics* **15**, 483–498 (2020).
96. Furchtgott, E., Dees, J. W. & Weckhin, S. Open-field exploration as a function of age. *J. Comp. Physiol. Psychol.* **54**, 386–388 (1961).
97. Werboff, J. & Havlena, J. The effects of aging on open-field behavior. *Psychol. Rep.* **10**, 395–398 (1962).
98. Valle, F. P. Rats performance on repeated tests in open field as a function of age. *Psychon. Sci.* **23**, 333–335 (1971).
99. Franklin, T. B. *et al.* Epigenetic transmission of the impact of early stress across generations. *Biol. Psychiatry* **68**, 408–415 (2010).
100. McBirney, M. *et al.* Atrazine induced epigenetic transgenerational inheritance of disease, lean phenotype and sperm epimutation pathology biomarkers. *PLoS One* **12**, e0184306. <https://doi.org/10.1371/journal.pone.0184306> (2017).
101. Becker, J. B. & Chartoff, E. Sex differences in neural mechanisms mediating reward and addiction. *Neuropsychopharmacology* **44**, 166–183 (2019).
102. Rubinow, D. R. & Schmidt, P. J. Sex differences and the neurobiology of affective disorders. *Neuropsychopharmacology* **44**, 111–128 (2019).
103. Eriksson, K., Halkka, O., Lokki, J. & Saura, A. Enzyme polymorphism in feral, outbred and inbred rats (*Rattus norvegicus*). *Heredity* **37**, 341–349 (1976).
104. Connor, J. L. & Belucci, M. J. Natural selection resisting inbreeding depression in captive wild housemice (*Mus musculus*). *Evolution* **33**, 929–940 (1979).
105. Mina, N. S., Sheldon, B. L., Yoo, B. H. & Frankham, R. Heterozygosity at protein loci in inbred and outbred lines of chickens. *Poult. Sci.* **70**, 1864–1872 (1991).
106. Turissini, D. A., Gamez, S. & White, B. J. Genome-wide patterns of polymorphism in an inbred line of the African malaria mosquito *Anopheles gambiae*. *Genome Biol. Evol.* **6**, 3094–3104 (2014).
107. Gray, J. D. *et al.* Methylphenidate administration to juvenile rats alters brain areas involved in cognition, motivated behaviors, appetite, and stress. *J. Neurosci.* **27**, 7196–7207 (2007).
108. Marco, E. M. *et al.* Neurobehavioral adaptations to methylphenidate: the issue of early adolescent exposure. *Neurosci. Biobehav. Rev.* **35**, 1722–1739 (2011).
109. American Psychiatric Association. Attention-deficit/hyperactivity disorder. In *Diagnostic and Statistical Manual of Mental Disorders: DSM-5* (American Psychiatric Association, Philadelphia, 2014).
110. Novartis Pharmaceuticals Canada Inc. *Product monograph for Ritalin and Ritalin SR* (2017).
111. Brenhouse, H. C. & Andersen, S. L. Developmental trajectories during adolescence in males and females: a cross-species understanding of underlying brain changes. *Neurosci. Biobehav. Rev.* **35**, 1687–1703 (2011).
112. Houde, A. E. *Sex, Color, and Mate Choice in Guppies* (Princeton, Princeton University Press, 1997).
113. Yoshida, M., Nagamine, M. & Uematsu, K. Comparison of behavioral responses to a novel environment between three teleosts, bluegill *Lepomis macrochirus*, crucian carp *Carassius langsdorffii*, and goldfish *Carassius auratus*. *Fisheries Sci.* **71**, 314–319 (2005).
114. Blumstein, D. T., Evans, C. S. & Daniels, J. C. *JWatcher* (v. 1.0, 2006).
115. Ahmad, F. & Richardson, M. K. Exploratory behaviour in the open field test adapted for larval zebrafish: impact of environmental complexity. *Behav. Process.* **92**, 88–98 (2013).
116. Burns, J. G., Price, A. C., Thomson, J. D., Hughes, K. A. & Rodd, F. H. Environmental and genetic effects on exploratory behavior of high- and low-predation guppies (*Poecilia reticulata*). *Behav. Ecol. Sociobiol.* **70**, 1187–1196 (2016).
117. Marriott, A. S. The effects of amphetamine, caffeine and methylphenidate on the locomotor activity of rats in an unfamiliar environment. *Int. J. Neuropharmacol.* **7**, 487–491 (1968).
118. Dyne, L. J. & Hughes, R. N. Effects of methylphenidate on activity and reactions to novelty in rats. *Psychon. Sci.* **19**, 267–268 (1970).
119. R Core Team. *R: A Language and Environment for Statistical Computing* (Vienna, R Foundation for Statistical Computing, 2018).
120. Venables, W. N. & Ripley, B. D. *Modern Applied Statistics with S* (Springer, Berlin, 2002).
121. Volkow, N. D. *et al.* Dopamine transporters decrease with age. *J. Nucl. Med.* **37**, 554–559 (1996).
122. Andersen, S. L. & Teicher, M. H. Sex differences in dopamine receptors and their relevance to ADHD. *Neurosci. Biobehav. Rev.* **24**, 137–141 (2000).
123. Arvidsson, E., Viereckel, T., Mikulovic, S. & Wallén-Mackenzie, Å. Age- and sex-dependence of dopamine release and capacity for recovery identified in the dorsal striatum of C57/Bl6J mice. *PLoS One* **9**, e99592. <https://doi.org/10.1371/journal.pone.0099592> (2014).
124. Faraone, S. V., Biederman, J., Morley, C. P. & Spencer, T. J. Effect of stimulants on height and weight: a review of the literature. *J. Am. Acad. Child Adolesc. Psychiatry* **47**, 994–1009 (2008).
125. Tempelman, R. J. & Rosa, G. J. M. Empirical Bayes approaches to mixed model inference in quantitative genetics. In *Genetic Analysis of Complex Traits Using SAS* (ed. Saxton, A.) (SAS Institute, Cary, 2004).
126. Schielzeth, H. Simple means to improve the interpretability of regression coefficients. *Methods Ecol. Evol.* **1**, 103–113 (2010).
127. Littell, R. C., Milliken, G. A., Stroup, W. W., Wolfinger, R. D. & Schabenberger, O. *SAS for Mixed Models* (SAS Institute, Cary, 2006).

## Acknowledgements

This work was supported by a Natural Sciences and Engineering Research Doctoral Grant and Ontario Graduate Scholarship to AD, and NSERC Discovery Grants to FHR (grant numbers: RGPIN 216891 and RGPIN-2018-06402). We thank Marla Sokolowski and Vince Tropepe for their advice on experimental design; Ashley Bruce, Marla Sokolowski, Megan Frederickson, and Vince Tropepe for use of their lab space; Adam Cembrowski, Ashley Bruce, Ina Anreiter, and Oscar Vasquez for assistance with equipment; Don Jackson and John Stinchcombe for their advice with statistical analyses; David Crews for suggestions on the manuscript; Eric Chen and Wenjiang Zhang at the Princess Margaret Cancer Centre for performing HPLC; Emily Xie, Lisa St. Amant, and Nicole



Slavin for assistance with experimental set-up; and our numerous undergraduate assistants for their assistance with guppy husbandry.

### Author contributions

A.D. conceived, designed, and coordinated the study, administered treatments, performed behavioural assays, prepared raw data for analysis, carried out statistical analyses, prepared figures and tables, and drafted the manuscript; K.H. helped to design the study, provided statistical advice, and helped draft the manuscript; F.H.R. conceived and designed the study, and helped draft the manuscript. All authors gave final approval for publication and agree to be held accountable for the work performed therein.

### Competing interests

The authors declare no competing interests.

### Additional information

**Supplementary Information** The online version contains supplementary material available at <https://doi.org/10.1038/s41598-021-83448-x>.

**Correspondence** and requests for materials should be addressed to A.R.D.S.

**Reprints and permissions information** is available at [www.nature.com/reprints](http://www.nature.com/reprints).

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2021