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Myocardial Mitochondria at the Intersection of Health and Disease

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The air that we breathe and the food that we eat reach their confluency in the mitochondria to derive the energy that the cell is dependent upon. The heart is the most aerobic of organs and has little anaerobic reserve compared to the constant demands placed upon it. At rest the arterial-venous oxygen extraction from the blood is the greatest from the heart and this only goes up with exercise. Common to all cell types residing in the heart is the need for energy and mitochondrial dysfunction is a significant contributor to cell death across the spectrum of cardiac disorders. Mitochondrial failure within any one cell type creates varying complications that eventually manifest as heart failure.

The field of mitochondrial biology has undergone several paradigm shifts since the early nineties. The pioneering work of Krebs, Chance, and others had brought the field to a plateau with the focus on ATP generation. It was not until the implications of mitochondria's role in apoptosis and the cell's health, was there a strong resurgence of interest in mitochondrial biology [1,2]. Since then, a second major road of significance, dug predominantly by D. Wallace, has been in identifying the participation of mitochondria DNA (mtDNA) alterations and damage as a common pathology in unrelated symptoms [3,4]. And third avenue, that mitochondria are not just the isolated organelles pictured in electron micrographs, but dynamic entities that undergo significant morphological changes as a course of their normal function [5].

The mitochondrial genome is a circular double-stranded DNA of more than 16 Kb in humans. It codes for 37 genes including 13 of the more than 1000 proteins indigenous to the mammalian mitochondria. Mitochondrial disorders are a heterogeneous group of diseases that may be characterized by maternal inheritance, heteroplasmy, and threshold effect. Mitochondrial dysfunction and mtDNA damage has been reported in diabetes, alcoholism, cancer, skeletal muscle disorders, and neurodegenerative diseases such as Barth Syndrome, MELAS, ALS, or LHON [6–9]. As one example, several mtDNA mutations have been identified that represent a high risk for the development of diabetes [10–13]. Mutations may take the form of deletions, rearrangements, or missense mutations that interfere with protein synthesis. The etiology for the accumulation of mtDNA mutations and deletions is not completely understood [14–16].

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Separate from inborn errors, mutations and deletions of mitochondrial DNA (mtDNA) accumulate as a function of the aging process or from environmental influences and thought responsible for the decline in mitochondrial function [14,17]. Indeed the “mitochondrial theory of aging” is centered on the accumulation of mtDNA mutations and an increase in mitochondrial oxidative stress, creating a “vicious” circle that accelerates this process. Mitochondrial DNA is thought to be at greater risk for oxidant-induced damage due to its close proximity to the electron transport chain and high levels of superoxide production. Also, unlike genomic DNA, mtDNA contains little intron DNA which may serve to absorb damage from chronically elevated oxidative stress. In postmitotic cells, the mitochondrial genome continues to replicate about once a month [18]. Oxidant-induced mtDNA mutations that are not corrected by mitochondrial DNA repair mechanisms are fixed in the mitochondrial genome.

Mitochondrial dependent ROS (mtROS) generation has been accepted as the singular cause of mtDNA damage in different pathophysiologic states [19]. However, antioxidant therapy studies have yielded results that range from disappointing to a potentially detrimental effect of antioxidants [20–23]. Other approaches that raise or lower mitochondrial antioxidant capacity have also yielded conflicting results [24,25].

Historically, ROS has been viewed as a waste byproduct of aerobic metabolism. However the cell’s ability to generate specific different oxidant species indicates a useful purpose for normal cell function [26,27]. More recently, this has taken the concept of compartmentalization of signaling; by physical separation of ROS production as well as activation of distinct enzyme complexes [26,27]. Several lines of evidence point towards increased mtROS as a significant cause of mitochondrial dysfunction [28–34].

A limitation to some studies is that they examined mtROS changes only as an early event. Others that examined an extended timeframe point to a more complex interaction of ROS within the mitochondria and suggest that alternative pathways may mediate the effect of ROS on mitochondrial function and mtDNA integrity [35–37]. Mitochondria have both endogenous oxidant buffers and mtDNA repair capability but this failure suggests that there may be limits to functional recovery. It may be that the pathology generates a transient signal either internal or external to the mitochondria that is prolonged by activation of pathways that exacerbate the initial insult. Going forward, work to differentiate the different oxidant species and their sources in the normal and diseased heart will clarify their respective roles.

The concept that increased mtDNA mutation rates leads to a vicious cycle of increased oxidative stress has been challenged by investigations using transgenic mice that express a cardiac-specific “proof-reading deficient” mitochondrial DNA polymerase (mtDNA-Pol^{def}). In those studies cardiac cells accumulated mtDNA mutations at a rate of more than 20 fold compared to controls, demonstrated increased apoptosis, and presented with significant heart failure [38,39]. Despite this, there was not a significant change in mitochondrial function; the P/O ratio and respiratory control index were similar in mtDNA-Pol^{def} and controls. Significantly, markers for ROS did not increase suggesting that oxidative stress was not an obligate mediator of mtDNA mutations. This model suggests that any increase in mtDNA

mutations may serve as a signal for the initiation of apoptosis. Further, Herlein et al have argued that in mild diabetes or prediabetes, mitochondrial superoxide may not be elevated in contrast to its decided presence in more severe diabetic states [31]. More recently our investigations suggest that separate from a direct effect of mtROS on mtDNA, mitochondrial topoisomerase dysfunction increased mtDNA strand breakage [37,40]. Collectively these studies suggest that more than just mtROS promotes mtDNA damage leading to mitochondrial and cellular degradation within the heart. And it remains that preservation and protection of mtDNA becomes a focal point for novel clinical strategies.

Mitochondria are dynamic organelles and the fission/fusion processes have a significant role in the myocardium. The familiar pictures from electron microscopy showing distinct mitochondria interspersed amongst the sarcomere are somewhat misleading. It portrays the mitochondria as individual entities, but we now know they may form reticulated networks [5]. This dynamic exchange allows for sharing of mitochondrial contents. If sharing did not occur then it would be necessary for each mitochondrion to coordinate with the nucleus to import of all the proteins it could not be synthesized. Sharing also allows for the dilution of mtDNA mutations that would directly interfere with respiratory function; permitting greater mutational loads to be carried.

Importantly, sharing may allow for segregation of dysfunction mitochondria as an early step towards autophagy or apoptosis [41], increased fission resulting in smaller more numerous mitochondria has been associated with elevated caspase activity, an initial step in apoptosis [42]. To date several proteins critical for fusion and fission processes have been identified including Mfn1, Mfn2, OPA1, Drp1, & PINK1. Although mutations of OPA1 and Mfn2 are mostly associated with the neuropathy Charcot-Marie-Tooth disease, they also have a role in myocardial mitochondrial function [43,44]. Inhibition of Drp1 appeared to protect mitochondria during an ischemia/reperfusion challenge suggesting that control of mitochondrial morphology is clinically relevant [45]. Beyond the gain of function and loss of function experimental paradigms, the field is still developing the tools to study the role of fusion and fission in myocardial mitochondria. We know that both processes are essential for maintenance of the phenotype, but specific perturbations associated with different pathologies remain to be more fully explored.

Cardiovascular disease has a higher incidence of mortality in patients with diabetics, alcoholism, cancer, and survivors of cancer treatment compared to the general population. The continuously beating heart is the most metabolically active organ in the body. This would not be possible without the continuous support from the mitochondria. This view developed in the early part of the 20th century brought the field to a plateau of understanding. Apoptosis was originally described as a developmental step but its role in the maintenance of the phenotype cleared the path for mitochondria's participation. Beyond this has come the recognition of the many roles mitochondria has beyond ATP regeneration and their vulnerability in cardiac pathology. As we learn more, viable new clinical strategies will hopefully become apparent to protect mitochondrial function in the heart.

References

1. Kerr JF, Wyllie AH, Currie AR. Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *Br J Cancer*. 1972; 26:239–257. [PubMed: 4561027]
2. Hockenbery D, Nuñez G, Milliman C, Schreiber RD, Korsmeyer SJ. Bcl-2 is an inner mitochondrial membrane protein that blocks programmed cell death. *Nature*. 1990; 348:334–336. [PubMed: 2250705]
3. Wallace DC. Diseases of the mitochondrial DNA. *Annu Rev Biochem*. 1992; 61:1175–1212. [PubMed: 1497308]
4. Wallace DC. A mitochondrial paradigm of metabolic and degenerative diseases, aging, and cancer: a dawn for evolutionary medicine. *Annu Rev Genet*. 2005; 39:359–407. [PubMed: 16285865]
5. Bereiter-Hahn J, Vöth M. Dynamics of mitochondria in living cells: shape changes, dislocations, fusion, and fission of mitochondria. *Microsc Res Tech*. 1994; 27:198–219. [PubMed: 8204911]
6. Gokey NG, Cao Z, Pak JW, Lee D, McKiernan SH, et al. Molecular analyses of mtDNA deletion mutations in microdissected skeletal muscle fibers from aged rhesus monkeys. *Aging Cell*. 2004; 3:319–326. [PubMed: 15379855]
7. Domenech E, Gomez-Zaera M, Nunes V. Wolfram/DIDMOAD syndrome, a heterogenic and molecularly complex neurodegenerative disease. *Pediatr Endocrinol Rev*. 2006; 3:249–257. [PubMed: 16639390]
8. Kagan J, Srivastava S. Mitochondria as a target for early detection and diagnosis of cancer. *Crit Rev Clin Lab Sci*. 2005; 42:453–472. [PubMed: 16390681]
9. Scliacco M, Prele A, Fagiolari G, Bordoni A, Crimi M, et al. A case of CPT deficiency, homoplasmic mtDNA mutation and ragged red fibers at muscle biopsy. *J Neurol Sci*. 2005; 239:21–24. [PubMed: 16168441]
10. Liou CW, Huang CC, Lee CF, Lin TK, Wei YH. Low antioxidant content and mutation load in mitochondrial DNA A3243G mutation-related diabetes mellitus. *J Formos Med Assoc*. 2003; 102:527–533. [PubMed: 14569316]
11. Lin TK, Chen SD, Wang PW, Wei YH, Lee CF, et al. Increased oxidative damage with altered antioxidative status in type 2 diabetic patients harboring the 16189 T to C variant of mitochondrial DNA. *Ann N Y Acad Sci*. 2005; 1042:64–69. [PubMed: 15965046]
12. Kokaze A, Ishikawa M, Matsunaga N, Yoshida M, Makita R, et al. Longevity-associated mitochondrial DNA 5178 C/A polymorphism is associated with fasting plasma glucose levels and glucose tolerance in Japanese men. *Mitochondrion*. 2005; 5:418–425. [PubMed: 16271520]
13. Guo LJ, Oshida Y, Fuku N, Takeyasu T, Fujita Y, et al. Mitochondrial genome polymorphisms associated with type-2 diabetes or obesity. *Mitochondrion*. 2005; 5:15–33. [PubMed: 16060290]
14. Kadenbach B, Münscher C, Frank V, Müller-Höcker J, Napiwotzki J. Human aging is associated with stochastic somatic mutations of mitochondrial DNA. *Mutat Res*. 1995; 338:161–172. [PubMed: 7565871]
15. Wang E, Wong A, Cortopassi G. The rate of mitochondrial mutagenesis is faster in mice than humans. *Mutat Res*. 1997; 377:157–166. [PubMed: 9247611]
16. Cortopassi GA, Wang E. There is substantial agreement among interspecies estimates of DNA repair activity. *Mech Ageing Dev*. 1996; 91:211–218. [PubMed: 9055244]
17. Beckman KB, Ames BN. The free radical theory of aging matures. *Physiol Rev*. 1998; 78:547–581. [PubMed: 9562038]
18. Cortopassi G, Wang E. Modelling the effects of age-related mtDNA mutation accumulation; complex I deficiency, superoxide and cell death. *Biochim Biophys Acta*. 1995; 1271:171–176. [PubMed: 7599205]
19. Nishikawa T, Edelstein D, Du XL, Yamagishi S, Matsumura T, et al. Normalizing mitochondrial superoxide production blocks three pathways of hyperglycaemic damage. *Nature*. 2000; 404:787–790. [PubMed: 10783895]
20. Miller ER 3rd, Pastor-Barriuso R, Dalal D, Riemersma RA, Appel LJ, et al. Meta-analysis: high-dosage vitamin E supplementation may increase all-cause mortality. *Ann Intern Med*. 2005; 142:37–46. [PubMed: 15537682]

21. Eidelman RS, Hollar D, Hebert PR, Lamas GA, Hennekens CH. Randomized trials of vitamin E in the treatment and prevention of cardiovascular disease. *Arch Intern Med.* 2004; 164:1552–1556. [PubMed: 15277288]
22. Vivekananthan DP, Penn MS, Sapp SK, Hsu A, Topol EJ. Use of antioxidant vitamins for the prevention of cardiovascular disease: meta-analysis of randomised trials. *Lancet.* 2003; 361:2017–2023. [PubMed: 12814711]
23. Löhrike B, Xu J, Weitzel JM, Krüger B, Goldammer T, et al. N-acetylcysteine impairs survival of luteal cells through mitochondrial dysfunction. *Cytometry A.* 2010; 77:310–320. [PubMed: 20151456]
24. Larosche I, Choumar A, Fromenty B, Lettéron P, Abbey-Toby A, et al. Prolonged ethanol administration depletes mitochondrial DNA in MnSOD-overexpressing transgenic mice, but not in their wild type littermates. *Toxicol Appl Pharmacol.* 2009; 234:326–338. [PubMed: 19063909]
25. Wheeler MD, Nakagami M, Bradford BU, Uesugi T, Mason RP, et al. Overexpression of manganese superoxide dismutase prevents alcohol-induced liver injury in the rat. *J Biol Chem.* 2001; 276:36664–36672. [PubMed: 11477087]
26. Chen K, Craige SE, Keaney JF Jr. Downstream targets and intracellular compartmentalization in Nox signaling. *Antioxid Redox Signal.* 2009; 11:2467–2480. [PubMed: 19309256]
27. Ushio-Fukai M. Compartmentalization of redox signaling through NADPH oxidase-derived ROS. *Antioxid Redox Signal.* 2009; 11:1289–1299. [PubMed: 1899986]
28. Ma Q, Fang H, Shang W, Liu L, Xu Z, et al. Superoxide flashes: early mitochondrial signals for oxidative stress-induced apoptosis. *J Biol Chem.* 2011; 286:27573–27581. [PubMed: 21659534]
29. Wang W, Fang H, Groom L, Cheng A, Zhang W, et al. Superoxide flashes in single mitochondria. *Cell.* 2008; 134:279–290. [PubMed: 18662543]
30. Herlein JA, Fink BD, Henry DM, Yorek MA, Teesch LM, et al. Mitochondrial superoxide and coenzyme Q in insulin-deficient rats: increased electron leak. *Am J Physiol Regul Integr Comp Physiol.* 2011; 301:R1616–R1624. [PubMed: 21940403]
31. Herlein JA, Fink BD, Sivitz WI. Superoxide production by mitochondria of insulin-sensitive tissues: mechanistic differences and effect of early diabetes. *Metabolism.* 2010; 59:247–257. [PubMed: 19765776]
32. Mariappan N, Elks CM, Sriramula S, Guggilam A, Liu Z, et al. NF-kappaB-induced oxidative stress contributes to mitochondrial and cardiac dysfunction in type II diabetes. *Cardiovasc Res.* 2010; 85:473–483. [PubMed: 19729361]
33. Cai L, Li W, Wang G, Guo L, Jiang Y, et al. Hyperglycemia-induced apoptosis in mouse myocardium: mitochondrial cytochrome C-mediated caspase-3 activation pathway. *Diabetes.* 2002; 51:1938–1948. [PubMed: 12031984]
34. Block K, Gorin Y, Abboud HE. Subcellular localization of Nox4 and regulation in diabetes. *Proc Natl Acad Sci U S A.* 2009; 106:14385–14390. [PubMed: 19706525]
35. Rachek LI, Musiyenko SI, LeDoux SP, Wilson GL. Palmitate induced mitochondrial deoxyribonucleic acid damage and apoptosis in l6 rat skeletal muscle cells. *Endocrinology.* 2007; 148:293–299. [PubMed: 17023529]
36. Russell JW, Golovoy D, Vincent AM, Mahendru P, Olzmann JA, et al. High glucose-induced oxidative stress and mitochondrial dysfunction in neurons. *FASEB J.* 2002; 16:1738–1748. [PubMed: 12409316]
37. Medikayala S, Piteo B, Zhao X, Edwards JG. Chronically elevated glucose compromises myocardial mitochondrial DNA integrity by alteration of mitochondrial topoisomerase function. *Am J Physiol Cell Physiol.* 2011; 300:C338–C348. [PubMed: 21123731]
38. Zhang D, Mott JL, Chang SW, Denniger G, Feng Z, et al. Construction of transgenic mice with tissue-specific acceleration of mitochondrial DNA mutagenesis. *Genomics.* 2000; 69:151–161. [PubMed: 11031098]
39. Zhang D, Mott JL, Chang SW, Stevens M, Mikolajczak P, et al. Mitochondrial DNA mutations activate programmed cell survival in the mouse heart. *Am J Physiol Heart Circ Physiol.* 2005; 288:H2476–H2483. [PubMed: 15840907]

40. Hicks S, Labinskyy N, Piteo B, Laurent D, Mathew J, et al. Type II diabetes increases mitochondrial DNA (mtDNA) mutations in the left ventricle of the Goto-Kakizaki diabetic rat. *Am J Physiol Heart Circ Physiol*. 2013
41. Chan DC. Fusion and fission: interlinked processes critical for mitochondrial health. *Annu Rev Genet*. 2012; 46:265–287. [PubMed: 22934639]
42. Lee S, Jeong SY, Lim WC, Kim S, Park YY, et al. Mitochondrial fission and fusion mediators, hFis1 and OPA1, modulate cellular senescence. *J Biol Chem*. 2007; 282:22977–22983. [PubMed: 17545159]
43. Chen L, Gong Q, Stice JP, Knowlton AA. Mitochondrial OPA1, apoptosis, and heart failure. *Cardiovasc Res*. 2009; 84:91–99. [PubMed: 19493956]
44. Dorn GW 2nd, Clark CF, Eschenbacher WH, Kang MY, Engelhard JT, et al. MARF and Opa1 control mitochondrial and cardiac function in *Drosophila*. *Circ Res*. 2011; 108:12–17. [PubMed: 21148429]
45. Ong SB, Subrayan S, Lim SY, Yellon DM, Davidson SM, et al. Inhibiting mitochondrial fission protects the heart against ischemia/reperfusion injury. *Circulation*. 2010; 121:2012–2022. [PubMed: 20421521]