



D-Amino Acids and D-Amino Acid-Containing Peptides: Potential Disease Biomarkers and Therapeutic Targets?

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Abstract: In nature, amino acids are found in two forms, L and D enantiomers, except for glycine which does not have a chiral center. The change of one form to the other will lead to a change in the primary structure of proteins and hence may affect the function and biological activity of proteins. Indeed, several D-amino acid-containing peptides (DAACPs) were isolated from patients with cataracts, Alzheimer's and other diseases. Additionally, significant levels of free D-amino acids were found in several diseases, reflecting the disease conditions. Studying the molecular mechanisms of the DAACPs formation and the alteration in D-amino acids metabolism will certainly assist in understanding these diseases and finding new biomarkers and drug targets. In this review, the presence of DAACPs and free D-amino acids and their links with disease development and progress are summarized. Similarly, we highlight some recent advances in analytical techniques that led to improvement in the discovery and analysis of DAACPs and D-amino acids.

Keywords: D-amino acids; DAACPs; biomarker; therapeutic targets; LC-MS; D-Asp; D-Ser

1. Introduction

Amino acids, the building blocks of peptides and proteins, are found naturally in two forms (L and D enantiomers), except for glycine which does not have a chiral center. L and D enantiomers have different orientation of the four substituents attached to the α -carbon and known as non-superimposed mirror images. The tertiary structure of proteins is critical for their activity, therefore a change in one amino acid stereoisomerism may lead to dramatical change in their physicochemical properties and activity. It was a long-lasting belief that only the L-forms are incorporated in the synthesis of proteins, and this homochirality of L-amino acids in protein synthesis was one of the mysteries in life. Nevertheless, in living organisms, not only free forms of D-amino acids, but also several D-amino acid-containing peptides (DAACPs) have been isolated [1,2]. Among these DAACPs many are biologically active while their counterpart peptides synthesized from only L-amino acids in proteins may result from different mechanisms; non-enzymatic racemization linked with ageing or diseases or by enzymatic post-transnational modification (PTM) for the mRNA-encoded L-amino acids [2].

2. Sources of D-Amino Acids

2.1. Enzymatic Conversion of L-Amino Acids

As L-amino acids are present more predominantly in living organisms, they act as a substrate for D-amino acid synthesis. This conversion reaction occurs in the presence of the enzyme racemase, which changes the stereochemistry of the chiral α -carbon in these



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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). amino acids [4]. Several racemase enzymes have been studied in different organisms, such as alanine racemase which catalyzes the generation of D-Ala, a crucial component of the peptidoglycans, from L-Ala [5,6]. Likewise, glutamate racemase is responsible for the conversion of L-Glu to D-Glu [7]. The presence of these D-isomers is thought to protect the bacteria from the attack of different peptidases [8]. In mammals, serine racemase is the enzyme that inverts the stereochemistry of the α -carbon of L-serine to produce D-serine [9]. Notably, D-Ser, albeit found in low amounts compared to L-Ser, is a potent co-agonist of the N-methyl-D-aspartate (NMDA) glutamate receptor. Moreover, D-Asp and D-Ala are also found in the mammalian brain, and though found in lower amounts than D-serine they are still detectable [10,11]. Additionally, bacteria and other species also synthesized a group of compounds, known as antimicrobial peptides or host defense peptides, for their natural immunity and existence [12]. Since their discovery they have been viewed as an alternative to conventional antibiotics and several are already in clinical use, such as Gramicidin S and Bacitracin. Among the distinctive features of these peptides is the presence of D-amino acids in their structures which would make them more resistant to proteolysis [13]. Furthermore, the incorporation of D-amino acids in some peptides led to improvements in the antibacterial action [14–17]. Nevertheless, the excessive modification of L-amino acids with their D-form counterparts in the structure of the biologically active peptides is linked with potential risks of cytotoxicity and immunogenicity [13]. In addition to antibacterial peptides, there are other drug molecules that contain D-amino acids in their chemical structure, such as the antidiabetic nateglinide and the thrombin inhibitor PPACK [18,19]. The growing interest in the chemical and pharmaceutical industries for pure amino acid enantiomers necessitates establishing an efficient reaction system for their synthesis [20]. Accordingly, different L-amino acid deaminases and D-amino acid oxidases were extensively studied to obtain pure amino acid enantiomers [21-24]. The reader is strongly recommended to refer to Molla et al. and Nakano et al. for more comprehensive information in this point [20,25].

2.2. Spontaneous Inversion of Isomerism

Importantly, not only racemases are responsible for the inversion of L-amino acids into the D-form, but this process can occur spontaneously, despite being very slow, in long-lived proteins and tissues. The reasons behind these changes in stereoisomerism are of non-enzymatical origin, instead, they occur due to chemical reactions that resulted in inversion of the amino acid chirality. Indeed, oxidative stress and free radicals were proved to induce such modifications in proteins and peptides [26,27]. Even though these reactions are very slow in kinetics, nevertheless, they are valuable as a mirror to reflect the age of these proteins and tissues. Among the 19 chiral proteinogenic amino acids, L-Asp is the most exposed to racemization. The inversion of the isomerism of L-Asp mainly happens through a succinimide intermediate. L-succinimide can be formed when the lone pair of electrons of the amino acid adjacent to L-Asp attack the carboxyl group of L-Asp. Then, the formed L-succinimide can be converted to D-succinimide by keto-enol tautomerism. Both L/D-succinimide can be hydrolyzed to form four stereoisomers, L- α -Asp, L- β -Asp, $D-\alpha$ -Asp and $D-\beta$ -Asp (Figure 1). Additionally, L-Asn deamidation will result in the same four isomers of Asp [28]. It is known that L-Asp and L-Asn can be isomerized to D-form when the following amino acid has a small side chain, such as Ala, Gly and Ser.

2.3. Food

Food and dietary products, both natural and processed, contain different amounts of D-amino acids. In fact, several D-amino acids are present in fruits and vegetables such as apples, grapes and tomatoes [5]. Unlike human milk, the milk of cows, goats and ewes contain free D-Ala, D-Asp, D-Glu and D-Ser [5,29]. There have been numerous observations that food processing resulted in stereoisomerism inversion of the proteinogenic L-amino acids and the production of the D-counterparts. Accordingly, many researchers studied the effects of different conditions in food making processes. Several manufacturing processes

are involved in making foods, such as high temperatures, fermentation, strong acid and alkali treatments. Ultimately, this all together resulted in increase in the D-amino acids formation [30]. Of worth noting is that many D-amino acids have a more sweet taste than the L-enantiomers, for example the dipeptide Alitame which contains L-Asp and D-Ala is 200 times sweeter than sucrose [31]. Not surprisingly, this led to the increased use of them as an alternative to sucrose.



Figure 1. Scheme of the spontaneous isomerization of the four isomers of Asp through the succinimide intermediate.

2.4. Microbiota

Another source for the D-amino acids in mammals is microbiota that present in the gut. As mentioned earlier, D-amino acids are important constituents of the peptidoglycans in the bacterial cell wall. The two most common D-amino acids that are found in bacterial cell walls are D-Ala and D-Glu [8]. Nonetheless, in some other bacterial cell walls more D-amino acids have their position in peptidoglycans synthesis, such as D-Ser and D-Asp [32].

3. Catabolism of D-Amino Acids

Interestingly, the presence of the D-amino acid oxidase (DAO) enzyme corrects the amounts of D-amino acids by converting them to imino acid by oxidative deamidation. Although DAO was firstly discovered almost one hundred years ago, its biological importance is only recently well understood, and only after the discovery of D-Ser in human brain [33,34]. Likewise, another enzyme that is accountable for D-Asp degradation, D-aspartate oxidase (DDO), was discovered some time ago. DDO is a peroxisomal flavoen-zyme that oxidizes D-Asp in presence of H₂O and O₂, producing oxaloacetate, H₂O₂, and NH₄⁺. Notably, this enzyme cannot degrade D-Ser [6,35]. DAO and DDO control the levels of D-amino acids in the body to certain levels.

Despite the presence of these enzymes, it bears mentioning that significant increase in the levels of D-amino acids have been found in several disease conditions. Additionally, several DAACPs have been isolated from different tissues in several diseases [36–39]. The significant increase in certain D-amino acids together with the growth in the discovered DAACPs in these diseases surge the consideration of them as potential biomarkers [40–45]. Unveiling the molecular mechanisms behind the formation of DAACPs and the alteration in D-amino acid metabolism will inevitably assist in understanding these diseases and finding new biomarkers and drug targets. This is particularly the case with the recent advances in the analytical techniques that allow for the detection of very small amounts of analytes, and are able to accurately distinguish several compounds having very similar

chemistry. Consequently, this will accelerate the disease discovery and/or finding of new treatments. In this review, we summarize the recent identification and findings of different D-amino acids and DAACPs from several diseases, and their possible role in the disease development and progress. In addition, we highlight the recent advancements in analytical techniques that will eventually lead the way to more discovery of new DAACPs.

4. D-Amino Acids and DAACPs in Selected Diseases

4.1. Alzheimer's Disease

D-Asp and D-Ser have been found in brain tissues isolated from patients with Alzheimer's disease (AD) [2]. This disease is linked with oxidative stress, a situation that arises from the presence of reactive oxygen species (ROS) [46]. AD is as an age-related neurodegenerative disease, and it is the most common cause of dementia in the elderly. Histopathological hallmarks of AD are intracellular neurofibrillary tangles and extracellular formation of senile plaques composed of the amyloid-beta peptide (A β) in aggregated form along with metal-ions such as copper and other metals. Copper can catalyze the production of ROS, such as hydroxyl radical, when bound to the A β . The produced hydroxyl radical and other ROS may result in the oxidative damage on both the AB peptide itself and other proteins in the brain. Another hallmark of AD is the presence of intracellular neurofibrillary tangles, which are composed of hyperphosphorylated Tau protein [47–49]. The A β has 38 to 42 amino acids, and this peptide is formed from the amyloid β -protein precursor by the sequential cutting actions of β - and γ -secretases [50]. The truncated and toxic fragment A β 25–35/40 which contains D-Ser at position 26 is believed to damage neurons and may account for the neurodegeneration in AD [51]. Moreover, D-serine levels in both blood and cerebrospinal fluid are found significantly higher in AD patients; as a result D-serine is believed to be responsible for controlling the extent of NMDAR-mediated neurotoxic changes that lead to AD [52–55]. However, these findings are contradicted by other researchers and further studies are therefore needed [56]. A recently published study suggested elevated levels of D-Ser could predict worse symptoms of memory decline, in particular the domains for word recall task, orientation, comprehensive and word-finding difficulty [57]. The reader is encouraged to refer to an excellent review for the roles of D-Ser in brain [58]. D-Asp also found its place in the investigation of the possible roles of D-amino acids in the brain and the links with some CNS dysfunctions. Free D-Asp could have a role in the development and maturation of tissues and organs in mammals. Indeed, D-Asp is found in the endocrine glands and its levels increase after birth as the maturation of these organs proceed. On the contrary, in the brain, free D-Asp amounts strongly decline following birth, and this is recognized as a consequence of the postnatal of DDO. In the brain, D-Asp is found mostly concentrated in the synaptic vesicles of terminal axon, thus proposing its function as an endogenous neurotransmitter [59,60]. A significant decrease in the amounts of D-Asp has been identified in the prefrontal cortex of patients with schizophrenia, this was accompanied with an elevated expression of DDO mRNA [61]. As highlighted earlier, incorporation of D-amino acid (such as D-Asp) in proteins produces different side chain orientation. Eventually, this results in changes in the secondary and tertiary structure, as well as the quaternary arrangement of a protein, which leads to dysfunctional proteins [62,63]. Briefly, Asp isomerization might lead to abnormal aggregation and degradation and induce partial protein unfolding, such as the A β aggregations that characterizes AD. Certainly, the racemization of Asp at position 23 accelerated the peptide aggregation and fibril formation [64]. Of final note is that there are many neuropeptides isolated from different organisms that contain D-isomer in their chains and the discovered number is rising. More importantly, theses neuropeptides have potent biological activities compared to those with all L-amino acids [65,66]. Dermorphin represents an excellent example of such neuropeptides. This heptapeptide has a D-Ala in the second position and is superior to morphine [67].

4.2. Cataract

Different PTMs have been characterized in the proteins of human eyes, include phosphorylation, Cys and Met oxidation, methylation and racemization. The latter is thought to be more closely related to the development of cataracts in eyes compared to others [68]. Crystallins are important structural and functional proteins in the human lens, and they have pivotal role in lens transparency. In the lens there are three types of crystallins, assigned as α -, β -, and γ -crystallins. For the lens to remain transparent, these proteins should retain their structures, as accumulative precipitation of lens proteins eventually will result in cataracts [69,70]. Several D-amino acids have been identified in human lenses including D-Asp, D-Ser, D-Glu/Gln and D-Phe. Moreover, the amount of the racemization to D-Ser and D-Asp was significantly higher in cataract lenses than in age-matched normal lenses. Indeed, cataract lens proteins from patients at 40 years old showed comparable racemization of Asp as lenses of 80-year-old normal people [69]. Not surprisingly, Asp racemization in lenses is the most studied among the others as it is the most susceptible amino acid for inversion in chirality. Asp racemization was identified in the two subunits of α -crystallins, named α A-crystallins and α B-crystallins, and such modification in amino acids chirality might disrupt the polymeric state of α -crystallins. In α A-crystallins, different Asp positions were found isomerized including Asp58, Asp84 and Asp151. Unavoidably, this results in decreased solubility and impairs the function of this important component of the lens proteins, contributing to the development of cataracts [68,71,72]. Likewise, in another study, Asp4 and Asp96 of α B-crystallin and Asp37 of β A3-crystallin were found isomerized to the D-form. These findings may also specify the possibility that the inversion of Asp residues could lead to the dissociation of α B- and β A3-crystallins from the polymeric and oligometric states [70,73]. Finally, the quantification of aspartic acid racemization in eye proteins has importance in forensic science and can be used as a measure of chronological aging in humans and other organisms [74,75].

4.3. Chronic Kidney Disease

The presence of D-amino acids was intermittently reported in patients with kidney diseases, though recent studies unraveled the clinical significance of D-amino acids in chronic kidney diseases (CKD) [40,76]. CKD is well recognized globally as a significant clinical and public health problem and it is estimated to affect almost one billion humans [41]. The common pathological alteration in progressive CKD is tubulointerstitial fibrosis, which involves various independent and overlapping cellular and molecular pathways leading to reduction in the glomerular filtration ratio (GFR). The disease has high morbidity and mortality ratios, and end-stage patients need careful monitoring of diet, as dietary contents and their metabolites are linked to the disease progression [77,78]. A core issue in the management of CKD is the early diagnosis of kidney diseases. In this regard, the levels of some D-amino acids are identified to be in correlation with GFR. Indeed, the plasma D-Ser level was associated with creatinine-based estimated GFR, and this finding was thought to offer new biomarkers to reflect kidney function as the currently available kidney markers are affected by other factors such as muscular mass [76,79,80]. Notably, D-Ser is not the only D-amino acid that is associated with kidney diseases, indeed, D-Asn, D-Ala and D-Pro all have significant levels in patients with kidney diseases in comparison with healthy people. In essence, the levels of D-Pro and D-Asn were found to have a significant correlation with kidney function (estimated GFR). Moreover, the levels of D-Ala and D-Pro correlated well with patients age, while D-Asp and D-Pro were associated with co-presence of diabetes [40]. Recently, researchers successfully developed a three-dimensional HPLC method to detect the trace amounts of D-amino acids in patients with CKD and the correlation with GFR. Good association was determined between kidney function and the levels of D-Ser and D-Asn in particular [41].

4.4. Cancer

The possible correlation between D-amino acids and cancer was controversial for a period of time. The first report for the potential link between D-amino acids and cancer was published almost 80 years ago by Kögl and Erxleben. They reported the detection of some D-amino acids, in particular D-Glu, in tumor cell proteins, and concluded that the tumor development was dependent on the presence of these D-amino acids [81]. However, these findings were contraindicated later by many researchers as summarized in this review [82]. It is worthy to note the technical difficulties at that time to detect the presence of trace amounts of D-amino acids. This controversial argument continued for a while as some reports revealed insignificant differences of D-amino acids levels between healthy patients and patients with tumors, while others concluded no differences [83,84]. In gastric cancer patients who are also helicobacter pylori-positive there were high levels of D-Ala in their gastric juice, and this observation influenced other researchers to develop a non-invasive test to detect the presence of these D-amino acids in saliva and enhance the early detection of gastric cancer [84,85]. In contrast, the levels of D-Glu and D-Gln were lower in patients with hepatocellular carcinoma compared to healthy individuals [86]. One can conclude that D-amino acids levels could be different from one tumor condition to the other and there is no universal assumption. Truly, the altered amino acid profiles, particularly D-amino acids, may have great potential in the early detection of some tumors and could be recognized as potential oncometabolites [87-89]. Indeed, the levels of D-Ser and D-Asp were significantly higher in MCF-7 cancer cells compared to non-tumorigenic MCF-10A epithelial cells. The elevation of these D-amino acids could be a result of the upregulation of the enzyme racemases. These findings suggest possible roles of these D-amino acids in breast cancer proliferation and thus consider them as potential oncometabolites for breast cancer [87]. Although only D-Ser racemase has been identified in humans and responsible for D-Ser formation, a recent report suggests a role of this enzyme in the formation of D-Asp as well [90]. The reader is advised to a recently published excellent review on the possible roles of D-amino acids as cancer biomarkers and treatment targets [91].

5. Recent Advancement in Separation Techniques and the Impact on the Detections of D-Amino Acids and DAACPs

Generally speaking, D-amino acids and DAACPs are present in trace amounts in living organisms, which makes their identification and quantification a challenging task. In multiomics approaches such as proteomics and metabolomics, mass spectrometry (MS) plays a core role in the analysis of different proteins and peptides. MS analysis requires only limited amounts of the analytes, and it can also be coupled online with separation techniques such as liquid chromatography (LC) and capillary electrophoresis (CE). The availability of deconvolution software and improvements in bioinformatics made MS the main analytical technique for peptide sequencing and protein analysis, particularly when dealing with samples in complex mixtures. Truly, MS techniques were successful in determining many PTMs in proteins such as phosphorylation, acetylation and others [92,93]. However, the modification of amino acids in a peptide to the D-form will not result in a change in the m/z values, which challenges the MS detection of such modifications. Nonetheless, MS is capable of detecting the presence of these peptides, albeit with some difficulties. The physicochemical properties of L/D-forms of an amino acid are similar, it is only their reflection of the polarized light which is different. Therefore, in order to separate them, for example by LC, a derivatization step is first required with some chiral molecules and the detection with chiral chromatography. In contrast, diastereomeric peptides, where the chirality of one (or more) amino acid is inverted, have different physicochemical properties which permit their separation by LC without being derivatized. However, the fragmentation pattern in MS will likely be similar, and as a result the differentiation will be challenging. In fact, the Sweedler group proposed a detailed approach for the untargeted discovery of DAACPs, combining enzymatic screening, chiral amino acid analysis and LC–MS. By using this protocol, they were able to detect the presence of a novel bioactive



Figure 2. LC–MS analysis of DAACP isolated from *Aplysia californica*, the retention times of the indigenous (**A**) and synthetic DAACP (**C**) are identical and are different from the peptides with all L-amino acids (**B**). However, the MS profiles of the three peptides are similar. Reproduced with permission from [94].

Consequently, the presence of indigenous DAACPs could be missed if not compared with the previously synthesized one. Therefore, unless one has the synthetic peptides to compare the retention times with, one might miss the detection of the indigenous DAACPs. This raises a question of how many DAACPs have not been detected and instead were recognized as their all L-amino acids counterparts. What is more difficult is pointing to the specific site of the isomerization, as it is not practical to synthesize all possible DAACPs to compare the retention times. However, some fragmentation techniques such as electron-capture dissociation (ECD) and electron-transfer dissociation (ETD) produce unique fragmentation patterns when α -Asp or β -Asp is present [96–98]. Additionally, radical-directed dissociation (RDD) fragmentation was successful in discriminating DAACPs epimers as it is sensitive to the structure of the ions being fragmented. Certainly, RDD fragmentation was better than collision-induced dissociation (CID) in discriminating biologically-active peptides containing D-amino acids [99]. A comprehensive analysis protocol for differentiation of peptide isomers and epimers of lens crystallin using RDD was recently published [100]. Furthermore, the presence of the four Asp isomers in protein and peptides was identified using LC–MS, and three commercial enzymes without the need for synthesized reference peptides. The protein was first hydrolyzed by trypsin, then the tryptic peptides were further hydrolyzed by either Asp-N, Protein L-isoaspartyl methyltransferase (PIMT), or paenidase. These three enzymes hydrolyze the peptides that have L- α -Asp, L- β -Asp and D- α -Asp, respectively. Indeed, this method succeeded to identify all Asp isomers in tryptic peptides of aged lens proteins [43,101]. Notably, the presence of a D-amino acid in a peptide can resist the hydrolysis by certain proteases, and this fact was used to detect the presence of DAACPs. One of the enzymes used for this purpose is Aminopeptidase M (AMP) which hydrolyzes the peptide from the N-terminus. However, AMP digestion stops because of steric hinderance when it reaches a D-amino acid, and in fact some peptides can resist AMP digestion for several days. Nonetheless, the presence of Proline also resists AMP digestion, which will produce false positive results [65,94]. The current advancements in ion mobility spectrometry (IMS) make it an excellent choice for isomeric biomolecules separation [102]. Despite that the theory and principles of IMS were

stablished several decades ago, fundamental contributions only started very recently in omics sciences [103]. Moreover, as the coupling of IMS and MS will provide an additional separation dimension, this will allow the detection of isomeric and isobaric molecules in complex mixtures. Eventually, this will lead to greater understanding and characterization of the properties of biomolecules such as proteins and metabolites [104,105]. IMS measures the arrival times of ions in the gaseous phase under applied electric field and vacuum or atmospheric pressure. Ion mobility under these conditions is affected by size, charge and shape of the ions. Consequently, this technique separates molecules in the gaseous phase based on the differences of their mass to charge and shape [104]. Hence, the arrangement of the functional groups is different in DAACPs; therefore, this will change the cross-sectional area of the molecule, and as a result the arrival time of ions will be changed. Subsequently, IMS will permit the direct determination of the site where isomerization takes place (Figure 3). Indeed, a recently published method was successful in discriminating isomeric peptides and instantaneously pointed to the site of epimerization by combining LC–MS and IMS [106,107].



Figure 3. Identification of the epimerization site with LC–MS IMS by comparing the arrival times of the ions of deltorphin having L-Ala (labeled L-DTP) or D-Ala (labeled D-DTP). (**A**,**B**) LC–MS separation shows similar MS profiles and different retention times, indicating that they are isomeric peptides. (**C**) A shift in arrival time is detected for the L/D-forms. (**D**,**E**) Following IMS fragmentation, the arrival times of y_2^+ , y_3^+ , y_4^+ and y_5^+ are similar in both L-DTP and D-DTP, while the arrival times of the y_6^+ ions are different, therefore determining the epimerization site. *, interference ions. Reproduced with permission from [107].

Additionally, a recently developed IMS method was able to baseline separate four A β 17-28 tryptic peptide epimers on a rapid time scale [108]. In this study, sodium adduct ions, [M + H + Na]²⁺, allowed the complete separation of all A β epimer sets assessed, which were unachievable for their [M + 2H]²⁺ doubly protonated ions. IMS coupled with MS was also applied for the detection of free D-amino acids after derivatization with a chiral reagent to diastereomers. This novel analytical method was able to detect D-amino acids in the nanomolar range, and the analysis time including the derivatization step was less than 15 min [109]. Notably, several chiral derivatization reagents were used for the chromatographic separations of D-amino acids; however, Marfey's reagent is the mostly

used one [110–113]. Recently, a pair of stereodynamic chiral benzylicaldehyde probes were developed for the determination of amino acid configuration in peptides by MS. This method was accurate and the obtained results were well correlated when compared with Marfey's derivatization methods, without the need for tedious and time-consuming separation steps [114]. It is worth mentioning that not only the analytical techniques are used for the detection and quantification of D-amino acids. Additionally, there are reported protocols for the detection of D-amino acids by enzymatic assay methods, and the most exploited enzyme in these assays is D-amino oxidase. These enzymatic assays provide the advantages of high sensitivity and specificity which allow the development of low-cost protocols and avoid the tedious and time-consuming steps in the analytical techniques [115]. Moreover, there is ongoing research to develop enzymatic biosensors for the rapid detection of D-amino acids [116]. Eventually, several biosensors were fabricated for the detection of D-amino acids in biological samples, and they showed rapid response, good sensitivity and high recovery of the added D-amino acids [117,118]. Pundir et al. and Rosini et al. published excellent reviews on the use of biosensors for the detection of D-amino acids [116,119]. Similarly, for more deep details in the recent progress of the approaches for the detection of isomerism in peptides and proteins, see the review written by Erik T. Jansson [120]. In addition, the research on detecting these potential disease biomarkers is ongoing, and more lab-on-chip devices were developed or are on the road for disease detection [119,121].

6. Conclusions

The tertiary structure of proteins is crucial for their functions and activity, in the way that minor changes in the amino acid sequence may lead to significant influence on the activity. Homochirality in protein synthesis is one of life's mysteries, yet the presence of DAACPs in different tissues represent a new insight to unravel several molecular mechanisms in these tissues. The levels of some free D-amino acids in body are found to be linked with many diseases. Consequently, the presence of DAACPs and D-amino acids in significant levels in some disease conditions will make them potential biomarkers and therapeutic targets for those diseases. In the human body, most cells are replaced after a certain period of time; however, some cells of relatively long life spans, such as neurons, are more likely to accumulate more DAACPs. Though DAACPs and D-amino acids are present in trace amounts, the recent advances in analytical techniques permit more accurate analysis of them. Subsequently, this will assist in better understanding on DAACPs and D-amino acids and unravel their association with disease development and progress.

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References

- Grishin, D.V.; Zhdanov, D.D.; Pokrovskaya, M.V.; Sokolov, N.N. D-amino acids in nature, agriculture and biomedicine. *Front. Life* Sci. 2020, 13, 11–22. [CrossRef]
- Ollivaux, C.; Soyez, D.; Toullec, J.Y. Biogenesis of D-amino acid containing peptides/proteins: Where, when and how? J. Pept. Sci. 2014, 20, 595–612. [CrossRef] [PubMed]

- Koehbach, J.; Gruber, C.W.; Becker, C.; Kreil, D.P.; Jilek, A. MALDI TOF/TOF-Based Approach for the Identification of d- Amino Acids in Biologically Active Peptides and Proteins. J. Proteome Res. 2016, 15, 1487–1496. [CrossRef] [PubMed]
- 4. Yoshimura, T.; Esaki, N. Amino acid racemases: Functions and mechanisms. J. Biosci. Bioeng. 2003, 96, 103–109. [CrossRef]
- 5. Genchi, G. An overview on D-amino acids. Amino Acids 2017, 49, 1521–1533. [CrossRef] [PubMed]
- 6. Sasabe, J.; Suzuki, M. Emerging Role of D-Amino Acid Metabolism in the Innate Defense. Front. Microbiol. 2018, 9, 933. [CrossRef]
- Choi, S.Y.; Esaki, N.; Yoshimura, T.; Soda, K. Raction mechanism of Glutamte Racemase, A Pyridoxal Phosphate-independent Amino-acid Racemase. J. Biochem. 1992, 112, 139–142. [CrossRef] [PubMed]
- 8. Cava, F.; Lam, H.; de Pedro, M.A.; Waldor, M.K. Emerging knowledge of regulatory roles of d-amino acids in bacteria. *Cell. Mol. Life Sci.* 2011, *68*, 817–831. [CrossRef] [PubMed]
- 9. Wolosker, H.; Sheth, K.N.; Takahashi, M.; Mothet, J.P.; Brady, R.O.; Ferris, C.D.; Snyder, S.H. Purification of serine racemase: Biosynthesis of the neuromodulator D-serine. *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 721–725. [CrossRef]
- 10. MacKay, M.B.; Kravtsenyuk, M.; Thomas, R.; Mitchell, N.D.; Dursun, S.M.; Baker, G.B. D-Serine: Potential Therapeutic Agent and/or Biomarker in Schizophrenia and Depression? *Front. Psychiatry* **2019**, *10*, 25. [CrossRef]
- 11. Billard, J.-M. D-Amino acids in brain neurotransmission and synaptic plasticity. Amino Acids 2012, 43, 1851–1860. [CrossRef]
- 12. Li, S.; Wang, Y.; Xue, Z.; Jia, Y.; Li, R.; He, C.; Chen, H. The structure-mechanism relationship and mode of actions of antimicrobial peptides: A review. *Trends Food Sci. Technol.* **2021**, *109*, 103–115. [CrossRef]
- 13. Lu, J.; Xu, H.; Xia, J.; Ma, J.; Xu, J.; Li, Y.; Feng, J. D-and unnatural amino acid substituted antimicrobial peptides with improved proteolytic resistance and their proteolytic degradation characteristics. *Front. Microbiol.* **2020**, *11*, 30. [CrossRef]
- 14. Wongkaewkhiaw, S.; Taweechaisupapong, S.; Anutrakunchai, C.; Nazmi, K.; Bolscher, J.G.; Wongratanacheewin, S.; Kanthawong, S.J.B. D-LL-31 in combination with ceftazidime synergistically enhances bactericidal activity and biofilm destruction in *Burkholderia pseudomallei*. *Biofouling* **2019**, *35*, 573–584. [CrossRef]
- Kim, E.Y.; Rajasekaran, G.; Shin, S.Y. LL-37-derived short antimicrobial peptide KR-12-a5 and its d-amino acid substituted analogs with cell selectivity, anti-biofilm activity, synergistic effect with conventional antibiotics, and anti-inflammatory activity. *Eur. J. Med. Chem.* 2017, 136, 428–441. [CrossRef] [PubMed]
- 16. Silva, T.; Magalhães, B.; Maia, S.; Gomes, P.; Nazmi, K.; Bolscher, J.G.; Rodrigues, P.N.; Bastos, M.; Gomes, M.S. Killing of Mycobacterium avium by lactoferricin peptides: Improved activity of arginine-and D-amino-acid-containing molecules. *Antimicrob. Agents Chemother.* **2014**, *58*, 3461–3467. [CrossRef]
- 17. Jia, F.; Wang, J.; Peng, J.; Zhao, P.; Kong, Z.; Wang, K.; Yan, W.; Wang, R. D-amino acid substitution enhances the stability of antimicrobial peptide polybia-CP. *Acta Biochim. Biophys. Sin.* **2017**, *49*, 916–925. [CrossRef] [PubMed]
- 18. Zhang, D.; Jing, X.; Zhang, W.; Nie, Y.; Xu, Y. Highly selective synthesis of D-amino acids from readily available L-amino acids by a one-pot biocatalytic stereoinversion cascade. *RSC Adv.* **2019**, *9*, 29927–29935. [CrossRef]
- Palekar, R.U.; Jallouk, A.P.; Myerson, J.W.; Pan, H.; Wickline, S.A.J.A. Inhibition of thrombin with PPACK-nanoparticles restores disrupted endothelial barriers and attenuates thrombotic risk in experimental atherosclerosis. *Arterioscler. Thromb. Vasc. Biol.* 2016, 36, 446–455. [CrossRef]
- 20. Molla, G.; Melis, R.; Pollegioni, L. Breaking the mirror: L-amino acid deaminase, a novel stereoselective biocatalyst. *Biotechnol. Adv.* **2017**, *35*, 657–668. [CrossRef]
- 21. Nakano, S.; Minamino, Y.; Hasebe, F.; Ito, S. Deracemization and stereoinversion to aromatic d-amino acid derivatives with ancestral l-amino acid oxidase. *ACS Catal.* **2019**, *9*, 10152–10158. [CrossRef]
- 22. Rosini, E.; Melis, R.; Molla, G.; Tessaro, D.; Pollegioni, L. Deracemization and Stereoinversion of α-Amino Acids by l-Amino Acid Deaminase. *Adv. Synth. Catal.* **2017**, *359*, *3773–3781*. [CrossRef]
- 23. Parmeggiani, F.; Lovelock, S.L.; Weise, N.J.; Ahmed, S.T.; Turner, N.J. Synthesis of D-and l-phenylalanine derivatives by phenylalanine ammonia lyases: A multienzymatic cascade process. *Angew. Chem.* **2015**, 127, 4691–4694. [CrossRef]
- 24. Pollegioni, L.; Motta, P.; Molla, G. l-Amino acid oxidase as biocatalyst: A dream too far? *Appl. Microbiol. Biotechnol.* 2013, 97, 9323–9341. [CrossRef] [PubMed]
- 25. Nakano, S.; Kozuka, K.; Minamino, Y.; Karasuda, H.; Hasebe, F.; Ito, S. Ancestral L-amino acid oxidases for deracemization and stereoinversion of amino acids. *Commun. Chem.* **2020**, *3*, 1–11. [CrossRef]
- 26. Sheykhkarimli, D.; Choo, K.L.; Owen, M.; Fiser, B.; Jojart, B.; Csizmadia, I.G.; Viskolcz, B. Molecular ageing: Free radical initiated epimerization of thymopentin—A case study. *J. Chem. Phys.* **2014**, *140*, 8. [CrossRef]
- 27. Kuge, K.; Kitamura, K.; Nakaoji, K.; Hamada, K.; Fujii, N.; Saito, T.; Fujii, N. Oxidative Stress Induces the Formation of D-Aspartyl Residues in the Elastin Mimic Peptides. *Chem. Biodivers.* **2010**, *7*, 1408–1412. [CrossRef] [PubMed]
- 28. Geiger, T.; Clarke, S. Deamidation, isomerization, and racemization at asparaginyl and aspartyl residues in peptides. Succinimidelinked reactions that contribute to protein degradation. *J. Biol. Chem.* **1987**, *262*, 785–794. [CrossRef]
- 29. Tian, H.; Zheng, N.; Li, S.; Zhang, Y.; Zhao, S.; Wen, F.; Wang, J. Characterization of chiral amino acids from different milk origins using ultra-performance liquid chromatography coupled to ion-mobility mass spectrometry. *Sci. Rep.* 2017, *7*, 46289. [CrossRef]
- 30. Cartus, A.T. 12—d-Amino acids and cross-linked amino acids as food contaminants. In *Chemical Contaminants and Residues in Food*; Schrenk, D., Ed.; Woodhead Publishing: Sawston, UK, 2012; pp. 286–319.
- Chattopadhyay, S.; Raychaudhuri, U.; Chakraborty, R. Artificial sweeteners—A review. J. Food Sci. Technol. 2014, 51, 611–621. [CrossRef] [PubMed]

- Veiga, P.; Piquet, S.; Maisons, A.; Furlan, S.; Courtin, P.; Chapot-Chartier, M.-P.; Kulakauskas, S. Identification of an essential gene responsible for D-Asp incorporation in the Lactococcus lactis peptidoglycan crossbridge. *Mol. Microbiol.* 2006, 62, 1713–1724. [CrossRef] [PubMed]
- 33. Kondori, N.R.; Paul, P.; Robbins, J.P.; Liu, K.; Hildyard, J.C.W.; Wells, D.J.; de Belleroche, J.S. Focus on the Role of D-serine and D-amino Acid Oxidase in Amyotrophic Lateral Sclerosis/Motor Neuron Disease (ALS). *Front. Mol. Biosci.* 2018, *5*, 8. [CrossRef]
- Sasabe, J.; Suzuki, M. Distinctive Roles of D-Amino Acids in the Homochiral World: Chirality of Amino Acids Modulates Mammalian Physiology and Pathology. *Keio J. Med.* 2019, 68, 1–16. [CrossRef]
- 35. Errico, F.; Nuzzo, T.; Carella, M.; Bertolino, A.; Usiello, A. The Emerging Role of Altered D-Aspartate Metabolism in Schizophrenia: New Insights from Preclinical Models and Human Studies. *Front. Psychiatry* **2018**, *9*, 559. [CrossRef]
- Fujii, N.; Tajima, S.; Tanaka, N.; Fujimoto, N.; Takata, T.; Shimo-Oka, T. The presence of D-beta-aspartic acid-containing peptides in elastic fibers of sun-damaged skin: A potent marker for ultraviolet-induced skin aging. *Biochem. Biophys. Res. Commun.* 2002, 294, 1047–1051. [CrossRef]
- Fujii, N.; Takata, T.; Fujii, N.; Aki, K.; Sakaue, H. D-Amino acids in protein: The mirror of life as a molecular index of aging. Biochim. Biophys. Acta-Proteins Proteom. 2018, 1866, 840–847. [CrossRef] [PubMed]
- Sivan, S.S.; Van El, B.; Merkher, Y.; Schmelzer, C.E.H.; Zuurmond, A.M.; Heinz, A.; Wachtel, E.; Varga, P.P.; Lazary, A.; Brayda-Bruno, M.; et al. Longevity of elastin in human intervertebral disc as probed by the racemization of aspartic acid. *Biochim. Biophys. Acta-Gen. Subj.* 2012, 1820, 1671–1677. [CrossRef]
- 39. Powell, J.T.; Vine, N.; Crossman, M. On the accumulation of d-aspartate in elastin and other proteins of the aging aorta. *Atherosclerosis* **1992**, *97*, 201–208. [CrossRef]
- 40. Kimura, T.; Hamase, K.; Miyoshi, Y.; Yamamoto, R.; Yasuda, K.; Mita, M.; Rakugi, H.; Hayashi, T.; Isaka, Y. Chiral amino acid metabolomics for novel biomarker screening in the prognosis of chronic kidney disease. *Sci. Rep.* **2016**, *6*, 26137. [CrossRef]
- Furusho, A.; Koga, R.; Akita, T.; Mita, M.; Kimura, T.; Hamase, K. Three-Dimensional High-Performance Liquid Chromatographic Determination of Asn, Ser, Ala, and Pro Enantiomers in the Plasma of Patients with Chronic Kidney Disease. *Anal. Chem.* 2019, 91, 11569–11575. [CrossRef]
- 42. Ha, S.; Kim, I.; Takata, T.; Kinouchi, T.; Isoyama, M.; Suzuki, M.; Fujii, N. Identification of D-amino acid-containing peptides in human serum. *PLoS ONE* 2017, 12, e0189972. [CrossRef]
- 43. Maeda, H.; Takata, T.; Fujii, N.; Sakaue, H.; Nirasawa, S.; Takahashi, S.; Sasaki, H.; Fujii, N. Rapid survey of four Asp isomers in disease-related proteins by LC-MS combined with commercial enzymes. *Anal. Chem.* **2015**, *87*, 561–568. [CrossRef] [PubMed]
- 44. Ishii, C.; Akita, T.; Mita, M.; Ide, T.; Hamase, K. Development of an online two-dimensional high-performance liquid chromatographic system in combination with tandem mass spectrometric detection for enantiomeric analysis of free amino acids in human physiological fluid. *J. Chromatogr. A* **2018**, 1570, 91–98. [CrossRef]
- 45. Miyamoto, T.; Homma, H. Detection and quantification of d-amino acid residues in peptides and proteins using acid hydrolysis. *Biochim. Biophys. Acta Proteins Proteom.* **2018**, *1866*, 775–782. [CrossRef] [PubMed]
- 46. Cheignon, C.; Tomas, M.; Bonnefont-Rousselot, D.; Faller, P.; Hureau, C.; Collin, F. Oxidative stress and the amyloid beta peptide in Alzheimer's disease. *Redox Biol.* **2018**, *14*, 450–464. [CrossRef]
- 47. Goedert, M. Alzheimer's and Parkinson's diseases: The prion concept in relation to assembled A beta, tau, and alpha-synuclein. *Science* **2015**, *349*, 1255555. [CrossRef] [PubMed]
- 48. Livingston, G.; Sommerlad, A.; Orgeta, V.; Costafreda, S.G.; Huntley, J.; Ames, D.; Ballard, C.; Banerjee, S.; Burns, A.; Cohen-Mansfield, J.; et al. Dementia prevention, intervention, and care. *Lancet* **2017**, *390*, 2673–2734. [CrossRef]
- Jack, C.R., Jr.; Bennett, D.A.; Blennow, K.; Carrillo, M.C.; Dunn, B.; Haeberlein, S.B.; Holtzman, D.M.; Jagust, W.; Jessen, F.; Karlawish, J.; et al. NIA-AA Research Framework: Toward a biological definition of Alzheimer's disease. *Alzheimers Dement*. 2018, 14, 535–562. [CrossRef] [PubMed]
- 50. Atri, A. Current and Future Treatments in Alzheimer's Disease. Semin. Neurol. 2019, 39, 227-240. [CrossRef]
- Kubo, T.; Nishimura, S.; Kumagae, Y.; Kaneko, I. In vivo conversion of racemized beta-amyloid ([D-Ser 26]A beta 1-40) to truncated and toxic fragments ([D-Ser 26]A beta 25-35/40) and fragment presence in the brains of Alzheimer's patients. *J. Neurosci. Res.* 2002, 70, 474–483. [CrossRef]
- Hashimoto, K.; Fukushima, T.; Shimizu, E.; Okada, S.I.; Komatsu, N.; Okamura, N.; Koike, K.; Koizumi, H.; Kumakiri, C.; Imai, K.; et al. Possible role of D-serine in the pathophysiology of Alzheimer's disease. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* 2004, *28*, 385–388. [CrossRef] [PubMed]
- 53. Danysz, W.; Parsons, C.G. Alzheimer's disease, beta-amyloid, glutamate, NMDA receptors and memantine—Searching for the connections. *Br. J. Pharmacol.* **2012**, *167*, 324–352. [CrossRef] [PubMed]
- Madeira, C.; Lourenco, M.V.; Vargas-Lopes, C.; Suemoto, C.K.; Brandao, C.O.; Reis, T.; Leite, R.E.P.; Laks, J.; Jacob-Filho, W.; Pasqualucci, C.A.; et al. D-serine levels in Alzheimer's disease: Implications for novel biomarker development. *Transl. Psychiatry* 2015, 5, e564. [CrossRef]
- 55. Lin, C.-H.; Yang, H.-T.; Chiu, C.-C.; Lane, H.-Y. Blood levels of D-amino acid oxidase vs. D-amino acids in reflecting cognitive aging. *Sci. Rep.* 2017, 7, 14849. [CrossRef]
- 56. Biemans, E.A.L.M.; Verhoeven-Duif, N.M.; Gerrits, J.; Claassen, J.A.H.R.; Kuiperij, H.B.; Verbeek, M.M. CSF D-serine concentrations are similar in Alzheimer's disease, other dementias, and elderly controls. *Neurobiol. Aging* **2016**, *42*, 213–216. [CrossRef]

- 57. Lin, C.H.; Yang, H.T.; Lane, H.Y. D-glutamate, D-serine, and D-alanine differ in their roles in cognitive decline in patients with Alzheimer's disease or mild cognitive impairment. *Pharm. Biochem. Behav.* **2019**, *185*, 172760. [CrossRef] [PubMed]
- 58. Mothet, J.-P.; Billard, J.-M.; Pollegioni, L.; Coyle, J.T.; Sweedler, J.V. Investigating brain d-serine: Advocacy for good practices. *Acta Physiol.* **2019**, *226*, e13257. [CrossRef] [PubMed]
- 59. D'Aniello, A. D-Aspartic acid: An endogenous amino acid with an important neuroendocrine role. *Brain Res. Rev.* 2007, 53, 215–234. [CrossRef]
- 60. Fontanarosa, C.; Pane, F.; Sepe, N.; Pinto, G.; Trifuoggi, M.; Squillace, M.; Errico, F.; Usiello, A.; Pucci, P.; Amoresano, A. Quantitative determination of free D-Asp, L-Asp and N-methyl-D-aspartate in mouse brain tissues by chiral separation and Multiple Reaction Monitoring tandem mass spectrometry. *PLoS ONE* **2017**, *12*, e0179748. [CrossRef]
- Errico, F.; Napolitano, F.; Squillace, M.; Vitucci, D.; Blasi, G.; de Bartolomeis, A.; Bertolino, A.; D'Aniello, A.; Usiello, A. Decreased levels of D-aspartate and NMDA in the prefrontal cortex and striatum of patients with schizophrenia. *J. Psychiatry Res.* 2013, 47, 1432–1437. [CrossRef]
- 62. Sakaue, H.; Kinouchi, T.; Fujii, N.; Takata, T.; Fujii, N. Isomeric Replacement of a Single Aspartic Acid Induces a Marked Change in Protein Function: The Example of Ribonuclease A. *ACS Omega* **2017**, *2*, 260–267. [CrossRef]
- 63. Fujii, N.; Takata, T.; Fujii, N. Quantitative analysis of isomeric (l-alpha-, l-beta-, D-alpha-, D-beta-) aspartyl residues in proteins from elderly donors. *J. Pharm. Biomed. Anal.* **2015**, *116*, 25–33. [CrossRef] [PubMed]
- 64. Tomiyama, T.; Asano, S.; Furiya, Y.; Shirasawa, T.; Endo, N.; Mori, H. Racemization of ASP(23) residue affects the aggregation properties of alzheimer amyloid-beta protein analogs. *J. Biol. Chem.* **1994**, *269*, 10205–10208. [CrossRef]
- Ewing, M.A.; Wang, J.; Sheeley, S.A.; Sweedler, J.V. Detecting D-Amino acid-containing neuropeptides using selective enzymatic digestion. *Anal. Chem.* 2008, *80*, 2874–2880. [CrossRef]
- 66. Bai, L.; Romanova, E.V.; Sweedler, J.V. Distinguishing Endogenous D-Amino Acid-Containing Neuropeptides in Individual Neurons Using Tandem Mass Spectrometry. *Anal. Chem.* **2011**, *83*, 2794–2800. [CrossRef]
- 67. Hesselink, J.M.K.; Schatman, M.E. Rediscovery of old drugs: The forgotten case of dermorphin for postoperative pain and palliation. *J. Pain Res.* **2018**, *11*, 2991–2995. [CrossRef]
- 68. Zhu, X.J.; Zhang, K.K.; He, W.W.; Du, Y.; Hooi, M.; Lu, Y. Racemization at the Asp 58 residue in alphaA-crystallin from the lens of high myopic cataract patients. *J. Cell Mol. Med.* **2018**, *22*, 1118–1126. [CrossRef]
- 69. Hooi, M.Y.S.; Truscott, R.J.W. Racemisation and human cataract. d-Ser, d-Asp/Asn and d-Thr are higher in the lifelong proteins of cataract lenses than in age-matched normal lenses. *Age* **2011**, *33*, 131–141. [CrossRef]
- 70. Sakaue, H.; Takata, T.; Fujii, N.; Sasaki, H.; Fujii, N. Alpha B- and βA3-crystallins containing d-Aspartic acids exist in a monomeric state. *Biochim. Biophys. Acta (BBA)-Proteins Proteom.* **2015**, *1854*, 1–9. [CrossRef] [PubMed]
- 71. Takata, T.; Fujii, N. Isomerization of Asp residues plays an important role in alphaA-crystallin dissociation. *FEBS J.* **2016**, 283, 850–859. [CrossRef] [PubMed]
- Aki, K.; Fujii, N.; Fujii, N. Kinetics of isomerization and inversion of aspartate 58 of alphaA-crystallin peptide mimics under physiological conditions. *PLoS ONE* 2013, 8, e58515. [CrossRef]
- 73. Fujii, N.; Kawaguchi, T.; Sasaki, H.; Fujii, N. Simultaneous stereoinversion and isomerization at the Asp-4 residue in betaB2crystallin from the aged human eye lenses. *Biochemistry* **2011**, *50*, 8628–8635. [CrossRef]
- 74. Ritz-Timme, S.; Rochholz, G.; Schutz, H.W.; Collins, M.J.; Waite, E.R.; Cattaneo, C.; Kaatsch, H.J. Quality assurance in age estimation based on aspartic acid racemisation. *Int. J. Leg. Med.* **2000**, *114*, 83–86. [CrossRef]
- 75. Arany, S.; Ohtani, S.; Yoshioka, N.; Gonmori, K. Age estimation from aspartic acid racemization of root dentin by internal standard method. *Forensic Sci. Int.* 2004, 141, 127–130. [CrossRef]
- Hesaka, A.; Sakai, S.; Hamase, K.; Ikeda, T.; Matsui, R.; Mita, M.; Horio, M.; Isaka, Y.; Kimura, T. D-Serine reflects kidney function and diseases. *Sci. Rep.* 2019, *9*, 5104. [CrossRef] [PubMed]
- 77. Hasegawa, S.; Jao, T.-M.; Inagi, R. Dietary Metabolites and Chronic Kidney Disease. Nutrients 2017, 9, 358. [CrossRef] [PubMed]
- Li, S.; Xu, P.; Han, L.; Mao, W.; Wang, Y.; Luo, G.; Yang, N. Disease-syndrome combination modeling: Metabolomic strategy for the pathogenesis of chronic kidney disease. *Sci. Rep.* 2017, *7*, 8830. [CrossRef] [PubMed]
- 79. Stevens, L.A.; Schmid, C.H.; Greene, T.; Li, L.; Beck, G.J.; Joffe, M.M.; Froissart, M.; Kusek, J.W.; Zhang, Y.; Coresh, J.; et al. Factors other than glomerular filtration rate affect serum cystatin C levels. *Kidney Int.* **2009**, *75*, 652–660. [CrossRef]
- 80. Iwakawa, H.; Makabe, S.; Ito, T.; Yoshimura, T.; Watanabe, H. Urinary D-serine level as a predictive biomarker for deterioration of renal function in patients with atherosclerotic risk factors. *Biomarkers* **2019**, *24*, 159–165. [CrossRef]
- Kögl, F.; Erxleben, H. Zur Ätiologie der malignen Tumoren. 1. Mitteilung über die Chemie der Tumoren. Hoppe-Seyler'S Z. Physiol. Chem. 1939, 258, 57. [CrossRef]
- 82. Miller, J.A. Do tumor proteins contain d-amino acids? A review of the controversy. *Cancer Res.* **1950**, *10*, 65–72. [PubMed]
- 83. Fisher, G.H.; Torres, D.; Bruna, J.; Cerwinski, S.; Martin, T.; Bergljung, C.; Gruneiro, A.; Chou, S.J.; Man, E.H.; Pappatheodorou, S. Presence of D-aspartate and D-glutamate in tumor proteins. *Cancer Biochem. Biophys.* **1995**, *15*, 79–82. [PubMed]
- 84. Nagata, Y.; Sato, T.; Enomoto, N.; Ishii, Y.; Sasaki, K.; Yamada, T. High concentrations of D-amino acids in human gastric juice. *Amino Acids* 2007, *32*, 137–140. [CrossRef]
- 85. Zhang, Z.; Liu, Y.; Liu, P.; Yang, L.; Jiang, X.; Luo, D.; Yang, D. Non-invasive detection of gastric cancer relevant d-amino acids with luminescent DNA/silver nanoclusters. *Nanoscale* 2017, *9*, 19367–19373. [CrossRef]

- Han, M.; Xie, M.; Han, J.; Yuan, D.; Yang, T.; Xie, Y. Development and validation of a rapid, selective, and sensitive LC-MS/MS method for simultaneous determination of d and l-amino acids in human serum: Application to the study of hepatocellular carcinoma. *Anal. Bioanal. Chem.* 2018, 410, 2517–2531. [CrossRef] [PubMed]
- Du, S.; Wang, Y.; Alatrash, N.; Weatherly, C.A.; Roy, D.; MacDonnell, F.M.; Armstrong, D.W. Altered profiles and metabolism of Land D-amino acids in cultured human breast cancer cells vs. non-tumorigenic human breast epithelial cells. *J. Pharm. Biomed. Anal.* 2019, 164, 421–429. [CrossRef] [PubMed]
- 88. Bi, X.; Henry, C.J. Plasma-free amino acid profiles are predictors of cancer and diabetes development. *Nutr. Diabetes* **2017**, *7*, e249. [CrossRef] [PubMed]
- 89. DeBerardinis, R.J.; Chandel, N.S. Fundamentals of cancer metabolism. Sci. Adv. 2016, 2, e1600200. [CrossRef] [PubMed]
- 90. Ito, T.; Hayashida, M.; Kobayashi, S.; Muto, N.; Hayashi, A.; Yoshimura, T.; Mori, H. Serine racemase is involved in d-aspartate biosynthesis. *J. Biochem.* **2016**, *160*, 345–353. [CrossRef]
- 91. Bastings, J.J.A.J.; van Eijk, H.M.; Damink, S.W.O.; Rensen, S.S. d-amino Acids in Health and Disease: A Focus on Cancer. *Nutrients* 2019, *11*, 2205. [CrossRef]
- 92. Yakubu, R.R.; Nieves, E.; Weiss, L.M. The Methods Employed in Mass Spectrometric Analysis of Posttranslational Modifications (PTMs) and Protein-Protein Interactions (PPIs). *Adv. Exp. Med. Biol.* **2019**, *1140*, 169–198. [CrossRef]
- 93. Vidova, V.; Spacil, Z. A review on mass spectrometry-based quantitative proteomics: Targeted and data independent acquisition. *Anal. Chim Acta* **2017**, *964*, 7–23. [CrossRef]
- 94. Livnat, I.; Tai, H.-C.; Jansson, E.T.; Bai, L.; Romanova, E.V.; Chen, T.-t.; Yu, K.; Chen, S.-a.; Zhang, Y.; Wang, Z.-y.; et al. A D-Amino Acid-Containing Neuropeptide Discovery Funnel. *Anal. Chem.* **2016**, *88*, 11868–11876. [CrossRef]
- Tai, H.C.; Checco, J.W.; Sweedler, J.V. Non-targeted Identification of D-Amino Acid-Containing Peptides Through Enzymatic Screening, Chiral Amino Acid Analysis, and LC-MS. *Methods Mol. Biol.* 2018, 1719, 107–118. [CrossRef]
- Ni, W.; Dai, S.; Karger, B.L.; Zhou, Z.S. Analysis of isoaspartic Acid by selective proteolysis with Asp-N and electron transfer dissociation mass spectrometry. *Anal. Chem.* 2010, 82, 7485–7491. [CrossRef] [PubMed]
- 97. Adams, C.M.; Zubarev, R.A. Distinguishing and quantifying peptides and proteins containing D-amino acids by tandem mass spectrometry. *Anal. Chem.* 2005, 77, 4571–4580. [CrossRef] [PubMed]
- 98. Adams, C.M.; Kjeldsen, F.; Zubarev, R.A.; Budnik, B.A.; Haselmann, K.F. Electron capture dissociation distinguishes a single D-amino acid in a protein and probes the tertiary structure. *J. Am. Soc. Mass Spectrom.* **2004**, *15*, 1087–1098. [CrossRef]
- 99. Tao, Y.; Quebbemann, N.R.; Julian, R.R. Discriminating D-Amino Acid-Containing Peptide Epimers by Radical-Directed Dissociation Mass Spectrometry. *Anal. Chem.* **2012**, *84*, 6814–6820. [CrossRef] [PubMed]
- Lambeth, T.R.; Julian, R.R. Chapter Four—Differentiation of peptide isomers and epimers by radical-directed dissociation. In *Methods in Enzymology*; Garcia, B.A., Ed.; Academic Press: Cambridge, MA, USA, 2019; Volume 626, pp. 67–87.
- Fujii, N.; Takata, T.; Fujii, N.; Aki, K. Isomerization of aspartyl residues in crystallins and its influence upon cataract. *Biochim. Biophys. Acta* 2016, 1860, 183–191. [CrossRef]
- 102. Li, G.; Delafield, D.G.; Li, L. Improved structural elucidation of peptide isomers and their receptors using advanced ion mobility-mass spectrometry. *Trends Anal. Chem.* 2019. [CrossRef]
- 103. Gabelica, V.; Marklund, E. Fundamentals of ion mobility spectrometry. *Curr. Opin. Chem. Biol.* **2018**, *42*, 51–59. [CrossRef] [PubMed]
- 104. Uetrecht, C.; Rose, R.J.; van Duijn, E.; Lorenzen, K.; Heck, A.J. Ion mobility mass spectrometry of proteins and protein assemblies. *Chem Soc. Rev.* 2010, 39, 1633–1655. [CrossRef]
- Hernandez-Mesa, M.; Ropartz, D.; Garcia-Campana, A.M.; Rogniaux, H.; Dervilly-Pinel, G.; Le Bizec, B. Ion Mobility Spectrometry in Food Analysis: Principles, Current Applications and Future Trends. *Molecules* 2019, 24, 2706. [CrossRef]
- Pang, X.; Jia, C.; Chen, Z.; Li, L. Structural Characterization of Monomers and Oligomers of D-Amino Acid-Containing Peptides Using T-Wave Ion Mobility Mass Spectrometry. J. Am. Soc. Mass Spectrom. 2017, 28, 110–118. [CrossRef]
- Jia, C.; Lietz, C.B.; Yu, Q.; Li, L. Site-specific characterization of (D)-amino acid containing peptide epimers by ion mobility spectrometry. *Anal. Chem.* 2014, *86*, 2972–2981. [CrossRef]
- Nagy, G.; Kedia, K.; Attah, I.K.; Garimella, S.V.B.; Ibrahim, Y.M.; Petyuk, V.A.; Smith, R.D. Separation of beta-Amyloid Tryptic Peptide Species with Isomerized and Racemized I-Aspartic Residues with Ion Mobility in Structures for Lossless Ion Manipulations. *Anal. Chem.* 2019, *91*, 4374–4380. [CrossRef]
- Perez-Miguez, R.; Bruyneel, B.; Castro-Puyana, M.; Marina, M.L.; Somsen, G.W.; Dominguez-Vega, E. Chiral Discrimination of DL-Amino Acids by Trapped Ion Mobility Spectrometry after Derivatization with (+)-1-(9-Fluorenyl)ethyl Chloroformate. *Anal. Chem.* 2019, *91*, 3277–3285. [CrossRef] [PubMed]
- 110. Bhushan, R.; Bruckner, H. Marfey's reagent for chiral amino acid analysis: A review. Amino Acids 2004, 27, 231–247. [CrossRef]
- B'Hymer, C.; Montes-Bayon, M.; Caruso, J.A. Marfey's reagent: Past, present, and future uses of 1-fluoro-2,4-dinitrophenyl-5-Lalanine amide. J. Sep. Sci. 2003, 26, 7–19. [CrossRef]
- 112. Ayon, N.J.; Sharma, A.D.; Gutheil, W.G. LC-MS/MS-Based Separation and Quantification of Marfey's Reagent Derivatized Proteinogenic Amino Acid DL-Stereoisomers. *J. Am. Soc. Mass Spectrom.* **2019**, *30*, 448–458. [CrossRef]
- Li, Z.; Xing, Y.; Guo, X.; Cui, Y. Development of an UPLC-MS/MS method for simultaneous quantitation of 11 d-amino acids in different regions of rat brain: Application to a study on the associations of d-amino acid concentration changes and Alzheimer's disease. J. Chromatogr. B Anal. Technol Biomed. Life Sci. 2017, 1058, 40–46. [CrossRef]

- 114. Wang, L.; Jin, Z.; Wang, X.; Zeng, S.; Sun, C.; Pan, Y. Pair of Stereodynamic Chiral Benzylicaldehyde Probes for Determination of Absolute Configuration of Amino Acid Residues in Peptides by Mass Spectrometry. *Anal. Chem.* 2017, *89*, 11902–11907. [CrossRef] [PubMed]
- 115. Molla, G.; Piubelli, L.; Volontè, F.; Pilone, M.S. Enzymatic detection of D-amino acids. In *Unnatural Amino Acids*; Springer: Totowa, NJ, USA, 2012; pp. 273–289.
- 116. Rosini, E.; D'Antona, P.; Pollegioni, L. Biosensors for D-Amino acids: Detection methods and applications. *Int. J. Mol. Sci.* 2020, 21, 4574. [CrossRef] [PubMed]
- 117. Lata, S.; Pundir, C. Fabrication of an amperometric D-amino acid biosensor based on nickel hexacyanoferrate polypyrrole hybrid film deposited on glassy carbon electrode. *Bioprocess. Biosyst. Eng.* **2013**, *36*, 81–89. [CrossRef]
- 118. Shoja, Y.; Rafati, A.A.; Ghodsi, J. Enzymatic biosensor based on entrapment of D-amino acid oxidase on gold nanofilm/MWCNTs nanocomposite modified glassy carbon electrode by sol-gel network: Analytical applications for D-alanine in human serum. *Enzym. Microb. Technol.* 2017, 100, 20–27. [CrossRef] [PubMed]
- 119. Pundir, C.S.; Lata, S.; Narwal, V. Biosensors for determination of D and L- amino acids: A review. *Biosens. Bioelectron.* 2018, 117, 373–384. [CrossRef]
- 120. Jansson, E.T. Strategies for analysis of isomeric peptides. J. Sep. Sci. 2018, 41, 385–397. [CrossRef]
- 121. Batalla, P.; Martin, A.; Angel Lopez, M.; Cristina Gonzalez, M.; Escarpa, A. Enzyme-Based Microfluidic Chip Coupled to Graphene Electrodes for the Detection of D-Amino Acid Enantiomer-Biomarkers. *Anal. Chem.* **2015**, *87*, 5074–5078. [CrossRef] [PubMed]