An open randomized multicentre Phase 2 trial to assess the safety of DAV132 and its efficacy to protect gut microbiota diversity in hospitalized patients treated with fluoroquinolones

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Background: DAV132 (colon-targeted adsorbent) has prevented antibiotic-induced effects on microbiota in healthy volunteers.

Objectives: To assess DAV132 safety and biological efficacy in patients.

Patients and methods: An open-label, randomized [stratification: fluoroquinolone (FQ) indication] multicentre trial comparing DAV132 (7.5 g, 3 times a day, orally) with No-DAV132 in hospitalized patients requiring 5–21 day treatment with FQs and at risk of *Clostridioides difficile* infection (CDI). FQ and DAV132 were started simultaneously, DAV132 was administered for 48 h more, and patients were followed up for 51 days. The primary endpoint was the rate of adverse events (AEs) independently adjudicated as related to DAV132 and/or FQ. The planned sample size of 260 patients would provide a 95% CI of \pm 11.4%, assuming a 33% treatment-related AE rate. Plasma and faecal FQ concentrations, intestinal microbiota diversity, intestinal colonization with *C. difficile*, MDR bacteria and yeasts, and *ex vivo* resistance to *C. difficile* faecal colonization were assessed.

Results: Two hundred and forty-three patients (median age 71 years; 96% with chronic comorbidity) were included (No-DAV132, n=120; DAV132, n=123). DAV132- and/or FQ-related AEs did not differ significantly: 18 (14.8%) versus 13 (10.8%) in DAV132 versus No-DAV132 patients (difference 3.9%; 95% CI: -4.7 to 12.6). Day 4 FQ plasma levels were unaffected. DAV132 was associated with a >98% reduction in faecal FQ levels (Day 4 to end of treatment; P < 0.001), less impaired microbiota diversity (Shannon index; P = 0.003), increased ex vivo resistance to *C. difficile* colonization (P = 0.0003) and less frequent FQ-induced VRE acquisition (P = 0.01).

Conclusions: In FQ-treated hospitalized patients, DAV132 was well tolerated, and FQ plasma concentrations unaffected. DAV132 preserved intestinal microbiota diversity and *C. difficile* colonization resistance.

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Introduction

Antibiotics constitute a landmark of modern medicine. However, their use disrupts the colonic microbiota. The non-absorbed part of orally administered antibiotics, and the fraction of oral and parenteral antibiotics excreted into the bile that reach the colon, induce dysbiosis and a decrease in richness and diversity of the microbiota. Short-term consequences¹⁻⁶ include antibioticassociated diarrhoea (AAD), Clostridioides difficile infection (CDI) and selection of resistant bacteria.⁷⁻⁹ Long-term consequences include impacts on immune^{4,6} and metabolic regulations.^{5,6} Exacerbation of graft-versus-host disease in allogeneic HSCT recipients and increased mortality in cancer patients have been associated with antibiotic use.^{10,11} Various strategies to protect the intestinal microbiota have been developed.¹² Oral administration of β -lactamases hydrolysing β -lactams in the colon appears promising in Phase 2 studies,^{13–17} although limited to β-lactam antibiotics. Filling this gap, DAV132 has been developed for use with a broad range of antibiotics. It is made of millimetric beads consisting of a core of a specific activated charcoal surrounded by a polymer coating that is insoluble during transit through the stomach and most of the small intestine. In the Phase 1 studies programme, it was shown to dissolve in the distal ileum to liberate the charcoal, which then adsorbs and thereby inactivates antibiotics in the caecum/colon.¹⁸ In another Phase 1 study in healthy volunteers treated with oral moxifloxacin, DAV132 reduced faecal antibiotic concentrations by 99% and preserved intestinal microbiota diversity.¹⁹

As the charcoal contained in DAV132 is delivered in the late ileum and not before, there is no adsorption of oral medications in the upper part of the GI tract. Indeed, blood levels of drugs absorbed in the upper part of the small intestine, such as antibiotics like amoxicillin¹⁸ and moxifloxacin,¹⁹ or other medications like narrow therapeutic index drugs such as warfarin and clonazepam²⁰ are not impacted by co-administration of DAV132. Here, we report the safety of DAV132 in an open-label, randomized, Phase 2 clinical trial in hospitalized patients, mostly elderly with comorbidities, receiving systemic fluoroquinolones (FQs) for acute infections and DAV132 efficacy to protect intestinal microbiota diversity and preserve colonization resistance.

Materials and methods

Study design and oversight

We performed a parallel-arm randomized open-label multicentre clinical trial in hospitalized patients receiving FQs. The primary endpoint was the safety of DAV132 based on the occurrence of events of interest (definition in the Supplementary methods, available as Supplementary data at *JAC* Online). All other clinical and biological endpoints were secondary (Table S1). Patients were randomized to receive DAV132 or No-DAV132, and the 1:1 randomization was stratified according to FQ indication. The trial was performed in an open-label fashion, since no placebo with the ability to blacken the stools in the same manner as DAV132 could be identified. In response, an independent adjudication committee (IAC), made up of external pharmacologists and pharmacovigilance experts, performed a blinded assessment of the causality of predefined types of events (see *Safety assessment* section and Supplementary methods). Approvals from Health Authorities and Ethics Committees from each of the participating countries where the trial was run were

Ethics

The study was conducted in agreement with Good Clinical Practice and registered appropriately (NCT03710694 and EUDAMED #CIV-18-03-023465). Patients signed an informed consent prior to inclusion and randomization. Regulatory authority approvals were obtained on the first version and the final protocol in the participating countries (for details, see Supplementary methods).

Patients

Eligible patients were \geq 18 years old, hospitalized (excluding ICUs) for an expected stay of \geq 3 days, and treated for 5–21 days with a FQ monotherapy (moxifloxacin, levofloxacin or ciprofloxacin; oral or IV) for either a lower respiratory tract infection (LRTI), a complicated urinary tract infection (cUTI) or for prophylaxis of febrile neutropenia. Inclusion criteria also included risk factors for CDI (Table S2). Exclusion criteria included antibiotic exposure during the week preceding randomization, suspected or confirmed CDI at screening or anti-*C. difficile* treatment, use of probiotics or intestinal adsorbents, a history of faecal transplantation, or diarrhoea of any cause (Table S2). As the duration of the FQ treatment was variable, study visits were planned at fixed days following randomization, and at days based on the end of FQ treatment (see *Visits* in Supplementary methods and Figure S1).

Products and treatments

In the DAV132 arm, the product was given orally three times a day before meals, at a unitary dose of 7.5 g (i.e. 5.1 g of activated charcoal), for the entire duration of the antibiotic treatment, and for 2 days thereafter.

Study populations

The intent-to-treat set (ITTS) included all randomized patients managed in compliance with Good Clinical Practice throughout the study (Figure 1). The safety set (SS), on which safety was assessed, included patients from the DAV132 arm having received at least one dose of DAV132 and one dose of FQ for the No-DAV132 arm. The per protocol set (PPS), on which efficacy was assessed, included patients having received at least 5 days of DAV132 (if in the DAV132 arm) and of FQ, and without any major protocol deviations.

Safety assessment

Clinical examinations were performed at each study visit and as needed according to the investigator's judgment. Management of comorbidities and compliance with DAV132 treatment were documented throughout the study. Adverse events (AEs), serious AEs (SAEs) and treatment-emergent AEs (TEAEs) were described according to their prespecified definitions (see Supplementary methods). For predefined events such as those reported as related to DAV132 and/or to FQ by the investigators, AAD and modification of concomitant treatments of comorbidities, i.e. events of interest for the study, their causality was independently adjudicated by the IAC whilst blinded to treatment allocation. Assessment of the absence of interaction of DAV132 with antibiotic efficacy was based on the clinical cure of the original infection, and relied on the investigator's judgement and on the duration of hospitalization. The acceptability of DAV132 was assessed through questionnaires administered to patients, which used 1 (very bad) to 9 (very good) scales for the evaluation of taste, texture, ease of intake, ease of compliance with reconstitution and intake instructions, and ease of reconstitution.



Figure 1. Study populations flowchart. GCP, good clinical practice. This figure appears in colour in the online version of JAC and in black and white in the print version of JAC.

Efficacy assessment

CDI and AAD were defined as described.^{21,22} Blood, rectal swabs and stool were sampled at defined intervals after inclusion (Figure S1). Plasma and free faecal FQ concentrations were measured by reversedphase HPLC coupled with tandem MS detection. Faecal carriage of enterobacteria producing ESBL, of VRE and of yeasts was assessed qualitatively and semi-quantitatively, while that of *C. difficile* was assessed qualitatively only (all methods in Supplementary methods). Intestinal microbiota analysis was carried out by 16S rRNA gene sequencing on an Illumina MiSeq platform; α - and β -diversity indices were computed as described.²³ Resistance to *C. difficile* colonization in patients' faeces was assessed *ex vivo*, as described.²⁴

Study endpoints

The primary endpoint was the proportion of patients with at least one event of interest with causality considered related to DAV132 and/or to FQ as assessed by the IAC. Secondary endpoints included other safety endpoints, and clinical and biological efficacy endpoints such as the rate of AAD, the plasma and faecal levels of FQ, intestinal colonization with predefined bacteria, and diversity of the intestinal microbiota.

Statistics

The trial sample size, 260 patients, was based on the precision of the estimated AE rate. Assuming an observed IAC-adjudicated AE rate of 33% in each arm, the difference in rates would be 0%, with a 95% CI of -11.4% to +11.4%. This was considered a reasonable precision for a Phase 2a study. A blocked randomization list stratified by indication of FQ treatment was generated by computer. Randomization results were communicated through a Web Response System.

All data were managed blindly prior to database lock and statistical analyses performed with SAS[®] version 9.4 (Cary, NC, USA) or R version 3.6.1.²⁵ While the ITTS was predefined to be analysed as randomized, SS and PPS populations were predefined to be analysed as treated, i.e. considering the treatments actually received by the patient. Safety endpoints were analysed on the SS, and clinical and biological efficacy endpoints were analysed on the PPS. For the primary endpoint, the difference between treatments was calculated and the 95% CI for the difference presented using the method of Miettinen and Nurminen. Descriptive statistical analyses were performed for the AEs reported by the investigators. Statistical tests were two-sided with a 5% level of

significance. For the analysis of each intestinal microbiota index, correction for multiple testing at different timepoints was used as described.²⁶ No missing data were expected for the primary endpoints as the IAC reviewed relevant information on all patients. For secondary and exploratory endpoints, data were classified as missing and not imputed, and the number of missing values was clearly stated.

Results

Patients

From October 2018 to August 2019, 260 patients were enrolled at 24 study sites in Serbia (n=114 patients; 5 sites), Romania (n=34; 10 sites), Bulgaria (n=110; 8 sites) and Germany (n=110; 8 sites)2; 1 site). All 17 patients from one specific site were excluded, based on the site's non-compliance with Good Clinical Practice (mostly unavailability of patient charts for monitoring). The ITTS included 243 patients with 120 in the No-DAV132 arm and 123 in the DAV132 arm (Figure 1). For safety purposes, the SS analysed patients as actually treated; therefore, one patient from the No-DAV132 arm who did not receive FQs was excluded from the SS, whereas one patient from the DAV132 arm who did not receive DAV132 was considered for safety analysis in the No-DAV132 arm. Therefore, the SS included 242 patients from 23 participating centres. The PPS eventually comprised 199 patients. Baseline characteristics were similar between both arms (Table 1); in particular, the mean duration of treatment with FQs was 7.4 days in the No-DAV132 arm and 7.6 days in the DAV132 arm (median 7.0 days in both arms), and the treatment was initially administered by the IV route for 79.1% and 78.1% of patients, respectively. The duration of treatment and the ratio of IV route versus oral route of administration were similar between FQs (see Table S3).

Safety

Primary endpoint

The proportions of patients with events of interest related to DAV132 and/or FQ, as adjudicated by the IAC, were not different

between arms [20 events in 13 (10.8%) No-DAV132 patients, versus 25 events in 18 (14.8%) DAV132 patients; difference of the proportions: 3.9%; 95% CI: -4.7% to 12.6%]. No event of interest could be related to DAV132 only.

Secondary endpoints

Clinical safety Overall, 135 AEs were reported by the investigators in 74/242 patients (30.4%), without a clinically meaningful difference between arms (Table 2). None of the TEAEs leading to withdrawal from the study led to modification of the regimens of any concomitant drug. Overall, SAEs were reported in 17 patients, with a similar incidence between treatment arms, and

Table 1. Main characteristics of patients at baseline in the SS and PPS

none were considered related to DAV132 or FQs (Table 2). All patients with non-fatal SAEs recovered, except for one with a lung neoplasm. The most frequent TEAEs were gastrointestinal, mostly constipation in DAV132 patients (5.7% versus 0.8% of No-DAV132 patients), whereas diarrhoea events were more frequent in No-DAV132 patients (6.7% versus 3.3% in DAV132 patients). Most TEAEs were mild to moderate. The incidence of TEAEs leading to FQ discontinuation was similar between arms (Table 2).

No patient with DAV132 required modification of the regimen of concomitantly administered drugs.

The rate of cure of LRTI or cUTI was not different between the No-DAV132 and DAV132 arms (95.9% versus 94.2%).

		SS		PPS	
/ariables	No-DAV132 (n=120)	DAV132 (n=122)	No-DAV132 (n=104)	DAV132 (n=95)	
Age, years, median (SD)	72.2 (7.8)	72.0 (8.1)	72.8 (5.9)	72.6 (7.9)	
patients aged ≥ 65 years, n (%)	114 (95.0)	113 (92.6)	101 (97.1)	88 (92.6)	
Male sex, n (%)	60 (50.0)	60 (49.2)	53 (51.0)	43 (45.3)	
FQ indication, n (%)					
LRTI	96 (80.0)	96 (78.7)	84 (80.8)	73 (76.8)	
cUTI	20 (16.7)	22 (18.0)	17 (16.3)	19 (20.0)	
febrile neutropenia (prophylaxis)	4 (4.2)	4 (3.3)	3 (2.9)	3 (3.2)	
Fluoroquinolone administered ^a , n (%)					
moxifloxacin	25 (20.8)	17 (13.9)	23 (22.1)	16 (16.8)	
levofloxacin	48 (40.0)	57 (46.7)	43 (41.3)	47 (49.5)	
ciprofloxacin	47 (39.2)	48 (39.3)	38 (36.5)	32 (33.7)	
Chronic comorbidities					
at least one comorbidity, n (%)	113 (94.2)	118 (96.7)	99 (95.2)	93 (97.9)	
charlson comorbidity index, median (min-max)	2.0 (0–10)	2.0 (0-9)	2.0 (0-10)	2.0 (0–9)	
Comorbidities, n (%)					
severe cardiopulmonary condition ^b	85 (70.8)	91 (74.6)	70 (72.2)	69 (80.2)	
congestive heart failure	62 (51.7)	66 (54.1)	52 (53.6)	53 (61.6)	
COPD	71 (59.2)	63 (51.6)	68 (65.4)	50 (52.6)	
diabetes mellitus	35 (29.2)	49 (40.2)	29 (29.9)	38 (44.2)	
cerebrovascular disease	15 (12.5)	10 (8.2)	13 (12.5)	5 (5.3)	
solid tumour or haematological malignancy	12 (10.0)	9 (7.3)	11 (10.6)	8 (8.4)	
moderate to severe chronic kidney disease	9 (7.5)	4 (3.3)	7 (6.7)	2 (2.1)	
cirrhosis	3 (2.5)	4 (3.3)	3 (2.9)	3 (3.2)	
Recent history of CDI ^c , n (%)					
patients without any CDI	97 (80.8)	96 (78.7)	85 (81.7)	74 (77.9)	
1 or 2 recent episodes	4 (3.3)	9 (7.4)	3 (2.9)	8 (8.4)	
Previous hospitalization of more than 72 h and/or receiving long-term nursing care for more than 1 month within the last 90 days, n (%)	47 (39.2)	45 (36.9)	40 (38.5)	34 (35.8)	
Previous cumulated exposure of at least 5 days to any antibiotic within the last 90 days, n (%)	107 (89.2)	114 (93.4)	98 (94.2)	89 (93.7)	
β-Lactams, penicillins	20 (16.7)	15 (12.3)	18 (17.3)	10 (10.5)	
β -Lactams, cephalosporins and carbapenems	52 (43.3)	53 (43.4)	43 (41.3)	42 (44.2)	
quinolones	28 (23.3)	32 (26.2)	24 (23.1)	24 (25.3)	
macrolides, lincosamides and streptogramins	27 (22.5)	28 (23.0)	26 (25.0)	24 (25.3)	

^aMost commonly used FQ dose regimen: (a) moxifloxacin 400 mg once a day, IV route; (b) levofloxacin 500 mg once a day, IV route; (c) levofloxacin 500 mg once a day, oral route; (d) ciprofloxacin 200 mg twice a day, IV route.

^bSevere cardiopulmonary conditions included chronic congestive heart failure and severe arterial hypertension.

^cThe time interval for recent history of CDI was within the last 6 months prior to study inclusion.

Hospitalization duration was similar between arms: median (IQR): 8 (7–9) days in No-DAV132 versus 8 (7–10) in DAV132 patients.

The compliance with DAV132, defined as the proportion of patients who took 100% of the doses, was high (86.9%). Product acceptability was good with a median score of 6–7 for the various items (details in Table S4). Vitamin K blood levels at the end of FQ treatment were similar between groups: median (min-max): 180 (30–1449) versus 159 ng/L (30–1776) in No-DAV132 versus DAV132 patients, respectively, as were blood electrolytes (data not shown).

Efficacy

Biological safety Trough and peak plasma concentrations of the three tested FQs were not significantly different between arms for any regimen (Figure S2 and Table S5).

Stools and rectal swabs were collected at predefined timepoints, e.g. 86.9% of faecal samples at end-of-FQ visit and 87.9% at 10 ± 1 days for the pharmacokinetics analysis (Figure S1).

Table 2. Number of AEs (TEAE or not) and as reported by the investigators, and number of patients affected by these AEs/TEAEs in patients not receiving (No-DAV132) or receiving DAV132 in the SS

	No-DAV132 (n=120)	DAV132 (n=122)		
Characteristics	number of patients (%)	number of events	number of patients (%)	number of events	
At least one AE	33 (27.5)	62	41 (33.6)	73	
At least one TEAE	33 (27.5)	62	40 (32.8)	71	
AE leading to study withdrawal	2 (1.7)	2 (3.2)	4 (3.3)	4 (5.5)	
SAE	8 (6.7)	8 (12.9)	9 (7.4)	9 (12.3)	
AE leading to death	2 (1.7)	2 (3.2)	2 (1.6)	2 (2.7)	
Intensity of the AE					
mild	24 (20.0)	35	29 (23.8)	44	
moderate	13 (10.8)	23	12 (9.8)	19	
severe	3 (2.5)	4	6 (4.9)	8	
At least one TEAE related to DAV132	NA	NA	8 (6.6)	8 (11.3)	
At least one TEAE related to FQ	9 (7.5)	9 (14.5)	11 (9.0)	11 (15.5)	
At least one TEAE related to DAV132 only	NA	NA	0	0	
Any TEAE	33 (27.5)	62	40 (32.8)	71	
Gastrointestinal disorders	15 (12.5)	23	17 (13.9)	22	
diarrhoea	8 (6.7)	8	4 (3.3)	4	
constipation	1 (0.8)	1	7 (5.7)	7	
nausea	2 (1.7)	2	4 (3.3)	4	
abdominal pain	4 (3.3)	4	1 (0.8)	1	
General disorders and administration site conditions	5 (4.2)	6	8 (6.6)	8	
Infections and infestations	5 (4.2)	5	6 (4.9)	6	
Vascular disorders	3 (2.5)	3	5 (4.1)	6	
Cardiac disorders	4 (3.3)	4	4 (3.3)	5	
Nervous system disorders	3 (2.5)	3	4 (3.3)	4	
Psychiatric disorders	0	0	2 (1.6)	3	
Respiratory, thoracic and mediastinal disorders	7 (5.8)	7	3 (2.5)	3	
Musculoskeletal and connective tissue disorders	1 (0.8)	1	2 (1.6)	2	
Ear and labyrinth disorders	0	0	2 (1.6)	2	
Skin and subcutaneous tissue disorders	0	0	2 (1.6)	2	
Investigations	3 (2.5)	4	2 (1.6)	4	
Metabolism and nutrition disorders	3 (2.5)	5	1 (0.8)	1	
Renal and urinary disorders	1 (0.8)	1	1 (0.8)	1	
Neoplasms benign, malignant and unspecified	0	0	1 (0.8)	1	
Reproductive system and breast disorders	0	0	1 (0.8)	1	

Several AEs may have occurred in a single patient. An AE is any untoward medical occurrence, unintended disease or injury, or untoward clinical signs (including abnormal laboratory findings) in a patient participating in a clinical study whether or not the event is related to a treatment or procedure. A TEAE is an AE that occurs on or after the first administration or that is present prior to dosing but is exacerbated on or after the first administration. An SAE is defined as any untoward medical occurrence that, at any dose, results in death, is life-threatening, requires hospitalization or prolongation of existing hospitalization, results in persistent or significant disability/incapacity, or is a congenital anomaly/birth defect. NA, not applicable.

Faecal levels of fluoroquinolones

Co-administration of DAV132 led to a major reduction in the faecal levels of all FQs, at all times, with the geometric mean of free FQ faecal concentrations at the end-of-FQ visit reduced by 99.6%, 98.6% and 99.8% for levofloxacin, moxifloxacin and ciprofloxacin, respectively (Figure 2).

Clinical and biological endpoints

No CDI occurred in any patient. The proportion of AAD was not significantly different between treatment arms [between-treatment-arm difference in proportions of AAD: 2.6 (95% CI: -2.1 to 8.1); 5 patients (4.2%) in No-DAV132 arm versus 2 (1.6%) patients in DAV132 arm]. There was a trend towards decreased newly acquired *C. difficile* colonization during FQ treatment in DAV132 patients (5/101 versus 0/88; P=0.06). Furthermore, the *ex vivo*-tested resistance of patient faeces to proliferation of inoculated *C. difficile*, while similar between both arms at baseline, was significantly better preserved in patients who received DAV132: mean (95% CI) proliferation: 0.25 log cfu/mL (0.01-0.48) in DAV132 (n=46) versus 0.87 (0.53-1.21) in No-DAV132 (n=52); P=0.035 (Figure 3a).

The proportion of patients who acquired VRE during FQ treatment was similar between arms (No-DAV132, n=20 versus DAV132, n=16). However, in those newly colonized patients, the mean faecal counts of VRE per gram of faeces was reduced by >98% in those who received DAV132: mean (95% CI) log₁₀ of VRE count: 5.4 (4.41–6.39) in the No-DAV132 arm versus 3.6

(2.49–4.64) in the DAV132 arm; P = 0.025 (Figure 3b). By contrast, there was no statistically significant difference between arms in terms of acquisition of ESBL-producing Enterobacteriaceae or yeasts (data not shown).

Protection of intestinal microbiota diversity

In the No-DAV132 arm, FQ treatment induced a decreased intestinal microbiota α -diversity and richness, as assessed by the change at the end of FQ versus baseline for the Shannon index (mean \pm SEM: -0.44 ± 0.14) and the number of observed operational taxonomic units (OTUs) (mean \pm SEM: -23.5 ± 6.1). These were highly significantly prevented by DAV132, both for the Shannon index [difference of means (95% CI) for No-DAV132 versus DAV132: -0.42 (-0.71 to -0.12); P=0.024] and the number of observed OTUs [difference of means (95% CI) for No-DAV132 versus DAV132: -28.1 (-42.6 to -13.7); P= 0.001], respectively (Figure 4). In DAV132 patients, the mean (SEM) changes at the end of FQ versus baseline were -0.02(0.10) for the Shannon index and 2.4 (5.1) for the number of observed OTUs. Ten and 30 days after the end of FQ, microbiota diversity and richness were not significantly different between arms. Differences in composition of the microbiota were analysed by computing two B-diversity indices. Bray-Curtis dissimilarity and unweighted UniFrac distances (Figure 5). Both metrics show that the composition of the microbiota was significantly less altered when patients were co-administered DAV132 together with antibiotics.



Figure 2. Faecal free concentrations of FQs at successive timepoints since study initiation. Faecal free concentrations of FQs (mean \pm SEM) in PPS patients treated in the absence (red signs) or presence (blue signs) of DAV132 are shown. At each timepoint, the values from both arms were compared using a Wilcoxon rank sum test. MXF, moxifloxacin; LVX, levofloxacin; CIP, ciprofloxacin (by FQ, all dose regimens together for each FQ). This figure appears in colour in the online version of JAC and in black and white in the print version of JAC.



Figure 3. Microbiological assessment of the effect of DAV132 on the intestinal colonization of patients from the PPS in the DAV132 (blue) or No DAV132 (red) arm. (a) Ability to prevent the growth of *C. difficile* inoculated *ex vivo* into stools of patients collected at baseline (D1) or at the end of FQ treatment. (b) Quantification of the acquisition of VRE colonization at the end of FQ treatment among those not colonized at baseline. Each box shows the IQR (the bottom is Q1; the top is Q3) and the inner line is the median. In (a) and (b), values from both arms were compared using a Wilcoxon rank sum test. This figure appears in colour in the online version of *JAC* and in black and white in the print version of *JAC*.

Discussion

Our most important result in this first-in-patient trial with DAV132 was that the number of AEs was similar between both arms, providing reassurance for the safe use of the product. This result was observed in a challenging setting of mostly elderly patients with comorbidities, which is reassuring. The safety of DAV132 was confirmed by several secondary endpoints. First, none of the investigators felt it necessary to adjust any of the regimens of the concomitant treatments prescribed for comorbidities. This may suggest an absence of clinically meaningful drug interactions in the intestinal tract between DAV132 and other

treatments, because of DAV132's localized site of action in the caecum and colon. Nearly all drugs used here for the care of patients are absorbed proximally to the distal ileum; thus, their pharmacokinetics should not be modified, as shown for some probes in volunteers.^{18–20} Second, treatment with DAV132 was not modified by investigators after initiation, demonstrating good safety and acceptability. Third, antibiotic treatment success rates, as well as hospitalization duration, were not different between arms, regardless of the FQ type and regimen. FQ pharmacokinetics in plasma were preserved and similar in patients receiving DAV132 or not. Fourth, no biological parameter was significantly modified in patients receiving DAV132. In



Figure 4. Microbiota diversity at different days after the start of the FQ treatment in patients without (red) or with DAV132 (blue) in the PPS. (a) Shannon diversity index (mean \pm SEM) and (b) the number of observed OTUs (mean \pm SEM). At each timepoint, the values from both arms were compared using a Wilcoxon rank sum test and *P* values are reported for the significant differences. This figure appears in colour in the online version of *JAC* and in black and white in the print version of *JAC*.

particular, the serum level of vitamin K, which is produced by the intestinal microbiota and is instrumental for blood coagulation,²⁷ was not affected.

The efficacy of DAV132 in protecting the intestinal microbiota from FQ-induced changes was another important result: the impairment of metagenomic α -diversity indices, which are recognized as the major descriptive element of a healthy microbiota,²⁸ was significantly prevented. This is likely related to the almost complete elimination of FQ residues from the colon by DAV132, thereby sparing the microbiota from antibiotic exposure, as previously shown in volunteers.¹⁹

The trial results also suggest a functional benefit of DAV132 in preserving resistance to colonization by potentially pathogenic microorganisms. First, among patients who acquired VRE during

FQ, VRE counts were significantly lower at the end of FQ in DAV132 patients. Nosocomial infections can be preceded by VRE colonization and overgrowth,^{29,30} which may also promote spread to other patients,³¹ resulting in complications and costs.³² Thus, DAV132 may limit the spread of and subsequent infections with VRE. We did not observe the same effect for ESBL-producing Enterobacteriaceae. However, *in vivo* results in mice treated with a third-generation cephalosporin suggest that it might be different in other epidemiological settings.³³ Of note, a Phase 2 study assessing colonization resistance during co-administration of an orally administered β -lactamase together with an IV cephalosporin treatment yielded similarly discrepant results between Gram-negative and Gram-positive pathogens,¹⁷ just like faecal microbiota transfer treatments in



Figure 5. Microbiota β -diversity at different days after start of the FQ treatment in patients without (red) or with DAV132 (blue) in the PPS. (a) Bray-Curtis dissimilarity and (b) unweighted UniFrac distances. Each box shows the IQR (the bottom is Q1; the top is Q3) and the inner line is the median. At each timepoint, the values from both arms were compared using a Wilcoxon rank sum test and *P* values are reported for the significant differences. This figure appears in colour in the online version of JAC and in black and white in the print version of JAC.

eradicating VRE and/or MDR Gram-negative pathogens from the human gut. 34,35 Gram-negative pathogens seem to occupy an ecological niche that is more difficult to modify than that of VRE. 36,37

Second, the co-administration of DAV132 conferred improved colonization resistance against *C. difficile*. Patients taking DAV132 tended to be less frequently colonized by *C. difficile* and the *ex vivo* assay demonstrated the maintained growth-suppressive effect of faeces recovered from DAV132 recipients. This is in agreement with results in hamsters, which demonstrated the ability of activated charcoal to prevent CDI after a FQ^{38,39} or clindamycin.²³ Hence, there is consistency between the prevention of antibiotic-induced dysbiosis by DAV132 (assessed by α - and β -diversity indices) and the concomitant maintenance of resistance to colonization by potentially pathogenic bacteria such as VRE and *C. difficile*.

Our study has limitations. One is the lack of a placebo that can uniformly blacken the stools as DAV132 does. We minimized the consequences of this by keeping the IAC blinded to the randomization arm for analysing the causality of events of interest, and by performing all biological assays blind. Another limitation is the focus on FQs only. This enabled us to obtain a more homogeneous patient population than that of a study open to all antibiotics. Importantly, *in vitro*¹⁹ and *in vivo* studies^{23,33} have shown that DAV132 efficiently adsorbs antibiotics from non-FQ classes, hence the conclusions may be applicable to other antimicrobials. Finally, we conducted our study in four countries only; however, no country-specific effect was observed.

In conclusion, DAV132 was safe in mostly elderly hospitalized patients with comorbidities who received FQs for the treatment or prevention of infection, and effective in preventing antibiotic-induced changes in the intestinal microbiota. Given the increasing concern that antibiotic-induced intestinal changes are major drivers of a broad range of acute and chronic infectious and non-infectious diseases, as well as a factor that limits the efficacy of some major anticancer therapies, the results of this trial underline the importance of further clinical development of DAV132.

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Supplementary data

Supplementary methods, Tables S1 to S5 and Figures S1 and S2 are available as Supplementary data at JAC Online.

References

1 Jernberg C, Löfmark S, Edlund C *et al.* Long-term impacts of antibiotic exposure on the human intestinal microbiota. *Microbiology* 2010; **156**: 3216–23.

2 Dethlefsen L, Relman DA. Incomplete recovery and individualized responses of the human distal gut microbiota to repeated antibiotic perturbation. *Proc Natl Acad Sci U S A* 2011; **108** Suppl 1: 4554–61.

3 Pérez-Cobas AE, Gosalbes MJ, Friedrichs A *et al.* Gut microbiota disturbance during antibiotic therapy: a multi-omic approach. *Gut* 2013; **62**: 1591–601.

4 Belkaid Y, Hand TW. Role of the microbiota in immunity and inflammation. *Cell* 2014; **157**: 121–41.

5 Cox LM, Blaser MJ. Antibiotics in early life and obesity. *Nat Rev Endocrinol* 2015; **11**: 182–90.

6 Aversa Z, Atkinson EJ, Schafer MJ *et al*. Association of infant antibiotic exposure with childhood health outcomes. *Mayo Clin Proc* 2021; **96**: 66–77.

7 de Lastours V, Chau F, Tubach F *et al.* Independent behavior of commensal flora for carriage of fluoroquinolone-resistant bacteria in patients at admission. *Antimicrob Agents Chemother* 2010; **54**: 5193–200.

8 Johanesen P, Mackin K, Hutton M *et al.* Disruption of the gut microbiome: *Clostridium difficile* infection and the threat of antibiotic resistance. *Genes* 2015; **6**: 1347–60.

9 Leffler DA, Lamont JT. *Clostridium difficile* infection. *N Engl J Med* 2015; **372**: 1539–48.

10 Shono Y, Docampo MD, Peled JU *et al.* Increased GVHD-related mortality with broad-spectrum antibiotic use after allogeneic hematopoietic stem cell transplantation in human patients and mice. *Sci Transl Med* 2016; **8**: 339ra71.

11 Peled JU, Gomes ALC, Devlin SM *et al.* Microbiota as predictor of mortality in allogeneic hematopoietic-cell transplantation. *N Engl J Med* 2020; **382**: 822–34.

12 Andremont A, Cervesi J, Bandinelli P-A *et al.* Spare and repair the gut microbiota from antibiotic-induced dysbiosis: state-of-the-art. *Drug Discov Today* 2021; **26**: 2159–63.

13 Leonard F, Andremont A, Leclerq B *et al.* Use of beta-lactamase-producing anaerobes to prevent ceftriaxone from degrading intestinal resistance to colonization. *J Infect Dis* 1989; **160**: 274–80.

14 Stiefel U, Pultz NJ, Harmoinen J *et al.* Oral administration of β -lactamase preserves colonization resistance of piperacillin-treated mice. *J Infect Dis* 2003; **188**: 1605–9.

15 Pitout JDD. IPSAT P1A, a class A β -lactamase therapy for the prevention of penicillin-induced disruption to the intestinal microflora. *Curr Opin Investig Drugs* 2009; **10**: 838–44.

16 Connelly S, Bristol JA, Hubert S *et al.* SYN-004 (ribaxamase), an oral β -lactamase, mitigates antibiotic-mediated dysbiosis in a porcine gut microbiome model. *J Appl Microbiol* 2017; **123**: 66–79.

17 Kokai-Kun JF, Roberts T, Coughlin O *et al.* Use of ribaxamase (SYN-004), a β -lactamase, to prevent *Clostridium difficile* infection in β -lactam-treated patients: a double-blind, phase 2b, randomised placebo-controlled trial. *Lancet Infect Dis* 2019; **19**: 487–96.

18 de Gunzburg J, Ducher A, Modess C *et al.* Targeted adsorption of molecules in the colon with the novel adsorbent-based medicinal product, DAV132: a proof of concept study in healthy subject. *J Clin Pharmacol* 2015; **55**: 10–6.

19 de Gunzburg J, Ghozlane A, Ducher A *et al*. Protection of the human gut microbiome from antibiotics. *J Infect Dis* 2018; **217**: 628–36.

20 Pinquier J-L, Varastet M, Meyers D *et al.* A colon-targeted adsorbent (DAV132) does not affect the pharmacokinetics of warfarin or clonazepam in healthy subjects. *Clin Pharmacol Drug Dev* 2021; **10**: 908–17.

21 Crobach MJT, Planche T, Eckert C *et al.* European Society of Clinical Microbiology and Infectious Diseases: update of the diagnostic guidance document for *Clostridium difficile* infection. *Clin Microbiol Infect* 2016; **22** Suppl 4: S63–81.

22 Allen SJ, Wareham K, Wang D *et al.* Lactobacilli and bifidobacteria in the prevention of antibiotic-associated diarrhoea and *Clostridium difficile* diarrhoea in older inpatients (PLACIDE): a randomised, double-blind, placebo-controlled, multicentre trial. *Lancet* 2013; **382**: 1249–57.

23 Burdet C, Sayah-Jeanne S, Nguyen TT *et al.* Antibiotic-induced dysbiosis predicts mortality in an animal model of *Clostridium difficile* infection. *Antimicrob Agents Chemother* 2018; **62**: e00925-18.

24 Harris HC, Best EL, Normington C *et al*. Optimization of an assay to determine colonization resistance to *Clostridioides difficile* in fecal samples from healthy subjects and those treated with antibiotics. *Antimicrob Agents Chemother* 2020; **65**: e01401-20.

25 R Core Team. R: A Language and Environment for Statistical Computing. 2020. https://www.r-project.org/.

26 Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J R Stat Soc Ser B Methodol* 1995; **57**: 289–300.

27 LeBlanc JG, Milani C, de Giori GS *et al.* Bacteria as vitamin suppliers to their host: a gut microbiota perspective. *Curr Opin Biotechnol* 2013; **24**: 160–8.

28 Kim B-R, Shin J, Guevarra RB *et al.* Deciphering diversity indices for a better understanding of microbial communities. *J Microbiol Biotechnol* 2017; **27**: 2089–93.

29 Liss BJ, Vehreschild JJ, Cornely OA *et al.* Intestinal colonisation and blood stream infections due to vancomycin-resistant enterococci (VRE) and extended-spectrum β -lactamase-producing Enterobacteriaceae (ESBLE) in patients with haematological and oncological malignancies. *Infection* 2012; **40**: 613–9.

30 Freedberg DE, Zhou MJ, Cohen ME *et al.* Pathogen colonization of the gastrointestinal microbiome at intensive care unit admission and risk for subsequent death or infection. *Intensive Care Med* 2018; **44**: 1203–11.

31 Donskey CJ, Chowdhry TK, Hecker MT *et al*. Effect of antibiotic therapy on the density of vancomycin-resistant enterococci in the stool of colonized patients. *N Engl J Med* 2000; **343**: 1925–32.

32 Lucet J-C, Armand-Lefevre L, Laurichesse J-J *et al.* Rapid control of an outbreak of vancomycin-resistant enterococci in a French university hospital. *J Hosp Infect* 2007; **67**: 42–8.

33 Grall N, Massias L, Nguyen TT *et al.* Oral DAV131, a charcoal-based adsorbent, inhibits intestinal colonization by β -lactam-resistant *Klebsiella pneumoniae* in cefotaxime-treated mice. *Antimicrob Agents Chemother* 2013; **57**: 5423–5.

34 Huttner BD, de Lastours V, Wassenberg M *et al*. A 5-day course of oral antibiotics followed by faecal transplantation to eradicate carriage of multidrug-resistant Enterobacteriaceae: a randomized clinical trial. *Clin Microbiol Infect* 2019; **25**: 830–8.

35 Seong H, Lee SK, Cheon JH *et al*. Fecal microbiota transplantation for multidrug-resistant organism: efficacy and response prediction. *J Infect* 2020; **81**: 719–25.

36 Dubberke ER, Mullane KM, Gerding DN *et al.* Clearance of vancomycinresistant *Enterococcus* concomitant with administration of a microbiotabased drug targeted at recurrent *Clostridium difficile* infection. *Open Forum Infect Dis* 2016; **3**: ofw133.

37 Kuijper EJ, Vendrik KEW, Vehreschild MJGT. Manipulation of the microbiota to eradicate multidrug-resistant Enterobacteriaceae from the human intestinal tract. *Clin Microbiol Infect* 2019; **25**: 786–9.

38 Burdet C, Sayah-Jeanne S, Nguyen TT *et al.* Protection of hamsters from mortality by reducing fecal moxifloxacin concentration with DAV131A in a model of moxifloxacin-induced *Clostridium difficile* colitis. *Antimicrob Agents Chemother* 2017; **61**: e00543-17.

39 Saint-Lu N, Burdet C, Sablier-Gallis F *et al.* DAV131A protects hamsters from lethal *Clostridioides difficile* infection induced by fluoroquino-lones. *Antimicrob Agents Chemother* 2019; **64**: e01196-19.