



RESEARCH ARTICLE

3 OPEN ACCESS



Taxonomic Study of Sixteen Unrecorded and Five New Species of Hypocreales from the Korean Marine Environment

Wonjun Lee[‡] (D), Ji Seon Kim[‡] (D), Sumin Jo (D), Chang Wan Seo (D) and Young Woon Lim (D)

School of Biological Sciences and Institute of Biodiversity, Seoul National University, Seoul, Korea

ABSTRACT

The order *Hypocreales*, which belongs to the *Ascomycota* class *Sordariomycetes*, has a large number of species and occupies a variety of ecological niches, including saprophytic, symbiotic, and parasitic fungi. While much research has focused on terrestrial Hypocrealean fungi, there remains a significant gap in our understanding of their diversity and ecological roles in marine environments. In this study, we isolated 47 fungal strains from various marine habitats in South Korea. Through the polyphasic study, including phylogenetic analysis using multi-genetic markers (ITS, LSU, *TEF1*, *RPB2*, *TUB*, and *ACT*) and morphological analysis, we identified 21 species previously undiscovered in Korea, including 5 new and 16 unrecorded species. Our findings illustrate the species diversity of marine *Hypocreales*, highlighting the need for additional research into their ecological functions and potential in biotechnology and medicine.

ARTICLE HISTORY

Received 23 September 2024 Revised 15 October 2024 Accepted 15 October 2024

KEYWORDS

Hypocreales; Sordariomycetes; marine fungi; new species; unrecorded species

1. Introduction

order within Hypocreales is an the class Sordariomycetes of Ascomycota, comprising 14 families and 303 genera, making it the largest order in Sordariomycetes [1-4]. With its high species diversity, Hypocrealean fungi occupy a wide range of ecological niches, from saprophytic to symbiotic and parasitic fungi [5-7]. Many parasitic fungi in this order, particularly those affecting insects and plants, have garnered significant attention due to their economic impact. A notable example is the family Cordycipitaceae, which includes genera like Beauveria, Cordyceps, and Lecanicillium, known for their entomopathogenic species used as biocontrol agents (such as cyclosporine, pyridones, and fumosorinone) [8-10]. Furthermore, the other Hypocrealean fungi also directly affect plants, humans, and animals, which has been the subject of extensive research

Hypocrealean fungi are distributed globally across terrestrial and aquatic environments, but research has predominantly focused on their diversity and function in terrestrial ecosystems [7,14–17]. In contrast, studies on aquatic Hypocrealean fungi, particularly those in marine environments, have been relatively limited, with most attention directed toward

freshwater habitats [15,18-22]. This has left a gap in our understanding of their ecological roles and adaptations in these environments, challenging traditional concepts about their ecological niches [15,23-26]. Marine Hypocrealean fungi have been reported in various marine environments, from coastal sediments to deep-sea [27-30], with 168 species documented across 64 genera and 15 families (accessed September 09 2024, via https://www.marinefungi. org/) [31]. Although marine fungi are less studied compared to terrestrial fungi, they play vital roles in oceanic ecosystems, including organic matter decomposition, nutrient recycling, and symbiotic interactions with marine flora and fauna [23,32,33]. Exploring marine Hypocreales is especially interesting because they can make new bioactive compounds in response to the special physicochemical conditions of marine environments [10,34], which could be used in medicine, agriculture, and industry.

Despite growing interest, taxonomic research on marine *Hypocreales* is still in its early stages. Although Hypocrealean fungi are frequently found in various marine environments, such as macroalgae and sediment, sometimes constituting up to 30% of its fungal communities [27–30], they account for only 7% of the total marine fungi listed to date

families (accessed September 09 2024, via https:// www.marinefungi.org/) [31]. Morphological identification of marine Hypocreales is challenging due to the limited observable traits in cultured isolates, making DNA-based identification essential [4-7]. The nuclear ribosomal internal transcribed spacer (ITS), a universal fungal DNA barcode marker has been used to identify species within the Hypocreales. However, due to its low resolution, additional genetic markers, such as the partial actin (ACT), the RNA polymerase II subunit (RPB2), the translation elongation factor $1-\alpha$ (*TEF1*), and the beta-tubulin (*TUB*) have been applied in phylogenetic analyses [2,4,35,36]. Nevertheless, of the 168 documented marine Hypocreales species, only 88 have been identified based on DNA sequences (accessed September 09 2024, via https://www.marinefungi.org/) [31], highlighting the need for further molecular studies to confirm their taxonomic accuracy.

In South Korea, 262 species of Hypocreales across 8 families have been documented (National Institute of Biological Resources, accessed 2024.06.13). In comparison, only 131 species have been reported from marine environments (Marine Bio-Resource Information System, updated 2024.03.29). Over the past decade, our research has aimed to bridge the gap in marine fungal diversity, leading to the establishment of the Marine Fungal Resource Bank (MFRB), which isolates fungal strains from various marine environments [37-41]. As part of this effort, we recently applied polyphasic phylogenetic analyses using multi-genetic markers to newly isolated Hypocrealean fungal strains. This resulted in the discovery of 21 species, including 5 new species and 16 unrecorded species. Our findings underscore the need for further research into the diversity, ecological interactions, and adaptation mechanisms of marine Hypocreales to understand their roles in marine ecosystems better.

2. Materials and methods

2.1. Sampling and fungal isolation

Fungal strains were isolated from various substrates in marine environments in South Korea between 2015 and 2022 (Table 1). Sampling and isolation methods followed previously described protocols [37,38,41]. Marine organisms such as macroalgae, sandfish eggs, and sponges were collected and washed with sterilized seawater (SSW) at least three times. The samples were then cut into pieces and placed on different media used for fungal isolation. For marine sediment samples (mudflats, sand, and seabed sediment), samples were diluted to 1/10 or 1/100, and 100-200 µL of the diluted samples were inoculated onto fungal isolation media. Seawater samples were collected from the surface or bottom using Niskin bottles equipped with conductivity, temperature, and depth (CTD) rosettes. The sampled seawater was filtered through a 0.2 µm pore-sized PETE membrane filter using a vacuum pump, and the filters were placed on DRBC media. All plates were incubated at 25 °C or room temperature for 7-21 d. Each fungal colony grown on the isolation media was transferred to new PDA media supplemented with SSW. Fungal strains were preserved in 20% (v/v) glycerol with SSW at -80°C and deposited in the Seoul National University Fungus Collection.

2.2. Molecular analysis

Fresh mycelium from each strain was ground using Ruptor Elite Homogenizer (OMNI International, Kennesaw, GA). Genomic DNA was extracted using either a modified cetyltrimethylammonium bromide (CTAB) method [42] or the AccuPrep Genomic DNA Extraction Kit (Bioneer Co., Daejeon, Korea) following the manufacturer's protocols. The ITS region was amplified for all strains using the primer set ITS1F [43]/ITS4 [44]. To ensure accurate species identification within Hypocreales, additional genetic markers were selected based on reference studies Supplementary Table 2, and amplification was performed using the primer sets and conditions listed in Supplementary Table 2. PCR was performed using a C1000 thermal cycler (Bio-Rad, Richmond, CA), and the PCR products were confirmed by gel electrophoresis on a 1% agarose gel. Verified PCR products were purified using the PCR Purification Kit (GeneAll Biotechnology, Seoul, South Korea) or the ExoSAP-IT Express PCR Product Cleanup Kit (Thermo Fisher Scientific, Waltham, MA) according to the manufacturer's instructions. The purified PCR products were sequenced with the same primers used for PCR on an ABI Prism 3730xl Genetic Analyzer (Life Technologies, Gaithersburg, MD) at Macrogen (Seoul, South Korea).

Generated sequences were proofread and manually edited using Geneious Prime 2023.1 (Biomatters Ltd., San Diego, CA, https://www.geneious.com) [45]. Edited sequences were assembled using the de novo assembly function in Geneious Prime 2023.1. All sequences were deposited in GenBank (Table 1). Reference sequences were obtained from published studies and downloaded from GenBank using GenMine software [46] (Supplementary Table 1).

study.
the
.⊑
nsed
accessions
GenBank
and
information
Strain
-
Table

Charitae Cha	Chain	Location (in Remublic of Korea)	Cubetrate	ΣĽ	IISI	7561	RDR7	TIB	ACT
canado	Sualli	Location (iii nepublic of notea)	Jubbilate	2	000	1 - 1 - 1	Nr 02	201	2
Achroiostachys aurantispora	SFC20240607-M001	Incheon, Ganghwa-gun	Mud flat	PQ304492	PQ304459	PQ319913			
Acrostalagmus annulatus	SFC20240607-M002	Jeollanam-do, Suncheon-si	Sand	PQ304493	PQ304460	PQ319909	PQ319914		
	SFC20240607-M016	Jeollanam-do, Muan-gun	Mud flat	PQ304494	PQ304461	PQ319910	PQ319915		
Beauveria pseudobassiana	SFC20240607-M003	Jeollanam-do, Suncheon-si	Sand	PQ304495	PQ304462	PQ319886			
Cordyceps fumosorosea	SFC20240607-M004	Jeollanam-do, Suncheon-si	Sand	PQ304496	PQ304463	PQ319887	PQ319916	PQ319882	
	SFC20240607-M018	Gyeongsangbuk-do, Ulleung-gun	Unknown sponge	PQ304497	PQ304464	PQ319888	PQ319917	PQ319883	
Emericellopsis atlantica	SFC20240607-M005	Jeollanam-do, Muan-gun	Mud flat	PQ304498	PQ304465	PQ319908	PQ319918	PQ319884	
Fusarium concentricum	SFC20240607-M006	Jeollanam-do, Suncheon-si	Mud flat	PQ304499	PQ304466	PQ319912	PQ319919		
Lasionectriella arenuloides	SFC20240607-M007	Jeollanam-do, Suncheon-si	Sand	PO304500	PO304467	PO319892	PO319920		
I ecanicillium verrucum	SEC20240607-M029 T	Gveongsanghik-do Pohang-si	Sand	PO355558	PO355577	PO355481	,		
	SEC20240607-M030	lein-do Chuia-myeon	Unknown macroaldae	PO355559	PO355578	PO355482			
	SEC20240607 M031	Joju do, Chuis-myoon	The macroalgae	D0355560	DO355570	DO355/183			
	SFC20240007 - 1M031	Jeju-uo, Ciiuja-iiiyeoii	UINIUWII IIIACIOAIYAE	0001100	1000000	10000400			
	SFC20240807-IM032	Incheon, Jung-gu	Unknown	PQ355501	PQ555580	PQ355484	1,000,000		
Metapocnonia rubescens	SFC20160907-M17	Gangwon-do, Gangneung-si	Egg or Arcroscopus	PQ304501		PQ319896	PQ319921		
			<i>japonicus</i> (Sandfish)						
	SFC20240607-M010	Jeollanam-do, Suncheon-si	Sand	PQ304502			PQ319922	PQ319885	
Neoacremonium distortum	SFC20240607-M011	Incheon, Ganghwa-gun	Mud flat	PQ304503	PQ304468	PQ319897	PQ319923		
	SFC20240607-M034	Incheon, Ganghwa-gun	Sand	PO304504	PO304469	PO319899	PO319924		
	SFC20240607-M035	Jeollanam-do. Muan-dun	Sand	PO304505	PO304470	PO319902	PO319925		
	SEC20240607-M036	Incheon Gandhwa-dun	Mild flat	PO304506	PO304471	PO319900	PO319976		
	3FCZ 024000/ -1M1030	Inchedit, daligitwa-guii	Mud IIat	10304300	1,470	10319900	10319920		
	SFC20240607-M037	Incheon, Ganghwa-gun	Mud flat	PQ30450/	PQ304472	PQ319898	PQ31992/		
	SFC20240607-M038	Incheon, Ganghwa-gun	Mud flat	PQ304508	PQ3044/3	PQ319901	PQ319928		
Neocosmospora tuberculata	SFC20240607-M039 T	South sea of Korea	Seawater in 40 m	PQ355562	PQ355581	PQ355485	PQ355473		
			bottom						
	SFC20240607-M040	South sea of Korea	Seawater in 40 m	PQ355563	PQ355582	PQ355486	PQ355474		
			bottom						
	SFC20240607-M041	South sea of Korea	Seawater in 40 m	PO355564	PO355583	PO355487	PO355475		
			bottom						
	SFC20240607-M042	South sea of Korea	Seawater in 40 m	PO355565	PO355584	PO355488	PO355476		
			bottom	,					
Nieselia marinisedimenta	SEC20171120-M03	allo-delly ob-medelloel	Sand	DOSESER	DOSESSE	D0355/80	D0355/177	DOSESARO	
	SEC20240607-M014 T	loollanam do, maan gan	מנים לייני	DO355567	00355500	DO255400	DO255479	DO355470	
	SEC20240607 M043	Joshanam do Musa cun	ממוט למינט	000000	00055500	PO255401	0/50000	DO255470	
	SI CZ0240007 1M043	Jeonanam-do, Muan-gun	Mild flat	0000000	1000000	00255491	00711000	FQ333471	
::-::	SFC20240807 -1M044	licheon, Garagina-gun	Mud liat	10333309	0000000	PQ333492	r (2333400		
Parasarociaalum mabikii	SFC2U2406U/-IMU25	Incheon, Ganghwa-gun	Mud flat	PQ3555/0	PQ355589	PQ355493		Σ (70355463
	SFC20240607-M026	Incheon, Ganghwa-gun	Mud flat	PQ3555/1	PQ355590	PQ355494		2. 0	PQ355464
	SFC20240607-IM027	Incheon, Ganghwa-gun	Mud riat	PQ5555/2	PQ555591	PQ355495		Σ.	PQ355465
Parasarocladium	SFC20240607-M022	Jeju-do, Chuja-myeon	Unidentified seaweed	PQ355573	PQ355592	PQ355496		<u>a</u>	PQ355466
multimorphologicum									
	SFC20240607-M023	Jeju-do, Chuja-myeon	Unidentified seaweed	PQ355574	PQ355593	PQ355497		Δ.	PQ355467
	SFC20240607-M024 T	Jeju-do, Chuja-myeon	Grateloupia sp.	PQ355575	PQ355594	PQ355498		<u> </u>	PQ355468
Protocreopsis rutila	SFC20170718-M03	Incheon, Ganghwa-gun	Sand	PQ304509	PQ304474	PQ319911			
Purpureocillium lavendulum	SFC20240607-M017	South sea of Korea	Sediment in 40 m	PQ304510		PQ319889	PQ319929		
	SFC20240607-M033	Jeollanam-do, Muan-gun	Sand	PQ304511		PQ319890	PQ319930		
	SFC20240607-M015	Ganawon-do, Goseona-aun	Sand	PO304512		PO319891	PO319931		
Sarocladium bacillisporum	SFC20240607-M028	Gangwon-do, Goseong-gun	Sand	PQ304513	PQ304475	PQ319904	•	Δ.	PQ319877
Sarocladium terricola	SFC20240607-M008	Jeju-do, Jeju-si	Gelidium sp.	PQ304514	PQ304476	PQ319905		4	PQ319878

Table 1. Continued.									
Species	Strain	Location (in Republic of Korea)	Substrate	ITS	LSU	TEF1	RPB2	TUB	ACT
	SFC20240607-M009	Jeollanam-do, Muan-gun	Sand	PQ304515	PQ304477	PQ319906			PQ319879
	SFC20240607-M012	Gyeongsangbuk-do, Ulleung-gun	Unknown sponge	PQ304516	PQ304478	PQ319907			PQ319880
Sarocladium zeae	SFC20240607-M013	Jeollanam-do, Gangjin-jun	Fulvia mutica	PQ304517	PQ304479	PQ319903			PQ319881
Verruciconidia infuscata	SFC20240607-M019	Jeollanam-do, Suncheon-si	Sand	PQ304518	PQ304480	PQ319893	PQ319932		
Verruciconidia persicina	SFC20240607-M020	Jeju-do, Chuja-myeon	Sargassum thunbergii	PQ304519		PQ319894	PQ319933		
	SEC20240607-M021	Incheon Ganghwa-diin	Mud flat	PO304520	PO304481	PO319895	PO319934		

Due to the large dataset and computational requirements, the dataset was divided into four subsets following a previous study [4,7,21,35,36,47,48], and RAxML phylogenetic analysis was conducted for each.

The first dataset included Bionectriaceae and Sarocladiaceae, with phylogenetic analysis using six genetic markers (ITS, LSU, TEF1, RPB2, TUB, and ACT). ACT sequences were used for the analysis of only two genera (Sarocladium Parasarocladium) in the family Sarocladiaceae. The second dataset comprised sequences of four genetic markers (ITS, LSU, TEF1, and RPB2) from three families (Nectriaceae, Neoacremoniaceae, Stachybotryaceae). The third and fourth datasets included sequences of five genetic markers (ITS, LSU, TEF1, RPB2, and TUB) from three families (Clavicipitaceae, Hypocreaceae, and Ophiocordycipitaceae) and two families (Cordycipitaceae and Niessliaceae), respectively. All sequences for each genetic marker were aligned using MAFFT version 7.490 [49], and uninformative ends were trimmed in Geneious Prime 2023.1 (https://www.geneious.com) [45]. The alignments of each genetic marker were concatenated, and phylogenetic trees were constructed using the GTR+GAMMA model with 1000 replications through RAxML version 8 on Geneious Prime (https://www.geneious.com) [45].

2.3. Morphological observations

Each strain used for morphological observations was sub-cultured on potato dextrose agar (PDA; Difco, Pinellas Park, FL) supplemented with distilled water and inoculated on the following media: PDA, malt extract agar (MEA; Oxoid, Badhoevedorp, Netherlands), oatmeal agar (OA; Difco), corn meal agar (CMA; Difco), potato carrot agar (PCA; India), Himedia, Mumbai, and synthetic nutrient-poor agar (SNA; KH2PO4 1g, KNO3 1g, MgSO₄7H₂O 5 g, KCl 0.5 g, glucose 0.2 g, saccharose 0.2 g, and Bacto agar 20 g/L) [50]. Cultures were incubated for 7 d at 25 °C. Cultures for microscopic observation were incubated at 25°C for more than 7 d on media appropriate for each genus until the conidia formation was confirmed. The specific media used for each genus are indicated under each taxon's Taxonomy section. Conidial structures were observed using a Nikon 80i (Tokyo, Japan) or Leica DM2500 (Wetzlar, Germany) light microscope. When necessary, lactic acid or lactophenol blue dye was used for observation. More than 30 conidial structures were measured per strain using ImageJ software [51]. Colony color, including the surface and reverse side, was described using the Methuen Handbook of Colour [52].

3. Results

We isolated 47 strains from various marine substrates environments (macroalgae, animals, seawater, and sediments) and obtained 190 sequences for six genetic markers (ITS, LSU, TEF1, RPB2, TUB, and ACT) from these strains (Table 1). Phylogenetic analysis was performed using these genetic markers, along with 886 reference sequences. The dataset was divided into four subsets based on families to improve computational efficiency, and phylogenetic analysis was conducted using RAxML for each subset. The resulting phylogenetic trees are presented in Figures 1–4.

On the basis of the ITS phylogeny, the 47 strains were classified into 21 taxa, which were further assigned to 17 genera across 10 families. Different sets of genetic markers were applied depending on the family. For the six families, Bionectriaceae, Hypocreaceae, Clavicipitaceae, Cordycipitaceae, Niessliaceae, and Ophiocordycipitaceae, five markers (ITS, LSU, TEF1, RPB2, and TUB) were used. In the case of Sarocladiaceae, six markers (ITS, LSU, TEF1, were RPB2, TUB, and ACT) applied. Stachybotryaceae, Nectriaceae, and Neoacremoniaceae, four markers (ITS, LSU, TEF1, and RPB2) were used.

Through the phylogenetic analysis based on these multi-genetic markers, the 47 strains were identified as 21 species within the order Hypocreales. Of these, 16 are identified as described but previously unrecorded species in South Korea: Bionectriaceae (Emericellopsis atlantica, Lasionectriella arenuloides, Protocreopsis rutila, Verruciconidia infuscata, V. persicina), Clavicipitaceae (Metapochonia rubescens), Cordycipitaceae (Beauveria pseudobassiana, Cordyceps fumosorosea), Hypocreaceae (Acrostalagmus annula-Nectriaceae (Fusarium concentricum), Neoacremoniaceae (Neoacremonium distortum), Ophiocordycipitaceae (Purpureocillium lavendulum), Sarocladiaceae (Sarocladium bacillisporum, S. terricola, and S. zeae), and Stachybotryaceae (Achroiostachys aurantispora). Each species formed robust clades alongside sequences from described species. However, Lasionectriella arenuloides and La. marigotensis showed high similarity in both molecular and morphological characteristics [35,53]. Our strain was identified as La. arenuloides, with further details available in the species note in the Taxonomy section.

The remaining five taxa were confirmed as new species candidates, as they did not match any known

described species and formed distinct clades supported by high bootstrap values. The number of new species candidates identified in each family is as follows: Cordycipitaceae (1), Nectriaceae (1), Niessliaceae (1), and Sarocladiaceae (2). Detailed descriptions of these 21 species, including the 5 new (Lecanicillium verrucum sp. nov., Neocosmospora tuberculata sp. nov., Niesslia marinisedimenta sp. nov., Parasarocladium mabikii sp. nov., and Parasarocladium multimorphologicum sp. nov.) and 16 unrecorded species, are provided in the Taxonomy section.

4. Taxonomy

Achroiostachys aurantispora L. Lombard & Crous (Figure 5(A))

MycoBank: MB815917 **Family** *Stachybotryaceae*

Materials examined: South Korea. Incheon, Ganghwa-gun (37°36′36.3″N 126°31′13.8″E), July 2021, isolated from mudflat, M.S. Park, J.S. Kim & Y.W. Lim (SFC20240607-M001, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on MEA *Mycelia* smooth, septate, hyaline; *Conidiophores* mononematous, unbranched, erect, straight, 1–4-septate, hyaline, smooth, occasionally rough with nodules toward apex, $40-101\times2.5-5.4\,\mu\text{m}$, bearing a whorl of 5–6 conidiogenous cells. *Conidiogenous cells* terminal, ampulliform to ventricose, hyaline, smooth, $6.2-14\times2.9-4.6\,\mu\text{m}$. *Conidia* aseptate, ellipsoidal, smooth, hyaline, 1–2 guttules, $5-8.7\times3.2-4.6\,\mu\text{m}$ (av. $7.1\times3.9\,\mu\text{m}$).

Culture characteristics: Colonies on PDA reaching 56-60 mm diam in 7 d at 25 °C; flat, plain, floccose, sometimes felty, forward yellowish white (1A2), light grey (1C1); margin entire or irregular; reverse yellow (3B8), golden brown (5D7) at center. Colonies on OA reaching 53-58 mm diam in 7 d at 25 °C; flat, plain, floccose, granular, sometimes having patches with scarce aerial mycelia, forward orange white (5A2); margin entire; exudate light orange (5A4); reverse pastel yellow (3A4). Colonies on CMA: reaching 53-62 mm diam in 7 d at 25 °C; flat, plain, forward hyaline to reddish (or pinkish) white (7A2); few pustule-like formation; reverse hyaline. Colonies on SNA reaching 20-25 mm diam in 7 d at 25 °C; vegetative mycelial growth scarce, scattered, forward hyaline to yellowish white (4A2); reverse yellowish white (4A2).

Notes: The SFC20240607-M001 strain, isolated from a mudflat, is nearly identical to the holotype in morphology, with the following exceptions: it has more septa in its conidiophores compared to the

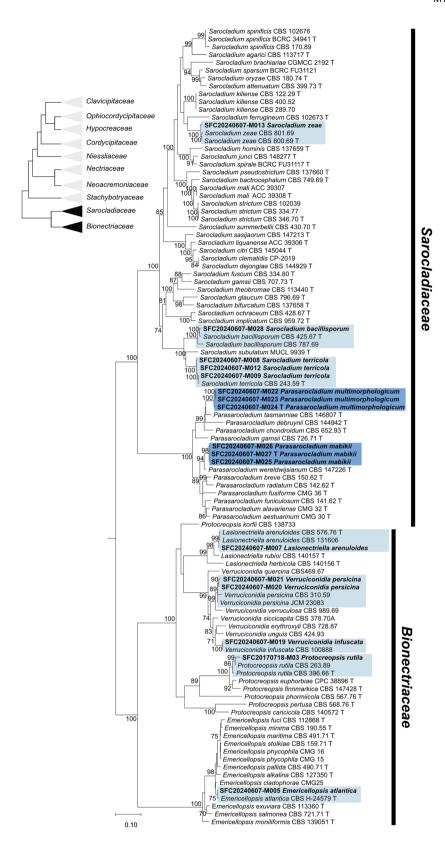


Figure 1. A phylogenetic tree of Bionectriaceae and Sarocladiaceae generated by RAXML analysis using ITS, LSU, TEF1, RPB2, TUB, and ACT. Bootstrap values over 70% are indicated at the branch nodes. Newly generated sequences from this study are represented in bold. The boxes indicate new species in blue and unrecorded in light blue. The summarized tree is illustrated in reference to Hou et al. [35].

holotype (1-2 septa) [54]. Additionally, its conidiophores are more widely distributed, but its conidiogenous cells and conidia are shorter than those of the holotype DAOM 695772 [54].

Acrostalagmus annulatus (Berk. & M.A. Curtis) Seifert (Figure 5(B))

MycoBank: MB518663 Family Hypocreaceae

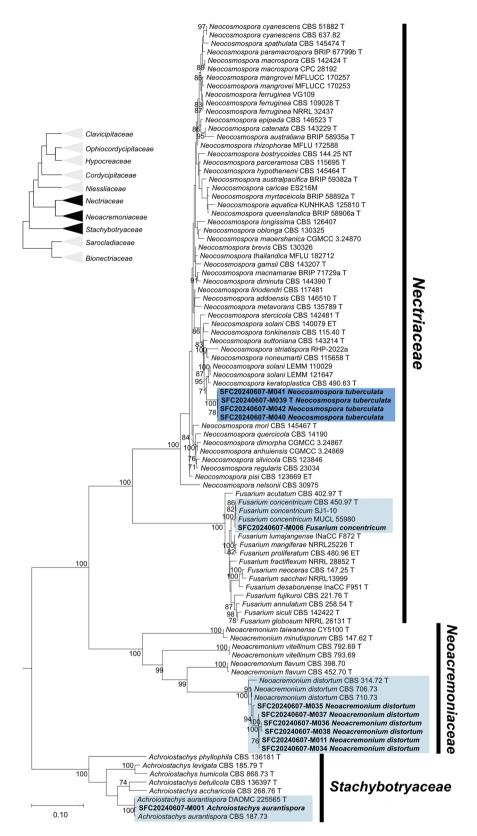


Figure 2. A phylogenetic tree of *Nectriaceae*, *Neoacremoniaceae*, and *Stachybotryaceae* generated by RAXML analysis using ITS, LSU, *TEF1*, and *RPB2*. Bootstrap values over 70% are indicated at the branch nodes. Newly generated sequences from this study are represented in bold. The boxes indicate new species in blue and unrecorded in light blue. The summarized tree is illustrated in reference to Hou et al. [35].

Basionym: Stilbum annulatum Berk. & M.A. Curtis, in Berkeley 1874

Materials examined: South Korea. Jeollanam-do, Suncheon-si (34°50′29.7″N 127°29′09.4″E), January

2020, isolated from sea sand, M.S. Park, J.S. Kim & Y.W. Lim (SFC20240607-M002, stored in a metabolically inactive state); Jeollanam-do, Muan-gun (35°1′38.57″N 126°25′17.06″E), January 2020,

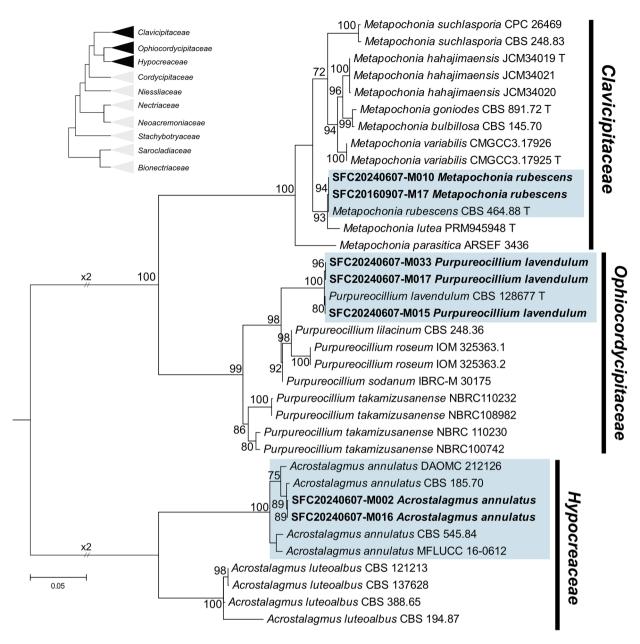


Figure 3. A phylogenetic tree of Clavicipitaceae, Ophiocordycipitaceae, and Hypocreaceae generated by RAxML analysis using ITS, LSU, TEF1, RPB2, and TUB. Bootstrap values over 70% are indicated at the branch nodes. Newly generated sequences from this study are represented in bold. The boxes in light blue indicate unrecorded. The summarized tree is illustrated in reference to Hou et al. [35].

isolated from mudflat, M.S. Park, J.S. Kim & Y.W. Lim (SFC20240607-M016, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on MEA Mycelia smooth, septate, hyaline; Conidiophores repeatedly branched, erect, septate, pale reddish brown, smooth, 2-4.2 µm wide. Conidiogenous cells phialidic, narrowly flask-shaped in the widest part, concolorous, smooth, collarette, $9-27\times1.7-3.9\,\mu\text{m}$. Conidia aseptate, ellipsoidal, cylindrical, smooth, concolorous, guttules, $4.9-7.6\times2.5-3.9\,\mu\text{m}$ (av. $6.1\times3.1\,\mu\text{m}$).

Culture characteristics: Colonies on PDA reaching 17-22 mm diam in 7 d at 25 °C; raised, plain, floccose, hairy, forward white, pale yellow (4A3); margin fimbriate; reverse pastel yellow (3A4).

Colonies on MEA reaching 19-26(-29) mm diam in 7 d at 25°C; flat, plain, hairy, forward white, yellowish white (3A2); margin fimbriate; reverse grayish orange (5B6). Colonies on OA reaching 18-24 mm diam in 7 d at 25°C; flat, plain, hairy, felty, forward white, orange (5B8); margin fimbriate; reverse pale yellow (4A3), light brown (6D8) at center. Colonies on CMA reaching 18-23 mm diam in 7 d at 25 °C; flat, plain, forward yellowish white (1A2), margin entire to irregular; exudate brownish red (8C7); reverse yellowish white (2A2).

Notes: Korean strains, isolated from marine sediments, share similar morphological characteristics with Acrostalagmus annulatus (MFLUCC 16-0612) isolated from decaying fruits [55]. However,



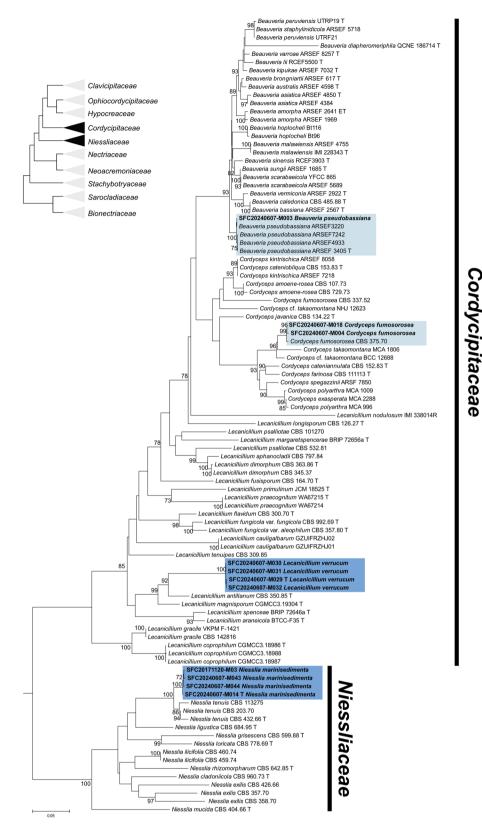


Figure 4. A phylogenetic tree of Cordycipitaceae and Niessliaceae generated by RAxML analysis using ITS, LSU, TEF1, RPB2, and TUB. Bootstrap values over 70% are indicated at the branch nodes. Newly generated sequences from this study are represented in bold. The boxes indicate new species in blue and unrecorded in light blue. The summarized tree is illustrated in reference to Hou et al. [35].

compared to the holotype [56], the Korean strain has slightly wider conidiophores and conidia.

Beauveria pseudobassiana S.A. Rehner **Humber** (Figure 5(C))

MycoBank: MB519125 Family Cordycipitaceae

Materials examined: South Korea. Jeollanam-do, (34°50′29.7″N, 127°29′09.4″E), Suncheon-si 2018, isolated from sea sand, M.S. Park & Y.W. Lim

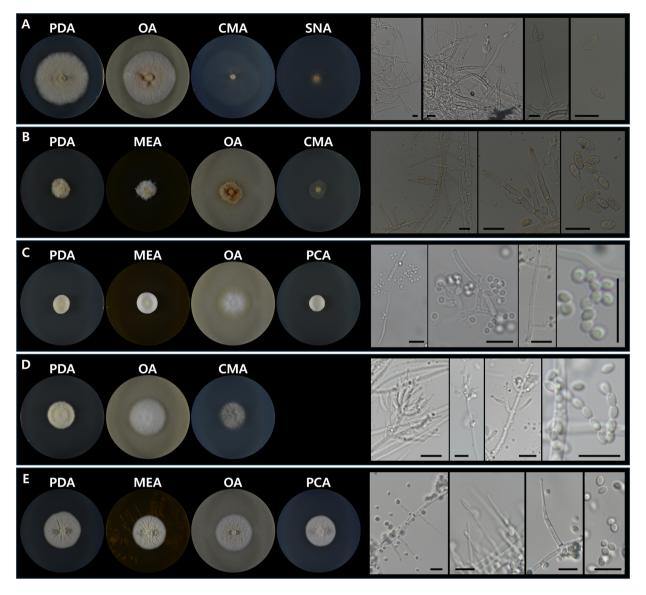


Figure 5. Cultural and conidial morphology. (A) Achroiostachys aurantispora. (B) Acrostalagmus annulatus. (C) Beauveria pseudobassiana. (D) Cordyceps fumosorosea. (E) Emericellopsis atlantica. Scale bar: 10 µm.

(SFC20240607-M003, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on PDA Mycelia smooth, septate, hyaline; Conidiophores branched, erect, septate, hyaline, smooth, 1.3-2.9 µm wide. Conidiogenous cells solitary but usually consisting of dense lateral clusters, base subspherical to ampulliform, hyaline, smooth, 3.8–8.1×1.5–2.8μm. Conidia aseptate, globose, subglobose, smooth, hyaline, $1.6-2.6\times1.2-2.4\mu m$ (av. $2.1\times1.9\mu m$).

Culture characteristics: Colonies on PDA reaching 17-20 mm diam in 7 d at 25 °C; flat to crateriform, plain, cottony, forward white, yellowish white (2A2) at center; margin entire; reverse light yellow (3A5). Colonies on MEA reaching 21-23 mm diam in 7 d at 25 °C; crateriform, radially sulcate, cottony, forward white, yellowish white (2A2) to grey (2B1) at center; margin entire; reverse butter yellow (4A5), brownish orange (7C4) at center. Colonies on OA

reaching 33-37 mm diam in 7 d at 25 °C; flat, floccose, forward yellowish white (3A2), white at center; margin entire; reverse yellowish white (3A2). Colonies on PCA reaching 15-17 mm diam in 7 d at 25 °C; crateriform, plain, cottony, forward white, yellowish white (3A2) at center, margin entire; reverse white, pale yellow (3A3) at center.

Notes: Beauveria pseudobassiana shows intraspecific variation in morphology [57,58]. Korean strains isolated from sea sand have long conidiogenous cells compared to other strains within the species [58].

Cordyceps fumosorosea (Wize) Kepler, B. Shrestha & Spatafora (Figure 5(D))

MycoBank: MB 820980 **Family** Cordycipitaceae

Basionym: Isaria fumosorosea Wize 1904

Materials examined: South Korea. Jeollanam-do, Suncheon-si (34°50'29.7"N 127°29'09.4"E), July 2021, isolated from sea sand, M.S. Park, J.S. Kim & Y.W. Lim (SFC20240607-M004, stored in a metabolically inactive state); Gyeongsangbuk-do, Ulleung-gun (37°32′2.05″N 130°49′24.87″E), unknown date in 2018, isolated from a sponge, M.S. Park & Y.W. Lim (SFC20240607-M018, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on PDA *Mycelia* smooth, septate, hyaline; *Conidiophores* branched, erect, septate, hyaline, smooth, $1.6-2.8\,\mu\text{m}$ wide. *Phialides* ampulliform, subcylindrical tapering into the apex, subulate in immature, hyaline, smooth, solitary or in whorls of 2-4 on each branch, up to five phialides on a whorl, $6.8-18\times1.4-3.4\,\mu\text{m}$. *Conidia* aseptate, ellipsoidal, smooth, hyaline, $1.9-3.6\times1.2-2.3\,\mu\text{m}$ (av. $2.8\times1.8\,\mu\text{m}$).

Culture characteristics: Colonies on PDA reaching 29–30 mm diam in 7 d at 25 °C; umbonate, raised plain, floccose, velvety at margin, forward white; margin entire; reverse butter yellow (4A5). Colonies on OA reaching 36–39 mm diam in 7 d at 25 °C; raised, floccose, forward white; margin entire; reverse yellowish white (4A2). Colonies on CMA reaching 27–32 mm diam in 7 d at 25 °C; flat, plain, floccose, felty, forward white to hyaline; margin filiform; reverse yellowish white (3A2).

Notes: Cordyceps fumosorosea, formerly Isaria fumosorosea [59], is recognized to infect many pest species [60]. It is very interesting that this species was isolated from sponge in the marine environment.

Emericellopsis atlantica L.W. Hou, Crous, Rämä & Hagestad (Figure 5(E))

MycoBank: MB838493 Family *Bionectriaceae*

Materials examined: South Korea. Jeollanam-do, Muan-gun (35°1′38.57″N 126°25′17.06″E), July 2021, isolated from mudflat, M.S. Park, J.S. Kim & Y.W. Lim (SFC20240607-M005, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on OA *Mycelia* smooth, septate, hyaline; *Conidiophores* repeatedly branched, erect, septate, hyaline, smooth, $1.3-3.6\,\mu m$ wide. *Conidiogenous cells* phialidic, narrowly flask-shaped in the widest part, hyaline, smooth, $16-44\times1.5-3.2\,\mu m$. *Conidia* aseptate, broadly ellipsoidal, sometimes obclavate, smooth, hyaline, mostly 1 globose, or subglobose, guttule, $2.7-4.2\times2-3.6\,\mu m$ (av. $3.5\times2.7\,\mu m$).

Culture characteristics: Colonies on PDA reaching 36–40 mm diam in 7 d at 25 °C; flat, radially sulcate, felty, velvety at center, forward yellowish white (1A2); margin entire; reverse pale yellow (3A3). Colonies on MEA reaching 34–36 mm diam

in 7 d at 25°C; flat, radially sulcate, velvety, felty at margin, forward white, yellowish white (4A2) at center; margin entire; exudate clear; reverse orange (6B7). Colonies on OA reaching 38–39 mm diam in 7 d at 25°C; flat, felty, forward white, yellowish white (4A2) at center; margin entire; exudate clear; reverse pastel yellow (3A4). Colonies on PCA reaching 30–32 mm diam in 7 d at 25°C; flat, slightly radially sulcate, felty; forward yellowish white (4A2), margin entire; reverse white, pale yellow (3A3) at center.

Notes: The Korean strain SFC20240607-M005 matches the holotype of *Emericellopsis atlantica*, a member of the marine clade within the genus. However, there are several differences in their conidial structures: the conidiogenous cells of SFC20240607-M005 are shorter than those of the holotype ($vs.\ 24.5-50(-64)\ \mu m$) [61]. Additionally, the conidia of SFC20240607-M005 are shorter and wider ($vs.\ 3-6(-9)\times 2-2.5\,\mu m$). The guttules in the conidia of SFC20240607-M005 are globose or subglobose, while those of the holotype are irregular in shape [61].

Fusarium concentricum Nirenberg & O'Donnell (Figure 6(A))

MycoBank: MB 444884 Family Nectriaceae

Materials examined: South Korea. Jeollanam-do, Suncheon-si (34°50′51.8″N 127°29′31.8″E), January 2017, isolated from mudflat, M.S. Park & Y.W. Lim (SFC20240607-M006, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on PCA Mycelia septate, smooth, hyaline; Conidiophores septate, branched, erect, smooth, hyaline, 1.5-3.3 µm wide. Phialides lateral or terminal, subulate to subcylindrical, cylindrical, hya- $6.6-32 \times 1.9-3 \,\mu m$ line, smooth, polyphialides observed. Microconidia 0-1 septate, subcylindrical, smooth, hyaline, $5-13\times1.5-3\,\mu\text{m}$, solitary or in *Sporodochia* 0-5(-6)-septate, straight slightly curved, broadest at the half and tapering toward both ends, foot-shaped, abundant guttules, $9.8-59\times1.9-4.3\,\mu\text{m}$. *Chlamydospores* not observed.

Culture characteristics: Colonies on PDA reaching 76–80 mm diam in 7 d at 25 °C; flat, plain, floccose, forward white; margin fimbriate; reverse grayish yellow (4B3). Colonies on OA reaching 90 mm diam in 7 d at 25 °C; flat to raised, plain, floccose, forward white; margin entire; reverse yellowish grey (4B2). Colonies on SNA reaching 67–73 mm diam in 7 d at 25 °C; flat, plain, felty, forward yellowish white (4A2); margin entire; reverse pale yellow (4A3).

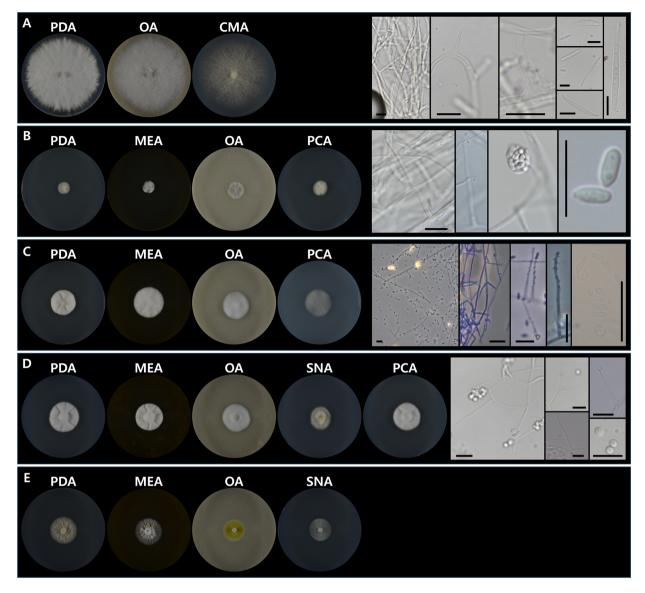


Figure 6. Cultural and conidial morphology. (A) Fusarium concentricum. (B) Lasionectriella arenuloides. (C) Lecanicillium verrucum sp. nov. (D) Metapochonia rubescens. (E) Neoacremonium distortum. Scale bar: 10 µm.

Notes: False heads are not observed in Korean strain SFC20240607-M006, but are observed in the holotype of this species [62].

Lasionectriella arenuloides (Samuels) L.W. Hou, L. Cai & Crous (Figure 6(B))

MycoBank: MB 845846 Family Bionectriaceae

Basionym: Nectria arenuloides Samuels, New Zealand J. Bot. 14: 254. 1976

Synonym: Hydropisphaera arenuloides (Samuels) Rossman & Samuels, Stud. Mycol. 42: 30. 1999.

Lasionectriella marigotensis (Lechat & J. Fourn.) L.W. Hou, L. Cai & Crous.

Materials examined: South Korea. Jeollanam-do, Suncheon-si (34°50′29.7"N 127°29′09.4"E), January 2020, isolated from sea sand, M.S. Park, J.S. Kim & Y.W. Lim (SFC20240607-M007, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on MEA Mycelia smooth, septate, hyaline; Conidiophores branched, erect, septate, hyaline, smooth, 1.3-2 µm wide. Phialides, subulate, hyaline, smooth, $24-50\times1.8-2.3\,\mu\text{m}$. Conidia aseptate, ellipsoidal to subcylindrical, smooth, hyaline, forming a mucous, rounded head at the tip of phialide, guttules, $3.1-5.8\times1.6-2.3\,\mu\text{m}$ (av. $4.4\times1.9\,\mu\text{m}$).

Culture characteristics: Colonies on PDA reaching 13-15 mm diam in 7 d at 25 °C; flat, plain, rugose, membranous, forward yellowish white (2A2); margin filiform; reverse pale yellow (2A3). Colonies on MEA reaching 11-13 mm diam in 7 d at 25 °C; raised, plain, felty to floccose, forward white; margin fimbriate; reverse light yellow (4A4). Colonies on OA reaching 17-19 mm diam in 7 d at 25 °C; flat, plain, felty, hairy, forward white; margin filiform; reverse yellowish white (2A2). Colonies on PCA reaching 15-17 mm diam in 7 d at 25 °C; flat, plain, rugose, hairy, membranous, forward white; margin entire to filiform; reverse pale yellow (2A3).

Notes: Lasionectriella arenuloides and L. marigotensis were recently transferred to the genus Lasionectriella and exhibited little differences in morphological features and four genetic markers (ITS, LSU, TEF1, and RPB2) [35]. In this study, we synonymized Lasionectriella marigotensis as Lasionectriella arenuloides because the polyphasic analysis produced similar results to those of the previous study [35]. The conidia of the Korean strain SFC20240607-M007 are bigger than those of CBS131606 (previous L. marigotensis) [53].

Lecanicillium verrucum Wonjun Lee & Y.W. Lim, sp. nov. (Figure 6(C))

MycoBank: MB 855663 **Family** *Cordycipitaceae*

Etymology: Referring to the phialides that form warts at the upper part.

Typus: South Korea. Gyeongsangbuk-do, Pohang-si (36°15′07″N 129°22′31″E), August 2015, isolated from sea sand, M.S. Park & Y.W. Lim (**holotype** SFC20240607-M029, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on PDA *Mycelia* smooth, septate, hyaline; *Conidiophores* branched, erect, septate, hyaline, 1–2.2 μm wide. *Phialides* narrowly flask-shaped in the widest part, hyaline, rough-walled with warts at the upper part, $11-27\times0.9-2$ μm, polyphialides observed. *Conidia* aseptate, ellipsoidal, oblong-ellipsoidal, smooth, hyaline, $2.9-5.4\times1.5-2.4$ μm (av. 4.1×1.9 μm).

Culture characteristics: Colonies on PDA reaching 25–29 mm diam in 7 d at 25 °C; raised, radially sulcate, velvety, forward white; margin irregular; exudate clear; reverse pastel yellow (2A4). Colonies on MEA reaching 31–33 mm diam in 7 d at 25 °C; raised, radially sulcate, velvety, forward white; margin entire; reverse light orange (5A4). Colonies on OA reaching 27–29 mm diam in 7 d at 25 °C; convex, plain, floccose, forward white; margin entire; reverse yellowish white (3A2). Colonies on SNA reaching 27–29 mm diam in 7 d at 25 °C; flat, plain, velvety, forward white; margin entire; reverse hyaline to white.

Additional materials examined: South Korea. Jeju-do, Chuja-myeon (33°56′31″N 126°18′50″E), 31 August 2021, isolated from macroalga, M.S. Park, J.S. Kim & Y.W. Lim (SFC20240607-M030, stored in a metabolically inactive state); *Ibid.* (SFC20240607-M031, stored in a metabolically inactive state); Incheon, Jung-gu (37°21′54.0″N 126°31′40.8″E), January 2019, isolated from unknown substrate, M.S. Park & Y.W.

Lim (SFC20240607-M032, stored in a metabolically inactive state).

Notes: *Lecanicillium verrucum* is phylogenetically related to *L. antillanum*. On PDA, *L. verrucum* colonies grow faster than those of *L. antillanum* CBS 350.85 (*vs.* 18mm in 10d) and have entire, regular margins compared to the irregular margins of *L. antillanum* [63]. *Lecanicillium verrucum* is characterized by rough phialides with warts and does not produce fusiform primary conidia, unlike *L. antillanum*.

Metapochonia rubescens (Zare, W. Gams & López-Llorca) Kepler, S.A. Rehner & Humber (Figure 6(D))

MycoBank: MB 806075 **Family** *Clavicipitaceae*

Basionym: *Pochonia rubescens* Zare, W. Gams & López-Llorca, Nova Hedwigia 73:69, 2001.

Materials examined: South Korea. Jeollanam-do, Suncheon-si (34°50′29.7″N 127°29′09.4″E), July 2021, isolated from sea sand, M.S. Park, J.S. Kim & Y.W. Lim (SFC20240607-M010, stored in a metabolically inactive state); Gangwon-do, Gangneung-si (37°51′30.82″N 128°51′15.90″E), January 2015, isolated from sandfish egg, M.S. Park & Y.W. Lim (SFC20160907-M17=SFC102204, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on PCA *Mycelia* smooth, septate, hyaline; *Conidiophores* branched, erect, septate, hyaline, smooth, $1.1-2.3 \,\mu\text{m}$ wide. *Phialides* terminal, lateral, narrowly flask-shaped in the widest part, hyaline, smooth, $15-28\times1.1-2.3 \,\mu\text{m}$, schizophialides observed. *Conidia* aseptate, globose, smooth, hyaline, $2.2-3.2\times2-3.1 \,\mu\text{m}$ (av. $2.7\times2.6 \,\mu\text{m}$).

Culture characteristics: Colonies on PDA: reaching 29-32 mm diam in 7 d at 25 °C; umbonate, radially sulcate, velvety, forward white; margin entire; reverse pale yellow (2A3). Colonies on MEA reaching 28-29 mm diam in 7 d at 25 °C; umbonate, radially sulcate, velvety, forward yellowish white (1A2) to white; margin entire; reverse orange (5B8). Colonies on OA reaching 26-29 mm diam in 7 d at 25°C; umbonate, plain, velvety, forward white; margin entire; reverse yellowish white (2A2). Colonies on SNA reaching 21-25 mm diam in 7 d at 25 °C; raised due to abundant aerial mycelium, plain, floccose, felty, forward white, pale yellow (2A3) at center; margin entire; reverse white, pale yellow (2A3). Colonies on PCA reaching 27-29 mm diam in 7 d at 25°C; umbonate, radially sulcate, velvety, forward white; margin entire; reverse pale yellow (2A3).

Notes: The strain SFC102204, isolated from a yellow sandfish egg, was initially identified as



Metapochonia suchlasporia based on TUB and was reported to have endoglucanase and gelatinase activities [41]. However, in this study, it was identified as M. rubescens through phylogenetic analysis using ITS, TEF1, and RPB2.

Neoacremonium distortum L.W. Hou, L. Cai & Crous (Figure 6(E))

MycoBank: MB 845811 Family Neoacremoniaceae

Materials examined: South Korea. Ganghwa-gun (37°36'36.3"N 126°31'13.8"E), January 2017, isolated from mudflat, M.S. Park & Y.W. Lim (SFC20240607-M011, stored in a metabolically inactive state); Ibid. July 2018, isolated from mudflat, M.S. Park & Y.W. Lim (SFC20240607-M038, stored in a metabolically inactive state); Ibid. April 2017, isolated from mudflat, M.S. Park & Y.W. Lim (SFC20240607-M036, stored in a metabolically inactive state); Ibid. (SFC20240607-M037, stored in a metabolically inactive state); Ganghwa-gun (37°35'33.72" 126°27'30.29"), October 2016, isolated from sea sand, M.S. Park, J.S. Kim & Y.W. Lim (SFC20240607-M034, stored in a metabolically inactive state); Jeollanam-do, Muan-gun (35°3'43.65" 126°20′13.91"), October 2016, isolated from sea sand, M.S. Park & Y.W. Lim (SFC20240607-M035, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph not observed on PDA, MEA, OA, SNA, and PCA.

Culture characteristics: Colonies on PDA reaching 27-29 mm diam in 7 d at 25 °C; flat, radially sulcate, rugose, forward white to butter yellow (4A5); margin fimbriate; reverse yellowish white (3A2). Colonies on MEA reaching 25-29 mm diam in 7 d at 25°C flat, radially sulcate, rugose, floccose at center, forward white to hyaline; margin fimbriate; reverse light orange (5A4). Colonies on OA reaching 22-25 mm diam in 7 d at 25 °C; flat, plane, sometimes radially sulcate, membranous, moist, forward vivid yellow (2A8), white at margin, sometimes partly vivid yellow (2A8); margin entire; reverse pale yellow (3A3). Colonies on PCA reaching 23-25 mm diam in 7 d at 25°C; flat, plain, hairy, forward white to hyaline; margin filiform; reverse yellowish white (1A2).

Notes: *Neoacremonium distortum* is known for its intraspecific variation [35]. On OA media, Korean strains form vivid yellow colonies, whereas the holotype CBS H-6647 forms white colonies with a buff pigment [35]. Korean strains exhibit fimbriate margins on PDA and MEA, and filiform margins on PCA, while the holotype has entire margins on OA, MEA, and PDA [35]. Korean strains also show radially sulcate colonies on PDA and MEA, whereas the holotype displays only a lightly radially sulcate trait on PDA as shown in [35]. The conidial structures of Korean strains were not observed on any of the cultured media (PDA, MEA, OA, SNA, and PCA). Korean strains are genetically distinct from the other strains. The variations are 4-7 bp for ITS (515 bp), 13-15 bp for TEF1 (808 bp), and 7 bp for RPB2 (750 bp), while LSU does not show significant differences between Korean strains and the others.

Neocosmospora tuberculata Wonjun Lee & Y.W. Lim \, (Figure 7(A))

MycoBank: MB 855664 Family Nectriaceae

Etymology: Referring to forming tuberculate chlamydospores.

Typus: South Korea. South Sea of Korea (34°27'42.4"N 128°15'34.7"E), August 16 2022, isolated from seawater in 40 m depth (bottom), Wonjun Lee, J.W. Lee & Y.W. Lim (holotype SFC20240607-M039, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on OA Mycelia smooth, septate, hyaline; Conidiophores branched, erect, septate, hyaline, smooth, 1.6-4.2 µm wide. Phialides subulate to subcylindrical, cylindrical, non-flared or flared collarette, hvaline, smooth, $38-148 \times 1.4-4.3 \,\mu\text{m}$ polyphialides observed. Conidia 0-3-septate, subcylindrical, cylindrical, ellipsoidal, smooth, hyaline, gently curved due to forming a false head on phialides, with or without papillate, $5.9-27\times2-6\,\mu\text{m}$. abundant, globose, subglobose, Chlamydospores mostly tuberculate, terminal or intercalary in hyphae, solitary, in chains, or in clusters, 5.4–12 μm diam. Sporodochia not observed.

Culture characteristics: Colonies on PDA reaching 66-71 mm diam in 7 d at 25 °C; flat, plain, floccose, hairy, felty forward white to dull blue (23E4), amber yellow (4B6) at margin, sometimes pale red (12A3) at center; margin entire to irregular, filiform; exudate yellow (2A7), sometimes pink (11A5), soluble pigment blond (4D4) when mature; reverse garnet brown (9D8) to buttercup yellow (4A7). Colonies on MEA reaching 42-46 mm diam in 7 d at 25 °C; flat to raised, plain, floccose, hairy, sometimes felty, forward grayish green (29B3), having blackish blue (20F7) to grayish green (29C4) patches, sometimes brownish red (10D6) patches; margin entire; soluble pigment some strains pastel red (10A5) when mature; reverse English red (8D8), partly reddish brown (8E8), light orange (5A4) at margin. Colonies on OA reaching 50-55 mm diam in 7 d at 25 °C; flat, raised, plain, floccose, hairy, felty, forward yellowish white

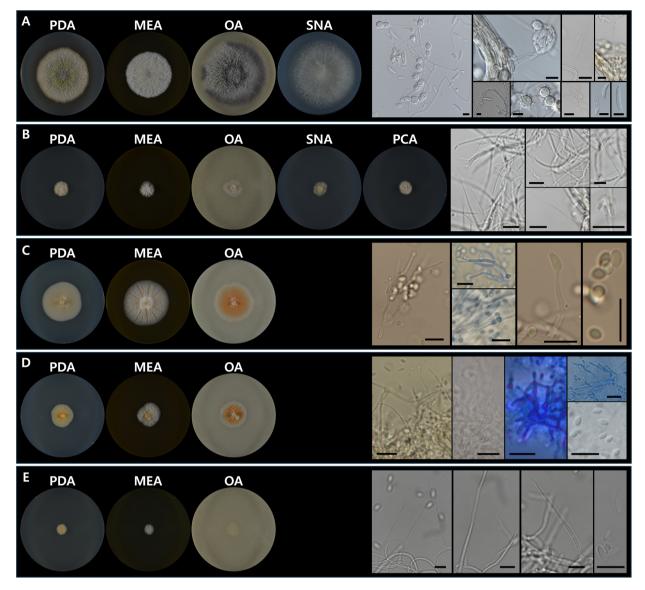


Figure 7. Cultural and conidial morphology. (A) *Neocosmospora tuberculata* sp. nov. (B) *Niesslia marinisedimenta* sp. nov. (C) *Parasarocladium mabikii* sp. nov. (D) *Parasarocladium multimorphologicum* sp. nov. (E) *Protocreopsis rutila*. Scale bar: 10 μm.

(3A2) to dark blue (21F4), pastel grey (15C1) to dull violet (15E4); margin entire; exudate pink (11A5) when mature; reverse golden brown (5D7) to dark brown (8F8), reddish brown (9D4). Colonies on SNA reaching 64–69 mm diam in 7 d at 25 °C; raised due to aerial mycelia, plain, hairy, felty, forward white to hyaline; margin entire; reverse hyaline to yellowish white (1A2).

Additional materials examined: South Korea. South Sea of Korea (34°27′42.4″N 128°15′34.7″E), August 16 2022, isolated from seawater in 40 m depth (bottom), Wonjun Lee, J.W. Lee & Y.W. Lim (SFC20240607-M040, stored in a metabolically inactive state); *Ibid.* (SFC20240607-M040, stored in a metabolically inactive state); *Ibid.* (SFC20240607-M041, stored in a metabolically inactive state); *Ibid.* (SFC20240607-M042, stored in a metabolically inactive state).

Notes: Neocosmospora tuberculata is phylogenetically related to N. solani and N. keratoplastica, but it differs from both species in several characteristics. N. tuberculata forms chlamydospores solitarily, in chains, or in clusters, whereas N. solani produces chlamydospores that are one or two-celled [64]. Additionally, N. tuberculata produces tuberculate chlamydospores, whereas those of N. keratoplastica are smooth to rough [65]. The chlamydospores of N. tuberculata are also larger, measuring 6.5–8.5 µm compared to 6–8 µm in N. solani [64] and N. keratoplastica [65].

Niesslia marinisedimenta Wonjun Lee & Y.W. Lim, (Figure 7(B))

MycoBank: MB 855665
Family Niessliaceae

Etymology: Referring to the marine sediments (mudflat and sea sand) from which it was isolated.



Typus: South Korea. Jeollanam-do, Muan-gun (35°3'43.65"N 126°20'13.91"E), October 2016, isolated from sea sand, M.S. Park & Y.W. Lim (holotype SFC20240607-M014, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on PCA Mycelia smooth, septate, hyaline; Conidiophores branched, erect, septate, hvaline, smooth, 1.2-2.6 µm wide. Phialide subulate, hyaline, smooth, $27-47 \times 1.1-2.4 \,\mu\text{m}$. Conidia aseptate, oblong-ellipsoidal, sometimes cylindrical, smooth, hyaline, $2.5-6.6\times0.9-1.8\,\mu\text{m}$ (av. $3.5\times1.3\,\mu\text{m}$).

Culture characteristics: Colonies on PDA reaching 15-17 mm diam in 7 d at 25 °C; raised, radially sulcate, velvety, forward white to grayish yellow (4B4); margin lobed; reverse pale yellow (3A3). Colonies on MEA reaching 15-20 mm diam in 7 d at 25 °C; raised, radially and concentrically sulcate, felty, downy, forward white, grayish yellow (4B3); margin lobed, filiform; reverse reddish orange (7B8). Colonies on OA reaching 15-20 mm diam in 7 d at 25°C; flat, plain, membranous, granular, forward white; margin irregular; reverse yellowish white (3A2). Colonies on SNA reaching 14-18 mm diam in 7 d at 25 °C; flat, plain, membranous, granular, forward white to lemon (3B8); margin entire; reverse pale yellow (3A3). Colonies on PCA reaching 15-17 mm diam in 7 d at 25 °C; flat, radially and concentrically sulcate, velvety, membranous, forward white to lemon (3B8); margin entire, fimbriate; reverse pale yellow (3A3).

Additional materials examined: South Korea. Jeollanam-do, Muan-gun (35°3'43.65"N 126°20'13.91"E), October 2016, isolated from sea sand, M.S. Park & Y.W. Lim (SFC20171120-M03, stored in a metabolically inactive state); Ibid. (SFC20240607-M043, stored in a metabolically inactive state); Ibid. Incheon, Ganghwa-gun (37°36'36.3"N 126°31′13.8"E), January 2019, isolated from mudflat, M.S. Park & Y.W. Lim (SFC20240607-M044, stored in a metabolically inactive state).

Notes: Niesslia marinisedimenta is phylogenetically closely related to N. tenuis but can be distinguished by the following characteristics. Niesslia marinisedimenta does not form white to pinkish colonies on MEA, but N. tenuis does [66]. Additionally, N. marinisedimenta does not sporulate, N. tenuis produces conidia on MEA [66].

Parasarocladium mabikii Wonjun Lee & Y.W. Lim, (Figure 7(C))

MycoBank: MB 855666 **Family** Sarocladiaceae

Etymology: Referring to the name of the National Marine Biodiversity Institute of Korea (MABIK).

MABIK is an institute that investigates Korean marine organisms and strives to unveil a variety of marine organisms.

Typus: South Korea. Incheon, Ganghwa-gun (37°36'36.3"N 126°31'13.8"E), January 2021, isolated from mudflat, M.S. Park, J.S. Kim & Y.W. Lim (holotype SFC20240607-M027, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on PDA Mycelia smooth, septate, hyaline, abundant guttules; Conidiophores solitary or aggregated, arising directly from aerial or substratal mycelium, branched, erect, straight or irregularly curved, septate, hyaline, smooth, bearing 1-2 levels with 1-3 phialides per node, showing conidiogenous cells as lateral, cylindrical, asymmetrical projections, guttules, 1.1-3 µm wide. Phialides lateral or terminal, narrowly flask-shaped in the widest part, subulate to cylindrical, hyaline, smooth, terminal or subterminal proliferation, monophialides, polyphialides with up to three conidiogenous loci, frequently trident-form, guttules, 9.4-48×1.4-3 μm. Conidia aseptate, ellipsoidal, obclavate, smooth, hyaline, eguttulate, truncated base, 2.8- $6.4\times2-3.5\,\mu\text{m}$ (av. $3.9\times2.7\,\mu\text{m}$). Chlamydospores not observed.

Culture characteristics: Colonies on PDA reaching 38-40 mm diam in 7 d at 25 °C; flat, plain, membranous, felty, forward orange white (5A2), white at margin; margin filiform; reverse pale yellow (3A3). Colonies on MEA reaching 44-48 mm diam in 7 d at 25°C; flat, radially sulcate, felty, cottony at center, forward orange white (5A2) to pinkish white (8A2), white at center due to aerial mycelia; margin filiform; reverse pale orange (5A3). Colonies on OA reaching 41-44 mm diam in 7 d at 25 °C; flat, plain, granular, moist, forward light orange (5A5), white at margin; margin entire; reverse yellowish white (4A2).

Additional materials examined: South Korea. Incheon, Ganghwa-gun (37°36′36.3"N 126°31′13.8"E), July 2016, isolated from mudflat, M.S. Park & Y.W. Lim (SFC20240607-M025, stored in a metabolically inactive state); Ibid. (SFC20240607-M026, stored in a metabolically inactive state).

Notes: Parasarocladium mabikii is closely related to P. wereldwijsianum but differs in several aspects. The former species grow faster on OA (vs 30-35 mm diam) and MEA (vs. 30mm diam) compared to P. wereldwijsianum [67]. The conidia of P. mabikii are shorter than those of *P. wereldwijsianum* (vs. 4–10 μm long) [67]. Additinally, P. mabikii contains distinct insertions in TEF1 genetic marker, which are absent in P. wereldwijsianum [67].

Parasarocladium multimorphologicum Wonjun Lee & Y.W. Lim, (Figure 7(D))

MycoBank: MB 855668 Family Sarocladiaceae

Etymology: Referring to the morphological characteristics of colonies on MEA media.

Typus: South Korea. Jeju-do, Chuja-myeon (33°57′11.88″N 126°18′07.56″E), August 31 2021, isolated from *Grateloupia* sp. (Macroalga), M.S. Park, J.S. Kim & Y.W. Lim (**holotype** SFC20240607-M024, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on PDA *Mycelia* smooth, septate, hyaline; *Conidiophores* reduced to conidiogenous cells, aggregated, arising directly from aerial or substratal mycelium, branched, erect, septate, hyaline, smooth, $10-34\times1.1-2.1\,\mu\text{m}$, polyphialidic structure observed. *Conidia* aseptate, ellipsoidal, oblong-ellipsoidal, smooth, hyaline, $2.3-4.7\times1.2-2\,\mu\text{m}$ (av. $3.3\times1.6\,\mu\text{m}$). *Chlamydospores* not observed.

Culture characteristics: Colonies on PDA reaching 22-25 mm diam in 7 d at 25°C; flat, radially sulcate, membranous, felty, cerebriform at center, sometimes rugose, forward light orange (5A5), yellowish white (3A2) to pale yellow (3A3); margin lobed; reverse pale yellow (4A3). Colonies on MEA reaching 17-24 mm diam in 7 d at 25 °C; raised, plain, radially sulcate in some part, rugose, felty, cerebriform, sometimes membranous, felty to floccose in SFC20240607-M023, forward pale yellow (2A3), white at margin, orange white (5A2) to white; margin irregular, lobed filiform; reverse light orange (6A4). Colonies on OA reaching 24-28 mm diam in 7 d at 25 °C; flat, plain, sometimes radially sulcate, rugose, felty, sometimes membranous, forward light orange (5A5), white at margin; margin entire, sometimes irregular; reverse orange white (5A2).

Additional materials examined: South Korea. Jeju-do, Chuja-myeon (33°56′31″N 126°18′50″E), August 31 2021, isolated from mudflat, M.S. Park, J.S. Kim & Y.W. Lim (SFC20240607-M023, stored in a metabolically inactive state); *Ibid.* (SFC20240607-M022, stored in a metabolically inactive state).

Notes: Parasarocladium multimorphologicum forms a distinct clade, separate from other species, such as *P. tasmanniae*, *P. debruynii*, *P. chondroidum*, and *P. gamsii* (Figure 1). The colonies of *P. multimorphologicum* grow slowly on OA and MEA compared to those of *P. debruynii* (vs. 45–55 mm and 38–50 mm diam, respectively) [68]. Among closely related species, *P. gamsii* is considered a marine fungus [16,69].

Protocreopsis rutila (W. Gams) L.W. Hou, L. Cai

& Crous (Figure 7(E)) MycoBank: MB 845853 Family Bionectriaceae Basionym: Acremonium rutilum W. Gams 1971

Materials examined: South Korea. Incheon, Ganghwa-gun (37°35′33.72″N 126°27′30.29″E), July 2016, isolated from sea sand, M.S. Park & Y.W. Lim (SFC20170718-M03, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on OA Mycelia smooth, septate, hyaline; Conidiophores branched, erect, septate, hyaline, smooth, 1.5-3.6 µm wide, degraded conidiophores not observed. Phialides subulate, hyaline, smooth, 39-70×2.2-3.8 µm at the base, polyphialides not observed. Conidia aseptate, ellipsoidal, oblong-ellipsoidal, $4-6.9 \times 1.4$ smooth, hyaline, $2.6 \,\mu \text{m}$ (av. $5.3 \times 2 \,\mu \text{m}$).

Culture characteristics: Colonies on PDA reaching 10–11 mm diam in 7 d at 25 °C; flat, radially sulcate, rugose, membranous, forward light yellow (4A4); margin lobed; reverse pastel yellow (3A4). Colonies on MEA reaching 10–11 mm diam in 7 d at 25 °C; flat, radially sulcate, rugose, membranous, forward yellowish white (3A2); margin entire; reverse light orange (5A4). Colonies on OA reaching 13–15 mm diam in 7 d at 25 °C; flat, plain, membranous, forward yellowish white (3A2); margin entire; reverse yellowish white (2A2).

Notes: Korean strain SFC20170718-M03 forms yellowish-white without pigments on PDA media. On the other hand, the holotype CBS 394.70 forms colonies with a sienna center, peach middle, and salmon periphery, producing a yellowish pigment [35]. Additionally, moist slimy heads of conidia and degenerated conidiophores, observed in CBS 394.70 [35], are absent in the Korean strain.

Purpureocillium lavendulum Perdomo, Dania García, Gené, Cano & Guarro (Figure 8(A))

MycoBank: MB 561126 **Family** *Ophiocordycipitaceae*

Materials examined: South Korea. South Sea of Korea (34°27'42.4"N 128°15'34.7"E), August 16 2022, isolated from seabed sediment 40 m deep, Wonjun Lee, J.W. Lee & Y.W. (SFC20240607-M017, stored in a metabolically inactive state); Jeollanam-do, Muan-gun (35°3'43.65"N 126°20′13.91"E), July 2021, isolated from sea sand, M.S. Park & Y.W. Lim (SFC20240607-M033, stored in a metabolically inactive state); Gangwon-do, Goseong-gun (38°28′53.0″N 128°26′18.1″E), January 2020, isolated from sea sand, M.S. Park, J.S. Kim & Y.W. Lim (SFC20240607-M015, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on MEA *Mycelia* smooth, septate, hyaline; *Conidiophores* erect, septate, hyaline, smooth

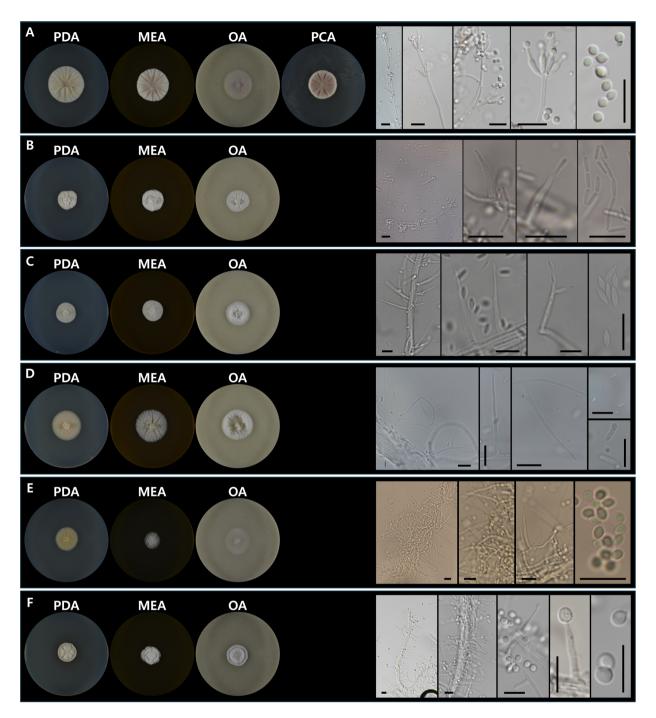


Figure 8. Cultural and conidial morphology. (A) Purpureocillium lavendulum. (B) Sarocladium bacillisporum. (C) Sarocladium terricola. (D) Sarocladium zeae. (E) Verruciconidia infuscata. (F) Verruciconidia persicina. Scale bar: 10 µm.

or rough-walled stipe, 1-2.2 µm wide, 3-6 phialides per whorl. Phialides terminal, lateral, cylindrical, ampulliform, hyaline, smooth, $6.3-14\times1.8-3\,\mu\text{m}$, Acremoniuim-like synanamorph observed. Conidia aseptate, globose, subglobose, sometimes broadly ellipsoidal, smooth, hyaline, $1.8-2.8\times1.7-2.5\,\mu m$ (av. $2.3 \times 2.1 \,\mu\text{m}$).

Culture characteristics: Colonies on PDA reaching 37-39 mm diam in 7 d at 25 °C; flat, radially sulcate, velvety, powdery at grayish Magenta center and sulcate, forward pale yellow (2A3), grayish Magenta (13B3) at center; margin entire; reverse light yellow (3A5), yellowish white (3A2) at margin, reddish lilac (14B4) in age. Colonies on MEA reaching 34-36 mm diam in 7 d at 25°C; flat, radially sulcate, velvety, forward reddish lilac (14B4), yellowish white (2A2) at middle, white at margin; margin entire; reverse golden yellow (5B7). Colonies on OA reaching 30-32 mm diam in 7 d at 25 °C; flat, plain, velvety, granular, forward purplish pink (14A4), yellowish white (3A2) thinly at middle, white at margin; margin entire; reverse pastel yellow (2A4). Colonies on PCA reaching 27-31 mm diam in 7 d at 25°C; flat, radially sulcate, velvety, powdery, forward reddish lilac (14B4), white at margin; margin entire; reverse lemon (3B8).

Notes: Purpureocillium lavendulum forms acremonium-like phialides and ampuliform phialides with longer necks at the apex compared to Penicillium [70]. The stipe ornamentation of *P. lavendulum* (CBS 128677) is well observed under an electron microscope [71], but the Korean strains were not clear visible by a light microscope. However, the overall growth morphology of the Korean strains was similar to CBS 128677 [70].

Sarocladium bacillisporum (Onions & G.L. Barron) Summerb. (Figure 8(B))

MycoBank: MB 519589 Family Sarocladiaceae

Basionym: *Paecilomyces bacillisporus* Onions & G.L. Barron 1967

Materials examined: South Korea. Gangwon-do, Goseong-gun (38°28′53.0″N 128°26′18.1″E), July 2018, isolated from sea sand, M.S. Park & Y.W. Lim (SFC20240607-M028, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on OA *Mycelia* smooth, septate, hyaline; *Conidiophores* solitary or aggregated, erect, septate, hyaline, smooth, $1.3-3.3 \,\mu\text{m}$ wide. *Phialides* subulate, hyaline, smooth, polyphialides not observed, $11-27\times1.6-3.3 \,\mu\text{m}$ at the base. *Conidia* aseptate, cylindrical, rod-shaped, straight, smooth, hyaline, eguttulate, truncate, arranged in chain, $3.7-5.4\times0.9-1.6 \,\mu\text{m}$ (av. $4.5\times1.2 \,\mu\text{m}$).

Culture characteristics: Colonies on PDA reaching 19–21 mm diam in 7 d at 25 °C; raised, radially sulcate, felty, floccose, forward white to yellowish white (3A2); margin fimbriate; exudate clear; reverse pale yellow (4A3). Colonies on MEA reaching 19–22 mm diam in 7 d at 25 °C verrucose, slightly radially sulcate, felty, floccose, forward white; margin fimbriate; exudate clear; reverse light orange (5A4). Colonies on OA reaching 20–23 mm diam in 7 d at 25 °C; flat, radially sulcate, granular, felty, forward white; margin entire; reverse yellowish white (4A2).

Notes: The Korean strain SFC20240607-M028 grows faster than the strains (20–24 mm diameter in 14 d) described by Giraldo et al. [36]. SFC20240607-M028 reaches 19–21 mm in diameter in 7 d on PDA media.

Sarocladium terricola (J.H. Mill., Giddens & A.A. Foster) A. Giraldo, Gené & Guarro (Figure 8(C))

MycoBank: MB 807950 **Family** *Sarocladiaceae*

Basionym: Fusidium terricola J.H. Mill., Giddens & A.A. Foster 1958

Materials examined: South Korea. Jeju-do, Jeju-si (33°23′53″N 126°14′24″E), August 15 2021, isolated

from Gelidium sp. (Macroalga), M.S. Park, J.S. Kim & Y.W. Lim (SFC20240607-M008, stored in a metabolically inactive state); Jeollanam-do, Muan-gun (35°3'43.65"N 126°20'13.91"E), July 17 2017, isolated M.S. Park & Y.W. from sea sand, (SFC20240607-M009, stored in a metabolically inactive state); Gyeongsangbuk-do, Ulleung-gun (37°32′2.05″N 30°49′24.87″E), unknown date 2018, isolated from sponge, M.S. Park & Y.W. Lim (SFC20240607-M012, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on OA Mycelia smooth, septate, hyaline; Conidiophores solitary or aggregated, erect, septate, hyaline, smooth, $1-2.7 \,\mu m$ wide. Phialides subulate, hyaline, smooth, schizophialides observed, adelophialides observed $11-32\times1.5-3.1 \,\mu m$ at the base. Conidia aseptate, fusiform, smooth, hyaline, eguttulate, truncate, arranged in chain, $3.1-6.3\times1.3-2.4 \,\mu m$ (av. $4.6\times1.8 \,\mu m$).

Culture characteristics: Colonies on PDA reaching 18–21 mm diam in 7 d at 25 °C; raised, plain, sometimes radially sulcate, felty, sometimes floccose at center, forward white to pale yellow (4A3); margin entire; reverse yellowish white (4A2). Colonies on MEA reaching 19–22 mm diam in 7 d at 25 °C raised, plain, sometimes radially sulcate, felty to floccose, forward white; margin entire; reverse light orange (5A4). Colonies on OA reaching 23–25 mm diam in 7 d at 25 °C; raised due to aerial mycelium, plain, felty, hairy to floccose, forward white; margin filiform; reverse yellowish white (3A2).

Notes: The morphological characteristics of the Korean strains resemble those described by Giraldo et al. [36]. While occurrences of the species have been commonly reported in terrestrial environments [36], in this study, the species was isolated from macroalgae, sea sand, and seawater. Notably, it was also discovered at a depth of 5,572 m in the western Pacific [72].

Sarocladium zeae (W. Gams & D.R. Sumner) Summerb. (Figure 8(D))

MycoBank: MB 519595 **Family** *Sarocladiaceae*

Basionym: Acremonium zeae W. Gams & D.R. Sumner, in Gams 1971

Materials examined: South Korea. Jeollanam-do, Gangjin-jun (34°26′49″N 126°49′07″E), 2016, isolated from *Fulvia mutica* (marine bivalve mollusks), M.S. Park & Y.W. Lim (SFC20240607-M013, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on OA Vegetative hyphae septate, hyaline, smooth- and thin-walled, 1.1–2.9 μm wide.

smooth-walled. Conidiophores hyaline, erect, Phialides arising directly from vegetative hyphae and aerial mycelia, acicular, subulate, 34-56×1.1-2.7 µm at the base, thin- and smooth-walled, hyaline; adelophialides and schizophialides not observed. Conidia unicellular, cylindrical, 3-9×0.9-2.4 µm, hyaline to subhyaline, smooth- and thin-walled.

Culture characteristics: Colonies on PDA reaching 28-30 mm diam in 7 d at 25 °C; flat, plain, felty, forward orange white (5A2), yellowish white (4A2) at margin; margin filiform; reverse pale yellow (4A3). Colonies on MEA reaching 31-33 mm diam in 7 d at 25°C flat, radially sulcate, rugose, felty, forward yellowish white (3A2), white at margin; margin fimbriate; reverse light orange (5A4). Colonies on OA reaching 29-33 mm diam in 7 d at 25 °C; crateriform due to the aerial mycelia at margin and middle, plain, floccose, felty, membranous, forward white to wax white (2B3); margin filiform; reverse yellowish white (4A2).

Notes: The Korean strain SFC20240607-M013, isolated from a marine bivalve (Fulvia mutica), clusters with strains of Sarocladium zeae in the phylogenetic tree. Compared to strains documented by Giraldo et al. [36], SFC20240607-M013 grows faster and produces longer spores on OA media.

Verruciconidia infuscata L.W. Hou, L. Cai & Crous (Figure 8(E))

MycoBank: MB845839 Family Bionectriaceae

Materials examined: South Korea. Jeollanam-do, (34°50′29.7′′N 127°29′09.4′′E), Suncheon-si 2021, isolated from sea sand, M.S. Park, J.S. Kim & Y.W. Lim (SFC20240607-M019, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on OA Mycelium branched, septate, hyaline; Sporulation abundant; Conidiophores solitaty or aggregate, unbranched or branched, erect, straight to flexuous, septate, smooth, hyaline, 1.2-3.3 µm wide, bearing 1-2 phialides per node. Phialides cylindrical or subulate, smooth-walled, hyaline, $26-48\times1.4-3\,\mu m$ at the base. Conidia aseptate, subglobose to broadly ellipsoidal, smooth, hyaline, 2.4- $3.5 \times 1.4 - 2.7 \,\mu\text{m}$ (av. $2.8 \times 1.9 \,\mu\text{m}$), conidia chains not observed. Chlamydospores not observed.

Culture characteristics: Colonies on PDA reaching 21-23 mm diam in 7 d at 25 °C; flat, radially sulcate, rugose, felty, forward pale yellow (2A3); margin entire; reverse pale yellow (3A3). Colonies on MEA reaching 16-17 mm diam in 7 d at 25 °C; flat, radially sulcate, rugose, membranous, forward white to hyaline; margin entire; reverse reddish orange (7B8) to light yellow (4A4). Colonies on OA

reaching 25-27 mm diam in 7 d at 25 °C; flat, plain, rugose, hairy, forward white to hyaline; margin filiform; reverse vellowish white (1A2).

Notes: The Korean strain SFC20240607-M019 does not produce pigment, whereas the holotype CBS H-24613 produces pigments on PDA, OA, and MEA media in old cultures [35]. Additionally, SFC20240607-M019 has shorter phialides conidia compared to the holotypes (38-50.5 and 3-5 μm long, respectively) [35].

Verruciconidia persicina (Nicot) L.W. Hou, L. Cai & Crous (Figure 8(F))

MycoBank: MB845840 **Family** Bionectriaceae

Basionym: Paecilomyces persicinus Nicot 1958

examined: Materials South Korea. (34°0′0.94"N 126°20′30.31″E), Chuja-myeon September 12 2017, isolated from Sargassum thunbergii (Macroalga), M.S. Park & Y.W. Lim (SFC20240607-M020, stored in a metabolically inactive state); Incheon, Ganghwa-gun (37°36'36.3"N 126°31′13.8″E), July 2021, isolated from mudflat, M.S. Park, J.S. Kim & Y.W. Lim (SFC20240607-M019, stored in a metabolically inactive state).

Description: Sexual morph undetermined. Asexual morph on OA Mycelium branched, septate, hyaline; Sporulation abundant; Conidiophores solitary or aggregate, unbranched or branched, erect, straight to flexuous, septate, smooth, hyaline, 1.2-3.3 µm wide, bearing 1-2(-5) phialides per node. Phialides subulate, smooth-walled rarely with a nodule, hyaline, $17-26 \times 1.5-2.8 \,\mu m$ at the base. Conidia aseptate, subglobose to broadly ellipsoidal, smooth, hyaline, $2.6-4.2\times2-3.2\,\mu\text{m}$ (av. $3.5\times2.7\,\mu\text{m}$), conidia chains not observed. Chlamydospores not observed.

Culture characteristics: Colonies on PDA reaching 17-19 mm diam in 7 d at 25 °C; flat, radially sulcate, velvety, felty, forward white to dull yellow (3B3), pale yellow (3A3); margin entire to slightly irregular; reverse butter yellow (4A5), pale orange (5A3), pale yellow (3A3) at margin. Colonies on MEA reaching 19-20 mm diam in 7 d at 25 °C; raised to convex, radially sulcate, floccose, felty, forward white; margin lobed; reverse light orange (5A5), grayish orange (5B5). Colonies on OA reaching 18-21 mm diam in 7 d at 25 °C; convex to umbonate due to abundant aerial mycelium, plain, felty, floccose, forward white, sometimes partly pastel yellow (3A4); margin entire to slightly irregular; exudate sometimes wax yellow (3B5); reverse pale orange (5A3) to pale yellow (4A3), sometimes dull red (8B3) at center.

Notes: The Korean strains, isolated from mudflat and green macroalga (Sargassum thunbergii), exhibit textures ranging from velvety to floccose compared to the dusty colonies of the holotype CBS H-6661 [35]. Additionally, the Korean strains have longer conidia, measuring $2.6-4.2 \,\mu\text{m}$, compared to those of the holotype $(4.2-6.2 \,\mu\text{m})$ [35].

5. Discussion

This study expanded our understanding of marine *Hypocreales* by identifying 21 species, including 5 new species and 16 unrecorded species in South Korea. Phylogenetic and morphological analyses strongly support that these five species are new to science, as they formed distinct clades separate from closely related species, with apparent morphological differences. Despite being based on a limited number of strains, this is undoubtedly remarkable, especially considering that only 131 species of *Hypocreales* have been previously reported from marine environments in South Korea (Marine Bio-Resource Information System, updated 2024.03.29).

In fungal taxonomy, standardized methods are limited for some groups, with most research focused on economically important genera like Penicillium and Trichoderma [70,73,74]. The International Code of Nomenclature for Fungi (ICNF) only requires confirmation of description for valid publication of new species, resulting in varying levels of taxonomic detail depending on the classification [75]. Some genera in this study, such as Purpureocillium and Metapochonia, had also limited taxonomic research, which led to fewer descriptive features being available [11,76]. To address the morphological plasticity of marine fungi, we used various media and multiple genetic markers to establish the starting point of a robust species description framework. Integrating detailed morphological observations with molecular data, this polyphasic approach provided a comprehensive dataset for marine Hypocreales. Some species exhibited morphological variability, which is often a response to environmental stressors, such as changes in size, growth rates, and textures [77,78]. Further studies on ecotypes and physiological adaptations are necessary to understand these phenomena better.

Most of the species identified in this study were isolated in sediments (Table 1), and only a few were associated with marine organisms such as marine animals and macroalgae. Ongoing debates question whether fungi found in sediments are "truly marine fungi" – meaning fungi actively functioning in marine environments [25,31,79]. However, recent meta-transcriptome data suggest that fungi, though not dominant, play roles like carbohydrate recycling in marine sediment [80–84]. Microscopy studies

have also provided evidence, such as the presence of branching cells, indicating fungal growth [85]. Although the specific roles of these fungi remain unclear, further research into their diversity, function, and whether they are invaders that have adapted to marine environments is essential.

This study is one of the first to apply a polyphasic approach to reveal the diversity and taxonomic placement of Hypocrealean fungi in Korean marine environments. Our findings highlight the high species diversity of Hypocreales in these environments and the limitations of relying solely on morphological characteristics or ITS-based single-marker analysis for species-level identification. This study also gives detailed descriptions of Hypocrealean fungi that have not been studied much before. These descriptions help us learn more about each species and are meant to be the basis for standardizing taxonomic studies for these groups. Furthermore, this research provides valuable insights for future studies on their ecological interactions, evolutionary relationships, and potential industrial applications.

Disclosure statement

The authors declare no conflict of interest.

Funding

This work was supported by the management of Marine Fishery Bio-resources Center (2024) funded by the National Marine Biodiversity Institute of Korea (MABIK).

ORCID

Wonjun Lee http://orcid.org/0000-0002-7227-0777
Ji Seon Kim http://orcid.org/0000-0003-1869-7347
Sumin Jo http://orcid.org/0009-0004-7236-1312
Chang Wan Seo http://orcid.org/0000-0002-5948-1836
Young Woon Lim http://orcid.org/0000-0003-2864-3449

References

- [1] Hyde KD, Norphanphoun C, Maharachchikumbura SSN, et al. Refined families of *Sordariomycetes*. Mycosphere. 2020;11(1):305–1059. doi: 10.5943/mycosphere/11/1/7.
- [2] Perera RH, Hyde KD, Jones EBG, et al. Profile of Bionectriaceae, Calcarisporiaceae, Hypocreaceae, Nectriaceae, Tilachlidiaceae, Ijuhyaceae fam. nov., Stromatonectriaceae fam. nov. and Xanthonectriaceae fam. nov. Fungal Divers. 2023;118(1):95–271. doi: 10.1007/s13225-022-00512-1.
- [3] Rogerson CT. The hypocrealean fungi (ascomycetes, Hypocreales). Mycologia. 1970;62(5):865–910. doi: 10.1080/00275514.1970.12019033.
- [4] Rossman AY, Seifert KA, Samuels GJ, et al. Genera in *Bionectriaceae*, *Hypocreaceae*, and *Nectriaceae* (*Hypocreales*) proposed for acceptance or rejection.

- IMA Fungus. 2013;4(1):41-51. doi: 10.5598/imafungus.2013.04.01.05.
- [5] Rossman AY. Towards monophyletic genera in the holomorphic Hypocreales. Stud Mycol. 2000;45:27-34.
- [6] Rossman AY, Samuels GJ, Rogerson CT, et al. Genera of Bionectriaceae, Hypocreaceae and Nectriaceae (Hypocreales, Ascomycetes). Stud Mycol. 1999;42:1-248.
- [7] Zhang W, Zhang X, Li K, et al. Introgression and gene family contraction drive the evolution of lifestyle and host shifts of hypocrealean fungi. Mycology. 2018;9(3):176-188. doi: 10.1080/21501203. 2018.1478333.
- [8] Dworecka-Kaszak B. Cordyceps fungi as natural killers, new hopes for medicine and biological control factors. Ann Parasitol. 2014;60(3):151-158.
- [9] Khan S, Guo L, Maimaiti Y, et al. Entomopathogenic fungi as microbial biocontrol agent. Mol Plant Breed. 2012;3(7):63-79. doi: 10.5376/mpb.2012.03.0007.
- [10] Zhang L, Fasoyin OE, Molnár I, et al. Secondary metabolites from hypocrealean entomopathogenic fungi: novel bioactive compounds. Nat Prod Rep. 2020;37(9):1181-1206. doi: 10.1039/c9np00065h.
- [11] Calvillo-Medina RP, Ponce-Angulo DG, Raymundo T, et al. Purpureocillium roseum sp. nov. A new ocular pathogen for humans and mice resistant to antifungals. Mycoses. 2021;64(2):162-173. doi: 10.1111/ myc.13198.
- [12] Perdomo H, Sutton DA, García D, et al. Spectrum of clinically relevant Acremonium species in the United States. J Clin Microbiol. 2011;49(1):243-256. doi: 10.1128/JCM.00793-10.
- [13] Salgado-Salazar C, Crouch JA. Genome resources for the stem and bark canker pathogens Corinectria fuckeliana, Neonectria hederae and N. punicea. Plant Dis. 2019;103(3):389-391. doi: 10.1094/PDIS-05-18-0904-A.
- [14] Shrestha B, Kubátová A, Tanaka E, et al. Spider-pathogenic fungi within Hypocreales (Ascomycota): their current nomenclature, diversity, and distribution. Mycol Prog. 2019;18(8):983-1003. doi: 10.1007/s11557-019-01512-3.
- [15] Calabon MS, Hyde KD, Jones EB, et al. Freshwater fungal numbers. Fungal Divers. 2022;114(1):3-235. doi: 10.1007/s13225-022-00503-2.
- [16] Calabon MS, Jones EBG, Pang KL, et al. Updates on the classification and numbers of marine fungi. Bot Mar. 2023;66(4):213-238. doi: 10.1515/bot-2023-0032.
- [17] Hu DM, Liu F, Cai L. Biodiversity of aquatic fungi China. Mycology. 2013;4(3):125-168. 10.1080/21501203.2013.835752.
- [18] Bao D-F, Hyde KD, Maharachchikumbura SSN, et al. Taxonomy, phylogeny and evolution of freshwater Hypocreomycetidae (Sordariomycetes). Fungal Divers. 2023;121(1):1-94. doi: 10.1007/s13225-023-00521-8.
- [19] Cai L, Hu D-M, Liu Fang Hyde KD, et al. The molecular phylogeny of freshwater Sordariomycetes and discomycetes. Freshwater mycology and fungal-like organisms. Berlin, Germany: Walter de Gruyer, GmbH; 2014.
- [20] Pang KL, Alias SA, Chiang MWL, et al. Sedecimiella taiwanensis gen. et sp. nov., a marine mangrove funthe Hypocreales (Hypocreomycetidae, Ascomycota). Bot Mar. 2010;53(6):493-498. doi: 10.1515/bot.2010.061.

- [21] Schoch CL, Sung GH, Volkmann-Kohlmeyer B, et al. Marine fungal lineages in the Hypocreomycetidae. Mycol Res. 2007;111(Pt 2):154-162. doi: 10.1016/j. mycres.2006.10.005.
- [22] Wurzbacher C, Kerr J, Grossart HP. Aquatic fungi.
- [23] Amend A, Burgaud G, Cunliffe M, et al. Fungi in the marine environment: open questions and unsolved problems. MBio. 2019;10(2):e01189-18. doi: 10.1128/mBio.01189-18.
- [24] Cunliffe M. Who are the marine fungi? Environ Microbiol. 2023;25(1):131-134. doi: 10.1111/1462-2920,16240.
- [25] Pang KL, Overy DP, Jones EBG, et al. Marine fungi'and 'marine-derived fungi'in natural product chemistry research: toward a new consensual definition. Fungal Biol Rev. 2016;30(4):163-175. doi: 10.1016/j.fbr.2016.08.001.
- [26] Wong MKM, Goh T-K, Hodgkiss IJ, et al. Role of fungi in freshwater ecosystems. Biodivers Conserv. 1998;7(9):1187-1206. doi: 10.1023/A:1008883716975.
- [27] da Silva MK, de Souza LMD, Vieira R, et al. Fungal and fungal-like diversity in marine sediments from the maritime Antarctic assessed using DNA metabarcoding. Sci Rep. 2022;12(1):21044. doi: 10.1038/ s41598-022-25310-2.
- [28] Hassett BT, Vonnahme TR, Peng X, et al. Global diversity and geography of planktonic marine fungi. Bot Mar. 2020;63(2):121-139. doi: 10.1515/bot-2018-0113.
- [29] Li P-D, Jeewon R, Aruna B, et al. Metabarcoding reveals differences in fungal communities between unflooded versus tidal flat soil in coastal saline ecosystem. Sci Total Environ. 2019;690:911-922. doi: 10.1016/j.scitotenv.2019.06.473.
- [30] Wong Chin JM, Puchooa D, Bahorun T, et al. Metabarcoding assessment of fungal diversity in brown algae and sponges of Mauritius. Front Microbiol. 2022;13:1003790. doi: 10.3389/ fmicb.2022.1003790.
- [31] Jones EB, Gareth Pang KL, Abdel-Wahab MA, et al. An online resource for marine fungi. Fungal Divers. 2019;96(1):347-433. doi: 10.1007/s13225-019-00426-5.
- [32] Hyde KD, Jones EBG, Leaño E, et al. Role of fungi marine ecosystems. **Biodivers** Conserv. 1998;7(9):1147-1161. doi: 10.1023/A:1008823515157.
- [33] Richards TA, Jones MDM, Leonard G, et al. Marine fungi: their ecology and molecular diversity. Ann Rev Mar Sci. 2012;4(1):495-522. doi: 10.1146/ annurev-marine-120710-100802.
- [34] Tian J, Lai D, Zhou L. Secondary metabolites from Acremonium fungi: diverse structures and bioactivities. Mini Rev Med Chem. 2017;17(7):603-632. doi: 10.2174/1389557516666160914194134.
- [35] Hou LW, Giraldo A, Groenewald JZ, et Redisposition of acremonium-like fungi Hypocreales. Stud Mycol. 2023;105(1):23-203. doi: 10.3114/sim.2023.105.02_supp.
- [36] Giraldo A, Gené J, Sutton DA, Madrid H, et al. Phylogeny of Sarocladium (Hypocreales). Persoonia-Mol Phylogen Evol Fungi. 2015;34(1):10-24.
- [37] Lee W, Kim JS, Seo CW, et al. Diversity of Cladosporium (Cladosporiales, Cladosporiaceae) species in marine environments and report on five new

- - species. MycoKeys. 2023b;98:87-111. doi: 10.3897/ mycokeys.98.101918.
- [38] Lee S, Park MS, Lee H, et al. Fungal diversity and enzyme activity associated with the macroalgae, Agarum clathratum. Mycobiology. 2019;47(1):50-58. doi: 10.1080/12298093.2019.1580464.
- [39] Lee S, Park MS, Lim YW. Diversity of marine-derived Aspergillus from tidal mudflats and sea sand in Mycobiology. 2016;44(4):237-247. 10.5941/MYCO.2016.44.4.237.
- [40] Lee IW, Seo CW, Lee W, et al. Diversity and dynamics of marine arenicolous fungi in three seasides of the Korean peninsula. J Microbiol. 2023a;61(1):63-82. doi: 10.1007/s12275-023-00011-1.
- [41] Park MS, Oh SY, Lee S, et al. Fungal diversity and enzyme activity associated with sailfin sandfish egg masses in Korea. Fungal Ecol. 2018;34:1-9. doi: 10.1016/j.funeco.2018.03.004.
- [42] Rogers SO, Bendich AJ. Extraction of total cellular DNA from plants, algae and fungi. Plant molecular biology manual. Dordrecht, the Netherlands: Kluwer Academic Publishers; 1994. p. 183-190.
- [43] Gardes M, Bruns TD. ITS primers with enhanced specificity for basidiomycetes-application to the identification of mycorrhizae and rusts. Mol Ecol. 1993;2(2):113-118. doi: 10.1111/j.1365-294x.1993. tb00005.x.
- [44] White TJ, Bruns Thomas Lee Sjwt, Taylor J. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. PCR protocol guide to methods and applications. Vol. 18. Cambridge (MA): Academic Press, Inc.; 1990. p. 315-322.
- [45] Kearse M, Moir R, Wilson A, et al. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. Bioinformatics. 2012;28(12):1647-1649. doi: 10.1093/bioinformatics/bts199.
- [46] Seo CW, Kim SH, Lim YW, et al. Re-identification on Korean Penicillium sequences in GenBank collected by software GenMine. Mycobiology. 2022;50(4):231-237. doi: 10.1080/12298093.2022.2116816.
- [47] Crous PW, Akulov A, Balashov S, et al. New and interesting fungi. 6. Fungal Syst Evol. 2023;11(1):109-156. doi: 10.3114/fuse.2023.11.09.
- [48] Xiao YP, Wang YB, Hyde KD, et al. Polycephalomycetaceae, a new family of clavicipitoid fungi segregates from Ophiocordycipitaceae. Fungal Divers. 2023;120(1):1-76. doi: 10.1007/s13225-023-00517-4.
- [49] Katoh K, Misawa K, Kuma KI, et al. MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform. Nucleic Acids Res. 2002;30(14):3059-3066. doi: 10.1093/nar/gkf436.
- [50] Nirengerg HI. Untersuchungen uber die morpholoigische und biologische Differnzierung in der Fusarium-Sektion Liseola. Mitteilungen aus der biologischen bundesanstalt für land- und forstwirtschaft Berlin-Dahlem. Berlin: Kommissionsverlag Paul Parey; Vol. 169. 1976. p. 1-117.
- [51] Schneider CA, Rasband WS, Eliceiri KW. NIH Image to ImageJ: 25 years of image analysis. Nat Methods. 2012;9(7):671-675. doi: 10.1038/nmeth.2089.
- [52] Kornerup A, Wanscher JH. Methuen handbook of colour. 3rd ed. London: E. Methuen and Co. Ltd; 1978.

- [53] Lechat C, Fournier J. Two new species of Lasionectria (Bionectriaceae, Hypocreales) from Guadeloupe and Martinique (French West Indies). Mycotaxon. 2013;121(1):275-280. doi: 10.5248/121.275.
- [54] Lombard L, Houbraken Jos Decock Cony Samson RA, Meijer Martin Réblová M, et al. Generic hyper-diversity in Stachybotriaceae. Persoonia-Mol Phylogen Evol Fungi. 2016;36(1):156-246.
- [55] Hyde KD, Tennakoon DS, Jeewon R, et al. Fungal diversity notes 1036-1150: taxonomic and phylogenetic contributions on genera and species of fungal taxa. Fungal Divers. 2019;96(1):1-242. doi: 10.1007/ s13225-019-00429-2.
- [56] Seifert KA. A monograph of stilbella and some allied hyphomycetes. The Netherlands: Centraalbureau voor Schimmelcultures Baarn; 1985.
- [57] Rehner SA, Minnis AM, Sung G-H, et al. Phylogeny and systematics of the anamorphic, entomopathogenic genus Beauveria. Mycologia. 2011;103(5):1055-1073. doi: 10.3852/10-302.
- [58] Wang Y, Tang DX, Duan DE, et al. Morphology, molecular characterization, and virulence of Beauveria pseudobassiana isolated from different hosts. J Invertebr Pathol. 2020;172:107333. doi: 10.1016/j. jip.2020.107333.
- [59] Kepler RM, Luangsa-Ard JJ, Hywel-Jones NL, et al. A phylogenetically-based nomenclature for Cordycipitaceae (Hypocreales). IMA Fungus. 2017;8(2):335-353. doi: 10.5598/imafungus.2017.08.02.08.
- [60] Zimmermann G. The entomopathogenic fungi Isaria farinosa (formerly Paecilomyces farinosus) and the Isaria fumosorosea species complex (formerly Paecilomyces fumosoroseus): biology, ecology and use in biological control. Biocontrol Sci Technol. 2008;18(9):865-901. doi: 10.1080/09583150802471812.
- [61] Hagestad OC, Hou L, Andersen JH, et al. Genomic characterization of three marine fungi, including Emericellopsis atlantica sp. nov. with signatures of a generalist lifestyle and marine biomass degradation. Fungus. 2021;12(1):21. doi: s43008-021-00072-0.
- Nirenberg HI, O'Donnell K. New Fusarium species and combinations within the Gibberella fujikuroi species complex. Mycologia. 1998;90(3):434-458. doi: 10.1080/00275514.1998.12026929.
- [63] Zare R, Gams W. The genera Lecanicillium and Simplicillium gen. nov. Nova_Hedwigia. 2001;73(1-2):1-50. doi: 10.1127/nova.hedwigia/73/2001/1.
- [64] Schroers H-J, Samuels GJ, Zhang N, et al. Epitypification of Fusisporium (Fusarium) solani and its assignment to a common phylogenetic species in the Fusarium solani species complex. Mycologia. 2016;108(4):806-819. doi: 10.3852/15-255.
- [65] Short DPG, O'Donnell K, Thrane U, et al. Phylogenetic relationships among members of the Fusarium solani species complex in human infections and the descriptions of F. keratoplasticum sp. nov. and F. petroliphilum stat. nov. Fungal Genet Biol. 2013;53:59-70. doi: 10.1016/j.fgb.2013.01.004.
- [66] Gams W, Stielow B, Gräfenhan T, et al. The ascomycete genus Niesslia and associated monocillium-like anamorphs. Mycol Prog. 2019;18(1-2):5-76. doi: 10.1007/s11557-018-1459-5.



- [67] Crous PW, Hernández-Restrepo M, Schumacher RK, et al. New and interesting fungi. 4. Fungal Syst Evol. 2021;7(1):255-343. doi: 10.3114/fuse.2021.07.13.
- [68] Crous PW, Luangsa-Ard JJ, Wingfield MJ, et al. Fungal Planet description sheets: 785-867. Persoonia-Mol Phylogen Evol Fungi. 2018;41(1):238-417. doi: 10.3767/persoonia.2018.41.12.
- [69] Kim JS, Kim SH, Lee W, et al. Five previously unrecorded fungal species isolated from marine plastic wastes in South Korea. Mycobiology. 2022;50(6):420-428. doi: 10.1080/12298093.2022.2152951.
- [70] Visagie CM, Houbraken J, Frisvad Jens Christian Hong SB, et al. Identification and nomenclature of the genus Penicillium. Stud Mycol. 2014;78(1):343-371. doi: 10.1016/j.simyco.2014.09.001.
- [71] Perdomo H, Cano Josep Gené J, García D, et al. Polyphasic analysis of Purpureocillium lilacinum isolates from different origins and proposal of the new species Purpureocillium lavendulum. Mycologia. 2013;105(1):151-161. doi: 10.3852/11-190.
- [72] Xie Y, Wang Y, Zhang K, et al. Saromacrophorins A-C, rare sesquiterpene-hydroquinone hybrids from the deep-sea-derived fungus Sarocladium terricola 494 a. Eur J Org Chem. 2024;27(5):e202301250. doi: 10.1002/ejoc.202301250.
- [73] Cai F, Druzhinina IS. In honor of John Bissett: authoritative guidelines on molecular identification of Trichoderma. Fungal Divers. 2021;107(1):1-69. doi: 10.1007/s13225-020-00464-4.
- [74] Samson RA, Visagie CM, Houbraken J, et al. Phylogeny, identification and nomenclature of the genus Aspergillus. Stud Mycol. 2014;78(1):141-173. doi: 10.1016/j.simyco.2014.07.004.
- [75] Turland NJ, Wiersema J, Harry Barrie FR, et al. International code of nomenclature for algae, fungi, and plants (Shenzhen Code) adopted by the International Nineteenth Botanical Congress Shenzhen, China, July 2017. Glashütten, Germany: Koeltz Botanical Books; 2018.
- [76] Kondo N, Tokiwa T, Sato K, et al. Metapochonia hahajimaensis (Clavicipitaceae, Hypocreales), a new species from soil in Hahajima Island, Tokyo, Japan.

- Mycoscience. 2020;61(6):337-341. doi: 10.1016/j. myc.2020.06.001.
- [77] Ahumada-Rudolph R, Novoa V, Becerra J. Morphological response to salinity, temperature, and pH changes by marine fungus Epicoccum nigrum. Environ Monit Assess. 2018;191(1):35. doi: 10.1007/ s10661-018-7166-5.
- [78] Francisco CS, Ma X, Zwyssig MM, et al. Morphological changes in response to environmental stresses in the fungal plant pathogen Zymoseptoria tritici. Sci Rep. 2019:9(1):9642. doi: 10.1038/ s41598-019-45994-3.
- [79] Jones EB, Gareth Ramakrishna S, Vikineswary S, et al. How do fungi survive in the sea and respond to climate change? JoF. 2022;8(3):291. doi: 10.3390/ jof8030291.
- [80] Broman E, Sachpazidou V, Dopson M, et al. Diatoms dominate the eukaryotic metatranscriptome during spring in coastal 'dead zone' sediments. Proc R Soc B. 2017;284(1864):20171617. doi: 10.1098/rspb.2017.1617.
- [81] Orsi W, Biddle JF, Edgcomb V. Deep sequencing of subseafloor eukaryotic rRNA reveals active fungi across marine subsurface provinces. PLoS One. 2013;8(2):e56335. doi: 10.1371/journal.pone.0056335.
- [82] Orsi WD, Edgcomb VP, Christman GD, et al. Gene expression in the deep biosphere. 2013;499(7457):205-208. doi: 10.1038/nature12230.
- [83] Orsi WD, Vuillemin A, Coskun ÖK, et al. Carbon assimilating fungi from surface ocean to subseafloor revealed by coupled phylogenetic and stable isotope analysis. ISME J. 2022;16(5):1245-1261. doi: 10.1038/ s41396-021-01169-5.
- [84] Shekarriz E, Chen J, Xu Z, et al. Disentangling the functional role of fungi in cold seep sediment. Microbiol Spectr. 2023;11(2):e01978-01922. 10.1128/spectrum.01978-22.
- Pachiadaki MG, Rédou V, Beaudoin DJ, et al. Fungal and prokaryotic activities in the marine subsurface biosphere at Peru Margin and Canterbury Basin inferred from RNA-based analyses and microscopy. Front Microbiol. 2016;7:846. doi: 10.3389/fmicb.2016.